

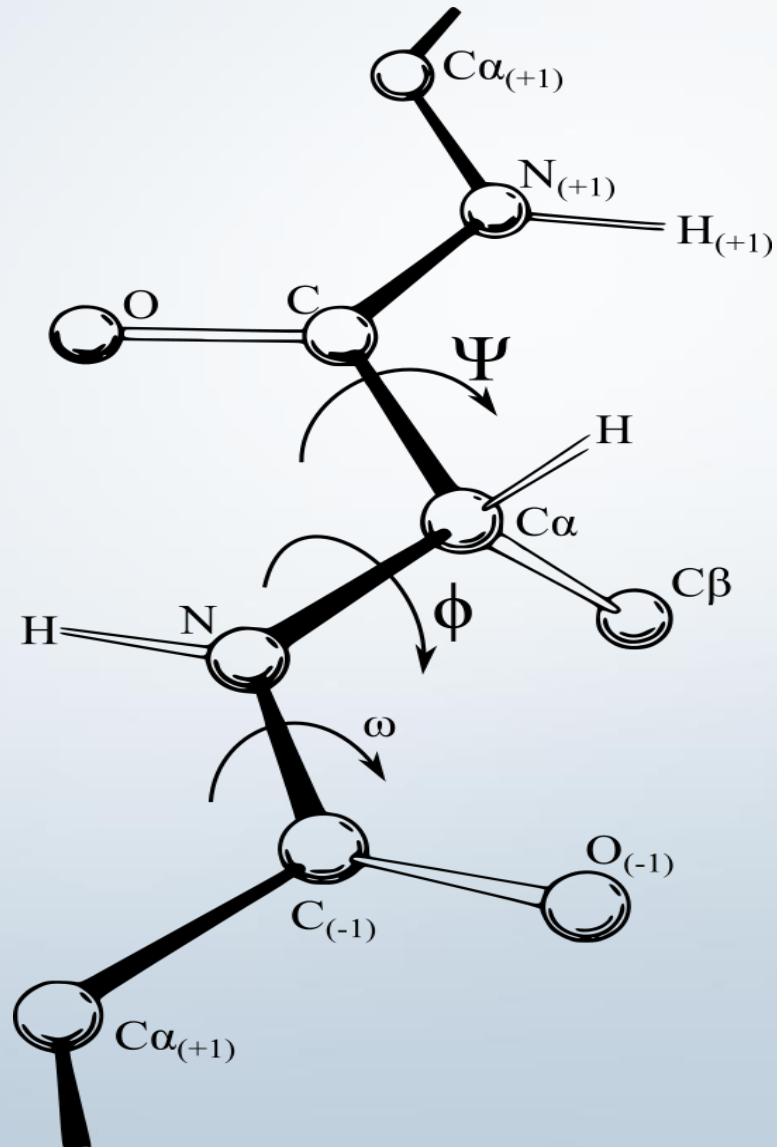


# Working with Proteins

Yazdan Asgari

2020

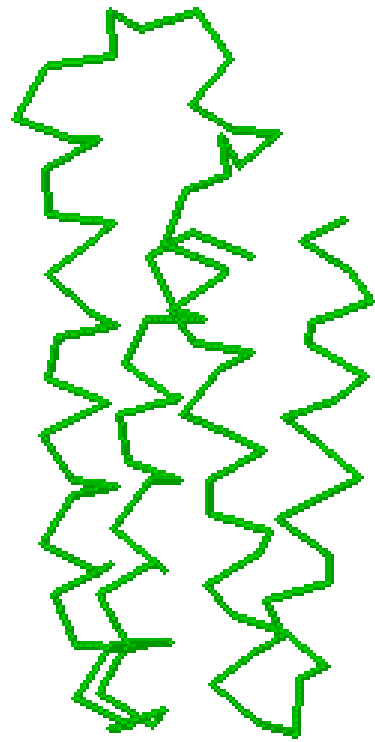
# Protein Backbone



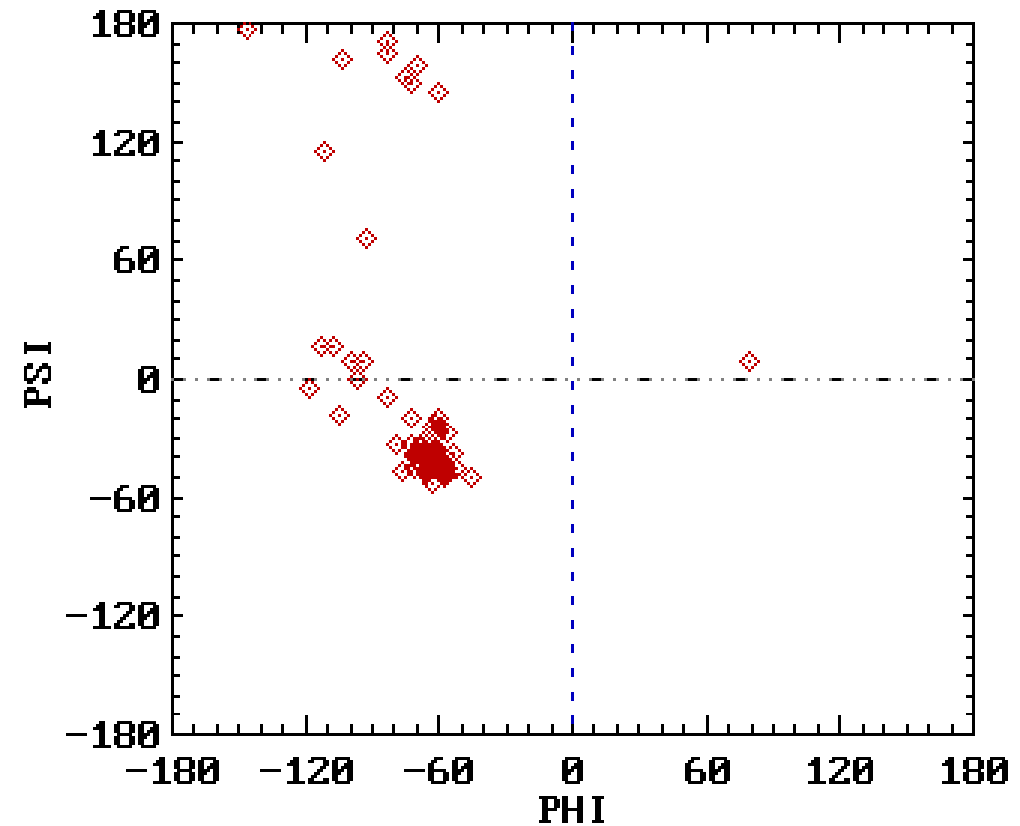
# Ramachandran Plot

- Plot of  $\phi$  vs.  $\psi$
- Repeating values of  $\phi$  and  $\psi$  along the chain result in regular structure
- For example, repeating values of  $\phi \sim -57^\circ$  and  $\psi \sim -47^\circ$  give a right-handed helical fold (the alpha-helix)
- The structure of cytochrome C-256 shows many segments of helix and the Ramachandran plot shows a tight grouping of  $\phi$ ,  $\psi$  angles near  $-50, -50$

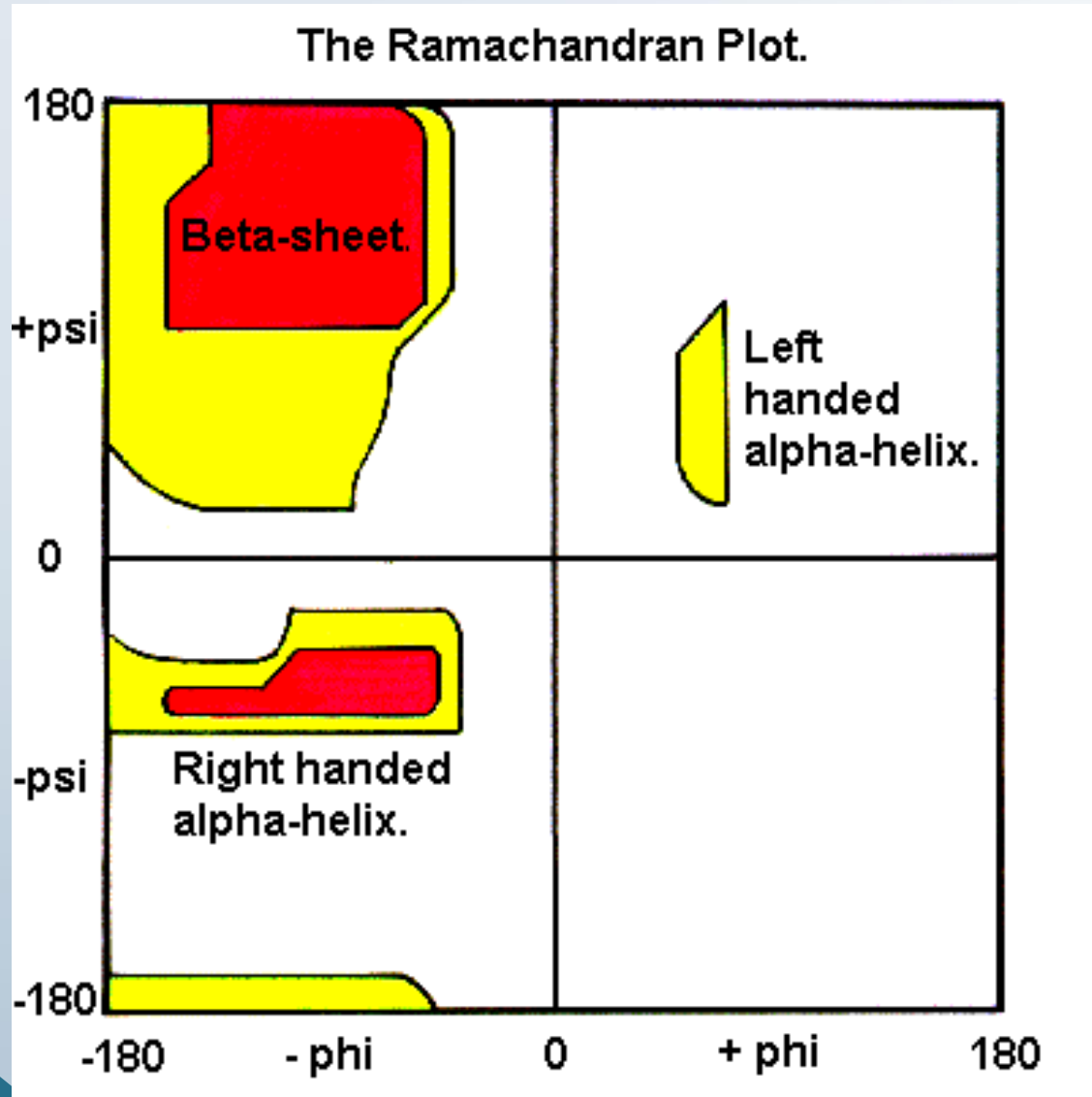
# The structure of cytochrome C-256



alpha-helix

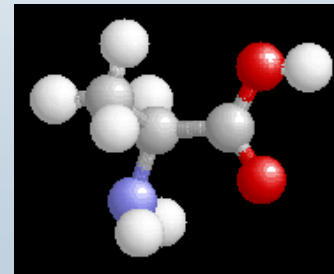
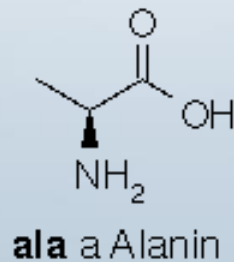
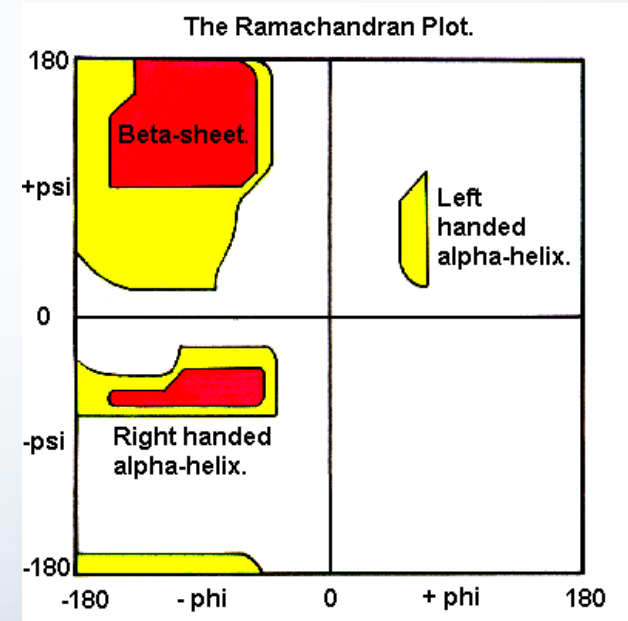
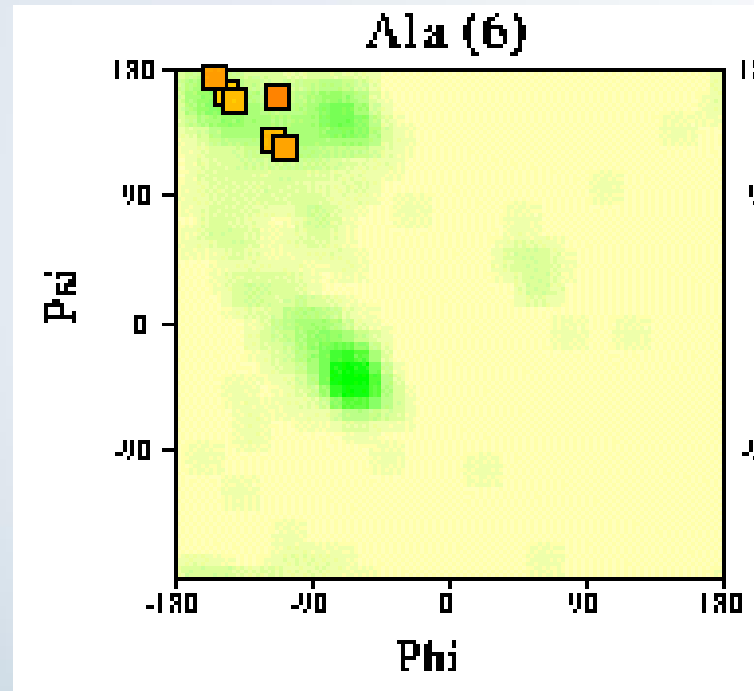


