

# Pseudo Code: Parkinson's Disease Motor Progression Prediction Model

## Stacking Regressor with Explainable AI (SHAP)

Version: 1.0

Date: November 2025

Performance:  $R^2=0.551$ , MAE=6.01 (Independent Clinical Test Set)

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## Overview

### System Architecture

Plain Text

#### TRAINING PIPELINE

[1] Load Data → [2] Preprocess → [3] Feature Engineering →  
[4] Split Data → [5] Optimize Hyperparameters →  
[6] Train Final Model → [7] Validate → [8] SHAP Analysis

## PREDICTION PIPELINE

[1] Load Patient Data → [2] Preprocess → [3] Scale Features →  
[4] Predict → [5] Inverse Transform → [6] Generate Report

## Model Components

- **Base Models:** XGBoost, LightGBM, CatBoost
- **Meta-Learner:** Huber Regressor
- **Explainability:** SHAP (SHapley Additive exPlanations)
- **Validation:** 7-Fold Cross-Validation + Independent Holdout

## Main Training Pipeline

### Algorithm 1: Complete Training Pipeline

Plain Text

ALGORITHM: TrainParkinsonsProgressionModel()

INPUT:

- clinical\_data: DataFrame with baseline clinical features
- rnaseq\_data: DataFrame with gene expression data (RNA-seq)
- random\_seed: Integer for reproducibility (default: 42)

OUTPUT:

- trained\_model: Optimized Stacking Regressor
- performance\_metrics: Dictionary with R<sup>2</sup>, MAE, RMSE
- shap\_values: SHAP importance values for features
- model\_artifacts: Scalers, transformers, feature names

PROCEDURE:

1. INITIALIZE
  - SET random\_seed = 42
  - SET n\_top\_genes = 100
  - SET n\_optuna\_trials = 30
  - SET n\_cv\_folds = 7
  - SET test\_size = 0.2

## 2. LOAD AND MERGE DATA

```
clinical_df ← LOAD_CSV("final_dataset.csv")
rnaseq_df ← LOAD_CSV("rnaseq_baseline_filtered.csv")
merged_df ← MERGE(clinical_df, rnaseq_df, on="PATNO")
```

```
PRINT "Loaded", LENGTH(merged_df), "patients"
```

## 3. REMOVE OUTLIERS

```
Q1 ← QUANTILE(merged_df["UPDRS_V04"], 0.25)
Q3 ← QUANTILE(merged_df["UPDRS_V04"], 0.75)
IQR ← Q3 - Q1
```

```
outlier_mask ← (UPDRS_V04 < Q1 - 1.5×IQR) OR (UPDRS_V04 > Q3 +
1.5×IQR)
clean_df ← merged_df[NOT outlier_mask]
```

```
PRINT "Removed", SUM(outlier_mask), "outliers"
```

## 4. SELECT TOP GENES

```
gene_correlations ← []
```

```
FOR EACH gene IN gene_columns:
```

```
  corr ← CORRELATION(clean_df[gene], clean_df["DELTA_UPDRS"])
```

```
  IF NOT IS_NAN(corr):
```

```
    APPEND (gene, ABS(corr)) TO gene_correlations
```

```
SORT gene_correlations BY correlation DESC
```

```
top_genes ← FIRST 100 genes FROM gene_correlations
```

```
PRINT "Selected top 100 genes with correlation range:",
      MAX(corr), "to", MIN(corr)
```

## 5. ENGINEER FEATURES

```
// Clinical features
```

```
clinical_features ← ["UPDRS_BL", "AGE", "GENDER"]
```

```
// PD risk genes
```

```
pd_genes ← ["PD_SNCA", "PD_LRRK2", "PD_GBA", "PD_PRKN",
            "PD_PINK1", "PD_PARK7", "PD_VPS35"]
```

```
// Pathway scores
```

```
pathway_features ← ["PATHWAY_Inflammation",
                    "PATHWAY_Mitochondrial",
                    "PATHWAY_Autophagy"]
```

```
// Create interaction features
```

```
clean_df["PINK1_x_PARK7"] ← clean_df["PD_PINK1"] ×
```

```

clean_df["PD_PARK7"]
  clean_df["AGE_x_PINK1"] ← clean_df["AGE"] × clean_df["PD_PINK1"]
  clean_df["UPDRS_BL_x_PINK1"] ← clean_df["UPDRS_BL"] ×
clean_df["PD_PINK1"]

  interaction_features ← ["PINK1_x_PARK7", "AGE_x_PINK1",
"UPDRS_BL_x_PINK1"]

  // Combine all features
  all_features ← clinical_features + top_genes + pd_genes +
    pathway_features + interaction_features

  PRINT "Total features:", LENGTH(all_features)

6. PREPARE DATA
  X ← clean_df[all_features]
  y_v04 ← clean_df["UPDRS_V04"] // Target: 12-month UPDRS
  y_clf ← (clean_df["DELTA_UPDRS"] >= 5) // For stratification

  // Transform target variable
  target_transformer ← PowerTransformer(method="yeo-johnson")
  y_transformed ← target_transformer.FIT_TRANSFORM(y_v04)

7. SPLIT DATA (Stratified)
  (X_trainval, X_test,
  y_trainval_trans, y_test_trans,
  y_trainval_orig, y_test_orig) ← TRAIN_TEST_SPLIT(
    X, y_transformed, y_v04,
    test_size=0.2,
    stratify=y_clf,
    random_state=random_seed
  )

  PRINT "Train+Val:", LENGTH(X_trainval), "Test:", LENGTH(X_test)

8. OPTIMIZE HYPERPARAMETERS
  best_params ← OPTIMIZE_HYPERPARAMETERS(
    X_trainval, y_trainval_trans,
    n_trials=30,
    n_folds=7
  )

  PRINT "Best CV R²:", best_params["cv_r2"]

9. TRAIN FINAL MODEL
  // Scale features
  scaler ← StandardScaler()
  X_trainval_scaled ← scaler.FIT_TRANSFORM(X_trainval)

