

Genome-wide Association and Longitudinal Analyses Reveal Genetic Loci Linking Pubertal Height Growth, Pubertal Timing, and Childhood Adiposity

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Abstract

The pubertal height growth spurt is a distinctive feature of childhood growth reflecting both the central onset of puberty and local growth factors. While little is known about the underlying genetics, growth variability during puberty correlates with adult risks for hormone-dependent cancer and adverse cardiometabolic health. The only gene so far associated with pubertal height growth, *LIN28B*, pleiotropically influences childhood growth, puberty, and cancer progression, pointing to shared underlying mechanisms.

To discover genetic loci influencing pubertal height and growth and place them in context of overall growth and maturation, we performed genome-wide association (GWA) meta-analyses in up to 18,737 European samples utilizing longitudinally collected height measurements. We found significant associations ($P < 1.67 \times 10^{-8}$) at 10 loci, including *LIN28B*. Five loci associated with pubertal timing, all impacting multiple aspects of growth. In particular, a novel variant correlated with expression of *MAPK3*, and associated both with increased prepubertal growth and earlier menarche. Another variant near *ADCY3-POMC* associated with increased BMI, reduced pubertal growth, and earlier puberty.

While epidemiological correlations suggest that early puberty marks a pathway from rapid prepubertal growth to reduced final height and adult obesity, our study shows that individual loci associating with pubertal growth have variable longitudinal growth patterns that may differ from epidemiological observations. Overall this study uncovers part of the complex genetic architecture linking pubertal height growth, the timing of puberty, and childhood obesity, and provides new information to pinpoint processes linking these traits.

Introduction

Postnatal height growth is a heritable complex process characterized by rapid infant growth, slowly diminishing mid-childhood growth, and a distinct pubertal height growth spurt.

While the genetics of adult stature has been evaluated in large-scale GWA analyses (1), few studies have addressed the molecular underpinnings of distinct growth phases. Moreover, specific growth patterns during childhood correlate with both altered pubertal timing and adult health risks. For example, increased height and BMI prior to puberty correlate with advanced pubertal onset (2,3,4), and early puberty associates with increased risk for adult obesity and related metabolic traits (5,6,7). Still, the specific mechanisms linking these traits remain elusive.

To elucidate part of the genetic architecture impacting adolescent growth, we focused primarily on the dynamic and highly variable pubertal growth spurt, which reflects both the activation of central puberty and local growth factors (8-9) while accounting for up to 15-20% of adult stature (10). Narrow-sense heritability estimates place the genetic contribution to variation in pubertal growth between 60%-90% (11,12,13), and twin studies suggest a substantial proportion of shared genetic variance with other phases of childhood growth (13). Specifically, we aimed to 1) identify genetic variants associated with the onset, total magnitude, and tail end of the pubertal growth spurt, and 2) investigate these variants' longitudinal effects on overall childhood growth and the timing of puberty (study design outlined in **Figure 1**).

Due to large variation in the timing and rate of the pubertal growth spurt (shown schematically in **Figure 2**), an accurate model typically requires frequent height measurements spanning a large age range, often difficult to obtain. Furthermore, girls enter puberty, and thus begin their growth spurt, an average of two years earlier than boys. Taking these challenges into consideration, we aimed to characterize loci influencing growth during puberty by leveraging heterogeneous height measurements taken at varied ages throughout childhood across participating cohorts to maximize statistical power. Therefore, we modeled the pubertal height growth spurt for GWA using three partially correlated simple measures (**Table S1; Figure 2**) that also partly reflect the timing of puberty (14). In Analysis I, we targeted the take-off phase of the growth spurt (height standard deviation

score (SDS) at 10 years in girls and 12 years in boys) by reasoning that increased height relative to the population mean in early puberty reflects either overall genetic height potential or entrance into the pubertal growth spurt. Because a large proportion of adult stature is achieved prior to the onset of puberty, we expected a significant part of the detected variants to associate with overall height growth potential, while a minority would have specific pubertal timing effects. In Analysis II, we assessed the overall contribution of growth across puberty to adult height (height change SDS between 8 years and adult), which reflects the total magnitude of growth during the pubertal growth spurt. Finally, in Analysis III, we approximated the timing of peak height velocity by looking at the height change SDS between age 14 years and adult, since early maturing individuals grow less during late adolescence than late-maturing individuals, who still have much of their remaining growth to achieve after age 14. Similar simple height measurements across puberty have previously proven robust in the GWA setting for detecting common genetic variation influencing both height growth and pubertal timing (15).

Results

Discovery and follow-up meta-analyses reveal 10 genome-wide significant loci associated with pubertal growth

Nine cohorts contributed partly overlapping population-based samples (**Table S2**) with childhood height measurements and approximately 2.5 million directly genotyped or imputed SNPs to three discovery GWA analyses (**Table S3**), in which we meta-analyzed data from males and females both separately and combined for the three models. We observed significant deviation from the expected distribution of *P*-values for all three combined-gender analyses (I, II, and III), males and females separately for Analysis I, and females only for Analysis II (**Figure S1a**).

All three models resulted in genome-wide significant loci, although we had most power (**Table S4**) to detect loci for Analysis I (height SDS at age 10 yrs in girls and 12 yrs in boys) (**Table 1**). In total, 9 loci contained markers that reached *P*-values below the genome-wide significance threshold corrected for testing three primary phenotypes ($P < 1.67 \times 10^{-8}$, after

genomic control). Of these, only rs7759938 nearby *LIN28B* was previously known to influence pubertal growth (15, 16).

Due to the requirement of Analyses II and III to have both childhood and adult height measurements for the same individuals, there were no additional samples available for follow-up of suggestive signals for these Analyses. Thus, we only performed follow-up for Analysis I. An additional six cohorts comprising up to 9,710 samples were available for follow-up of the 22 suggestive signals ($P < 1 \times 10^{-5}$) for Analysis I (**Table S5**). Joint analysis of discovery and follow-up stages for Analysis I robustly confirmed a single novel variant, rs4788196 ($P = 9.49 \times 10^{-11}$, $n = 18,737$; **Table 1**), thus bringing the number of loci reaching the genome-wide significance threshold to 10, of which 7 were associated with Analysis I.

eQTL analysis links rs4788196 (G) to decreased expression of nearby gene *MAPK3* and pathway analyses highlight the TGF-beta signaling pathway and pathways in cancer

To link the identified association signals with putative biological processes, we tested all significantly associated gene regions for association with leukocyte gene expression levels and performed gene pathway analyses. eQTL analysis in whole blood (17) linked rs960273 with the gene *GNA12*, as previously reported (1), as well as highlighting a previously unknown role for the extracellular signal-regulated kinase 1 (*MAPK3*, also known as *ERK1*) in prepubertal height growth (**Table S6**). More specifically, the adolescent height-increasing allele (G) at rs4788196 on 16p11.2 (Analysis I) correlated with decreased expression of *MAPK3*, consistent with previous studies linking deactivation of the gene with increased bone growth in mice (18). We subsequently performed pathway analyses using the g:Profiler Gene Group Functional Profiler tool (g:GOS (19); **Table S7a**) and MAGENTA Gene Set Enrichment Analyses (GSEA (20); **Table S7b**), which commonly highlighted the TGF-beta signaling pathway and pathways in cancer for loci identified in Analysis I. Whereas g:Profiler identified the MAPK-pathway, the GSEA showed enrichment of lower than expected P -values for genes belonging to the *TOB1* pathway, although the individual implicated gene regions were only suggestively associated in Analysis I.