cytochrome C-256 Ramachandran plot

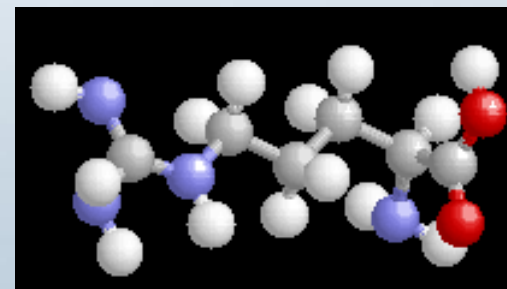
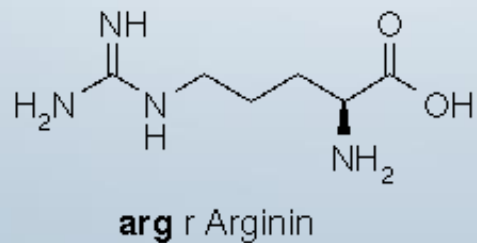
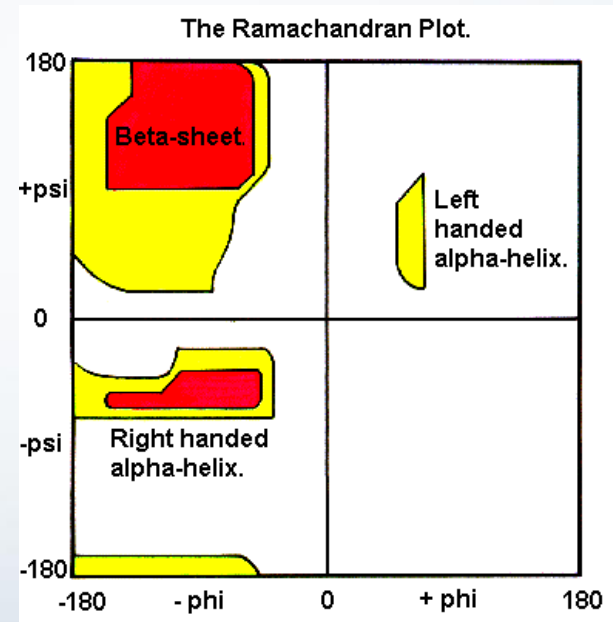
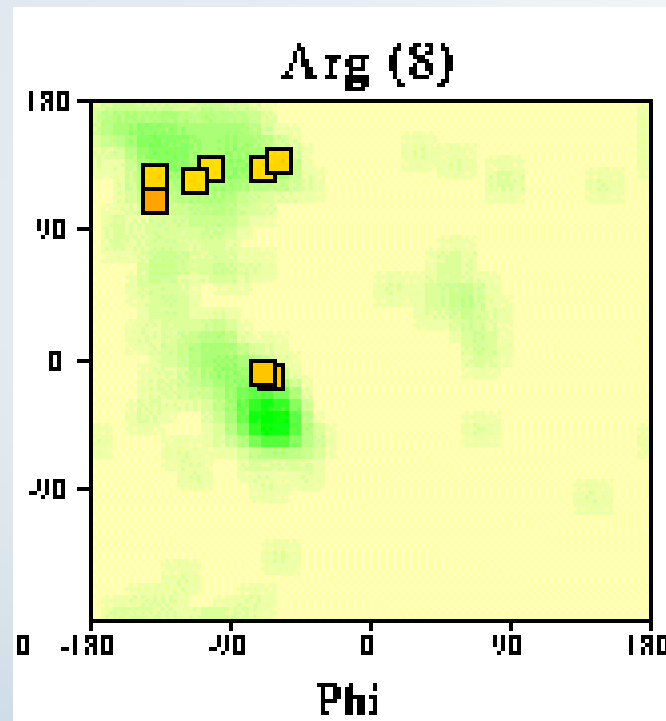


- White = Sterically disallowed conformations (atoms in the polypeptide come closer than the sum of their van der Waals radii)
- Red = Sterically allowed regions (namely right-handed alpha helix and beta sheet)
- Yellow = Sterically allowed if shorter radii are used (i.e. atoms allowed closer together; brings out left-handed helix)

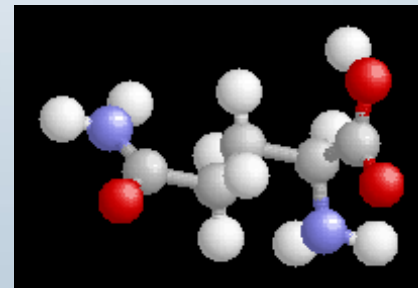
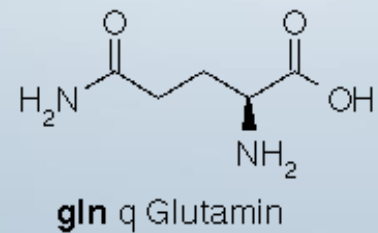
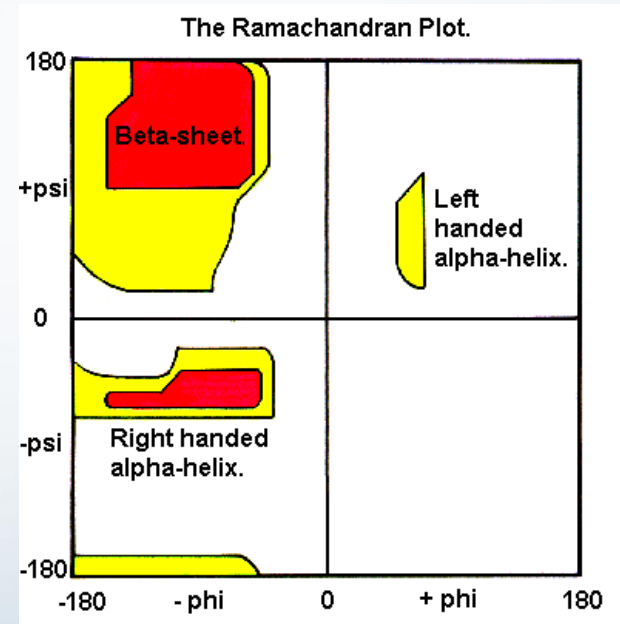
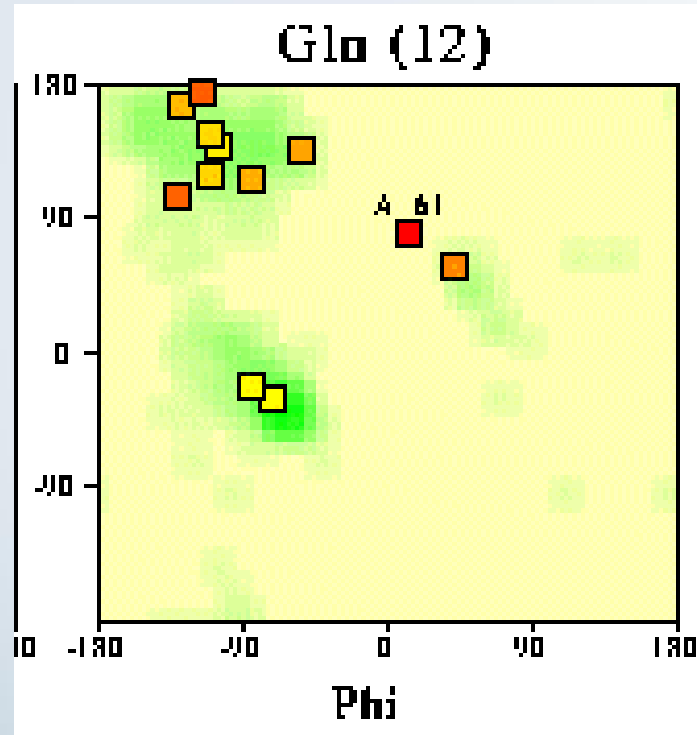
# Alanine Ramachandran Plot



# Arginine Ramachandran Plot

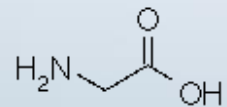
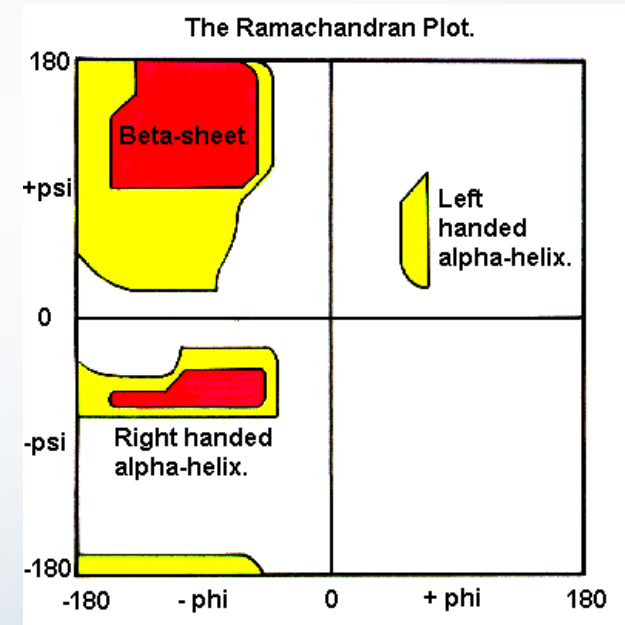
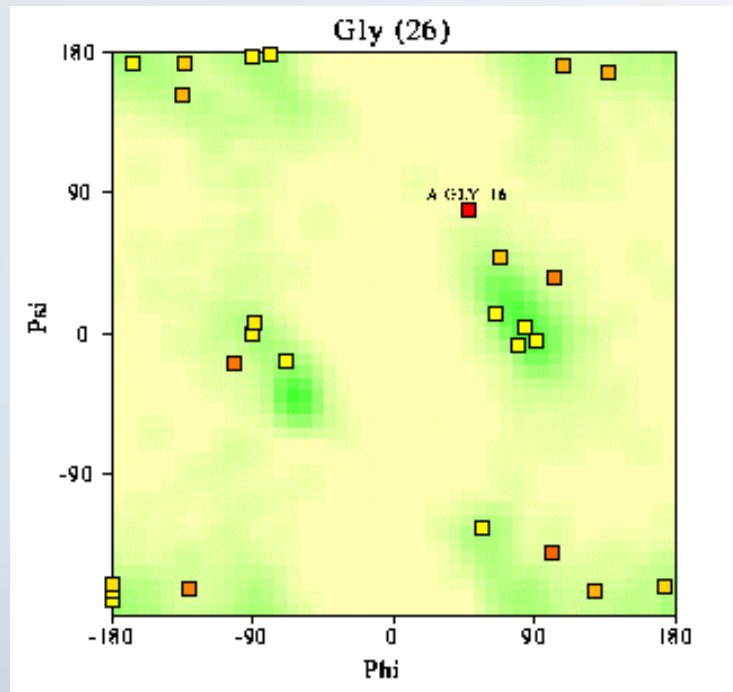


# Glutamine Ramachandran Plot

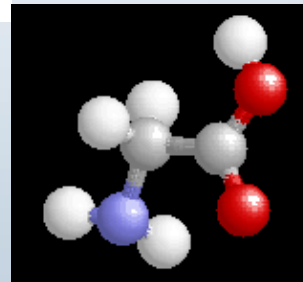




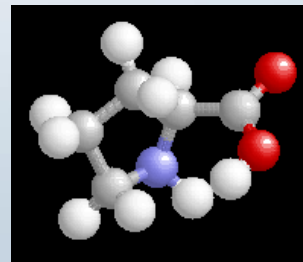
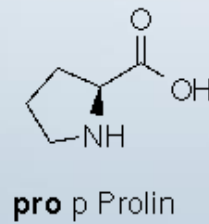
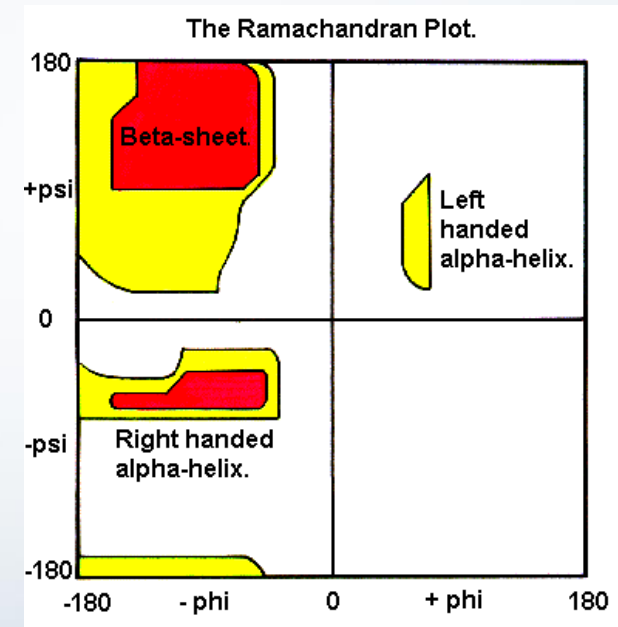
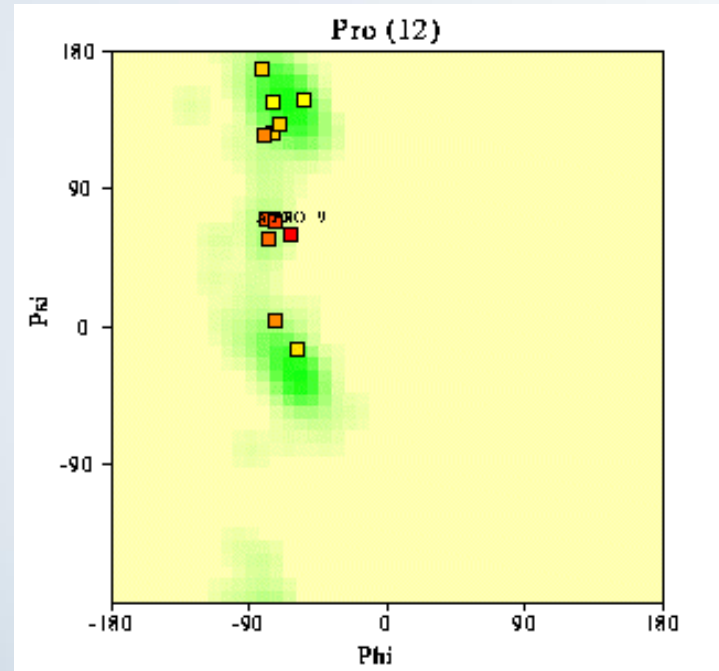
# Glycine Ramachandran Plot



gly g Glycin



# Proline Ramachandran Plot



# Regular Secondary Structure Conformations

Secondary Structure Element	Torsional Angle (°)		Residue/turn	Translational distance per residue (Å)
	$\phi$	$\psi$		
$\alpha$ helix	-57	-47	3.6	1.50
$3_{10}$ helix	-49	-26	3.0	2.00
$\pi$ helix	-57	-70	4.4	1.15
Parallel $\beta$ strand	-139	+135	2.0	3.20
Antiparallel $\beta$ strand	-119	+113	2.0	3.40
Poly(Pro) I	-83	+158	3.3	1.90
Poly(Pro) II	-78	+149	3.0	3.12

# Physicochemical Properties

- Online servers exist to determine many properties of your protein sequences
  - Molecular weight
  - Extinction coefficients
  - Half-life
  - Isoelectric point
- It is also possible to simulate protease digestion

# Molecular Weight

- Measured in daltons (Da)
  - Same as unified atomic mass unit
  - Equivalent to grams/mol
    - But it is mass of one molecule, not a mole
- Technically measure of mass, not weight
  - Like grams, kilograms (mass units)
  - Not like ounces, pounds (weight units)
- Still common to call it molecular weight

# Isoelectric Point (pI)

- Defined as pH at which entire protein has no net charge
  - If  $pI < 7$ , protein is acidic
  - If  $pI > 7$ , protein is basic
- Acidic proteins are negatively charged at neutral pH
- Basic proteins are positively charged at neutral pH



# Protoparam

## Protparam tool

**Protparam** ([References](#) / [Documentation](#)) is a tool which allows the computation of various physical and chemical parameters for a given protein stored in [Swiss-Prot](#) or [TrEMBL](#) or for a user entered protein sequence. The computed parameters include the molecular weight, theoretical pI, amino acid composition, atomic composition, extinction coefficient, estimated half-life, instability index, aliphatic index and grand average of hydropathicity (GRAVY) ([Disclaimer](#)).

