

# Package ‘metevalue’

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**Type** Package

**Title** E-Value in the Omics Data Association Studies

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**Description** In the omics data association studies, it is common to conduct the p-value corrections to control the false significance. Among those p-value correction methods, E-value is recently studied based on V. Vovk and R. Wang (2021) <doi:10.1214/20-AOS2020>. This package provides e-value calculation for several types of omics data association studies. Currently, five data formats are supported: BiSeq, MDRfinder, methylKit, metilene and RNAseq data. Other DNA methylation tools are also supported. The relevant references are listed below: Katja Hebestreit and Hans-Ulrich Klein (2022) <doi:10.18129/B9.bioc.BiSeq>; Al-tuna Akalin et.al (2012) <doi:10.18129/B9.bioc.methylKit>.

**License** Apache License (>= 2)

**RoxygenNote** 7.2.1

**Depends** sqldf, psych, dplyr, R (>= 3.5.0)

**Encoding** UTF-8

**Suggests** rmarkdown, prettydoc, knitr, ggplot2, tidyr, testthat (>= 3.0.0)

**VignetteBuilder** knitr

**LazyData** true

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**NeedsCompilation** no

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demo_biseq_DMR	<i>DMR BiSeq Demo Dataset</i>
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### Description

The BiSeq dataset for demo purpose. The data are dummy data. It includes 9 columns:

The dummy output for BiSeq illustrating purpose. It is dummy.

### Details

- seqnames: Chromosome
- start: The positions of the start sites of the corresponding region
- end: The positions of the end sites of the corresponding region
- strand: Strand
- median.p
- median.meth.group1
- median.meth.group2
- median.meth.diff
- seqnames
- start
- end
- width
- strand
- median.p
- median.meth.group1
- median.meth.group2
- median.meth.diff

Notice that there are "NaN" within the feature columns.

Please check the vignette "metevalvalue" for details.

---

demo_biseq_methyrate	<i>BiSeq Methyrate Demo Dataset</i>
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**Description**

The methyrate for BiSeq illustrating purpose. It is dummy.

**Details**

The data includes 12 columns.

- chr: string Chromosome

- pos: int Position

- g1~g2: methylation rate data in groups, repeat 5 times. Notice that there are "NaN" within the feature columns.

Please check the vignette "metevale" for details.

---

demo_desq_out	<i>DESeq Output Dataset</i>
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---

**Description**

The output dummy data for "RNA" meythod illustrating purpose.

**Details**

The data includes 10 columns.

- treated1fb:

- treated2fb:

- treated3fb:

- untreated1fb:

- untreated2fb:

- untreated3fb:

- untreated4fb:

This data contains 8166 rows and 7 columns.

Please check the vignette "metevale" for details.

## Examples

```
# library("pasilla")
# pasCts <- system.file("extdata",
#                       "pasilla_gene_counts.tsv",
#                       package="pasilla", mustWork=TRUE)
# pasAnno <- system.file("extdata",
#                       "pasilla_sample_annotation.csv",
#                       package="pasilla", mustWork=TRUE)
# cts <- as.matrix(read.csv(pasCts, sep="\t", row.names="gene_id"))
# coldata <- read.csv(pasAnno, row.names=1)
# coldata <- coldata[,c("condition", "type")]
# coldata$condition <- factor(coldata$condition)
# coldata$type <- factor(coldata$type)
#
# library("DESeq2")
# colnames(cts)=paste0(colnames(cts), 'fb')
# cts = cts[,rownames(coldata)]
# dds <- DESeqDataSetFromMatrix(countData = cts,
#                               colData = coldata,
#                               design = ~ condition)
# dds <- DESeq(dds)
#
#
# dat <- t(t(cts)/(dds$sizeFactor))
# dat.out <- dat[rowSums(dat >5)>=0.8*ncol(dat),]
#
# demo_desq_out <- log(dat.out)
```

---

demo_DMRfinder_DMRs	<i>DMRfinder Output Demo Dataset</i>
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---

## Description

The output dummy dataset for DMRfinder illustrating purpose.

## Details

The data includes 6 columns.