The novel locus near *MAPK3* associates transiently with height growth in childhood and earlier menarche

While there are no published studies implicating *MAPK3* in human height growth, rare recurrent CNVs near *MAPK3* on chromosome 16p11.2 have been shown to associate with early onset obesity (21, 22). Nonetheless, the adolescent height effect that we observed did not appear to be CNV-mediated (**Table S8**). To characterize the *MAPK3* variant in more depth, we evaluated the longitudinal height and BMI effects of rs4788196. We plotted the effect size (beta) against 6 age bins across puberty from 8 years to adult (**Figure 3a**), and investigated early height yearly from ages 1 to 4 (**Table S9**). These analyses revealed a transient effect on height growth for the G allele from age 4 in both males and females that was diluted by adulthood, with no apparent effect on BMI (**Figure S2**). Finally, since rapid growth during childhood and early adolescence may correlate with early timing of sexual maturation (8), we also tested rs4788196 for association with age at menarche (AAM) (23), and found that the height-increasing allele associated with earlier AAM ($P=1.42 \times 10^{-4}$, surpassing the significance threshold of 0.007, which corresponds to a Bonferroni-correction accounting for follow-up of 7 loci not previously associated with AAM; **Table S10**).

Five of the discovered pubertal growth loci are also associated with pubertal timing, eight are adult stature loci, and one is a BMI locus

Even though the *MAPK3*-locus associated with both prepubertal height growth and the timing of puberty, it showed little evidence for association with adult anthropometric traits (**Table 2**). Nonetheless, epidemiological data support phenotypic correlations between earlier pubertal timing, increased adult obesity and decreased final height. To better understand how the loci we detected contribute to a genetic link underlying these traits, we performed a systematic analysis of all leading SNPs significantly associated with pubertal height and growth and noted substantial evidence for overlap (**Figure 4**). Assessing the pubertal timing effect, both based on published GWA of age at menarche (23) and by *in silico* meta-analysis of leading signals not previously implicated in the timing of menarche, as described above, showed that all three discovery Analysis approaches detected pubertal timing loci (near *MAPK3*, *PXMP3*, *VGLL3*, *ADCY3-POMC*, and *LIN28B*). Moreover, 8 signals overlapped with adult stature loci (1), of which *ADCY3-POMC* has further been implicated in

childhood (24) and adult obesity (25). However, two of our signals within 1 Mb of reported height loci showed partial linkage disequilibrium ($r^2 < 0.6$) with previously published SNPs. Conditioning for the previously reported marker revealed evidence for a further independent association nearby *CABLES1* (rs6507528; $P=0.00011$) (**Table S11**). This variant may represent allelic heterogeneity or partially tag the same causative variant (26).

Longitudinal analyses across puberty shows that the identified pubertal growth loci represent both overall growth potential and pubertal timing

To further evaluate the leading signals, we compared their height effects longitudinally across puberty, revealing multiple distinct growth trajectories. This approach divided the loci associated with various measures of pubertal height and growth into two groups based on association with pubertal timing. One group of loci (near *ZBTB38*, *EFEMP1*, *CABLES1*, *ADAMTSL3*, and *GNA12*), not associated with pubertal timing, all impacted height SDS across multiple growth phases, strongly and steadily from prepuberty to adulthood (**Figure 3c**). Thus, these loci likely reflect overall growth potential, rather than puberty-specific effects. In contrast, the five pubertal timing-associated variants displayed diverse effects on the timing and tempo of growth, both before and during puberty (**Table 2**; **Figure 3a and Figure 3b**).

The locus associating with increased childhood BMI also associates with decreased pubertal growth but not with prepubertal height

Of the pinpointed loci associating with pubertal growth and timing, only one had previously been associated with BMI (**Figure S2**). The strong correlation between childhood obesity and prepubertal height (2,3,4) and between prepubertal height and AAM (27) predicts that the BMI-associated marker, rs1172294 (*ADCY3-POMC*), would associate with increased prepubertal stature. However, the variant showed no association with stature before puberty (**Figure 3b**). Nonetheless, the BMI-increasing allele (G) was associated with earlier menarche ($P=8.64 \times 10^{-6}$) and a decline in pubertal growth in both males and females, as expected. Consistently, other variants previously associated with childhood obesity (24) showed a parallel between elevated BMI and diminished growth across puberty (**Table S12**). We also found that rs3817334 (*MTCH2*), previously associated with adult (25) but not childhood BMI, also associated with the same decrease in overall pubertal growth.

Discussion

Taken together, the simple approach used in this study to model the pubertal growth spurt for GWAS in more than 18,000 study subjects of European descent identified 10 significantly associated pubertal growth loci. Utilizing unique longitudinal childhood measurements, we described the distinct height growth and BMI effects of these variants across puberty, and noted significant longitudinal associations at each locus. More specifically, half of the identified loci also associated with pubertal timing, and provided evidence linking a robust novel growth and menarche locus with *MAPK3* expression levels. Despite prior association between several genes in the MAPK-pathway and skeletal growth syndromes (28, 29, 30, 31) *MAPK3* has not been associated with human height before. Interestingly, gonadotropin-releasing hormone (GnRH), crucial for regulating the onset of puberty, activates *MAPK3* (32), providing a putative biological link between rs4788196 and pubertal timing.

This study has several strengths. It is based on a large dataset of rather unique longitudinal height data from multiple well-characterized study cohorts. The two-stage approach applied in this study enabled wide-ranging characterization of growth and maturation phenotypes associated with the 10 leading association signals. Nonetheless, one limitation is that the height data available for analysis varied between the cohorts. Furthermore, the height measurements available in most of the cohorts were not frequent enough to allow detailed modeling of the pubertal growth spurt. In order to overcome the lack of very frequent height measurements and the variability of height assessments between cohorts, we chose to adopt an analysis strategy aiming to maximize the number of study subjects. By utilizing three simple and robust height growth estimates to model the pubertal growth spurt, in addition to applying rigorous statistical significance thresholds, we were successfully able to identify and characterize novel loci significantly associated with pubertal height growth. As expected, a proportion of these loci also associated with pubertal timing, as assessed by age of menarche, and with adult height.

