

Pseudo Code: Parkinson's Disease Motor Progression Prediction Model

Stacking Regressor with Explainable AI (SHAP)

Version: 1.0

Date: November 2025

Performance: $R^2=0.551$, $MAE=6.01$ (Independent Clinical Test Set)

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Overview

System Architecture

Plain Text

TRAINING PIPELINE

```
[1] Load Data → [2] Preprocess → [3] Feature Engineering →  
[4] Split Data → [5] Optimize Hyperparameters →  
[6] Train Final Model → [7] Validate → [8] SHAP Analysis
```

PREDICTION PIPELINE

```
[1] Load Patient Data → [2] Preprocess → [3] Scale Features →  
[4] Predict → [5] Inverse Transform → [6] Generate Report
```

Model Components

- **Base Models:** XGBoost, LightGBM, CatBoost
- **Meta-Learner:** Huber Regressor
- **Explainability:** SHAP (SHapley Additive exPlanations)
- **Validation:** 7-Fold Cross-Validation + Independent Holdout

Main Training Pipeline

Algorithm 1: Complete Training Pipeline

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ALGORITHM: TrainParkinsonsProgressionModel()

INPUT:

- clinical_data: DataFrame with baseline clinical features
- rnaseq_data: DataFrame with gene expression data (RNA-seq)
- random_seed: Integer for reproducibility (default: 42)

OUTPUT:

- trained_model: Optimized Stacking Regressor
- performance_metrics: Dictionary with R², MAE, RMSE
- shap_values: SHAP importance values for features
- model_artifacts: Scalers, transformers, feature names

PROCEDURE:

1. INITIALIZE

```
SET random_seed = 42
SET n_top_genes = 100
SET n_optuna_trials = 30
SET n_cv_folds = 7
SET test_size = 0.2
```

```

2. LOAD AND MERGE DATA
  clinical_df ← LOAD_CSV("final_dataset.csv")
  rnaseq_df ← LOAD_CSV("rnaseq_baseline_filtered.csv")
  merged_df ← MERGE(clinical_df, rnaseq_df, on="PATNO")

  PRINT "Loaded", LENGTH(merged_df), "patients"

3. REMOVE OUTLIERS
  Q1 ← QUANTILE(merged_df["UPDRS_V04"], 0.25)
  Q3 ← QUANTILE(merged_df["UPDRS_V04"], 0.75)
  IQR ← Q3 - Q1

  outlier_mask ← (UPDRS_V04 < Q1 - 1.5×IQR) OR (UPDRS_V04 > Q3 +
1.5×IQR)
  clean_df ← merged_df[NOT outlier_mask]

  PRINT "Removed", SUM(outlier_mask), "outliers"

4. SELECT TOP GENES
  gene_correlations ← []

  FOR EACH gene IN gene_columns:
    corr ← CORRELATION(clean_df[gene], clean_df["DELTA_UPDRS"])
    IF NOT IS_NAN(corr):
      APPEND (gene, ABS(corr)) TO gene_correlations

  SORT gene_correlations BY correlation DESC
  top_genes ← FIRST 100 genes FROM gene_correlations

  PRINT "Selected top 100 genes with correlation range:",
        MAX(corr), "to", MIN(corr)

5. ENGINEER FEATURES
  // Clinical features
  clinical_features ← ["UPDRS_BL", "AGE", "GENDER"]

  // PD risk genes
  pd_genes ← ["PD_SNCA", "PD_LRRK2", "PD_GBA", "PD_PRKN",
              "PD_PINK1", "PD_PARK7", "PD_VPS35"]

  // Pathway scores
  pathway_features ← ["PATHWAY_Inflammation",
                      "PATHWAY_Mitochondrial",
                      "PATHWAY_Autophagy"]

  // Create interaction features
  clean_df["PINK1_x_PARK7"] ← clean_df["PD_PINK1"] ×

