# The Aging Brain

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### **Key Words**

Alzheimer's disease, amyloid, DNA damage, memory, microarray, mitochondria, oxidative stress

#### **Abstract**

Aging is accompanied by cognitive decline in a major segment of the population and is the primary risk factor for Alzheimer's disease and other prevalent neurodegenerative disorders. Despite this central role in disease pathogenesis and morbidity, the aging of the brain has not been well understood at a molecular level. This review seeks to integrate what is known about age-related cognitive and neuroanatomical changes with recent advances in understanding basic molecular mechanisms that underlie aging. An important issue is how normal brain aging transitions to pathological aging, giving rise to neurodegenerative disorders. Toxic protein aggregates have been identified as potential contributory factors, including amyloid β-protein in Alzheimer's disease, tau in frontotemporal dementia, and α-synuclein in Parkinson's disease. However, current models of pathogenesis do not explain the origin of the common sporadic forms of these diseases or address the critical nexus between aging and disease. This review discusses potential approaches to unifying the systems biology of the aging brain with the pathogenesis of neurodegeneration.

AD: Alzheimer's disease

Cognitive decline:
age-related cognitive
decline may include
deficits in short-term
recall, spatial
memory, and
naming, whereas
neurodegenerative
disorders may
involve more global
and profound deficits
in memory,
language, reasoning,
and behavior

#### INTRODUCTION

Neurodegenerative diseases share a common predisposing factor, the aging of the brain, which poses several basic questions. First, how can a human neuron survive for 100 years or more and remain functionally competent? Second, do human neurons possess unique mechanisms for the repair of DNA and protein damage and protection against toxic free radicals? And finally, how and when do these quality control and repair systems break down? The answers to these questions are likely to provide new insights into the aging process and may suggest directions for treating neurodegenerative diseases. This review discusses recent advances in understanding the aging brain by integrating multiple levels of analysis, from neuropsychology and pathology to molecular genetics.

# COGNITIVE DECLINE IN THE AGING POPULATION

# Memory Loss and Altered Activation of the Prefrontal Cortex and Hippocampus

Cross-sectional studies suggest that delayed recall of verbal information declines significantly in the normal aging human population (1). In addition, working memory and shortterm recall, as well as the speed of processing information, gradually decline throughout the adult life span (2). A longitudinal study of the same individuals from 20 to 60 years of age showed that processing speed was the most affected modality (3), which may explain why normal aging individuals take longer to learn new information (4). Another feature of age-related memory loss that is highly conserved among mammalian species is reduced spatial memory, documented in aged humans (5), monkeys (6), dogs (7), and mice (8).

Long-term memory of life history and implicit memory, the unconscious response to previously encountered information, are well preserved through the normal aging process. Other age-stable measures of cognitive function include attention span, vocabulary, and verbal knowledge. Some cognitive processes may change or even improve with aging. For example, the emotional components of memories may receive greater emphasis in aged individuals than among young adults (9). Emotional stability, however, may improve with age, especially after age 65, possibly as a result of changing physiological responses of the medial prefrontal cortex (10).

Functional magnetic resonance imaging and positron emission tomography studies suggest that age-related memory changes may relate to altered functional activation of the prefrontal cortex and hippocampus. When presented with a task that involves executive function, regions of the prefrontal cortex activated in young adults typically exhibit reduced activation in aged adults (11, 12). In addition, aged adults often exhibit a broader area of activated prefrontal cortex and, in contrast to young adults, also activate the contralateral hemisphere—a phenomenon known as loss of hemispheric asymmetry. This may represent a normal compensatory response in the aging brain that is lost in mild cognitive impairment and Alzheimer's disease (AD) (13). Activation of the hippocampus is also reduced when healthy aged adults perform memoryrelated tasks. A cross-species study used functional magnetic resonance imaging to map hippocampal blood flow in aging rhesus monkeys, and in situ hybridization to map expression of Arc, an indicator of neuronal activity, in aging rats. This study suggested that the dentate gyrus was the hippocampal region most affected by aging in both species (14).

Normal age-related memory loss is distinguished from pathological memory loss by both the degree of impairment and the rate of cognitive decline. A structural correlate of pathological memory loss is volume loss in the medial temporal lobes, particularly the entorhinal cortex. This volume loss can appear at the earliest stages of mild cognitive impairment, progressing to severe atrophy in AD, but generally is not observed in normal aged individuals. In contrast, volume loss

in the prefrontal cortex can appear in normal aged individuals. There is increasing evidence, therefore, that altered brain activation on functional imaging and the appearance of early pathological changes in medial temporal lobe structures may distinguish incipient dementia from normal aging (15).

### Loss of Neural Circuits and Synaptic Plasticity

Early studies suggested that substantial neuronal loss occurs in the aging neocortex and hippocampus. However, stereological methods of neuronal quantification developed in the early 1990s showed that neuronal loss was not significant in most regions of the aging neocortex and hippocampus (16). Similarly, early reports also suggested significant loss of dendritic branching in the aging hippocampus. More recent studies, however, suggest that these early reports were confounded by the inclusion of both normal and dementia cases in the analysis, and that dendritic branching could actually increase in some hippocampal regions in aged individuals (17). In contrast, the aging prefrontal cortex shows variable changes in dendritic branching patterns (16).

Aging also affects white matter density, with the greatest reductions occurring in the prefrontal cortex and anterior corpus callosum (15, 18). Reduction in white matter density, as measured by diffusion tensor imaging, correlates with changes in executive function, short-term recall, and processing speed (19). It has been suggested that white matter pathology in the aging frontal cortex might compromise circuits that integrate the prefrontal cortex, hippocampus, and striatum (15).

Loss of synaptic function is likely a contributing factor in age-related cognitive decline. Age-related changes in dendritic spine and synapse number vary among different regions of the aging neocortex and hippocampus. Decreased synapse density has been observed in the frontal cortex of aged humans (20) and monkeys (21), and correlates with reduced activation of the prefrontal cortex when performing executive processing tasks. One of the most compelling examples of age-related synapse loss is in the hippocampal dentate gyrus in aged rats (22), a structural change that correlates with reduced excitatory post-synaptic potentials (23). Synapse loss in the dentate gyrus may also account for the spatial memory deficit in aged rats (24).

An important question is whether agerelated changes in cognitive function correlate with changes in long-term potentiation (LTP) and long-term depression, electrophysiological correlates of learning. Impaired induction and maintenance of LTP have been observed in a number of studies of the aged rat hippocampus, but the nature of the defect varies among the different hippocampal subregions (23–25). Aged rats also show increased susceptibility to the induction of long-term depression (26).

Synaptic plasticity is critically dependent upon the regulation of neuronal calcium fluxes and calcium-mediated signaling pathways. It has been proposed that altered calcium homeostasis in the aged brain might contribute to altered synaptic plasticity. Voltage-activated calcium influx is increased in hippocampal CA1 neurons from aged rats and relates to increased L-type calcium channels (27). In addition to changes in calcium channels, impaired intraneuronal calcium buffering capacity may increase cytoplasmic free calcium levels. Reduced immunoreactivity for calbindin 1, a major neuronal calcium-buffering protein, has been demonstrated in basal forebrain cholinergic and cortical neurons in aging humans and nonhuman primates (28). Moreover, reduced mRNA expression of calbindin 1 and 2, calcium channel subunits, and critical calcium signaling proteins, such as calmodulin 1, have been demonstrated in the aging prefrontal cortex (29). Altered gene expression may therefore affect calcium homeostasis and synaptic plasticity in the aging brain. Moreover, the age-related decline in expression of calbindin and other calcium-binding proteins

LTP: long-term potentiation

Stress response: a general category of biological mechanisms used by cells to protect against the damaging effects of toxic agents such as ROS

may render neurons more vulnerable to a variety of toxic insults mediated by calcium, such as excitotoxicity, contributing to neuronal loss in AD and other neurodegenerative disorders (28, 30, 31).

### GENE EXPRESSION AND THE SYSTEMS BIOLOGY OF BRAIN AGING

A growing number of microarray studies have been conducted to monitor genome-wide changes in gene expression associated with aging, particularly in the brain. Studies of Caenorhabditis elegans (32, 33) and Drosophila (34, 35) have examined the transcriptional effects of aging on the entire organism, whereas brain-specific studies have been conducted in mice (36, 37), rats (38), chimpanzees (39), and humans (29, 39, 40). Two primary themes arise from comparing the aging process in these six species (Table 1). First, age-related expression changes account for only a small fraction of the genes monitored in each of the six species. This observation lends support to the idea that specific biological pathways are being altered as a result of the aging process, as opposed to a genome-wide dysregulation of transcription. Second, although the underlying cause has not been identified, there is an age-associated induction of stress response genes that is common to all six species. Microarray studies in four out of the six species also detected a significant reduction in the expression of mitochondrial genes, suggesting that mitochondrial dysfunction may be a source of increased stress. Some of these studies were underpowered and therefore unable to detect all of the age-related expression changes. However, the fact that some biological changes, such as the increased stress response, are detected by all of the studies, despite differing experimental designs and analyses, suggests that these changes are robust (**Table 2**).

Microarray-based studies in *Drosophila* suggest that a large number of age-related gene expression changes can be reproduced

by exposing flies to oxidative stress (34). In addition, many of the single gene mutations found to increase the life span of various model organisms, including superoxide dismutase and p66she in Drosophila (41–44), are associated with an increased resistance to oxidative stress. Likewise, caloric restriction in mice extends life span together with increased oxidative stress resistance in the brain (45). Furthermore, a subset of agerelated gene expression changes in the aging mouse brain can be reversed by caloric restriction (36). Oxidative stress may therefore contribute to many of the gene expression changes associated with aging.