In fact, our data affirm a complex genetic architecture underlying growth, pubertal timing, and adiposity. In particular, specific genetic effects may contradict epidemiological correlations. Epidemiological studies have observed a developmental pattern linking taller

prepubertal stature to earlier puberty, accelerated skeletal maturation and short adult stature due to early cessation of growth (2,3). While the majority of loci we assessed showed the expected parallel association between early menarche and decreased overall pubertal height growth, their prepubertal height effects varied. Three variants (near *MAPK3*, *PXMP3*, and *VGLL3*) followed the expected pattern, linking taller prepubertal stature with earlier AAM, while the early puberty-associated allele (T) at rs7759938 (*LIN28B*) correlated with shorter prepubertal childhood height, as reported previously (15).

The relationship between puberty and adult stature is similarly complex; whereas epidemiological studies show a correlation between early puberty and reduced adult height (3), a genetic association study found that early puberty alleles may associate with either increased or decreased adult stature (23). An example is rs7846385 (*PXMP3*), for which, contradictory to the predicted pattern, the early-menarche allele associates with increased adult height. Our results show that tall adult height is achieved because the early-menarche allele (C) also influences tall childhood height and a limited reduction of total pubertal growth. These data thus agree with a recent candidate gene study suggesting that loci associated with adult height may have a stronger influence on prepubertal growth than during the pubertal growth spurt (16). However, utilizing a genome-wide approach with greater sample sizes, our study identifies loci previously missed that specifically target pubertal growth, and we find that they are associated with diverse and unique longitudinal growth patterns. We also find that not all loci influencing pubertal growth also impact adult stature.

Additionally, epidemiological studies link increased childhood adiposity with advanced puberty and increased prepubertal height. While all childhood BMI-increasing alleles assessed in this study also showed an association with decreased overall pubertal growth, at the *ADCY3-POMC* locus, the same allele associated with both earlier puberty and increased childhood BMI, but not with prepubertal stature. The correlation between obesity and pubertal growth may be consequential of hormonal changes associated with childhood adiposity. However, since the same association pattern was also present at a locus uniquely associated with adult BMI (*MTCH2*), an underlying shared genetic effect remains likely.

Given the complexity of the relationships between these developmental traits, tracking unique gene effects across multiple growth periods may help elucidate specific pathways linking childhood events to adult outcomes, as illustrated here with height growth, pubertal timing, and adult stature. While epidemiological studies have described correlations between distinct childhood growth events and adult health, genetically defined association patterns may pinpoint molecular processes linking these traits. Characterization of these pathways may thus provide new insight towards a better understanding of the relationships between early growth patterns, pubertal timing, and adult disease risk.

Materials and Methods

Phenotypes and study subjects. Discovery study subjects were included from cohorts participating in the Early Growth Genetics (EGG) Consortium, namely the Avon Longitudinal Study of Parents and Children (ALSPAC), 1958 British Birth Cohort (BC58-T1DGC and BC58-WTCCC), Cardiovascular Risk in Young Finns Study (YFS), Helsinki Birth Cohort Study (HBCS), Lifestyle- Immune System- Allergy Plus Environment and Genetics Study (LISApplus), Northern Finland Birth Cohort 1966 (NFBC1966), Queensland Institute of Medical Research (QIMR), and Western Australia Pregnancy Study (RAINE). Cohort-specific details for all analyses can be found in **Table S3**. The data annotation, exchange and storage have been facilitated by the SIMBioMS platform (33).

Three primary phenotypes were analyzed, which were defined as follows:

Analysis I: Single height: Girls with height measurements available at age 10 (± 1 y) and boys with height measured at age 12 (± 1 y) were included. Sex-specific SD scores were calculated within each study by dividing the within-population height mean by the SD. A total of 14,040 samples (7,161 males and 6,879 females) from 9 contributing cohorts were included.

Analysis II: Total pubertal growth: Individuals with a childhood height measurement at age 8 (± 1 y) and at adulthood (≥ 18 years of age) were included. Height difference was calculated between the two measurements and sex-specific SD scores of this difference were calculated within each study by dividing the within-population mean difference in

height by the SD. 6 cohorts with up to 10,799 samples (5,043 males and 5,756 females) contributed to the Analysis.

Analysis III: Late pubertal growth: Subjects with a height measurement in adolescence at age 14 (± 1 y) and at adulthood (≥ 18 years of age) were included. Height difference was calculated between the two measurements and logarithm-transformed sex-specific Z-scores were calculated within each study. Log transformation was performed prior to SD score calculation, again by dividing the population-specific mean log-transformed height difference by the SD. 5 cohorts with up to 9,228 subjects (4,282 males and 4,946 females) were included in this analysis.

Genotyping and quality control. Genome-wide genotypes were obtained using high-density SNP arrays on Illumina and Affymetrix platforms. Before imputation, SNPs with minor allele frequency of $<1\%$, call rate $<95\%$, or Hardy-Weinberg Equilibrium $p < 1 \times 10^{-6}$ were excluded. Samples were also excluded if they contained duplicates, excess heterozygosity, non-European ancestry, or ambiguous gender. Imputation was performed using IMPUTE (34) or MACH (35) for roughly 2.5 million SNPs against HapMap Phase II (release 21/22). Imputed SNPs were filtered prior to meta-analysis to exclude poorly imputed SNPs (IMPUTE filter PROPER INFO < 0.4 , MACH filter $r^2 < 0.3$).

Genome-wide association analyses. Within each cohort, association analyses were performed by linear regression using an additive model across genotyped and imputed SNPs (dosages), for males and females separately. For all analyses, age at adolescent measurement was included as a covariate where available, and the first two principal components were adjusted within each study sample if necessary. For the association tests, PLINK (36), ProbABEL (27), SNPtest (34), or MACH2QTL(35) were used.

Meta-analyses. A fixed effects inverse-variance meta-analysis model was used to test the effect of each variant on height, total pubertal growth, or late pubertal growth separately for males and females. Sex-specific results from each study were also meta-analyzed for each phenotype in three combined-gender analyses. The R package MetABEL (38) (v.2.11.1) was used to perform all meta-analyses. MetABEL corrects each individual result for its respective genomic inflation factor (λ) according to the genomic control method for population stratification. Subsequently, an additional genomic control correction was

applied using the overall genomic inflation factor calculated for each of the nine meta-analyzed results. The threshold for genome-wide significance was set at a conservative Bonferroni-corrected threshold of $P < 1.67 \times 10^{-8}$, accounting for testing three primary phenotypes. A further significance threshold of 0.002 (accounting for examining 10 loci in males and females) was applied to all follow-up analyses unless otherwise stated.

