

Adaptive evolution with aneuploidy and mutation

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Abstract

Aneuploidy is common in eukaryotes, often leading to decreased cell growth and fitness. However, evidence from yeast and fungi, as well as human tumour cells, suggests that aneuploidy can be beneficial under stressful conditions and lead to elevated growth rates and adaptation. Because aneuploidy differs from mutation in rate, expected effect, and reversibility, it is crucial to develop a quantitative theory for the role of aneuploidy in adaptive evolution. Here, we develop evolutionary models for adaptive evolution with both mutation and aneuploidy. These models are used within an approximate Bayesian computation framework to estimate the formation rate and fitness effect of aneuploidy and mutation from empirical results of experiments in which *Saccharomyces cerevisiae* adapted to heat stress. The experimental population first acquired chromosome duplications, only to later revert back to a euploid state. We also analyze our models to estimate the effect of the aneuploidy and mutation rates on the expected adaptation time and the probability for adaptation via aneuploidy. Our results suggest that aneuploidy is a transient adaptive solution, which can decelerate adaptation in a non-intuitive manner. By creating an evolutionary conflict between the individual and the population, aneuploidy further complicates the process of adaptation in cell populations.

Introduction

Aneuploidy is common in eukaryotes. Aneuploidy is an imbalance in the number of chromosomes in the cell: an incorrect karyotype. Evidence suggests aneuploidy is very common in eukaryotes, e.g. animals (Santaguida and Amon, 2015; Naylor and van Deursen, 2016; Bakhoum and Landau, 2017), and fungi (Pavelka et al., 2010; Zhu et al., 2016; Robbins et al., 2017; Todd et al., 2017). Aneuploidy has been implicated in cancer formation and progression (Boveri, 2008; Schwartzman et al., 2010): 90% of solid tumours and 50% of blood cancers are aneuploid (Santaguida and Amon, 2015). Aneuploidy is also linked to the emergence of drug resistance (Selmecki et al., 2009) and virulence (Möller et al., 2018) in fungal pathogens, which are under-studied (Rodrigues and Albuquerque, 2018) despite infecting close to a billion people per year, causing serious infections and significant morbidity in >150 million people per year and killing >1.5 million people per year (Selmecki et al., 2009; Rodrigues and Albuquerque, 2018). In addition, aneuploidy is common in protozoan pathogens of the *Leishmania* genus, a major global health concern (Mannaert et al., 2012).

Aneuploidy is generally deleterious. The molecular and genetic mechanisms involved in aneuploidy have been explored (Musacchio and Salmon, 2007; Sheltzer and Amon, 2011; Chen et al., 2012; Rancati and Pavelka, 2013; Gerstein et al., 2015; Shor and Perlin, 2015). Experiments with human and mouse embryos found that aneuploidy is usually lethal. It is also associated with developmental defects and lethality in other multicellular organisms (Sheltzer and Amon, 2011). For example, aneuploid mouse embryonic cells grow slower than euploid cells (Williams et al., 2008). Similarly, in unicellular eukaryotes growing in benign conditions, aneuploidy usually leads to slower growth and decreased overall fitness (Niwa et al., 2006; Torres et al., 2007; Pavelka et al., 2010; Sheltzer and Amon, 2011; Kasuga et al., 2016), in part due to proteotoxic stress caused by increased expression in aneuploid cells (Pavelka et al., 2010; Santaguida et al., 2015; Zhu et al., 2018) and hypo-osmotic-like stress (Tsai et al., 2019).

Aneuploidy can lead to adaptation. However, aneuploidy can be beneficial under stressful conditions due to the wide range of phenotypes it can produce, some of which are advantageous (Pavelka et al., 2010). Thus, aneuploidy can lead to rapid adaptation in unicellular eukaryotes (Gerstein et al., 2015; Torres et al., 2010; Hong and Gresham, 2014; Rancati et al., 2008), as well as to rapid growth of somatic tumour cells (Schwartzman et al., 2010; Sheltzer et al., 2017). For example, aneuploidy in *S. cerevisiae* facilitates adaptation to a variety of stressful conditions like heat and pH (Yona et al., 2012), copper (Covo et al., 2014; Gerstein et al., 2015), salt (Dhar et al., 2011), and nutrient limitation (Dunham et al., 2002; Gresham et al., 2008). Importantly, aneuploidy can also lead to drug resistance in pathogenic fungi such as *Candida albicans* (Selmecki et al., 2008, 2010; Gerstein and Berman, 2018) and *Cryptococcus neoformans* (Sionov et al., 2010), which cause candidiasis and meningoencephalitis, respectively.