Transcriptional profiling of the aging human frontal cortex in a group of 30 individuals ranging from 26 to 106 years of age showed that approximately 4% of the genes expressed in the brain are age regulated (29). Age-related changes in gene expression became apparent in middle age and most pronounced after 70 years of age. Genes involved in synaptic functions that mediate memory and learning were significantly age downregulated, including glutamate receptor subunits, synaptic vesicle proteins, and members of the major signal transduction systems that mediate LTP. Notably, the synaptic calcium signaling system appears to be particularly affected with reduced expression of calmodulin 1 and 3, CAM kinase IIά and IV, calcineurin Bά, and multiple protein kinase C isoforms. Other prominent categories of age-downregulated genes include genes involved in vesicle-mediated protein transport and mitochondrial function. Genes involved in stress responses constitute the largest category of age-upregulated genes, including antioxidant defense, DNA repair, and immune function. These findings were confirmed in another study that showed a similar expression profile in several different cortical areas of the aging human brain (39). However, extracortical regions, such as cerebellum and caudate, showed different patterns. For cerebellum, this difference was due to fewer age-related changes, particularly in age-downregulated genes, rather than to a

Table 1 Age-related biological pathways

Caenorhabditis elegans	C. elegans and Drosphila*	D. melanogaster	Rattus norvegicus	Mus musculus	Homo sapiens
Whole organism (32)	Whole organism (33)	Thorax and abdomen	Hippocampus (CA1) (38)	Neocortex and cerebellum (36)	Prefrontal cortex (29)
Stress response genes Insulin signaling pathway Tc3 transposons Heat shock genes (up then down)	91 ± 2 Pa. 41 ± 1	Stress response genes (34, 35) Oxidative stress genes (34) Proteases (35) Mitochondrial genes (34, 35) Metabolism (34, 35) Reproduction (34)  sapiens (human) In troglodytes (chimpanzee) musculus (mouse) norvegicus (Norway rat)	Oxidative stress response Protein processing Inflammatory response Glial/structure Myelin-related proteins Metal ion homeostasis Growth/maintenance Signal transduction (Ca²+ related) Negative transcriptional regulation Mitochondrial and metabolism Synaptic/neurite plasticty Biosynthesis Signaling (extracellularly regulated) Extracellular matrix/structure Protein trafficking	Stress response genes Oxidative stress response Heat shock genes Proteases Inflammatory response Cathepsins Neural plasticity CNS development Hypothalamus (37)  Proteases Mitochondrial genes Stress response genes Neural plasticity Synaptic transmission ATPases  Cortex (37)  Proteases	Stress response genes DNA repair genes Inflammatory response Metal ion homeostasis Myelin-related proteins Synaptic function Vesicular transport Neuronal survival Mitochondrial function Amino acid modification Ca²+ homeostasis  Prefrontal cortex (40)  Glial enriched Immune response/cellular defense Growth factors Microtubule structure & function  Neuronal enriched Synaptic transmission
D. melanogaster (fruit fly)				Mitochondrial genes	Ca <sup>2+</sup> homeostasis
Induced Repressed	c.	elegans (nematode)		Stress response genes Neural plasticity Synaptic transmission ATPases	Voltage-gated ion channels G protein—coupled receptors Microtubule structure and function Kinase-phosphatase

Age-upregulated (red) and age-downregulated (blue) gene categories are shown for microarray studies of aging in species from *Caenorhabditis elegans* to man. The insert shows the phylogenetic relationship and estimated time of the last common ancestor between species in millions of years (green), as previously described (163).

Table 2 Experimental designs of microarray studies of aging

Species—Tissue	Platform	Sample size	Primary analysis	Reference
C. elegans—whole body	cDNA arrays	6 time points (3, 4, 6–7, 9–11, 12–14, 16–19 days)	One-way ANOVA	(32)
C. elegans—whole body	Spotted arrays	2 time points w/4 repeats each (0, 6 day adults)	Interspecies correlations of young-old	(33)
Drosophila—whole body	Affymetrix arrays	2 time points w/4 repeats each (3, 23 days)	Expression differences	(33)
Drosophila—thorax and abdomen	cDNA arrays	7 time points w/2 repeats each (3, 10, 15, 25, 30, 40, 50 days)	Fold change cutoff	(34)
Drosophila—whole body	Affymetrix arrays	6 time points w/5 repeats each (7, 18, 23, 28, 42, 47 days)	ANOVA and weighted regression	(35)
Rat—hippocampus (CA1)	Affymetrix arrays	3 time points w/10 repeats each (4, 14, 24 months)	One-way ANOVA and correlation analysis	(38)
Mouse—neocortex and cerebellum	Affymetrix arrays	2 time points w/3 repeats each (5, 30 months)	Fold change cutoff	(36)
Mouse—hypothalamus and cortex	Affymetrix arrays	2 time points w/2 repeats each (2, 22 months)	Fold change cutoff	(37)
Human—prefrontal cortex	Affymetrix arrays	30 samples (26–106 years)	Significance analysis of microarrays	(29)
Human—prefrontal cortex (BA9 and BA47)	Affymetrix arrays	39 samples (13–79 years)	Correlation and multifactorial analysis	(40)

qualitatively different pattern. A similar agerelated reduction in genes involved in synaptic function was reported for the aging rat hippocampus and was associated with cognitive impairment (38). These studies suggest that systems involved in higher-order cognitive functions may be compromised in the aging mammalian brain.

An important question is whether gene expression changes play a role in the susceptibility of the aging brain to neurodegenerative disorders such as AD. A microarray study of AD demonstrated a substantial number of expression changes that correlated with pathological markers and cognitive test scores; these included upregulation of signaling and tumor suppressor genes and downregulation of protein folding, metabolism, and energy-related genes (46). Profiling of several affected brain regions suggested that expression of the retromer trafficking gene *VPS35* correlated closely with the spatiotemporal pattern of AD (47). Cell culture studies using small inter-

fering RNAs to *VPS35* suggested that *VPS35* might regulate levels of the Aβ peptide.

Gene expression profiling can also be used to monitor the effects of therapeutic interventions in animal models. Microarray analysis of a transgenic mouse model expressing AD-linked amyloid precursor protein (APP) and presenilin-1 variants identified specific changes associated with placing the mice in an enriched environment, including upregulation of genes involved in synaptic plasticity, neurogenesis, neuronal survival, and Aβ degradation (48). These expression changes correlated with reduced cortical Aß levels and amyloid deposits. Another study showed that pharmacologic inhibitors of histone deacetylases, which activate silenced genes, restore memory function and induce synapse formation in the p25/cdk5 mouse model of neurodegeneration (49). There is increasing evidence, therefore, that neurodegeneration and cognitive decline may be associated with specific changes in gene expression.

**APP:** amyloid precursor protein

#### **DNA DAMAGE**

# DNA Repair and Accelerated Aging Syndromes

The association of human syndromes of accelerated aging with inherited mutations in DNA repair genes strongly implicates DNA damage in the human aging process. These disorders, known as segmental progeroid syndromes, are characterized by accelerated onset of a subset of human aging phenotypes that frequently include neurodegeneration (50). Mutations in genes involved in singleor double-strand DNA break repair result in cerebellar degenerative syndromes known as ataxias, which are manifested by movement disorders. The continued proliferation of cerebellar granule cells during postnatal development may underlie the vulnerability of the cerebellum to inherited deficits in genome stability. In contrast, inherited mutations in DNA helicases, such as Werner and Rothmund-Thomson syndromes, give rise to features of accelerated aging that often do not include nervous system dysfunction. This may reflect the role of RecQ-like helicases in recombinant events in replicating cells. Inherited mutations in enzymes involved in nucleotide and base excision repair, including xeroderma pigmentosum and Cockayne syndrome, are characterized by accelerated aging phenotypes that include neurodegeneration, mental retardation, and delayed psychomotor development (50). A new human progeroid syndrome that is caused by a loss of function mutation in the XPF-ERCC1 endonuclease that repairs helix-distorting DNA lesions was recently described. Mice deficient in ERCC1 recapitulate the progeroid features and exhibit a gene expression profile in the liver that overlaps with that of normal aging mice (correlation coefficient 0.32), suggesting that this type of DNA damage may contribute to the aging process (51). Segmental progerias typically have a short life span of less than 20 years, which may account for the absence of Alzheimer-type neuropathological

changes. However, individuals with Werner syndrome, a longer-lived progeroid syndrome, can have variable neuropathology, with one 57-year-old case reportedly showing unusually high levels of amyloid β-protein deposition in the brain (52).

# Consequences of Unrepaired DNA Damage

The role of DNA double-strand breaks (DSBs) in the aging of the brain is just beginning to be explored. DSBs are repaired by two major pathways: homologous recombination, which occurs during DNA replication, and nonhomologous end joining (NHEI), the predominant pathway in postmitotic cells such as neurons. NHEJ is mediated by a number of core factors, four of which are highly conserved from yeast to mammals, including Ku80, Ku70, Ligase IV, and XRCC4 (53). Targeted knockouts of the NHEJ factors in mice result in embryonic lethality, genomic instability, and apoptosis of neurons in the brain soon after postmitotic differentiation. Apoptosis was prevented by reducing p53 expression, suggesting that it is a manifestation of the DNA damage response (54). A broader role for p53 in neuronal aging was suggested by selectively reducing p53 activity in *Drosophila* neurons, which increased life span and resistance to oxidative stress (55).

The predominant class of oxidative DNA lesions in the aging brain are single base modifications, such as 8-oxoguanine, which increase with aging in rodent models and humans (29, 56–58). In addition, oxidative base damage to DNA and RNA is increased further in AD (59, 60). A dynamic role of DNA damage in the transcriptional regulation of genes in the aging brain was suggested by a recent study showing that oxidative DNA damage accumulates in the promotors of a subset of age-downregulated genes (29). Reduced transcription of genes involved in synaptic function, protein transport, and mitochondrial function occurred together with DNA

Double-strand break (DSB): a severe form of DNA damage involving scission of both DNA strands, usually induced by ionizing radiation or ROS

**NHEJ:** nonhomologous end joining

damage to these genes starting in middle age. The vulnerability of particular genes to DNA damage was recapitulated in cultures of primary neurons and neuroblastoma cells subjected to a mild oxidative stress (29). Another study also provided evidence that particular regions of the genome may be selectively vulnerable to oxidative DNA damage by showing that in situ staining of 8-oxoguanine is concentrated in chromosomal regions corresponding to recombination hot spots (61). Previous studies using chromatographic methods had normalized 8-oxoguanine levels to the total content of genomic DNA, concluding that median values of 0.5–4.0 oxoguanines per megabase of DNA appeared in cells (62). The implicit assumption in these measurements was that oxidative damage is randomly distributed

throughout the genome. It will be important to confirm, therefore, whether oxidative DNA damage is nonrandom, resulting in regions of the genome with concentrated damage.

Transcriptional repression may be a mechanism whereby aging neurons can silence damaged regions of the genome, enabling them to survive in the presence of unrepaired DNA damage (Figure 1). In dividing cells, unrepaired DNA damage predisposes to neoplastic transformation. Hence, mammalian cells have evolved to remove persistently damaged cells, particularly those with unrepaired DSBs, through p53-mediated apoptosis. Postmitotic cells, however, do not undergo neoplastic transformation. Hence, it may be advantageous for the aging brain to prevent the apoptosis of irreplaceable neurons, even in the

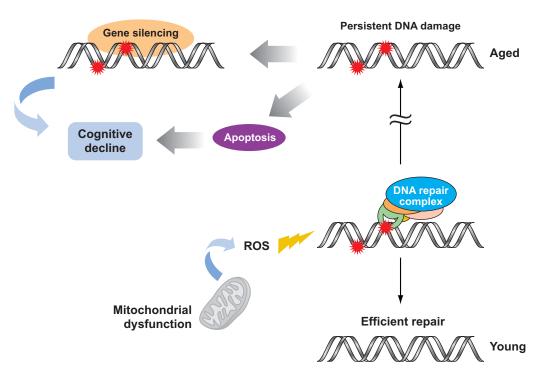


Figure 1

DNA damage and brain aging. Oxidative damage of DNA may be mediated by reactive oxygen species (ROS) derived from aging mitochondria. DNA damage is repaired efficiently in the young adult brain, but persists in the aged brain. During normal aging, this may result in the silencing of genes involved in synaptic plasticity, mitochondrial function, and protein trafficking, potentially contributing to cognitive decline. In neurodegenerative diseases, DNA damage may additionally compromise neuronal survival.

presence of unrepaired DNA damage. It will be important, therefore, to achieve a greater understanding of the regulation of the DNA damage response in the brain.