Conditional analyses. To determine whether our signals represent independent effects on growth during puberty from previously reported related phenotypes, we performed linear regression using an additive model on the primary pubertal growth phenotypes, adjusting each of the six markers (imputed genotype dose) for the previously reported marker nearby our signal. As in the primary analysis, age at adolescent measurement (where available), and optional adjustment for population substructure, were included as covariates (**Table S3**).

Follow-up analyses of suggestive association signals. Genetic markers yielding association P -values of 1×10^{-5} to 1.67×10^{-8} in Analysis I and not previously associated with related traits adult stature, AAM, or BMI, were selected for follow-up genotyping ($n=22$). Additional cohorts participated in the follow-up analyses, including *in-silico* analyses by Avon Longitudinal Study of Parents and Children (ALSPAC; follow-up sample), Children's Hospital of Philadelphia (CHOP), Finnish Twin Cohort Study (FTCS), Genome-Wide Population-Based Association Study of Extremely Overweight Young Adults (GOYA), and Lifestyle- Immune System- Allergy study & German Infant Study on the influence of Nutrition Intervention plus environment and genetics (LISApplus & GINIplus; follow-up sample). Association results for a marker showing borderline significance, rs281379, were also provided by Netherlands Twin Registry (NTR). *De novo* genotyping was done for selected markers (success rate >98%) from Northern Finland Birth Cohort 85-86 (NFBC8586) with TaqMan Pre-Designed SNP Genotyping Assays on LightCycler 480 Real-Time PCR System (Roche) according to the manufacturer's instructions at the Finnish Genome Center (Helsinki, Finland).

Statistical analysis in replication samples was performed similarly as in the discovery analyses with PLINK (36), ProbABEL (37), or SNPtest (34), using linear regression models for each of the 22 markers under an additive model, with age at adolescent measurement and correction for population substructure as optional covariates. Genomic control-corrected

discovery results were meta-analyzed together with the individual linear regression results from contributing cohorts for each SNP, using the MetABEL (38) package of R (v.2.11.1).

CNV analysis of 16p11.2. CNVs were genotyped using signal intensity distributions and B-allele frequency of the genotyping probes with PennCNV software (39) and adjusted for genomic waves according to genomic GC content, as previously described (40). The CNV scan was completed for 2,310 individuals in YFS and 4,931 in NFBC1966 (41).

Expression quantitative trait loci (eQTLs). We queried significant SNPs from the Dietary, Lifestyle, and Genetic determinants of Obesity and Metabolic syndrome (DILGOM) study, an extension of FINRISK 2007. The eQTL methods are previously published (17). Briefly, whole blood was extracted from 518 unrelated individuals and genotyped on the Illumina 610-Quad SNP array. In parallel, mRNA expression was quantified with Illumina HumanHT-12 Expression BeadChips. Linear regression was used to test association between transcript expression levels and each SNP.

Pathway analyses. We entered the nearest gene to all signals at $P < 1 \times 10^{-4}$ (one per locus) into the g:Profiler Gene Group Functional Profiler tool (g:GOST) (19), a webtool that, briefly, queries databases of biological pathways for enrichment of user-entered genes. For Analysis I, we also entered *MAPK3* because the gene was implicated by eQTL evidence to be functionally relevant ($n=93$, corresponding to 0.0003% of all discovery signals for Analysis I). We also ran GSEA using MAGENTA (20), a program that calculates a P -value for each gene in the genome based on GWA results, and then searches biological databases for pathways showing an enrichment of genes with lower than expected P -values. Analysis II and III data are not reported here due to the lack of significant findings.

Association analyses of age of menarche. To assess the relevance of the pubertal growth-associated variants for pubertal timing, leading signals not previously implicated in the timing of menarche were queried from *in silico* meta-analysis data of 87,802 women published by the ReproGen Consortium (23).

Cross-sectional height and BMI analyses. Height or BMI measurements from childhood to adulthood were divided into 6 age bins: 1) Pre-puberty (6.5-8.5 years old), 2) early puberty (8.6-10.5 yrs old), 3) mid-puberty for females (10.6-12.5 yrs old), 4) mid-puberty for males

(12.6-14.5 yrs old), 5) late puberty (14.6-17.5 yrs old) and finally 6) adult (>17.6 yrs old). In each cohort, each marker of interest (imputed genotype dosage) was tested for association with sex-specific height or BMI SDS for all age bins available, using linear regression assuming an additive model and adjusting for exact age at measurement (to the nearest month), along with optional correction for population stratification. A single measurement was included per study subject per bin, with the age closest to the mean used when more than one measurement was available. Altogether 23 SNPs were analyzed for height and BMI across pubertal growth (only significantly associated markers are reported here). Summary statistics were meta-analyzed like the primary analyses in each age bin, separately for males and females, for both height and BMI distinctly. Effect sizes were plotted versus age.

Early growth analyses. Cohorts with height measurements available at 1, 2, 3, or 4 years were included, namely The Children's Hospital of Philadelphia (CHOP), Copenhagen Study on Asthma in Childhood (COPSAC), Generation R Study (Generation R), Helsinki Birth Cohort Study (HBCS), Infancia y Medio Ambiente (Environment and Childhood) Project (INMA), Lifestyle – Immune System – Allergy Study & German Infant Study on the influence of Nutrition Intervention plus environment and genetics (LISApplus&GINIpplus), Netherlands Twin Register (NTR), Northern Finland Birth Cohort 1966 (NFBC1966), Prevention and Incidence of Asthma and Mite Allergy birth cohort study (PIAMA), and Western Australian Pregnancy study (RAINE). Length was measured at 12 months (range 6-18 months) and height at 24 (range 18-30), 36 (range 30-42) and 48 (range 42-54) months. If multiple measurements per individual were available, those closest to 12, 24, 36 or 48 months were used. Sex- and age-adjusted SD scores were calculated using Growth Analyser 3.0 (Dutch Growth Research Foundation, Rotterdam, the Netherlands) in each study separately (42). The sex-specific association between each marker genotype and length or height SDS was assessed using linear regression, assuming an additive model. Imputed genotypes were used where directly-assayed genotypes were unavailable. We meta-analyzed the within-cohort sex stratified linear regression results using the inverse-variance method. A fixed-effects model was assumed, using RMeta in R (v.2.7.0).

URLs. SIMBioMS, <http://www.simbioms.org/>; R, <http://www.r-project.org/>; PLINK, <http://pngu.mgh.harvard.edu/~purcell/plink/>; IMPUTE, <http://mathgen.stats.ox.ac.uk/impute/impute.html>; MACH, <http://www.sph.umich.edu/csg/abecasis/MACH/index.html>; The International HapMap Project, <http://hapmap.ncbi.nlm.nih.gov/>; GenABEL, <http://www.genabel.org/>; SNPtest, https://mathgen.stats.ox.ac.uk/genetics_software/snpTest/snpTest.html; g:Profiler, <http://biit.cs.ut.ee/gprofiler/>; MAGENTA, <http://www.broadinstitute.org/mpg/magenta/>; Growth Analyser, <http://www.growthanalyser.org>.