```

```

X_test_scaled ← scaler.TRANSFORM(X_test)

// Build ensemble with best hyperparameters
final_model ← BUILD_STACKING_ENSEMBLE(best_params)

// Train on full training set
final_model.FIT(X_trainval_scaled, y_trainval_trans)

PRINT "Final model trained"

10. VALIDATE ON TEST SET
// Predict on test set
y_test_pred_trans ← final_model.PREDICT(X_test_scaled)

// Inverse transform to original scale
y_test_pred ← target_transformer.INVERSE_TRANSFORM(y_test_pred_trans)

// Calculate metrics
test_r2 ← R2_SCORE(y_test_orig, y_test_pred)
test_mae ← MEAN_ABSOLUTE_ERROR(y_test_orig, y_test_pred)
test_rmse ← SQRT(MEAN_SQUARED_ERROR(y_test_orig, y_test_pred))

PRINT "Test R²:", test_r2
PRINT "Test MAE:", test_mae
PRINT "Test RMSE:", test_rmse

11. COMPUTE SHAP VALUES
shap_values ← COMPUTE_SHAP_VALUES(
    final_model,
    X_test_scaled,
    feature_names=all_features
)

PRINT "SHAP analysis complete"

12. SAVE MODEL ARTIFACTS
model_package ← {
    "ensemble_model": final_model,
    "scaler": scaler,
    "target_transformer": target_transformer,
    "feature_names": all_features,
    "n_features": LENGTH(all_features),
    "cv_results": best_params,
    "test_results": {
        "r2": test_r2,
        "mae": test_mae,
        "rmse": test_rmse
    },
},

```

```
        "shap_values": shap_values
    }

    SAVE_PICKLE(model_package, "lightweight_optimized_model.pkl")

    PRINT "Model saved successfully"

13. RETURN
    RETURN model_package

END ALGORITHM
```

## Data Preprocessing

### Algorithm 2: Data Loading and Cleaning

Plain Text

```
ALGORITHM: LoadAndCleanData()

INPUT:
  - clinical_file_path: Path to clinical data CSV
  - rnaseq_file_path: Path to RNA-seq data CSV

OUTPUT:
  - clean_data: Preprocessed DataFrame
  - n_outliers_removed: Number of outliers removed

PROCEDURE:
  1. LOAD DATA
    clinical_df ← READ_CSV(clinical_file_path)
    rnaseq_df ← READ_CSV(rnaseq_file_path)

    PRINT "Clinical data:", SHAPE(clinical_df)
    PRINT "RNA-seq data:", SHAPE(rnaseq_df)

  2. CONVERT PATIENT IDS TO STRING
    clinical_df["PATNO"] ← TO_STRING(clinical_df["PATNO"])
    rnaseq_df["PATNO"] ← TO_STRING(rnaseq_df["PATNO"])

  3. EXTRACT GENE COLUMNS
    gene_columns ← [col FOR col IN rnaseq_df.columns
                     IF col STARTS_WITH "ENSG"]

    rnaseq_genes ← rnaseq_df[["PATNO"] + gene_columns]
```

```

    PRINT "Gene columns:", LENGTH(gene_columns)

4. MERGE DATASETS
    merged_df ← INNER_JOIN(clinical_df, rnaseq_genes, on="PATNO")

    PRINT "Merged patients:", LENGTH(merged_df)

5. HANDLE MISSING VALUES IN TARGET
    IF ANY(IS_NULL(merged_df["UPDRS_V04"])):
        median_updrs ← MEDIAN(merged_df["UPDRS_V04"])
        merged_df["UPDRS_V04"].FILL_NA(median_updrs)

    PRINT "Filled", SUM(IS_NULL), "missing UPDRS values"

6. DETECT AND REMOVE OUTLIERS (IQR Method)
    Q1 ← QUANTILE(merged_df["UPDRS_V04"], 0.25)
    Q3 ← QUANTILE(merged_df["UPDRS_V04"], 0.75)
    IQR ← Q3 - Q1

    lower_bound ← Q1 - 1.5 × IQR
    upper_bound ← Q3 + 1.5 × IQR

    outlier_mask ← (merged_df["UPDRS_V04"] < lower_bound) OR
                  (merged_df["UPDRS_V04"] > upper_bound)

    n_outliers ← SUM(outlier_mask)
    clean_df ← merged_df[NOT outlier_mask]

    PRINT "Outliers removed:", n_outliers
    PRINT "Clean data size:", LENGTH(clean_df)

7. RETURN
    RETURN clean_df, n_outliers