- chr: string Chromosome

- pos: int Position

- g1~g2: methylation rate data in groups, repeat 2 times. Notice that there are "NaN" within the feature columns.

Please check the vignette "metvalue" for details.

---

demo\_DMRfinder\_rate\_combine*DMRfinder Methyrate Demo Dataset*

---

**Description**

The methyrate for BiSeq illustrating purpose. It is dummy.

**Details**

The data includes 6 columns.

- chr: string Chromosome

- pos: int Position

- g1~g2: methylation rate data in groups, repeat 2 times. Notice that there are "NaN" within the feature columns.

Please check the vignette "metevale" for details.

---

demo\_methylkit\_methyrate*Methyrate Dataset*

---

**Description**

The methyrate dataset samples "myCpG" data from the methylKit (a bioconductor package) for illustrating purpose.

**Details**

The data includes 6 columns.

- chr: string Chromosome

- pos: int Position

- g1~g2: methylation rate data in groups (4 columns)

Please check the vignette "metevale" for details.

**References**

Akalin, Altuna, et al. "methylKit: a comprehensive R package for the analysis of genome-wide DNA methylation profiles." *Genome biology* 13.10 (2012): 1-9. doi: [10.1186/gb20121310r87](https://doi.org/10.1186/gb20121310r87)

---

demo\_methylkit\_met\_all

*Methyrate output dataset from methylKit*


---

### Description

The methyrate dataset samples "myCpG" data from the methylKit (a bioconductor package) for illustrating purpose.

### Details

The data includes 7 columns:

- chr: Chromosome
- start: The positions of the start sites of the corresponding region
- end: The positions of the end sites of the corresponding region
- strand: Strand
- pvalue: The adjusted p-value based on BH method in MWU-test
- qvalue: cutoff for qvalue of differential methylation statistic
- methyl.diff: The difference between the group means of methylation level

Please check the vignette "metevalue" for details.

### References

Akalin, Altuna, et al. "methylKit: a comprehensive R package for the analysis of genome-wide DNA methylation profiles." *Genome biology* 13.10 (2012): 1-9. doi: [10.1186/gb20121310r87](https://doi.org/10.1186/gb20121310r87)

---

demo\_metilene\_input

*Metilene Methyrate Demo Dataset*


---

### Description

The methyrate for metilene illustrating purpose. It is dummy.

### Details

The data includes 18 columns.

- chr: string Chromosome
- pos: int Position
- g1~g2: methylation rate data in groups.

Notice that there are "NaN" within the feature columns.

Please check the vignette "metevalue" for details.

---

demo_metilene_out	<i>Metilene Demo Output Dataset</i>
-------------------	-------------------------------------

---

### Description

The output dummy data for "metilene" meythod illustrating purpose.

### Details

The data includes 10 columns.

- V1: string Chromosome
- V2: The positions of the start sites of the corresponding region
- V3: The positions of the end sites of the corresponding region
- V4- V10: data value.

Please check the vignette "metevalue" for details.

---

evalue_buildin_sql	<i>Build-in data process function</i>
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---

### Description

Build-in data process function

### Usage

```
evalue_buildin_sql(a, b, method = "metilene")
```

### Arguments

- |        |  |
|--------|--|
| a      | data frame of the methylation rate                             |
| b      | data frame of output data corresponding to the "method" option |
| method | "metilene" or "biseq", "DMRfinder" or "methylKit"              |

### Value

a data frame combines data frame a and b corresponding to the "method" option

### Examples

```
data("demo_metilene_out")
data("demo_metilene_input")
result = evaluate_buildin_var_fmt_nm(demo_metilene_input,
                                     demo_metilene_out, method="metilene")
result_sql = evaluate_buildin_sql(result$a, result$b, method="metilene")
```

---

```
evaluate_buildin_var_fmt_nm
```

*Build-in check file format function Perform the format check and data clean for the "metilene" or "biseq", "DMRfinder" or "methylKit" method correspondingly.*

---

### Description

Build-in check file format function Perform the format check and data clean for the "metilene" or "biseq", "DMRfinder" or "methylKit" method correspondingly.

