

Machine-learning accelerated geometry optimization in molecular simulation support information

There are three parts in this support information:

- Example to use `NN_ensemble_relaxer` for molecule geometry optimization
- Figures for configurations used as the example in the manuscript
- Code to access the datasets for reproducing the main figures in the manuscript

1 Example of NN_ensemble_relaxer utilization

1.1 Installation

We first download the NN_ensemble_relaxer repo from the github and compile the required file. The module to calculate the symmetry function is modified based on the functions from SimpleNN.[?]

```
1 git clone https://github.com/yilinyang1/NN_ensemble_relaxer.git
2 cd utils && python libsymb_builder
```

1.2 Instantiate NN_ensemble_relaxer and run the relaxation

Assume we have a ASE database called db contains the configurations need to be optimized, we could just feed it into the NN_ensemble_relaxer to conduct the relaxation.

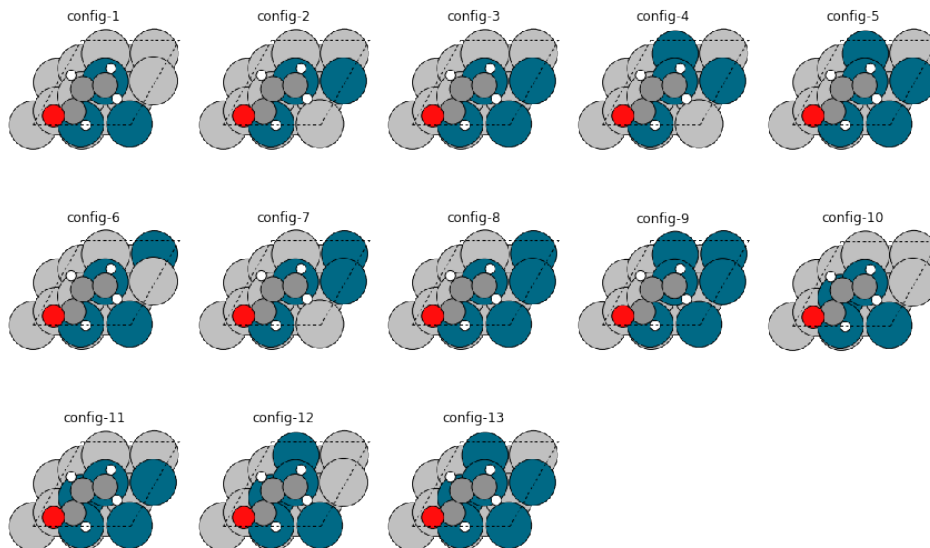
```
1 from nn_optimize import Ensemble_Relaxer
2
3 # NN hyperparameters
4 nn_params = {'layer_nodes': [40, 40], 'activations': ['tanh', 'tanh'], 'lr': 1}
5
6 # confidence coefficient used to control in what extend we trust the NN model
7 alpha = 2.0
8
9 # feed ASE database db, set ground truth calculator,
10 # specify the folder name to store intermediate models and data
11 relaxer = Ensemble_Relaxer(db=db, calculator=EMT(), jobname='AuPd-nano-test',
12                             ensemble_size=10, alpha=alpha, nn_params=nn_params)
13
14 # relaxer.run() returns a ase-db containing relaxed configurations
15 relaxed_db = relaxer.run(fmax=0.05, steps=50)
```

Above is a basic example, more details could be found in the github repo.

2 Figures for configurations used as the example in the manuscript

In second part of the result section, we use 13 Acrolein/AgPd configurations as the example to show the advantage of multiple configurations during geometry optimization. Here, we provides the figures for these 13 configurations.

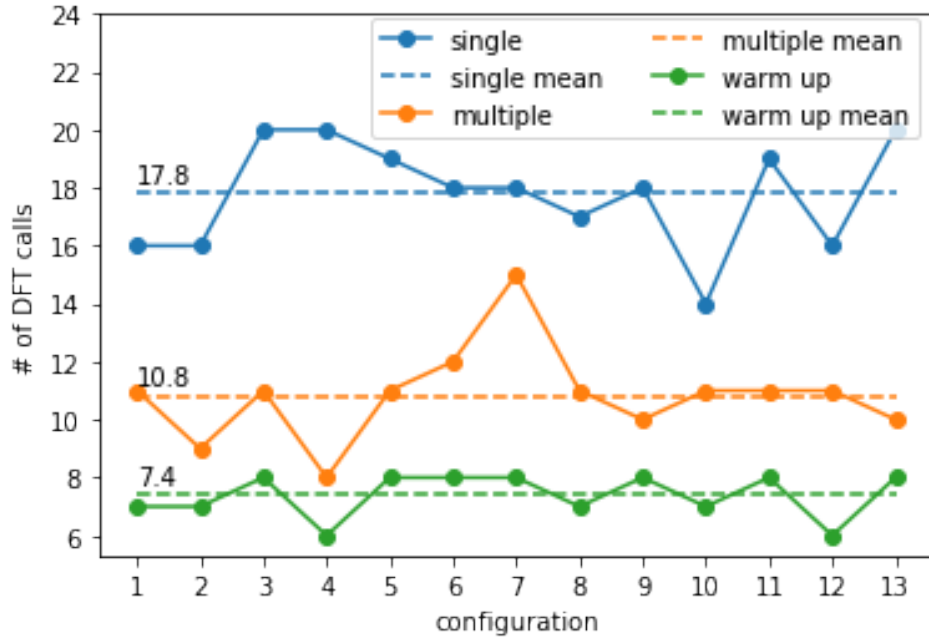
```
1 from ase.db import connect
2 from ase.io import write
3 from ase.visualize import view
4 import matplotlib.image as mpimg
5 from matplotlib import pyplot as plt
6
7 path = 'Acrolein-AgPd-single-multiple-configs'
8 data_path = f'./{path}/initial-configs.db'
9 db = connect(data_path)
10 i = 1
11 for entry in db.select():
12     write(f'./{path}/images/image-{i}.png', entry.toatoms())
13     i += 1
14
15 fig, axes = plt.subplots(3, 5, figsize=(12, 8))
16 for i in range(15):
17     row, col = i // 5, i % 5
18     if i < 13:
19         tmp_image = mpimg.imread(f'./{path}/images/image-{i+1}.png')
20         axes[row][col].imshow(tmp_image)
21         axes[row][col].set_title(f'config-{i+1}')
22         axes[row][col].axis('off')
23 fig.tight_layout()
```