```

END ALGORITHM

## Algorithm 3: Gene Selection by Correlation

Plain Text

ALGORITHM: SelectTopGenes(data, n\_top=100)

INPUT:

- data: DataFrame with gene expression and DELTA\_UPDRS
- n\_top: Number of top genes to select (default: 100)

OUTPUT:

- top\_genes: List of top gene names
- correlations: Dictionary mapping genes to correlation values

PROCEDURE:

1. IDENTIFY GENE COLUMNS

```
gene_columns ← [col FOR col IN data.columns  
                 IF col STARTS_WITH "ENSG"]
```

```
PRINT "Total genes available:", LENGTH(gene_columns)
```

2. COMPUTE CORRELATIONS

```
gene_correlations ← []
```

```
FOR EACH gene IN gene_columns:
```

```
    IF gene IN data.columns:
```

```
        // Compute Pearson correlation with progression
```

```
        corr_matrix ← CORRELATION(data[[gene, "DELTA_UPDRS"]])
```

```
        corr_value ← corr_matrix[0, 1]
```

```
        IF NOT IS_NAN(corr_value):
```

```
            abs_corr ← ABSOLUTE_VALUE(corr_value)
```

```
            APPEND (gene, abs_corr, corr_value) TO gene_correlations
```

```
PRINT "Valid correlations computed:", LENGTH(gene_correlations)
```

3. SORT BY ABSOLUTE CORRELATION

```
SORT gene_correlations BY abs_corr DESCENDING
```

4. SELECT TOP N GENES

```
top_genes ← []
```

```
correlations ← {}
```

```
FOR i FROM 0 TO n_top-1:
```

```
    gene_name ← gene_correlations[i][0]
```

```
    corr_value ← gene_correlations[i][2]
```

```
    APPEND gene_name TO top_genes
```

```
    correlations[gene_name] ← corr_value
```

```
PRINT "Top gene correlation range:",
```

```
    gene_correlations[0][1], "to", gene_correlations[n_top-1][1]
```

5. RETURN

```
RETURN top_genes, correlations
```

END ALGORITHM



# Feature Engineering

## Algorithm 4: Feature Engineering Pipeline

Plain Text

ALGORITHM: EngineerFeatures(data, top\_genes)

INPUT:

- data: Clean DataFrame
- top\_genes: List of top 100 selected genes

OUTPUT:

- feature\_matrix: Matrix of engineered features
- feature\_names: List of feature names
- feature\_categories: Dictionary categorizing features

PROCEDURE:

1. DEFINE CLINICAL FEATURES

```
clinical_features ← ["UPDRS_BL", "AGE", "GENDER"]
```

2. DEFINE PD RISK GENES

```
pd_genes ← [  
    "PD_SNCA",      // α-synuclein  
    "PD_LRRK2",     // Leucine-rich repeat kinase 2  
    "PD_GBA",       // Glucocerebrosidase  
    "PD_PRKN",      // Parkin  
    "PD_PINK1",     // PTEN-induced kinase 1  
    "PD_PARK7",     // DJ-1  
    "PD_VPS35"      // Vacuolar protein sorting 35  
]
```

3. DEFINE PATHWAY SCORES

```
pathway_features ← [  
    "PATHWAY_Inflammation", // Neuroinflammation  
    "PATHWAY_Mitochondrial", // Mitochondrial dysfunction  
    "PATHWAY_Autophagy"      // Autophagy/mitophagy  
]
```

4. CREATE INTERACTION FEATURES

```
// Gene-gene interaction (mitophagy pathway)  
data["PINK1_x_PARK7"] ← data["PD_PINK1"] × data["PD_PARK7"]  
  
// Age-gene interaction  
data["AGE_x_PINK1"] ← data["AGE"] × data["PD_PINK1"]
```

```
// Clinical-gene interaction (most important feature!)
data["UPDRS_BL_x_PINK1"] ← data["UPDRS_BL"] × data["PD_PINK1"]

interaction_features ← [
  "PINK1_x_PARK7",
  "AGE_x_PINK1",
  "UPDRS_BL_x_PINK1"
]
```

#### 5. COMBINE ALL FEATURES

```
all_features ← clinical_features +
  top_genes +
  pd_genes +
  pathway_features +
  interaction_features
```

```
// Filter to existing columns
final_features ← [f FOR f IN all_features IF f IN data.columns]
```