## MITOCHONDRIAL DYSFUNCTION

A substantial body of evidence suggests that progressive degeneration and dysfunction of mitochondria contribute to the aging process and in particular the aging of postmitotic tissues such as brain and muscle (Figure 2). The projection neurons of the cerebral cortex, which degenerate in Alzheimer's dis-

ease, are highly dependent upon mitochondrial oxidative phosphorylation to support energy-intensive ion fluxes and axonal transport across long distances in the brain. Hence, this neuronal population is also quite vulnerable to mitochondrial dysfunction. Two major sites of mitochondrial damage during aging are the respiratory chain enzymes and mitochondrial DNA (63).

# Generation of Reactive Oxygen Species

Inefficient electron transport through the mitochondrial respiratory complexes can lead

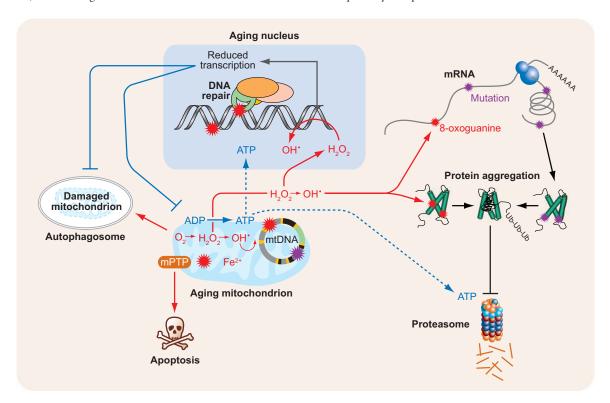


Figure 2

Global impact of mitochondrial aging. In the aging brain, reduced autophagic clearance of degenerating mitochondria and increased mitochondrial DNA (mtDNA) damage may reduce ATP levels and elevate the level of reactive oxygen species. Reactive oxygen species can further damage nuclear and mitochondrial DNA, resulting in reduced transcription, and damage RNA and protein, giving rise to protein misfolding and aggregation. Aggregated proteins may accumulate in the aging brain as a result of inefficient clearance through the autophagic and ubiquitin-proteasome pathways. Toxic protein aggregates may also lower the mitochondrial permeability transition pore (mPTP) threshold for inducing apoptosis.

Reactive oxygen species (ROS): free radicals derived from oxygen that damage macromolecules by introducing unpaired electrons

to reduced ATP synthesis and the generation of superoxide radical as a byproduct. Mitochondria are protected against endogenously generated reactive oxygen species (ROS) by a number of antioxidant defenses, including manganese superoxide dismutase, periredoxins, and redox reactions mediated by cytochrome C and cytochrome oxidase. However, there is considerable evidence that these defenses are overcome during aging, resulting in local oxidative damage to mitochondrial proteins and DNA. In addition, hydrogen peroxide, generated by the action of superoxide dismutase on superoxide radicals in mitochondria, is a stable molecule that can diffuse to other parts of the cell. Upon interaction with transition metals, hydrogen peroxide is converted by the Fenton reaction to the highly reactive and damaging hydroxyl radical. The destructive potential of this oxidative pathway is controlled through enzymatic deactivation of hydrogen peroxide by cytoplasmic glutathione peroxidase or peroxisomal catalase.

In addition to the generation of superoxide and hydrogen peroxide, the availability of redox-active iron is a major determinant of ROS-mediated cellular damage. Elevated levels of redox-active iron accumulate in the normal aging brain and in several neurodegenerative diseases (64), and may be derived in part from degenerating mitochondria. Iron is highly concentrated in mitochondria, where it is sequestered in heme and in iron sulfur clusters that serve as cofactors for respiratory enzymes. Recent studies suggest that damaged mitochondria are removed and degraded by the autophagic pathway (65). Autophagic function may be compromised in the aging human brain by reduced expression of beclin-1, a key regulator of autophagy (66), and other autophagy-related genes (T. Lu & B.A. Yankner, unpublished results). This may, in turn, lead to the accumulation of dysfunctional or degenerating mitochondria, resulting in increased ROS generation and the release of redox-active iron.

#### Mitochondrial DNA Mutations

A potentially important mechanism of agerelated mitochondrial damage is mutation of the mitochondrial DNA. Inherited mitochondrial DNA mutations give rise to multisystem disorders that predominantly affect brain and muscle (67, 68). Furthermore, somatic mitochondrial mutations appear in the normal aging brain and in AD, together with increased oxidative DNA damage, and may be related to the proximity of mitochondrial DNA to ROS-generating respiratory enzymes and the absence of protective histones (29, 56, 59, 69– 71). Decreased mitochondrial base excision repair in the aging brain may also be contributory (72). Mitochondrial DNA mutations are not randomly distributed, but rather accumulate in control regions where they impair the transcription and replication of mitochondrial DNA (71) and reduce the activity of respiratory chain enzymes (70).

To obtain greater insight into the role of mitochondrial DNA mutations in aging, two groups recently created knock-in mice expressing a mutated version of mitochondrial DNA polymerase  $\gamma$  (POLG) that was deficient in DNA proofreading but retained DNA polymerase activity. Mice with proofreadingdeficient POLG accumulated high levels of mitochondrial DNA mutations and showed reduced respiratory chain activity (73, 74). These mutator mice were initially normal, but by 6–9 months of age showed signs of accelerated aging, including weight loss, alopecia, kyphosis, osteoporosis, anemia, cardiomegaly, and shortened life span. This was accompanied by markers of apoptosis in a variety of tissues, including activated caspase-3 and DNA fragmentation (74). However, there was a distinct absence of markers of oxidative stress or damage, suggesting that the accelerated aging phenotype might be related to increased apoptotic cell death but not to oxidative stress. This surprising result is at odds with the widely held view that mitochondrial mutations result in respiratory chain dysfunction, which causes cellular

damage through elevated ROS generation. Support for this view was provided by another study in which transgenic mouse lines were created that selectively targeted human catalase to mitochondria, peroxisomes, or the nucleus (75). Catalase targeted to mitochondria extended mouse life span by  $\sim 20\%$  and also reduced age-related cardiac pathology and cataract development. Targeting catalase to the nucleus had no effect, although it was unclear whether this manipulation effectively reduced oxidative DNA damage. The mutator mouse and catalase targeting studies did not report any adverse effects on the nervous system, although it was unclear whether detailed neuropathological or behavioral studies were performed.

The expression of multiple nuclearencoded mitochondrial genes declines in the aging brain, suggesting a potential nuclear contribution to age-related mitochondrial dysfunction (29, 38, 39). Reducing the expression of one of these genes, the  $\alpha$  subunit of the F1-ATP synthase, by using small interfering RNA in neuroblastoma SH-SY5Y cells, reduced mitochondrial ATP synthesis and led to nuclear DNA damage (29). DNA damage was partially reversed by vitamin E, suggesting a role for ROS generation. These observations suggest that impaired mitochondrial function can lead to nuclear DNA damage that may, in turn, reduce the expression of nuclear-encoded mitochondrial genes, setting up a deleterious feedback loop in the aging brain.

### PATHOGENESIS OF NEURODEGENERATION

## The Pathology of Alzheimer's Disease

Since the identification of neurofibrillary tangles (NFTs) and amyloid plaques as pathological hallmarks of Alzheimer's disease, the role of these lesions in the pathogenesis of dementia has been debated. Early studies suggested that amyloid plaques correlated with cognitive decline (76). Subsequent studies, however, suggested that plaques were not the closest correlate and that NFT number and synapse loss correlated more closely with cognitive test scores in individuals with dementia (77). With the advent of stereological methods for neuronal quantification, it has become apparent that neuronal loss, particularly in the CA1 field of the hippocampus and layer 2 of entorhinal cortex, distinguishes normal aging from the earliest stages of cognitive decline and Alzheimer's disease (Figure 3) (78–80). Although synapse loss also parallels cognitive decline (81), this may in part be secondary to neuronal loss. In particular, synapse loss in the dentate gyrus of the hippocampus, an

**NFT:** neurofibrillary tangle

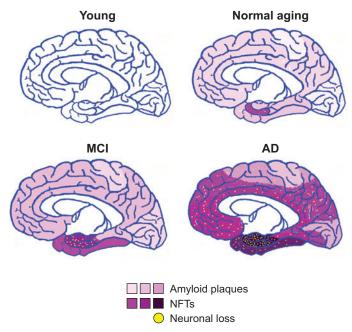


Figure 3

Progression of neuropathology in aging and Alzheimer's disease. Shown is the neuroanatomical distribution of amyloid plaques, neurofibrillary tangles (NFTs), and neuronal loss during normal aging, mild cognitive impairment (MCI), and Alzheimer's disease (AD). In cognitively intact aging individuals, amyloid plaques can appear in the neocortex and hippocampus, whereas NFTs are localized predominantly to the entorhinal cortex. MCI is marked by the appearance of neuronal loss in layer 2 of the entorhinal cortex and the CA1 region of the hippocampus, and is often accompanied by an increase in the number and distribution of plaques and NFTs. Plaques and NFTs are generally more widespread in AD, although this is variable. However, the extent of neuronal and synaptic loss correlates with dementia.

**apoE:** apolipoprotein E

early site of pathology, may be secondary to loss of afferent projection neurons in layer 2 of the entorhinal cortex (82). Taken together, these studies suggest that amyloid deposits and NFTs may not be sufficient to cause dementia, but may need to be accompanied by an as yet unidentified process that leads to neuronal cell death and the loss of synaptic connections.

#### Genetics of Alzheimer's Disease

Major advances in our understanding of AD and other age-related neurodegenerative disorders have come from the identification of inherited disease-causing mutations. The first genetic clue to the etiology of AD came from the observation that individuals with Down syndrome (trisomy 21) have a very high incidence of AD with early onset (83). Hence, it was suspected that a pathogenic gene was present on chromosome 21. The localization of the APP gene to chromosome 21 (84) suggested that APP might be a site of diseasecausing mutations. The first disease-causing mutation in APP was identified within the amyloid β-protein domain of APP in patients with a disorder known as hereditary cerebral hemorrhage with amyloidosis, Dutch type (HCHWA-D) (85, 86). In contrast to AD, this disorder is characterized by the deposition of amyloid β-protein predominantly in blood vessel walls, resulting in mortality from multiple cerebral hemorrhages. In 1992, the first APP mutation associated with familial AD was identified (87). This mutation and others were located proximate to the C-terminal end of the Aβ peptide and altered its proteolytic generation such that the relative amount of a slightly longer form of Aβ (Aβ 42) was increased (88). This longer form of the Aß peptide has a greater propensity to aggregate, potentially accounting for the pathogenicity of these APP mutations (89). Another APP mutation was identified proximate to the N terminus of Aß that increased Aß generation by augmenting the β-secretase cleavage (90, 91).

The second disease-causing gene identified in families with autosomal dominant inheritance of AD was presentlin-1 on chromosome 14 (92). Disease-causing mutations span the presenilin-1 gene and are the most common known cause of early-onset autosomal dominant AD. Mutations in a homologous gene, presenilin-2, account for a much smaller number of cases (93–96). Presenilin-1 and -2 are part of the  $\gamma$ -secretase complex that mediates the intramembrane proteolysis of APP that liberates the C terminus of the Aß peptide (97–99). γ-Secretase is a novel complex comprising four core components: presenilin, nicastrin, Aph-1, and Pen-2. Each of these is required for enzymatic activity (100). Disease-causing mutations in presentilin-1 or -2 increase the level of the longer Aβ42 peptide relative to the shorter A $\beta$ 40 peptide (101). When these pathogenic variants of APP and presenilin-1 are overexpressed in transgenic mice, they result in age-dependent formation of amyloid deposits, providing support for the idea that altered APP metabolism can give rise to an early-onset form of AD (102, 103).

