Package 'metevalue'

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Type Package

Title E-value in the Omics Data Association Studies

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Description
In the omics data association studies, it is common to conduct the p-value corrections to control the false significance. Beyond the P-value corrections, E-value is recently studied to facilitate multiple testing correction based on V. Vovk and R. Wang (2021) <doi:10.1214/20-AOS2020>. This package provides E-value calculation for DNA methylation data and RNA-seq data. Currently, five data formats are supported: DNA methylation levels using DMR detection tools (BiSeq, MDRfinder, methylKit, metilene and other DNA methylation tools) and RNA-seq data. The relevant references are listed below: Katja Hebestreit and Hans-Ulrich Klein (2022) <doi:10.18129/B9.bioc.BiSeq>; Altuna Akalin et.al (2012) <doi:10.18129/B9.bioc.methylKit>.

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RoxygenNote 7.2.1

Depends sqldf, psych, dplyr, R (>= 3.5.0)

Encoding UTF-8

Suggests rmarkdown, prettydoc, knitr, ggplot2, tidyr, testthat (>= 3.0.0)

VignetteBuilder knitr

LazyData true

Config/testthat/edition 3

NeedsCompilation no

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R topics documented:

`demo_biseq_DMR` *BiSeq Output Demo Dataset*

Description

The dummy output for BiSeq illustrating purpose. It is dummy.

Details

- seqnames: Chromosome

- start: The positions of the start sites of the corresponding region

- end: The positions of the end sites of the corresponding region

- width

- strand: Strand

- median.p

- median.meth.group1

- median.meth.group2

- median.meth.diff

Notice that there are "NaN" within the feature columns.

Please check the vignette "metevalue" for details.

demo_biseq_methyrate *BiSeq Methyrate Demo Dataset*

Description

The methyrate for BiSeq illustrating purpose. It is dummy.

Details

The data includes 12 columns.

- chr: string Chromosome

- pos: int Position

- g1~g2: methylation rate data in groups, repeat 5 times. Notice that there are "NaN" within the feature columns.

Please check the vignette "metevalue" for details.

demo_desq_out *DESeq Output Dataset*

Description

The output dummy data for "RNA" meythod illustrating purpose.

Details

The data includes 10 columns.

- treated1fb:

- treated2fb:

- treated3fb:

- untreated1fb:

- untreated2fb:

- untreated3fb:

- untreated4fb:

This data contains 8166 rows and 7 columns.

Please check the vignette "metevalue" for details.

Examples

```
# library("pasilla")
# pasCts <- system.file("extdata",
#                       "pasilla_gene_counts.tsv",
#                       package="pasilla", mustWork=TRUE)
# pasAnno <- system.file("extdata",
#                        "pasilla_sample_annotation.csv",
#                        package="pasilla", mustWork=TRUE)
# cts <- as.matrix(read.csv(pasCts,sep="\t",row.names="gene_id"))
# coldata <- read.csv(pasAnno, row.names=1)
# coldata <- coldata[,c("condition","type")]
# coldata$condition <- factor(coldata$condition)
# coldata$type <- factor(coldata$type)
#
# library("DESeq2")
# colnames(cts)=paste0(colnames(cts),'fb')
# cts = cts[,rownames(coldata)]
# dds <- DESeqDataSetFromMatrix(countData = cts,
#                               colData = coldata,
#                               design = ~ condition)
# dds <- DESeq(dds)
#
#
# dat <- t(t(cts)/(dds$sizeFactor))
# dat.out <- dat[rowSums(dat >5)>=0.8*ncol(dat),]
#
# demo_desq_out <- log(dat.out)
```

`demo_DMRfinder_DMRs` *DMRfinder Output Demo Dataset*

Description

The output dummy dataset for DMRfinder illustrating purpose.

Details

The data includes 6 columns.

- chr: string Chromosome

- pos: int Position

- g1~g2: methylation rate data in groups, repeat 2 times. Notice that there are "NaN" within the feature columns.

Please check the vignette "metevalue" for details.

demo_DMRfinder_rate_combine

DMRfinder Methyrate Demo Dataset

Description

The methyrate for BiSeq illustrating purpose. It is dummy.

