

Figure 1:

- In the brain, the perforant pathway is a neuronal route from the entorhinal cortex to all the fields of the hippocampal formation, including the dentate gyrus, CA fields (cornu ammonis), and the subiculum. To detect visually activation of the neurons within this pathway the researchers use c-Fos labeling (red dot points in the various images of figure 1). The proto-oncogene c-Fos can be readily detected with immunofluorescence to map groups of neurons that display changes in their activity, and is used in the paper to show levels of neuronal activation in various regions of the hippocampus [1] [2].
- Also, In the *in-vivo* experiment illustrated by figure 1 and pictures B-I, the researchers want to show that unilateral optogenetically stimulated mice show levels of neuronal activation (green color), detected by release of extra-cellular  $\text{Ca}^{2+}$ , on the same side of the entorhinal cortex (ipsi-LEC or ipsi-DG) are higher compared to the levels of neuronal activation of the contra-side of the entorhinal cortex (contra-LEC and contra-DG). In addition, we can see that SSFO-infected, optogenetically induced mice compared to the EYFP-injected mice for the same regions of the hippocampus, show higher level of activation: activation in green (EYFP), images B and C vs. F and G, and the neuronal activation is particularly visible on image C. The researchers also point that non-stimulated neuron in LEC (ipsi or contra-LEC pictures E, G or I) still show some levels of activation detected by c-Fos labeling.

To perform this experiment, the researchers inject mice with either an ontogenetically induced activator SSFO and a genetic encoded protein EYFP or only the GEI EYFP; both under control of a  $\text{CaMKII}\alpha$  promoter ( $\text{Ca}^{2+}$ /calmodulin-dependent protein kinase II alpha) using an AAV vector described by picture A:

- One group of mice is injected with an adeno-associated virus (AAV) expressing SSFO-EYFP. Stable step-function opsin (SSFO) is a mutated version of channelrhodopsin (hChR2(C128SS)) that puts neurons in a state of excitability up to 30mins after activation by a pulse light. The last effect can be reversed by a brief pulse of a deactivation light (Figure A) [3].
- The other group is injected with an AAV vector with a similar design to the one used for the first group; the difference being that it does not contain the optogenetic activator SSO (Figure A).

Figure 2

- The entorhinal cortex is traditionally the first area affected in AD, and more specifically the lateral entorhinal cortex (LEC). Amyloid  $\beta$  ( $\text{A}\beta$ ) plaques are hallmarks of Alzheimer's disease (AD).  $\text{A}\beta_{40}$  and  $\text{A}\beta_{42}$  are the two main types of  $\text{A}\beta$  polymers which have direct role in plaque formation. Compared to  $\text{A}\beta_{40}$ ,  $\text{A}\beta_{42}$  is less abundant, highly insoluble but severely neurotoxic [4]. Using the same experiment described above (figure 1) the researchers measure in-vivo, every hour the average level of  $\text{A}\beta_{42}$ , detected using microdialysis technique, into the interstitial fluid (ISF) of the APP Tg mice (a transgenic mice that overexpresses APP) injected with the SSFO activator, 3 hours prior to optogenetic stimulation, up to 5 hours after the stimulation. The curve shows a rapid increase of the ISF  $\text{A}\beta_{42}$  within an hour after the stimulation, which slowly reaches a peak, 2 hours after optical stimulation, and thereafter slowly decreases, and back, 5 hours after the stimulation, but not quite, to the initial level at the time of the stimulation. Therefore, they show that the optogenetic stimulation neurons of the entorhinal cortex increases significantly the level of ISF  $\text{A}\beta_{42}$ . (About 23% increase) for a long period of time (5 hours) (Figure 2 – A).
- Also, the authors of the paper run a statistical analysis to test the hypothesis that ISF  $\text{A}\beta_{42}$  level in the hippocampus of SSFO-injected mice, mice with hippocampus optogenetically stimulated, were significantly higher at 1 hour of stimulation than that for the mice non-optogenetically induced (EYFP-injected). They use a t-test and obtain a p value less than their alpha level of 0.05 which imply they can accept this hypothesis (Figure 2 – B).
- Finally, the graph (Figure 2 – B) shows that no significant increase of % ISF  $\text{A}\beta_{42}$  of the AAV-EYFP light stimulated mice is observed compared to AAV-SSFO mice.

In conclusion, the authors of the paper want to show 1) the applications of optogenetic techniques in-vivo using APP Tg mice to study AD and 2) a causal relationship between hyper activation of the neurons in the perforant pathway and the increase of  $\text{A}\beta$  levels

## References

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