```

```

clean_df["PD_PARK7"]
    clean_df["AGE_x_PINK1"] ← clean_df["AGE"] × clean_df["PD_PINK1"]
    clean_df["UPDRS_BL_x_PINK1"] ← clean_df["UPDRS_BL"] ×
clean_df["PD_PINK1"]

    interaction_features ← ["PINK1_x_PARK7", "AGE_x_PINK1",
"UPDRS_BL_x_PINK1"]

    // Combine all features
    all_features ← clinical_features + top_genes + pd_genes +
pathway_features + interaction_features

PRINT "Total features:", LENGTH(all_features)

6. PREPARE DATA
    X ← clean_df[all_features]
    y_v04 ← clean_df["UPDRS_V04"] // Target: 12-month UPDRS
    y_clf ← (clean_df["DELTA_UPDRS"] >= 5) // For stratification

    // Transform target variable
    target_transformer ← PowerTransformer(method="yeo-johnson")
    y_transformed ← target_transformer.FIT_TRANSFORM(y_v04)

7. SPLIT DATA (Stratified)
    (X_trainval, X_test,
     y_trainval_trans, y_test_trans,
     y_trainval_orig, y_test_orig) ← TRAIN_TEST_SPLIT(
        X, y_transformed, y_v04,
        test_size=0.2,
        stratify=y_clf,
        random_state=random_seed
    )

    PRINT "Train+Val:", LENGTH(X_trainval), "Test:", LENGTH(X_test)

8. OPTIMIZE HYPERPARAMETERS
    best_params ← OPTIMIZE_HYPERPARAMETERS(
        X_trainval, y_trainval_trans,
        n_trials=30,
        n_folds=7
    )

    PRINT "Best CV R2:", best_params["cv_r2"]

9. TRAIN FINAL MODEL
    // Scale features
    scaler ← StandardScaler()
    X_trainval_scaled ← scaler.FIT_TRANSFORM(X_trainval)

```

```

X_test_scaled ← scaler.TRANSFORM(X_test)

// Build ensemble with best hyperparameters
final_model ← BUILD_STACKING_ENSEMBLE(best_params)

// Train on full training set
final_model.FIT(X_trainval_scaled, y_trainval_trans)

PRINT "Final model trained"

10. VALIDATE ON TEST SET
    // Predict on test set
    y_test_pred_trans ← final_model.PREDICT(X_test_scaled)

    // Inverse transform to original scale
    y_test_pred ← target_transformer.INVERSE_TRANSFORM(y_test_pred_trans)

    // Calculate metrics
    test_r2 ← R2_SCORE(y_test_orig, y_test_pred)
    test_mae ← MEAN_ABSOLUTE_ERROR(y_test_orig, y_test_pred)
    test_rmse ← SQRT(MEAN_SQUARED_ERROR(y_test_orig, y_test_pred))

    PRINT "Test R2:", test_r2
    PRINT "Test MAE:", test_mae
    PRINT "Test RMSE:", test_rmse

11. COMPUTE SHAP VALUES
    shap_values ← COMPUTE_SHAP_VALUES(
        final_model,
        X_test_scaled,
        feature_names=all_features
    )

    PRINT "SHAP analysis complete"

12. SAVE MODEL ARTIFACTS
    model_package ← {
        "ensemble_model": final_model,
        "scaler": scaler,
        "target_transformer": target_transformer,
        "feature_names": all_features,
        "n_features": LENGTH(all_features),
        "cv_results": best_params,
        "test_results": {
            "r2": test_r2,
            "mae": test_mae,
            "rmse": test_rmse
        },
    }

```

```

        "shap_values": shap_values
    }

SAVE_PICKLE(model_package, "lightweight_optimized_model.pkl")

PRINT "Model saved successfully"

13. RETURN
    RETURN model_package

END ALGORITHM

```

Data Preprocessing

Algorithm 2: Data Loading and Cleaning

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```

ALGORITHM: LoadAndCleanData()

INPUT:
- clinical_file_path: Path to clinical data CSV
- rnaseq_file_path: Path to RNA-seq data CSV

OUTPUT:
- clean_data: Preprocessed DataFrame
- n_outliers_removed: Number of outliers removed

PROCEDURE:
1. LOAD DATA
    clinical_df ← READ_CSV(clinical_file_path)
    rnaseq_df ← READ_CSV(rnaseq_file_path)

    PRINT "Clinical data:", SHAPE(clinical_df)
    PRINT "RNA-seq data:", SHAPE(rnaseq_df)

2. CONVERT PATIENT IDs TO STRING
    clinical_df["PATNO"] ← TO_STRING(clinical_df["PATNO"])
    rnaseq_df["PATNO"] ← TO_STRING(rnaseq_df["PATNO"])

3. EXTRACT GENE COLUMNS
    gene_columns ← [col FOR col IN rnaseq_df.columns
                    IF col STARTS_WITH "ENSG"]

    rnaseq_genes ← rnaseq_df[["PATNO"] + gene_columns]

```

```

PRINT "Gene columns:", LENGTH(gene_columns)

4. MERGE DATASETS
merged_df ← INNER_JOIN(clinical_df, rnaseq_genes, on="PATNO")

PRINT "Merged patients:", LENGTH(merged_df)

5. HANDLE MISSING VALUES IN TARGET
IF ANY(IS_NULL(merged_df["UPDRS_V04"])):
    median_updrs ← MEDIAN(merged_df["UPDRS_V04"])
    merged_df["UPDRS_V04"].FILL_NA(median_updrs)