### Usage

```
evaluate_buildin_var_fmt_nm(a, b, method = "metilene")
```

### Arguments

a	data frame of the methylation rate
b	data frame of output data corresponding to the "method" option
method	"metilene" or "biseq", "DMRfinder" or "methylKit"

### Value

list(a, b) which contains the cleaned data correspondingly

### Examples

```
data("demo_metilene_out")
data("demo_metilene_input")
evaluate_buildin_var_fmt_nm(demo_metilene_input,
                           demo_metilene_out, method="metilene")
```

---

```
metevalue.biseq
```

*Calculate E-value of the BiSeq data format*

---

### Description

Please check vignette "metevalue" for details.

### Usage

```
metevalue.biseq(
  methyrate,
  BiSeq.output,
  adjust.methods = "BH",
  sep = "\t",
  bheader = FALSE
)
```



**Arguments**

methyrate	is the methyrate file. The columns are (in order): - chr: Chromosome - pos: int Position - g1~g2: methylation rate data in groups
BiSeq.output	is the output file of BiSeq. The columns are (in order): - seqnames: Chromosome - start: The positions of the start sites of the corresponding region - end: The positions of the end sites of the corresponding region - width: The number of CpG sites within the corresponding region - strand: Strand - median.p: The median p-value among CpG sites within the corresponding region - median.meth.group1: The median methylation rate in the first group among CpG sites within the corresponding region - median.meth.group2: The median methylation rate in the second group among CpG sites within the corresponding region - median.meth.diff: The median methylation difference between groups among CpG sites within the corresponding region
adjust.methods	is the adjust methods of e-value. It can be 'bonferroni', 'hochberg', 'holm', 'hommel', 'BH', 'BY'
sep	separator, default is the TAB key.
bheader	a logical value indicating whether the BiSeq.output file contains the names of the variables as its first line. By default, bheader = FALSE.

**Value**

a dataframe, the columns are (in order):

- chr: Chromosome
- start: The positions of the start sites of the corresponding region
- end: The positions of the end sites of the corresponding region
- q-value: The adjusted p-value based on BH method in MWU-test
- methyl.diff: The difference between the group means of methylation level
- CpGs: The number of CpG sites within the corresponding region
- p : p-value based on MWU-test
- p2: p-value based on 2D KS-test
- m1: The absolute mean methylation level for the corresponding segment of group 1
- m2: The absolute mean methylation level for the corresponding segment of group 2
- e\_value: The e-value of the corresponding region

**Examples**

```
#\donttest{
#data("demo_biseq_methyrate")
#data("demo_biseq_DMR")
#example_tempfiles = tempfile(c("demo_biseq_methyrate", "demo_biseq_DMR"))
```

```

#tempdir()
#### write to temp file ####
#write.table(demo_bisec_methyrate, file=example_tempfiles[1],row.names=FALSE,
#           col.names=TRUE, quote=FALSE, sep='\t')
#write.table(demo_bisec_DMR, file=example_tempfiles[2],
#           sep ="\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
#### compute e-value and its adjustment ####
#result = metevalue.bisec(example_tempfiles[1],
#           example_tempfiles[2], bheader = TRUE)
#}

```

---

metevalue.bisec.chk      *Check the BiSeq data format*

---

## Description

Check the BiSeq data format

## Usage

```

metevalue.bisec.chk(
  input_filename_a,
  input_filename_b,
  sep = "\t",
  bheader = FALSE
)

```

## Arguments

**input\_filename\_a**  
 metilene input file path. This file is a sep (e.g. TAB) separated file with two key columns and several value columns: For example:  
 chr pos g1 g1 g1 g1 g1 g1 g1 g2 g2 g2 g2 g2 g2 g2  
 - chr and pos are keys;  
 - g1~g2: methylation rate data in groups.

**input\_filename\_b**  
 metilene input file path. This file should stored as a sep(e.g. TAB) separated file with two key columns and several value columns: The columns are (in order):  
 - chr: Chromosome  
 - start: The position of the start site of the corresponding region  
 - end: The position of the end site of the corresponding region  
 - range: The range of the corresponding region  
 - strand: Strand  
 - median.p: The median of p-values in the corresponding region  
 - median.meth.group1 : The median of methylation level for the corresponding segment of group 1  
 - median.meth.group2 : The median of methylation level for the corresponding segment of group 2  
 - median.meth.diff: The median of the difference between the methylation level

**sep**  
 separator, default is the TAB key.

**bheader**  
 a logical value indicating whether the input\_filename\_b file contains the names of the variables as its first line. By default, bheader = FALSE.