Please note that you may only fill out **one** of the following fields at a time.

Enter a Swiss-Prot/TrEMBL accession number (AC) (for example **P05130**) or a sequence identifier (ID) (for example **KPC1\_DROME**):

Or you can paste your own amino acid sequence (in one-letter code) in the box below:

# Example (P32851 STX1A\_RAT)

## ProtParam

### Selection of endpoints on the sequence

#### STX1A\_RAT (P32851)

Syntaxin-1A (Neuron-specific antigen HPC-1) (Synaptotagmin-associated 35 kDa protein) (P35A) *Rattus norvegicus* (Rat).

Please select one of the following features by clicking on a pair of endpoints, and the computation complete sequence is used.

**Note:** Only the features corresponding to subsequences of at least 5 residues are highlighted.

FT	CHAIN	1-288	Syntaxin-1A.
FT	TOPO_DOM	1-265	Cytoplasmic. {ECO:0000255}.
FT	TRANSMEM	266-288	Helical; Anchor for type IV membrane
FT	DOMAIN	192-254	t-SNARE coiled-coil homology.
FT	COILED	68-109	{ECO:0000255}.
FT	COMPBias	13-19	Asp-rich (acidic).
FT	TURN	6-8	{ECO:0000244 PDB:3C98}.
FT	HELIX	28-63	{ECO:0000244 PDB:1EZ3}.
FT	STRAND	64-66	{ECO:0000244 PDB:4JEU}.
FT	HELIX	69-104	{ECO:0000244 PDB:1EZ3}.
FT	TURN	105-107	{ECO:0000244 PDB:3C98}.
FT	HELIX	111-146	{ECO:0000244 PDB:1EZ3}.
FT	HELIX	162-170	{ECO:0000244 PDB:4JEH}.
FT	HELIX	176-180	{ECO:0000244 PDB:4JEH}.
FT	STRAND	183-185	{ECO:0000244 PDB:4JEH}.
FT	HELIX	192-254	{ECO:0000244 PDB:1N7S}.
FT	HELIX	261-284	{ECO:0000244 PDB:2M8R}.



# Physicochemical Properties - Example

Number of amino acids: 288

Molecular weight: 33067.48

Theoretical pI: 5.14

Amino acid composition: [CSV format](#)

Ala (A)	16	5.6%
Arg (R)	22	7.6%
Asn (N)	8	2.8%
Asp (D)	22	7.6%
Cys (C)	3	1.0%
Gln (Q)	11	3.8%
Glu (E)	35	12.2%
Gly (G)	11	3.8%
His (H)	5	1.7%
Ile (I)	30	10.4%
Leu (L)	16	5.6%
Lys (K)	23	8.0%
Met (M)	12	4.2%
Phe (F)	8	2.8%
Pro (P)	3	1.0%
Ser (S)	26	9.0%
Thr (T)	16	5.6%
Trp (W)	0	0.0%
Tyr (Y)	5	1.7%
Val (V)	16	5.6%
Pyl (O)	0	0.0%
Sec (U)	0	0.0%

(B)	0	0.0%
(Z)	0	0.0%
(X)	0	0.0%

Total number of negatively charged residues (Asp + Glu): 57

Total number of positively charged residues (Arg + Lys): 45

Atomic composition:

Carbon	C	1419
Hydrogen	H	2334
Nitrogen	N	406
Oxygen	O	469
Sulfur	S	15

Formula:  $C_{1419}H_{2334}N_{406}O_{469}S_{15}$

Total number of atoms: 4643

Extinction coefficients:

This protein does not contain any Trp residues. Experience shows that this could result in more than 10% error in the computed extinction coefficient.

Extinction coefficients are in units of  $M^{-1} cm^{-1}$ , at 280 nm measured in water.

Ext. coefficient 7575

Abs 0.1% (=1 g/l) 0.229, assuming all pairs of Cys residues form cystines

Ext. coefficient 7450

Abs 0.1% (=1 g/l) 0.225, assuming all Cys residues are reduced

Estimated half-life:

The N-terminal of the sequence considered is M (Met).

The estimated half-life is: 30 hours (mammalian reticulocytes, in vitro).

>20 hours (yeast, in vivo).

>10 hours (Escherichia coli, in vivo).

Instability index:

The instability index (II) is computed to be 48.79

This classifies the protein as unstable.

Aliphatic index: 83.96

Grand average of hydropathicity (GRAVY): -0.604

# Compute\_pi

## Compute pI/Mw tool

**Compute pI/Mw** is a tool which allows the computation of the theoretical pI (isoelectric point) and Mw (molecular weight) for a list of [UniProt Knowledgebase \(Swiss-Prot or TrEMBL\)](#) entries or for user entered sequences [\[reference\]](#).

[Documentation](#) is available.

### Compute pI/Mw for Swiss-Prot/TrEMBL entries or a user-entered sequence

Please enter one or more UniProtKB/Swiss-Prot protein identifiers (ID) (e.g. *ALBU\_HUMAN*) or UniProt Knowledgebase accession numbers (AC) (e.g. *P04406*), separated by spaces, tabs or newlines. Alternatively, enter a protein sequence in single letter code. The theoretical *pI* and *Mw* (molecular weight) will then be computed.

Or upload a file from your computer, containing one Swiss-Prot/TrEMBL ID/AC or one sequence per line:  No file chosen

Resolution: ☒ Average or ☐ Monoisotopic

# Example – (P68871 HBB\_HUMAN)

## Compute pI/Mw

### HBB\_HUMAN (P68871)

Hemoglobin subunit beta (Beta-globin) (Hemoglobin beta chain) [Contains: LVV-hemorphin-7; Spinorphin]  
Homo sapiens (Human).

**The parameters have been computed for the following feature:**

FT CHAIN 2 147 Hemoglobin subunit beta.

Considered sequence fragment:

1	11	21	31	41	51	
1	VHLTPEEK	AVTALWGKVN	VDEVGGEALG	RLLVVYPWTQ	RFFESFGDLS	TPDAVMGNPK
61	VKAHGKKVLG	AFSDGLAHL	NLKGTFATLS	ELHCDKLHVD	PENFRLLGNV	LVCVLAHHFG
121	KEFTPPVQAA	YQKVVAGVAN	ALAHKYH			

» [Fasta](#)

**Molecular weight (Da):** 15867.22 (average mass), 15857.25 (monoisotopic mass)

**Theoretical pI:** 6.81

# Protein Localization

- Where does a protein go after translation?
- Secretory pathway
  - Includes ER, Golgi, plasma membrane
- Cytoplasm
- Other organelles
  - Nucleus, mitochondrion, peroxisome, lysosome
  - Chloroplast (plants only)

# Localization Prediction

- General predictors
  - Input protein sequence, output location
  - Some list several possibilities
- Specialized predictors
  - MITOPRED predicts mitochondrial proteins
  - Output is likelihood protein localizes to the mitochondrion

## Some localization servers

- Proloc-GO
- SignalP
- TargetP
- Predotar
- PSORT
- CELLO
- MultiLoc2
- Euk-mPLoc
- LocTree

# Example — (TPA: Pex8p [Saccharomyces cerevisiae S288c])

NCBI Resources How To

Protein Protein Advanced

NCBI is phasing out sequence GI numbers in September 2016. Please use accession.version! [Read more...](#)

GenPept

## Pex8p [Saccharomyces cerevisiae S288c]

NCBI Reference Sequence: NP\_011591.1

[Identical Proteins](#) [FASTA](#) [Graphics](#)

Go to:

LOCUS	NP_011591	589 aa	linear	PLN 11-MAR-2016
DEFINITION	Pex8p [Saccharomyces cerevisiae S288c].			
ACCESSION	NP_011591			
VERSION	NP_011591.1 GI:6321514			
DBLINK	BioProject: <a href="#">PRJNA128</a>			
DBSOURCE	REFSEQ: accession <a href="#">NM_001181206.1</a>			
KEYWORDS	RefSeq.			
SOURCE	Saccharomyces cerevisiae S288c			
ORGANISM	<a href="#">Saccharomyces cerevisiae S288c</a> Eukaryota; Fungi; Dikarya; Ascomycota; Saccharomycotina; Saccharomycetes; Saccharomycetales; Saccharomycetaceae; Saccharomyces.			
REFERENCE	1 (residues 1 to 589)			
AUTHORS	Tettelin,H., Agostoni Carbone,M.L., Albermann,K., Albers,M., Arroyo,J., Backes,U., Barreiros,T., Bertani,I., Bjourson,A.J., Bruckner,M., Bruschi,C.V., Carignani,G., Castagnoli,L., Cerdan,E., Clemente,M.L., Coblentz,A., Coglievina,M., Coissac,E., Defoor,E.,			



# CELLO

← → ↻ cello.life.nctu.edu.tw

**MBC** Molecular Bioinformatics Center  
National Chiao Tung University

About CELLO

**CELLO v.2.5: subCELLular Localization predictor**

1. Choose Eukaryotes

ORGANISMS	SEQUENCES
<input type="radio"/> Gram negative	<input type="radio"/> DNA
<input type="radio"/> Gram positive	<input checked="" type="radio"/> Protein
<input checked="" type="radio"/> Eukaryotes	

2. Delete Example Protein!

Paste the query sequences in FASTA format below

```
>1084005(Genbank)Outer membrane/Extracellular (Autotransporter) (major ring-  
forming surface protein precursor  
MTWISDVQENSTLGRKXSSILRRTFFQFLIATTLAFSLASSFVAACAGHAGAPVMAESITVTYQANTATVSGW  
GMAETTFINSANTTVGFIADAVTAPSTVMIANTVGHFTVGGFAGACAGCAEAKPVLHFTFGGSIASSTAKITFL  
SLGGAGMANALTSGLSLGAQATLHTHTGSIASGGFVINWVDAFFPEFTSGGATHTGNTVGTHTETSOTGTITTF  
GGWQIPNHSGLNGGTAVTGGACATVLTGWTSTYGGTHNVTFEKLGGGILAGHATGQSLGGFVTFEQQVHTTG  
VLASITGGVNTLHFGATVATGGHTLIHQGGTTFHTHTGVTGFTLTHATTFHAAAGGFAATVFGHINSAVQ  
GHTLHFTFAKLESTRAGWVFAAGATITATGGAHIVTTGGLVGGTLSTLQGSINTLVGTHNTVTHVILLTGW  
VHTFGWASHTLLFGHSTESTGGHMQTLNVAIVPFAAGGFFCAIFSTYGAFTNLAQWQAGGLHAAGAGA  
HAFAGCAVQGGQGLNFTGHSAGATNLATGGGTATVLAHTTILANLFGAAQWTVVGGHSSASIVLEAFNA  
SATITGGVYLGQGTSTVFGGSGHTSPVLIFAAGHGTPTLHATGSSITVSCAFGGFHCAGAGWLOVTVQHG  
IQHLSGQNTLQGGHLYSGGFGAFFGAILANAVDSNAKFAFGGIFLNVSLVSGHSTSPGGGHSFVHITLES  
VAVGSITALHQAFTSHHGTSTGVLVLAGDGVLLSTIAGHNGHLSHGLNQGAKLILQHSAGTGGVALHNT  
IASGHHHGGHAAITFGGFFVFGHQAHTALQHTVTLATGGGHHVSRTHFLLTVGAGHSTTTASGGQA  
SGGHHNALFVYVAGAGQGGHAGGGHATLNGHSTHSGGLYGHSTEDVIVTGGTGHFCGRISPHHGHKSYVVR  
YGGGTERAGVAVATVGGGGCAFTHTTIVGGVIGFTVFAKLTATWNAVGTETHHAGHAGHSTRAGLOSIPLOG  
TGGTGGGHTGGGQAGAQVITTFIFGAVANTHAGQLATALASHTVLYAHIDSLHGGHGLASHPHSGPHGGK
```

Or upload from file:  No file chosen



## CELLO RESULTS

SeqID: gi|6321514|ref|NP\_011591.1| Pex8p [Saccharomyces cerevisiae S288c]

### Analysis Report:

SVM	LOCALIZATION	RELIABILITY
Amino Acid Comp.	Peroxisomal	0.520
N-peptide Comp.	Peroxisomal	0.922
Partitioned seq. Comp.	PlasmaMembrane	0.511
Physico-chemical Comp.	Peroxisomal	0.810
Neighboring seq. Comp.	PlasmaMembrane	0.664

### CELLO Prediction:

Peroxisomal	2.934 *
PlasmaMembrane	1.590
Nuclear	0.098
Mitochondrial	0.093
Extracellular	0.076
Cytoplasmic	0.075
Golgi	0.038
ER	0.033
Lysosomal	0.026
Chloroplast	0.021
Cytoskeletal	0.009
Vacuole	0.008

# LOCTREE 3

## Protein Subcellular Localization Prediction System

Domain: Eukaryota

Details	Protein ID	Score	Expected Accuracy	Localization Class	Gene Ontology Terms	Annotation Type
<a href="#">Details</a>	<a href="#">gi 6321514 ref NP_011591.1 </a>	13	81%	peroxisome	<a href="#">peroxisomal matrix GO:0005782(IEA);</a>	PSI-BLAST



Mouse click on Details/Protein ID leads to the **detailed** description of a prediction.