Transient adaptive solution. Aneuploidy differs from mutation due to its distinct properties. Chromosome duplication usually occurs more often than mutation and on average produces larger fitness effects. Yet, because it affects many genes on a whole chromosome or a chromosome fragment, aneuploidy also carries fitness costs. Thus, aneuploidy can be a *transient adaptive solution*: it can rapidly occur and fix in the population under stressful conditions, and can be rapidly lost when the cost outweighs the benefit—when stress is removed or after beneficial mutations occur. Experimental evidence of such a transient role of aneuploidy was demonstrated by Yona et al. (2012). They evolved populations of *S. cerevisiae* under strong heat or pH stress. The populations adapted to the heat and pH stress within 450 and 150 generations, and this adaptation was determined to be due to

chromosome duplications. Much later, after more than 1500 and 750 generations, for the heat and pH stress, respectively, the populations reverted back to an euploid state, while remaining adapted to the stress and accumulating multiple mutations. However, under gradual heat stress, aneuploidy was not observed. Yona et al. (2012) concluded that aneuploidy serves as a transient adaptive solution, or a “quick fix”, which is expected to facilitate adaptation.

The present study. Here, we develop evolutionary-genetic models that include the effects of natural selection, genetic drift, aneuploidy, and mutation to examine the role of aneuploidy in adaptive evolution. These models follow a population of cells characterised by both their ploidy and their genotype. We fit these models to the experimental results of Yona et al. (2012) using an *approximate Bayesian computation* framework (Sisson et al., 2007; Klinger et al., 2018) to infer model parameters, including selection coefficients and rates of aneuploidy and mutation, and to perform model selection between different models, thereby testing different hypotheses about the evolutionary process. Furthermore, we analyze these models to estimate the effects of parameters on the adaptation time and the probability for adaptation via aneuploidy. We find that

Models and Methods

Evolutionary Models. We developed two models: a single-locus model and a multi-locus model. Both models are based on the Wright-Fisher model (Otto and Day, 2007), assuming non-overlapping generations and including the effects of natural selection, genetic drift, aneuploidy, and mutation. We focus on beneficial mutations, neglecting the effects of deleterious and neutral mutations. Both models allow for a single aneuploid karyotype (e.g., chromosome III duplication). While the single-locus model allows for only a single mutation to occur, the multi-locus model allows for multiple mutations to accumulate in the genome (Figure 1).

Single-locus model. This model assumes a constant population size N and follows four genotypes (Figure 1A): euploid wild-type, $2n$, the initial genotype; euploid mutant, $2n^*$, with the standard karyotype and a single beneficial mutation; aneuploid wild-type, $2n+1$, with an extra chromosome, e.g. following chromosome duplication; and aneuploid mutant, $2n+1^*$, with an extra chromosome and a beneficial mutation.

Transitions between the genotypes occur as follows (Figure 1A): Beneficial mutations from $2n$ to $2n^*$ occur with probability μ , the mutation rate, and from $2n+1$ to $2n+1^*$ with probability $\tau\mu$, where τ is the fold-change in the rate of beneficial mutations in aneuploid cells. By default, we assume $\tau = 33/32$, as the wild-type *S. cerevisiae* strains in the experiments by (Yona et al., 2012) are diploid,

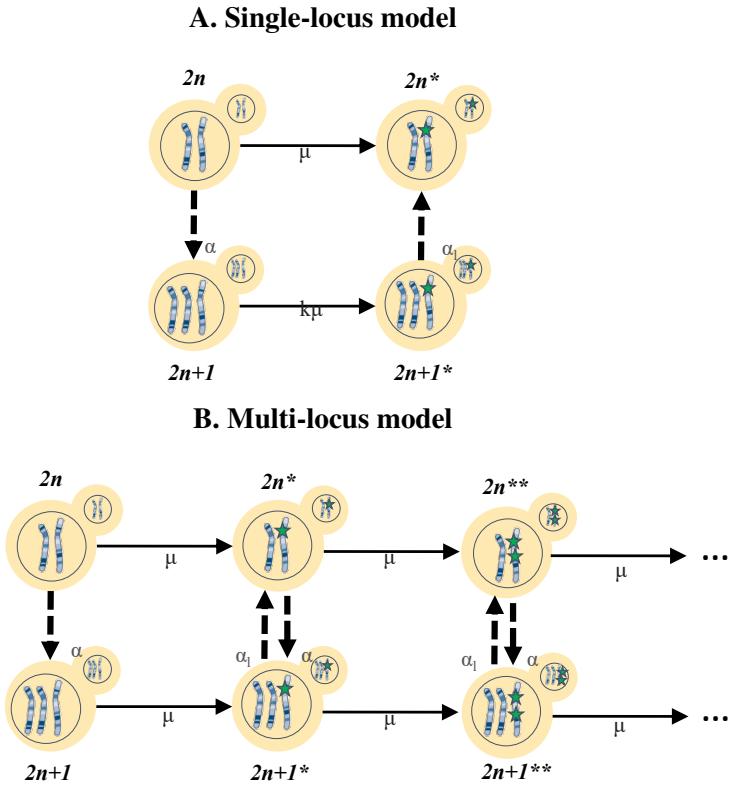


Figure 1: Model illustrations. **(A)** In the single-locus model, the four genotypes are: euploid wild-type, $2n$; euploid mutant, $2n^*$; aneuploid wild-type, $2n+1$; and aneuploid mutant, $2n+1^*$. Overall there are two possible trajectories from $2n$ to $2n^*$. **(B)** In the multi-locus model, each genotype is characterized by its karyotype, $2n$ or $2n+1$, and the number of accumulated beneficial mutations, denoted by stars. In both panels arrows denote transitions between genotypes, with transition rates: μ , mutation rate; δ , aneuploidy gain rate; δ_L , aneuploidy loss rate.