PRINT "Filled", SUM(IS_NULL), "missing UPDRS values"

6. DETECT AND REMOVE OUTLIERS (IQR Method)
Q1 ← QUANTILE(merged_df["UPDRS_V04"], 0.25)
Q3 ← QUANTILE(merged_df["UPDRS_V04"], 0.75)
IQR ← Q3 - Q1

lower_bound ← Q1 - 1.5 × IQR
upper_bound ← Q3 + 1.5 × IQR

outlier_mask ← (merged_df["UPDRS_V04"] < lower_bound) OR
                (merged_df["UPDRS_V04"] > upper_bound)

n_outliers ← SUM(outlier_mask)
clean_df ← merged_df[NOT outlier_mask]

PRINT "Outliers removed:", n_outliers
PRINT "Clean data size:", LENGTH(clean_df)

7. RETURN
RETURN clean_df, n_outliers

END ALGORITHM

```

Algorithm 3: Gene Selection by Correlation

Plain Text

ALGORITHM: SelectTopGenes(data, n_top=100)

INPUT:

- data: DataFrame with gene expression and DELTA_UPDRS
- n_top: Number of top genes to select (default: 100)

OUTPUT:

- top_genes: List of top gene names
- correlations: Dictionary mapping genes to correlation values

PROCEDURE:

1. IDENTIFY GENE COLUMNS

```
gene_columns ← [col FOR col IN data.columns  
IF col STARTS_WITH "ENSG"]
```

```
PRINT "Total genes available:", LENGTH(gene_columns)
```

2. COMPUTE CORRELATIONS

```
gene_correlations ← []
```

```
FOR EACH gene IN gene_columns:
```

```
IF gene IN data.columns:
```

```
// Compute Pearson correlation with progression
```

```
corr_matrix ← CORRELATION(data[[gene, "DELTA_UPDRS"]])
```

```
corr_value ← corr_matrix[0, 1]
```

```
IF NOT IS_NAN(corr_value):
```

```
abs_corr ← ABSOLUTE_VALUE(corr_value)
```

```
APPEND (gene, abs_corr, corr_value) TO gene_correlations
```

```
PRINT "Valid correlations computed:", LENGTH(gene_correlations)
```

3. SORT BY ABSOLUTE CORRELATION

```
SORT gene_correlations BY abs_corr DESCENDING
```

4. SELECT TOP N GENES

```
top_genes ← []
```

```
correlations ← {}
```

```
FOR i FROM 0 TO n_top-1:
```

```
gene_name ← gene_correlations[i][0]
```

```
corr_value ← gene_correlations[i][2]
```

```
APPEND gene_name TO top_genes
```

```
correlations[gene_name] ← corr_value
```

```
PRINT "Top gene correlation range:",
```

```
gene_correlations[0][1], "to", gene_correlations[n_top-1][1]
```

5. RETURN

```
RETURN top_genes, correlations
```

END ALGORITHM

Feature Engineering

Algorithm 4: Feature Engineering Pipeline

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```
ALGORITHM: EngineerFeatures(data, top_genes)

INPUT:
- data: Clean DataFrame
- top_genes: List of top 100 selected genes

OUTPUT:
- feature_matrix: Matrix of engineered features
- feature_names: List of feature names
- feature_categories: Dictionary categorizing features

PROCEDURE:
1. DEFINE CLINICAL FEATURES
    clinical_features ← ["UPDRS_BL", "AGE", "GENDER"]

2. DEFINE PD RISK GENES
    pd_genes ← [
        "PD_SNCA",           // α-synuclein
        "PD_LRRK2",          // Leucine-rich repeat kinase 2
        "PD_GBA",            // Glucocerebrosidase
        "PD_PRKN",           // Parkin
        "PD_PINK1",           // PTEN-induced kinase 1
        "PD_PARK7",           // DJ-1
        "PD_VPS35"           // Vacuolar protein sorting 35
    ]

3. DEFINE PATHWAY SCORES
    pathway_features ← [
        "PATHWAY_Inflammation",      // Neuroinflammation
        "PATHWAY_Mitochondrial",     // Mitochondrial dysfunction
        "PATHWAY_Autophagy"          // Autophagy/mitophagy
    ]

4. CREATE INTERACTION FEATURES
    // Gene-gene interaction (mitophagy pathway)
    data["PINK1_X_PARK7"] ← data["PD_PINK1"] × data["PD_PARK7"]

    // Age-gene interaction
    data["AGE_X_PINK1"] ← data["AGE"] × data["PD_PINK1"]
```

```

// Clinical-gene interaction (most important feature!)
data["UPDRS_BL_x_PINK1"] ← data["UPDRS_BL"] × data["PD_PINK1"]

interaction_features ← [
    "PINK1_x_PARK7",
    "AGE_x_PINK1",
    "UPDRS_BL_x_PINK1"
]

5. COMBINE ALL FEATURES
all_features ← clinical_features +
    top_genes +
    pd_genes +
    pathway_features +
    interaction_features

// Filter to existing columns
final_features ← [f FOR f IN all_features IF f IN data.columns]

6. HANDLE MISSING VALUES
FOR EACH feature IN final_features:
    IF ANY(IS_NULL(data[feature])):
        median_value ← MEDIAN(data[feature])
        data[feature].FILL_NA(median_value)

    PRINT "Filled missing values in", feature

7. CREATE FEATURE MATRIX
X ← data[final_features].TO_NUMPY()

8. CATEGORIZE FEATURES
feature_categories ← {
    "clinical": [f FOR f IN clinical_features IF f IN
final_features],
    "top_genes": [f FOR f IN top_genes IF f IN final_features],
    "pd_genes": [f FOR f IN pd_genes IF f IN final_features],
    "pathways": [f FOR f IN pathway_features IF f IN final_features],
    "interactions": [f FOR f IN interaction_features IF f IN
final_features]
}

PRINT "Feature breakdown:"
FOR category, features IN feature_categories:
    PRINT " ", category, ":", LENGTH(features)

PRINT "Total features:", LENGTH(final_features)

9. RETURN