Please cite:

Goldberg T, Hecht M, Hamp T, Karl T, Yachdav G, Nielsen H, Rost B et al. LocTree3 prediction of localization. *Nucleic Acids Research* 2014. PMID: [24848019](#)  
Goldberg T, Hamp T and Rost B: LocTree2 predicts localization for all domains of life. *Bioinformatics* 2012, 28:i458-i465. PMID: [22962467](#)

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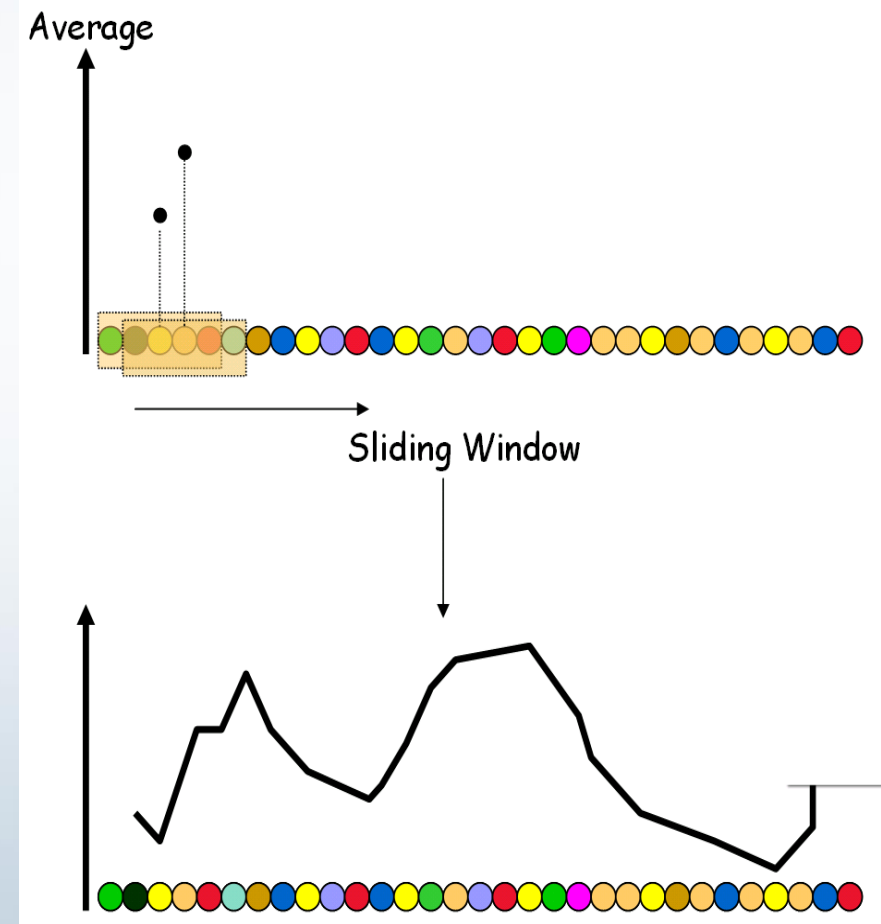
Copyright © 2016 ROSTLAB all rights reserved.

# Analyzing Local Properties

- Many local properties are important for the function of your protein
  - Hydrophobic regions are potential transmembrane domains
  - Coiled-coiled regions are potential protein-interaction domains
  - Hydrophilic stretches are potential loops
- You can discover these regions
  - Using sliding-window techniques (easy)
  - Using prediction methods such as hidden Markov Models (more sophisticated)

# Sliding-window Techniques

- Ideal for identifying strong signals
- Very simple methods
  - Few artifacts
  - Not very sensitive
- Make the window the same size as the feature you're looking for



# ProtScale

## ProtScale

**ProtScale** [[Reference](#) / [Documentation](#)] allows you to compute and represent the profile produced by any amino acid scale on a selected protein.

An **amino acid scale** is defined by a numerical value assigned to each type of amino acid. The most frequently used scales are the hydrophobicity or hydrophilicity scales and the secondary structure conformational parameters scales, but many other scales exist which are based on different chemical and physical properties of the amino acids. This program provides 57 predefined scales entered from the literature.

Enter a [UniProtKB/Swiss-Prot](#) or [UniProtKB/TrEMBL](#) accession number (AC) (e.g. **P05130**) or a sequence identifier (ID) (e.g. **KPC1\_DROME**):

Or you can paste your own sequence in the box below:

Please choose an amino acid scale from the following list. To display information about a scale (author, reference, amino acid scale values) you can click on its name.

- |  |   |
|--|---|
| <input type="radio"/> Molecular weight                     | <input type="radio"/> Number of codon(s)        |
| <input type="radio"/> Bulkiness                            | <input type="radio"/> Polarity / Zimmerman      |
| <input type="radio"/> Polarity / Grantham                  | <input type="radio"/> Refractivity              |
| <input type="radio"/> Recognition factors                  | <input type="radio"/> Hphob. / Eisenberg et al. |
| <input type="radio"/> Hphob. OMH / Sweet et al.            | <input type="radio"/> Hphob. / Hopp & Woods     |
| <input checked="" type="radio"/> Hphob. / Kyte & Doolittle | <input type="radio"/> Hphob. / Manavalan et al. |
| <input type="radio"/> Hphob. / Abraham & Leo               | <input type="radio"/> Hphob. / Black            |
| <input type="radio"/> Hphob. / Bull & Breese               | <input type="radio"/> Hphob. / Fauchere et al.  |

# ProtScale – Example (P78588)

FT CHAIN 1 669 Probable ferric reductase transmembrane

The computation has been carried out on the complete sequence (669 amino acids).

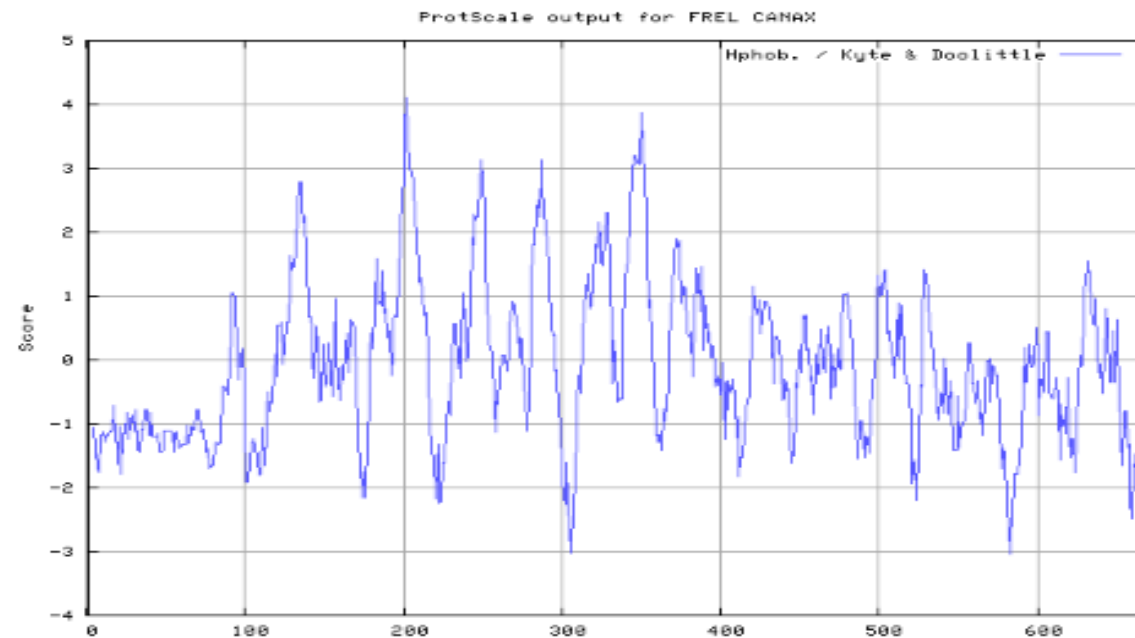
SEQUENCE LENGTH: 669

Using the scale **Hphob. / Kyte & Doolittle**, the individual values for the 20 amino acids are:

Ala: 1.800	Arg: -4.500	Asn: -3.500	Asp: -3.500	Cys: 2.500	Gln: -3.500
Glu: -3.500	Gly: -0.400	His: -3.200	Ile: 4.500	Leu: 3.800	Lys: -3.900
Met: 1.900	Phe: 2.800	Pro: -1.600	Ser: -0.800	Thr: -0.700	Trp: -0.900
Tyr: -1.300	Val: 4.200	: -3.500	: -3.500	: -0.400	

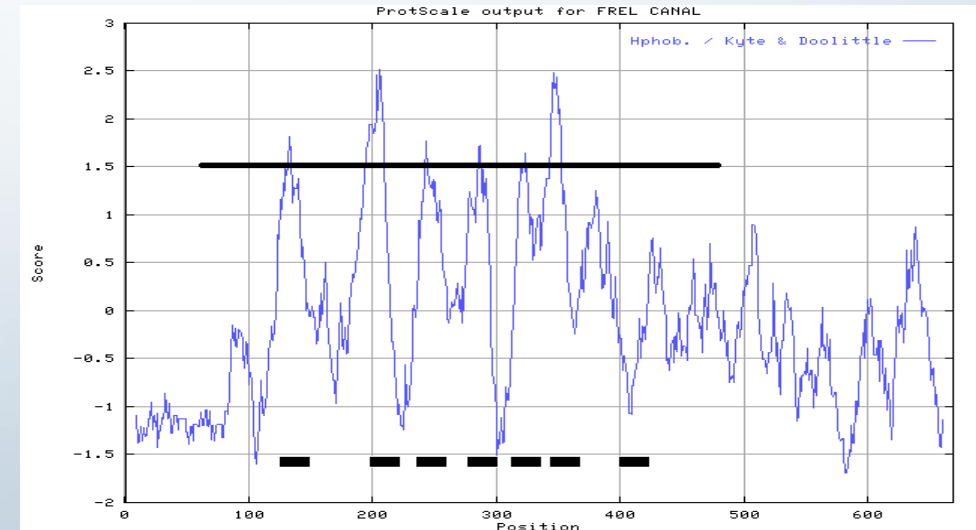
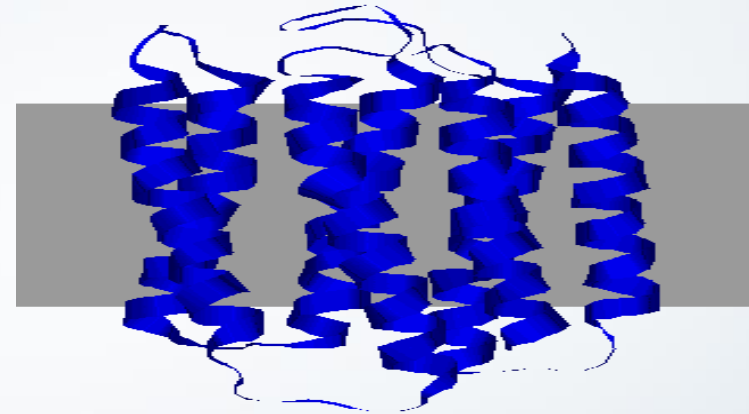
Weights for window positions 1,...,9, using **linear weight variation model**:

1	2	3	4	5	6	7	8	9
1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
edge				center				edge



# Transmembrane Domains

- Discovering a transmembrane domain tells you a lot about your protein
- Many important receptors have 7 transmembrane domains
- Transmembrane segments can be found using ProtScale
- The most accurate predictions come from using TMHMM



# Transmembrane Domains – TMHMM Method

- TMHMM is the best method for predicting transmembrane domains
- TMHMM uses an HMM
- Its principle is very different from that of ProtScale
- TMHMM output is a prediction

**TMHMM Server v. 2.0**  
Prediction of transmembrane helices in proteins

[Instructions](#)

**SUBMISSION**

Submission of a local file in **FASTA** format (HTML 3.0 or higher)  
 No file chosen

OR by pasting sequence(s) in **FASTA** format:

```
>sp|P78588|FREL_CANAX Probable ferric reductase transmembrane  
component OS=Candida albicans GN=CFL1 PE=3 SV=1  
MTESKFHAKYDKIQAEFKTNGTEYAKMTTKSSSGSKTSTSASKSSKSTGSSNASKSSTNA  
HGSNSSTSTSSSSSKSGKNSGTSTTETITPLLDYKKFTPYKDAYQMSNNFNLSIN  
YGSGLLGYWAGILAIIFANMIKKMFPSLTNNLSGSISNLFKRHLFLPATFRKKKAQEF
```

**Output format:**

☒ Extensive, with graphics  
☐ Extensive, no graphics  
☐ One line per protein

**Other options:**

☐ Use old model (version 1)

**Restrictions:**  
At most 10,000 sequences and 4,000,000 amino acids per submission; each sequence not more than 8,000 amino acids.

