RWorksheet_Gerona#4b

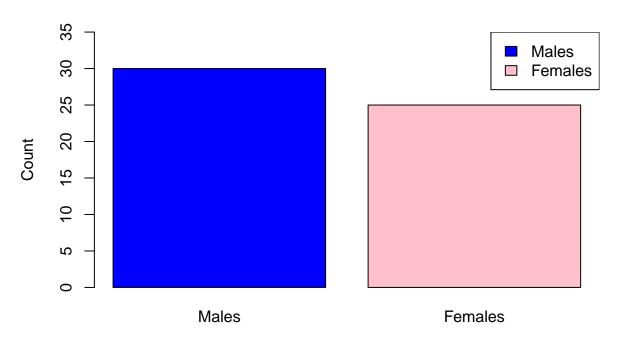
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```
#1. Create a 5x5 Matrix with for Loop
vectorA \leftarrow c(1, 2, 3, 4, 5)
matrixA <- matrix(0, nrow = 5, ncol = 5)</pre>
for (i in 1:5) {
  for (j in 1:5) {
    matrixA[i, j] <- abs(vectorA[i] - vectorA[j])</pre>
}
print(matrixA)
##
        [,1] [,2] [,3] [,4] [,5]
## [1,]
           0
                 1
                       2
                            2
                                 3
## [2,]
                 0
                       1
            1
## [3,]
           2
                                 2
                 1
                       0
                            1
## [4,]
            3
                       1
                                 1
## [5,]
            4
                 3
                       2
                            1
                                  0
#2. Print star pattern using for Loop
# Prints a triangle with 5 rows of stars
for(i in 1:5) {
  for(j in 1:i) {
    cat("*")
  }
  cat("\n")
}
## *
## **
## ***
## ****
## ****
#3. Fibonacci Sequence up to 500
fibonacci <- function(n) {</pre>
  a <- 0
  b <- 1
  repeat {
    cat(a, " ")
    temp <- a + b
    a <- b
    b <- temp
  if (a > 500) break
```

```
}
}
fibonacci()
## 0 1 1 2 3 5 8 13 21 34 55 89 144 233 377
#4. Importing and Analyzing Dataset #a. Import CSV file and display the first 6 rows
library(readr)
shoe_sizes <- read_csv("Shoe Sizes.csv")</pre>
## Rows: 28 Columns: 3
## -- Column specification
## Delimiter: ","
## chr (1): Gender
## dbl (2): Shoe size, Height
## i Use `spec()` to retrieve the full column specification for this data.
## i Specify the column types or set `show_col_types = FALSE` to quiet this message.
head(shoe_sizes)
## # A tibble: 6 x 3
     `Shoe size` Height Gender
##
           <dbl> <dbl> <chr>
## 1
             6.5
                    66 F
## 2
             9
                    68 F
## 3
             8.5
                    64.5 F
## 4
             8.5
                    65 F
## 5
            10.5
                    70
                         Μ
## 6
             7
                    64
                         F
#b. Create a subset for gender and count observations:
male <- subset(shoe_sizes, Gender == "M")</pre>
female <- subset(shoe_sizes, Gender == "F")</pre>
num_males <- nrow(male)</pre>
num females <- nrow(female)</pre>
cat("Number of Male observations:", num_males, "\n")
## Number of Male observations: 14
cat("Number of Female observations:", num_females, "\n")
## Number of Female observations: 14
#c. Barplot for Male and Female Counts
gender_counts <- c(Males = 30, Females = 25)</pre>
barplot(gender_counts,
        main = "Number of Males and Females in Households",
        xlab = "Gender",
        ylab = "Count",
        col = c("blue", "pink"),
        ylim = c(0, max(gender_counts) + 5),
        beside = TRUE)
```

```
legend("topright",
    legend = names(gender_counts),
    fill = c("blue", "pink"))
```

Number of Males and Females in Households



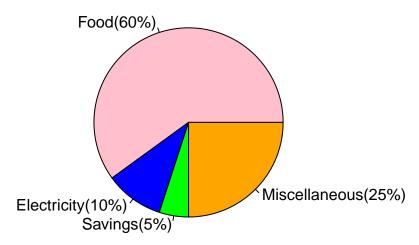
Gender

#5. Dela Cruz Family Monthly Income Pie Chart