3 Code to access the datasets for reproducing the main figures in the manuscript

3.1 Acceleration of the geometry optimization for Acrolein/AgPd

```
1 from ase.io.trajectory import Trajectory
2 import numpy as np
3
4 single_steps = []
5 multi_steps = []
6 warm_steps = []
7
8 path = 'Acrolein-AgPd-single-multiple-configs'
9 sub_path1 = 'single-config-scratch-trajs'
10 sub_path2 = 'multi-config-scratch-trajs'
11 sub_path3 = 'multi-config-warmup-trajs'
12
13
14 for i in range(13):
15     single_traj = Trajectory(f'./{path}/{sub_path1}/config-{i+1}.traj')
16     single_steps.append(len(single_traj))
17     multi_traj = Trajectory(f'./{path}/{sub_path2}/config-{i+1}.traj')
18     multi_steps.append(len(multi_traj))
19     warm_traj = Trajectory(f'./{path}/{sub_path3}/config-{i+1}.traj')
20     warm_steps.append(len(warm_traj))
21
22 steps = [single_steps, multi_steps, warm_steps]
23 labels = ['single', 'multiple', 'warm up']
24 mean_labels = ['single mean', 'multiple mean', 'warm up mean']
25 xs = range(1, 14)
26
27
28 fig = plt.figure(figsize=(6, 4))
29 ax = fig.add_subplot(111)
30 for i in range(3):
31     ax.plot(xs, steps[i], '-o', label = labels[i])
32     means = [round(np.mean(steps[i]), 1)] * len(xs)
33     ax.plot(xs, means, '--', color=f'C{i}', label = mean_labels[i])
34     ax.text(1, means[0] + 0.4, str(means[0]))
35
36 ax.legend(bbox_to_anchor=(0.31, 0.75), ncol=2)
37 ax.set_xticks(range(1, 14))
38 ax.set_yticks(range(6, 25, 2))
39 ax.set_xlabel('configuration')
40 ax.set_ylabel('# of DFT calls')
```