The only gene that has been unequivocally linked to the prevalent late-onset form of AD is apolipoprotein E (apoE) (104). The E4 allele of apoE is a significant risk factor for the development of AD, increasing the risk by as much as 10–12-fold when present in the homozygous state (105). In contrast, another variant, the E-2 allele, protects against the development of AD. The presence of an apoE4 allele correlates with increased A\beta deposition in the human brain, and apoE4 induces Aβ fibrillization in APP transgenic mice (106). In addition, apoE may have neurotrophic and antioxidant activity and may facilitate recovery after a variety of neuronal insults (107). However, the mechanism by which apoE4 increases the risk of AD remains an active area of investigation.

### The Amyloid Hypothesis

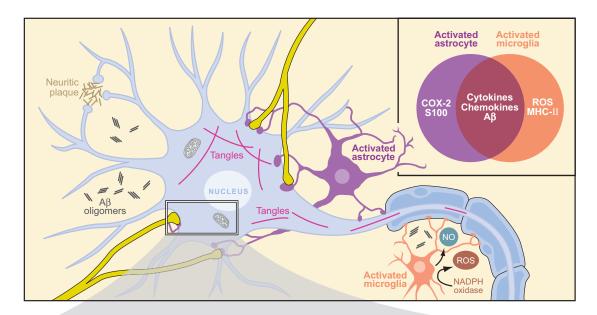
The initial studies of  $A\beta$  in cell culture showed that aggregated forms of the peptide

are neurotoxic, suggesting that Aβ could be a proximate cause of neurodegeneration in AD (108–110). In contrast, early analysis of transgenic mouse models that overexpressed mutant forms of APP that cause familial AD and develop AB deposits did not show widespread cytoskeletal pathology or neuronal loss (102, 111). A subsequent study, however, using confocal microscopy with a sophisticated cross-correlation mapping of neuronal density showed that significant neuronal loss occurred in direct contiguity to fibrillar thioflavin S-positive Aβ deposits (112). Nonetheless, these mouse models did not demonstrate the widespread neuronal loss that typically occurs in AD. A potential explanation came from studies of fibrillar AB injection into the brains of aging nonhuman primates. When a plaque-equivalent concentration of fibrillar A\beta was injected into the brains of aged but not young rhesus monkeys, it resulted in substantial neuronal degeneration, tau pathology, and microgliosis (113). However, injection of AB into the aging rat brain had no detectable pathological effects at a plaque-equivalent concentration. These results suggest that aging renders the brain vulnerable to Aβ toxicity and that a species barrier may reduce A\beta toxicity in the aging rodent brain.

More recent studies have focused on the toxic effects of lower molecular weight oligomeric forms of A\beta that may be more toxic than larger fibrillar aggregates (114). The initial observation of oligomer toxicity was made by Klein and associates, who showed that small diffusible A\beta oligomers referred to as ADDLs killed mature neurons in organotypic cultures at nanomolar concentrations. In addition, these oligomeric Aβ peptides inhibited hippocampal LTP, raising the possibility that they could impair synaptic plasticity and memory (115). Subsequent studies showed that smaller dimeric and trimeric forms of Aß synthesized by cells in culture could also impair LTP and learned behavior following intracerebral injection in the rat (116, 117). However, in contrast to the larger ADDL-type oligomers, there is no evidence that the smaller Aβ dimers or trimers can induce synaptic or neuronal loss. A recent study correlated the appearance of a larger 56-kDa Aβ oligomer with cognitive deficits in the Tg2576 transgenic mouse model of AD (118). Isolation of this oligomeric Aβ species from the transgenic mouse brain and infusion into the brains of rats resulted in a transient deficit in memory retention. These studies raise the possibility that particular oligomeric species may impair synaptic plasticity and learning. It will be important to determine whether these Aß species are present at functionally significant levels in the brains of AD patients, as Aβ oligomers are unstable and rapidly aggregate to higher molecular weight forms.

The toxic effects of A\beta have been associated with a variety of cellular mechanisms, including activation of caspases and calpain, stimulation of microglial inflammatory pathways, and compromise of the vascular endothelium (Figure 4) (110, 119–121). Recent studies suggest that AB can also alter glutamatergic neurotransmission by promoting the endocytosis of NMDA and AMPA receptors (122, 123). The clinical efficacy of the drug memantine, an NMDA receptor antagonist, suggests a role for altered glutamatergic neurotransmission in AD (124). Another potentially important mechanism of Aß toxicity is the activation of cell cycle reentry in postmitotic neurons. Aberrant cell cycle activation and DNA synthesis have been detected in neurons in the brains of patients with mild cognitive impairment and AD (125). Moreover, cell cycle reentry occurs when neurons are exposed to toxic doses of Aβ in culture (126).

The role of  $A\beta$  as the sole or dominant cause of cognitive decline in AD has recently been questioned by a study in which a mutation in the cytoplasmic domain of APP (Asp-664) was knocked into APP transgenic mice. This mutation did not alter total  $A\beta$  levels or plaque formation, but prevented the degenerative features of the transgenic model, including synaptic loss, astrogliosis, and cognitive



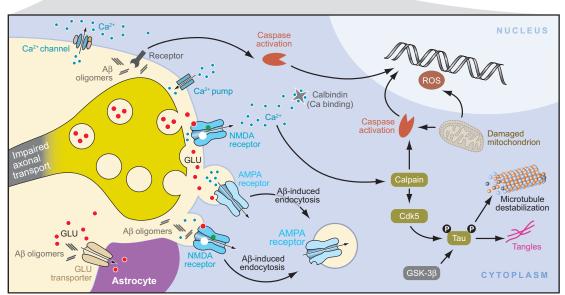


Figure 4

Neurodegenerative mechanisms in Alzheimer's disease. (*Upper panel*) Abnormal accumulation of  $A\beta$  and tau as oligomers, neuritic plaques, and neurofibrillary tangles may impair neuronal function. In addition,  $A\beta$  aggregates can induce the proliferation and activation of astrocytes and microglia, leading to the elaboration of neurotoxic cytokines and reactive oxygen species (ROS). (*Lower panel*) The inset in the upper panel is expanded in the lower panel to show mechanisms of synaptic dysfunction, including  $A\beta$ -induced endocytosis of AMPA and NMDA receptors, increased calcium influx through L-type calcium channels, and impaired glutamate (GLU) reuptake by astrocytes. In addition,  $A\beta$  aggregates can activate caspases through several pathways, including cell death receptors (FAS and the p75 NGFR), calpain activation and mitochondrial damage, leading to neuronal apoptosis. Hyperphosphorylation and aggregation of tau can destabilize microtubules and impair axonal transport, compromising synaptic function and giving rise to neurofibrillary tangles.

deficits (127). One potential model is that the mutation prevents the proteolytic release of a toxic APP C-terminal fragment (128–131). Another possibility is that altered regulation of the biological functions of the APP holoprotein may contribute to the neurodegenerative process.

### Tau Mutations and Neurodegeneration

Neurofibrillary tangles (NFTs) are a characteristic feature of the normal aging human brain that can become widespread in hippocampal, neocortical, and limbic neurons in AD. NFTs are composed predominantly of posttranslationally modified forms of tau, a microtubule-associated protein. Phosphorylation of tau at multiple sites is associated with its dissociation from microtubules and aggregation into oligomeric and fibrillar forms. This may destabilize microtubules and impair antegrade axonal transport of essential macromolecules and organelles to synaptic endings. Although tau-related pathology is characteristic of AD, the role of tau in neurodegeneration was firmly established in another disorder, frontotemporal dementia with Parkinsonism linked to chromosome 17 (FTDP-17). Frontotemporal dementias constitute a spectrum of clinical disorders characterized by the onset of a dementing illness in which language and behavioral deficits are the presenting symptoms rather than deficits in episodic memory characteristic of AD. At the histological level, these disorders are characterized by widespread neuronal cell death in the frontal and temporal lobes and the appearance of a variety of aggregated forms of tau, ranging from oligomeric forms to NFTs (132). Mutations in two genes, tau and progranulin, have been identified as causes of familial FTDP-17 (133-135). Tau mutations may act through three pathologic mechanisms: to alter splicing leading to abnormal gene expression, to alter the ability of tau to stabilize microtubules, or to increase tau protein aggregation. The expression of the P301L tau mutation in

transgenic mice recapitulates major features of human FTDP-17, including the age-dependent appearance of NFTs, neuronal loss, and behavioral deficits (136, 137).

Overexpression of AD-causing APP or presenilin mutations in transgenic mice does not give rise to the formation of NFTs or the extensive tau pathology characteristic of AD (102, 103). However, creation of a triple transgenic mouse that overexpressed pathogenic variants of APP, presenilin-1, and tau gave rise to progressive formation of amyloid plaques and NFTs, as well as synaptic dysfunction (138). Synaptic dysfunction appeared at four months of age and correlated with hippocampal accumulation of intraneuronal Aβ, but preceded the appearance of plaques and tangles. Treatment of these animals with an Aβ antibody improved cognitive function only under circumstances in which both soluble A\beta and tau were reduced, suggesting the importance of an A\beta-tau interaction (139). This idea was supported by crossing another APP transgenic mouse model with tau-deficient mice, which substantially improved Aβ-related deficits in spatial memory (140). Cognitive improvement was not related to changes in tau phosphorylation or neuritic dystrophy, but correlated with protection against excitotoxicity. It was suggested that tau might contribute to neuronal dysfunction and toxicity by modulating sensitivity to excitatory neurotransmission.

# PREVENTION OF COGNITIVE DECLINE AND NEURODEGENERATION

The prevalent comorbid medical conditions of aging, including hypertension, hyperlipidemia, and diabetes, are major contributing factors for stroke and white matter ischemic changes during aging, and may also predispose to AD. Hence, management of these conditions through medical and lifestyle interventions is likely to benefit cognitive function (141). A substantial body of literature suggests that physical exercise not only

### FTDP-17: frontotemporal dementia with Parkinsonism linked

to chromosome 17

Sirtuins: a family of proteins related to the histone deacetylase SIR2 that regulates longevity in yeast

reduces vascular disease, but may also directly benefit cognitive function through a number of mechanisms, including increased production of brain-derived neurotrophic factor and increased neurogenesis (142, 143). Aerobic exercise increased cerebral blood volume in the dentate gyrus of the human hippocampus in a study of 11 subjects ranging in age from 21–45 years (144). This was accompanied by improvement in immediate recall on cognitive testing. Another study showed that long-term physical activity was associated with improved cognitive function in a large population of women ranging in age from 70–81 years (145). An important question, however, is whether exercise can prevent or slow the onset of neurodegenerative diseases. In a transgenic mouse model of AD, increased exercise on a running wheel was associated with reduced amyloid deposition and improved cognitive function (48). Other studies suggest that placing mice in an enriched, stimulating environment may have a greater effect on spatial memory than exercise alone (146).

