Supplemental information #4: A new strategy to characterize the domain architecture structure of proteins of the innate inmune system in tunicate species

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Didemnum vexillum re-annotation

Annotation of coding regions with Augustus

Ernesto and Clara are working on that.

Mapping previous ncRNA annnotation on new assembly

Previous ncRNA annotation was retrieved from Velandia-Huerto, et~al~[] in fasta format. All the contigs which have been reported an ncRNA have been obtained from the first reported assembly of the D. vexillum genome¹. This multifasta file was mapped onto the new genome with lastz:

Alignent files were retrieved in maf format and were parsed with Bio::AlignIO Bioperl library. The criteria to obtain the best genome coordinates was choosen based on the relation between the length of the mapped region into the new genome (m) and the original size of the query contig in the old genome (s). The relation was defined as $R = \frac{m}{s}$, and were defined as the best mapping candidates those ones reported R = 1, but in order to retrieve the maximum number of mapping between the two genome versions, $R \geq 0.90$ was also considered.

From 247 contigs, was possible to map 212 in the raw results after the mapping stage with lastz. After considering the R relation, those results were parsed, resulting in: 64 (R=1), 35 ($0.95 \le R < 1$), 39 ($0.90 \le R < 0.95$) and 32 ($0.85 \le R < 0.90$), in total 170 contigs that reported high score mapping into the new genome.

Best candidates was choosen based on the final alignment score. For those contigs that reported 1:many relations, those set of positions in the new assembly was also considered for the following analysis.

Sequences from ncRNAs was obtained and mapped against the new *D. vexillum* assembly with blast, as follows:

```
blastall -p blastb -d <DB> -i <QUERY> -F F -e 10e-5 -m 8 -o <OUT>
```

According to the set of blast parameters the number of contigs were increasing into the new genome. At the same time, if one contig reported more than one candidates into the new genome, was choosen this/those one (s) that reported the highest bitscore. Having this previous information as an additional source of information in order to clean the true position of the annotated ncRNAs in the new genomes. After mapping all the candidates with blast, the true locations were obtained after applying those filters:

¹ http://tunicata.bioinf.uni-leipzig.de/Download.html

- Identity have to be $\geq 85\%$.
- E-value $< 10^{-10}$.
- Relation of sizes between the homology region of the query (r_h) and their calculated size (r_s) have to be $\frac{r_h}{r_s} \ge 0.9$

An additional confirmation step was performed using the Covariance Models from RFAMv.11 onto the retrieved fasta sequences, using infernal package:

$$cmsearch -g -Z < NT number (Mb) > --toponly < FASTA > < CM >$$

True candidates was obtained as reported in []. Following this methodology was possible to obtain 67 candidates that passed all the filters, and additional 15 candidates were included after a manual curation, based on their reported E-value and bitscore. Most of these, included candidates that failed to pass all the applied filters, due they have been detected as truncated sequences. So, in this group that was possible to mapped directly on the new *D. vexillum* 77 ncRNAs. Please discuss about the presence of the same candidate multiple times in the new genome.

The other 170 candidates did not reported a crossing with the old genome assembly in the pairwise-alignent. For that reason, the obtained blast mapping allowed to retrieve candidate coordinates into the new genome assembly. Fasta sequences were retrieved and next, evaluated by secondary alignments with correspondent CMs. From those candidates, final GFF3 file reported additional 86 mapped ncRNAs into the new assembly.

From the 264 reported loci of ncRNAs, 163 were retrieved in the new genome, GFF file is reported along with all the coding and non-coding regions on Supplemental File X.

After mapping the complete set of reported sequences on the new genome assembly some ncRNAs failed to be searched by blast. In this case, in order to determine if this ncRNA family is present, homology searches using Hidden Markov Models (HMMs) was applied directly with the missing 51 families.