Details

The data includes 6 columns.

- chr: string Chromosome

- pos: int Position

- g1~g2: methylation rate data in groups, repeat 2 times. Notice that there are "NaN" within the feature columns.

Please check the vignette "metevalue" for details.

demo_methylkit_methyrate

Methyrate Dataset

Description

The methyrate dataset samples "myCpG" data from the methylKit (a bioconductor package) for illustrating purpose.

Details

The data includes 6 columns.

- chr: string Chromosome

- pos: int Position

- g1~g2: methylation rate data in groups (4 columns)

Please check the vignette "metevalue" for details.

References

Akalin, Altuna, et al. "methylKit: a comprehensive R package for the analysis of genome-wide DNA methylation profiles." Genome biology 13.10 (2012): 1-9. doi: 10.1186/gb20121310r87

`demo_methylkit_met_all`

Methyrate output dataset from methylKit

Description

The methyrate dataset samples "myCpG" data from the methylKit (a bioconductor package) for illustrating purpose.

Details

The data includes 7 columns:

- chr: Chromosome

- start: The positions of the start sites of the corresponding region

- end: The positions of the end sites of the corresponding region

- strand: Strand

- pvalue: The adjusted p-value based on BH method in MWU-test

- qvalue: cutoff for qvalue of differential methylation statistic

- methyl.diff: The difference between the group means of methylation level

Please check the vignette "metevalue" for details.

References

Akalin, Altuna, et al. "methylKit: a comprehensive R package for the analysis of genome-wide DNA methylation profiles." Genome biology 13.10 (2012): 1-9. doi: 10.1186/gb20121310r87

`demo_metilene_input` *Metilene Methyrate Demo Dataset*

Description

The methyrate for metilene illustrating purpose. It is dummy.

Details

The data includes 18 columns.

- chr: string Chromosome

- pos: int Position

- g1~g2: methylation rate data in groups.

Notice that there are "NaN" within the feature columns.

Please check the vignette "metevalue" for details.

demo_metilene_out *Metilene Demo Output Dataset*

Description

The output dummy data for "metilene" meythod illustrating purpose.

Details

The data includes 10 columns.

- V1: string Chromosome
- V2: The positions of the start sites of the corresponding region
- V3: The positions of the end sites of the corresponding region
- V4- V10: data value.

Please check the vignette "metevalue" for details.

evalue_buildin_sql *Build-in data process function*

Description

Build-in data process function

Usage

```
evalue_buildin_sql(a, b, method = "metilene")
```

Arguments

a	data frame of the methylation rate
b	data frame of output data corresponding to the "method" option
method	"metilene" or "biseq", "DMRfinder" or "methylKit"

Value

a data frame combines data frame a and b corresponding to the "method" option

Examples

```
data("demo_metilene_out")
data("demo_metilene_input")
result = evalue_buildin_var_fmt_nm(demo_metilene_input,
                                   demo_metilene_out, method="metilene")
result_sql = evalue_buildin_sql(result$a, result$b, method="metilene")
```

`evalue_buildin_var_fmt_nm` *Build-in check file format function Perform the format check and data clean for the "metilene" or "biseq", "DMRfinder" or "methylKit" method correspondingly.*

Description

Build-in check file format function Perform the format check and data clean for the "metilene" or "biseq", "DMRfinder" or "methylKit" method correspondingly.

Usage

```
evalue_buildin_var_fmt_nm(a, b, method = "metilene")
```

Arguments

a	data frame of the methylation rate
b	data frame of output data corresponding to the "method" option
method	"metilene" or "biseq", "DMRfinder" or "methylKit"

Value

list(a, b) which contains the cleaned data correspondingly

Examples

```
data("demo_metilene_out")
data("demo_metilene_input")
evalue_buildin_var_fmt_nm(demo_metilene_input,
                          demo_metilene_out, method="metilene")
```

`metevalue.biseq` *Calculate E-value of the BiSeq data format*

Description

Please check vignette "metevalue" for details.