#### 6. HANDLE MISSING VALUES

```
FOR EACH feature IN final_features:
  IF ANY(IS_NULL(data[feature])):
    median_value ← MEDIAN(data[feature])
    data[feature].FILL_NA(median_value)

    PRINT "Filled missing values in", feature
```

#### 7. CREATE FEATURE MATRIX

```
X ← data[final_features].TO_NUMPY()
```

#### 8. CATEGORIZE FEATURES

```
feature_categories ← {
  "clinical": [f FOR f IN clinical_features IF f IN
final_features],
  "top_genes": [f FOR f IN top_genes IF f IN final_features],
  "pd_genes": [f FOR f IN pd_genes IF f IN final_features],
  "pathways": [f FOR f IN pathway_features IF f IN final_features],
  "interactions": [f FOR f IN interaction_features IF f IN
final_features]
}
```

```
PRINT "Feature breakdown:"
FOR category, features IN feature_categories:
  PRINT "  ", category, ":", LENGTH(features)
```

```
PRINT "Total features:", LENGTH(final_features)
```

#### 9. RETURN

```
RETURN X, final_features, feature_categories
```

```
END ALGORITHM
```

## Model Training

### Algorithm 5: Build Stacking Ensemble

Plain Text

```
ALGORITHM: BuildStackingEnsemble(hyperparameters)
```

INPUT:

- hyperparameters: Dictionary with optimized hyperparameters

OUTPUT:

- ensemble\_model: Configured Stacking Regressor

PROCEDURE:

1. CONFIGURE XGBoost

```
xgb_model ← XGBRegressor(  
    objective = "reg:pseudohubererror", // Robust to outliers  
    n_estimators = hyperparameters["xgb_n_estimators"],  
    max_depth = hyperparameters["xgb_max_depth"],  
    learning_rate = hyperparameters["xgb_learning_rate"],  
    subsample = hyperparameters["xgb_subsample"],  
    colsample_bytree = hyperparameters["xgb_colsample_bytree"],  
    min_child_weight = hyperparameters["xgb_min_child_weight"],  
    reg_alpha = hyperparameters["xgb_reg_alpha"], // L1
```

regularization

```
    reg_lambda = hyperparameters["xgb_reg_lambda"], // L2
```

regularization

```
    random_state = 42,  
    n_jobs = 2
```

```
)
```

2. CONFIGURE LightGBM

```
lgbm_model ← LGBMRegressor(  
    objective = "huber", // Robust loss function  
    n_estimators = hyperparameters["lgbm_n_estimators"],  
    max_depth = hyperparameters["lgbm_max_depth"],  
    learning_rate = hyperparameters["lgbm_learning_rate"],  
    subsample = hyperparameters["lgbm_subsample"],  
    colsample_bytree = hyperparameters["lgbm_colsample_bytree"],  
    min_data_in_leaf = hyperparameters["lgbm_min_data_in_leaf"],
```

```

        reg_alpha = hyperparameters["lgbm_reg_alpha"],
        reg_lambda = hyperparameters["lgbm_reg_lambda"],
        random_state = 42,
        n_jobs = 2
    )

```

### 3. CONFIGURE CatBoost

```

catboost_model ← CatBoostRegressor(
    loss_function = "RMSE",
    iterations = hyperparameters["cat_iterations"],
    depth = hyperparameters["cat_depth"],
    learning_rate = hyperparameters["cat_learning_rate"],
    subsample = hyperparameters["cat_subsample"],
    reg_lambda = hyperparameters["cat_reg_lambda"],
    random_state = 42,
    thread_count = 2,
    verbose = False
)

```

### 4. DEFINE BASE MODELS

```

base_models ← [
    ("xgb", xgb_model),
    ("lgbm", lgbm_model),
    ("catboost", catboost_model)
]

```

### 5. CONFIGURE META-LEARNER (Huber Regressor)

```

meta_learner ← HuberRegressor(
    epsilon = hyperparameters["meta_epsilon"], // Robustness
parameter    alpha = hyperparameters["meta_alpha"], // Regularization
    max_iter = 300
)

```

### 6. BUILD STACKING ENSEMBLE

```

ensemble_model ← StackingRegressor(
    estimators = base_models,
    final_estimator = meta_learner,
    cv = 5, // Internal cross-validation for meta-features
    n_jobs = 2
)

```

```

PRINT "Stacking ensemble configured:"
PRINT "  Base models: XGBoost, LightGBM, CatBoost"
PRINT "  Meta-learner: Huber Regressor"

```

### 7. RETURN

```

RETURN ensemble_model

```

END ALGORITHM

# Hyperparameter Optimization

## Algorithm 6: Bayesian Hyperparameter Optimization (Optuna)

Plain Text

ALGORITHM: OptimizeHyperparameters( $X_{\text{train}}$ ,  $y_{\text{train}}$ ,  $n_{\text{trials}}=30$ ,  $n_{\text{folds}}=7$ )

INPUT:

- $X_{\text{train}}$ : Training feature matrix
- $y_{\text{train}}$ : Training target (transformed)
- $n_{\text{trials}}$ : Number of Optuna trials (default: 30)
- $n_{\text{folds}}$ : Number of CV folds (default: 7)