## Value

list(file\_a, file\_b, file\_a\_b) returns a list with three pr-handled data.frames corresponding to the input\_filename\_a, input\_filename\_b file and a A JOIN B file.

## Examples

```
#data("demo_biseq_methyrate")
#data("demo_biseq_DMR")
#example_tempfiles = tempfile(c("demo_biseq_methyrate", "demo_biseq_DMR"))
#tempdir()
#write.table(demo_biseq_methyrate, file=example_tempfiles[1],row.names=FALSE,
#            col.names=TRUE, quote=FALSE, sep='\t')
#write.table(demo_biseq_DMR, file=example_tempfiles[2],
#            sep = "\t", row.names = FALSE, col.names = TRUE, quote = FALSE)
#### compute e-value and its adjustment ####
#result = metevalue.biseq.chk(example_tempfiles[1],
#                             example_tempfiles[2], bheader = TRUE)
```

---

metevalue.DMRfinder	<i>Calculate E-value of the DMRfinder data format</i>
---------------------	---

---

## Description

Calculate E-value of the DMRfinder data format

## Usage

```
metevalue.DMRfinder(
  methyrate,
  DMRfinder.output,
  adjust.methods = "BH",
  sep = "\t",
  bheader = FALSE
)
```

## Arguments

methyrate	is the methyrate file. - chr: Chromosome - pos: int Position - g1~g2: methylation rate data in groups
DMRfinder.output	is the output file of DMRfinder. - chr: Chromosome - start: The positions of the start sites of the corresponding region - end: The positions of the end sites of the corresponding region - CpG: The number of CpG sites within the corresponding region - Control.mu: The average methylation rate in control group - Expt1.mu: The average methylation rate in experiment group - Control.Expt1.diff: The methylation difference between control and experiment groups - Control.Expt1.pval: P-value based on Wald-test.

adjust.methods is the adjust methods of e-value. It can be 'bonferroni', 'hochberg', 'holm', 'hommel', 'BH', 'BY'

sep separator, default is the TAB key.

bheader a logical value indicating whether the DMRfinder.output file contains the names of the variables as its first line. By default, bheader = FALSE.

### Value

a dataframe, the columns are (in order):

- chr: Chromosome
- start: The positions of the start sites of the corresponding region
- end: The positions of the end sites of the corresponding region
- q-value: The adjusted p-value based on BH method in MWU-test
- methyl.diff: The difference between the group means of methylation level
- CpGs: The number of CpG sites within the corresponding region
- p : p-value based on MWU-test
- p2: p-value based on 2D KS-test
- m1: The absolute mean methylation level for the corresponding segment of group 1
- m2: The absolute mean methylation level for the corresponding segment of group 2
- e\_value: The e-value of the corresponding region

### Examples

```
data(demo_DMRfinder_rate_combine)
data(demo_DMRfinder_DMRs)
#example_tempfiles = tempfile(c("rate_combine", "DMRfinder_out"))
#tempdir()
#write.table(demo_DMRfinder_rate_combine, file=example_tempfiles[1],
#            row.names=FALSE, col.names=TRUE, quote=FALSE, sep='\t')
#write.table(demo_DMRfinder_DMRs, file=example_tempfiles[2],
#            sep = "\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
#result = metevalue._DMRfinder(example_tempfiles[1], example_tempfiles[2],
#                               bheader = TRUE)
#head(result)
```