**Confidentiality:**  
The sequences are kept confidential and will be deleted after processing.



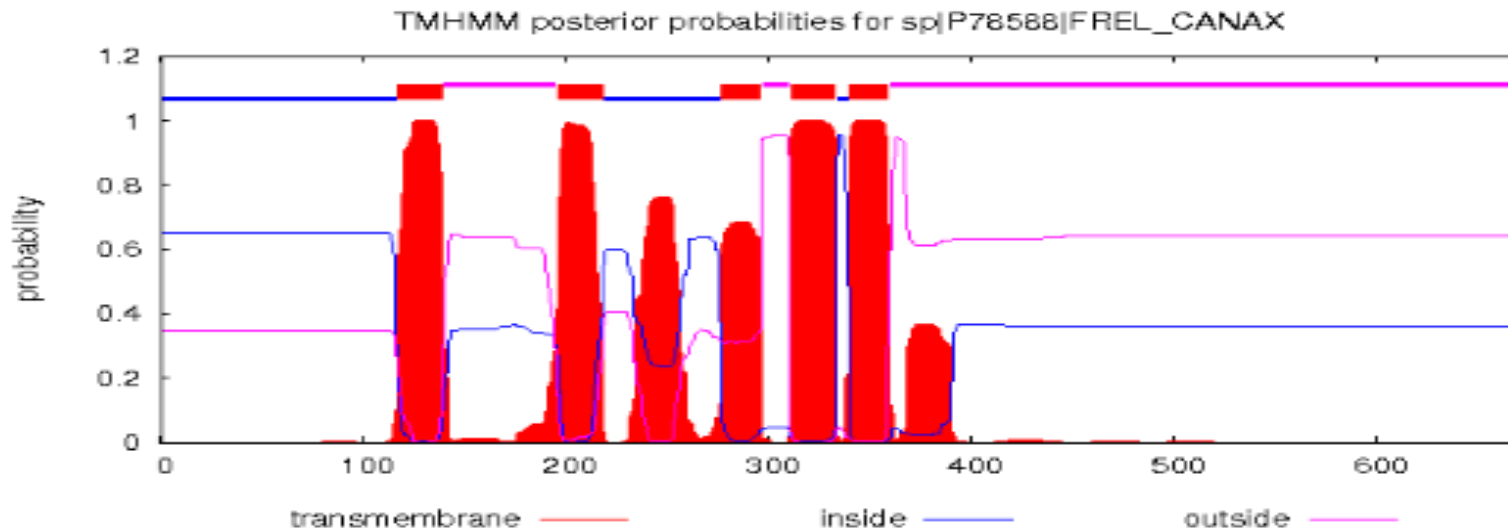
# Predicting Transmembrane using TMHMM

## Example (P78588)

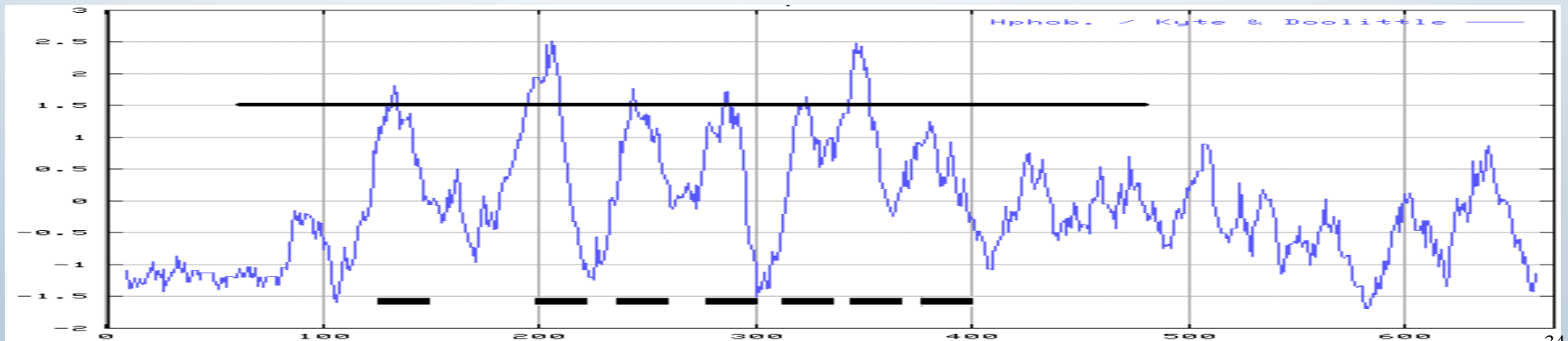
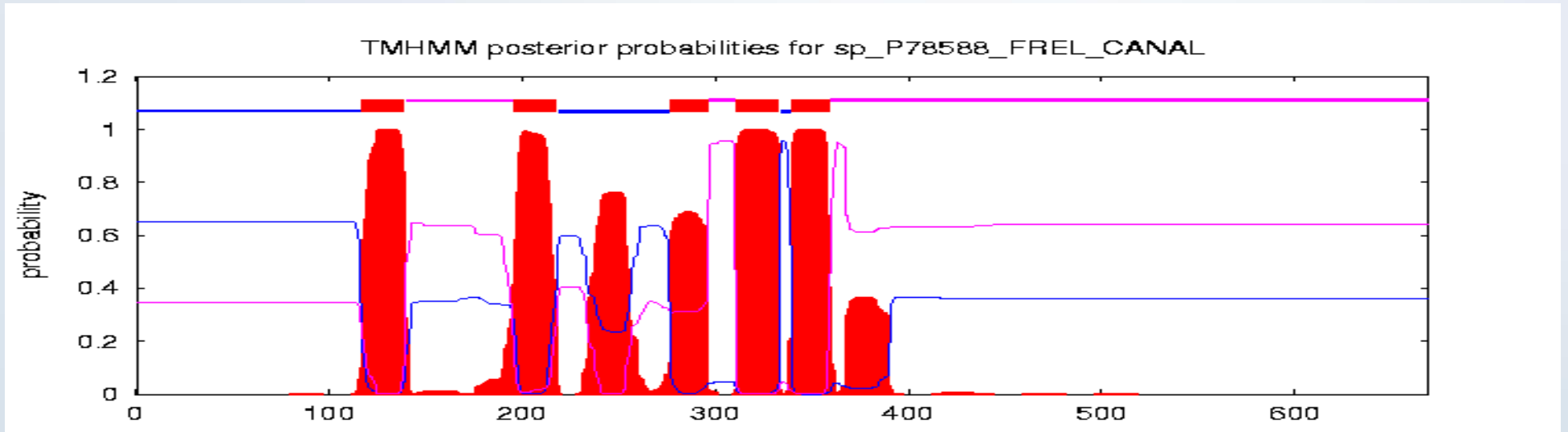
### TMHMM result

[HELP](#) with output formats



```
# sp|P78588|FREL_CANAX Length: 669
# sp|P78588|FREL_CANAX Number of predicted TMHs: 5
# sp|P78588|FREL_CANAX Exp number of AAs in TMHs: 126.67871
# sp|P78588|FREL_CANAX Exp number, first 60 AAs: 0
# sp|P78588|FREL_CANAX Total prob of N-in: 0.65191
sp|P78588|FREL_CANAX TMHMM2.0 inside 1 116
sp|P78588|FREL_CANAX TMHMM2.0 TMhelix 117 139
sp|P78588|FREL_CANAX TMHMM2.0 outside 140 195
sp|P78588|FREL_CANAX TMHMM2.0 TMhelix 196 218
sp|P78588|FREL_CANAX TMHMM2.0 inside 219 276
sp|P78588|FREL_CANAX TMHMM2.0 TMhelix 277 296
sp|P78588|FREL_CANAX TMHMM2.0 outside 297 310
sp|P78588|FREL_CANAX TMHMM2.0 TMhelix 311 333
sp|P78588|FREL_CANAX TMHMM2.0 inside 334 339
sp|P78588|FREL_CANAX TMHMM2.0 TMhelix 340 359
sp|P78588|FREL_CANAX TMHMM2.0 outside 360 669
```



# TMHMM vs. ProtScale



# Potential Cleavage Sites

 **Expasy**  
Bioinformatics Resource Portal

PeptideCutter

Home | [Contact](#)

---

## PeptideCutter

**PeptideCutter** [\[references\]](#) / [\[documentation\]](#) predicts potential cleavage sites cleaved by proteases or chemicals in a given protein sequence. PeptideCutter returns the query sequence with the possible cleavage sites mapped on it and /or a table of cleavage site positions.

Enter a UniProtKB (Swiss-Prot or TrEMBL) protein identifier, ID (e.g. ALBU\_HUMAN), or accession number, AC (e.g. P04406), **or** an amino acid sequence (e.g. 'SERVELAT'):

the cleavage of the protein.  the fields.

---

**Please, select**

☒ all available enzymes and chemicals

☐ only the following selection of **enzymes and chemicals**

<input type="checkbox"/> Arg-C proteinase	<input type="checkbox"/> Asp-N endopeptidase	<input type="checkbox"/> Asp-N endopeptidase + N-terminal Glu
<input type="checkbox"/> BNPS-Skatole	<input type="checkbox"/> Caspase1	<input type="checkbox"/> Caspase2
<input type="checkbox"/> Caspase3	<input type="checkbox"/> Caspase4	<input type="checkbox"/> Caspase5
<input type="checkbox"/> Caspase6	<input type="checkbox"/> Caspase7	<input type="checkbox"/> Caspase8

Start

## Example (plant protein) - Results

[illegible]

# Motifs – Patterns

## DESCRIBING MOTIFS

MOTIF: BIOLOGICALLY IMPORTANT REGION OF PROTEIN  
BASED ON STRUCTURE OR FUNCTION

PROFILE: QUANTATIVE DESCRIPTION OF MOTIF

PATTERN: QUALITATIVE DESCRIPTION OF MOTIF

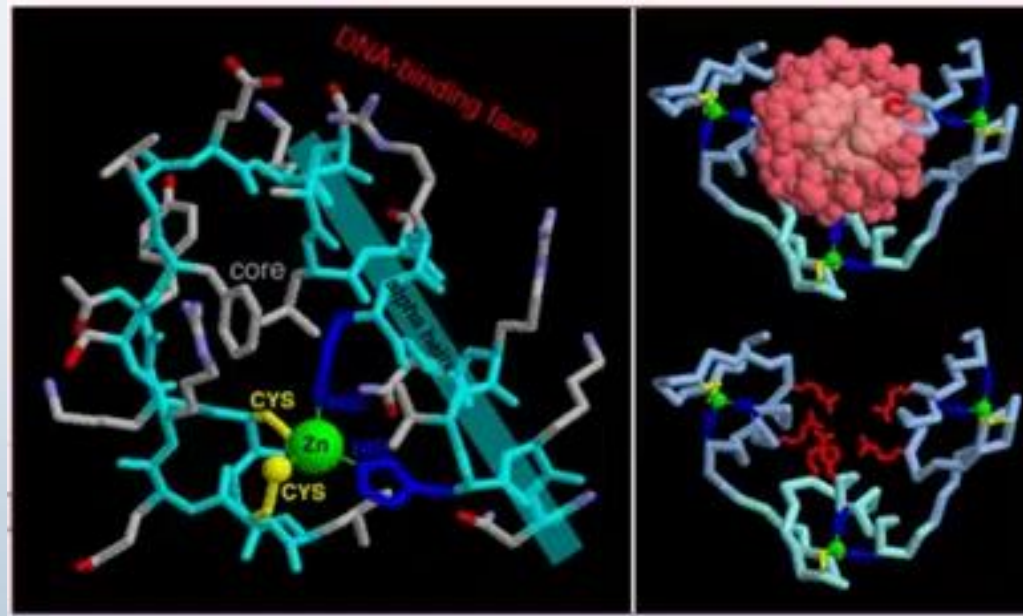
# Pattern expressions

- Patterns: qualitative descriptions
  - Represented by regular expressions
- Protein phosphorylation motif
  - [ST]-X-[RK]
  - Serine or threonine, followed by any amino acid, followed by arginine or lysine
- Cracking code
  - E-X(2)-[FHM]-X(4)-{P}-L
  - E, then any 2, then F, H or M, then any 4, then anything but P, then L
  - x(2,4) means x-x or x-x-x or x-x-x-x

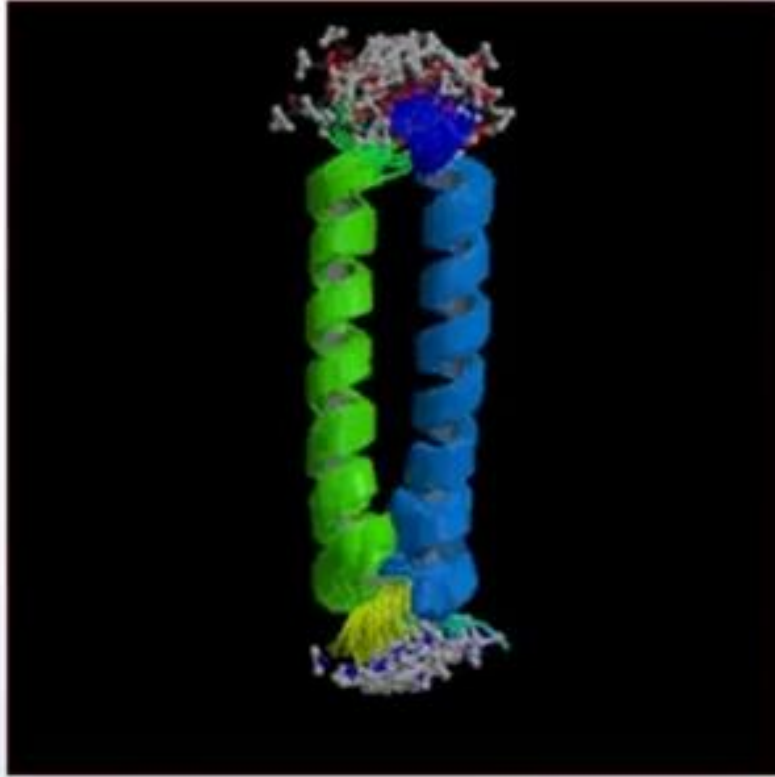


# Zinc finger domain (motif?)

- 2 cys, 2 his
  - Separated by somewhat specific distances
  - Four amino acids bind one zinc molecule
- Aligns with major groove of DNA



# Leucine zipper motif



PDB# 2A93

- Antiparallel  $\alpha$ -helices
  - Held together by hydrophobic interaction between leucines
- Leucines present at every second turn
- Can hold subunits together, bind DNA
- L-x(6)-L-x(6)-L



# Motifs - PROSITE

## PROSITE PROFILES

DESCRIBE MOTIFS USING PSSMs

POSITION-SPECIFIC SCORING MATRIX

MATRIX ALGEBRA USED TO REPRESENT FREQUENCY  
OF AMINO ACIDS AT POSITIONS WITHIN MOTIF

REQUIRES MANY PROTEINS WITH THAT MOTIF

PROSITE USES PSSMs, OTHER DATABASES USE  
HIDDEN MARKOV MODELS (HMMs)

# Example (NP\_180737.3)

### Search

### Browse

- by documentation entry
- by ProRule description
- by taxonomic scope
- by number of positive hits

### Quick Scan mode of ScanProsite

Quickly find matches of your protein sequences to PROSITE signatures (max. 10 sequences). [\[?\]](#) [Examples](#)

YRYFLRSYVEDGKKMKWCPSPGCEYAVEFGVNGSSSYDVSCLCSYKFCWNCCEDAHS  
PVDCE TVSKWLLK  
NKDESENMINWILAKTKPCPKCKRPIEKNTGCNHMSCSAPCRHYFCWACLQPLSDHKA  
CNAFKADNEDETK  
RKRAKDAIDRYTHFYERWAFNQSSRLKAMSDLEKWQSVELKQLSDIQSTPETQLSFT  
VDANLQIIECRRV  
LKWTYAYGGYILSQERNKRVFAS

☐ Exclude motifs with a high probability of occurrence from the scan

For more scanning options go to [ScanProsite](#)

### Other tools

- **PRATT** - allows to interactively generate conserved patterns from a series of unaligned proteins.
- **MyDomains - Image Creator** - allows to generate custom domain figures.

Custom

Images

of

DOMAINS

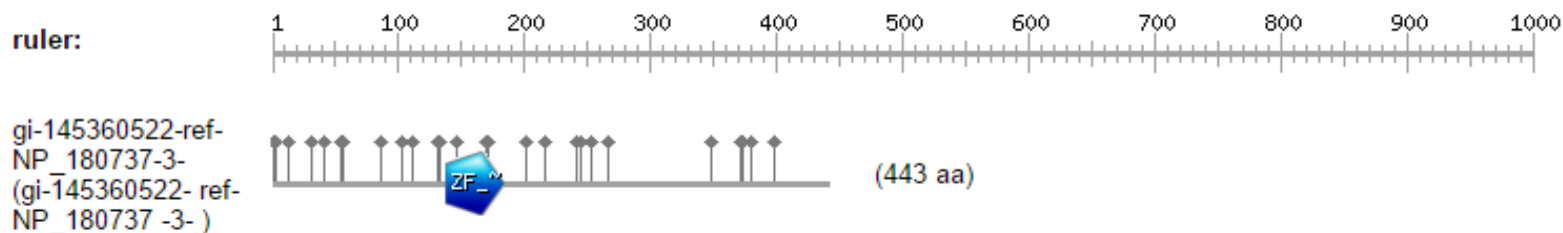
<http://prosite.expasy.org/>

42

### hits by profiles: [1 hit (by 1 profile) on 1 sequence]

Upper case represents match positions, lower case insert positions, and the '-' symbol represents deletions relative to the matching profile.

