Metabolomic Data Analysis with MetaboAnalyst 6.0

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1 Background

MSEA or Metabolite Set Enrichment Analysis is a way to identify biologically meaningful patterns that are significantly enriched in quantitative metabolomic data. In conventional approaches, metabolites are evaluated individually for their significance under conditions of study. Those compounds that have passed certain significance level are then combined to see if any meaningful patterns can be discerned. In contrast, MSEA directly investigates if a set of functionally related metabolites without the need to preselect compounds based on some arbitrary cut-off threshold. It has the potential to identify subtle but consistent changes among a group of related compounds, which may go undetected with the conventional approaches.

Essentially, MSEA is a metabolomic version of the popular GSEA (Gene Set Enrichment Analysis) software with its own collection of metabolite set libraries as well as an implementation of user-friendly web-interfaces. GSEA is widely used in genomics data analysis and has proven to be a powerful alternative to conventional approaches. For more information, please refer to the original paper by Subramanian A, and a nice review paper by Nam D, Kim SY. ¹. ²

2 MSEA Overview

Metabolite set enrichment analysis consists of four steps - data input, data processing, data analysis, and results download. Different analysis procedures are performed based on different input types. In addition, users can also browse and search the metabolite set libraries as well as upload their self-defined metabolite sets for enrichment analysis. Users can also perform metabolite name mapping between a variety of compound names, synonyms, and major database identifiers.

3 Data Input

There are three enrichment analysis algorithms offered by MSEA. Accordingly, three different types of data inputs are required by these three approaches:

- A list of important compound names entered as a one column data (Over Representation Analysis (ORA));
- A single measured biofluid (urine, blood, CSF) sample- entered as tab separated two-column data with the first column for compound name, and the second for concentration values (Single Sample Profiling (SSP));

¹Subramanian Gene set enrichment analysis: A knowledge-based approach for interpreting genome-wide expression profiles., Proc Natl Acad Sci USA. 2005 102(43): 15545-50

²Nam D, Kim SY. Gene-set approach for expression pattern analysis, Briefings in Bioinformatics. 2008 9(3): 189-197.

• A compound concentration table - entered as a comma separated (.csv) file with the each sample per row and each metabolite concentration per column. The first column is sample names and the second column for sample phenotype labels (Quantitative Enrichment Analysis (QEA))

You selected Over Representation Analysis (ORA) which requires a list of compound names as input.

4 Data Process

The first step is to standardize the compound labels. It is an essential step since the compound labels will be subsequently compared with compounds contained in the metabolite set library. MSEA has a built-in tool to convert between compound common names, synonyms, identifiers used in HMDB ID, PubChem, ChEBI, BiGG, METLIN, KEGG, or Reactome. **Table 1** shows the conversion results. Note: 1 indicates exact match, 2 indicates approximate match, and θ indicates no match. A text file contain the result can be found the downloaded file name map.csv