with 32 chromosomes, and the aneuploid strains are trisomic, with 33 chromosomes. Aneuploidy is formed by chromosome mis-segregation, so that cells transition from $2n$ to $2n+1$ with probability δ , the aneuploidy gain rate. Aneuploidy is lost, transitioning cells from $2n+1^*$ to $2n^*$ with probability δ_L , the aneuploidy loss rate. The fitness values of the four genotypes are given by Table 1.

Table 1: Single-locus model fitness values.

Genotype i	$2n$	$2n + 1$	$2n + 1^*$	$2n^*$
Fitness w_i	1	$1 - c + b$	$(1 - c)(1 + s) + b$	$1 + s$

$s \geq 0$ is the selection coefficient of a beneficial mutation; $b \geq 0$ is the selection coefficient of aneuploidy; and $0 \leq c \leq 1$ is the fitness cost of aneuploidy.

The first generation is initialized with N cells with genotype $2n$. The effect of natural selection on the frequency f_i of genotype $i = 2n, 2n + 1, 2n + 1^*$, or $2n^*$ is given by

$$f_i^s = \frac{f_i w_i}{\bar{w}}, \quad (1)$$

where the fitness values w_i are given in Table 1 and $\bar{w} = \sum_j f_j w_j$ is the population mean fitness. The effect of mutation and aneuploidy on genotype frequencies is given by

$$\begin{aligned} f_{2n}^m &= (1 - \delta - \mu) f_{2n}^s, \\ f_{2n+1}^m &= \delta f_{2n}^s + (1 - \tau \mu) f_{2n+1}^s, \\ f_{2n+1^*}^m &= \tau \mu f_{2n+1}^s + (1 - \delta_L) f_{2n+1^*}^s, \\ f_{2n^*}^m &= \mu f_{2n}^s + \delta_L f_{2n+1}^s + f_{2n^*}^s. \end{aligned} \quad (2)$$

Finally, random genetic drift is modeled using a multinomial distribution (Otto and Day, 2007),

$$\mathbf{f}' \sim \frac{1}{N} \text{Mult}(N, \mathbf{f}^m), \quad (3)$$

where $\mathbf{f}^m = (f_{2n}^m, f_{2n+1}^m, f_{2n+1^*}^m, f_{2n^*}^m)$ are the frequencies of the genotypes after mutation and aneuploidy, \mathbf{f}' are the genotype frequencies in the next generation, and $\text{Mult}(N, \mathbf{f})$ is a multinomial distribution parameterized by the population size N and the genotype frequencies \mathbf{f} . Overall, the change in genotype frequencies from one generation to the next is given by the transformation $f_i \rightarrow f'_i$.

Multi-locus model. This model expands the single-locus model by allowing for (i) the accumulation of beneficial mutations in the genome, and (ii) a fluctuating population size.

A genotype is characterized by its karyotype, $2n$ or $2n+1$, and the number of accumulated beneficial mutations, which can be zero or more. The selection coefficient of the i -th accumulated mutation in each individual, s_i , is drawn from an exponential distribution with expected value s , $s_i \sim \text{Exp}(s)$. The rest of the parameters ($N, \mu, \tau, \delta, \delta_L, b, c$) are the same as in the single-locus model. The fitness of the different genotypes is the same as in the single-locus model (Table 1), except that the fitness contribution of k beneficial mutations is the product of their independent effects, $\prod_{i=1}^k (1 + s_i)$, instead of the contribution of the single mutation allowed in the single-locus model, $(1 + s)$, see Table 2. Therefore, aneuploidy loss would be favored by selection only if there are enough beneficial mutations and/or the selection coefficients s_i are large enough. The intuition is that when the benefit of the accumulated beneficial mutations is small, the benefit of aneuploidy has a large effect; when the benefit of the accumulated beneficial mutations benefit is large, then aneuploidy is no longer advantageous because of its significant cost.

In contrast to the single-locus model, in the multi-locus model the population size changes in order to model serial-transfer experiment protocol (Yona et al., 2012): the population is serially diluted by transferring a fraction of the population (1/120) to a fresh medium, starting a new growth cycle. In this model, the population initial size is $N_0 = N$, and the population size is doubled every generation, $N_1 = 2N, N_2 = 4N, \dots$, and diluted back to N every eight generations, $N_8 = N$.

The change in frequencies due to selection is exactly the same as in the single-locus model (Equation 1), only applied using the fitness values in Table 2. The change due to random genetic drift is also the same as in Equation 3, except that the frequencies vector is $\mathbf{f} = (f_{2n}, f_{2n+1}, f_{2n^*}, f_{2n+1^*}, f_{2n^{**}}, f_{2n+1^{**}}, \dots)$ and that the population size changes between generations, as described above.

