The cell biology of aging

Race DiLoreto and Coleen T. Murphy

Department of Molecular Biology, Lewis-Sigler Institute for Integrative Genomics, Princeton University, Princeton, NJ 08544

ABSTRACT One of the original hypotheses of organismal longevity posits that aging is the natural result of entropy on the cells, tissues, and organs of the animal—a slow, inexorable slide into nonfunctionality caused by stochastic degradation of its parts. We now have evidence that aging is instead at least in part genetically regulated. Many mutations have been discovered to extend lifespan in organisms of all complexities, from yeast to mammals. The study of metazoan model organisms, such as Caenorhabditis elegans, has been instrumental in understanding the role of genetics in the cell biology of aging. Longevity mutants across the spectrum of model organisms demonstrate that rates of aging are regulated through genetic control of cellular processes. The regulation and subsequent breakdown of cellular processes represent a programmatic decision by the cell to either continue or abandon maintenance procedures with age. Our understanding of cell biological processes involved in regulating aging have been particularly informed by longevity mutants and treatments, such as reduced insulin/IGF-1 signaling and dietary restriction, which are critical in determining the distinction between causes of and responses to aging and have revealed a set of downstream targets that participate in a range of cell biological activities. Here we briefly review some of these important cellular processes.

Monitoring Editor William Bement University of Wisconsin

Received: Aug 12, 2015 Revised: Aug 20, 2015 Accepted: Aug 21, 2015

INTRODUCTION

To achieve lifespan extension in humans, we must understand which cellular programs are responsible for aging and how their dysregulation directs senescence and decline. In many degenerative diseases associated with aging, the specific, proximal etiologies of the disease, such as protein aggregation, stem from dysregulation of processes responsible for regulation of healthy aging, such as autophagy and proteostasis. Addressing the role of such cellular processes not only with age but also in the context of Alzheimer's disease and Parkinson's disease will provide valuable insight into the fundamental biology of aging, as well as directly aid in increasing the quality of life of a population aging at high risk for these diseases.

DOI:10.1091/mbc.E14-06-1084

Address correspondence to: Coleen T. Murphy (ctmurphy@Princeton.edu).

Abbreviations used: CMA, chaperone-mediated autophagy; DMQ, demethoxyubiquinone; DR, dietary restriction; ECM, extracellular matrix; ER, endoplasmic reticulum; ERAD, ER-associated decay; ETC, electron transport chain; IGF-1, insulin-like growth factor 1; IIS, insulin/IGF-1 signaling; NPC, nuclear pore complex; ROS, reactive oxygen species; SASP, senescence-associated secretory phenotype; TOR, target of rapamycin; UPR, unfolded protein response.

© 2015 DiLoreto and Murphy. This article is distributed by The American Society for Cell Biology under license from the author(s). Two months after publication it is available to the public under an Attribution–Noncommercial–Share Alike 3.0 Unported Creative Commons License (http://creativecommons.org/licenses/by-nc-sa/3.0).

"ASCB®," "The American Society for Cell Biology®," and "Molecular Biology of the Cell®" are registered trademarks of The American Society for Cell Biology.

Here we review a number of crucial processes responsible for regulating cellular health in aging and the link between many of those diseases and aging. Cellular health is controlled at various points in the cell, starting in the nucleus through chromosome structure/organization, transcriptional regulation, and nuclear export/import, ranging outward to protein translation and quality control, autophagic recycling of organelles, maintenance of cytoskeletal structure, and finally maintenance of the extracellular matrix and extracellular signaling (Figure 1). Each regulatory system receives information from every other system, resulting in an intricate interplay of regulation controlling the aging of the cell.

CHROMOSOME AND TELOMERE REGULATION

Telomeres cap each chromosome with a repetitive sequence, protecting chromosomes from damage caused by shortening (due to end-replication problems on the lagging strand and oxidative damage; Richter and von Zglinicki, 2007) at each replicative cycle (Richter and von Zglinicki, 2007). Telomere length, which is inversely correlated with lifespan (Gomes et al., 2011), and rate of accumulation of short telomeres are predictors of lifespan in zebra finch and mice (Heidinger et al., 2012; Vera et al., 2012), and the longest-lived organisms have long telomeres (Heidinger et al., 2012). Long telomeres are linked to increased stress resistance in Caenorhabditis elegans (Park et al., 2010), and increased stress in humans leads to increased telomere shortening (Shalev et al., 2013), suggesting that

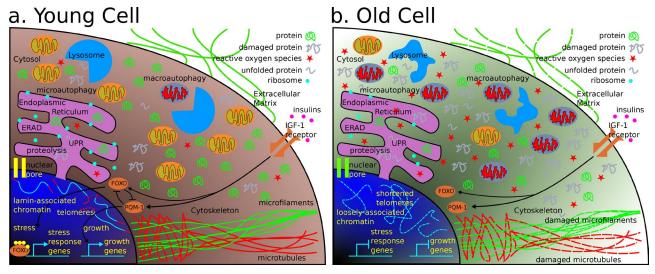


FIGURE 1: Major features of cellular aging. As the cell ages, translational defects and entropy progressively increase the amount of cellular damage, and clearance and quality control mechanisms grow less effective. (a) In a young cell, most organelles are very healthy, and when proteins are translated and misfolded or acquire damage in the cytosol, they are cleared either by ERAD (in the ER) or autophagy (in the cytosol). When organelles become too damaged, they are degraded to component parts by macroautophagy. (b) In an older cell, accumulated damage leads to a less healthy cell. ROS build up from damaged mitochondria and contribute to a greater fraction of the proteome consisting of damaged proteins and protein aggregates.

regulation of telomere length is predictive of longevity and influenced by longevity-related factors. Mammalian cell cultures enter senescence after 40–60 divisions (Hayflick, 1965), known as the Hayflick limit or replicative senescence (Hayflick and Moorhead, 1961). Shortening of the telomeres at each replication (Olovnikov, 1996; Shay and Wright, 2000) leads to cellular senescence (Harley et al., 1992). This telomere shortening has been implicated in many age-related phenotypes, such as decline in innate immunity (Effros and Pawelec, 1997; Effros et al., 2005), and even correlates with protein diseases, such as Alzheimer's disease (Panossian et al., 2003).

The sirtuins SIR3 and SIR4 are associated with lifespan and nuclear organization in Saccharomyces cerevisiae (Kaeberlein et al., 1999), regulating nucleolus fragmentation in aging yeast (Lewinska et al., 2014) and leading to a relocation of the telomeres to the nucleolus in old cells (Lo et al., 2006) and progressive dysregulation of the cell. Nuclear shape degrades, and heterochromatin becomes dissociated from the periphery in aged worms (Haithcock et al., 2005) but these processes are slowed in long-lived insulin/insulin-like growth factor 1 (IGF-1) signaling (IIS) mutants (Haithcock et al., 2005).

TRANSCRIPTIONAL REGULATION

Transcriptional regulation is key in coordinating the activation of many genes to extend lifespan. Most cellular processes that affect longevity are regulated at the transcriptional level through highly conserved signaling pathways (Kenyon, 2010a) including the IIS (Kenyon et al., 1993; Ogg et al., 1997; Lin et al., 2001; Lee et al., 2003; McElwee et al., 2003; Murphy et al., 2003) and target of rapamycin (TOR) pathways (Jia et al., 2004; Kaeberlein and Shamieh, 2010), which regulate gene expression in response to stress and nutrient availability stimuli (Jia et al., 2004). Regulation by the IIS pathway in *C. elegans* involves primarily the PQM-1 and DAF-16/FOXO transcription factors, which localize to the nucleus in a mutually exclusive manner (Tepper et al., 2013) and promote either growth/development or stress response/longevity, respectively (Kenyon et al., 1993; Lin et al., 1997, 2001; Ogg et al., 1997; Lee et al., 2003;

McElwee et al., 2003; Murphy et al., 2003; Tepper et al., 2013); their mutually exclusive nuclear localization breaks down with age (Tepper et al., 2013). The downstream targets of these pathways include genes implicated in regulation of cellular health (Murphy et al., 2003). The heat shock factor HSF-1 is responsible for regulation of cytoskeletal integrity (Baird et al., 2014), heat stress resistance (Hsu et al., 2003; Morley and Morimoto, 2004), and protein quality control (Morley and Morimoto, 2004), all of which contribute to its effect on *C. elegans* longevity (Hsu et al., 2003; Morley and Morimoto, 2004; Baird et al., 2014). The Nrf/SKN-1 transcription factor mediates longevity (Tullet et al., 2008; Robida-Stubbs et al., 2012), as well as regulation of extracellular collagen matrices (Ewald et al., 2015).