OUTPUT:

- $\text{best\_params}$ : Dictionary with best hyperparameters
- $\text{best\_cv\_score}$ : Best cross-validation  $R^2$  score

PROCEDURE:

1. DEFINE OBJECTIVE FUNCTION

    FUNCTION objective(trial):

        // Sample hyperparameters for XGBoost

        xgb\_params ← {

            "n\_estimators": trial.SUGGEST\_INT("xgb\_n\_estimators", 100,

250),

            "max\_depth": trial.SUGGEST\_INT("xgb\_max\_depth", 3, 7),

            "learning\_rate": trial.SUGGEST\_FLOAT("xgb\_learning\_rate",

0.01, 0.1, log=True),

            "subsample": trial.SUGGEST\_FLOAT("xgb\_subsample", 0.6, 0.9),

            "colsample\_bytree":

trial.SUGGEST\_FLOAT("xgb\_colsample\_bytree", 0.6, 0.9),

            "min\_child\_weight":

trial.SUGGEST\_INT("xgb\_min\_child\_weight", 1, 8),

            "reg\_alpha": trial.SUGGEST\_FLOAT("xgb\_reg\_alpha", 0.01, 1.0,

log=True),

            "reg\_lambda": trial.SUGGEST\_FLOAT("xgb\_reg\_lambda", 0.1,

5.0, log=True)

        }

        // Sample hyperparameters for LightGBM

        lgbm\_params ← {

            "n\_estimators": trial.SUGGEST\_INT("lgbm\_n\_estimators", 100,

```

250),
    "max_depth": trial.SUGGEST_INT("lgbm_max_depth", 3, 7),
    "learning_rate": trial.SUGGEST_FLOAT("lgbm_learning_rate",
0.01, 0.1, log=True),
    "subsample": trial.SUGGEST_FLOAT("lgbm_subsample", 0.6, 0.9),
    "colsample_bytree":
trial.SUGGEST_FLOAT("lgbm_colsample_bytree", 0.6, 0.9),
    "min_data_in_leaf":
trial.SUGGEST_INT("lgbm_min_data_in_leaf", 5, 25),
    "reg_alpha": trial.SUGGEST_FLOAT("lgbm_reg_alpha", 0.01,
1.0, log=True),
    "reg_lambda": trial.SUGGEST_FLOAT("lgbm_reg_lambda", 0.1,
5.0, log=True)
}

// Sample hyperparameters for CatBoost
catboost_params ← {
    "iterations": trial.SUGGEST_INT("cat_iterations", 100, 250),
    "depth": trial.SUGGEST_INT("cat_depth", 3, 7),
    "learning_rate": trial.SUGGEST_FLOAT("cat_learning_rate",
0.01, 0.1, log=True),
    "subsample": trial.SUGGEST_FLOAT("cat_subsample", 0.6, 0.9),
    "reg_lambda": trial.SUGGEST_FLOAT("cat_reg_lambda", 0.1,
5.0, log=True)
}

// Sample meta-learner hyperparameters
meta_alpha ← trial.SUGGEST_FLOAT("meta_alpha", 0.01, 1.0,
log=True)
meta_epsilon ← trial.SUGGEST_FLOAT("meta_epsilon", 1.0, 2.0)

// Build ensemble with sampled hyperparameters
ensemble ← BUILD_STACKING_ENSEMBLE({
    **xgb_params, **lgbm_params, **catboost_params,
    "meta_alpha": meta_alpha, "meta_epsilon": meta_epsilon
})

// Perform k-fold cross-validation
cv ← KFold(n_splits=n_folds, shuffle=True, random_state=42)
cv_scores ← []

FOR train_idx, val_idx IN cv.SPLIT(X_train):
    X_train_fold ← X_train[train_idx]
    y_train_fold ← y_train[train_idx]
    X_val_fold ← X_train[val_idx]
    y_val_fold ← y_train[val_idx]

    // Scale features

```

```

        scaler ← StandardScaler()
        X_train_scaled ← scaler.FIT_TRANSFORM(X_train_fold)
        X_val_scaled ← scaler.TRANSFORM(X_val_fold)

        // Train and evaluate
        ensemble.FIT(X_train_scaled, y_train_fold)
        y_val_pred ← ensemble.PREDICT(X_val_scaled)

        // Inverse transform predictions
        y_val_pred_orig ←
target_transformer.INVERSE_TRANSFORM(y_val_pred)
        y_val_orig ← target_transformer.INVERSE_TRANSFORM(y_val_fold)

        // Calculate R2
        r2 ← R2_SCORE(y_val_orig, y_val_pred_orig)
        APPEND r2 TO cv_scores

        // Return mean CV score
        RETURN MEAN(cv_scores)

    END FUNCTION

```

## 2. CREATE OPTUNA STUDY

```

study ← optuna.CREATE_STUDY(
    direction = "maximize", // Maximize R2
    study_name = "lightweight_optimization"
)

```

## 3. RUN OPTIMIZATION

```

PRINT "Starting Bayesian optimization with", n_trials, "trials..."

study.OPTIMIZE(
    objective,
    n_trials = n_trials,
    show_progress_bar = True
)

PRINT "Optimization complete!"