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References

1. Lango Allen, H., Estrada, K., Lettre, G., Berndt, S.I., Weedon, M.N., Rivadeneira, F., Willer, C.J., Jackson, A.U., Vedantam, S., Raychaudhuri, S., *et al.* (2010) Hundreds of variants clustered in genomic loci and biological pathways affect human height. *Nature*, **467**, 832-838.
2. Cooper, C., Kuh, D., Egger, P., Wadsworth, M. and Barker, D. (1996) Childhood growth and age at menarche. *Br. J. Obstet. Gynaecol.*, **103**, 814-817.
3. He, Q. and Karlberg, J. (2001) BMI in childhood and its association with height gain, timing of puberty, and final height. *Pediatr. Res.*, **49**, 244-251.
4. Aksglaede, L., Juul, A., Olsen, L.W. and Sørensen, T.I.A. (2009) Age at puberty and the emerging obesity epidemic. *PLoS ONE* **4**, e8450.
5. Silventoinen, K., Baker, J.L. and Sørensen, T.I.A. (2012) Growth in height in childhood and risk of coronary heart disease in adult men and women. *PLoS ONE*, **7**, e30476.
6. Widén, E., Silventoinen, K., Sovio, U., Ripatti, S., Cousminer, D.L., Hartikainen, A.L., Laitinen, J., Pouta, A., Kaprio, J., Järvelin, M.R., *et al.* (2012) Pubertal timing and growth influences cardiometabolic risk factors in adult males and females. *Diabetes Care* **35**, 850-856.
7. Lakshman, R., Forouhi, N., Luben, R., Bingham, S., Khaw, K., Wareham, N. and Ong, K.K. (2008) Association between age at menarche and risk of diabetes in adults: Results from the EPIC-norfolk cohort study. *Diabetologia*, **51**, 781-786.
8. Tanner, J.M., Whitehouse, R.H., Marubini, E. and Resele, L.F. (1976) The adolescent growth spurt of boys and girls of the Harpenden growth study. *Ann. Hum. Biol.*, **3**, 109-126.
9. Perry, R.J., Farquharson, C. and Ahmed, S.F. (2008) The role of sex steroids in controlling pubertal growth. *Clin. Endocrinol.*, **68**, 4-15.
10. Luo, Z.C. and Karlberg, J. (2000) Critical growth phases for adult shortness. *Am. J. Epidemiol.*, **152**, 125-131.
11. Sharma, J.C. (1983) The genetic contribution to pubertal growth and development studied by longitudinal growth data on twins. *Ann. Hum. Biol.*, **10**, 163-171.
12. Morris, D.H., Jones, M.E., Schoemaker, M.J., Ashworth, A. and Swerdlow, A.J. (2011) Familial concordance for age at menarche: Analyses from the Breakthrough Generations Study. *Paediatr. Perinat. Epidemiol.*, **25**, 306-311.
13. Silventoinen, K., Pietiläinen, K.H., Tynelius, P., Sørensen, T.I., Kaprio, J. and Rasmussen, F. (2008) Genetic regulation of growth from birth to 18 years of age: The Swedish Young Male Twins Study. *Am. J. Hum. Biol.*, **20**, 292-298.

14. Wehkalampi, K., Silventoinen, K., Kaprio, J., Dick, D.M., Rose, R.J., Pulkkinen, L. and Dunkel, L. (2008) Genetic and environmental influences on pubertal timing assessed by height growth. *Am. J. Hum. Biol.*, **20**, 417-423.
15. Widén, E., Ripatti, S., Cousminer, D.L., Surakka, I., Lappalainen, T., Järvelin, M.R., Eriksson, J.G., Raitakari, O., Salomaa, V., Sovio, U., *et al.* (2010) Distinct variants at LIN28B influence growth in height from birth to adulthood. *Am. J. Hum. Genet.*, **86**, 773-782.
16. Vrieze, S.I., McGue, M., Miller, M.B., LeGrand, L.N., Schork, N.J., and Iacono, W.G. (2011) An assessment of the individual and collective effects of variants on height using twins and a developmentally informative study design. *PLoS Genet.*, **7**, e1002413.
17. Inouye, M., Kettunen, J., Soininen, P., Silander, K., Ripatti, S., Kumpula, L.S., Hämäläinen, E., Jousilahti, P., Kangas, A.J., Männistö, S., *et al.* (2010) Metabonomic, transcriptomic, and genomic variation of a population cohort. *Mol. Syst. Biol.*, **6**, 441.
18. Sebastian, A., Matsushita, T., Kawanami, A., Mackem, S., Landreth, G.E. and Murakami, S. (2011) Genetic inactivation of ERK1 and ERK2 in chondrocytes promotes bone growth and enlarges the spinal canal. *J. Orthop. Res.*, **29**, 375-379.
19. Reimand, J., Kull, M., Peterson, H., Hansen, J. and Vilo, J. (2007) g:Profiler--a web-based toolset for functional profiling of gene lists from large-scale experiments. *Nucleic Acids Res.*, **35**, W193-200.
20. Segre, A.V., DIAGRAM Consortium, MAGIC investigators, Groop, L., Mootha, V.K., Daly, M.J. and Altshuler, D. (2010) Common inherited variation in mitochondrial genes is not enriched for associations with type 2 diabetes or related glycemic traits. *PLoS Genet.*, **6**, e1001058.
21. Walters, R.G., Jacquemont, S., Valsesia, A., de Smith, A.J., Martinet, D., Andersson, J., Falchi, M., Chen, F., Andrieux, J., Lobbens, S., *et al.* (2010) A new highly penetrant form of obesity due to deletions on chromosome 16p11.2. *Nature*, **463**, 671-675.
22. Jacquemont, S., Reymond, A., Zufferey, F., Harewood, L., Walters, R.G., Kutalik, Z., Martinet, D., Shen, Y., Valsesia, A., Beckmann, N.D., *et al.* (2011) Mirror extreme BMI phenotypes associated with gene dosage at the chromosome 16p11.2 locus. *Nature*, **478**, 97-102.
23. Elks, C.E., Perry, J.R., Sulem, P., Chasman, D.I., Franceschini, N., He, C., Lunetta, K.L., Visser, J.A., Byrne, E.M., Cousminer, D.L., *et al.* (2010) Thirty new loci for age at menarche identified by a meta-analysis of genome-wide association studies. *Nat. Genet.*, **42**, 1077-1085.
24. Bradfield, J.P., Taal, H.R., Timpson, N.J., Scherag, A., Lecoeur, C., Warrington, N.M., Hyponen, E., Holst, C., Valcarcel, B., Thiering, E., *et al.* (2012) A genome-wide association meta-analysis identifies new childhood obesity loci. *Nat. Genet.*, **44**, 526-531.