NUCLEAR TRAFFICKING AND ORGANIZATION

The eukaryotic nuclear pore complex (NPC), one of the most complex molecular devices, serves an essential role in exporting messages and proteins into and out of the nucleus and is critical to many aspects of cellular regulation and health (Nigg, 1997; D'Angelo and Hetzer, 2008; Toyama et al., 2013), including tumor suppression (Pinkston-Gosse and Kenyon, 2007). mRNA is shuttled to the cytoplasm through the NPC (Cole and Scarcelli, 2006), and nuclear trafficking decreases with cellular senescence, leading to hyporesponsiveness to cellular stresses (Kim et al., 2010). NPC proteins are long lived (Rout et al., 2000), rendering them susceptible to age-related damage. Progressive degradation of nucleoporins further contributes to aging through leaking of proteins and messages (D'Angelo et al., 2009).

Organization inside the nucleus is also important for cellular health. Incorrect organization of lamins at the nuclear envelope (Muchir et al., 2000; Mounkes et al., 2003), which spatially organize the genome, cause laminopathies, including "premature-aging" diseases (Broers et al., 2006); for example, Hutchinson–Gilford progeria patients display extreme aging phenotypes while very young (Scaffidi and Misteli, 2005). Genome instability caused by laminopathies renders DNA sensitive to damaging agents, causing higher rates of breaks, relocations, and aneuploidies (Liu et al., 2005a). Proper

regulation of nuclear lamins is essential for maintenance of tissues in healthy adults (Mounkes et al., 2003; Hutchison and Worman, 2004), and lamin damage results in increased sensitivity to reactive oxygen species (ROS; Pekovic et al., 2011), leading to oxidative damage (Pekovic et al., 2011). Laminopathy-driven altered nuclear architectures are also observed in patients with cardiomyopathies (Muchir et al., 2004; Nikolova-Krstevski et al., 2011) and in aged (Zuo et al., 2012) and damaged (Park et al., 2011) stem cells.

PROTEIN TRANSLATION

Protein translation is a critical control mechanism in longevity regulation; down-regulation of translation upon reduced nutrient availability (Evans et al., 2011) extends lifespan in many organisms (Syntichaki et al., 2007), including worms (Hansen et al., 2007) and flies (Partridge et al., 2011), via TOR signaling in dietary restriction (DR) regimes (Hansen et al., 2007; Katewa and Kapahi, 2011) and IIS/FOXO signaling (Hansen et al., 2007). Loss of the *C. elegans* eukaryotic initiation factor 4F (eIF-4F)/ife-2 extends lifespan (Hansen et al., 2007), as does loss of ribosomal-protein S6 kinase (S6K)/rsks-1 (Hansen et al., 2007). Loss of TOR signaling, eIF-4E/ife-2, or S6K/rsks-1 also increases heat stress resistance (Hansen et al., 2007).

PROTEOSTASIS

Maintenance of protein quality, or proteostasis, is critical for the health and longevity of the cell. Proteostasis ensures a supply of high-quality protein by culling misfolded and damaged proteins from the cellular pool and replacing them with newly formed proteins (Powers et al., 2009). Molecular chaperones direct amino acid chains to the correctly folded state (Hartl and Hayer-Hartl, 2002; Dobson, 2003; Kim et al., 2013), direct misfolded proteins to degradation pathways (Bukau et al., 2006; Hartl et al., 2011), and refold misfolded proteins (Wickner et al., 1999) before they enter the wider population of cellular proteins. Disruption of proteostasis alters the proteome (David et al., 2010; Walther et al., 2015), is hastened by stress (Ben-Zvi et al., 2009), and signals organismal aging in C. elegans (Ben-Zvi et al., 2009; David et al., 2010) but is forestalled by HSF-1 and DAF-16/FOXO activity (Ben-Zvi et al., 2009). (IS and FOXO are critical regulators of healthy aging across eukaryotes (Demontis and Perrimon, 2010; Haigis and Yankner, 2010), increasing resistance to oxygenation damage (Adachi et al., 1998), decreasing accumulation of carbonylated proteins (Adachi et al., 1998), and slowing proteome decline (Walther et al., 2015). Chaperones, including small heat shock proteins, as well as superoxide dismutase and catalase, are upregulated in long-lived daf-2 mutants by DAF-16/FOXO (Murphy et al., 2003) and HSF-1 (Hsu et al., 2003; Morley and Morimoto, 2004; Ben-Zvi et al., 2009). Increased proteostasis is necessary for the longevity of daf-2 mutants (Matilainen et al., 2013). Protein aggregation is decreased in human cells with reduced IIS (O'Neill et al., 2012), and reducing IGF-1 signaling in mice slows the onset of dysregulation of proteostasis, increasing healthy aging (Ben-Zvi et al., 2009).

Many age-related diseases, including Parkinson's disease, Alzheimer's disease, and amyotrophic lateral sclerosis, are associated with aggregated and misfolded proteins. Conformational defects allow the seeding of protein aggregates (Polymeropoulos et al., 1997), which can expand into macroscopic plaques (Jarrett et al., 1993), leading to tissue degeneration. Thus proper regulation of proteostasis is strongly correlated with healthy aging (Balch et al., 2008).

UNFOLDED PROTEIN RESPONSE

The unfolded protein response (UPR) monitors quality of unfolded amino acid chains primarily in the endoplasmic reticulum (ER), where much of translation occurs (Ron and Walter, 2007), and is

linked with ER-associated protein quality regulation processes, particularly the ER-associated degradation (ERAD) pathway (Travers et al., 2000). UPR dysregulation is linked to several diseases (Walter and Ron, 2011), especially age-related protein conformational diseases such as Alzheimer's and Parkinson's (Yoshida, 2007). Low nutrient sensing through the TOR pathway (Vellai et al., 2003), rapamycin treatment (Harrison et al., 2009), and low insulin signaling regulate the UPR (Jia et al., 2004), whereas the protein/histone deacetylase SIR-2.1 acts in insulin signaling-dependent (Berdichevsky et al., 2006) and -independent manners to regulate the UPR (Viswanathan et al., 2005). (In C. elegans IIS mutants, the ER stress response is up-regulated by XBP-1 and the DAF-16/FOXO (Henis-Korenblit et al., 2010), leading to better clearance of damaged proteins by UPR. ER stress in mouse fibroblasts also increases the UPR through Xbp-1 (Akha et al., 2011).

AUTOPHAGY

In much the same way as proteostasis is maintained through degradation and resynthesis of proteins, organelles are consumed by the cell through autophagy (Klionsky and Emr, 2000). In macroautophagy, the damaged organelle is encased in a double membrane to form the autophagosome, which traffics to the lysosome, where the organelle is broken down (Cesen et al., 2012). In microautophagy, cytosol is taken up in bulk directly to the lysosome (Massey et al., 2006), and in chaperone-mediated autophagy (CMA), specific proteins are targeted for transport to the lysosome (Cuervo et al., 2005; Massey et al., 2006; Mizushima et al., 2008).

Autophagy is required for longevity paradigms across many species (Gelino and Hansen, 2012), including DR (Jia and Levine, 2007; Hansen et al., 2008), and IIS mutants (Melendez et al., 2003), and inhibition of autophagy generates hallmarks of aging at an accelerated rate (Cuervo et al., 2005; Rubinsztein et al., 2011). Dependence on autophagy for longevity suggests that autophagic clearing of damaged proteins, protein aggregates, organelles, lipids, and other cargo is required to provide new raw material (Kenyon, 2010b) for a healthy cell.