A potentially important therapeutic approach to aging is the prevention of oxidative damage to DNA and other macromolecules. However, there is surprisingly little evidence yet for the efficacy of this approach in the human population. This may relate to the fact that free radical scavengers act stoichiometrically on their targets, requiring high doses to significantly impact oxidative stress in the brain. Epidemiological studies suggest that dietary intake of vitamin E might reduce the incidence of AD (147). In addition, treatment of AD patients with high doses of vitamin E or selegiline, a selective monoamine oxidase inhibitor, had a modest effect on functional indices of cognitive decline (148). The evidence for this approach is more compelling in aged rats that show mitochondrial degeneration and oxidative damage to DNA and RNA in the brain. Feeding aged rats the mitochondrial metabolite acetyl-L-carnitine and the antioxidant lipoic acid improved spatial memory and prevented structural decay of mitochondria in the hippocampus (58). Similarly, administration of coenzyme  $Q_{10}$ , a cofactor of the mitochondrial electron transport chain and an antioxidant, showed neuroprotective effects in rats (149). Initial studies in patients with Parkinson's disease also suggest that coenzyme  $Q_{10}$  may reduce the rate of functional decline (150).

Increased inflammation is a feature of brain aging that is conserved from mouse to man (Table 1). Epidemiologic studies suggested that individuals treated with nonsteroidal anti-inflammatory agents for arthritis may have a lower incidence of AD (151). This observation led to clinical trials of cyclooxygenase-2 inhibitors in patients with AD that did not demonstrate efficacy (152). Supplementation with dietary omega-3 fatty acids has received considerable attention because of their positive vascular and antiinflammatory effects. Epidemiologic studies show that individuals with a high dietary intake of fish and, in particular, the omega-3 fatty acid docosahexaenoic acid, have a reduced incidence of AD (153). Moreover, treatment of transgenic mouse models of AD with omega-3 fatty acids can improve cognitive function and reduce AB levels and deposition (154, 155).

Caloric restriction has been shown to significantly increase life span and promote resistance to a broad range of age-related pathology in worms, flies, and mice. Some of the effects of caloric restriction may be mediated through the sirtuin family of genes, as exemplified by SIR2, which prolongs life span in veast (156). It was found that resveratrol, a naturally occurring compound in grapes and red wine, can activate sirtuins and increase longevity in yeast, worms, and flies (157, 158). Moreover, treatment with high doses of resveratrol can significantly increase the survival of mice on a high-fat diet and increase insulin sensitivity (159, 160). The role of sirtuins in the brain is still relatively unexplored. It has been reported that SIRT1, the mammalian ortholog of the yeast SIR2 gene, is involved in a biosynthetic pathway for NAD that

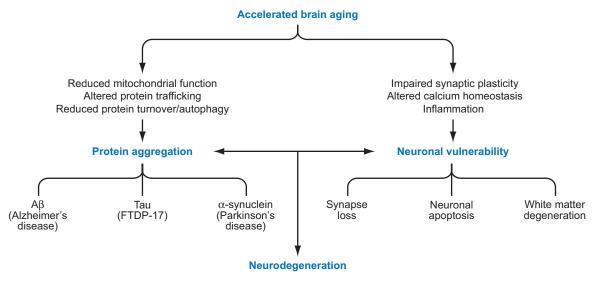


Figure 5

Neurodegeneration and accelerated brain aging. Genetic or environmental factors that accelerate underlying mechanisms of brain aging may affect biological systems that regulate protein aggregation and neuronal survival. Pathological protein aggregation and age-dependent neuronal vulnerability may act together to induce neurodegeneration.

protects axons from degeneration in a mouse model known as Wallerian degeneration slow (161). It remains to be determined whether activation of sirtuins can protect against cognitive decline and neurodegeneration, possibly by activating global anti-apoptotic and stress resistance pathways.

#### CONCLUSION

The question of how the brain ages at a molecular level is complex and has recently been addressed by a systems biology approach. Genome-scale transcriptional profiling studies in multiple species suggest that a conserved set of biological systems involved in synaptic function, mitochondrial energy metabolism, and stress resistance change in the aging brain (Table 1). A subset of agerelated transcriptional changes may be caused by DNA damage and genomic instability, an established etiologic factor in cancer and accelerated aging syndromes (Figure 1). There is also increasing evidence for a role of mitochondrial dysfunction as a source of oxidative

stress and neuronal dysfunction in the aging brain. Moreover, feeding rats mitochondrial metabolites can partially reverse age-related cognitive decline (58). Hence, nuclear and mitochondrial dysfunction may coordinately alter the systems biology of the brain, leading to a spectrum of age-related cognitive changes (Figure 1 and Figure 2).

A central issue is the relationship of normal to pathological aging and the mechanisms that underlie this transition. The risk of AD increases 14-fold from age 65-85, afflicting as much as 47% of individuals over age 85 (162). The onset of AD is therefore closely associated with the aging process. Moreover, normal age-related changes appear to be increased in AD at several levels, including the spread of plaque and tangle pathology (**Figure 3**), changes in gene expression, and exaggerated age-related changes in autophagy, mitochondrial function, and protein trafficking (Figure 5). What appears to distinguish AD, frontotemporal dementia, and Parkinson's disease from normal aging is the extreme degree of neuronal loss, which is minimal in most brain regions during normal aging. This distinguishing feature is not phenocopied in mouse transgenic models of AD and Parkinson's disease, suggesting that key features of human brain

aging may be missing. Thus, a greater understanding of the normal aging brain may be necessary before we can fully understand the causes of pathological aging and cognitive decline.

#### **DISCLOSURE STATEMENT**

The authors are not aware of any biases that might be perceived as affecting the objectivity of this review.

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#### LITERATURE CITED

- Albert M, Duffy FH, Naeser M. 1987. Nonlinear changes in cognition with age and their neuropsychologic correlates. Can. J. Psychol. 41:141–57
- Craik FI, Moscovitch M, McDowd JM. 1994. Contributions of surface and conceptual information to performance on implicit and explicit memory tasks. J. Exp. Psychol. Learn. Mem. Cogn. 20:864–75
- Zelinski EM, Burnight KP. 1997. Sixteen-year longitudinal and time lag changes in memory and cognition in older adults. Psychol. Aging 12:503–13
- Petersen RC, Smith G, Kokmen E, Ivnik RJ, Tangalos EG. 1992. Memory function in normal aging. Neurology 42:396–401
- Montgomery EB Jr, Koller WC, LaMantia TJ, Newman MC, Swanson-Hyland E, et al. 2000. Early detection of probable idiopathic Parkinson's disease: I. Development of a diagnostic test battery. *Mov. Disord.* 15:467–73
- Lai ZC, Moss MB, Killiany RJ, Rosene DL, Herndon JG. 1995. Executive system dysfunction in the aged monkey: spatial and object reversal learning. Neurobiol. Aging 16:947–54
- 7. Head E, Mehta R, Hartley J, Kameka M, Cummings BJ, et al. 1995. Spatial learning and memory as a function of age in the dog. *Behav. Neurosci.* 109:851–58
- Bach ME, Barad M, Son H, Zhuo M, Lu YF, et al. 1999. Age-related defects in spatial memory are correlated with defects in the late phase of hippocampal long-term potentiation in vitro and are attenuated by drugs that enhance the cAMP signaling pathway. *Proc. Natl. Acad. Sci. USA* 96:5280–85
- 9. Cartensen II, Fung HH, Charles ST. 2003. Socioemotional selectivity theory and the regulation of emotion in the second half of life. *Motiv. Emot.* 27:103–23
- 10. Williams LM, Brown KJ, Palmer D, Liddall BJ, Kemp AH, et al. 2006. The mellow years?: Neural basis of improving emotional stability over age. *7. Neurosci.* 26:6422–30
- Persson J, Sylvester CY, Nelson JK, Welsh KM, Jonides J, Reuter-Lorenz PA. 2004.
   Selection requirements during verb generation: differential recruitment in older and younger adults. *Neuroimage* 23:1382–90

- Logan JM, Sanders AL, Snyder AZ, Morris JC, Buckner RL. 2002. Under-recruitment and nonselective recruitment: dissociable neural mechanisms associated with aging. Neuron 33:827–40
- Remy F, Mirrashed F, Campbell B, Richter W. 2004. Mental calculation impairment in Alzheimer's disease: a functional magnetic resonance imaging study. *Neurosci. Lett.* 358:25–28
- Small SA, Chawla MK, Buonocore M, Rapp PR, Barnes CA. 2004. Imaging correlates
  of brain function in monkeys and rats isolates a hippocampal subregion differentially
  vulnerable to aging. *Proc. Natl. Acad. Sci. USA* 101:7181–86
- Hedden T, Gabrieli JD. 2004. Insights into the ageing mind: a view from cognitive neuroscience. Nat. Rev. Neurosci. 5:87–96
- Burke SN, Barnes CA. 2006. Neural plasticity in the ageing brain. Nat. Rev. Neurosci. 7:30–40
- Buell SJ, Coleman PD. 1979. Dendritic growth in the aged human brain and failure of growth in senile dementia. Science 206:854–56
- Bartzokis G, Cummings JL, Sultzer D, Henderson VW, Nuechterlein KH, Mintz J. 2003.
   White matter structural integrity in healthy aging adults and patients with Alzheimer disease: a magnetic resonance imaging study. *Arch. Neurol.* 60:393–98
- 19. Gunning-Dixon FM, Raz N. 2000. The cognitive correlates of white matter abnormalities in normal aging: a quantitative review. *Neuropsychology* 14:224–32
- Liu X, Erikson C, Brun A. 1996. Cortical synaptic changes and gliosis in normal aging, Alzheimer's disease and frontal lobe degeneration. *Dementia* 7:128–34
- Bourgeois JP, Rakic P. 1996. Synaptogenesis in the occipital cortex of macaque monkey devoid of retinal input from early embryonic stages. Eur. J. Neurosci. 8:942–50
- 22. Geinisman Y, de Toledo-Morrell L, Morrell F. 1986. Loss of perforated synapses in the dentate gyrus: morphological substrate of memory deficit in aged rats. *Proc. Natl. Acad. Sci. USA* 83:3027–31
- Barnes CA. 1979. Memory deficits associated with senescence: a neurophysiological and behavioral study in the rat. J. Comp. Physiol. Psychol. 93:74–104
- 24. Landfield PW, Lynch G. 1977. Impaired monosynaptic potentiation in in vitro hip-pocampal slices from aged, memory-deficient rats. *J. Gerontol.* 32:523–33
- 25. Barnes CA, Rao G, Houston FP. 2000. LTP induction threshold change in old rats at the perforant path–granule cell synapse. *Neurobiol. Aging* 21:613–20
- Norris CM, Korol DL, Foster TC. 1996. Increased susceptibility to induction of longterm depression and long-term potentiation reversal during aging. J. Neurosci. 16:5382–92
- 27. Thibault O, Landfield PW. 1996. Increase in single L-type calcium channels in hippocampal neurons during aging. *Science* 272:1017–20
- 28. Geula C, Bu J, Nagykery N, Scinto LF, Chan J, et al. 2003. Loss of calbindin-D28k from aging human cholinergic basal forebrain: relation to neuronal loss. *J. Comp. Neurol.* 455:249–59
- 29. Lu T, Pan Y, Kao SY, Li C, Kohane I, et al. 2004. Gene regulation and DNA damage in the ageing human brain. *Nature* 429:883-91
- Iacopino AM, Christakos S. 1990. Specific reduction of calcium-binding protein (28-kDa calbindin-D) gene expression in aging and neurodegenerative diseases. *Proc. Natl. Acad. Sci. USA* 87:4078–82
- Mattson MP, Rychlik B, Chu C, Christakos S. 1991. Evidence for calcium-reducing and excito-protective roles for the calcium-binding protein calbindin-D28k in cultured hippocampal neurons. *Neuron* 6:41–51

27. Calcium influx may increase in the aging hippocampus owing to more functional L-type calcium channels.

29. Expression of genes involved in learning, memory, and neuronal survival is reduced in the aging human brain and is associated with DNA damage.