25. Speliotes EK, Willer CJ, Berndt SI, Monda KL, Thorleifsson G, et al. (2010) Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. *Nat Genet* 42(11): 937-948.
26. Wood, A.R., Hernandez, D.G., Nalls, M.A., Yaghootkar, H., Gibbs, J.R., Harries, L.W., Chong, S., Moore, M., Weedon, M.N., Guralnik, J.M., et al. (2011) Allelic heterogeneity and more detailed analyses of known loci explain additional phenotypic variation and reveal complex patterns of association. *Hum. Mol. Genet.*, **20**, 4082-4092.
27. Okasha, M., McCarron, P., McEwen, J. and Smith, G.D. (2001) Age at menarche: secular trends and association with adult anthropometric measures. *Ann. Hum. Biol.*, **28**, 68-78.
28. Rousseau, F., Bonaventure, J., Legeai-Mallet, L., Pelet, A., Rozet, J.M., Maroteaux, P., Le Merrer, M. and Munnich, A. (1994) Mutations in the gene encoding fibroblast growth factor receptor-3 in achondroplasia. *Nature*, **371**, 252-254.
29. Shiang, R., Thompson, L.M., Zhu, Y.Z., Church, D.M., Fielder, T.J., Bocian, M., Winokur, S.T. and Wasmuth, J.J. (1994) Mutations in the transmembrane domain of FGFR3 cause the most common genetic form of dwarfism, achondroplasia. *Cell*, **78**, 335-342.
30. Bellus, G.A., McIntosh, I., Smith, E.A., Aylsworth, A.S., Kaitila, I., Horton, W.A., Greenhaw, G.A., Hecht, J.T. and Francomano, C.A. (1995) A recurrent mutation in the tyrosine kinase domain of fibroblast growth factor receptor 3 causes hypochondroplasia. *Nat. Genet.*, **10**, 357-359.
31. Aoki, Y., Niihori, T., Kawame, H., Kurosawa, K., Ohashi, H., Tanaka, Y., Filocamo, M., Kato, K., Suzuki, Y., Kure, S., et al. (2005) Germline mutations in HRAS proto-oncogene cause Costello syndrome. *Nat. Genet.*, **37**, 1038-1040.
32. Liu, F., Austin, D.A., Mellon, P.L., Olefsky, J.M. and Webster, N.J. (2002) GnRH activates ERK1/2 leading to the induction of c-fos and LHBeta protein expression in LbetaT2 cells. *Mol. Endocrinol.*, **16**, 419-434.
33. Kretyaninova, M., Zarins, A., Viksna, J., Kurbatova, N., Rucevskis, P., Neogi, S.G., Gostev, M., Perheentupa, T., Knuuttila, J., Barrett, A., et al. (2009) A system for information management in BioMedical studies--SIMBioMS. *Bioinformatics*, **25**, 2768-2769.
34. Marchini, J., Howie, B., Myers, S., McVean, G. and Donnelly, P. (2007) A new multipoint method for genome-wide association studies by imputation of genotypes. *Nat. Genet.*, **39**, 906-913.
35. Li, Y., Willer, C.J., Ding, J., Scheet, P. and Abecasis, G.R. (2010) MaCH: Using sequence and genotype data to estimate haplotypes and unobserved genotypes. *Genet. Epidemiol.*, **34**, 816-834.

36. Purcell, S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M.A., Bender, D., Maller, J., Sklar, P., de Bakker, P.I., Daly, M.J., *et al.* (2007) PLINK: A tool set for whole-genome association and population-based linkage analyses. *Am. J. Hum. Genet.*, **81**, 559-575.
37. Aulchenko, Y.S., Struchalin, M.V. and van Duijn, C.M. (2010) ProbABEL package for genome-wide association analysis of imputed data. *BMC Bioinformatics*, **11**, 134.
38. Aulchenko, Y.S., Ripke, S., Isaacs, A. and van Duijn, C.M. (2007) GenABEL: An R library for genome-wide association analysis. *Bioinformatics*, **23**, 1294-1296.
39. Wang, K., Li, M., Hadley, D., Liu, R., Glessner, J., Grant, S.F., Hakonarson, H. and Bucan, M. (2007) PennCNV: An integrated hidden markov model designed for high-resolution copy number variation detection in whole-genome SNP genotyping data. *Genome Res.*, **17**, 1665-1674.
40. Diskin, S.J., Li, M., Hou, C., Yang, S., Glessner, J., Hakonarson, H., Bucan, M., Maris, J.M. and Wang, K. (2008) Adjustment of genomic waves in signal intensities from whole-genome SNP genotyping platforms. *Nucleic Acids Res.*, **36**, e126.
41. Pietilainen, O.P., Rehnstrom, K., Jakkula, E., Service, S.K., Congdon, E., Tilgmann, C., Hartikainen, A.L., Taanila, A., Heikura, U., Paunio, T., *et al.* (2011) Phenotype mining in CNV carriers from a population cohort. *Hum. Mol. Genet.*, **20**, 2686-2695.
42. Fredriks, A.M., van Buuren, S., Burgmeijer, R.J., Meulmeester, J.F., Beuker, R.J., Brugman, E., Roede, M.J., Verloove-Vanhorick, S.P., Wit, J.M. (2000) Continuing positive secular growth change in the Netherlands 1955-1997. *Pediatr. Res.*, **47**, 316-323.

Figure Legends

Figure 1. Study design. We performed a 2-stage study to detect and characterize loci influencing the pubertal phase of childhood growth. Stage 1 consisted of locus-mapping using a genome-wide association approach on three related simple measurements of the pubertal growth spurt, and joint analysis of discovery and follow-up studies of novel variants for height SDS at age 10 yrs in girls and 12 yrs in boys (Analysis I). Validated loci were then characterized using a variety of methods in Stage 2, including genetic characterization (conditioned analysis on previously reported nearby SNPs in low or partial pairwise linkage disequilibrium (LD) with pubertal growth signals), functional characterization (eQTL analysis of pubertal growth SNPs on the expression levels of nearby genes and pathway analysis on biological pathways using g:Profiler and MAGENTA), characterization of the growth and maturational effects of the identified loci (on height and body mass index across puberty, on the timing of menarche for those signals not previously associated with age at menarche (AAM), and on early height from 1 to 4 years for loci which influence the timing of menarche and pubertal growth) and the association between BMI-increasing alleles and total pubertal growth.