Autophagy plays a complex role in neurodegenerative disease as well (Nixon, 2013). In Parkinson's disease brains, autophagy is down-regulated (Alvarez-Erviti et al., 2010) and CMA is dysregulated (Massey et al., 2006). However, autophagy is up-regulated in the brains of patients with Alzheimer's disease (Lipinski et al., 2010) and ALS (Sasaki, 2011). Regulatory differences occur at different steps in the autophagy pathway, suggesting a complex role of autophagy in maintaining health.

MITOCHONDRIAL FUNCTION, BIOGENESIS, AND MITOPHAGY

Maintenance of mitochondrial function is critical for cellular and organismal health, due to both the critical nature of their primary role in energy generation for the cell (McBride et al., 2006) and toxic by-products and side and intermediate products of energy generation (Richter et al., 1988), generating free radicals that cause damage (Harman, 1972). Mitochondria have their own mechanisms for proteostasis and regulation of unfolded proteins (Haynes and Ron, 2010), through chaperones and ubiquitin-mediated degradation (Kriegenburg et al., 2012). Mitophagy (mitochondrial autophagy) and mitochondrial biogenesis are coordinated and extended with age in *C. elegans daf-2* mutants (Pinkston-Gosse and Kenyon, 2007) through the activity of DCT-1, a DAF-16/FOXO target, and the Nrf2 homologue SKN-1 (Palikaras et al., 2015).

Defects in the oxidative phosphorylation chain extend lifespan in *C. elegans* (Kayser et al., 2004) and mice (Lapointe and Hekimi, 2008). Mutants in mitochondrial function, such as the DMQ

hydroxylase clk-1 in C. elegans (Ewbank et al., 1997), affect biological timing, including longevity, development, and reproduction (Branicky et al., 2000; Stepanyan et al., 2006), which is also true in mice (Liu et al., 2005b). Mutations in the C. elegans clk-1 gene lead to modest lifespan gains at the expense of small and unhealthy mutants (Stepanyan et al., 2006) in a manner independent of more robust lifespan gains from insulin signaling mutations (Stepanyan et al., 2006). The mechanism is hypothesized to be related to ROS (Harman, 1972) but is incompletely understood (Shibata et al., 2003). However, increased expression of certain electron transport chain (ETC) components increases lifespan in Drosophila (Zid et al., 2009). This extension seems to be linked to the relative expression of different ETC components and the buildup of oxidative intermediates in the mitochondria (Rea et al., 2007). By contrast, C. elegans IIS mutants display healthier organelles overall by well-regulated autophagy (Melendez et al., 2003; Rubinsztein et al., 2011), resulting in overall better health than clk-1 mutants.

Mitochondria have been associated with diseases stemming from direct pathology (Boffoli et al., 1994), aging-related diseases (de Grey, 2004), and neurological diseases such as Alzheimer's and Parkinson's (Devi and Anandatheerthavarada, 2010). Defects in autophagy are linked to multiple myopathies such as Pompe and Danon diseases (Masiero et al., 2009), primarily due to the highly toxic nature of decaying mitochondria (Wu et al., 2009) enriched in mammalian muscle (Wenz et al., 2009; Wu et al., 2009). Beyond the consequences of disruption of energy production, mitochondrial decay puts the cell at risk of leaking ROS and mitochondria-specific enzymes into the cytosol, generating stress that may lead to apoptosis (Green et al., 2011).

CYTOSKELETAL INTEGRITY

The cytoskeleton is critical in maintaining cell shape and integrity, and its dysregulation is an indicator of cellular aging (Gourlay and Ayscough, 2005). (In yeast, actin turnover increases with increased ROS (Gourlay et al., 2004) and disruption of the Ras pathway with age (Ho and Bretscher, 2001). ROS (Gourlay and Ayscough, 2005), ischemia (Genesca et al., 2006), ultraviolet treatment (Kulms et al., 2002), or introduction of a toxin (Korsnes et al., 2007) can lead to cytoskeletal stress, which activates apoptosis (Ashkenazi and Dixit, 1998). The SM22 actin filament cross-linking protein (Prinjha et al., 1994) has been identified as a biomarker of aging across a range of organisms, including yeast, *Drosophila*, and humans (Prinjha et al., 1994; Camoretti-Mercado et al., 1998). (In worms, loss of pat-10/troponin leads to cytoskeletal collapse, whereas overexpression leads to enhanced cytoskeletal stability and stress resistance (Baird et al., 2014).

Cytoskeletal disruptions can cause degenerative neural diseases characteristic of aging. The apolipoprotein E4 (apoE4) is a risk indicator for early-onset Alzheimer's disease in humans (Sando et al., 2008). ApoE4 is proteolyzed in neurons, forming toxic fragments that interact with the actin cytoskeleton (Mahley et al., 2006), hastening cell aging and oxidative stress (Aksenov et al., 2001), and eventually triggering decline and apoptosis. Par-4, which is increased in neurons of Alzheimer's patients (Guo et al., 1998), interacts with Dlk to form stress fibers and cause apoptosis (DiazMeco et al., 1996; Sells et al., 1997; Vetterkind et al., 2005). Hyperphosphorylation of the microtubule-associated tau protein leads to formation of neurofibrillary tangles (Braak and Braak, 1991), another hallmark of Alzheimer's disease, and inhibits proper proteasome activity (Keller et al., 2000a,b), allowing tangles to grow unchecked (Keck et al., 2003).

THE CELL MEMBRANE AND THE EXTRACELLULAR MATRIX

Extracellular signals pass into and out of the cell through the membrane, providing critical context for the health of the cell. Cells acquire geometric aberrations, which are driven by degradation and stress on the cytoskeleton (Kulms et al., 2002; Vetterkind et al., 2005), as they age (Kulms et al., 2002; Vetterkind et al., 2005), which can change surface texture in aging erythrocytes (Girasole et al., 2010). Cell signaling molecule recycling in the cell membrane becomes dysregulated with age (Meissner et al., 2004), leading to disruption of longevity-promoting cell signaling pathways (Meissner et al., 2004; Samuelson et al., 2007).

The extracellular matrix (ECM) is an important contributor to health and longevity and is also an indicator of the health inside the cell. Regulation of collagens, an ECM component, is shared from invertebrates to humans (Myllyharju and Kivirikko, 2004). Collagen expression in *C. elegans* declines with age (Tullet et al., 2008; Ewald et al., 2015), and regulation of specific collagens is required for daf-2-mediated lifespan extension (Ewald et al., 2015). Aging humans also experience glycosylation and other proteomic damage to the ECM proteins (Brownlee, 1995; Kristic et al., 2014), a process that is accelerated in type 2 diabetes patients (Evans et al., 2002; Giacco and Brownlee, 2010) due to buildup of oxidative damage products and ROS (Giacco and Brownlee, 2010).

REPLICATIVE AGING, SENESCENCE, AND RENEWAL

In addition to decline of health with time, cells also experience aging linked to the number of divisions they have undergone (replicative aging). S. cerevisiae reproduces by budding a new cell off of the mother cell and can undergo ~26 such divisions before succumbing to detrimental effects of age (Kaeberlein, 2010), whereas the daughter is not limited by the number of previous divisions of the mother cell (Kaeberlein, 2010), due to renewal of its replicative potential. Replicative aging can be modified by disruption of TOR signaling (Kaeberlein et al., 2005), dietary restriction (Kaeberlein et al., 2005), and modifications to intercellular pH (Schleit et al., 2013). During replicative aging, oxidative damage products such as carbonylated proteins and accumulated cellular damage build up in the yeast mother cell (Reverter-Branchat et al., 2004; Unal et al., 2011; Cabiscol et al., 2014) and are retained by the mother cell through a Sir2dependent mechanism, allowing the newborn daughter cell to be born without this hallmark of aging (Aguilaniu et al., 2003; Wood et al., 2004). (Although carbonylated protein clearance also occurs in C. elegans oocytes [Goudeau and Aquilaniu, 2010], this process is not dependent on sir-2 [Viswanathan et al., 2005; Berdichevsky et al., 2006].) Resetting of replicative potential has clear parallels in mammalian and invertebrate gametogenesis. Stem cells undergo asymmetric divisions that segregate new mitochondria to the daughter cell, affecting their ability to maintain "stemness" (Katajisto et al., 2015).