```

```
    RETURN X, final_features, feature_categories  
END ALGORITHM
```

Model Training

Algorithm 5: Build Stacking Ensemble

Plain Text

```
ALGORITHM: BuildStackingEnsemble(hyperparameters)  
  
INPUT:  
    - hyperparameters: Dictionary with optimized hyperparameters  
  
OUTPUT:  
    - ensemble_model: Configured Stacking Regressor  
  
PROCEDURE:  
    1. CONFIGURE XGBoost  
        xgb_model ← XGBRegressor(  
            objective = "reg:pseudohubererror", // Robust to outliers  
            n_estimators = hyperparameters["xgb_n_estimators"],  
            max_depth = hyperparameters["xgb_max_depth"],  
            learning_rate = hyperparameters["xgb_learning_rate"],  
            subsample = hyperparameters["xgb_subsample"],  
            colsample_bytree = hyperparameters["xgb_colsample_bytree"],  
            min_child_weight = hyperparameters["xgb_min_child_weight"],  
            reg_alpha = hyperparameters["xgb_reg_alpha"], // L1  
            regularization  
                reg_lambda = hyperparameters["xgb_reg_lambda"], // L2  
            regularization  
                random_state = 42,  
                n_jobs = 2  
        )  
  
    2. CONFIGURE LightGBM  
        lgbm_model ← LGBMRegressor(  
            objective = "huber", // Robust loss function  
            n_estimators = hyperparameters["lgbm_n_estimators"],  
            max_depth = hyperparameters["lgbm_max_depth"],  
            learning_rate = hyperparameters["lgbm_learning_rate"],  
            subsample = hyperparameters["lgbm_subsample"],  
            colsample_bytree = hyperparameters["lgbm_colsample_bytree"],  
            min_data_in_leaf = hyperparameters["lgbm_min_data_in_leaf"],
```

```

        reg_alpha = hyperparameters["lgbm_reg_alpha"],
        reg_lambda = hyperparameters["lgbm_reg_lambda"],
        random_state = 42,
        n_jobs = 2
    )

3. CONFIGURE CatBoost
    catboost_model ← CatBoostRegressor(
        loss_function = "RMSE",
        iterations = hyperparameters["cat_iterations"],
        depth = hyperparameters["cat_depth"],
        learning_rate = hyperparameters["cat_learning_rate"],
        subsample = hyperparameters["cat_subsample"],
        reg_lambda = hyperparameters["cat_reg_lambda"],
        random_state = 42,
        thread_count = 2,
        verbose = False
    )

4. DEFINE BASE MODELS
    base_models ← [
        ("xgb", xgb_model),
        ("lgbm", lgbm_model),
        ("catboost", catboost_model)
    ]

5. CONFIGURE META-LEARNER (Huber Regressor)
    meta_learner ← HuberRegressor(
        epsilon = hyperparameters["meta_epsilon"], // Robustness
parameter
        alpha = hyperparameters["meta_alpha"], // Regularization
        max_iter = 300
    )

6. BUILD STACKING ENSEMBLE
    ensemble_model ← StackingRegressor(
        estimators = base_models,
        final_estimator = meta_learner,
        cv = 5, // Internal cross-validation for meta-features
        n_jobs = 2
    )

    PRINT "Stacking ensemble configured:"
    PRINT "  Base models: XGBoost, LightGBM, CatBoost"
    PRINT "  Meta-learner: Huber Regressor"

7. RETURN
    RETURN ensemble_model

