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Every cell in our body contains a copy of our genome, over 20,000 genes, 3 billion letters of DNA.			
DNA consists of two strands, twisted into a double helix and held together by a simple pairing rule. A pairs with T, and G pairs with C. Our genes shape who we are as individuals and as a species.			

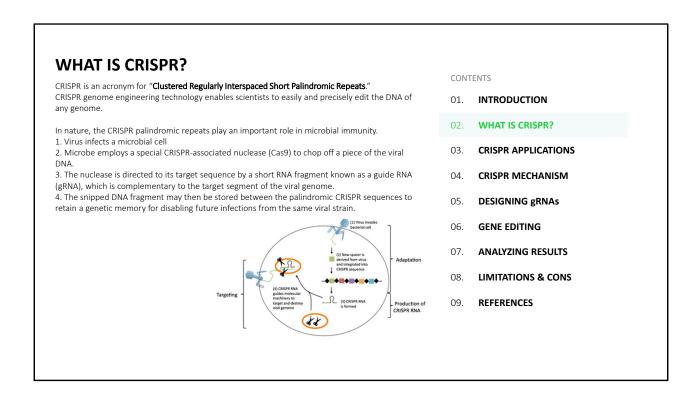
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Genes also have profound effects on health, and thanks to advances in DNA sequencing, researchers have identified thousands of genes that affect our risk of disease. To understand how genes work, researchers need ways to control them. Changing genes in living cells is not easy, but recently a new method has been developed that promises to dramatically improve our ability to edit the DNA of any species, including humans.



Introducing CRISPR, a system that has revolutionized the field of genome engineering with limitless applications in disease therapeutics, drug discovery, agriculture, biofuels, and much more. This revolutionary breakthrough has completely change the way science is performed, and it will soon change many aspects of our everyday lives.



What is CRISPR?

CRISPR is an acronym for "Clustered Regularly Interspaced Short Palindromic Repeats." CRISPR genome engineering technology enables scientists to easily and precisely edit the DNA of any genome.

In nature, the CRISPR palindromic repeats play an important role in microbial immunity. When a virus infects a microbial cell, the microbe employs a special CRISPR-associated nuclease (Cas9) to chop off a piece of the viral DNA. The nuclease is directed to its target sequence by a short RNA fragment known as a guide RNA (gRNA), which is complementary to the target segment of the viral genome. The snipped DNA fragment may then be stored between the palindromic CRISPR sequences to retain a genetic memory for disabling future infections from the same viral strain.

Once scientists learned how the CRISPR system worked in bacteria, they figured out how to reprogram it to allow efficient editing in any species.

DEFINITION

CRISPR gene editing is a genetic engineering technique in molecular biology by which the genomes of living organisms may be modified. It is based on a simplified version of the bacterial CRISPR antiviral defense system. By delivering the Cas9 nuclease complexed with a synthetic guide RNA (gRNA) into a cell, the cell's genome can be cut at a desired location, allowing existing genes to be removed and/or new ones added in vivo.

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CRISPR APPLICATIONS

CRISPR has already shown promising results for treating several diseases and has also had a huge impact in editing plant genomes, facilitating several beneficial agricultural applications. The CRISPR technology has also shown potential in other areas such as developing disease models and biofuels.

During the past years, the technology has been used to

- Functionally inactivate genes in human cell lines and cells
- Study Candida albicans
- Modify yeasts used to make biofuels
- Genetically modify crop strains.
- Studies on changing mosquitos so they cannot transmit diseases such as malaria.
- CRISPR based approaches utilizing Cas12a have recently been utilized in the successful
 modification of a broad number of plant species.

In July 2019, doctors in Tennessee, United States, used CRISPR to experimentally treat a patient with a genetic disorder. The patient was a 34-year-old woman with sickle cell disease

In the future, CRISPR gene editing could be used to create new species or revive extinct species from closely related ones.

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CRISPR MECHANISM

The beauty of CRISPR lies in its simplicity. CRISPR relies on just two components:

- 1) Molecular Scissors, a CRISPR-associated (Cas) nuclease.
- 2) The GPS guiding it to the appropriate site, the guide RNA (gRNA).

In natural, the gRNA consists of two distinct segments of RNA: CRISPR RNA (crRNA) and transactivating CRISPR RNA (tracrRNA). Synthetically, they can be engineered as one seamless fusion sequence known as single guide RNA (sgRNA). Why do we want it?

- Improved Editing Efficiency: The editing efficiency in cells transfected with synthetic sgRNA has been experimentally confirmed to be higher than that of non-synthetically derived sgRNA.
- Minimum Risk Delivery System: The continual expression of guide RNAs inside the cell could
 result in unwanted effects in random or unexpected places in the genome. Introducing the
 CRISPR machinery in the ribonucleoprotein format into cells alleviates these concerns as the
 RNP exists transiently inside the cell and shows reduced toxicity and off-target effects.
- Increased Experimental Convenience: Even if an experiment is complicated, its preparation does not need to be.
- Compatibility With Chemically Modified Guides