As cells age, they communicate their internal status—DNA damage (Kuilman et al., 2010), oncogene activation (Kuilman et al., 2010), and proteomic dysregulation (Walther et al., 2015)—to their neighbors by the senescence-associated secretory phenotype (SASP; Coppe et al., 2010; Kuilman et al., 2010). SASP has been reported in flies, mice, and humans (Coppe et al., 2008; Neves et al., 2015), but particular SASP profiles vary based on cell type and context (Gu and Kitamura, 2012). SASP regulates tumor formation (Lehmann et al., 2008) and may play a role in regulating organismal aging (Childs et al., 2014), as well as in age-related pathologies (van Deursen, 2014; Demaria et al., 2015), including neurodegenerative aging diseases (Chinta et al., 2015).

Dysregulation of replicative lifespan is characteristic of diseases of aging, particularly cancer (Reya et al., 2001). Stem cells maintain a balance between multipotency and tumorogenicity by careful internal regulation (Thomson et al., 1998; Pittenger et al., 1999) and by sensing external stimuli (Engler et al., 2006). In cells induced to recover their stem cell-like state through coordinated factor expression (Takahashi and Yamanaka, 2006; Takahashi et al., 2007), chromosomal abnormalities quickly arise (Mayshar et al., 2010), as in normally aging cells (Aubert and Lansdorp, 2008) and immortalized cell lines (Landry et al., 2013). This tendency toward tumor formation suggests that aging and senescence plays an important role in limiting the chance of runaway errors that compromise the overall health of the organism.

CONCLUSION

Cellular health is regulated by a number of cell biological processes. Conserved gene-regulatory pathways coordinate separate processes of cellular aging to maintain cellular health. Because cellular health is regulated across a wide range of scales from molecular to cellular and across every spatial division of the cell, the processes of regulating cellular health are linked: poor protein quality leads to defective organelles, defective organelles lead to increased ROS, and increased ROS leads to further low protein quality. Each of these is linked to regulation of aging on the cellular level, ultimately impinging on the control of aging of the whole organism.

The deeply conserved nature of the IIS pathway, the TOR pathway, and dietary restriction–induced longevity across eukaryotes suggests that regulation of aging is fundamental to eukaryotic evolution. The links between longevity, nutrient availability, and reproduction suggest that longevity pathways evolved to link somatic health to delayed reproduction in periods of low nutrient availability (Luo and Murphy, 2011). The robust, healthy-longevity phenotypes of DR and reduced IIS are the result of the systemic, coordinated change of many cell biological processes that together extend longevity.

Understanding the global regulatory processes that control the cell's health will lead to a greater understanding of aging, which may allow us to better treat and prevent aging-related degenerative diseases such as Alzheimer's and Parkinson's, diseases of incorrect senescence or failure to senesce, such as cancer, and slow aging itself, improving quality of life with age.

ACKNOWLEDGMENTS

We thank Malene Hansen and members of the Murphy lab for discussion. C.T.M. is the Director of the Glenn Center for Aging Research at Princeton, and R.D. is supported by National Institutes of Health Grant T32 GM 007388.

REFERENCES

- Adachi H, Fujiwara Y, Ishii N (1998). Effects of oxygen on protein carbonyl and aging in Caenorhabditis elegans mutants with long (age-1) and short (mev-1) life spans. J Gerontol A Biol Sci Med Sci 53, B240–B244.
- Aguilaniu H, Gustafsson L, Rigoulet M, Nystrom T (2003). Asymmetric inheritance of oxidatively damaged proteins during cytokinesis. Science 299, 1751–1753.
- Akha AAS, Harper JM, Salmon AB, Schroeder BA, Tyra HM, Rutkowski DT, Miller RA (2011). Heightened induction of proapoptotic signals in response to endoplasmic reticulum stress in primary fibroblasts from a mouse model of longevity. J Biol Chem 286, 30344–30351.
- Aksenov MY, Aksenova MV, Butterfield DA, Geddes JW, Markesbery WR (2001). Protein oxidation in the brain in Alzheimer's disease. Neuroscience 103, 373–383.
- Alvarez-Erviti L, Rodriguez-Oroz MC, Cooper JM, Caballero C, Ferrer I, Obeso JA, Schapira AHV (2010). Chaperone-mediated autophagy markers in Parkinson disease brains. Arch Neurol 67, 1464–1472.

- Ashkenazi A, Dixit VM (1998). Death receptors: signaling and modulation. Science 281, 1305–1308.
- Aubert G, Lansdorp PM (2008). Telomeres and aging. Physiol Rev 88, 557–579.
- Baird NA, Douglas PM, Simic MS, Grant AR, Moresco JJ, Wolff SC, Yates JR, Manning G, Dillin A (2014). HSF-1-mediated cytoskeletal integrity determines thermotolerance and life span. Science 346, 360–363.
- Balch WE, Morimoto RI, Dillin A, Kelly JW (2008). Adapting proteostasis for disease intervention. Science 319, 916–919.
- Ben-Zvi A, Miller EA, Morimoto RI (2009). Collapse of proteostasis represents an early molecular event in Caenorhabditis elegans aging. Proc Natl Acad Sci USA 106, 14914–14919.
- Berdichevsky A, Viswanathan M, Horvitz HR, Guarente L (2006). C. elegans SIR-2.1 interacts with 14–3-3 proteins to activate DAF-16 and extend life span. Cell 125, 1165–1177.
- Boffoli D, Scacco SC, Vergari R, Solarino G, Santacroce G, Papa S (1994). Decline with age of the respiratory chain activity in human skeletal muscle. Biochim Biophys Acta 1226, 73–82.
- Braak H, Braak E (1991). Neuropathological staging of Alzheimer-related changes. Acta Neuropathol 82, 239–259.
- Branicky R, Benard C, Hekimi S (2000). clk-1, mitochondria, and physiological rates. Bioessays 22, 48–56.
- Broers JLV, Ramaekers FCS, Bonne G, Ben Yaou R, Hutchison CJ (2006). Nuclear lamins: laminopathies and their role in premature ageing. Physiol Rev 86, 967–1008.
- Brownlee M (1995). Advanced protein glycosylation in diabetes and aging. Annu Rev Med 46, 223–234.
- Bukau B, Weissman J, Horwich A (2006). Molecular chaperones and protein quality control. Cell 125, 443–451.
- Cabiscol E, Tamarit J, Ros J (2014). Protein carbonylation: proteomics, specificity and relevance to aging. Mass Spectrom Rev 33, 21–48.
- Camoretti-Mercado B, Forsythe SM, LeBeau MM, Espinosa R, Vieira JE, Halayko AJ, Willadsen S, Kurtz B, Ober C, Evans GA, et al. (1998). Expression and cytogenetic localization of the human SM22 gene (TAGLN). Genomics 49, 452–457.
- Cesen MH, Pegan K, Spes A, Turk B (2012). Lysosomal pathways to cell death and their therapeutic applications. Exp Cell Res 318, 1245–1251.
- Childs BG, Baker DJ, Kirkland JL, Campisi J, van Deursen JM (2014). Senescence and apoptosis: dueling or complementary cell fates? EMBO Rep 15, 1139–1153.
- Chinta SJ, Woods G, Rane A, Demaria M, Campisi J, Andersen JK (2015). Cellular senescence and the aging brain. Exp Gerontol 68, 3–7.
- Cole CN, Scarcelli JJ (2006). Transport of messenger RNA from the nucleus to the cytoplasm. Curr Opin Cell Biol 18, 299–306.
- Coppe JP, Desprez PY, Krtolica A, Campisi J (2010). The senescence-associated secretory phenotype: the dark side of tumor suppression. Annu Rev Pathol 5, 99–118.
- Coppe JP, Patil CK, Rodier F, Sun Y, Munoz DP, Goldstein J, Nelson PS, Desprez PY, Campisi J (2008). Senescence-associated secretory phenotypes reveal cell-nonautonomous functions of oncogenic RAS and the p53 tumor suppressor. PLoS Biol 6, 2853–2868.
- Cuervo AM, Bergamini E, Brunk UT, Droge W, Ffrench M, Terman A (2005). Autophagy and aging—the importance of maintaining "clean" cells. Autophagy 1, 131–140.
- D'Angelo MA, Hetzer MW (2008). Structure, dynamics and function of nuclear pore complexes. Trends Cell Biol 18, 456–466.
- D'Angelo MA, Raices M, Panowski SH, Hetzer MW (2009). Age-dependent deterioration of nuclear pore complexes causes a loss of nuclear integrity in postmitotic cells. Cell 136, 284–295.
- David DC, Ollikainen N, Trinidad JC, Cary MP, Burlingame AL, Kenyon C (2010). Widespread protein aggregation as an inherent part of aging in C. elegans. PLoS Biol 8, 23.
- de Grey A (2004). Mitochondrial mutations in mammalian aging: an overhasty about-turn? Rejuvenation Res 7, 171–174.
- Demaria M, Desprez PY, Campisi J, Velarde MC (2015). Cell autonomous and non-autonomous effects of senescent cells in the skin. J Invest Dermatol 135, 1722–1726.
- Demontis F, Perrimon N (2010). FOXO/4E-BP signaling in Drosophila muscles regulates organism-wide proteostasis during aging. Cell 143, 813–825.
- Devi L, Anandatheerthavarada HK (2010). Mitochondrial trafficking of APP and alpha synuclein: Relevance to mitochondrial dysfunction in Alzheimer's and Parkinson's diseases. Biochim Biophys Acta 1802, 11–19.
- DiazMeco MT, Municio MM, Frutos S, Sanchez P, Lozano J, Sanz L, Moscat J (1996). The product of par-4, a gene induced during apoptosis, interacts selectively with the atypical isoforms of protein kinase C. Cell 86, 777–786.