```

```
END ALGORITHM
```

Hyperparameter Optimization

Algorithm 6: Bayesian Hyperparameter Optimization (Optuna)

Plain Text

```
ALGORITHM: OptimizeHyperparameters(X_train, y_train, n_trials=30, n_folds=7)

INPUT:
    - X_train: Training feature matrix
    - y_train: Training target (transformed)
    - n_trials: Number of Optuna trials (default: 30)
    - n_folds: Number of CV folds (default: 7)

OUTPUT:
    - best_params: Dictionary with best hyperparameters
    - best_cv_score: Best cross-validation R2 score

PROCEDURE:
    1. DEFINE OBJECTIVE FUNCTION
        FUNCTION objective(trial):
            // Sample hyperparameters for XGBoost
            xgb_params ← {
                "n_estimators": trial.SUGGEST_INT("xgb_n_estimators", 100,
250),
                "max_depth": trial.SUGGEST_INT("xgb_max_depth", 3, 7),
                "learning_rate": trial.SUGGEST_FLOAT("xgb_learning_rate",
0.01, 0.1, log=True),
                "subsample": trial.SUGGEST_FLOAT("xgb_subsample", 0.6, 0.9),
                "colsample_bytree":
                    trial.SUGGEST_FLOAT("xgb_colsample_bytree", 0.6, 0.9),
                "min_child_weight":
                    trial.SUGGEST_INT("xgb_min_child_weight", 1, 8),
                "reg_alpha": trial.SUGGEST_FLOAT("xgb_reg_alpha", 0.01, 1.0,
log=True),
                "reg_lambda": trial.SUGGEST_FLOAT("xgb_reg_lambda", 0.1,
5.0, log=True)
            }

            // Sample hyperparameters for LightGBM
            lgbm_params ← {
                "n_estimators": trial.SUGGEST_INT("lgbm_n_estimators", 100,
```

```

250),
    "max_depth": trial.SUGGEST_INT("lgbm_max_depth", 3, 7),
    "learning_rate": trial.SUGGEST_FLOAT("lgbm_learning_rate",
0.01, 0.1, log=True),
    "subsample": trial.SUGGEST_FLOAT("lgbm_subsample", 0.6, 0.9),
    "colsample_bytree":
trial.SUGGEST_FLOAT("lgbm_colsample_bytree", 0.6, 0.9),
    "min_data_in_leaf":
trial.SUGGEST_INT("lgbm_min_data_in_leaf", 5, 25),
    "reg_alpha": trial.SUGGEST_FLOAT("lgbm_reg_alpha", 0.01,
1.0, log=True),
    "reg_lambda": trial.SUGGEST_FLOAT("lgbm_reg_lambda", 0.1,
5.0, log=True)
}

// Sample hyperparameters for CatBoost
catboost_params ← {
    "iterations": trial.SUGGEST_INT("cat_iterations", 100, 250),
    "depth": trial.SUGGEST_INT("cat_depth", 3, 7),
    "learning_rate": trial.SUGGEST_FLOAT("cat_learning_rate",
0.01, 0.1, log=True),
    "subsample": trial.SUGGEST_FLOAT("cat_subsample", 0.6, 0.9),
    "reg_lambda": trial.SUGGEST_FLOAT("cat_reg_lambda", 0.1,
5.0, log=True)
}

// Sample meta-learner hyperparameters
meta_alpha ← trial.SUGGEST_FLOAT("meta_alpha", 0.01, 1.0,
log=True)
meta_epsilon ← trial.SUGGEST_FLOAT("meta_epsilon", 1.0, 2.0)

// Build ensemble with sampled hyperparameters
ensemble ← BUILD_STACKING_ENSEMBLE({
    **xgb_params, **lgbm_params, **catboost_params,
    "meta_alpha": meta_alpha, "meta_epsilon": meta_epsilon
})

// Perform k-fold cross-validation
cv ← KFold(n_splits=n_folds, shuffle=True, random_state=42)
cv_scores ← []

FOR train_idx, val_idx IN cv.SPLIT(X_train):
    X_train_fold ← X_train[train_idx]
    y_train_fold ← y_train[train_idx]
    X_val_fold ← X_train[val_idx]
    y_val_fold ← y_train[val_idx]

    // Scale features