The effects of mutation and aneuploidy on genotype frequencies is more elaborate than in the single-locus model. Genotype i is classified according to their karyotype ($2n$ or $2n+1$), the number of accumulated beneficial mutations ($k \geq 0$), and their fitness (w_i). Each offspring cell inherits these properties from its mother cell. Then, with probability μ or $\tau\mu$ for euploid and aneuploid cells, respectively, a new beneficial mutation is accumulated, such that the number of mutations is $k + 1$, and its effect s_{k+1} is drawn from an exponential distribution with expected value s , such that the contribution of the mutations to the fitness is $\prod_{j=0}^{k+1} (1 + s_j)$. Next, euploid offspring become aneuploid with probability δ , and aneuploid offspring become euploid with probability δ_L .

Table 2: Multi-locus model fitness values.

Genotype i	$2n$	$2n + 1$	$2n + 1^{*k}$	$2n^{*k}$
Fitness w_i	1	$1 - c + b$	$(1 - c) \prod_{j=1}^k (1 + s_i) + b$	$\prod_{j=1}^k (1 + s_i)$

k is the number of accumulated beneficial mutations in the genome; $s \geq 0$ is the selection coefficient of a beneficial mutation; $b \geq 0$ is the selection coefficient of aneuploidy; and $0 \leq c \leq 1$ is the fitness cost of aneuploidy.

Empirical evidence. Our inference procedure uses empirical data from evolutionary experiments performed by Yona et al. (2012). In the heat-stress experiment, four populations of *S. cerevisiae* evolved under 39 °C. Aneuploidy fixed in all four population in the first 450 generations (hereafter, fixation or elimination of a genotype by generation t means that more than 95% or less than 5% of the population carry the genotype at generation t , and possibly earlier). The experiment continued with two populations, in which aneuploidy was eliminated by generation 1700 and 2350. In the pH-stress experiment, four populations of *S. cerevisiae* evolved under high-pH stress (8.6). Aneuploidy fixed during the first 150 generations. It was fully eliminated in two populations and partly eliminated in two populations by generation 750. These empirical results were published by Yona et al. (2012).

Likelihood function. Denote by A_t the fixation of aneuploidy at generation t , by L_t the loss of aneuploidy at generation t , and by $L_{t,T}^*$ the loss of aneuploidy at generation t conditioned on no loss by generation T . The model likelihood function for parameter vector θ for the heat-stress experiment is

$$\begin{aligned} \mathcal{L}(\theta | A_{450}, L_{1700}, L_{2350,1700}^*) = \\ P^4(A_{450}) \cdot \left(1 - P^4(\neg L_{1700} | A_{450}) - P^4(\neg L_{2350,1700}^* | A_{450}) + P^4(\neg L_{1700} \wedge \neg L_{2350,1700}^* | A_{450}) \right), \end{aligned} \quad (4)$$

where $P(X)$ is the probability of even X and \neg means *not*. The likelihood function of the model with parameter set θ for the pH-stress experiment is

$$\mathcal{L}(\theta | A_{150}, L_{750}) = P^4(L_{450}) \cdot 6 \cdot P^4(L_{750} | A_{150}) \cdot P^4(\neg L_{750} | A_{150}). \quad (5)$$

Because of the complexity of the model, this likelihood function is intractable. Therefore, given specific values of the model parameters θ , we simulate the model in many replicates to approximate the value of the likelihood function $\mathcal{L}(\theta)$.

Parameter inference. To infer model parameters, we use approximate Bayesian computation (Sunnåker et al., 2013) with a sequential Monte-Carlo scheme, or ABC-SMC (Sisson et al., 2007), implemented in the pyABC¹ Python package (Klinger et al., 2018). Briefly, this approach uses numerical stochastic simulations of the model to infer a posterior distribution over the model parameters. It is a method of likelihood-free, simulation-based inference (Cranmer et al., 2020), that is, for estimating a posterior distribution when a likelihood function cannot be computed. It is therefore suitable to our case, in which the likelihood function can only be approximated from simulations, and cannot be directly computed.

Prior distributions. The prior distributions of $w_{2n+1} = 1 - c + b$, $w_{2n+1^*} = (1 + s)(1 - c) + b$ and $w_{2n^*} = 1 + s$ were obtained by estimating $1 - c$, $1 - c + b$ and $1 + s$ from growth curves data. These growth curve data were previously obtained by Yona et al. (2012) using mono-culture growth experiments. The raw growth curves data were not published before, but were used to produce figures 3C, 4A, and S2 in Yona et al. (2012). We used Curveball², a dedicated method for predicting results of competition experiments from growth curve data (Ram et al., 2019). Curveball takes growth curves of two strains growing separately in mono-culture and predicts how they would grow in a mixed culture, that is, it predicts the results of a competition assay. From these predictions, relative fitness values can be computed. We used growth curves of $2n + 1$ and $2n$ in 39 °C and in 30 °C to estimate $1 - c + b$ and $1 - c$, respectively. The latter assumes that the cost of aneuploidy is the same in 39 °C and 30 °C; this might be incorrect, but we only assume this to generate a prior distribution for the fitness values. We also used growth curves of $2n + 1$ and $2n^*$ in 39 °C to estimate $1 + s$. Because Curveball uses a maximum-likelihood approach to estimate model parameters, we were able to estimate a distribution of relative fitness values by sampling from a truncated multivariate normal distribution defined by the maximum-likelihood covariance matrix. Thus, we sampled 10,000 values for $1 - c + b$, $1 - c$, and $1 + s$, which we used as prior distributions. Note that the first and last values correspond to w_{2n+1} and w_{2n} , whereas $w_{2n+1^*} = (1 - c)(w_{2n^*} - 1) + w_{2n+1}$. See Figures S1 and S2.