Usage

```
metevalue.biseq(
  methyrate,
  BiSeq.output,
  adjust.methods = "BH",
  sep = "\t",
  bheader = FALSE
)
```

Arguments

`methyrate`	is the methyrate file. The columns are (in order): - chr: Chromosome - pos: int Position - g1~g2: methylation rate data in groups
`BiSeq.output`	is the output file of BiSeq. The columns are (in order): - seqnames: Chromosome - start: The positions of the start sites of the corresponding region - end: The positions of the end sites of the corresponding region - width: The number of CpG sites within the corresponding region - strand: Strand - median.p: The median p-value among CpG sites within the corresponding region - median.meth.group1: The median methylation rate in the first group among CpG sites within the corresponding region - median.meth.group2: The median methylation rate in the second group among CpG sites within the corresponding region - median.meth.diff: The median methylation difference between groups among CpG sites within the corresponding region
`adjust.methods`	is the adjust methods of e-value. It can be 'bonferroni', 'hochberg', 'holm', 'hommel', 'BH', 'BY'
`sep`	seperator, default is the TAB key.
`bheader`	a logical value indicating whether the BiSeq.output file contains the names of the variables as its first line. By default, bheader = FALSE.

Value

a dataframe, the columns are (in order):

- chr: Chromosome
- start: The positions of the start sites of the corresponding region
- end: The positions of the end sites of the corresponding region
- q-value: The adjusted p-value based on BH method in MWU-test
- methyl.diff: The difference between the group means of methylation level
- CpGs: The number of CpG sites within the corresponding region
- p : p-value based on MWU-test
- p2: p-value based on 2D KS-test
- m1: The absolute mean methylation level for the corresponding segment of group 1
- m2: The absolute mean methylation level for the corresponding segment of group 2
- e_value: The e-value of the corresponding region

Examples

```
#\donttest{
#data("demo_biseq_methyrate")
#data("demo_biseq_DMR")
#example_tempfiles = tempfile(c("demo_biseq_methyrate", "demo_biseq_DMR"))
```

```
#tempdir()
#### write to temp file ####
#write.table(demo_biseq_methyrate, file=example_tempfiles[1],row.names=FALSE,
#            col.names=TRUE, quote=FALSE, sep='\t')
#write.table(demo_biseq_DMR, file=example_tempfiles[2],
#             sep ="\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
#### compute e-value and its adjustment ####
#result = metevalue.biseq(example_tempfiles[1],
#                         example_tempfiles[2], bheader = TRUE)
#}
```

`metevalue.biseq.chk` *Check the BiSeq data format*

Description

Check the BiSeq data format

Usage

```
metevalue.biseq.chk(
  input_filename_a,
  input_filename_b,
  sep = "\t",
  bheader = FALSE
)
```

Arguments

`input_filename_a`
: metilene input file path. This file is a sep (e.g. TAB) separated file with two key columns and several value columns: For exampe:
chr pos g1 g1 g1 g1 g1 g1 g1 g1 g2 g2 g2 g2 g2 g2 g2 g2
- chr and pos are keys;
- g1~g2: methylation rate data in groups.

`input_filename_b`
: metilene input file path. This file should stored as a sep(e.g. TAB) separated file with two key columns and several value columns: The columns are (in order):
- chr: Chromosome
- start: The position of the start site of the corresponding region
- end: The position of the end site of the corresponding region
- range: The range of the corresponding region
- strand: Strand
- median.p: The median of p-values in the corresponding region
- median.meth.group1 : The median of methylation level for the corresponding segment of group 1
- median.meth.group2 : The median of methylation level for the corresponding segment of group 2
- median.meth.diff: The median of the difference between the methylation level

`sep`
: separator, default is the TAB key.

`bheader`
: a logical value indicating whether the input_filename_b file contains the names of the variables as its first line. By default, bheader = FALSE.

Value

list(file_a, file_b, file_a_b) returns a list with three pr-handled data.frames corresponding to the input_filename_a, input_filename_b file and a A JOIN B file.