3.2 Performance for various systems

```

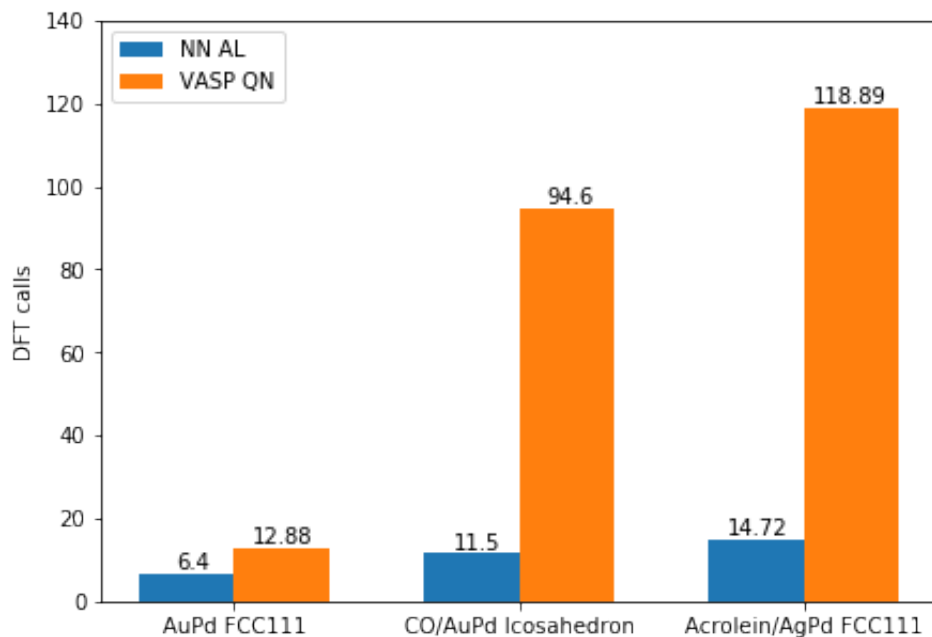
1 from matplotlib import pyplot as plt
2 import pickle
3 import numpy as np
4
5 path = 'more-geometry-optimization-data'
6
7 with open(f'./{path}/AuPd-slab-vasp-data-25.pkl', 'rb') as f:
8     slab_vasp_data = pickle.load(f)
9
10 with open(f'./{path}/AuPd-slab-nn-data-25.pkl', 'rb') as f:
11     slab_nn_data = pickle.load(f)
12
13 with open(f'./{path}/Acrolein-AgPd-vasp-data-100.pkl', 'rb') as f:
14     ads_vasp_data = pickle.load(f)
15
16 with open(f'./{path}/Acrolein-AgPd-nn-data-scratch-100.pkl', 'rb') as f:
17     ads_nn_data = pickle.load(f)
18
19 with open(f'./{path}/CO-AuPd-Ico-vasp-data-10.pkl', 'rb') as f:
20     nano_vasp_data = pickle.load(f)
21
22 with open(f'./{path}/CO-AuPd-Ico-nn-data-10.pkl', 'rb') as f:
23     nano_nn_data = pickle.load(f)
24
25 ads_nn_step_mean = np.mean(ads_nn_data['steps'])
26 ads_vasp_step_mean = np.mean(ads_vasp_data['steps'])
27
28 slab_nn_step_mean = np.mean(slab_nn_data['steps'])
29 slab_vasp_step_mean = np.mean(slab_vasp_data['steps'])
30

```

```

31 nano_nn_step_mean = np.mean(nano_nn_data['steps'])
32 nano_vasp_step_mean = np.mean(nano_vasp_data['steps'])
33
34 step_mean = np.array([slab_nn_step_mean, slab_vasp_step_mean, nano_nn_step_mean,
35                       nano_vasp_step_mean, ads_nn_step_mean, ads_vasp_step_mean])
36
37 nn_steps = step_mean[[0, 2, 4]]
38 vasp_steps = step_mean[[1, 3, 5]]
39 nn_xs = [1, 2.5, 4]
40 vasp_xs = [1.5, 3, 4.5]
41
42 fig = plt.figure(figsize=(7, 5))
43 ax = fig.add_subplot(111)
44 ax.bar(nn_xs, nn_steps, width=0.5, label='NN AL')
45 ax.bar(vasp_xs, vasp_steps, width=0.5, label='VASP QN')
46 ax.set_ylabel('DFT calls')
47 ax.set_yticks(range(0, 160, 20))
48 ax.set_xticks([1.25, 2.75, 4.25])
49 ax.set_xticklabels(['AuPd FCC111', 'CO/AuPd Icosahedron', 'Acrolein/AgPd FCC111'])
50 ax.legend(loc='upper left')
51 ax.text(0.95, 7.5, str(nn_steps[0]))
52 ax.text(1.35, vasp_steps[0] + 1.1, str(vasp_steps[0]))
53 ax.text(2.4, nn_steps[1] + 1.1, str(nn_steps[1]))
54 ax.text(2.9, vasp_steps[1] + 1.1, str(vasp_steps[1]))
55 ax.text(3.85, nn_steps[2] + 1.1, str(nn_steps[2]))
56 ax.text(4.3, vasp_steps[2] + 1.1, str(vasp_steps[2]))

```



```

1  from ase.db import connect
2  from matplotlib import pyplot as plt
3
4  init_db = connect('./Acetylene-hydrogenation-NEB/Acetylene-hydro-initial-configs.db')
5  vasp_db = connect('./Acetylene-hydrogenation-NEB/Acetylene-hydro-vasp-cnvg.db')
6  nn_db = connect('./Acetylene-hydrogenation-NEB/Acetylene-hydro-nn-cnvg.db')
7
8  init_nrgs = [entry.energy for entry in init_db.select()]
9  vasp_nrgs = [entry.energy for entry in vasp_db.select()]
10 nn_nrgs = [entry.energy for entry in nn_db.select()]
11 xs = range(len(vasp_nrgs))
12
13 fig = plt.figure()
14 ax = fig.add_subplot(111)
15 ax.plot(xs, nn_nrgs, '-o')
16 ax.plot(xs, vasp_nrgs, '-o')
17 ax.set_xlabel('Reaction coordinate')
18 ax.set_ylabel('energy (eV)')
19 ax.legend(['NN ensemble', 'Vasp'])
20 vasp_act = vasp_nrgs[5] - vasp_nrgs[0]
21 nn_act = nn_nrgs[5] - nn_nrgs[0]
22 ax.text(3.5, -204.6, 'Activation energy:')
23 ax.text(3.5, -204.65, f'NN ensemble: {round(nn_act, 3)} eV')
24 ax.text(3.5, -204.7, f'Vasp: {round(vasp_act, 3)} eV')

```