```
# Data for Dela Cruz family spending
categories <- c("Food", "Electricity", "Savings", "Miscellaneous")
spending <- c(60, 10, 5, 25)

# Create a pie chart
pie(spending,
    labels = paste(categories, "(", spending, "%)", sep=""),
    col = c("pink", "blue", "green", "orange"),
    main = "Dela Cruz Family Monthly Spending Distribution")</pre>
```

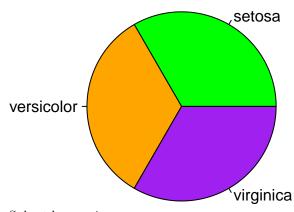
Dela Cruz Family Monthly Spending Distribution



#6. Iris Dataset Analysis #a. Check structure of the iris dataset:

```
data(iris)
str(iris)
## 'data.frame':
                    150 obs. of 5 variables:
## $ Sepal.Length: num 5.1 4.9 4.7 4.6 5 5.4 4.6 5 4.4 4.9 ...
## $ Sepal.Width : num 3.5 3 3.2 3.1 3.6 3.9 3.4 3.4 2.9 3.1 ...
## $ Petal.Length: num 1.4 1.4 1.3 1.5 1.4 1.7 1.4 1.5 1.4 1.5 ...
## $ Petal.Width : num 0.2 0.2 0.2 0.2 0.4 0.3 0.2 0.2 0.1 ...
                  : Factor w/ 3 levels "setosa", "versicolor", ...: 1 1 1 1 1 1 1 1 1 1 1 ...
## $ Species
\#b. Mean of Sepal and Petal dimensions:
mean_values <- colMeans(iris[, 1:4])</pre>
print(mean_values)
## Sepal.Length Sepal.Width Petal.Length Petal.Width
       5.843333
                    3.057333
                                 3.758000
                                               1.199333
#c. Species Distribution Pie Chart
pie(table(iris$Species), main = "Species Distribution in Iris Dataset", col = c("green", "orange", "pur
```

Species Distribution in Iris Dataset



#d. Subset by Species and Display Last 6 Rows: #

Subset by species

```
setosa <- subset(iris, Species == "setosa")
versicolor <- subset(iris, Species == "versicolor")
virginica <- subset(iris, Species == "virginica")
tail(setosa)</pre>
```

##		Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species
##	45	5.1	3.8	1.9	0.4	setosa
##	46	4.8	3.0	1.4	0.3	setosa
##	47	5.1	3.8	1.6	0.2	setosa
##	48	4.6	3.2	1.4	0.2	setosa
##	49	5.3	3.7	1.5	0.2	setosa
##	50	5.0	3.3	1.4	0.2	setosa

tail(versicolor)

##		Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species
##	95	5.6	2.7	4.2	1.3	versicolor
##	96	5.7	3.0	4.2	1.2	versicolor
##	97	5.7	2.9	4.2	1.3	versicolor
##	98	6.2	2.9	4.3	1.3	versicolor
##	99	5.1	2.5	3.0	1.1	versicolor
##	100	5.7	2.8	4.1	1.3	versicolor

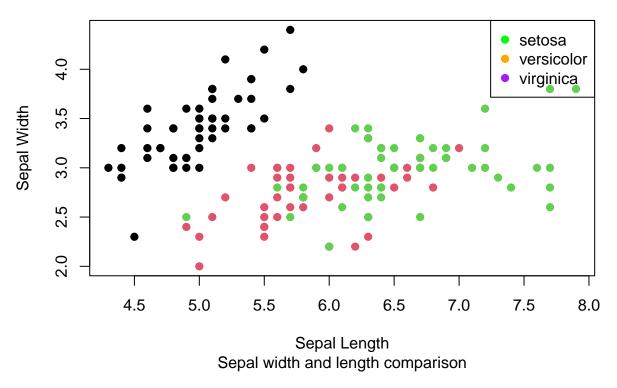
tail(virginica)

```
Sepal.Length Sepal.Width Petal.Length Petal.Width
##
                                                             Species
                                          5.7
## 145
                6.7
                             3.3
                                                      2.5 virginica
## 146
                6.7
                             3.0
                                          5.2
                                                      2.3 virginica
## 147
                6.3
                             2.5
                                          5.0
                                                      1.9 virginica
## 148
                6.5
                             3.0
                                          5.2
                                                      2.0 virginica
## 149
                6.2
                                          5.4
                             3.4
                                                      2.3 virginica
                                          5.1
## 150
                5.9
                             3.0
                                                      1.8 virginica
```

#e. Scatter Plot for Sepal Dimensions by Species: # Scatter plot for Sepal.Length and Sepal.Width