¹<https://pyabc.readthedocs.io>

²<https://curveball.yoavram.com>

Model comparison.

Results

Discussion

Aneuploidy is not just another type of mutation. The published data indicate that, like mutation, aneuploidy can be both deleterious and beneficial (Pavelka et al., 2010; Sheltzer and Amon, 2011). Nevertheless, there are important and fundamental differences between adaptation by aneuploidy and adaptation by beneficial mutations (Yona et al., 2015), which make aneuploidy a unique mechanism for generating genetic variation. First, the aneuploidy rate (i.e. the frequency of mis-segregation events) is significantly higher than the mutation rate (Santaguida and Amon, 2015). Thus, everything else being equal, adaptation by aneuploidy will be faster and more frequent. Second, fitness effects of aneuploidy are larger than those of the majority of mutations, on average, and are rarely neutral (Pavelka et al., 2010; Yona et al., 2012; Sunshine et al., 2015), allowing selection to quickly sort deleterious and beneficial genotypes. Third, the number of different karyotypes is considerably smaller than the number of different genotypes, and different karyotypes are likely to have different phenotypes (Pavelka et al., 2010). Therefore, exploration of the phenotype space by aneuploidy requires smaller populations and a shorter time span. Fourth, aneuploidy is a reversible state, as the rate of chromosome loss is high and the cost of aneuploidy is significant (Niwa et al., 2006). Indeed, aneuploidy often provides a transient solution: under short-term stress conditions, aneuploidy reverts (chromosome number returns to normal) when the stress subsides; under long-term stress conditions, aneuploidy reverts when refined solutions, generated by beneficial mutations, take over (Yona et al., 2012). Finally, aneuploidy results in increased genome instability, potentially increasing genetic variation by a positive feedback loop (Rancati and Pavelka, 2013; Bouchonville et al., 2009; Zhu et al., 2012), while also increasing its own transience.

Evolutionary theory of aneuploidy. The role of aneuploidy in adaptation has only recently been observed (Sionov et al., 2010; Yona et al., 2012; Gerstein et al., 2015), and is largely missing from the literature on evolution and adaptation: the introductory textbook *Evolution* by Bergstrom and Dugatkin (2012) does not mention the word aneuploidy, and the graduate-level book *Mutation-Driven Evolution* by Nei (2013) only briefly mentions aneuploidy in the context of speciation, but not adaptation. In recent reviews of the literature, aneuploidy is suggested to play an important role in fungal adaptation (Robbins et al., 2017; Todd et al., 2017) and cancer evolution (Santaguida and Amon, 2015; Naylor and van Deursen, 2016; Sansregret and Swanton, 2017), yet these reviews cite no theoretical studies nor any quantitative models. Indeed, evolutionary, ecological, and epidemiological studies mostly assume adaptation occurs via beneficial mutations, recombination, and sex. Therefore, there is a critical need to develop an evolutionary theory of aneuploidy like the evolutionary theories of other mechanisms for generation of genetic variation, e.g. mutation (Lynch, 2010), recombination (Hartfield and Keightley, 2012), and sex (Otto, 2009). An evolutionary theory of aneuploidy will be central to the interpretation of experimental and clinical observations and design of new hypotheses, experiments, and treatments (Carja et al., 2014). For example, despite the lack of theoretical models, aneuploidy has been invoked in a new strategy to combat pathogens and tumour cells by setting “evolutionary traps” (Gerstein et al., 2015; Chen et al., 2015), in which a condition that predictably leads to emergence of aneuploidy is applied, followed by a condition that specifically selects against aneuploid cells.