- Dobson CM (2003). Protein folding and misfolding. Nature 426, 884-890. Effros RB, Dagarag M, Spaulding C, Man J (2005). The role of CD8(+) T-cell replicative senescence in human aging. Immunol Rev 205, 147-157
- Effros RB, Pawelec G (1997). Replicative senescence of T cells: does the Hayflick limit lead to immune exhaustion? Immunol. Today 18, 450-454. Engler AJ, Sen S, Sweeney HL, Discher DE (2006). Matrix elasticity directs
- stem cell lineage specification. Cell 126, 677-689.
- Evans JL, Goldfine ID, Maddux BA, Grodsky GM (2002). Oxidative stress and stress-activated signaling pathways: a unifying hypothesis of type 2diabetes. Endocr Rev 23, 599-622.
- Evans DS, Kapahi P, Hsueh WC, Kockel L (2011). TOR signaling never gets old: aging, longevity and TORC1 activity. Ageing Res Rev 10, 225-237.
- Ewald CY, Landis JN, Abate JP, Murphy CT, Blackwell TK (2015). Dauerindependent insulin/IGF-1-signalling implicates collagen remodelling in longevity. Nature 519, U97-U212.
- Ewbank JJ, Barnes TM, Lakowski B, Lussier M, Bussey H, Hekimi S (1997). Structural and functional conservation of the Caenorhabditis elegans timing gene clk-1. Science 275, 980-983.
- Gelino S, Hansen M (2012). Autophagy—an emerging anti-aging mechanism. J Clin Exp Pathol Suppl 4, 006.
- Genesca M, Sola A, Hotter G (2006). Actin cytoskeleton derangement induces apoptosis in renal ischemia/reperfusion. Apoptosis 11, 563-571.
- Giacco F, Brownlee M (2010). Oxidative stress and diabetic complications. Circ Res 107, 1058-1070.
- Girasole M, Pompeo G, Cricenti A, Longo G, Boumis G, Bellelli A, Amiconi S (2010). The how, when, and why of the aging signals appearing on the human erythrocyte membrane: an atomic force microscopy study of surface roughness. Nanomed Nanotechnol Biol Med 6, 760–768.
- Gomes NMV, Ryder OA, Houck ML, Charter SJ, Walker W, Forsyth NR, Austad SN, Venditti C, Pagel M, Shay JW, Wright WE (2011). Comparative biology of mammalian telomeres: hypotheses on ancestral states and the roles of telomeres in longevity determination. Aging Cell 10, 761-768
- Goudeau J, Aguilaniu H (2010). Carbonylated proteins are eliminated during reproduction in C-elegans. Aging Cell 9, 991-1003.
- Gourlay CW, Ayscough KR (2005). The actin cytoskeleton: a key regulator of apoptosis and ageing? Nat Rev Mol Cell Biol 6, U583-U585
- Gourlay CW, Carpp LN, Timpson P, Winder SJ, Ayscough KR (2004). A role for the actin cytoskeleton in cell death and aging in yeast. J Cell Biol 164, 803-809.
- Green DR, Galluzzi L, Kroemer G (2011). Mitochondria and the autophagy-inflammation-cell death axis in organismal aging. Science 333, 1109-1112
- Gu LB, Kitamura M (2012). Sensitive detection and monitoring of senescence-associated secretory phenotype by SASP-RAP assay. PLoS One
- Guo Q, Fu WM, Xie J, Luo H, Sells SF, Geddes JW, Bondada V, Rangnekar VM, Mattson MP (1998). Par-4 is a mediator of neuronal degeneration associated with the pathogenesis of Alzheimer disease. Nat Med 4, 957-962.
- Haigis MC, Yankner BA (2010). The aging stress response. Mol Cell 40, 333-344.
- Haithcock E, Dayani Y, Neufeld E, Zahand AJ, Feinstein N, Mattout A, Gruenbaum Y, Liu J (2005). Age-related changes of nuclear architecture in Caenorhabditis elegans. Proc Natl Acad Sci USA 102, 16690–16695.
- Hansen M, Chandra A, Mitic LL, Onken B, Driscoll M, Kenyon C (2008). A role for autophagy in the extension of lifespan by dietary restriction in C-elegans. PLos Genet 4, e24.
- Hansen M, Taubert S, Crawford D, Libina N, Lee S-J, Kenyon C (2007). Lifespan extension by conditions that inhibit translation in Caenorhabditis elegans. Aging Cell 6, 95-110.
- Harley CB, Vaziri H, Counter CM, Allsopp RC (1992). The telomere hypothesis of cellular aging. Exp Gerontol 27, 375-382.
- Harman D (1972). The biologic clock—the mitochondria? J Am Geriatr Soc 20, 145-147.
- Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, Nadon NL, Wilkinson JE, Frenkel K, Carter CS, et al. (2009). Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. Nature 460, 392-395.
- Hartl FU, Bracher A, Hayer-Hartl M (2011). Molecular chaperones in protein folding and proteostasis. Nature 475, 324-332.
- Hartl FU, Hayer-Hartl M (2002). Protein folding-molecular chaperones in the cytosol: from nascent chain to folded protein. Science 295, 1852-1858.
- Hayflick L (1965). The limited in vitro lifetime of human diploid cell strains. Exp Cell Res 37, 614-636.