Examples

```
#data("demo_biseq_methyrate")
#data("demo_biseq_DMR")
#example_tempfiles = tempfile(c("demo_biseq_methyrate", "demo_biseq_DMR"))
#tempdir()
#write.table(demo_biseq_methyrate, file=example_tempfiles[1],row.names=FALSE,
#            col.names=TRUE, quote=FALSE, sep='\t')
#write.table(demo_biseq_DMR, file=example_tempfiles[2],
#             sep ="\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
#### compute e-value and its adjustment ####
#result = metevalue.biseq.chk(example_tempfiles[1],
#                         example_tempfiles[2], bheader = TRUE)
```

`metevalue.DMRfinder` *Calculate E-value of the DMRfinder data format*

Description

Calculate E-value of the DMRfinder data format

Usage

```
metevalue.DMRfinder(
  methyrate,
  DMRfinder.output,
  adjust.methods = "BH",
  sep = "\t",
  bheader = FALSE
)
```

Arguments

`methyrate` is the methyrate file.
- chr: Chromosome
- pos: int Position
- g1~g2: methylation rate data in groups

`DMRfinder.output` is the output file of DMRfinder.
- chr: Chromosome
- start: The positions of the start sites of the corresponding region
- end: The positions of the end sites of the corresponding region
- CpG: The number of CpG sites within the corresponding region
- Control.mu: The average methylation rate in control group
- Expt1.mu: The average methylation rate in experiment group
- Control.Expt1.diff: The methylation difference between control and experiment groups
- Control.Expt1.pval: P-value based on Wald-test.

adjust.methods	is the adjust methods of e-value. It can be 'bonferroni', 'hochberg', 'holm', 'hommel', 'BH', 'BY'
sep	seperator, default is the TAB key.
bheader	a logical value indicating whether the DMRfinder.output file contains the names of the variables as its first line. By default, bheader = FALSE.

Value

a dataframe, the columns are (in order):

- chr: Chromosome
- start: The positions of the start sites of the corresponding region
- end: The positions of the end sites of the corresponding region
- q-value: The adjusted p-value based on BH method in MWU-test
- methyl.diff: The difference between the group means of methylation level
- CpGs: The number of CpG sites within the corresponding region
- p : p-value based on MWU-test
- p2: p-value based on 2D KS-test
- m1: The absolute mean methylation level for the corresponding segment of group 1
- m2: The absolute mean methylation level for the corresponding segment of group 2
- e_value: The e-value of the corresponding region

Examples

```
data(demo_DMRfinder_rate_combine)
data(demo_DMRfinder_DMRs)
#example_tempfiles = tempfile(c("rate_combine", "DMRfinder_out"))
#tempdir()
#write.table(demo_DMRfinder_rate_combine, file=example_tempfiles[1],
#      row.names=FALSE, col.names=TRUE, quote=FALSE, sep='\t')
#write.table(demo_DMRfinder_DMRs, file=example_tempfiles[2],
#      sep ="\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
#result = metevalue._DMRfinder(example_tempfiles[1], example_tempfiles[2],
#      bheader = TRUE)
#head(result)
```

---

metevalue.DMRfinder.chk

Check the DMRfinder data format

Description

Check the DMRfinder data format

Usage

```
metevalue.DMRfinder.chk(
  input_filename_a,
  input_filename_b,
  sep = "\t",
  bheader = FALSE
)
```

Arguments

input_filename_a	the combined data of methylation rate file. This file is a sep (e.g. TAB) separated file with two key columns and several value columns: For exampe: chr pos g1 g1 g1 g1 g1 g1 g1 g1 g2 g2 g2 g2 g2 g2 g2 g2 - chr and pos are keys; - g1~g2: methylation rate data in groups.
input_filename_b	the output file of DMRfinder. The columns are (in order): - chr: Chromosome - start: The position of the start sites of the corresponding region - end: The position of the end sites of the corresponding region - CpG: The number of CpG sites within the corresponding region - 'Control:mu': The absolute mean methylation level for the corresponding segment of the control group - 'Exptl:mu': The absolute mean methylation level for the corresponding segment of the experimental group - 'Control->Exptl:diff': The difference between the group means of methylation level - p: p-value
sep	separator, default is the TAB key.
bheader	a logical value indicating whether the input_filename_b file contains the names of the variables as its first line. By default, bheader = FALSE.

Value

list(file_a, file_b, file_a_b) returns a list with three pr-handled data.frames corresponding to the input_filename_a, input_filename_b file and a A JOIN B file.