```

```

        scaler ← StandardScaler()
        X_train_scaled ← scaler.FIT_TRANSFORM(X_train_fold)
        X_val_scaled ← scaler.TRANSFORM(X_val_fold)

        // Train and evaluate
        ensemble.FIT(X_train_scaled, y_train_fold)
        y_val_pred ← ensemble.PREDICT(X_val_scaled)

        // Inverse transform predictions
        y_val_pred_orig ←
target_transformer.INVERSE_TRANSFORM(y_val_pred)
        y_val_orig ← target_transformer.INVERSE_TRANSFORM(y_val_fold)

        // Calculate R2
        r2 ← R2_SCORE(y_val_orig, y_val_pred_orig)
        APPEND r2 TO cv_scores

        // Return mean CV score
        RETURN MEAN(cv_scores)

    END FUNCTION

2. CREATE OPTUNA STUDY
study ← optuna.CREATE_STUDY(
    direction = "maximize", // Maximize R2
    study_name = "lightweight_optimization"
)

3. RUN OPTIMIZATION
PRINT "Starting Bayesian optimization with", n_trials, "trials..."

study.OPTIMIZE(
    objective,
    n_trials = n_trials,
    show_progress_bar = True
)

PRINT "Optimization complete!"

4. EXTRACT BEST PARAMETERS
best_params ← study.best_params
best_cv_score ← study.best_value

PRINT "Best CV R2:", best_cv_score
PRINT "Best hyperparameters:"
FOR param, value IN best_params:
    PRINT " ", param, ":", value

```

```

5. RETURN
    RETURN best_params, best_cv_score

END ALGORITHM

```

Model Validation

Algorithm 7: Model Validation on Independent Test Set

Plain Text

```

ALGORITHM: ValidateModel(model, X_test, y_test, scaler, target_transformer)

INPUT:
- model: Trained ensemble model
- X_test: Test feature matrix (unscaled)
- y_test: Test target (original scale)
- scaler: Fitted StandardScaler
- target_transformer: Fitted PowerTransformer

OUTPUT:
- metrics: Dictionary with R2, MAE, RMSE, Pearson r
- predictions: Array of predicted values
- residuals: Array of residuals (actual - predicted)

PROCEDURE:
1. SCALE TEST FEATURES
    X_test_scaled ← scaler.TRANSFORM(X_test)

2. MAKE PREDICTIONS (Transformed Space)
    y_test_pred_trans ← model.PREDICT(X_test_scaled)

3. INVERSE TRANSFORM TO ORIGINAL SCALE
    y_test_pred ← target_transformer.INVERSE_TRANSFORM(
        y_test_pred_trans.RESHAPE(-1, 1)
    ).FLATTEN()

4. CALCULATE REGRESSION METRICS
    // R2 (coefficient of determination)
    r2 ← R2_SCORE(y_test, y_test_pred)

    // MAE (mean absolute error)
    mae ← MEAN_ABSOLUTE_ERROR(y_test, y_test_pred)

    // RMSE (root mean squared error)

```

```

mse ← MEAN_SQUARED_ERROR(y_test, y_test_pred)
rmse ← SQRT(mse)

// Pearson correlation coefficient
pearson_r ← PEARSON_CORRELATION(y_test, y_test_pred)

// Spearman rank correlation
spearman_rho ← SPEARMAN_CORRELATION(y_test, y_test_pred)

5. CALCULATE RESIDUALS
residuals ← y_test - y_test_pred

// Residual statistics
mean_residual ← MEAN(residuals)
std_residual ← STD(residuals)

PRINT "Mean residual:", mean_residual
PRINT "Std residual:", std_residual

6. PRINT RESULTS
PRINT "=" * 80
PRINT "INDEPENDENT TEST SET VALIDATION RESULTS"
PRINT "=" * 80
PRINT "R2 Score:", r2
PRINT "MAE:", mae, "UPDRS points"
PRINT "RMSE:", rmse, "UPDRS points"
PRINT "Pearson r:", pearson_r
PRINT "Spearman ρ:", spearman_rho
PRINT "=" * 80

7. PACKAGE METRICS
metrics ← {
    "r2": r2,
    "mae": mae,
    "rmse": rmse,
    "pearson_r": pearson_r,
    "spearman_rho": spearman_rho,
    "mean_residual": mean_residual,
    "std_residual": std_residual,
    "n_samples": LENGTH(y_test)
}

8. RETURN
RETURN metrics, y_test_pred, residuals

END ALGORITHM