Hits by [PS50089](#) **ZF\_RING\_2** *Zinc finger RING-type profile* :

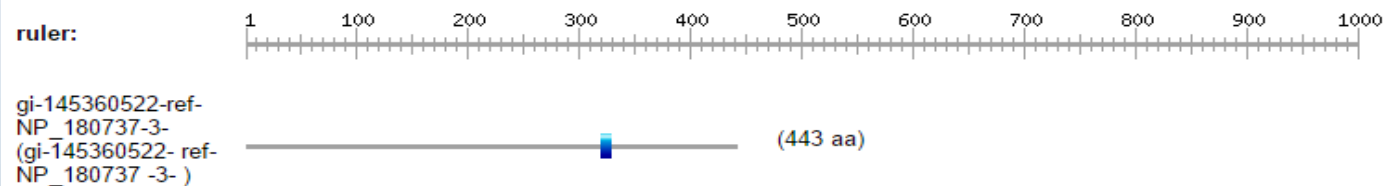


139 - 185: score = 9.421

CGICFESYTRKEIARVSCGHPYCKTCWTGYittkiedGPGCLRVKCP---

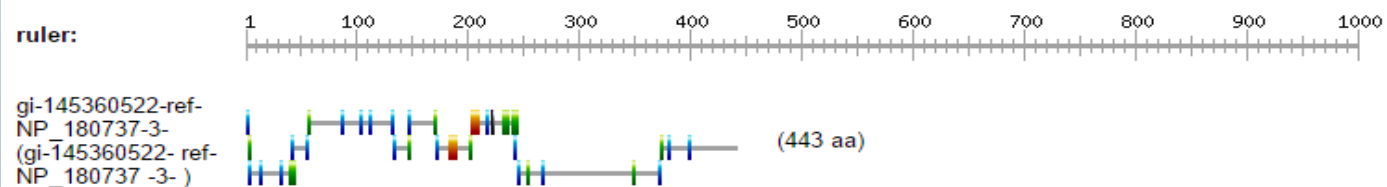
### hits by patterns: [1 hit (by 1 pattern) on 1 sequence]

Hits by [PS00518](#) **ZF\_RING\_1** *Zinc finger RING-type signature* :



320 - 329: [confidence level: (0)] CrHyFCwaCL

### hits by patterns with a high probability of occurrence or by user-defined patterns: [34 hits (by 6 distinct patterns) on 1 sequence]



# Result

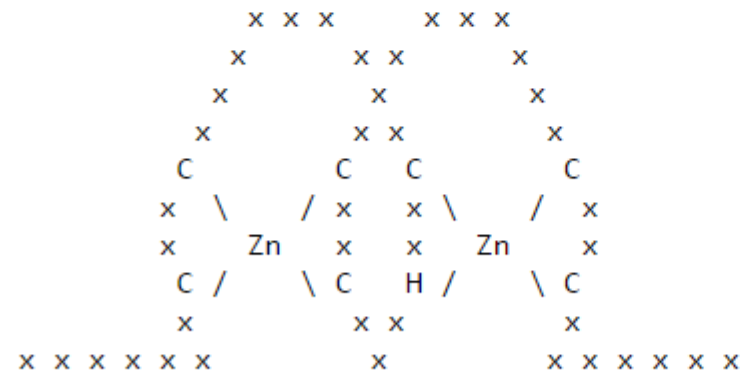
## Zinc finger RING-type signature and profile

Description	Technical section	References	Copyright	Miscellaneous
-------------	-------------------	------------	-----------	---------------

### Description

A number of eukaryotic and viral proteins contain a conserved cysteine-rich domain of 40 to 60 residues (called C3HC4 zinc-finger or 'RING' finger) [1] that binds two atoms of zinc. There are two different variants, the C3HC4-type and the C3H2C3-type, which is clearly related despite the different cysteine/histidine pattern. The latter type is sometimes referred to as "RING-H2 finger".

The 3D structure [2] of the zinc ligation system is referred to as the "cross-brace" motif. This atypical conformation is also shared by the FYVE (see <PDOC50178>) and PHD (see <PDOC50016>) domains. The way the "cross-brace" motif is binding two atoms of zinc is illustrated in the following schematic representation:



'C': conserved cysteine involved zinc binding.  
'H': conserved histidine involved in zinc binding.  
'Zn': zinc atom.

# Result

ZF\_RING\_1, [PS00518](#); Zinc finger RING-type signature (PATTERN)

- Consensus pattern:  
C-x-H-x-[LIVMFY]-C-x(2)-C-[LIVMYA]
- Sequences in UniProtKB/Swiss-Prot known to belong to this class: 1660
  - detected by PS00518: [769](#) (true positives)
  - undetected by PS00518: 891 ([890](#) false negatives and [1](#) 'partial')
- Other sequence(s) in UniProtKB/Swiss-Prot detected by PS00518:  
[6](#) false positives.
- Retrieve an alignment of UniProtKB/Swiss-Prot true positive hits:  
[Clustal format, color, condensed view](#) / [Clustal format, color](#) / [Clustal format, plain text](#) / [Fasta format](#)
- Retrieve the sequence logo from the alignment
- Taxonomic distribution of all UniProtKB (Swiss-Prot + TrEMBL) entries matching PS00518
- Retrieve a list of all UniProtKB (Swiss-Prot + TrEMBL) entries matching PS00518
- Scan UniProtKB (Swiss-Prot and/or TrEMBL) entries against PS00518
- View ligand binding statistics of PS00518
- Matching PDB structures: [1BOR](#) [1CHC](#) [1FBV](#) [1G25](#) ... [\[ALL\]](#)



Link to view the Structure in JSmol

# Result

**PS00005** PKC\_PHOSPHO\_SITE *Protein kinase C phosphorylation site* :

4 - 6: SdR

**Predicted feature:**

MOD\_RES      4                      Phosphoserine                      [condition: S]

PKC\_PHOSPHO\_SITE, [PS00005](#); Protein kinase C phosphorylation site (PATTERN with a high probability of occurrence!)

- Consensus pattern:  
[ST]-x-[RK]  
S or T is the phosphorylation site
- [Scan UniProtKB \(Swiss-Prot and/or TrEMBL\) entries against PS00005](#)
- [View ligand binding statistics of PS00005](#)



# Patterns and Domains

- Patterns are usually the most striking feature of the more general motifs (called domains)
- Domains are less conserved than patterns but usually longer
- In proteins, domain analysis is gradually replacing pattern analysis

# Different Definitions

## InterPro definitions

**TABLE 10-1** Definitions from InterPro Database of Protein Families and Related Terms

Term	Definition
Family	An InterPro family is a group of evolutionarily related proteins that share one or more domains/repeats in common. An InterPro entry of "type = family" may contain a signature for a small conserved region that is representative of the family and therefore need not necessarily cover the whole protein.
Domain	A domain is defined as an independent structural unit which can be found alone or in conjunction with other domains or repeats. Domains are evolutionarily related. Even though the structure of a domain is not always known it is still possible to define the boundaries in many cases from sequence alone. Therefore, sequence criteria can be used to define domain boundaries.
Repeat	An InterPro repeat is a region that is not expected to fold into a globular domain on its own. For example, six to eight copies of the <i>WD40</i> repeat are needed to form a single globular domain. There also many other short repeat motifs that probably do not form a globular fold that have "type = repeat."
Posttranslational modification	A posttranslational modification includes, for example, an N-glycosylation site. The sequence motif is defined by the molecular recognition of this region in a cell. This may group together proteins that need not be evolutionarily related.

Source: Adapted from ► [http://www.ebi.ac.uk/interpro/user\\_manual.html](http://www.ebi.ac.uk/interpro/user_manual.html).



# Different Definitions

## SMART definitions

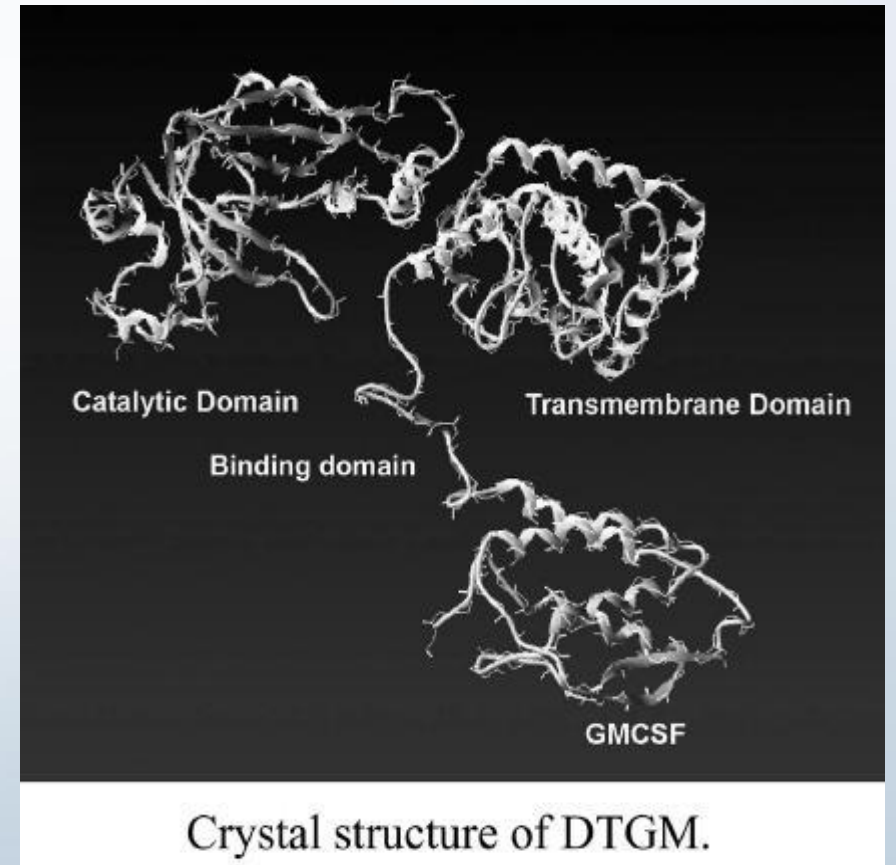
**TABLE 10-2** Definitions of Protein Domains and Motifs from SMART Database

Term	Definition
Domain	Conserved structural entities with distinctive secondary structure content and a hydrophobic core. In small disulfide-rich and $\text{Zn}^{2+}$ -binding or $\text{Ca}^{2+}$ -binding domains, the hydrophobic core may be provided by cystines and metal ions, respectively. Homologous domains with common functions usually show sequence similarities.
Domain composition	Proteins with the same domain composition have at least one copy of each domain of the query.
Domain organization	Proteins having all the domains as the query in the same order (additional domains are allowed).
Motif	Sequence motifs are short conserved regions of polypeptides. Sets of sequence motifs need not necessarily represent homologs.
Profile	A profile is a table of position-specific scores and gap penalties, representing an homologous family that may be used to search sequence databases (Bork and Gibson, 1996).

Source: Adapted from ► [http://smart.embl-heidelberg.de/help/smart\\_glossary.shtml](http://smart.embl-heidelberg.de/help/smart_glossary.shtml).  
SMART is a tool to allow automatic identification and annotation of domains in user-supplied protein sequences (see Chapter 6).

# Protein Domains

- Proteins are usually made of domains
- A domain is an autonomous folding unit
- Domains are more than 50 amino acids long
- It's common to find these together:
  - A regulatory domain
  - A binding domain
  - A catalytic domain



# Discovering Domains

- Researchers discover domains by
  - Comparing proteins that have similar functions
  - Aligning those proteins
  - Identifying conserved segments
- A domain is a multiple-sequence alignment formulated as a profile
- For each column, a domain indicates which amino acid is more likely to occur

# Domain Collections

- Scientists have been discovering and characterizing protein domains for many years
- 8 collections of domains have been established
  - Manual collections are very precise but small
  - Automatic collections are very extensive but less informative
- These collections
  - Overlap
  - Have been assembled by different scientists
  - Have different strengths and weaknesses