Examples

```
data(demo_DMRfinder_rate_combine)
data(demo_DMRfinder_DMRs)
#example_tempfiles = tempfile(c("rate_combine", "DMRfinder_out"))
#tempdir()
#write.table(demo_DMRfinder_rate_combine, file=example_tempfiles[1],
#      row.names=FALSE, col.names=TRUE, quote=FALSE, sep='\t')
#write.table(demo_DMRfinder_DMRs, file=example_tempfiles[2],
#      sep ="\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
#result = metevalue.DMRfinder.chk(example_tempfiles[1], example_tempfiles[2],
#      bheader = TRUE)
```

---

metevalue.methylKit *Calculate E-value of the methylKit data format*

Description

Calculate E-value of the methylKit data format

Usage

```
metevalue.methylKit(
  methyrate,
  methylKit.output,
  adjust.methods = "BH",
  sep = "\t",
  bheader = FALSE
)
```

Arguments

`methyrate`	is the data of methylation rates of each sites and group, the columns are (in order): - chr: Chromosome - pos: int Position - g1~g2: methylation rate data in groups
`methylKit.output`	is the output data with e-value of each region - chr: Chromosome - start: The positions of the start sites of the corresponding region - end: The positions of the end sites of the corresponding region - strand: Strand - pvalue: The adjusted p-value based on BH method in MWU-test - qvalue: cutoff for qvalue of differential methylation statistic - methyl.diff: The difference between the group means of methylation level
`adjust.methods`	is the adjust methods of e-value. It can be 'bonferroni', 'hochberg', 'holm', 'hommel', 'BH', 'BY'
`sep`	seperator, default is the TAB key.
`bheader`	a logical value indicating whether the input_filename_b file contains the names of the variables as its first line. By default, bheader = FALSE.

Value

a dataframe, the columns are (in order):

- chr: Chromosome

- start: The positions of the start sites of the corresponding region

- end: The positions of the end sites of the corresponding region

- q-value: The adjusted p-value based on BH method in MWU-test

- methyl.diff: The difference between the group means of methylation level

- CpGs: The number of CpG sites within the corresponding region

- p : p-value based on MWU-test

- p2: p-value based on 2D KS-test

- m1: The absolute mean methylation level for the corresponding segment of group 1

- m2: The absolute mean methylation level for the corresponding segment of group 2

- e_value: The e-value of the corresponding region

Examples

```
data(demo_methylkit_methyrate)
data(demo_methylkit_met_all)
example_tempfiles = tempfile(c("rate_combine", "methylKit_DMR_raw"))
tempdir()
write.table(demo_methylkit_methyrate, file=example_tempfiles[1],
      row.names=FALSE, col.names=TRUE, quote=FALSE, sep='\t')
write.table(demo_methylkit_met_all, file=example_tempfiles[2],
      sep ="\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
result = metevalue.methylKit(example_tempfiles[1], example_tempfiles[2],
      bheader = TRUE)
str(result)
```

`metevalue.methylKit.chk` *Check the methylKit data format*

Description

Check the methylKit data format

Usage

```
metevalue.methylKit.chk(
  input_filename_a,
  input_filename_b,
  sep = "\t",
  bheader = FALSE
)
```

Arguments

`input_filename_a`	the combined data of methylation rate file. This file is a sep (e.g. TAB) separated file with two key columns and several value columns: For exampe: chr pos g1 g1 g1 g1 g1 g1 g1 g1 g2 g2 g2 g2 g2 g2 g2 g2 - chr and pos are keys; - g1~g2: methylation rate data in groups.
`input_filename_b`	the output file of methylKit. a methylDiff or methylDiffDB object containing the differential methylated locations satisfying the criteria. The columns are (in order): - chr: Chromosome - start: The position of the start sites of the corresponding region - end: The position of the end sites of the corresponding region - strand: Strand - p: p-value - qvalue: The adjusted p-value based on BH method - meth.diff : The difference between the group means of methylation level
`sep`	separator, default is the TAB key.
`bheader`	a logical value indicating whether the input_filename_b file contains the names of the variables as its first line. By default, bheader = FALSE.