- 32. Lund J, Tedesco P, Duke K, Wang J, Kim SK, Johnson TE. 2002. Transcriptional profile of aging in *C. elegans. Curr. Biol.* 12:1566–73
- 33. McCarroll SA, Murphy CT, Zou S, Pletcher SD, Chin CS, et al. 2004. Comparing genomic expression patterns across species identifies shared transcriptional profile in aging. *Nat. Genet.* 36:197–204
- 34. Zou S, Meadows S, Sharp L, Jan LY, Jan YN. 2000. Genome-wide study of aging and oxidative stress response in *Drosophila melanogaster*. *Proc. Natl. Acad. Sci. USA* 97:13726–31
- 35. Pletcher SD, Macdonald SJ, Marguerie R, Certa U, Stearns SC, et al. 2002. Genome-wide transcript profiles in aging and calorically restricted *Drosophila melanogaster*. Curr. Biol. 12:712–23
- 36. Lee CK, Weindruch R, Prolla TA. 2000. Gene-expression profile of the ageing brain in mice. *Nat. Genet.* 25:294–97
- Jiang CH, Tsien JZ, Schultz PG, Hu Y. 2001. The effects of aging on gene expression in the hypothalamus and cortex of mice. *Proc. Natl. Acad. Sci. USA* 98:1930–34
- Blalock EM, Chen KC, Sharrow K, Herman JP, Porter NM, et al. 2003. Gene microarrays in hippocampal aging: statistical profiling identifies novel processes correlated with cognitive impairment. *J. Neurosci.* 23:3807–19
- 39. Fraser HB, Khaitovich P, Plotkin JB, Paabo S, Eisen MB. 2005. Aging and gene expression in the primate brain. *PLoS Biol.* 3:e274
- Erraji-Benchekroun L, Underwood MD, Arango V, Galfalvy H, Pavlidis P, et al. 2005.
   Molecular aging in human prefrontal cortex is selective and continuous throughout adult life. *Biol. Psychiatry* 57:549–58
- 41. Orr WC, Sohal RS. 1994. Extension of life-span by overexpression of superoxide dismutase and catalase in *Drosophila melanogaster*. Science 263:1128–30
- 42. Lin YJ, Seroude L, Benzer S. 1998. Extended life-span and stress resistance in the *Drosophila* mutant methuselah. *Science* 282:943–46
- Parkes TL, Elia AJ, Dickinson D, Hilliker AJ, Phillips JP, Boulianne GL. 1998. Extension of *Drosophila* lifespan by overexpression of human SOD1 in motorneurons. *Nat. Genet*. 19:171–74
- 44. Migliaccio E, Giorgio M, Mele S, Pelicci G, Reboldi P, et al. 1999. The p66shc adaptor protein controls oxidative stress response and life span in mammals. *Nature* 402:309–13
- 45. Hyun DH, Emerson SS, Jo DG, Mattson MP, de Cabo R. 2006. Calorie restriction upregulates the plasma membrane redox system in brain cells and suppresses oxidative stress during aging. *Proc. Natl. Acad. Sci. USA* 103:19908–12
- Blalock EM, Geddes JW, Chen KC, Porter NM, Markesbery WR, Landfield PW. 2004. Incipient Alzheimer's disease: microarray correlation analyses reveal major transcriptional and tumor suppressor responses. *Proc. Natl. Acad. Sci. USA* 101:2173–78
- 47. Small SA, Kent K, Pierce A, Leung C, Kang MS, et al. 2005. Model-guided microarray implicates the retromer complex in Alzheimer's disease. *Ann. Neurol.* 58:909–19
- Lazarov O, Robinson J, Tang YP, Hairston IS, Korade-Mirnics Z, et al. 2005. Environmental enrichment reduces Aβ levels and amyloid deposition in transgenic mice. *Cell* 120:701–13
- 49. Fischer A, Sananbenesi F, Wang X, Dobbin M, Tsai LH. 2007. Recovery of learning and memory is associated with chromatin remodeling. *Nature* 447:178–82
- 50. Hasty P, Campisi J, Hoeijmakers J, van Steeg H, Vijg J. 2003. Aging and genome maintenance: lessons from the mouse? *Science* 299:1355–59
- Niedernhofer LJ, Garinis GA, Raams A, Lalai AS, Robinson AR, et al. 2006. A new progeroid syndrome reveals that genotoxic stress suppresses the somatotroph axis. *Nature* 444:1038–43

- 52. Leverenz JB, Yu CE, Schellenberg GD. 1998. Aging-associated neuropathology in Werner syndrome. *Acta Neuropathol. (Berl.)* 96:421–24
- 53. Lombard DB, Chua KF, Mostoslavsky R, Franco S, Gostissa M, Alt FW. 2005. DNA repair, genome stability, and aging. *Cell* 120:497–512
- Ferguson DO, Alt FW. 2001. DNA double strand break repair and chromosomal translocation: lessons from animal models. Oncogene 20:5572–79
- Bauer JH, Poon PC, Glatt-Deeley H, Abrams JM, Helfand SL. 2005. Neuronal expression of p53 dominant-negative proteins in adult *Drosophila melanogaster* extends life span. *Curr. Biol.* 15:2063–68
- Mecocci P, MacGarvey U, Kaufman AE, Koontz D, Shoffner JM, et al. 1993. Oxidative damage to mitochondrial DNA shows marked age-dependent increases in human brain. *Ann. Neurol.* 34:609–16
- 57. Hamilton ML, Van Remmen H, Drake JA, Yang H, Guo ZM, et al. 2001. Does oxidative damage to DNA increase with age? *Proc. Natl. Acad. Sci. USA* 98:10469–74
- 58. Liu J, Head E, Gharib AM, Yuan W, Ingersoll RT, et al. 2002. Memory loss in old rats is associated with brain mitochondrial decay and RNA/DNA oxidation: partial reversal by feeding acetyl-L-carnitine and/or *R*-α-lipoic acid. *Proc. Natl. Acad. Sci. USA* 99:2356–61
- 59. Mecocci P, MacGarvey U, Beal MF. 1994. Oxidative damage to mitochondrial DNA is increased in Alzheimer's disease. *Ann. Neurol.* 36:747–51
- 60. Honda K, Smith MA, Zhu X, Baus D, Merrick WC, et al. 2005. Ribosomal RNA in Alzheimer disease is oxidized by bound redox-active iron. *J. Biol. Chem.* 280:20978–86
- 61. Ohno M, Miura T, Furuichi M, Tominaga Y, Tsuchimoto D, et al. 2006. A genome-wide distribution of 8-oxoguanine correlates with the preferred regions for recombination and single nucleotide polymorphism in the human genome. *Genome Res.* 16:567–75
- Eur. Stand. Comm. Oxid. DNA Damage. 2003. Measurement of DNA oxidation in human cells by chromatographic and enzymic methods. Free Radic. Biol. Med. 34:1089– 99
- 63. Wallace DC. 2005. A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: a dawn for evolutionary medicine. *Annu. Rev. Genet.* 39:359–407
- 64. Zecca L, Youdim MB, Riederer P, Connor JR, Crichton RR. 2004. Iron, brain ageing and neurodegenerative disorders. *Nat. Rev. Neurosci.* 5:863–73
- 65. Brunk UT, Terman A. 2002. The mitochondrial-lysosomal axis theory of aging: accumulation of damaged mitochondria as a result of imperfect autophagocytosis. *Eur. J. Biochem.* 269:1996–2002
- 66. Shibata M, Lu T, Furuya T, Degterev A, Mizushima N, et al. 2006. Regulation of intracellular accumulation of mutant Huntingtin by Beclin 1. 7. Biol. Chem. 281:14474–85
- 67. Holt IJ, Harding AE, Morgan-Hughes JA. 1988. Deletions of muscle mitochondrial DNA in patients with mitochondrial myopathies. *Nature* 331:717–19
- 68. Wallace DC, Zheng XX, Lott MT, Shoffner JM, Hodge JA, et al. 1988. Familial mitochondrial encephalomyopathy (MERRF): genetic, pathophysiological, and biochemical characterization of a mitochondrial DNA disease. *Cell* 55:601–10
- 69. Richter C, Park JW, Ames BN. 1988. Normal oxidative damage to mitochondrial and nuclear DNA is extensive. *Proc. Natl. Acad. Sci. USA* 85:6465–67
- Lin MT, Simon DK, Ahn CH, Kim LM, Beal MF. 2002. High aggregate burden of somatic mtDNA point mutations in aging and Alzheimer's disease brain. *Hum. Mol. Genet.* 11:133–45

58. Feeding aging rats mitochondrial metabolites and antioxidants can partially reverse DNA and RNA damage and ameliorate memory loss.

73. Excessive mutation of the mitochondrial genome accelerates aging.

- 84. The amyloid β-protein precursor gene is identified and localized to chromosome 21, implicating it as a potential etiologic factor in AD.
- 87. Identification of the first mutation in APP associated with autosomal dominant inheritance of AD, providing genetic evidence for the amyloid hypothesis.