# 8 Domain Collections

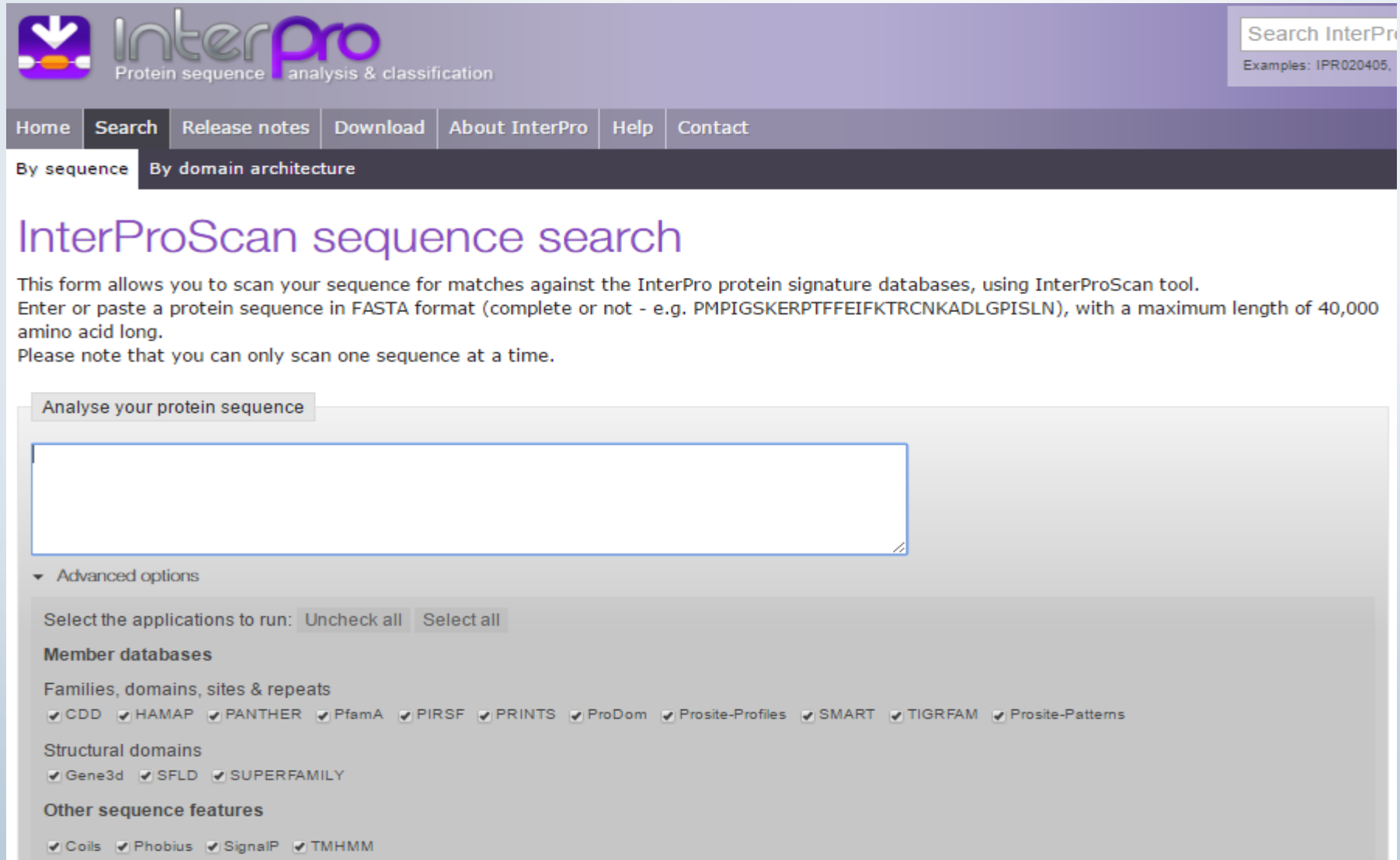
<i><b>Name</b></i>	<i><b>Web Address</b></i>	<i><b>Size</b></i>	<i><b>Generation</b></i>
PROSITE-Profile (IP)	<a href="http://www.expasy.org/prosite">www.expasy.org/prosite</a>	616	Manual
PfamA (IP)	<a href="http://www.sanger.ac.uk/Software/Pfam">www.sanger.ac.uk/Software/Pfam</a>	7973	Manual
PRINTs (IP)	<a href="http://www.bioinf.man.ac.uk/dbbrowsers/PRINTS">www.bioinf.man.ac.uk/dbbrowsers/PRINTS</a>	1900	Manual
PRODOM (IP)	<a href="http://protein.toulouse.inra.fr/prodom/current/html/home.php">protein.toulouse.inra.fr/prodom/current/html/home.php</a>	736000	Automatic
SMART (IP)	<a href="http://smart.embl-heidelberg.de">smart.embl-heidelberg.de</a>	685	Manual
COGs	<a href="http://www.ncbi.nlm.nih.gov/COG/new/">www.ncbi.nlm.nih.gov/COG/new/</a>	4852	Manual
TIGRFAM (IP)	<a href="http://www.tigr.org/TIGRFAMs">www.tigr.org/TIGRFAMs</a>	2453	Manual
BLOCKS	<a href="http://blocks.fhcrc.org/">blocks.fhcrc.org/</a>	12542	Automatic

- Pfam is the most extensive manual collection and is often used as a reference
- **Note:** Some addresses may not work. Please search via the internet for new ones.

# Searching Domain Collections

- Domains in Pfam often include known functions
- A match between your protein and a domain is desirable
  - A match is a potential indication of a function
  - This is **VERY** informative for further research!
- Three servers exist to compare proteins and domain collections:
  - InterProScan
  - CD-Search
  - Motif Scan

# InterProScan



The image shows the InterProScan web interface. At the top, there is a purple header with the InterPro logo and the text 'Protein sequence analysis & classification'. To the right of the logo is a search bar with the placeholder text 'Search InterPro' and examples: 'IPR020405'. Below the header is a navigation bar with links: 'Home', 'Search', 'Release notes', 'Download', 'About InterPro', 'Help', and 'Contact'. Below the navigation bar is a dark purple bar with two tabs: 'By sequence' (selected) and 'By domain architecture'. The main content area has a heading 'InterProScan sequence search'. Below the heading is a paragraph explaining the tool: 'This form allows you to scan your sequence for matches against the InterPro protein signature databases, using InterProScan tool. Enter or paste a protein sequence in FASTA format (complete or not - e.g. PMPIGSKERPTFFEIFKTRCNKADLGPISLN), with a maximum length of 40,000 amino acid long. Please note that you can only scan one sequence at a time.' Below this paragraph is a large text input field with the placeholder text 'Analyse your protein sequence'. Below the input field is a section titled 'Advanced options' with a dropdown arrow. This section contains three sub-sections: 'Select the applications to run:' with 'Uncheck all' and 'Select all' buttons; 'Member databases' with a list of databases and their status (checked/unchecked); and 'Other sequence features' with a list of features and their status (checked/unchecked).

InterPro  
Protein sequence analysis & classification

Search InterPro  
Examples: IPR020405,

Home Search Release notes Download About InterPro Help Contact

By sequence By domain architecture

## InterProScan sequence search

This form allows you to scan your sequence for matches against the InterPro protein signature databases, using InterProScan tool. Enter or paste a protein sequence in FASTA format (complete or not - e.g. PMPIGSKERPTFFEIFKTRCNKADLGPISLN), with a maximum length of 40,000 amino acid long. Please note that you can only scan one sequence at a time.

Analyse your protein sequence

Advanced options

Select the applications to run: Uncheck all Select all

**Member databases**

Families, domains, sites & repeats

☒ CDD ☒ HAMAP ☒ PANTHER ☒ PfamA ☒ PIRSF ☒ PRINTS ☒ ProDom ☒ Prosite-Profiles ☒ SMART ☒ TIGRFAM ☒ Prosite-Patterns

Structural domains

☒ Gene3d ☒ SFLD ☒ SUPERFAMILY

**Other sequence features**

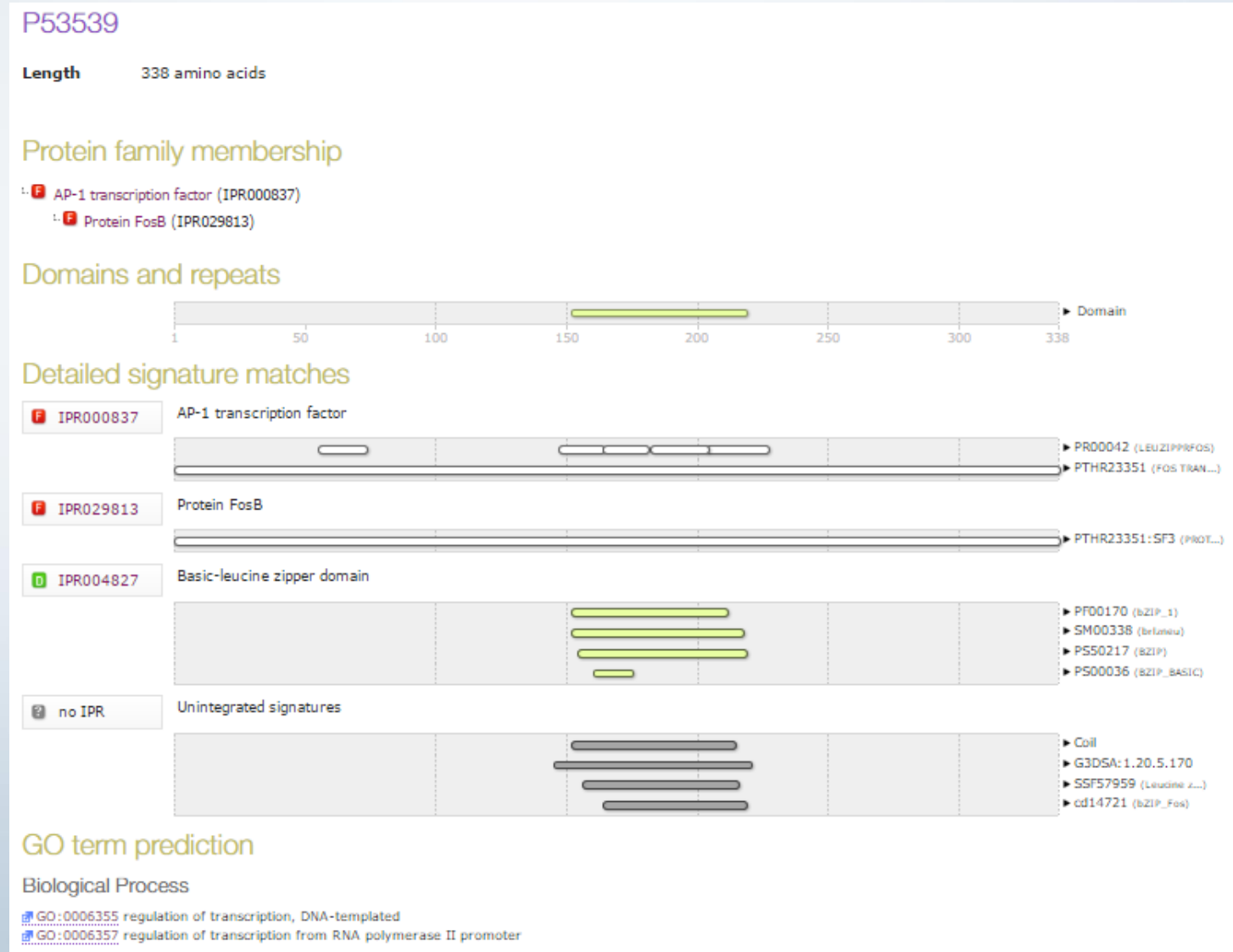
☒ Coils ☒ Phobius ☒ SignalP ☒ TMHMM

<https://www.ebi.ac.uk/interpro/search/sequence-search>




# InterProScan – Example (P53539)

- InterProScan is the most comprehensive search engine for domain databases
- Makes it possible to compare alternative results on most collections
- Does not provide a statistical score



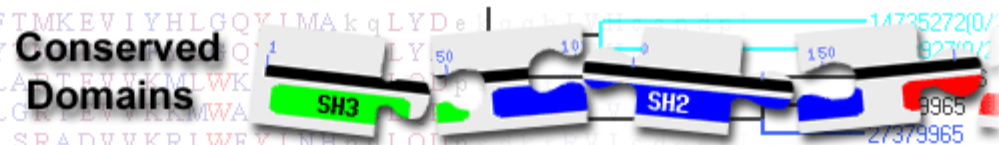


# CD-Search



HOMESEARCHGUIDEStructure Home3D Macromolecular StructuresConserved DomainsPubchemBioSystems

Conserved Domains



NCBI will be testing https on public web servers from 1:00-4:00 PM EDT (17:00-20:00 UTC) on Monday, October 24. You may experience problems with NCBI services, especially file downloads, during that time. Please plan accordingly. [Read more.](#)

Search for Conserved Domains within a protein or coding nucleotide sequence

**NEW!** Use **Batch CD-search** to submit multiple query proteins at once!

Enter **protein** or **nucleotide** query as accession, gi, or sequence in [FASTA format](#) ?

**OPTIONS**

Search against database ? : CDD v3.15 - 48963 PSSMs ▾

Expect Value ? threshold: 0.010000

Apply low-complexity filter ? ☐

Composition based statistics adjustment ? ☒

Force live search ? ☐

Rescue borderline hits ☐ Suppress weak overlapping hits ☐

Maximum number of hits ? 500

Result mode ☒ Concise ? ☐ Standard ? ☐ Full ?

Submit

Reset

<https://www.ncbi.nlm.nih.gov/Structure/cdd/wrpsb.cgi>


# CD-Search - Example (P53539)


- CD search is less extensive than that of InterProScan
- Results come with a statistical evaluation (E-value)
  - $10^{-23}$  **Low E-value**      Good match


**Conserved domains on** [sp|P53539] View Concise Results ?

FOSB\_HUMAN Protein fosB OS=Homo sapiens GN=FOSB PE=2 SV=1

**Graphical summary** ☐ Zoom to residue level show extra options ?

Query seq. 

**Specific hits** 

**Superfamilies** 

Search for similar domain architectures ? Refine search ?

**List of domain hits** ?

	Name	Accession	Description	Interval	E-value
[+]	bZIP_Fos	cd14721	Basic leucine zipper (bZIP) domain of the oncogene Fos (Fos): a DNA-binding and dimerization ...	165-218	4.10e-23

# Motif Scan

## Motif Scan

Motif scanning means finding all known motifs that occur in a sequence. This form lets you paste a protein sequence, select the collections of motifs to scan for, and launch the search.

A [document](#) deals with the interpretation of the match scores. You should consult the home pages of [Prosites](#) on ExPASy, [Pfam](#) and [InterPro](#) for additional information.

If your proteins of interest are already in the sequence databases (see [list](#)), the [Query by Protein](#) form is much faster, and the [Protein Hub](#) provides a collection of tools that you might find useful.

Protein Identifiers  
or Protein Sequence

examples ▼

mot\_source

- ☒ perox - PeroxiBase profiles
- ☒ hamap - HAMAP profiles
- ☒ pat - PROSITE patterns
- ☒ freq\_pat - PROSITE patterns (frequent match producers)
- ☒ prf - PROSITE profiles
- ☒ pre - More profiles
- ☒ pfam\_fs - Pfam HMMs (local models)
- ☒ pfam\_ls - Pfam HMMs (global models)

The scan might take  
a few minutes.

[http://myhits.isb-sib.ch/cgi-bin/motif\\_scan](http://myhits.isb-sib.ch/cgi-bin/motif_scan)

# Motif Scan - Example (P53539)

## Motif Scan Results

**Query Protein** temporarily stored [here](#).

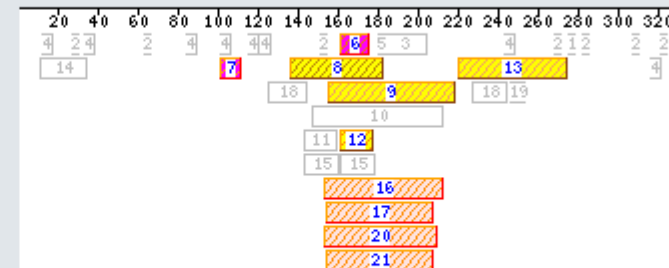
**Database of motifs** PeroxiBase profiles [perox], HAMAP profiles [hamap], PROSITE patterns [pat], More profiles [pre], Pfam HMMs (local models) [pfam\_fs], Pfam HMMs (global models) [pfam\_ls], PROSITE patterns (frequent match producers) [freq\_pat], PROSITE profiles [prf].

searching PeroxiBase profiles  
searching HAMAP profiles  
searching PROSITE patterns  
searching PROSITE patterns (frequent match producers)  
searching PROSITE profiles  
searching More profiles  
searching Pfam HMMs (local models)  
searching Pfam HMMs (global models)  
postprocessing

### Summary

**Original output** [perox](#), [hamap](#), [pat](#), [freq\\_pat](#), [prf](#), [pre](#), [pfam\\_fs](#), [pfam\\_ls](#).