Figure 2. Schematic picture of postnatal height and the three partly correlated GWA phenotypes describing pubertal growth. Childhood and pubertal growth rates at the 3rd, 50th and 97th percentile are shown for girls in the left panel and boys in the right panel. Growth rates during puberty vary as a consequence of variable timing of the growth spurt. The black growth curves illustrate a growth pattern representing the mean timing of the pubertal growth spurt, whereas two standard deviations early (-2SD) and late (+2SD) timing of the pubertal growth spurt are shown in dark vs. light orange in girls and dark vs. light blue in boys. The three genome-wide analysis strategies are illustrated in the bottom of each panel. Analysis I aims at capturing the take-off phase of the pubertal growth spurt and includes a single height measurement relative to the population mean (height SDS) at age 10 yrs in girls and 12 yrs in boys. Analyses II measures the change in relative height between age 8 yrs and adulthood, and targets the total magnitude of pubertal growth. Analysis III, measuring the relative height change between age 14 yrs and adult, targets the late end of the pubertal spurt in height growth, which varies depending on the timing of peak height velocity of the growth spurt.

Figure 3. Height across multiple growth periods for 10 pubertal growth loci. The effect size of linear regression of height SDS against genotype at 6 age bins from prepuberty to adulthood were plotted longitudinally across adolescence for males (blue) and females (red). a) *MAPK3* b) Pubertal growth loci not associated with the timing of menarche are shown for the height-increasing allele. All of these variants are also associated with adult stature. c) Pubertal growth loci associated with pubertal timing, shown for the menarche-advancing allele. These loci show divergent association to prepubertal growth. The x-axis represents age and the y-axis is the effect size (beta) of the association between the indicated allele and relative height at each age. * $P < 0.002$

Figure 4. Loci associated with pubertal height and their overlap with partially correlated phenotypes. The 10 genome-wide significant loci were assessed for their association with the correlated traits adult stature, pubertal timing (age at menarche), and adiposity (body mass index) by examining previously published literature. Additionally, for loci not previously associated with pubertal timing, we queried the leading SNPs in genome-wide association data of age at menarche by the ReproGen Consortium. All of the pubertal growth loci showed pleiotropic associations to one or more related phenotype.

Table 1. Pubertal growth loci reaching genome-wide significance ($P < 1.67 \times 10^{-8}$)

							Discovery		Follow-up			Joint Discovery + Follow-up			
SNP	Nearby gene(s)	Previously associated related trait [Ⓜ]	Chr.	Position (bp) [Ⓜ]	Effect allele/ Other allele [Ⓜ]	Effect allele freq.	Relative height change β (S.E.) ^c	<i>P</i> ^d	<i>P</i> _{het}	<i>P</i> _{sex-het}	Relative height change β (S.E.) ^c	<i>P</i>	Relative height change β (S.E.) ^c	<i>P</i>	<i>n</i>
Analysis I: Single height															
Height SDS at age 10 in females and age 12 in males combined															
rs6764769	ZBTB38, RASA2	H	3	142582970	G/A	0.45	0.08 (0.012)	4.6 x 10 ⁻¹⁰	0.52	0.35					13960
rs7846385	PXMP3, PKIA	H, AAM	8	78322734	C/T	0.26	0.09 (0.014)	5.27 x 10 ⁻¹⁰	0.13	0.13					13942
rs1346789	EFEMP1	H	2	55945556	C/T	0.22	-0.08 (0.015)	1.15 x 10 ⁻⁸	0.41	0.85					13960
rs6507528	CABLES1	H	18	19025578	G/A	0.55	-0.09 (0.016)	1.31 x 10 ⁻⁸	0.44	0.2					13160
rs1365198	ADAMTSL3	H	15	82189907	G/T	0.76	0.08 (0.014)	1.5 x 10 ⁻⁸	0.66	0.41					13946
rs4788196	MAPK3	Novel	16	29874935	G/A	0.44	0.06 (0.012)	6.38 x 10 ⁻⁷	0.51	0.43	0.08 (0.021)	4.25 x 10 ⁻⁵	0.07 (0.011)	9.49 x 10 ⁻¹¹	18737
Height SDS at age 12 in males only															
rs960273	GNA12	H	7	2824402	C/T	0.32	-0.1 (0.018)	2.51 x 10 ⁻⁸	0.96	5.17x10 ⁻⁴					6986
Analysis II: Pubertal growth															
Height change SDS (8-adult) in females and males combined															
rs1172294	ADCY3, DNAJC27, POMC	H, BMI	2	25022704	G /A	0.45	-0.08 (0.014)	1.02 x 10 ⁻⁸	0.87	0.92					10799
Height change SDS (8-adult) in females only															
rs7628864	VGLL3	AAM	3	86933308	G/A	0.38	-0.11 (0.019)	3.17 x 10 ⁻⁹	0.76	6.83x10 ⁻⁶					5756
Analysis III: Late pubertal growth															
Height change SDS (14-adult) in females and males combined															
rs7759938	LIN28B	H, AAM	6	105485647	C/T	0.32	0.11 (0.016)	3.87 x 10 ⁻⁹	0.23	0.23					8863

[Ⓜ] Previously associated related traits are adult stature (H), age at menarche (AAM), or body mass index (BMI).

[Ⓜ] Marker position reported according to Build 36 and allele coding based on the positive strand.

^c Effect sizes are change in height or growth SDS score.

^d P value adjusted for genomic control.

^e P value assessed by t-test for sexual heterogeneity.

Table 2. Association of pubertal growth variants with height growth across puberty