- Hayflick L, Moorhead PS (1961). The serial cultivation of human diploid cell strains. Exp Cell Res 25, 585-621.
- Haynes CM, Ron D (2010). The mitochondrial UPR-protecting organelle protein homeostasis. J Cell Sci 123, 3849-3855.
- Heidinger BJ, Blount JD, Boner W, Griffiths K, Metcalfe NB, Monaghan P (2012). Telomere length in early life predicts lifespan. Proc Natl Acad Sci USA 109, 1743-1748.
- Henis-Korenblit S, Zhang PC, Hansen M, McCormick M, Lee SJ, Cary M, Kenyon C (2010). Insulin/IGF-1 signaling mutants reprogram ER stress response regulators to promote longevity. Proc Natl Acad Sci USA 107,
- Ho J, Bretscher A (2001). Ras regulates the polarity of the yeast actin cytoskeleton through the stress response pathway. Mol Biol Cell 12, 1541-1555.
- Hsu AL, Murphy CT, Kenyon C (2003). Regulation of aging and age-related disease by DAF-16 and heat-shock factor. Science 300, 1142-1145.
- Hutchison CJ, Worman HJ (2004). A-type lamins: guardians of the soma? Nat Cell Biol 6, 1062-1067.
- Jarrett JT, Berger EP, Lansbury PT (1993). The carboxy terminus of the beta-amyloid protein is critical for the seeding of amyloid formation implications for the pathogenesis of alzheimers-disease. Biochemistry 32, 4693-4697
- Jia K, Chen D, Riddle DL (2004). The TOR pathway interacts with the insulin signaling pathway to regulate C. elegans larval development, metabolism and life span. Development 131, 3897-3906.
- Jia KL, Levine B (2007). Autophagy is required for dietary restriction-mediated life span extension in C-elegans. Autophagy 3, 597-599.
- Kaeberlein M (2010). Lessons on longevity from budding yeast. Nature 464, 513-519.
- Kaeberlein M, McVey M, Guarente L (1999). The SIR2/3/4 complex and SIR2 alone promote longevity in Saccharomyces cerevisiae by two different mechanisms. Genes Dev 13, 2570-2580.
- Kaeberlein M, Powers RW, Steffen KK, Westman EA, Hu D, Dang N, Kerr EO, Kirkland KT, Fields S, Kennedy BK (2005). Regulation of yeast replicative life span by TOR and Sch9 in response to nutrients. Science 310. 1193-1196.
- Kaeberlein M, Shamieh LS (2010). The Role of TOR Signaling in Aging, Berlin: Springer-Verlag.
- Katajisto P, Dohla J, Chaffer CL, Pentinmikko N, Marjanovic N, Igbal S, Zoncu R, Chen W, Weinberg RA, Sabatini DM (2015). Asymmetric apportioning of aged mitochondria between daughter cells is required for stemness. Science 348, 340-343.
- Katewa SD, Kapahi P (2011). Role of TOR signaling in aging and related biological processes in Drosophila melanogaster. Exp Gerontol 46, 382-390.
- Kayser EB, Sedensky MM, Morgan PG, Hoppel CL (2004). Mitochondrial oxidative phosphorylation is defective in the long-lived mutant clk-1. J Biol Chem 279, 54479-54486
- Keck S, Nitsch R, Grune T, Ullrich O (2003). Proteasome inhibition by paired helical filament-tau in brains of patients with Alzheimer's disease. J Neurochem 85, 115-122.
- Keller JN, Hanni KB, Markesbery WR (2000a). Impaired proteasome function in Alzheimer's disease. J Neurochem 75, 436-439.
- Keller JN, Hanni KB, Markesbery WR (2000b). Possible involvement of proteasome inhibition in aging: implications for oxidative stress. Mech Ageing Dev 113, 61–70.
- Kenyon CJ (2010a). The genetics of ageing. Nature 464, 504-512.
- Kenyon CJ (2010b). The genetics of ageing [Erratum]. Nature 467, 622.
- Kenyon C, Chang J, Gensch E, Rudner A, Tabtiang R (1993). A C. elegans mutant that lives twice as long as wild-type. Nature 366, 461-464
- Kim SY, Ryu SJ, Ahn HJ, Choi HR, Kang HT, Park SC (2010). Senescencerelated functional nuclear barrier by down-regulation of nucleo-cytoplasmic trafficking gene expression. Biochem Biophys Res Commun 391, 28-32.
- Kim YE, Hipp MS, Bracher A, Hayer-Hartl M, Hartl FU (2013). Molecular chaperone functions in protein folding and proteostasis. Annu Rev Biochem 82, 323-355.
- Klionsky DJ, Emr SD (2000). Cell biology—autophagy as a regulated pathway of cellular degradation. Science 290, 1717-1721.
- Korsnes MS, Hetland DL, Espenes A, Aune T (2007). Cleavage of tensin during cytoskeleton disruption in YTX-induced apoptosis. Toxicol Vitro 21, 9-15
- Kriegenburg F, Ellgaard L, Hartmann-Petersen R (2012). Molecular chaperones in targeting misfolded proteins for ubiquitin-dependent degradation. FEBS J 279, 532-542.

- Kristic J, Vuckovic F, Menni C, Klaric L, Keser T, Beceheli I, Pucic-Bakovic M, Novokmet M, Mangino M, Thaqi K, et al. (2014). Glycans are a novel biomarker of chronological and biological ages. J Gerontol A Biol Sci Med Sci 69, 779–789.
- Kuilman T, Michaloglou C, Mooi WJ, Peeper DS (2010). The essence of senescence. Genes Dev 24, 2463–2479.
- Kulms D, Dussmann H, Poppelmann B, Stander S, Schwarz A, Schwarz T (2002). Apoptosis induced by disruption of the actin cytoskeleton is mediated via activation of CD95 (Fas/APO-1). Cell Death Differ 9, 598–608.
- Landry JJM, Pyl PT, Rausch T, Zichner T, Tekkedil MM, Stutz AM, Jauch A, Aiyar RS, Pau G, Delhomme N, et al. (2013). The genomic and transcriptomic landscape of a HeLa cell line. G3 (Bethesda) 3, 1213–1224.
- Lapointe J, Hekimi S (2008). Early mitochondrial dysfunction in long-lived Mclk1(+/-) mice. J Biol Chem 283, 26217–26227.
- Lee SS, Kennedy S, Tolonen AC, Ruvkun G (2003). DAF-16 target genes that control C-elegans life-span and metabolism. Science 300, 644–647.
- Lehmann BD, Paine MS, Brooks AM, McCubrey JA, Renegar RH, Wang R, Terrian DM (2008). Senescence-associated exosome release from human prostate cancer cells. Cancer Res 68, 7864–7871.
- Lewinska A, Miedziak B, Kulak K, Molon M, Wnuk M (2014). Links between nucleolar activity, rDNA stability, aneuploidy and chronological aging in the yeast Saccharomyces cerevisiae. Biogerontology 15, 289–316.
- Lin K, Dorman JB, Rodan A, Kenyon C (1997). daf-16: an HNF-3/forkhead family member that can function to double the life-span of Caenorhab-ditis elegans. Science 278, 1319–1322.
- Lin K, Hsin H, Libina N, Kenyon C (2001). Regulation of the Caenorhabditis elegans longevity protein DAF-16 by insulin/IGF-1 and germline signaling. Nat Genet 28, 139–145.
- Lipinski MM, Zheng B, Lu T, Yan ZY, Py BF, Ng A, Xavier RJ, Li C, Yankner BA, Scherzer CR, Yuan JY (2010). Genome-wide analysis reveals mechanisms modulating autophagy in normal brain aging and in Alzheimer's disease. Proc Natl Acad Sci USA 107, 14164–14169.
- Liu BH, Wang JM, Chan KM, Tjia WM, Deng W, Guan XY, Huang JD, Li KM, Chau PY, Chen DJ, et al. (2005a). Genomic instability in laminopathy-based premature aging. Nat Med 11, 780–785.
- Liu XX, Jiang N, Hughes B, Bigras E, Shoubridge E, Hekimi S (2005b). Evolutionary conservation of the clk-1-dependent mechanism of longevity: loss of mclk1 increases cellular fitness and lifespan in mice. Genes Dev 19, 2424–2434.
- Lo SJ, Lee CC, Lai HJ (2006). The nucleolus: reviewing oldies to have new understandings. Cell Res 16, 530–538.
- Luo SJ, Murphy CT (2011). Caenorhabditis elegans reproductive aging: regulation and underlying mechanisms. Genesis 49, 53–65.
- Mahley RW, Weisgraber KH, Huang YD (2006). Apolipoprotein E4: a causative factor and therapeutic target in neuropathology including Alzheimer's disease. Proc Natl Acad Sci USA 103, 5644–5651.
- Masiero E, Agatea L, Mammucari C, Blaauw B, Loro E, Komatsu M, Metzger D, Reggiani C, Schiaffino S, Sandri M (2009). Autophagy is required to maintain muscle mass. Cell Metab 10, 507–515.
- Massey AC, Kiffin R, Cuervo AM (2006). Autophagic defects in aging—looking for an "emergency exit"? Cell Cycle 5, 1292–1296.
- Matilainen O, Arpalahti L, Rantanen V, Hautaniemi S, Holmberg CI (2013). Insulin/IGF-1 signaling regulates proteasome activity through the deubiquitinating enzyme UBH-4. Cell Rep 3, 1980–1995.
- Mayshar Y, Ben-David U, Lavon N, Biancotti JC, Yakir B, Clark AT, Plath K, Lowry WE, Benvenisty N (2010). Identification and classification of chromosomal aberrations in human induced pluripotent stem cells. Cell Stem Cell 7, 521–531.
- McBride HM, Neuspiel M, Wasiak S (2006). Mitochondria: more than just a powerhouse. Curr Biol 16, R551–R560.
- McElwee J, Bubb K, Thomas JH (2003). Transcriptional outputs of the Caenorhabditis elegans forkhead protein DAF-16. Aging Cell 2, 111–121.
- Meissner B, Boll M, Daniel H, Baumeister R (2004). Deletion of the intestinal peptide transporter affects insulin and TOR signaling in Caenorhabditis elegans. J Biol Chem 279, 36739–36745.
- Melendez A, Talloczy Z, Seaman M, Eskelinen EL, Hall DH, Levine B (2003). Autophagy genes are essential for dauer development and life-span extension in C-elegans. Science 301, 1387–1391.
- Mizushima N, Levine B, Cuervo AM, Klionsky DJ (2008). Autophagy fights disease through cellular self-digestion. Nature 451, 1069–1075.
- Morley JF, Morimoto RI (2004). Regulation of longevity in Caenorhabditis elegans by heat shock factor and molecular chaperones. Mol Biol Cell
- Mounkes LC, Kozlov S, Hernandez L, Sullivan T, Stewart CL (2003). A progeroid syndrome in mice is caused by defects in A-type lamins. Nature 423, 298–301.