**Matches map**  
(features from query are  
above the ruler, matches of  
the motif scan are below the  
ruler)



Legends: 1, freq\_pat:ASN\_GLYCOSYLATION [?]; 2, freq\_pat:CK2\_PHOSPHO\_SITE [?]; 3, freq\_pat:LEUCINE\_ZIPPER [?]; 4, freq\_pat:MYRISTYL [?]; 5, freq\_pat:PKC\_PHOSPHO\_SITE [?]; 6, pat:BZIP\_BASIC [!]; 7, pat:SUBTILASE\_SER [!]; 8, prf:ARG\_RICH [!]; 9, prf:BZIP [!]; 10, prf:GLU\_RICH [?]; 11, prf:NLS\_BP [?]; 12, prf:NLS\_BP [!]; 13, prf:PRO\_RICH [!]; 14, prf:SER\_RICH [?]; 15, pre:NLS\_BP [?]; 16, pfam\_fs:bZIP\_1 [!]; 17, pfam\_fs:bZIP\_2 [!]; 18, pfam\_ls:OGFr\_III [?]; 19, pfam\_ls:Octapeptide [?]; 20, pfam\_ls:bZIP\_1 [!]; 21, pfam\_ls:bZIP\_2 [!].

# Looking into the Details

- Catalytic residues are normally highly conserved in domains
- Motif Scan makes it possible to check whether these important residues are conserved in your sequence
  - **High bar above 0** = Highly conserved residues
  - Green = Your sequence has an expected residue
  - Red = Your sequence has an unexpected residue



# Looking into the Details



- ✓ R (Arginine) is highly expected at this position  
High bar  
Potential active site
- ✓ If your protein has an arginine on this position . . .  
Bar is filled with green  
Your protein could be active

# Predicting Post-translational Modifications

- Post-translational modifications often occur on similar motifs in different proteins
- PROSITE is a database containing a list of known motifs, each associated with a function or a post-translational modification
- You can search PROSITE by looking for each motif it contains in your protein
- PROSITE entries come with an extensive documentation on each function of the motif

# Predicting Functions with Domains

- Finding a match with a domain having a catalytic function is good news . . . but what, exactly, does it mean?
- A match indicates that your sequence has the domain structure . . . but does it also have the function?
- You cannot say before looking into these details:
  - Where are the catalytic residues on the domain?
  - Does your sequence have the right residues at these positions?



# Function Prediction Methods

- Homology-based methods
- Sequence motif/domain-based methods
- Structure-based methods
- Genomic context-based methods
  - Gene fusion
  - Co-location/co-expression
- Computational Solvent Mapping
- Network-based methods

# SIFTER – Example (P0C871)

**SIFTER Protein Function Prediction**[Home](#)[About](#)[Download](#)[Contacts](#)[Help](#)

## Phylogeny-based Protein Function Prediction

**SIFTER** (**S**tatistical **I**nference of **F**unction **T**hrough **E**volutionary **R**elationships) is a statistical approach to predicting protein function that uses a protein family's phylogenetic tree, as the natural structure for representing protein relationships.

[Learn more »](#)

## Search SIFTER

Find SIFTER predictions for your proteins, species, or functions.

Enter any Uniprot ID or Accession, Go term ID, Function name, Species Name or ID, etc.

[Example 1](#) [Example 2](#) [Example 3](#) [Example 4](#)

[Submit](#) [Reset](#)

### Quick Search

### Advanced Search

Predict by **Protein ID**

Predict for all proteins of a **Species**

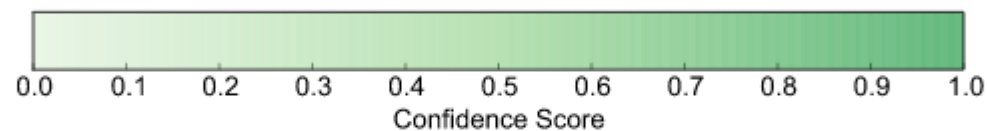
Find proteins that have given **Functions**

Predict for homologs of given **Sequences**

- SIFTER was recently honored as the best-performing sequence-based protein function prediction method in the [Critical Assessment of Function Annotation](#).
- SIFTER webserver is updated on Aug 12, 2015:**
  - Results are obtained using the family data from [Pfam](#) v27.0, the gene ontology data from [GO](#) (update 03/31/2015), and the annotation data from [UniProt-GOA](#) (update 03/31/2015).
  - Currently SIFTER has predictions for **17,692,644** proteins across **331,365** species.

# Example (PA24B\_MOUSE - P0C871)

- **Job ID:** 8262450
- **Query Mode:** by\_protein
- **Number of Query Proteins:** 1
- **Number of Proteins with Predictions:** 1
- **SIFTER Scheme:** EXP-Model
- **Submission Date:** Oct. 25, 2016



SIFTER Predictions for Job ID: 8262450

[Download results](#)

PA24B_MOUSE (See domain details)		<i>Mus musculus</i>	Confidence Score
GO:0047498	calcium-dependent phospholipase A2 activity		0.89
GO:0047499	calcium-independent phospholipase A2 activity		0.58
GO:0004622	lysophospholipase activity		0.55
GO:0005509	calcium ion binding		0.55
GO:0005544	calcium-dependent phospholipid binding		0.55
GO:0035035	histone acetyltransferase binding		0.53

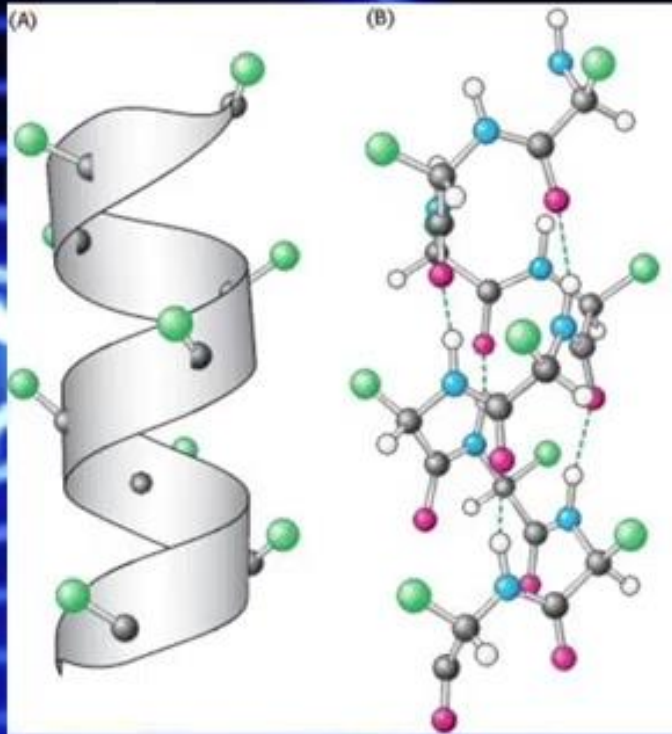
# Protein Function Prediction – General Protocol

1. Similarity search: First start with Blastp, if your sequence is less than 40% identity go for PSI-Blast
2. Domain search: Do domain search using Interproscan, Pfam or CDART
3. Search for signal peptide and transmembrane (TM): search for signal peptide using signalp and TM using TMHMM, phobius
4. Comparative modelling: Do homology modelling using swiss model, if your sequence less than 40% identity from blast result go for ab-intio modelling using I-Tasser
5. Gene ontology classification: You can search sequence for GO classification using blast2go or STRAP
6. Functional association prediction: Try searching sequence using STRING search



# Secondary Structure Prediction

# ALPHA-HELICES



BERG, *BIOCHEMISTRY*, 5TH ED.

MOST ABUNDANT SECONDARY STRUCTURE

3.6 AMINO ACIDS PER TURN

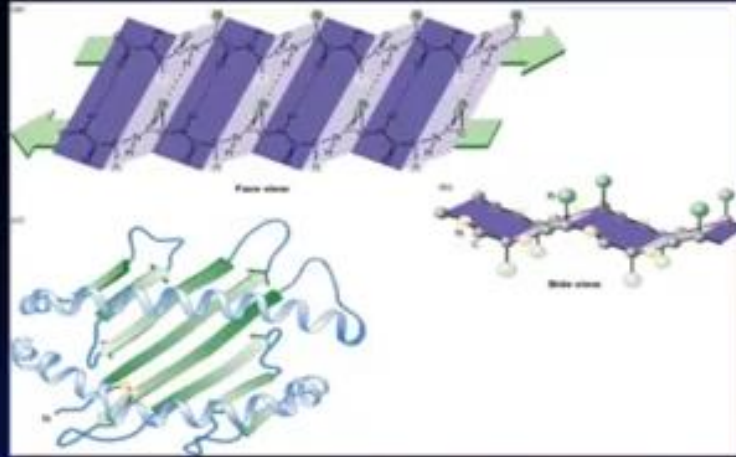
AVERAGE LENGTH: 10aa (VARIES FROM 5-40aa)

INNER-FACING SIDE CHAINS HYDROPHOBIC

3RD OF EVERY 4 AMINO ACIDS HYDROPHOBIC



# BETA-SHEETS (STRANDS)



LODISH ET AL.,  
*MOLECULAR CELL BIOLOGY*

H-BONDS BETWEEN 2 SEPARATE REGIONS OF CHAIN  
~5-10AA; EACH REGION:  $\beta$ -STRAND

PARALLEL - CHAINS RUN IN SAME DIRECTION  
N TO C TERMINAL

ANTI-PARALLEL - CHAINS RUN OPPOSITE

# Secondary Structure Prediction Method

- **HNN** (Hierarchical Neural Network )

*[https://npsa-prabi.ibcp.fr/cgi-bin/npsa\\_automat.pl?page=/NPSA/npsa\\_hnn.html](https://npsa-prabi.ibcp.fr/cgi-bin/npsa_automat.pl?page=/NPSA/npsa_hnn.html)*

- **PHD** (Profile network from Heidelberg)

*[https://npsa-prabi.ibcp.fr/cgi-bin/npsa\\_automat.pl?page=/NPSA/npsa\\_phd.html](https://npsa-prabi.ibcp.fr/cgi-bin/npsa_automat.pl?page=/NPSA/npsa_phd.html)*



# Example

## SEQUENCES


>plant protein - eight beta strands

```
GSSGSSGPHGTLEVVLVSAKGLEDADFLNNMDPYVQLTCRTQDQKSNVAEGMGTTPEWNETFIFTVSEGT  
TELKAKIFDKDVGTEDDAVGEATIPLEPVFVEGSIPPTAYNVVKDEEYKGEIWVALSFKPSGPSSG
```

>Thermus thermophilus Hb8 - histone fold - six alpha helices

```
XLXKVAEFERLFRQAAGLDVDKNDLKRVSDFLRNKLYDLLAVAERNAKYNGRDLIFEPDLPIAKGLQETL  
QEFRXDTALELKPVLDALAALPPLDLEVAEDVRNLLPELAGALVVAYARVLKELDPALKNPQTEHHERA  
ERVFNLL
```

# HNN Algorithm



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## HNN SECONDARY STRUCTURE PREDICTION METHOD

[\[Abstract\]](#) [\[NPS@ help\]](#) [\[Original server\]](#)

Sequence name (optional) :

Paste a protein sequence below : [help](#)

```
GSSGSSGPHGTLEVVLVSAKGLEDADFLNNMDPYVQLTCRTQDQKSNV
AEGMGTTPEWNETFIFTVSEGT
TELKAKIFDKDVGTEDDAVGEATIPLEPVFVEGSIPPTAYNVVKDEEYKG
EIWVALSFKPSGPSSG
```

Output width :

[https://npsa-prabi.ibcp.fr/cgi-bin/npsa\\_automat.pl?page=/NPSA/npsa\\_hnn.html](https://npsa-prabi.ibcp.fr/cgi-bin/npsa_automat.pl?page=/NPSA/npsa_hnn.html)

# HNN Algorithm

[illegible]

Sequence length : 136

```

HNN :
Alpha helix      (Hh) :    17 is  12.50%
310 helix      (Gg) :     0 is   0.00%
Pi helix         (Ii) :     0 is   0.00%
Beta bridge      (Bb) :     0 is   0.00%
Extended strand (Ee) :    33 is  24.26%
Beta turn        (Tt) :     0 is   0.00%
Bend region      (Ss) :     0 is   0.00%
Random coil      (Cc) :    86 is  63.24%
Ambiguous states (?) :     0 is   0.00%
Other states      :     0 is   0.00%

```

## Actual Locations

3-15

29-32

37-39

51-61


66-71

84-89

98-108

111-124

# PHD Algorithm



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## PHD SECONDARY STRUCTURE PREDICTION METHOD

[\[Abstract\]](#) [\[NPS@ help\]](#) [\[Original server\]](#)

Sequence name (optional) :

Paste a protein sequence below : [help](#)

```
XLXKVAEFERLFRQAAGLDVDKNDLKRVSDFLRNKLYDLLAVAERNAK
YNGRDLIFEPDLPIAKGLQETL
QEFRRXDTALELKPVLDAALPPLDLEVAEDVRNLLPELAGALVVAYAR
VLKELDPALKNPQTEHHERA
ERVFNLLL
```

Output width :

[https://npsa-prabi.ibcp.fr/cgi-bin/npsa\\_automat.pl?page=/NPSA/npsa\\_phd.html](https://npsa-prabi.ibcp.fr/cgi-bin/npsa_automat.pl?page=/NPSA/npsa_phd.html)

# PHD Algorithm

10 20 30 40 50 60 70  
| | | | | | |  
XLXKVAEFERLFRQAAGLDVDKNDLKRVSDFLRNKLYDLLAVAERNAKYNGRDLIFEPDLPIAKGLQETL  
ChHHHHHHHHHHHHHHhhccccHHHHHHHHHHHHHHHHHHHHHHHHHHHHcCCceEEcCCCchHHHHHHHHH  
QEFRRXDTALELKPVLDAALPPLDLEVAEDVRNLLPELAGALVVAYARVLKELDPALKNPQTEHHERA  
HHHHHhHHHHHHHHHHHHHHhhcCCCCchHHHHHHHHHHHHHHHHHHHHHHHHHHHHHHhCCCCchHHHHH  
ERVFNLLL  
HHHHHHHC

Sequence length : 148

PHD :

Alpha helix	(Hh)	:	120	is	81.08%
3 <sub>10</sub> helix	(Gg)	:	0	is	0.00%
Pi helix	(Ii)	:	0	is	0.00%
Beta bridge	(Bb)	:	0	is	0.00%
Extended strand	(Ee)	:	3	is	2.03%
Beta turn	(Tt)	:	0	is	0.00%
Bend region	(Ss)	:	0	is	0.00%
Random coil	(Cc)	:	25	is	16.89%
Ambiguous states (?)		:	0	is	0.00%
Other states		:	0	is	0.00%

## Actual Locations

5-16

25-49

64-74

82-90

101-125

134-147