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References

- Bakhoum, S. F. and Landau, D. A. (2017), ‘Chromosomal instability as a driver of tumor heterogeneity and evolution’, *Cold Spring Harb. Perspect. Med.* **7**(6), 1–14.
- Bergstrom, C. T. and Dugatkin, L. A. (2012), *Evolution*, 1 edn, W. W. Norton & Company, New York, NY.
- Bouchonville, K., Forche, A., Tang, K. E. S., Semple, C. a. M. and Berman, J. (2009), ‘Aneuploid chromosomes are highly unstable during DNA transformation of *Candida albicans*.’, *Eukaryot. Cell* **8**(10), 1554–66.
- Boveri, T. (2008), ‘Concerning the Origin of Malignant Tumours’, *J. Cell Sci.* **121**(Supplement 1), 1–84.
- Carja, O., Liberman, U. and Feldman, M. W. (2014), ‘Evolution in changing environments: Modifiers of mutation, recombination, and migration’, *Proc. Natl. Acad. Sci.* p. 201417664.
- Chen, G., Mulla, W. A., Kucharavy, A., Tsai, H.-J., Rubinstein, B., Conkright, J., McCroskey, S., Bradford, W. D., Weems, L., Haug, J. S., Seidel, C. W., Berman, J. and Li, R. (2015), ‘Targeting the Adaptability of Heterogeneous Aneuploids’, *Cell* **160**(4), 771–784.
- Chen, G., Rubinstein, B. and Li, R. (2012), ‘Whole chromosome aneuploidy: Big mutations drive adaptation by phenotypic leap’, *BioEssays* **34**(10), 893–900.
- Covo, S., Puccia, C. M., Argueso, J. L., Gordenin, D. A. and Resnick, M. A. (2014), ‘The sister chromatid cohesion pathway suppresses multiple chromosome gain and chromosome amplification.’, *Genetics* **196**(2), 373–384.
- Cranmer, K., Brehmer, J. and Louppe, G. (2020), ‘The frontier of simulation-based inference’, *Proc. Natl. Acad. Sci. U. S. A.* p. 201912789.
- Dhar, R., Sägesser, R., Weikert, C., Yuan, J. and Wagner, A. (2011), ‘Adaptation of *Saccharomyces cerevisiae* to saline stress through laboratory evolution.’, *J. Evol. Biol.* **24**(5), 1135–53.
- Dunham, M. J., Badrane, H., Ferea, T., Adams, J., Brown, P. O., Rosenzweig, F. and Botstein, D. (2002), ‘Characteristic genome rearrangements in experimental evolution of *Saccharomyces cerevisiae*’, *Proc. Natl. Acad. Sci.* **99**(25), 16144–16149.
- Gerstein, A. C. and Berman, J. (2018), ‘Diversity of acquired adaptation to fluconazole is influenced by genetic background and ancestral fitness in *Candida albicans*’, *bioRxiv* p. 360347.
- Gerstein, A. C., Ono, J., Lo, D. S., Campbell, M. L., Kuzmin, A. and Otto, S. P. (2015), ‘Too much of a good thing: the unique and repeated paths toward copper adaptation.’, *Genetics* **199**(2), 555–71.
- Gresham, D., Desai, M. M., Tucker, C. M., Jenq, H. T., Pai, D. A., Ward, A., DeSevo, C. G., Botstein, D. and Dunham, M. J. (2008), ‘The repertoire and dynamics of evolutionary adaptations to controlled nutrient-limited environments in yeast’, *PLoS Genet.* **4**(12).
- Hartfield, M. and Keightley, P. D. (2012), ‘Current hypotheses for the evolution of sex and recombination.’, *Integr. Zool.* **7**(2), 192–209.