- Coskun PE, Beal MF, Wallace DC. 2004. Alzheimer's brains harbor somatic mtDNA control-region mutations that suppress mitochondrial transcription and replication. *Proc.* Natl. Acad. Sci. USA 101:10726–31
- Imam SZ, Karahalil B, Hogue BA, Souza-Pinto NC, Bohr VA. 2006. Mitochondrial and nuclear DNA-repair capacity of various brain regions in mouse is altered in an agedependent manner. *Neurobiol. Aging* 27:1129–36
- Trifunovic A, Wredenberg A, Falkenberg M, Spelbrink JN, Rovio AT, et al. 2004.
   Premature ageing in mice expressing defective mitochondrial DNA polymerase.
   Nature 429:417–23
- 74. Kujoth GC, Hiona A, Pugh TD, Someya S, Panzer K, et al. 2005. Mitochondrial DNA mutations, oxidative stress, and apoptosis in mammalian aging. *Science* 309:481–84
- Schriner SE, Linford NJ, Martin GM, Treuting P, Ogburn CE, et al. 2005. Extension of murine life span by overexpression of catalase targeted to mitochondria. Science 308:1909– 11
- Roth M, Tomlinson BE, Blessed G. 1966. Correlation between scores for dementia and counts of 'senile plaques' in cerebral gray matter of elderly subjects. *Nature* 209:109–10
- 77. Terry RD, Masliah E, Salmon DP, Butters N, DeTeresa R, et al. 1991. Physical basis of cognitive alterations in Alzheimer's disease: Synapse loss is the major correlate of cognitive impairment. *Ann. Neurol.* 30:572–80
- 78. West MJ, Coleman PD, Flood DG, Troncoso JC. 1994. Differences in the pattern of hippocampal neuronal loss in normal ageing and Alzheimer's disease. *Lancet* 344:769–72
- 79. Price JL, Ko AI, Wade MJ, Tsou SK, McKeel DW, Morris JC. 2001. Neuron number in the entorhinal cortex and CA1 in preclinical Alzheimer disease. *Arch. Neurol.* 58:1395–402
- 80. Gomez-Isla T, Price JL, McKeel DW Jr, Morris JC, Growdon JH, Hyman BT. 1996. Profound loss of layer II entorhinal cortex neurons occurs in very mild Alzheimer's disease. 7. Neurosci. 16:4491–500
- Ingelsson M, Fukumoto H, Newell KL, Growdon JH, Hedley-Whyte ET, et al. 2004.
   Early Aβ accumulation and progressive synaptic loss, gliosis, and tangle formation in AD brain. Neurology 62:925–31
- 82. Lassmann H, Fischer P, Jellinger K. 1993. Synaptic pathology of Alzheimer's disease. *Ann. N. Y. Acad. Sci.* 695:59–64
- Lai F, Williams RS. 1989. A prospective study of Alzheimer disease in Down syndrome. *Arch. Neurol.* 46:849–53
- 84. Kang J, Lemaire HG, Unterbeck A, Salbaum JM, Masters CL, et al. 1987. The precursor of Alzheimer's disease amyloid A4 protein resembles a cell-surface receptor. *Nature* 325:733–36
- Van Broeckhoven C, Haan J, Bakker E, Hardy JA, Van Hul W, et al. 1990. Amyloid beta protein precursor gene and hereditary cerebral hemorrhage with amyloidosis (Dutch). Science 248:1120–22
- 86. Levy E, Carman MD, Fernandez-Madrid IJ, Power MD, Lieberburg I, et al. 1990. Mutation of the Alzheimer's disease amyloid gene in hereditary cerebral hemorrhage, Dutch type. *Science* 248:1124–26
- Goate A, Chartier-Harlin MC, Mullan M, Brown J, Crawford F, et al. 1991. Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. *Nature* 349:704–6
- 88. Suzuki N, Cheung TT, Cai XD, Odaka A, Otvos LJr, et al. 1994. An increased percentage of long amyloid beta protein secreted by familial amyloid beta protein precursor (beta APP717) mutants. *Science* 264:1336–40

- 89. Jarrett JT, Lansbury PT Jr. 1993. Seeding "one-dimensional crystallization" of amyloid: a pathogenic mechanism in Alzheimer's disease and scrapie? *Cell* 73:1055–58
- 90. Citron M, Oltersdorf T, Haass C, McConlogue L, Hung AY, et al. 1992. Mutation of the beta-amyloid precursor protein in familial Alzheimer's disease increases beta-protein production. *Nature* 360:672–74
- Cai XD, Golde TE, Younkin SG. 1993. Release of excess amyloid beta protein from a mutant amyloid beta protein precursor. Science 259:514–16
- 92. Sherrington R, Rogaev EI, Liang Y, Rogaeva EA, Levesque G, et al. 1995. Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's disease. *Nature* 375:754–60
- 93. Levy-Lahad E, Lahad A, Wijsman EM, Bird TD, Schellenberg GD. 1995. Apolipoprotein E genotypes and age of onset in early-onset familial Alzheimer's disease. *Ann. Neurol.* 38:678–80
- 94. Levy-Lahad E, Wasco W, Poorkaj P, Romano DM, Oshima J, et al. 1995. Candidate gene for the chromosome 1 familial Alzheimer's disease locus. *Science* 269:973–77
- 95. Rogaev EI, Sherrington R, Rogaeva EA, Levesque G, Ikeda M, et al. 1995. Familial Alzheimer's disease in kindreds with missense mutations in a gene on chromosome 1 related to the Alzheimer's disease type 3 gene. *Nature* 376:775–78
- Li J, Ma J, Potter H. 1995. Identification and expression analysis of a potential familial Alzheimer disease gene on chromosome 1 related to AD3. Proc. Natl. Acad. Sci. USA 92:12180–84
- 97. De Strooper B, Saftig P, Craessaerts K, Vanderstichele H, Guhde G, et al. 1998. Deficiency of presenilin-1 inhibits the normal cleavage of amyloid precursor protein. *Nature* 391:387–90
- Wolfe MS, Xia W, Ostaszewski BL, Diehl TS, Kimberly WT, Selkoe DJ. 1999. Two transmembrane aspartates in presenilin-1 required for presenilin endoproteolysis and γ-secretase activity. *Nature* 398:513–17
- 99. Li YM, Xu M, Lai MT, Huang Q, Castro JL, et al. 2000. Photoactivated γ-secretase inhibitors directed to the active site covalently label presenilin 1. *Nature* 405:689–94
- De Strooper B. 2003. Aph-1, Pen-2, and Nicastrin with Presentilin generate an active γ-Secretase complex. Neuron 38:9–12
- 101. Scheuner D, Eckman C, Jensen M, Song X, Citron M, et al. 1996. Secreted amyloid beta-protein similar to that in the senile plaques of Alzheimer's disease is increased in vivo by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease. Nat. Med. 2:864–70
- 102. Games D, Adams D, Alessandrini R, Barbour R, Berthelette P, et al. 1995. Alzheimertype neuropathology in transgenic mice overexpressing V717F beta-amyloid precursor protein. *Nature* 373:523–27
- 103. Borchelt DR, Ratovitski T, van Lare J, Lee MK, Gonzales V, et al. 1997. Accelerated amyloid deposition in the brains of transgenic mice coexpressing mutant presenilin 1 and amyloid precursor proteins. *Neuron* 19:939–45
- 104. Strittmatter WJ, Saunders AM, Schmechel D, Pericak-Vance M, Enghild J, et al. 1993. Apolipoprotein E: high-avidity binding to beta-amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer disease. *Proc. Natl. Acad. Sci. USA* 90:1977–81
- 105. Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, et al. 1993. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. Science 261:921–23

92. Identification of presenilin-1 as a genetic cause of early-onset familial AD.

104. Identification of the E4 allele of apolipoprotein E as a prevalent genetic risk factor for late-onset AD.

108. Demonstra-

tion that the Aß

peptide can cause neuronal cell death.

- 106. Holtzman DM, Bales KR, Tenkova T, Fagan AM, Parsadanian M, et al. 2000. Apolipoprotein E isoform-dependent amyloid deposition and neuritic degeneration in a mouse model of Alzheimer's disease. *Proc. Natl. Acad. Sci. USA* 97:2892–97
- Laskowitz DT, Horsburgh K, Roses AD. 1998. Apolipoprotein E and the CNS response to injury. 7. Cereb. Blood Flow Metab. 18:465–71
- 108. Yankner BA, Duffy LK, Kirschner DA. 1990. Neurotrophic and neurotoxic effects of amyloid beta protein: reversal by tachykinin neuropeptides. Science 250:279–82
- Pike CJ, Burdick D, Walencewicz AJ, Glabe CG, Cotman CW. 1993. Neurodegeneration induced by beta-amyloid peptides in vitro: the role of peptide assembly state. J. Neurosci. 13:1676–87
- Yankner BA. 1996. Mechanisms of neuronal degeneration in Alzheimer's disease. Neuron 16:921–32
- 111. Irizarry MC, Soriano F, McNamara M, Page KJ, Schenk D, et al. 1997. Aβ deposition is associated with neuropil changes, but not with overt neuronal loss in the human amyloid precursor protein V717F (PDAPP) transgenic mouse. J. Neurosci. 17:7053–59
- Urbanc B, Cruz L, Le R, Sanders J, Ashe KH, et al. 2002. Neurotoxic effects of thioflavin S-positive amyloid deposits in transgenic mice and Alzheimer's disease. *Proc. Natl. Acad.* Sci. USA 99:13990–95
- 113. Geula C, Wu CK, Saroff D, Lorenzo A, Yuan M, Yankner BA. 1998. Aging renders the brain vulnerable to amyloid beta-protein neurotoxicity. *Nat. Med.* 4:827–31
- Deshpande A, Mina E, Glabe C, Busciglio J. 2006. Different conformations of amyloid beta induce neurotoxicity by distinct mechanisms in human cortical neurons. J. Neurosci. 26:6011–18
- 115. Lambert MP, Barlow AK, Chromy BA, Edwards C, Freed R, et al. 1998. Diffusible, nonfibrillar ligands derived from Aβ1-42 are potent central nervous system neurotoxins. Proc. Natl. Acad. Sci. USA 95:6448–53
- 116. Walsh DM, Klyubin I, Fadeeva JV, Cullen WK, Anwyl R, et al. 2002. Naturally secreted oligomers of amyloid beta protein potently inhibit hippocampal long-term potentiation in vivo. *Nature* 416:535–39
- 117. Cleary JP, Walsh DM, Hofmeister JJ, Shankar GM, Kuskowski MA, et al. 2005. Natural oligomers of the amyloid-beta protein specifically disrupt cognitive function. *Nat. Neurosci.* 8:79–84
- 118. Lesne S, Koh MT, Kotilinek L, Kayed R, Glabe CG, et al. 2006. A specific amyloid-beta protein assembly in the brain impairs memory. *Nature* 440:352–57
- 119. Lee MS, Kwon YT, Li M, Peng J, Friedlander RM, Tsai LH. 2000. Neurotoxicity induces cleavage of p35 to p25 by calpain. *Nature* 405:360–64
- Nakagawa T, Zhu H, Morishima N, Li E, Xu J, et al. 2000. Caspase-12 mediates endoplasmic-reticulum-specific apoptosis and cytotoxicity by amyloid-beta. *Nature* 403:98–103
- 121. Troy CM, Rabacchi SA, Friedman WJ, Frappier TF, Brown K, Shelanski ML. 2000. Caspase-2 mediates neuronal cell death induced by beta-amyloid. J. Neurosci. 20:1386–92
- 122. Snyder EM, Nong Y, Almeida CG, Paul S, Moran T, et al. 2005. Regulation of NMDA receptor trafficking by amyloid-beta. *Nat. Neurosci.* 8:1051–58
- 123. Hsieh H, Boehm J, Sato C, Iwatsubo T, Tomita T, et al. 2006. AMPAR removal underlies Aβ-induced synaptic depression and dendritic spine loss. Neuron 52:831–43
- 124. Cummings JL. 2004. Treatment of Alzheimer's disease: current and future therapeutic approaches. *Rev. Neurol. Dis.* 1:60–69