- Muchir A, Bonne G, van der Kool AJ, van Meegen M, Baas F, Bolhuis PA, de Visser M, Schwartz K (2000). Identification of mutations in the gene encoding lamins A/C in autosomal dominant limb girdle muscular dystrophy with atrioventricular conduction disturbances (LGMD1B). Hum Mol Genet 9, 1453–1459.
- Muchir A, Medioni J, Laluc M, Massart C, Arimura T, Van Der Kooi AJ, Desguerre I, Mayer M, Ferrer X, Briault S, et al. (2004). Nuclear envelope alterations in fibroblasts from patients with muscular dystrophy, cardiomyopathy, and partial lipodystrophy carrying lamin A/C gene mutations. Muscle Nerve 30, 444–450.
- Murphy CT, McCarroll SA, Bargmann CI, Fraser A, Kamath RS, Ahringer J, Li H, Kenyon C (2003). Genes that act downstream of DAF-16 to influence the lifespan of Caenorhabditis elegans. Nature 424, 277–284.
- Myllyharju J, Kivirikko KI (2004). Collagens, modifying enzymes and their mutations in humans, flies and worms. Trends Genet 20, 33–43.
- Neves J, Demaria M, Campisi J, Jasper H (2015). Of flies, mice, and men: evolutionarily conserved tissue damage responses and aging. Dev Cell 32. 9–18.
- Nigg EA (1997). Nucleocytoplasmic transport: signals, mechanisms and regulation. Nature 386, 779–787.
- Nikolova-Krstevski V, Leimena C, Xiao XH, Kesteven S, Tan JC, Yeo LS, Yu ZY, Zhang QP, Carlton A, Head S, et al. (2011). Nesprin-1 and actin contribute to nuclear and cytoskeletal defects in lamin A/C-deficient cardiomy
- Nixon RA (2013). The role of autophagy in neurodegenerative disease. Nat Med 19, 983–997.
- Ogg S, Paradis S, Gottlieb S, Patterson GI, Lee L, Tissenbaum HA, Ruvkun G (1997). The Fork head transcription factor DAF-16 transduces insulinlike metabolic and longevity signals in C-elegans. Nature 389, 994–999.
- Olovnikov AM (1996). Telomeres, telomerase, and aging: origin of the theory. Exp Gerontol 31, 443–448.
- O'Neill C, Kiely AP, Coakley MF, Manning S, Long-Smith CM (2012). Insulin and IGF-1 signalling: longevity, protein homoeostasis and Alzheimer's disease. Biochem Soc Trans 40, 721–727.
- Palikaras K, Lionaki E, Tavernarakis N (2015). Coordination of mitophagy and mitochondrial biogenesis during ageing in C. elegans. Nature 521, 525–528.
- Panossian LA, Porter VR, Valenzuela HF, Zhu X, Reback E, Masterman D, Cummings JL, Effros RB (2003). Telomere shortening in T cells correlates with Alzheimer's disease status. Neurobiol Aging 24, 77–84.
- Park MC, Park D, Lee EK, Park T, Lee J (2010). Genomic analysis of the telomeric length effect on organismic lifespan in Caenorhabditis elegans. Biochem Biophys Res Commun 396, 382–387.
- Park NH, Lee CW, Bae JH, Na YJ (2011). Protective effects of amentoflavone on Lamin A-dependent UVB-induced nuclear aberration in normal human fibroblasts. Bioorg Med Chem Lett 21, 6482–6484.
- Partridge L, Alic N, Bjedov I, Piper MDW (2011). Ageing in Drosophila: the role of the insulin/Igf and TOR signalling network. Exp Gerontol 46, 376–381.
- Pekovic V, Gibbs-Seymour I, Markiewicz E, Alzoghaibi F, Benham AM, Edwards R, Wenhert M, von Zglinicki T, Hutchison CJ (2011). Conserved cysteine residues in the mammalian lamin A tail are essential for cellular responses to ROS generation. Aging Cell 10, 1067–1079.
- Pinkston-Gosse J, Kenyon C (2007). DAF-16/FOXO targets genes that regulate tumor growth in Caenorhabditis elegans. Nat Genet 39, 1403–1409.
- Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, Moorman MA, Simonetti DW, Craig S, Marshak DR (1999). Multilineage potential of adult human mesenchymal stem cells. Science 284, 143–147.
- Polymeropoulos MH, Lavedan C, Leroy E, Ide SE, Dehejia A, Dutra A, Pike B, Root H, Rubenstein J, Boyer R, *et al.* (1997). Mutation in the alphasynuclein gene identified in families with Parkinson's disease. Science 276, 2045–2047.
- Powers ET, Morimoto RI, Dillin A, Kelly JW, Balch WE (2009). Biological and chemical approaches to diseases of proteostasis deficiency. Annu Rev Biochem 78, 959–991.
- Prinjha RK, Shapland CE, Hsuan JJ, Totty NF, Mason IJ, Lawson D (1994). Cloning and sequencing of cdnas encoding the actin cross-linking protein transgelin defines a new family of actin-associated proteins. Cell Motil Cytoskeleton 28, 243–255.
- Rea SL, Ventura N, Johnson TE (2007). Relationship between mitochondrial electron transport chain dysfunction, development, and life extension in Caenorhabditis elegans. PLoS Biol 5, 2312–2329.
- Reverter-Branchat G, Cabiscol E, Tamarit J, Ros J (2004). Oxidative damage to specific proteins in replicative and chronological-aged Saccharomyces cerevisiae—common targets and prevention by calorie restriction. J Biol Chem 279, 31983–31989.