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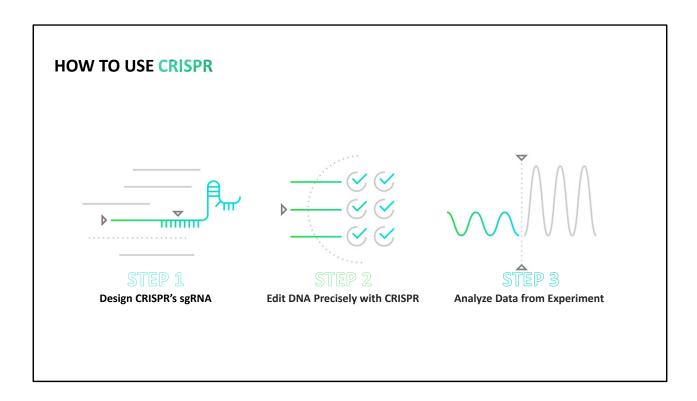
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- **Increased Experimental Convenience:** Even if an experiment is complicated, its preparation does not need to be.
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Successfully incorporating CRISPR into your research involves several steps: selecting the correct components to achieve your experimental goals, introducing these components into cells to perform the editing, and analyzing the results of the CRISPR experiments. Broadly, these steps can be categorized under design, edit, and analyze. First, let's explore how to design guide RNAs for CRISPR

STEP 1: DESIGNING gRNA

Choose the best CRISPR gRNA design tool, from the market, according to the type of research you are holding. The following are the most common types CRISPR is used in:

Designing gRNA for Gene Knockouts

Most widely adopted use of CRISPR is to knock out (KO) specific genes in the genome of an organism to test their function. A successful genetic knockout generally results in a loss of protein function and often (but not always) manifests as a change in phenotype.

Designing gRNA for Gene Knock-ins

A knock-in (KI) experiment aims to edit the target genome by inserting a new DNA fragment into the genome. In this case, we divert the cell from using its Non-homologous end joining (NHEJ) repair pathway (which repairs double-strand breaks in DNA) to another more sophisticated repair pathway, referred to as the homology-directed repair (HDR) pathway. In HDR, the cell copies the sequence of a donor DNA template to fill in the broken piece accurately. In knock-in experiments, researchers supply a surplus of template DNA bearing their desired genomic change, improving their chances that the cell will pick this DNA to use for the repair.

Designing gRNA for CRISPRA (Activator) and CRISPRI (Inhibitor)

These experiments involve gene activation or inhibition by targeting the promoter of the target gene. The location target DNA range for guide RNA is therefore quite narrow, necessitating a balance of complementary sequence and optimized location while designing.

While designing, ensure the on-target activity of the gRNA and try to minimize most of its off-target effects.

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STEP 2: GENE EDITING

Once the sgRNA is designed and synthesized, you can precisely edit the desired DNA sequence in any genome. Depending on the application and specific experimental needs, a variety of alternative nucleases exist for every job.

<u>Note</u>: you need to ensure that the sequence you want to edit contains the PAM (Protospacer Adjacent Motif) sequence that the nuclease absolutely needs to cut.

Cas9: The Workhorse CRISPR-Associated Nuclease

Cas9 is currently the most widely known nuclease in CRISPR experiments, specifically the Cas9 variant isolated from the bacterium Streptococcus pyogenes (SpCas9). Once Cas9 recognizes the PAM sequence (5'-NGG-3 in case of SpCas9), where N is any nucleotide), it creates a double-strand break (DSB) at the target locus. Cas9 activity is a collective effort of two parts of the protein: the recognition lobe and the nuclease lobe. One senses the complementary DNA sequence, and the other has nuclease domains which cleave the DNA.

Cas9 Alternatives

CasX, CasY, Cas12a (Cpf1), Cas14a, High-Fidelity Cas9, eSpCas9, SpCas9-HF1, HypaCas9, Fokl-Fused dCas9, Catalytically Dead Cas9 (dCas9), xCas9: Expanded PAM Recognition.

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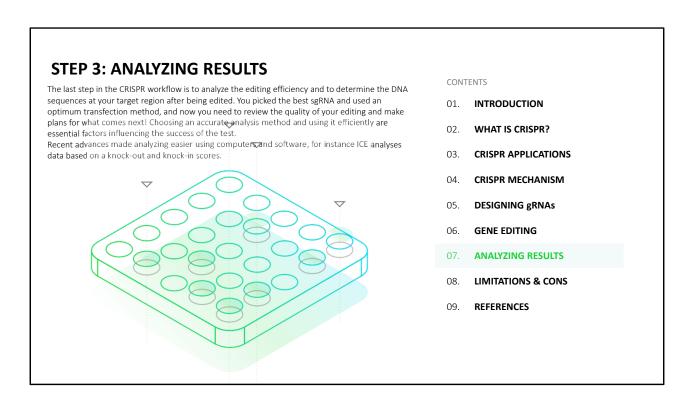
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Some test requires a nuclease other than CAS9 here are the alternatives.



The last step in the CRISPR workflow is to analyze the editing efficiency and to determine the DNA sequences at your target region after being edited. You picked the best sgRNA and used an optimum transfection method, and now you need to review the quality of your editing and make plans for what comes next! Choosing an accurate analysis method and using it efficiently are essential factors influencing the success of the test.

Recent advances made analyzing easier using computers and software, for instance ICE analyses data based on a knock-out and knock-in scores.

LIMITATIONS

One potential limitation of CRISPR technology is that the approach may create off-target effects. These off-target effects might play a role in recognizing and destroying hypervariable viral nucleic acids or plasmid DNA, which is beneficial to bacteria and archaea.

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Thank you yebnl3rs