Value

list(file_a, file_b, file_a_b) returns a list with three pr-handled data.frames corresponding to the input_filename_a, input_filename_b file and a A JOIN B file.

Examples

```
data(demo_methylkit_methyrate)
data(demo_methylkit_met_all)
example_tempfiles = tempfile(c("rate_combine", "methylKit_DMR_raw"))
tempdir()
write.table(demo_methylkit_methyrate, file=example_tempfiles[1],
      row.names=FALSE, col.names=TRUE, quote=FALSE, sep='\t')
write.table(demo_methylkit_met_all, file=example_tempfiles[2],
      sep ="\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
result = metevalue.methylKit.chk(example_tempfiles[1], example_tempfiles[2],
      bheader = TRUE)
```

---

metevalue.metilene *Calculate E-value of the Metilene data format*

Description

Calculate E-value of the Metilene data format

Usage

```
metevalue.metilene(
  methyrate,
  metilene.output,
  adjust.methods = "BH",
  sep = "\t",
  bheader = FALSE
)
```

Arguments

methyrate	metilene input file path. This file is a sep (e.g. TAB) separated file with two key columns and several value columns: For exampe: chr pos g1 g1 g1 g1 g1 g1 g1 g1 g2 g2 g2 g2 g2 g2 g2 g2 - chr and pos are keys; - g1~g2: methylation rate data in groups.
metilene.output	metilene input file path. This file should stored as a sep(e.g. TAB) separated file with two key columns and several value columns: The columns are (in order): - chr: Chromosome - start: The positions of the start sites of the corresponding region - end: The positions of the end sites of the corresponding region - q-value: The adjusted p-value based on BH method in MWU-test - methyl.diff: The difference between the group means of methylation level - CpGs: The number of CpG sites within the corresponding region

- p : p-value based on MWU-test

- p2: p-value based on 2D KS-test

- m1: The absolute mean methylation level for the corresponding segment of group 1

- m2: The absolute mean methylation level for the corresponding segment of group 2

`adjust.methods` is the adjust methods of e-value. It can be 'bonferroni', 'hochberg', 'holm', 'hommel', 'BH', 'BY'

`sep` seperator, default is the TAB key.

`bheader` a logical value indicating whether the metilene.output file contains the names of the variables as its first line. By default, bheader = FALSE.

Value

a dataframe, the columns are (in order):

- chr: Chromosome

- start: The positions of the start sites of the corresponding region

- end: The positions of the end sites of the corresponding region

- q-value: The adjusted p-value based on BH method in MWU-test

- methyl.diff: The difference between the group means of methylation level

- CpGs: The number of CpG sites within the corresponding region

- p : p-value based on MWU-test

- p2: p-value based on 2D KS-test

- m1: The absolute mean methylation level for the corresponding segment of group 1

- m2: The absolute mean methylation level for the corresponding segment of group 2

- e_value: The e-value of the corresponding region

Examples

```
#### metilene example ####'
data(demo_metilene_input)
data(demo_metilene_out)
#example_tempfiles = tempfile(c("metilene_input", "metilene_out"))
#tempdir()
#write.table(demo_metilene_input, file=example_tempfiles[1],
#      row.names=FALSE, col.names=TRUE, quote=FALSE, sep='\t')
#write.table(demo_metilene_out, file=example_tempfiles[2],
#      sep ="\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
#result = metevalue.metilene(example_tempfiles[1], example_tempfiles[2],
#      bheader = TRUE)
#head(result)
```

`metevalue.metilene.chk` *Check the Metilene data format*

Description

Check the Metilene data format

Usage

```
metevalue.metilene.chk(
  input_filename_a,
  input_filename_b,
  sep = "\t",
  bheader = FALSE
)
```