```

## 4. EXTRACT BEST PARAMETERS

```

best_params ← study.best_params
best_cv_score ← study.best_value

PRINT "Best CV R2:", best_cv_score
PRINT "Best hyperparameters:"
FOR param, value IN best_params:
    PRINT "  ", param, ":", value

```

```
5. RETURN  
   RETURN best_params, best_cv_score
```

```
END ALGORITHM
```

## Model Validation

### Algorithm 7: Model Validation on Independent Test Set

Plain Text

```
ALGORITHM: ValidateModel(model, X_test, y_test, scaler, target_transformer)
```

INPUT:

- model: Trained ensemble model
- X\_test: Test feature matrix (unscaled)
- y\_test: Test target (original scale)
- scaler: Fitted StandardScaler
- target\_transformer: Fitted PowerTransformer

OUTPUT:

- metrics: Dictionary with  $R^2$ , MAE, RMSE, Pearson r
- predictions: Array of predicted values
- residuals: Array of residuals (actual - predicted)

PROCEDURE:

1. SCALE TEST FEATURES  
X\_test\_scaled  $\leftarrow$  scaler.TRANSFORM(X\_test)
2. MAKE PREDICTIONS (Transformed Space)  
y\_test\_pred\_trans  $\leftarrow$  model.PREDICT(X\_test\_scaled)
3. INVERSE TRANSFORM TO ORIGINAL SCALE  
y\_test\_pred  $\leftarrow$  target\_transformer.INVERSE\_TRANSFORM(  
y\_test\_pred\_trans.RESHAPE(-1, 1)  
) .FLATTEN()
4. CALCULATE REGRESSION METRICS  
//  $R^2$  (coefficient of determination)  
r2  $\leftarrow$  R2\_SCORE(y\_test, y\_test\_pred)  
  
// MAE (mean absolute error)  
mae  $\leftarrow$  MEAN\_ABSOLUTE\_ERROR(y\_test, y\_test\_pred)  
  
// RMSE (root mean squared error)



```
mse ← MEAN_SQUARED_ERROR(y_test, y_test_pred)
rmse ← SQRT(mse)

// Pearson correlation coefficient
pearson_r ← PEARSON_CORRELATION(y_test, y_test_pred)

// Spearman rank correlation
spearman_rho ← SPEARMAN_CORRELATION(y_test, y_test_pred)
```

5. CALCULATE RESIDUALS

```
residuals ← y_test - y_test_pred

// Residual statistics
mean_residual ← MEAN(residuals)
std_residual ← STD(residuals)

PRINT "Mean residual:", mean_residual
PRINT "Std residual:", std_residual
```

6. PRINT RESULTS

```
PRINT "=" * 80
PRINT "INDEPENDENT TEST SET VALIDATION RESULTS"
PRINT "=" * 80
PRINT "R2 Score:", r2
PRINT "MAE:", mae, "UPDRS points"
PRINT "RMSE:", rmse, "UPDRS points"
PRINT "Pearson r:", pearson_r
PRINT "Spearman ρ:", spearman_rho
PRINT "=" * 80
```

7. PACKAGE METRICS

```
metrics ← {
  "r2": r2,
  "mae": mae,
  "rmse": rmse,
  "pearson_r": pearson_r,
  "spearman_rho": spearman_rho,
  "mean_residual": mean_residual,
  "std_residual": std_residual,
  "n_samples": LENGTH(y_test)
}
```

8. RETURN

```
RETURN metrics, y_test_pred, residuals
```

END ALGORITHM

# SHAP Analysis

## Algorithm 8: SHAP Feature Importance Analysis

Plain Text

```
ALGORITHM: ComputeSHAPValues(model, X_test, feature_names)
```

INPUT:

- model: Trained ensemble model
- X\_test: Test feature matrix (scaled)
- feature\_names: List of feature names

OUTPUT:

- shap\_values: SHAP importance values for each feature
- feature\_importance: Sorted list of (feature, importance) tuples

PROCEDURE:

1. CREATE SHAP EXPLAINER  
    // Use TreeExplainer for gradient boosting models  
    explainer ← shap.TreeExplainer(model)  
  
    PRINT "SHAP explainer created"
2. COMPUTE SHAP VALUES  
    PRINT "Computing SHAP values (this may take a few minutes)..."  
  
    shap\_values ← explainer.SHAP\_VALUES(X\_test)  
  
    PRINT "SHAP values computed for", LENGTH(X\_test), "samples"
3. CALCULATE MEAN ABSOLUTE SHAP VALUES  
    mean\_abs\_shap ← []  
  
    FOR i FROM 0 TO LENGTH(feature\_names)-1:  
        feature\_name ← feature\_names[i]  
        shap\_column ← shap\_values[:, i]  
  
        mean\_abs\_value ← MEAN(ABSOLUTE\_VALUE(shap\_column))  
  