```

SHAP Analysis

Algorithm 8: SHAP Feature Importance Analysis

Plain Text

```
ALGORITHM: ComputeSHAPValues(model, X_test, feature_names)

INPUT:
- model: Trained ensemble model
- X_test: Test feature matrix (scaled)
- feature_names: List of feature names

OUTPUT:
- shap_values: SHAP importance values for each feature
- feature_importance: Sorted list of (feature, importance) tuples

PROCEDURE:
1. CREATE SHAP EXPLAINER
   // Use TreeExplainer for gradient boosting models
   explainer ← shap.TreeExplainer(model)

   PRINT "SHAP explainer created"

2. COMPUTE SHAP VALUES
   PRINT "Computing SHAP values (this may take a few minutes)..."

   shap_values ← explainer.SHAP_VALUES(X_test)

   PRINT "SHAP values computed for", LENGTH(X_test), "samples"

3. CALCULATE MEAN ABSOLUTE SHAP VALUES
   mean_abs_shap ← []

   FOR i FROM 0 TO LENGTH(feature_names)-1:
      feature_name ← feature_names[i]
      shap_column ← shap_values[:, i]

      mean_abs_value ← MEAN(ABSOLUTE_VALUE(shap_column))

      APPEND (feature_name, mean_abs_value) TO mean_abs_shap

4. SORT BY IMPORTANCE
   SORT mean_abs_shap BY mean_abs_value DESCENDING

5. PRINT TOP 20 FEATURES
   PRINT "=" * 80
```

```

PRINT "TOP 20 FEATURES BY SHAP IMPORTANCE"
PRINT "=" * 80

FOR i FROM 0 TO 19:
    feature, importance ← mean_abs_shap[i]
    PRINT i+1, ".", feature, ":", importance

PRINT "=" * 80

6. CATEGORIZE FEATURES
// Separate by feature type
clinical_shap ← []
gene_shap ← []
pd_gene_shap ← []
pathway_shap ← []
interaction_shap ← []

FOR feature, importance IN mean_abs_shap:
    IF feature IN ["UPDRS_BL", "AGE", "GENDER"]:
        APPEND (feature, importance) TO clinical_shap
    ELSE IF feature STARTS_WITH "ENSG":
        APPEND (feature, importance) TO gene_shap
    ELSE IF feature STARTS_WITH "PD__":
        APPEND (feature, importance) TO pd_gene_shap
    ELSE IF feature STARTS_WITH "PATHWAY__":
        APPEND (feature, importance) TO pathway_shap
    ELSE IF feature CONTAINS "_x__":
        APPEND (feature, importance) TO interaction_shap

7. PRINT CATEGORY SUMMARIES
PRINT "SHAP IMPORTANCE BY CATEGORY:"
PRINT " Clinical features:", SUM([imp FOR _, imp IN clinical_shap])
PRINT " Top genes:", SUM([imp FOR _, imp IN gene_shap[:20]])
PRINT " PD risk genes:", SUM([imp FOR _, imp IN pd_gene_shap])
PRINT " Pathways:", SUM([imp FOR _, imp IN pathway_shap])
PRINT " Interactions:", SUM([imp FOR _, imp IN interaction_shap])

8. RETURN
RETURN shap_values, mean_abs_shap

END ALGORITHM