SNP (allele) Nearby Gene	PREPUBERTAL HEIGHT			HEIGHT AT EARLY PUBERTY			TOTAL PUBERTAL GROWTH			LATE ADOLESCENT GROWTH			ADULT HEIGHT ^b			GIANT height ^c
	Height SDS at 7-8 yrs			Analysis I Height SDS at 10 yrs in females and 12 yrs in males			Analysis II Height SDS change 8 yrs-adult			Analysis III Height SDS change 14 yrs-adult			Adult stature SDS			
	Beta	SE	P	Beta	SE	P ^a	Beta	SE	P ^a	Beta	SE	P ^a	Beta	SE	P	P, N
rs7846385 (C) <i>PXMP3</i>																
Females	0.090	0.016	3.73 x 10⁻⁸	0.096	0.018	8.71 x 10⁻⁸	-0.057	0.021	0.006	-0.043	0.023	0.068	0.03	0.019	0.114	2.988 x 10 ⁻¹⁰ , 133772
Males	0.058	0.016	0.031	0.067	0.019	5.46 x 10⁻⁴	-0.009	0.009	0.696	-0.068	0.026	0.008	0.038	0.02	0.055	
Combined	0.074	0.012	1.48 x 10⁻¹⁰	0.082	0.013	3.04 x 10⁻¹⁰	-0.035	0.015	0.022915	-0.054	0.017	0.002	0.034	0.014	0.014	
rs7628864 (G) <i>VGLL3</i>																
Females	0.051	0.015	0.001	0.044	0.016	0.007	-0.112	0.019	3.17 x 10⁻⁹	-0.059	0.021	0.006	-0.029	0.017	0.087	0.6118, 133795
Males	0.009	0.015	0.549	0.023	0.018	0.201	0.011	0.020	0.578	-0.001	0.022	0.969	0.028	0.017	0.113	
Combined	0.030	0.011	0.004	0.035	0.012	0.004	-0.054	0.014	9.41 x 10⁻⁵	-0.031	0.015	0.043	0.00	0.012	0.967	
rs7759938 (T) <i>LIN28B</i>																
Females	-0.042	0.014	0.003	0.004	0.017	0.792	-0.027	0.020	0.169	-0.112	0.022	2.89 x 10⁻⁷	-0.054	0.018	0.002	8.691 x 10 ⁻¹⁸ , 133774
Males	-0.046	0.014	0.001	-0.011	0.018	0.547	-0.020	0.021	0.327	-0.074	0.023	0.002	-0.045	0.018	0.011	
Combined	-0.044	0.010	8.80 x 10⁻⁶	-0.003	0.012	0.804	-0.024	0.014	0.097	-0.094	0.016	3.87 x 10⁻⁹	-0.05	0.013	1.0 x 10⁻⁴	
rs4788196 (G) <i>MAPK3</i>																
Females	0.047	0.015	0.001	0.059	0.017	8.72 x 10⁻⁴	-0.0003	0.018	0.958	-0.025	0.021	0.231	0.03	0.017	0.081	0.003336, 133818
Males	0.061	0.015	4.51 x 10⁻⁵	0.066	0.017	1.49 x 10⁻⁴	0.003	0.02	0.867	-0.046	0.023	0.04	0.039	0.017	0.023	
Combined	0.054	0.011	3.56 x 10⁻⁷	0.063	0.012	6.38 x 10⁻⁷	0.001	0.014	0.921	-0.035	0.015	0.023	0.035	0.012	0.004	
rs1172294 (G) <i>ADCY3</i>																
Females	-0.001	0.013	0.945	-0.006	0.017	0.718	-0.078	0.019	3.16 x 10⁻⁵	-0.046	0.021	0.028	-0.047	0.017	0.005	1.7 x 10 ⁻¹³ , 133780
Males	-0.016	0.014	0.246	-0.045	0.017	0.01	-0.08	0.02	5.97 x 10⁻⁵	0.01	0.023	0.673	-0.045	0.017	0.008	
Combined	-0.008	0.01	0.404	-0.026	0.012	0.04	-0.079	0.014	1.02 x 10⁻⁸	-0.02	0.015	0.184	-0.046	0.012	1.3 x 10⁻⁴	

^aP-values taken from genome-wide association discovery analyses, genomic-control corrected.^bMeta-analysis of association results for adult stature taken from the cohorts participating in the longitudinal analyses.^cAssociation to adult stature taken from publically available results of the GIANT Consortium adult height meta-analysis

(http://www.broadinstitute.org/collaboration/giant/index.php/GIANT_consortium_data_files.) Beta estimates were not available for public download.

All effect sizes are given for the menarche-advancing allele. In bold are associations reaching a P -value threshold of 0.002 (Bonferroni-corrected threshold accounting for follow-up analysis of 5 markers in 5 analyses).

Abbreviations

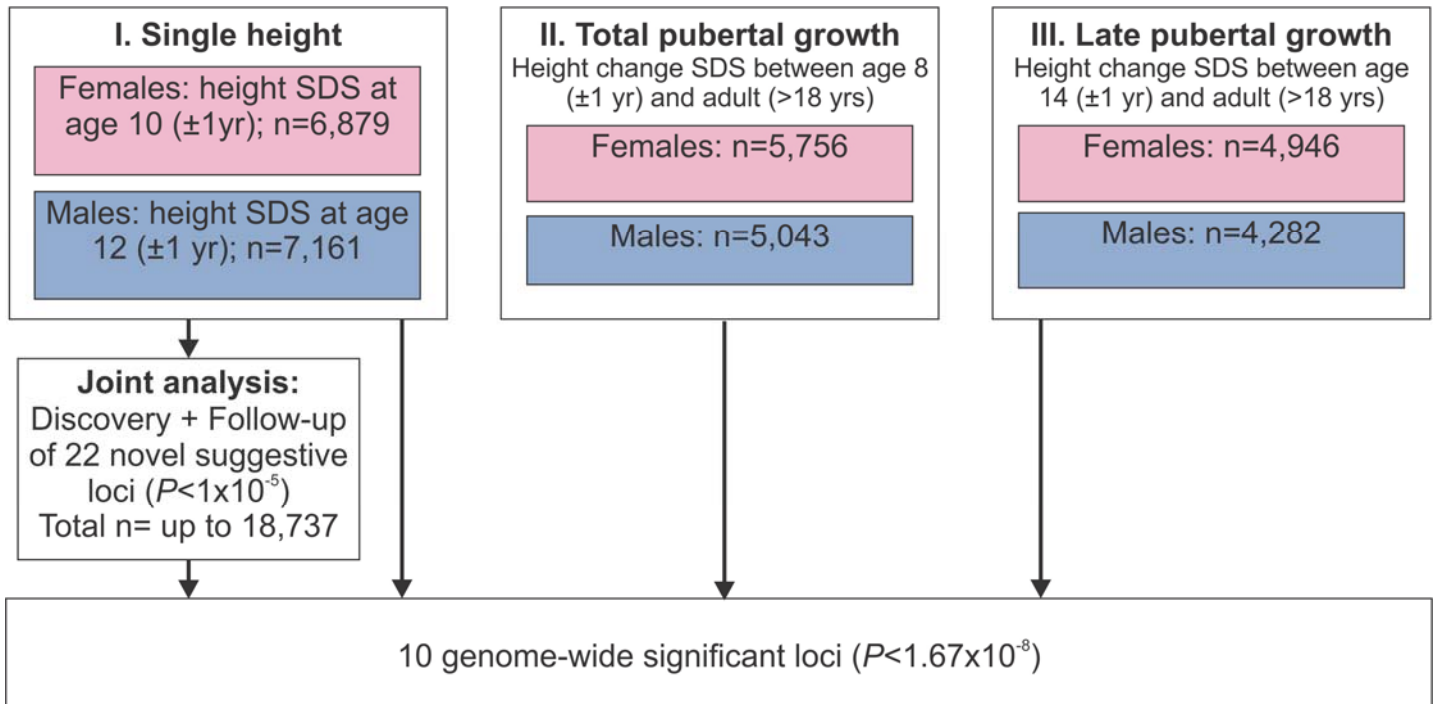
AAM Age at menarche

BMI Body mass index

GWAS Genome-wide association study

SDS Standard deviation score

Stage I. Discovery Genome-Wide Association Meta-Analyses



Stage II. Characterization of Identified Loci