- Reya T, Morrison SJ, Clarke MF, Weissman IL (2001). Stem cells, cancer, and cancer stem cells. Nature 414, 105–111.
- Richter C, Park JW, Ames BN (1988). Normal oxidative damage to mitochondrial and nuclear-DNA is extensive. Proc Natl Acad Sci USA 85, 6465–6467.
- Richter T, von Zglinicki T (2007). A continuous correlation between oxidative stress and telomere shortening in fibroblasts. Exp Gerontol 42, 1039–1042.
- Robida-Stubbs S, Glover-Cutter K, Lamming DW, Mizunuma M, Narasimhan SD, Neumann-Haefelin E, Sabatini DM, Blackwell TK (2012). TOR signaling and rapamycin influence longevity by regulating SKN-1/Nrf and DAF-16/FoxO. Cell Metab 15, 713–724.
- Ron D, Walter P (2007). Signal integration in the endoplasmic reticulum unfolded protein response. Nat Rev Mol Cell Biol 8, 519–529.
- Rout MP, Aitchison JD, Suprapto A, Hjertaas K, Zhao YM, Chait BT (2000). The yeast nuclear pore complex: composition, architecture, and transport mechanism. J Cell Biol 148, 635–651.
- Rubinsztein DC, Marino G, Kroemer G (2011). Autophagy and aging. Cell 146, 682–695.
- Samuelson AV, Carr CE, Ruvkun G (2007). Gene activities that mediate increased life span of C. elegans insulin-like signaling mutants. Genes Dev 21, 2976–2994.
- Sando SB, Melquist S, Cannon A, Hutton ML, Sletvold O, Saltvedt I, White LR, Lydersen S, Aasly JO (2008). APOE epsilon 4 lowers age at onset and is a high risk factor for Alzheimer's disease; a case control study from central Norway. BMC Neurol 8, 7.
- Sasaki S (2011). Autophagy in spinal cord motor neurons in sporadic amyotrophic lateral sclerosis. J Neuropathol Exp Neurol 70, 349–359.
- Scaffidi P, Misteli T (2005). Reversal of the cellular phenotype in the premature aging disease Hutchinson-Gilford progeria syndrome. Nat Med 11, 440–445.
- Schleit J, Johnson SC, Bennett CF, Simko M, Trongtham N, Castanza A, Hsieh EJ, Moller RM, Wasko BM, Delaney JR, et al. (2013). Molecular mechanisms underlying genotype-dependent responses to dietary restriction. Aging Cell 12, 1050–1061.
- Sells SF, Han SS, Muthukkumar S, Maddiwar N, Johnstone R, Boghaert E, Gillis D, Liu GH, Nair P, Monnig S, et al. (1997). Expression and function of the leucine zipper protein par-4 in apoptosis. Mol Cell Biol 17, 3823–3832
- Shalev C, Entringer S, Wadhwa PD, Wolkowitz OM, Puterman E, Lin J, Epel ES (2013). Stress and telomere biology: a lifespan perspective. Psychoneuroendocrinology 38, 1835–1842.
- Shay JW, Wright WE (2000). Hayflick, his limit, and cellular ageing. Nat Rev Mol Cell Biol 1, 72–76.
- Shibata Y, Branicky R, Landaverde IO, Hekimi S (2003). Redox regulation of germline and vulval development in Caenorhabditis elegans. Science 302, 1779–1782.
- Stepanyan Z, Hughes B, Cliche DO, Camp D, Hekimi S (2006). Genetic and molecular characterization of CLK-1/mCLK1 a conserved determinant of the rate of aging. Exp Gerontol 41, 940–951.
- Syntichaki P, Troulinaki K, Tavernarakis N (2007). Protein synthesis is a novel determinant of aging in Caenorhabditis elegans. In: Molecular Mechanisms and Models of Aging, Vol. 1119, ed. ES Gonos, IP Trougakos, and N Chondrogianni, Oxford, UK: Blackwell, 289–295.
- Takahashi K, Tanabe K, Ohnuki M, Narita M, Ichisaka T, Tomoda K, Yamanaka S (2007). Induction of pluripotent stem cells from adult human fibroblasts by defined factors. Cell 131, 861–872.
- Takahashi K, Yamanaka S (2006). Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. Cell 126, 663–676.

- Tepper RG, Ashraf J, Kaletsky R, Kleemann G, Murphy CT, Bussemaker HJ (2013). PQM-1 complements DAF-16 as a key transcriptional regulator of DAF-2-mediated development and longevity. Cell 154, 676–690.
- Thomson JA, Itskovitz-Eldor J, Shapiro SS, Waknitz MA, Swiergiel JJ, Marshall VS, Jones JM (1998). Embryonic stem cell lines derived from human blastocysts. Science 282, 1145–1147.
- Toyama BH, Savas JN, Park SK, Harris MS, Ingolia NT, Yates JR, Hetzer MW (2013). Identification of long-lived proteins reveals exceptional stability of essential cellular structures. Cell 154, 971–982.
- Travers KJ, Patil CK, Wodicka L, Lockhart DJ, Weissman JS, Walter P (2000). Functional and genomic analyses reveal an essential coordination between the unfolded protein response and ER-associated degradation. Cell 101, 249–258.
- Tullet JMA, Hertweck M, An JH, Baker J, Hwang JY, Liu S, Oliveira RP, Baumeister R, Blackwell TK (2008). Direct inhibition of the longevitypromoting factor SKN-1 by insulin-like signaling in C. elegans. Cell 132, 1025–1038.
- Unal E, Kinde B, Amon A (2011). Gametogenesis eliminates age-induced cellular damage and resets life span in yeast. Science 332, 1554–1557.
- van Deursen JM (2014). The role of senescent cells in ageing. Nature 509, 439–446.
- Vellai T, Takacs-Vellai K, Zhang Y, Kovacs AL, Orosz L, Muller F (2003). Genetics— influence of TOR kinase on lifespan in C-elegans. Nature 426, 620–620
- Vera E, de Jesus BB, Foronda M, Flores JM, Blasco MA (2012). The rate of increase of short telomeres predicts longevity in mammals. Cell Rep 2, 732–737
- Vetterkind S, Illenberger S, Kubicek J, Boosen M, Appel S, Naim HY, Scheidtmann KH, Preuss U (2005). Binding of Par-4 to the actin cytoskeleton is essential for Par-4/Dlk-mediated apoptosis. Exp Cell Res 305, 392–408
- Viswanathan M, Kim SK, Berdichevsky A, Guarente L (2005). A role for SIR-2.1 regulation of ER stress response genes in determining C-elegans life span. Dev Cell 9, 605–615.
- Walter P, Ron D (2011). The unfolded protein response: from stress pathway to homeostatic regulation. Science 334, 1081–1086.
- Walther DM, Kasturi P, Zheng M, Pinkert S, Vecchi G, Ciryam P, Morimoto RI, Dobson CM, Vendruscolo M, Mann M, Hartl FU (2015). Widespread proteome remodeling and aggregation in aging C-elegans. Cell 161, 919–932.
- Wenz T, Rossi SG, Rotundo RL, Spiegelman BM, Moraes CT (2009). Increased muscle PGC-1 alpha expression protects from sarcopenia and metabolic disease during aging. Proc Natl Acad Sci USA 106, 20405–20410.
- Wickner S, Maurizi MR, Gottesman S (1999). Posttranslational quality control: folding, refolding, and degrading proteins. Science 286, 1888–1893.
- Wood JG, Rogina B, Lavu S, Howitz K, Helfand SL, Tatar M, Sinclair D (2004). Sirtuin activators mimic caloric restriction and delay ageing in metazoans. Nature 430, 686–689.
- Wu JJ, Quijano C, Chen E, Liu HJ, Cao L, Fergusson MM, Rovira II, Gutkind S, Daniels MP, Komatsu M, Finkel T (2009). Mitochondrial dysfunction and oxidative stress mediate the physiological impairment induced by the disruption of autophagy. Aging (Albany NY) 1, 425–437.
- Yoshida H (2007). ER stress and diseases. FEBS J 274, 630-658.
- Zid BM, Rogers AN, Katewa SD, Vargas MA, Kolipinski MC, Lu TA, Benzer S, Kapahi P (2009). 4E-BP extends lifespan upon dietary restriction by enhancing mitochondrial activity in Drosophila. Cell 139, 149–160.
- Zuo BF, Yang J, Wang F, Wang L, Yin Y, Dan JM, Liu N, Liu L (2012). Influences of lamin A levels on induction of pluripotent stem cells. Biol Open 1, 1118–1127.

Volume 26 December 15, 2015 Cell biology of aging | 4531