```

Clinical Prediction

Algorithm 9: Predict for New Patient

Plain Text

```
ALGORITHM: PredictNewPatient(patient_data, model_package)

INPUT:
- patient_data: Dictionary with baseline clinical data
{
    "PATNO": "PATIENT_001",
    "UPDRS_BL": 20.0,
    "AGE": 68.0,
    "GENDER": 1.0 // 0=Female, 1=Male
}
- model_package: Loaded model artifacts

OUTPUT:
- prediction_report: Dictionary with predictions and interpretation

PROCEDURE:
1. EXTRACT MODEL ARTIFACTS
    model ← model_package["ensemble_model"]
    scaler ← model_package["scaler"]
    target_transformer ← model_package["target_transformer"]
    feature_names ← model_package["feature_names"]
    n_features ← model_package["n_features"]
    test_mae ← model_package["test_results"]["mae"]

2. CREATE PATIENT DATAFRAME
    patient_df ← DataFrame([patient_data])

    PRINT "Patient ID:", patient_data["PATNO"]
    PRINT "Baseline UPDRS:", patient_data["UPDRS_BL"]
    PRINT "Age:", patient_data["AGE"]
    PRINT "Gender:", "Male" IF patient_data["GENDER"]==1 ELSE "Female"

3. IMPUTE MISSING FEATURES
    // For clinical prediction, we only have baseline clinical data
    // All gene expression and pathway features are imputed with 0

    FOR feature IN feature_names:
        IF feature NOT IN patient_df.columns:
            patient_df[feature] ← 0.0

        // Ensure correct column order
        patient_df ← patient_df[feature_names]

    PRINT "Warning: Missing", n_features - 4, "features (imputed with
zeros)"
```

```

4. SCALE FEATURES
    X_patient ← patient_df.TO_NUMPY()
    X_patient_scaled ← scaler.TRANSFORM(X_patient)

5. MAKE PREDICTION (Transformed Space)
    y_pred_trans ← model.PREDICT(X_patient_scaled)

6. INVERSE TRANSFORM TO UPDRS SCALE
    y_pred_updrs ← target_transformer.INVERSE_TRANSFORM(
        y_pred_trans.RESHAPE(-1, 1)
    ).FLATTEN()[0]

    PRINT "Predicted UPDRS at 12 months:", y_pred_updrs

7. CALCULATE PREDICTED CHANGE
    baseline_updrs ← patient_data["UPDRS_BL"]
    predicted_change ← y_pred_updrs - baseline_updrs

    PRINT "Predicted change:", predicted_change, "points"

8. CALCULATE CONFIDENCE INTERVAL
    // Use MAE as uncertainty estimate
    lower_bound ← y_pred_updrs - test_mae
    upper_bound ← y_pred_updrs + test_mae

    PRINT "95% Confidence Interval: [", lower_bound, ", ", upper_bound,
"]"

9. CATEGORIZE PROGRESSION RISK
    IF predicted_change < 0:
        risk_category ← "Improvement"
        confidence ← "Low" // Unusual, low confidence
    ELSE IF predicted_change < 3:
        risk_category ← "Stable"
        confidence ← "High"
    ELSE IF predicted_change < 5:
        risk_category ← "Mild Progression"
        confidence ← "High"
    ELSE IF predicted_change < 10:
        risk_category ← "Moderate Progression"
        confidence ← "High"
    ELSE:
        risk_category ← "Rapid Progression"
        confidence ← "High"

    PRINT "Progression Risk:", risk_category
    PRINT "Confidence:", confidence

```

```

10. GENERATE CLINICAL INTERPRETATION
    IF risk_category == "Stable":
        interpretation ← "Patient likely to remain stable over 12 months
(" +
                    predicted_change + " points change)."

    ELSE IF risk_category == "Mild Progression":
        interpretation ← "Mild progression expected (" +
predicted_change +
                    " points). Standard monitoring recommended."

    ELSE IF risk_category == "Moderate Progression":
        interpretation ← "Moderate progression expected (" +
predicted_change +
                    " points). Consider treatment adjustment."

    ELSE IF risk_category == "Rapid Progression":
        interpretation ← "Rapid progression expected (" +
predicted_change +
                    " points). Urgent clinical review recommended."

    ELSE: // Improvement
        interpretation ← "Patient shows predicted improvement (" +
predicted_change + " points). Monitor for
accuracy."


    PRINT "Clinical Interpretation:", interpretation

11. PACKAGE PREDICTION REPORT
    prediction_report ← {
        "patient_id": patient_data["PATNO"],
        "baseline_updrs": baseline_updrs,
        "predicted_updrs_12m": y_pred_updrs,
        "predicted_change": predicted_change,
        "lower_bound_12m": lower_bound,
        "upper_bound_12m": upper_bound,
        "progression_risk": risk_category,
        "confidence_level": confidence,
        "clinical_interpretation": interpretation
    }

12. RETURN
    RETURN prediction_report

END ALGORITHM

```

Summary Statistics

Key Performance Metrics

Plain Text

MODEL PERFORMANCE SUMMARY

Training Set (n=312, 7-Fold CV):

R² = 0.513 ± 0.052

MAE = 6.15 ± 0.25 UPDRS points

RMSE = 7.82 ± 0.31 UPDRS points

Independent Test Set (n=78):

R² = 0.551 (55.1% variance explained)

MAE = 6.01 UPDRS points

RMSE = 7.45 UPDRS points

Pearson r = 0.74

Top 3 Features (SHAP):

1. UPDRS_BL × PINK1 = 0.283

2. UPDRS_BL = 0.258

3. ENSG00000243053 = 0.025

Computational Complexity

Plain Text

COMPUTATIONAL COMPLEXITY

Training Phase:

Time Complexity: O(n × m × t × k)

n = number of samples (312)

m = number of features (116)

t = number of trees per model (~200)

k = number of CV folds (7)

Space Complexity: O(n × m + t × m)

Actual Training Time: ~1 hour (30 Optuna trials)

Prediction Phase:

Time Complexity: $O(m \times t)$

Space Complexity: $O(m)$

Actual Prediction Time: <1 second per patient

Model Size: 582 KB (compressed)

End of Pseudo Code Documentation

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For implementation details, see the actual Python code in `lightweight_optimization.py`