---

metevalue.DMRfinder.chk

*Check the DMRfinder data format*

---

### Description

Check the DMRfinder data format

### Usage

```
metevalue.DMRfinder.chk(
  input_filename_a,
  input_filename_b,
  sep = "\t",
  bheader = FALSE
)
```

**Arguments**

<code>input_filename_a</code>	the combined data of methylation rate file. This file is a sep (e.g. TAB) separated file with two key columns and several value columns: For example: chr pos g1 g1 g1 g1 g1 g1 g1 g2 g2 g2 g2 g2 g2 g2 - chr and pos are keys; - g1~g2: methylation rate data in groups.
<code>input_filename_b</code>	the output file of DMRfinder. The columns are (in order): - chr: Chromosome - start: The position of the start sites of the corresponding region - end: The position of the end sites of the corresponding region - CpG: The number of CpG sites within the corresponding region - 'Control:mu': The absolute mean methylation level for the corresponding segment of the control group - 'Exptl:mu': The absolute mean methylation level for the corresponding segment of the experimental group - 'Control->Exptl:diff': The difference between the group means of methylation level - p: p-value
<code>sep</code>	separator, default is the TAB key.
<code>bheader</code>	a logical value indicating whether the <code>input_filename_b</code> file contains the names of the variables as its first line. By default, <code>bheader = FALSE</code> .

**Value**

`list(file_a, file_b, file_a_b)` returns a list with three pre-handled data.frames corresponding to the `input_filename_a`, `input_filename_b` file and a A JOIN B file.

**Examples**

```
data(demo_DMRfinder_rate_combine)
data(demo_DMRfinder_DMRs)
#example_tempfiles = tempfile(c("rate_combine", "DMRfinder_out"))
#tempdir()
#write.table(demo_DMRfinder_rate_combine, file=example_tempfiles[1],
#           row.names=FALSE, col.names=TRUE, quote=FALSE, sep='\t')
#write.table(demo_DMRfinder_DMRs, file=example_tempfiles[2],
#           sep="\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
#result = metevalue.DMRfinder.chk(example_tempfiles[1], example_tempfiles[2],
#           bheader = TRUE)
```

---

metevalue.methylKit     *Calculate E-value of the methylKit data format*

---

**Description**

Calculate E-value of the methylKit data format

**Usage**

```
metevalue.methylKit(
  methyrate,
  methylKit.output,
  adjust.methods = "BH",
  sep = "\t",
  bheader = FALSE
)
```

**Arguments**

**methyrate** is the data of methylation rates of each sites and group, the columns are (in order):

- chr: Chromosome
- pos: int Position
- g1~g2: methylation rate data in groups

**methylKit.output** is the output data with e-value of each region

- chr: Chromosome
- start: The positions of the start sites of the corresponding region
- end: The positions of the end sites of the corresponding region
- strand: Strand
- pvalue: The adjusted p-value based on BH method in MWU-test
- qvalue: cutoff for qvalue of differential methylation statistic
- methyl.diff: The difference between the group means of methylation level

**adjust.methods** is the adjust methods of e-value. It can be 'bonferroni', 'hochberg', 'holm', 'hommel', 'BH', 'BY'

**sep** separator, default is the TAB key.

**bheader** a logical value indicating whether the input\_filename\_b file contains the names of the variables as its first line. By default, bheader = FALSE.

**Value**

a dataframe, the columns are (in order):

- chr: Chromosome
- start: The positions of the start sites of the corresponding region
- end: The positions of the end sites of the corresponding region
- q-value: The adjusted p-value based on BH method in MWU-test
- methyl.diff: The difference between the group means of methylation level
- CpGs: The number of CpG sites within the corresponding region
- p : p-value based on MWU-test
- p2: p-value based on 2D KS-test
- m1: The absolute mean methylation level for the corresponding segment of group 1
- m2: The absolute mean methylation level for the corresponding segment of group 2
- e\_value: The e-value of the corresponding region

**Examples**

```
data(demo_methylkit_methyrate)
data(demo_methylkit_met_all)
example_tempfiles = tempfile(c("rate_combine", "methylKit_DMR_raw"))
tempdir()
write.table(demo_methylkit_methyrate, file=example_tempfiles[1],
            row.names=FALSE, col.names=TRUE, quote=FALSE, sep='\t')
write.table(demo_methylkit_met_all, file=example_tempfiles[2],
            sep = "\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
result = metevalue.methylKit(example_tempfiles[1], example_tempfiles[2],
                             bheader = TRUE)
str(result)
```