        APPEND (feature\_name, mean\_abs\_value) TO mean\_abs\_shap
4. SORT BY IMPORTANCE  
    SORT mean\_abs\_shap BY mean\_abs\_value DESCENDING
5. PRINT TOP 20 FEATURES  
    PRINT "=" \* 80

```

PRINT "TOP 20 FEATURES BY SHAP IMPORTANCE"
PRINT "=" * 80

FOR i FROM 0 TO 19:
    feature, importance ← mean_abs_shap[i]
    PRINT i+1, ".", feature, ":", importance

PRINT "=" * 80

```

#### 6. CATEGORIZE FEATURES

```

// Separate by feature type
clinical_shap ← []
gene_shap ← []
pd_gene_shap ← []
pathway_shap ← []
interaction_shap ← []

FOR feature, importance IN mean_abs_shap:
    IF feature IN ["UPDRS_BL", "AGE", "GENDER"]:
        APPEND (feature, importance) TO clinical_shap
    ELSE IF feature STARTS_WITH "ENSG":
        APPEND (feature, importance) TO gene_shap
    ELSE IF feature STARTS_WITH "PD_":
        APPEND (feature, importance) TO pd_gene_shap
    ELSE IF feature STARTS_WITH "PATHWAY_":
        APPEND (feature, importance) TO pathway_shap
    ELSE IF feature CONTAINS "_x_":
        APPEND (feature, importance) TO interaction_shap

```

#### 7. PRINT CATEGORY SUMMARIES

```

PRINT "SHAP IMPORTANCE BY CATEGORY:"
PRINT "  Clinical features:", SUM([imp FOR _, imp IN clinical_shap])
PRINT "  Top genes:", SUM([imp FOR _, imp IN gene_shap[:20]])
PRINT "  PD risk genes:", SUM([imp FOR _, imp IN pd_gene_shap])
PRINT "  Pathways:", SUM([imp FOR _, imp IN pathway_shap])
PRINT "  Interactions:", SUM([imp FOR _, imp IN interaction_shap])

```

#### 8. RETURN

```

RETURN shap_values, mean_abs_shap

```

END ALGORITHM

## Clinical Prediction

### Algorithm 9: Predict for New Patient

## Plain Text

ALGORITHM: PredictNewPatient(patient\_data, model\_package)

### INPUT:

- patient\_data: Dictionary with baseline clinical data
  - {
    - "PATNO": "PATIENT\_001",
    - "UPDRS\_BL": 20.0,
    - "AGE": 68.0,
    - "GENDER": 1.0 // 0=Female, 1=Male
- model\_package: Loaded model artifacts

### OUTPUT:

- prediction\_report: Dictionary with predictions and interpretation

### PROCEDURE:

#### 1. EXTRACT MODEL ARTIFACTS

```
model ← model_package["ensemble_model"]
scaler ← model_package["scaler"]
target_transformer ← model_package["target_transformer"]
feature_names ← model_package["feature_names"]
n_features ← model_package["n_features"]
test_mae ← model_package["test_results"]["mae"]
```

#### 2. CREATE PATIENT DATAFRAME

```
patient_df ← DataFrame([patient_data])

PRINT "Patient ID:", patient_data["PATNO"]
PRINT "Baseline UPDRS:", patient_data["UPDRS_BL"]
PRINT "Age:", patient_data["AGE"]
PRINT "Gender:", "Male" IF patient_data["GENDER"]==1 ELSE "Female"
```

#### 3. IMPUTE MISSING FEATURES

```
// For clinical prediction, we only have baseline clinical data
// All gene expression and pathway features are imputed with 0
```

```
FOR feature IN feature_names:
  IF feature NOT IN patient_df.columns:
    patient_df[feature] ← 0.0
```

```
// Ensure correct column order
patient_df ← patient_df[feature_names]
```

```
PRINT "Warning: Missing", n_features - 4, "features (imputed with zeros)"
```

#### 4. SCALE FEATURES

```
X_patient ← patient_df.TO_NUMPY()  
X_patient_scaled ← scaler.TRANSFORM(X_patient)
```

#### 5. MAKE PREDICTION (Transformed Space)

```
y_pred_trans ← model.PREDICT(X_patient_scaled)
```

#### 6. INVERSE TRANSFORM TO UPDRS SCALE

```
y_pred_updrs ← target_transformer.INVERSE_TRANSFORM(  
  y_pred_trans.RESHAPE(-1, 1)  
) .FLATTEN()[0]
```

```
PRINT "Predicted UPDRS at 12 months:", y_pred_updrs
```

#### 7. CALCULATE PREDICTED CHANGE

```
baseline_updrs ← patient_data["UPDRS_BL"]  
predicted_change ← y_pred_updrs - baseline_updrs
```

```
PRINT "Predicted change:", predicted_change, "points"
```