- 125. Yang Y, Mufson EJ, Herrup K. 2003. Neuronal cell death is preceded by cell cycle events at all stages of Alzheimer's disease. *7. Neurosci.* 23:2557–63
- 126. Kruman II, Wersto RP, Cardozo-Pelaez F, Smilenov L, Chan SL, et al. 2004. Cell cycle activation linked to neuronal cell death initiated by DNA damage. *Neuron* 41:549–61
- 127. Galvan V, Gorostiza OF, Banwait S, Ataie M, Logvinova AV, et al. 2006. Reversal of Alzheimer's-like pathology and behavior in human APP transgenic mice by mutation of Asp664. Proc. Natl. Acad. Sci. USA 103:7130–35
- 128. Yankner BA, Dawes LR, Fisher S, Villa-Komaroff L, Oster-Granite ML, Neve RL. 1989. Neurotoxicity of a fragment of the amyloid precursor associated with Alzheimer's disease. Science 245:417–20
- Lorenzo A, Yuan M, Zhang Z, Paganetti PA, Sturchler-Pierrat C, et al. 2000. Amyloid beta interacts with the amyloid precursor protein: a potential toxic mechanism in Alzheimer's disease. *Nat. Neurosci.* 3:460–64
- 130. Lu DC, Rabizadeh S, Chandra S, Shayya RF, Ellerby LM, et al. 2000. A second cytotoxic proteolytic peptide derived from amyloid beta-protein precursor. *Nat. Med.* 6:397–404
- 131. Shaked GM, Kummer MP, Lu DC, Galvan V, Bredesen DE, Koo EH. 2006. Aβ induces cell death by direct interaction with its cognate extracellular domain on APP (APP 597–624). *FASEB* 7. 20:1254–56
- Skovronsky DM, Lee VM, Trojanowski J. 2006. Neurodegenerative diseases: new concepts of pathogenesis and their therapeutic implications. *Annu. Rev. Pathol. Mech. Dis.* 1:151–70
- 133. Hutton M, Lendon CL, Rizzu P, Baker M, Froelich S, et al. 1998. Association of missense and 5'-splice-site mutations in tau with the inherited dementia FTDP-17. *Nature* 393:702-5
- 134. Baker M, Mackenzie IR, Pickering-Brown SM, Gass J, Rademakers R, et al. 2006. Mutations in progranulin cause tau-negative frontotemporal dementia linked to chromosome 17. *Nature* 442:916–19
- 135. Cruts M, Gijselinck I, van der Zee J, Engelborghs S, Wils H, et al. 2006. Null mutations in progranulin cause ubiquitin-positive frontotemporal dementia linked to chromosome 17q21. Nature 442:920–24
- 136. Lewis J, McGowan E, Rockwood J, Melrose H, Nacharaju P, et al. 2000. Neurofibrillary tangles, amyotrophy and progressive motor disturbance in mice expressing mutant (P301L) tau protein. *Nat. Genet.* 25:402–5
- 137. Santacruz K, Lewis J, Spires T, Paulson J, Kotilinek L, et al. 2005. Tau suppression in a neurodegenerative mouse model improves memory function. *Science* 309:476–81
- 138. Oddo S, Caccamo A, Shepherd JD, Murphy MP, Golde TE, et al. 2003. Triple-transgenic model of Alzheimer's disease with plaques and tangles: intracellular Aβ and synaptic dysfunction. *Neuron* 39:409–21
- 139. Oddo S, Vasilevko V, Caccamo A, Kitazawa M, Cribbs DH, Laferla FM. 2006. Reduction of soluble Aβ and tau, but not soluble Aβ alone, ameliorates cognitive decline in transgenic mice with plaques and tangles. J. Biol. Chem. 281:39413–23
- Roberson ED, Scearce-Levie K, Palop JJ, Yan F, Cheng IH, et al. 2007. Reducing endogenous tau ameliorates amyloid beta-induced deficits in an Alzheimer's disease mouse model. Science 316:750–54
- Fillit HM, Butler RN, O'Connell AW, Albert MS, Birren JE, et al. 2002. Achieving and maintaining cognitive vitality with aging. Mayo Clin. Proc. 77:681–96
- van Praag H, Shubert T, Zhao C, Gage FH. 2005. Exercise enhances learning and hip-pocampal neurogenesis in aged mice. J. Neurosci. 25:8680–85

133. Identification of mutations in tau as a cause of familial frontotemporal dementia, linking tau pathology to neurodegeneration.

- Kramer AF, Erickson KI, Colcombe SJ. 2006. Exercise, cognition, and the aging brain.
   Appl. Physiol. 101:1237–42
- 144. Pereira AC, Huddleston DE, Brickman AM, Sosunov AA, Hen R, et al. 2007. An in vivo correlate of exercise-induced neurogenesis in the adult dentate gyrus. *Proc. Natl. Acad. Sci. USA* 104:5638–43
- 145. Weuve J, Kang JH, Manson JE, Breteler MM, Ware JH, Grodstein F. 2004. Physical activity, including walking, and cognitive function in older women. *JAMA* 292:1454–61
- 146. Wolf SA, Kronenberg G, Lehmann K, Blankenship A, Overall R, et al. 2006. Cognitive and physical activity differently modulate disease progression in the amyloid precursor protein (APP)-23 model of Alzheimer's disease. *Biol. Psychiatry* 60:1314–23
- Morris MC, Evans DA, Bienias JL, Tangney CC, Wilson RS. 2002. Vitamin E and cognitive decline in older persons. Arch. Neurol. 59:1125–32
- 148. Sano M, Ernesto C, Thomas RG, Klauber MR, Schafer K, et al. 1997. A controlled trial of selegiline, alpha-tocopherol, or both as treatment for Alzheimer's disease. The Alzheimer's Disease Cooperative Study. N. Engl. J. Med. 336:1216–22
- Matthews RT, Yang L, Browne S, Baik M, Beal MF. 1998. Coenzyme Q10 administration increases brain mitochondrial concentrations and exerts neuroprotective effects. *Proc.* Natl. Acad. Sci. USA 95:8892–97
- Shults CW, Oakes D, Kieburtz K, Beal MF, Haas R, et al. 2002. Effects of coenzyme Q10 in early Parkinson disease: evidence of slowing of the functional decline. *Arch. Neurol.* 59:1541–50
- McGeer PL, Schulzer M, McGeer EG. 1996. Arthritis and anti-inflammatory agents as possible protective factors for Alzheimer's disease: a review of 17 epidemiologic studies. *Neurology* 47:425–32
- 152. Firuzi Ö, Pratico D. 2006. Coxibs and Alzheimer's disease: Should they stay or should they go? *Ann. Neurol.* 59:219–28
- 153. Morris MC. 2006. Docosahexaenoic acid and Alzheimer disease. Arch. Neurol. 63:1527–28
- 154. Calon F, Lim GP, Yang F, Morihara T, Teter B, et al. 2004. Docosahexaenoic acid protects from dendritic pathology in an Alzheimer's disease mouse model. *Neuron* 43:633–45
- 155. Lim GP, Calon F, Morihara T, Yang F, Teter B, et al. 2005. A diet enriched with the omega-3 fatty acid docosahexaenoic acid reduces amyloid burden in an aged Alzheimer mouse model. J. Neurosci. 25:3032–40
- 156. Haigis MC, Guarente LP. 2006. Mammalian sirtuins—emerging roles in physiology, aging, and calorie restriction. *Genes Dev.* 20:2913–21
- 157. Howitz KT, Bitterman KJ, Cohen HY, Lamming DW, Lavu S, et al. 2003. Small molecule activators of sirtuins extend *Saccharomyces cerevisiae* lifespan. *Nature* 425:191–96
- Wood JG, Rogina B, Lavu S, Howitz K, Helfand SL, et al. 2004. Sirtuin activators mimic caloric restriction and delay ageing in metazoans. *Nature* 430:686–89
- 159. Baur JA, Pearson KJ, Price NL, Jamieson HA, Lerin C, et al. 2006. Resveratrol improves health and survival of mice on a high-calorie diet. *Nature* 444:337–42
- Lagouge M, Argmann C, Gerhart-Hines Z, Meziane H, Lerin C, et al. 2006. Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1α. Cell 127:1109–22
- 161. Araki T, Sasaki Y, Milbrandt J. 2004. Increased nuclear NAD biosynthesis and SIRT1 activation prevent axonal degeneration. *Science* 305:1010–13
- 162. Hebert LE, Scherr PA, Beckett LA, Albert MS, Pilgrim DM, et al. 1995. Age-specific incidence of Alzheimer's disease in a community population. *JAMA* 273:1354–59
- 163. Hedges SB. 2002. The origin and evolution of model organisms. Nat. Rev. Genet. 3:838-49



Annual Review of Pathology: Mechanisms of Disease

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# Contents

The Relevance of Research on Red Cell Membranes to the
Understanding of Complex Human Disease: A Personal Perspective  Vincent T. Marchesi
Molecular Mechanisms of Prion Pathogenesis  Adriano Aguzzi, Christina Sigurdson, and Mathias Heikenwalder
The Aging Brain Bruce A. Yankner, Tao Lu, and Patrick Loerch
Gene Expression Profiling of Breast Cancer  Maggie C.U. Cheang, Matt van de Rijn, and Torsten O. Nielsen
The Inflammatory Response to Cell Death  Kenneth L. Rock and Hajime Kono
Molecular Biology and Pathogenesis of Viral Myocarditis  Mitra Esfandiarei and Bruce M. McManus
Pancreatic Cancer Anirban Maitra and Ralph H. Hruban
Kidney Transplantation: Mechanisms of Rejection and Acceptance  Lynn D. Cornell, R. Neal Smith, and Robert B. Colvin
Metastatic Cancer Cell  Marina Bacac and Ivan Stamenkovic
Pathogenesis of Thrombotic Microangiopathies  X. Long Zheng and J. Evan Sadler
Anti-Inflammatory and Proresolving Lipid Mediators  Charles N. Serhan, Stephanie Yacoubian, and Rong Yang
Modeling Morphogenesis and Oncogenesis in Three-Dimensional Breast Epithelial Cultures
Christy Hebner, Valerie M. Weaver, and Jayanta Debnath313

The Origins of Medulloblastoma Subtypes  *Richard J. Gilbertson and David W. Ellison	341
Molecular Biology and Pathology of Lymphangiogenesis  Terhi Karpanen and Kari Alitalo	367
Endoplasmic Reticulum Stress in Disease Pathogenesis  Jonathan H. Lin, Peter Walter, and T.S. Benedict Yen	399
Autophagy: Basic Principles and Relevance to Disease  Mondira Kundu and Craig B. Thompson	427
The Osteoclast: Friend or Foe?  Deborah V. Novack and Steven L. Teitelbaum	457
Applications of Proteomics to Lab Diagnosis  Raghothama Chaerkady and Akhilesh Pandey	485
The Pathology of Influenza Virus Infections  *Jeffrey K. Taubenberger and David M. Morens	499
Airway Smooth Muscle in Asthma  Marc B. Hershenson, Melanie Brown, Blanca Camoretti-Mercado, and Julian Solway	523
Molecular Pathobiology of Gastrointestinal Stromal Sarcomas  Christopher L. Corless and Michael C. Heinrich	557
Notch Signaling in Leukemia  Jon C. Aster, Warren S. Pear, and Stephen C. Blacklow	587
The Role of Hypoxia in Vascular Injury and Repair  Tony E. Walshe and Patricia A. D'Amore	615
Indexes	
Cumulative Index of Contributing Authors, Volumes 1–3	645
Cumulative Index of Chapter Titles, Volumes 1–3	647

### Errata

An online log of corrections to *Annual Review of Pathology: Mechanisms of Disease* articles may be found at http://pathol.annualreviews.org