Table 1: Result from Compound Name M

	Query	Match	HMDB	PubChem	KEGG	SMILES
1	C07005	Flunisolide	${ m HMDB0014326}$	82153	C07005	[H][C@@]12C[C@@]3([H])[C@]4([H])C[C@H](F)C[C@H
2	C00307	Citicoline	${ m HMDB0001413}$	13805	C00307	C[N+](C)(C)CCOP(O)(=O)OP(O)(=O)OC[C@H
3	C00127	Oxidized glutathione	${ m HMDB0003337}$	65359	C00127	N[C@@H](CCC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](CSSC[C@H](NC(=O)N[C@@H](NC(=O)M[C@@H](NC(=O)M[C@@H](NC(=O)M[C@@H](NC(=O)M[C@@H](NC(=O)M[C@@H](NC(=O)M[C](ASSC[C@H](NC(=O)M[C](ASSC[C@H](NC(=O)M[C](ASSC[C@H](NC(=O)M[C](ASSC[C@H](NC(=O)M[C](ASSC[C@H](ASSC[CASC[C@H](ASSC[C@H](ASSC[C@H](ASSC[C@H](ASSC[C@H](ASSC[C@H](ASSC[C@H](A
4	C00105	Uridine 5'-monophosphate	${ m HMDB0000288}$	6030	C00105	O[C@H]1[C@@H](O)[C@@H](O[C@@H]1COP(O)
5	C05635	5-Hydroxyindoleacetic acid	${ m HMDB0000763}$	1826	C05635	OC(=O)CC1=CNC2=C1C=C(O)C=C2
6	C02301	O-Acylcarnitine		5355	C02301	
7	C00319	Sphingosine	${ m HMDB0000252}$	5280335	C00319	CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC
8	C00836	Sphinganine	${ m HMDB0000269}$	91486	C00836	CCCCCCCCCCCCC[C@@H](O)[C@@H](N)C
9	C04100	NA	NA	NA	NA	NA
10	C02990	Palmitoylcarnitine	${ m HMDB0000222}$	11953816	C02990	CCCCCCCCCCCCCCC(=O)O[C@H](CC(O)=O)
11	C03546	D-myo-Inositol 4-phosphate	HMDB0001313	440043	C03546	O[C@@H]1[C@H](O)[C@H](O)[C@@H](OP(O)(O)
12	C00052	Uridine diphosphategalactose	${ m HMDB0000302}$	18068	C00052	OC[C@H]1O[C@H](OP(O)(=O)OP(O)(=O)OC[C
13	C00158	Citric acid	${ m HMDB0000094}$	311	C00158	OC(=O)CC(O)(CC(O)=O)C(O)=O
14	C03889	NA	NA	NA	NA	NA
15	C19434	NA	NA	NA	NA	NA
16	C02862	Butyrylcarnitine	${ m HMDB0002013}$	213144	C02862	CCCC(=O)O[C@H](CC([O-])=O)C[N+](C)(C)C
17	C00029	Uridine diphosphate glucose	${ m HMDB0000286}$	8629	C00029	OC[C@H]1O[C@H](OP(O)(=O)OP(O)(=O)OC[C
18	C02305	Phosphocreatine	HMDB0001511	587	C02305	CN(CC(O)=O)C(=N)NP(O)(O)=O
19	C00570	CDP-ethanolamine	${ m HMDB0001564}$	123727	C00570	NCCOP(O)(=O)OP(O)(=O)OC[C@H]1O[C@H](
20	C00167	Uridine diphosphate glucuronic acid	${ m HMDB0000935}$	17473	C00167	O[C@@H]1[C@@H](COP(O)(=O)OP(O)(=O)O[CO]
21	C05526	S-Glutathionyl-L-cysteine	METPA0607		C05526	
_22	C00103	Glucose 1-phosphate	HMDB0001586	65533	C00103	OC[C@H]1O[C@H](OP(O)(O)=O)[C@H](O)[C@G

The second step is to check concentration values. For SSP analysis, the concentration must be measured in umol for blood and CSF samples. The urinary concentrations must be first converted to $umol/mmol_creatinine$ in order to compare with reported concentrations in literature. No missing or negative values are allowed in SSP analysis. The concentration data for QEA analysis is more flexible. Users can upload either the original concentration data or normalized data. Missing or negative values are allowed (coded as NA) for QEA.

5 Selection of Metabolite Set Library

Before proceeding to enrichment analysis, a metabolite set library has to be chosen. There are seven built-in libraries offered by MSEA:

- Metabolic pathway associated metabolite sets (currently contains 99 entries);
- Disease associated metabolite sets (reported in blood) (currently contains 344 entries);
- Disease associated metabolite sets (reported in urine) (currently contains 384 entries)
- Disease associated metabolite sets (reported in CSF) (currently contains 166 entries)
- Metabolite sets associated with SNPs (currently contains 4598 entries)
- Predicted metabolite sets based on computational enzyme knockout model (currently contains 912 entries)
- Metabolite sets based on locations (currently contains 73 entries)
- Drug pathway associated metabolite sets (currently contains 461 entries)

In addition, MSEA also allows user-defined metabolite sets to be uploaded to perform enrichment analysis on arbitrary groups of compounds which researchers want to test. The metabolite set library is simply a two-column comma separated text file with the first column for metabolite set names and the second column for its compound names (must use HMDB compound name) separated by "; ". Please note, the built-in libraries are mainly from human studies. The functional grouping of metabolites may not be valid. Therefore, for data from subjects other than human being, users are suggested to upload their self-defined metabolite set libraries for enrichment analysis.

6 Enrichment Analysis

Over Representation Analysis (ORA) is performed when a list of compound names is provided. The list of compound list can be obtained through conventional feature selection methods, or from a clustering algorithm, or from the compounds with abnormal concentrations detected in SSP, to investigate if some biologically meaningful patterns can be identified.

ORA was implemented using the *hypergeometric test* to evaluate whether a particular metabolite set is represented more than expected by chance within the given compound list. One-tailed p values are provided after adjusting for multiple testing. **Figure 2** below summarizes the result.