#### 8. CALCULATE CONFIDENCE INTERVAL

```
// Use MAE as uncertainty estimate  
lower_bound ← y_pred_updrs - test_mae  
upper_bound ← y_pred_updrs + test_mae
```

```
PRINT "95% Confidence Interval: [", lower_bound, ",", upper_bound,
```

```
"]"
```

#### 9. CATEGORIZE PROGRESSION RISK

```
IF predicted_change < 0:  
  risk_category ← "Improvement"  
  confidence ← "Low" // Unusual, low confidence  
ELSE IF predicted_change < 3:  
  risk_category ← "Stable"  
  confidence ← "High"  
ELSE IF predicted_change < 5:  
  risk_category ← "Mild Progression"  
  confidence ← "High"  
ELSE IF predicted_change < 10:  
  risk_category ← "Moderate Progression"  
  confidence ← "High"  
ELSE:  
  risk_category ← "Rapid Progression"  
  confidence ← "High"
```

```
PRINT "Progression Risk:", risk_category
```

```
PRINT "Confidence:", confidence
```

```

10. GENERATE CLINICAL INTERPRETATION
    IF risk_category == "Stable":
        interpretation ← "Patient likely to remain stable over 12 months
(" +
                        predicted_change + " points change)."

        ELSE IF risk_category == "Mild Progression":
            interpretation ← "Mild progression expected (" +
predicted_change +
                        " points). Standard monitoring recommended."

        ELSE IF risk_category == "Moderate Progression":
            interpretation ← "Moderate progression expected (" +
predicted_change +
                        " points). Consider treatment adjustment."

        ELSE IF risk_category == "Rapid Progression":
            interpretation ← "Rapid progression expected (" +
predicted_change +
                        " points). Urgent clinical review recommended."

        ELSE: // Improvement
            interpretation ← "Patient shows predicted improvement (" +
                        predicted_change + " points). Monitor for
accuracy."

        PRINT "Clinical Interpretation:", interpretation

11. PACKAGE PREDICTION REPORT
    prediction_report ← {
        "patient_id": patient_data["PATNO"],
        "baseline_updrs": baseline_updrs,
        "predicted_updrs_12m": y_pred_updrs,
        "predicted_change": predicted_change,
        "lower_bound_12m": lower_bound,
        "upper_bound_12m": upper_bound,
        "progression_risk": risk_category,
        "confidence_level": confidence,
        "clinical_interpretation": interpretation
    }

12. RETURN
    RETURN prediction_report

END ALGORITHM

```

# Summary Statistics

## Key Performance Metrics

Plain Text		
<table><tr><th>MODEL PERFORMANCE SUMMARY</th></tr><tr><td>Training Set (n=312, 7-Fold CV): R<sup>2</sup> = 0.513 ± 0.052 MAE = 6.15 ± 0.25 UPDRS points RMSE = 7.82 ± 0.31 UPDRS points  Independent Test Set (n=78): R<sup>2</sup> = 0.551 (55.1% variance explained) MAE = 6.01 UPDRS points RMSE = 7.45 UPDRS points Pearson r = 0.74  Top 3 Features (SHAP): 1. UPDRS_BL × PINK1 = 0.283 2. UPDRS_BL = 0.258 3. ENSG00000243053 = 0.025</td></tr></table>	MODEL PERFORMANCE SUMMARY	Training Set (n=312, 7-Fold CV): R <sup>2</sup> = 0.513 ± 0.052 MAE = 6.15 ± 0.25 UPDRS points RMSE = 7.82 ± 0.31 UPDRS points  Independent Test Set (n=78): R <sup>2</sup> = 0.551 (55.1% variance explained) MAE = 6.01 UPDRS points RMSE = 7.45 UPDRS points Pearson r = 0.74  Top 3 Features (SHAP): 1. UPDRS_BL × PINK1 = 0.283 2. UPDRS_BL = 0.258 3. ENSG00000243053 = 0.025
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## Computational Complexity

Plain Text		
<table><tr><th>COMPUTATIONAL COMPLEXITY</th></tr><tr><td>Training Phase: Time Complexity: <math>O(n \times m \times t \times k)</math> n = number of samples (312) m = number of features (116) t = number of trees per model (~200) k = number of CV folds (7)  Space Complexity: <math>O(n \times m + t \times m)</math>  Actual Training Time: ~1 hour (30 Optuna trials)</td></tr></table>	COMPUTATIONAL COMPLEXITY	Training Phase: Time Complexity: $O(n \times m \times t \times k)$ n = number of samples (312) m = number of features (116) t = number of trees per model (~200) k = number of CV folds (7)  Space Complexity: $O(n \times m + t \times m)$  Actual Training Time: ~1 hour (30 Optuna trials)
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Prediction Phase:

Time Complexity:  $O(m \times t)$

Space Complexity:  $O(m)$

Actual Prediction Time: <1 second per patient

Model Size: 582 KB (compressed)

## End of Pseudo Code Documentation

**Version:** 1.0

**Last Updated:** November 15, 2025

**Author:** Clinical ML Research Team

For implementation details, see the actual Python code in `lightweight_optimization.py`