---

metevalue.methylKit.chk

*Check the methylKit data format*


---

**Description**

Check the methylKit data format

**Usage**

```
metevalue.methylKit.chk(
  input_filename_a,
  input_filename_b,
  sep = "\t",
  bheader = FALSE
)
```

**Arguments**

**input\_filename\_a**  
the combined data of methylation rate file. This file is a sep (e.g. TAB) separated file with two key columns and several value columns: For example:  
chr pos g1 g1 g1 g1 g1 g1 g1 g2 g2 g2 g2 g2 g2 g2  
- chr and pos are keys;  
- g1~g2: methylation rate data in groups.

**input\_filename\_b**  
the output file of methylKit. a methylDiff or methylDiffDB object containing the differential methylated locations satisfying the criteria. The columns are (in order):  
- chr: Chromosome  
- start: The position of the start sites of the corresponding region  
- end: The position of the end sites of the corresponding region  
- strand: Strand  
- p: p-value  
- qvalue: The adjusted p-value based on BH method  
- meth.diff : The difference between the group means of methylation level

**sep**  
separator, default is the TAB key.

**bheader**  
a logical value indicating whether the input\_filename\_b file contains the names of the variables as its first line. By default, bheader = FALSE.

**Value**

list(file\_a, file\_b, file\_a\_b) returns a list with three pr-handled data.frames corresponding to the input\_filename\_a, input\_filename\_b file and a A JOIN B file.

**Examples**

```
data(demo_methylkit_methyrate)
data(demo_methylkit_met_all)
example_tempfiles = tempfile(c("rate_combine", "methylKit_DMR_raw"))
tempdir()
write.table(demo_methylkit_methyrate, file=example_tempfiles[1],
            row.names=FALSE, col.names=TRUE, quote=FALSE, sep='\t')
write.table(demo_methylkit_met_all, file=example_tempfiles[2],
            sep = "\t", row.names = FALSE, col.names = TRUE, quote = FALSE)
result = metevalue.methylKit.chk(example_tempfiles[1], example_tempfiles[2],
                                bheader = TRUE)
```

---

metevalue.metilene	<i>Calculate E-value of the Metilene data format</i>
--------------------	--

---

**Description**

Calculate E-value of the Metilene data format

**Usage**

```
metevalue.metilene(
  methyrate,
  metilene.output,
  adjust.methods = "BH",
  sep = "\t",
  bheader = FALSE
)
```

**Arguments**

methyrate	metilene input file path. This file is a sep (e.g. TAB) separated file with two key columns and several value columns: For example: chr pos g1 g1 g1 g1 g1 g1 g1 g2 g2 g2 g2 g2 g2 g2 - chr and pos are keys; - g1~g2: methylation rate data in groups.
metilene.output	metilene input file path. This file should stored as a sep(e.g. TAB) separated file with two key columns and several value columns: The columns are (in order): - chr: Chromosome - start: The positions of the start sites of the corresponding region - end: The positions of the end sites of the corresponding region - q-value: The adjusted p-value based on BH method in MWU-test - methyl.diff: The difference between the group means of methylation level - CpGs: The number of CpG sites within the corresponding region





---

metevalue.metilene.chk

*Check the Metilene data format*


---

## Description

Check the Metilene data format

## Usage

```
metevalue.metilene.chk(
  input_filename_a,
  input_filename_b,
  sep = "\t",
  bheader = FALSE
)
```

## Arguments

**input\_filename\_a**  
metilene input file path. This file is a sep (e.g. TAB) separated file with two key columns and several value columns: For example:  
chr pos g1 g1 g1 g1 g1 g1 g1 g2 g2 g2 g2 g2 g2 g2 g2  
- chr and pos are keys;  
- g1~g2: methylation rate data in groups.