- Hong, J. and Gresham, D. (2014), ‘Molecular Specificity, Convergence and Constraint Shape Adaptive Evolution in Nutrient-Poor Environments’, *PLoS Genet.* **10**(1).
- Kasuga, T., Bui, M., Bernhardt, E., Swiecki, T., Aram, K., Cano, L. M., Webber, J., Brasier, C., Press, C., Grünwald, N. J., Rizzo, D. M. and Garbelotto, M. (2016), ‘Host-induced aneuploidy and phenotypic diversification in the Sudden Oak Death pathogen *Phytophthora ramorum*’, *BMC Genomics* **17**(1), 1–17.
- Klinger, E., Rickert, D. and Hasenauer, J. (2018), ‘pyABC: distributed, likelihood-free inference’, *Bioinformatics* (May), 1–3.
- Lynch, M. (2010), ‘Evolution of the mutation rate.’, *Trends Genet.* **26**(8), 345–352.
- Mannaert, A., Downing, T., Imamura, H. and Dujardin, J. C. (2012), ‘Adaptive mechanisms in pathogens: Universal aneuploidy in *Leishmania*’, *Trends Parasitol.* **28**(9), 370–376.
- Möller, M., Habig, M., Freitag, M. and Stukenbrock, E. H. (2018), ‘Extraordinary Genome Instability and Widespread Chromosome Rearrangements During Vegetative Growth’, *Genetics* **210**(2), 517–529.
- Musacchio, A. and Salmon, E. D. (2007), ‘The spindle-assembly checkpoint in space and time’, *Nat. Rev. Mol. Cell Biol.* **8**(5), 379–393.
- Naylor, R. M. and van Deursen, J. M. (2016), ‘Aneuploidy in Cancer and Aging’, *Annu. Rev. Genet.* **50**(1), 45–66.
- Nei, M. (2013), *Mutation-Driven Evolution*, 1st edn, Oxford University Press, Oxford.
- Niwa, O., Tange, Y. and Kurabayashi, A. (2006), ‘Growth arrest and chromosome instability in aneuploid yeast’, *Yeast* **23**(13), 937–950.
- Otto, S. P. (2009), ‘The Evolutionary Enigma of Sex’, *Am. Nat.* **174**(July), S1–S14.
- Otto, S. P. and Day, T. (2007), *A biologist’s guide to mathematical modeling in ecology and evolution*, Princeton University Press.
- Pavelka, N., Rancati, G., Zhu, J., Bradford, W. D., Saraf, A., Florens, L., Sanderson, B. W., Hattem, G. L. and Li, R. (2010), ‘Aneuploidy confers quantitative proteome changes and phenotypic variation in budding yeast.’, *Nature* **468**(7321), 321–5.
- Ram, Y., Dellus-Gur, E., Bibi, M., Karkare, K., Obolski, U., Feldman, M. W., Cooper, T. F., Berman, J. and Hadany, L. (2019), ‘Predicting microbial growth in a mixed culture from growth curve data’, *Proc. Natl. Acad. Sci. U. S. A.* **116**(29), 14698–14707.
- Rancati, G. and Pavelka, N. (2013), ‘Karyotypic changes as drivers and catalyzers of cellular evolvability: A perspective from non-pathogenic yeasts’, *Semin. Cell Dev. Biol.* **24**(4), 332–338.
- Rancati, G., Pavelka, N., Fleharty, B., Noll, A., Trimble, R., Walton, K., Perera, A., Staehling-Hampton, K., Seidel, C. W. and Li, R. (2008), ‘Aneuploidy Underlies Rapid Adaptive Evolution of Yeast Cells Deprived of a Conserved Cytokinesis Motor’, *Cell* **135**(5), 879–893.
- Robbins, N., Caplan, T. and Cowen, L. E. (2017), ‘Molecular Evolution of Antifungal Drug Resistance’, *Annu. Rev. Microbiol.* **71**(1), 753–775.
- Rodrigues, M. L. and Albuquerque, P. C. (2018), ‘Searching for a change: The need for increased support for public health and research on fungal diseases’, *PLoS Negl. Trop. Dis.* **12**(6), 1–5.
- Sansregret, L. and Swanton, C. (2017), ‘The Role of Aneuploidy in Cancer Evolution’, *Cold Spring Harb. Perspect. Med.* **7**(1), a028373.

- Santaguida, S. and Amon, A. (2015), ‘Short- and long-term effects of chromosome mis-segregation and aneuploidy’, *Nat. Rev. Mol. Cell Biol.* **16**(8), 473–485.
- Santaguida, S., Vasile, E., White, E. and Amon, A. (2015), ‘Aneuploidy-induced cellular stresses limit autophagic degradation’, *Genes Dev.* **29**(19), 2010–2021.
- Schvartzman, J. M., Sotillo, R. and Benezra, R. (2010), ‘Mitotic chromosomal instability and cancer: Mouse modelling of the human disease’, *Nat. Rev. Cancer* **10**(2), 102–115.
- Selmecki, A. M., Dulmage, K., Cowen, L. E., Anderson, J. B. and Berman, J. (2009), ‘Acquisition of Aneuploidy Provides Increased Fitness during the Evolution of Antifungal Drug Resistance’, *PLoS Genet.* **5**(10), e1000705.
- Selmecki, A. M., Forche, A. and Berman, J. (2010), ‘Genomic Plasticity of the Human Fungal Pathogen *Candida albicans*’, *Eukaryot. Cell* **9**(7), 991–1008.
- Selmecki, A. M., Gerami-Nejad, M., Paulson, C., Forche, A. and Berman, J. (2008), ‘An isochromosome confers drug resistance in vivo by amplification of two genes, ERG11 and TAC1’, *Mol. Microbiol.* **68**(3), 624–641.
- Sheltzer, J. M. and Amon, A. (2011), ‘The aneuploidy paradox: Costs and benefits of an incorrect karyotype’, *Trends Genet.* **27**(11), 446–453.
- Sheltzer, J. M., Ko, J. H., Replogle, J. M., Habibe Burgos, N. C., Chung, E. S., Meehl, C. M., Sayles, N. M., Passerini, V., Storchova, Z. and Amon, A. (2017), ‘Single-chromosome Gains Commonly Function as Tumor Suppressors’, *Cancer Cell* **31**(2), 240–255.
- Shor, E. and Perlin, D. S. (2015), ‘Coping with Stress and the Emergence of Multidrug Resistance in Fungi’, *PLOS Pathog.* **11**(3), e1004668.
- Sionov, E., Lee, H., Chang, Y. C. and Kwon-Chung, K. J. (2010), ‘*Cryptococcus neoformans* Overcomes Stress of Azole Drugs by Formation of Disomy in Specific Multiple Chromosomes’, *PLoS Pathog.* **6**(4), e1000848.
- Sisson, S. A., Fan, Y. and Tanaka, M. M. (2007), ‘Sequential Monte Carlo without likelihoods’, *Proc. Natl. Acad. Sci.* **104**(6), 1760–1765.
- Sunnåker, M., Busetto, A. G., Numminen, E., Corander, J., Foll, M. and Dessimoz, C. (2013), ‘Approximate Bayesian Computation’, *PLoS Comput. Biol.* **9**(1), e1002803.
- Sunshine, A. B., Payen, C., Ong, G. T., Liachko, I., Tan, K. M. and Dunham, M. J. (2015), ‘The Fitness Consequences of Aneuploidy Are Driven by Condition-Dependent Gene Effects’, *PLOS Biol.* **13**(5), e1002155.
- Todd, R. T., Forche, A. and Selmecki, A. M. (2017), ‘Ploidy Variation in Fungi: Polyploidy, Aneuploidy, and Genome Evolution’, *Microbiol. Spectr.* **5**(4), 1–20.
- Torres, E. M., Dephoure, N., Panneerselvam, A., Tucker, C. M., Whittaker, C. A., Gygi, S. P., Dunham, M. J. and Amon, A. (2010), ‘Identification of aneuploidy-tolerating mutations’, *Cell* **143**(1), 71–83.
- Torres, E. M., Sokolsky, T., Tucker, C. M., Chan, L. Y., Boselli, M., Dunham, M. J. and Amon, A. (2007), ‘Effects of Aneuploidy on Cellular Physiology and Cell Division in Haploid Yeast’, *Science (80-.).* **317**(5840), 916–924.
- Tsai, H. J., Nelliat, A. R., Choudhury, M. I., Kucharavy, A., Bradford, W. D., Cook, M. E., Kim, J., Mair, D. B., Sun, S. X., Schatz, M. C. and Li, R. (2019), ‘Hypo-osmotic-like stress underlies general cellular defects of aneuploidy’, *Nature* .