```
sub = "Sepal width and length comparison", col.main = "blue")
legend("topright", legend = levels(iris$Species), col = c("green", "orange", "purple"), pch = 19)
```

Iris Dataset



#6f. Interpretation of the result. The scatterplot illustrates the relationship between Sepal.Length and Sepal.Width across three species of flowers. Setosa points, shown in orange, typically have shorter sepal lengths and widths, clearly distinguishing them from the other species. Versicolor points, depicted in pink, are more widely distributed and exhibit slight overlap with Virginica points in yellow, indicating some similarities between these two species. In contrast, Virginica points tend to have larger sepal lengths and widths, further distinguishing them from Setosa while overlapping with some Versicolor points. Overall, the scatterplot effectively highlights the distinct separation of Setosa from the other species, with Versicolor and Virginica demonstrating some degree of overlap.

#7a. Import the alexa-file.xlsx, Clean Up Variants, and Rename

```
library(readxl)
data <- read_excel("/cloud/project/Worksheet#4/Worksheet#4b/alexa-file.xlsx")

data$variation <- gsub("Black Dot", "Black Dot", data$variation)
data$variation <- gsub("White Plus", "White Plus", data$variation)

head(data$variation)

## [1] "Black Dot" "Black Dot" "Black Dot" "Black Dot" "Black Dot"

#b. Get the total number of each variations and save it into another object.

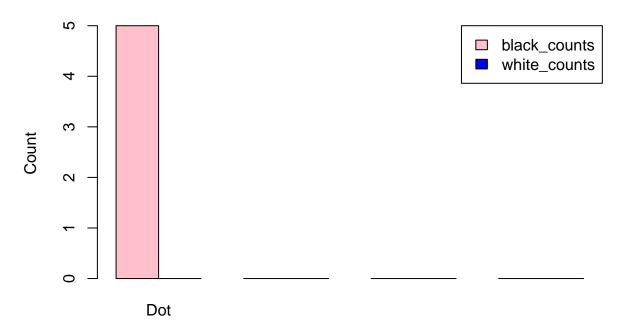
library(dplyr)

##

## Attaching package: 'dplyr'</pre>
```

```
## The following objects are masked from 'package:stats':
##
##
       filter, lag
## The following objects are masked from 'package:base':
##
##
       intersect, setdiff, setequal, union
RObject <- data.frame(columnName = c("A", "B", "A", "C", "B", "A", "D"))</pre>
variations <- RObject %>%
  count(columnName)
save(variations, file = "variations.RData")
print(variations)
     columnName n
## 1
              АЗ
## 2
              B 2
## 3
              C 1
## 4
              D 1
#7c
library(readxl)
library(dplyr)
alexa_file <- "/cloud/project/Worksheet#4/Worksheet#4b/alexa-file.xlsx"</pre>
data <- read_excel(alexa_file)</pre>
data$variation <- gsub("Black Dot", "Black Dot", data$variation)</pre>
data$variation <- gsub("Black Plus", "Black Plus", data$variation)</pre>
data$variation <- gsub("Black Show", "Black Show", data$variation)</pre>
data$variation <- gsub("Black Spot", "Black Spot", data$variation)</pre>
data$variation <- gsub("White Dot", "White Dot", data$variation)</pre>
data$variation <- gsub("White Plus", "White Plus", data$variation)</pre>
data$variation <- gsub("White Show", "White Show", data$variation)</pre>
data$variation <- gsub("White Spot", "White Spot", data$variation)
black_white_data <- data %>%
  filter(grepl("Black|White", variation))
variation_count <- black_white_data %>%
  count(variation)
black_variations <- variation_count %>% filter(grepl("Black", variation))
white variations <- variation count %>% filter(grepl("White", variation))
black_counts <- setNames(black_variations$n, gsub("Black ", "", black_variations$variation))
white_counts <- setNames(white_variations$n, gsub("White ", "", white_variations$variation))
common_variations <- c("Dot", "Plus", "Show", "Spot")</pre>
black_counts <- black_counts[common_variations]</pre>
black_counts[is.na(black_counts)] <- 0</pre>
```

Count of Black and White Variations



Variation Type