**input\_filename\_b**  
metilene input file path. This file should stored as a sep(e.g. TAB) separated file with two key columns and several value columns: The columns are (in order):  
- chr: Chromosome  
- start: The position of the start sites of the corresponding region  
- end: The position of the end sites of the corresponding region  
- q-value: The adjusted p-value based on BH method in MWU-test  
- methyl.diff: The difference between the group means of methylation level  
- CpGs: The number of CpG sites within the corresponding region  
- p : p-value based on MWU-test  
- p2: p-value based on 2D KS-test  
- m1: The absolute mean methylation level for the corresponding segment of group 1  
- m2: The absolute mean methylation level for the corresponding segment of group 2

**sep**  
separator, default is the TAB key.

**bheader**  
a logical value indicating whether the input\_filename\_b file contains the names of the variables as its first line. By default, bheader = FALSE.

## Value

list(file\_a, file\_b, file\_a\_b) returns a list with three pr-handled data.frames corresponding to the input\_filename\_a, input\_filename\_b file and a A JOIN B file.

**Examples**

```
#data(demo_metilene_input)
#data(demo_metilene_out)
#example_tempfiles = tempfile(c("metilene_input", "metilene_out"))
#tempdir()
#write.table(demo_metilene_input, file=example_tempfiles[1],
#            row.names=FALSE, col.names=TRUE, quote=FALSE, sep='\t')
#write.table(demo_metilene_out, file=example_tempfiles[2],
#            sep="\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
#result = metevalue.metilene.chk(example_tempfiles[1], example_tempfiles[2],
#                                bheader = TRUE)
```

---

metevalue.RNA\_general *A general method to calculate the e-value for RNA-seq data.*

---

**Description**

A general method to calculate the e-value for RNA-seq data.

**Usage**

```
metevalue.RNA_general(rna, group1_name, group2_name)
```

**Arguments**

rna	data.frame: A data.frame object of RNAseq data. For example: <pre>treated1fb treated2fb untreated1fb untreated2fb TAG1 4.449648 4.750104 4.392285 4.497514 TAG2 8.241116 8.302852 8.318125 8.488796 ...</pre> Row names (TAG1 and TAG2 in the above example) is also suggested.
group1_name	charactor: The name of the first group. For example, "treated" in the example.
group2_name	charactor: The name of the second group. For example, "untreated" in the example.

**Value**

evaluate

**Examples**

```
data("demo_desq_out")
evaluate = metevalue.RNA_general(demo_desq_out, 'treated', 'untreated')
```

---

```
varevalue.single_general
```

*A general method to calculate the e-value for other DNA methylation tools not described above. The input data is the DNA methylation rates using the same format with Metilene.*

---

## Description

The data file could be pre-handled by the `metevalue.[types].chk` function. The Chromosome name, start and end sites should be specified.

## Usage

```
varevalue.single_general(
  methyrate,
  group1_name = "g1",
  group2_name = "g2",
  chr,
  start,
  end
)
```

## Arguments

<code>methyrate</code>	data.frame: A data.frame object of methylation rates, the columns should be (name of groups can be self-defined) chr pos group1_name group1_name ... group1_name group2_name group2_name
<code>group1_name</code>	character: The name of the first group. For example, "treated" in the above example.
<code>group2_name</code>	character: The name of the second group. For example, "untreated" in the above example.
<code>chr</code>	character: The Chromosome name. Typically, it is a string like "chr21" and so on.
<code>start</code>	integer: The position of the start site of the corresponding region
<code>end</code>	integer: The position of the end site of the corresponding region

## Value

evaluate

## Examples

```
#data("demo_metilene_input")
#varevalue.single_general(demo_metilene_input, chr = "chr21", start = 9437432, end = 9437540)
# [1] 2.626126e+43

#### Compare to `metevalue.metilene` ####
data(demo_metilene_out)
#example_tempfiles = tempfile(c("metilene_input", "metilene_out"))
#tempdir()
```

```
#write.table(demo_metilene_input, file=example_tempfiles[1],
#            row.names=FALSE, col.names=TRUE, quote=FALSE, sep='\t')
#write.table (demo_metilene_out, file=example_tempfiles[2],
#            sep = "\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
#result = metevalue.metilene(example_tempfiles[1], example_tempfiles[2],
#                             bheader = TRUE)
# result[with(result, chr == 'chr21' & start == '9437432' & end == '9437540'), ncol(result)]
# [1] 2.626126e+43
```

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