Williams, B. R., Prabhu, V. R., Hunter, K. E., Glazier, C. M., Whittaker, C. a., Housman, D. E. and Amon, A. (2008), ‘Aneuploidy Affects Proliferation and Spontaneous Immortalization in Mammalian Cells’, *Science* (80-.). **322**(5902), 703–709.

Yona, A., Frumkin, I. and Pilpel, Y. (2015), ‘A Relay Race on the Evolutionary Adaptation Spectrum’, *Cell* **163**(3), 549–559.

Yona, A., Manor, Y. S., Herbst, R. H., Romano, G. H., Mitchell, A., Kupiec, M., Pilpel, Y. and Dahan, O. (2012), ‘Chromosomal duplication is a transient evolutionary solution to stress.’, *Proc. Natl. Acad. Sci.* **109**(51), 21010–5.

Zhu, J., Pavelka, N., Bradford, W. D., Rancati, G. and Li, R. (2012), ‘Karyotypic determinants of chromosome instability in aneuploid budding yeast’, *PLoS Genet.* **8**(5).

Zhu, J., Tsai, H.-J., Gordon, M. R. and Li, R. (2018), ‘Cellular Stress Associated with Aneuploidy’, *Dev. Cell* **44**(4), 420–431.

Zhu, Y. O., Sherlock, G. and Petrov, D. A. (2016), ‘Whole Genome Analysis of 132 Clinical *Saccharomyces cerevisiae* Strains Reveals Extensive Ploidy Variation’, *G3 Genes, Genomes, Genet.* **6**(8), 2421–2434.

Supplementary Material

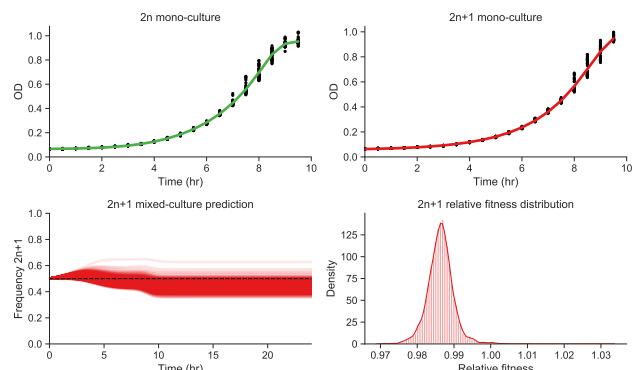


Figure S1: Fitness estimation from 30 °C.

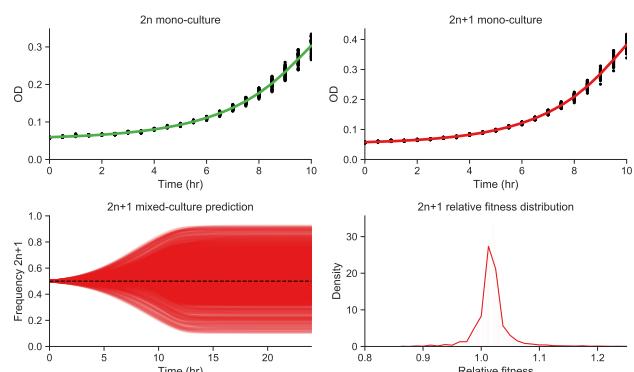


Figure S2: Fitness estimation from 39 °C.