Arguments

`input_filename_a`	metilene input file path. This file is a sep (e.g. TAB) separated file with two key columns and several value columns: For exampe: chr pos g1 g1 g1 g1 g1 g1 g1 g1 g2 g2 g2 g2 g2 g2 g2 g2 - chr and pos are keys; - g1~g2: methylation rate data in groups.
`input_filename_b`	metilene input file path. This file should stored as a sep(e.g. TAB) separated file with two key columns and several value columns: The columns are (in order): - chr: Chromosome - start: The position of the start sites of the corresponding region - end: The position of the end sites of the corresponding region - q-value: The adjusted p-value based on BH method in MWU-test - methyl.diff: The difference between the group means of methylation level - CpGs: The number of CpG sites within the corresponding region - p : p-value based on MWU-test - p2: p-value based on 2D KS-test - m1: The absolute mean methylation level for the corresponding segment of group 1 - m2: The absolute mean methylation level for the corresponding segment of group 2
`sep`	separator, default is the TAB key.
`bheader`	a logical value indicating whether the input_filename_b file contains the names of the variables as its first line. By default, bheader = FALSE.

Value

list(file_a, file_b, file_a_b) returns a list with three pr-handled data.frames corresponding to the input_filename_a, input_filename_b file and a A JOIN B file.

Examples

```
#data(demo_metilene_input)
#data(demo_metilene_out)
#example_tempfiles = tempfile(c("metilene_input", "metilene_out"))
#tempdir()
#write.table(demo_metilene_input, file=example_tempfiles[1],
#      row.names=FALSE, col.names=TRUE, quote=FALSE, sep='\t')
#write.table(demo_metilene_out, file=example_tempfiles[2],
#      sep ="\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
#result = metevalue.metilene.chk(example_tempfiles[1], example_tempfiles[2],
#      bheader = TRUE)
```

`metevalue.RNA_general` *A general method to calculate the e-value for RNA-seq data.*

Description

A general method to calculate the e-value for RNA-seq data.

Usage

```
metevalue.RNA_general(rna, group1_name, group2_name)
```

Arguments

`rna`	data.frame: A data.frame object of RNAseq data. For example: treated1fb treated2fb untreated1fb untreated2fb TAG1 4.449648 4.750104 4.392285 4.497514 TAG2 8.241116 8.302852 8.318125 8.488796 ... Row names (TAG1 and TAG2 in the above example) is also suggested.
`group1_name`	charactor: The name of the first group. For example, "treated" in the example.
`group2_name`	charactor: The name of the second group. For example, "untreated" in the example.

Value

evalue

Examples

```
data("demo_desq_out")
evalue = metevalue.RNA_general(demo_desq_out, 'treated','untreated')
```

varevalue.single_general

A general method to calculate the e-value for other DNA methylation tools not described above. The input data is the DNA methylation rates using the same format with Metilene.

Description

The data file could be pre-handled by the metevalue.[types].chk function. The Chromosome name, start and end sits shoule be specified.

Usage

```
varevalue.single_general(
  methyrate,
  group1_name = "g1",
  group2_name = "g2",
  chr,
  start,
  end
)
```

Arguments

methyrate	data.frame: A data.frame object of methylation rates, the columns should be(name of groups can be self-defined) chr pos group1_name group1_name ... group1_name group2_name group2_name
group1_name	charactor: The name of the first group. For example, "treated" in the above example.
group2_name	charactor: The name of the second group. For example, "untreated" in the above example.
chr	charactor: The Chromosome name. Typically, it is a string like "chr21" and so on.
start	integer: The position of the start site of the corresponding region
end	integer: The position of the end site of the corresponding region

Value

evalue

Examples

```
#data("demo_metilene_input")
#varevalue.single_general(demo_metilene_input, chr = "chr21", start = 9437432, end = 9437540)
# [1] 2.626126e+43

#### Compare to `metevalue.metilene`  ####
data(demo_metilene_out)
#example_tempfiles = tempfile(c("metilene_input", "metilene_out"))
#tempdir()
```

```
#write.table(demo_metilene_input, file=example_tempfiles[1],
#      row.names=FALSE, col.names=TRUE, quote=FALSE, sep='\t')
#write.table (demo_metilene_out, file=example_tempfiles[2],
#      sep ="\t", row.names =FALSE, col.names =TRUE, quote =FALSE)
#result = metevalue.metilene(example_tempfiles[1], example_tempfiles[2],
#      bheader = TRUE)
# result[with(result, chr == 'chr21' & start == '9437432' & end == '9437540'), ncol(result)]
# [1] 2.626126e+43
```

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