Metabolite Sets Enrichment Overview

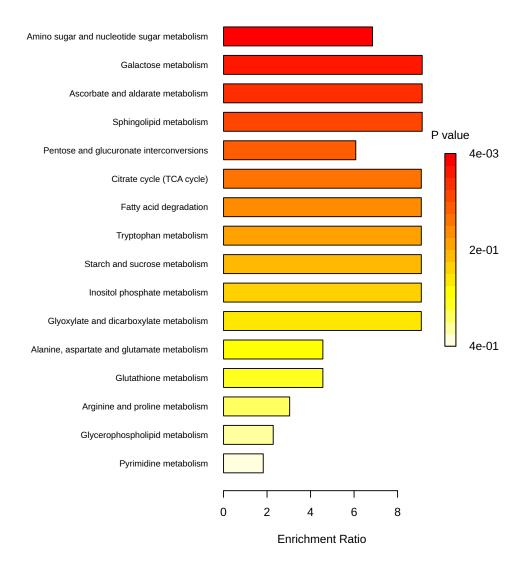


Figure 1: Summary Plot for Over Representation Analysis (ORA)

Table 2: Result from Over Representation Analysis

	total	expected	hits	Raw p	Holm p	FDR
Amino sugar and nucleotide sugar	4	0.44	3	4.11E-03	3.29E-01	2.27E-01
$\operatorname{metabolism}$						
Galactose metabolism	2	0.22	2	1.13E-02	8.96E-01	2.27E-01
Ascorbate and aldarate metabolism	2	0.22	2	1.13E-02	8.96E-01	2.27E-01
Sphingolipid metabolism	2	0.22	2	1.13E-02	8.96E-01	2.27E-01
Pentose and glucuronate interconversions	3	0.33	2	3.18E-02	1.00E+00	5.09E-01
Citrate cycle (TCA cycle)	1	0.11	1	1.10E-01	1.00E+00	7.97E-01
Fatty acid degradation	1	0.11	1	1.10E-01	1.00E+00	7.97E-01
Tryptophan metabolism	1	0.11	1	1.10E-01	1.00E+00	7.97E-01
Starch and sucrose metabolism	1	0.11	1	1.10E-01	1.00E+00	7.97E-01
Inositol phosphate metabolism	1	0.11	1	1.10E-01	1.00E+00	7.97E-01
Glyoxylate and dicarboxylate	1	0.11	1	1.10E-01	1.00E+00	7.97E-01
$\operatorname{metabolism}$						
Alanine, aspartate and glutamate	2	0.22	1	2.08E-01	1.00E+00	1.00E + 00
$\operatorname{metabolism}$						
Glutathione metabolism	2	0.22	1	2.08E-01	1.00E+00	1.00E + 00
Arginine and proline metabolism	3	0.33	1	2.96E-01	1.00E+00	1.00E + 00
Glycerophospholipid metabolism	4	0.44	1	3.75E-01	1.00E+00	1.00E + 00
Pyrimidine metabolism	5	0.55	1	4.45E-01	1.00E+00	1.00E+00

7 Appendix: R Command History

```
[1] "mSet<-InitDataObjects(\"conc\", \"msetora\", FALSE)"
[2] "cmpd.vec<-c(\"C07005\",\"C00307\",\"C00127\",\"C00105\",\"C05635\",\"C02301\",\"C00319\",\"C00
[3] "mSet<-Setup.MapData(mSet, cmpd.vec);"
[4] "mSet<-CrossReferencing(mSet, \"kegg\");"
[5] "mSet<-CreateMappingResultTable(mSet)"
[6] "mSet<-Setup.HMDBReferenceMetabolome(mSet, \"FCCP.txt\");"
[7] "mSet<-SetMetabolomeFilter(mSet, T);"
[8] "mSet<-SetCurrentMsetLib(mSet, \"kegg_pathway\", 2);"
[9] "mSet<-CalculateHyperScore(mSet)"
[10] "mSet<-PlotORA(mSet, \"ora_0_\", \"net\", \"png\", 72, width=NA)"
[11] "mSet<-PlotEnrichDotPlot(mSet, \"ora\", \"ora_dot_0_\", \"png\", 72, width=NA)"
[12] "mSet<-CalculateHyperScore(mSet)"
[13] "mSet<-PlotORA(mSet, \"ora_1_\", \"net\", \"png\", 72, width=NA)"
[14] "mSet<-PlotEnrichDotPlot(mSet, \"ora\", \"png\", 72, width=NA)"
[15] "mSet<-SaveTransformedData(mSet)"
[16] "mSet<-PreparePDFReport(mSet, \"guest7826037707225041567\")\n"</pre>
```

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