

Aneuploidy can be an evolutionary detour on the path to adaptation

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Abstract

16 Aneuploidy is common in eukaryotes, often leading to decreased growth and fitness. How-
18 ever, evidence from yeast and other fungi, as well as human tumour cells, suggests that specific
20 aneuploidies can be beneficial under stressful conditions and facilitate adaptation. In a prominent
22 example, an evolutionary experiment with yeast, populations evolving under heat stress had be-
come aneuploid (chromosome III), only to later revert back to euploid after genetic mutations have
accumulated. It has therefore been suggested that aneuploidy serves as a "stepping stone" on the
path to adaptation.

24 Here, we test this hypothesis. First, we apply DNA sequencing to show that mutant alleles
common in aneuploid cells are uncommon in the evolved euploid population. Second, we develop
26 an evolutionary model with both aneuploidy and mutation, and fit it to the results of the experiment
using a Bayesian inference framework. We then predict the genotype frequency dynamics during
the experiment, demonstrating that the majority of the evolved euploid population likely did not
28 descend from aneuploid cells, but rather directly from the euploid wild-type population. Our
model shows how the beneficial mutation supply—the product of population size and beneficial
30 mutation rate—determine the evolutionary dynamics: with a low mutation supply, a large fraction
of the evolved population may descend from aneuploid cells; but with a high mutation supply,
32 beneficial mutations are generated before fixation of aneuploidy, and can outcompete aneuploidy
due to the latter's inherent fitness cost.

34 Together, our results suggest that aneuploidy can be an evolutionary "detour" rather than a
"stepping stone": it can delay, rather than facilitate, the adaptation of the population, and cells that
36 become aneuploid may leave less descendants compared to cells that remain diploid.

Introduction

38 Aneuploidy is an imbalance in the number of chromosomes in the cell: an incorrect karyotype.
Evidence suggests aneuploidy is very common in eukaryotes, e.g. animals^{42,33,2}, and fungi^{36,65,40,54}.
40 Aneuploidy has been implicated in cancer formation, progression, and drug resistance^{4,44,42,21}. It
is also common in protozoan pathogens of the *Leishmania* genus, a major global health concern³¹,
42 and contributes to the emergence of drug resistance⁴⁵ and virulence³² in fungal pathogens, which
are under-studied⁴¹, despite infecting a billion people per year, causing significant morbidity in >150
44 million and death in >1.5 million people per year^{45,41}.

Experiments with human and mouse embryos found that most germline aneuploidies are lethal. An-
46 euploidies are also associated with developmental defects and lethality in other multicellular organ-
isms⁴⁸. For example, aneuploid mouse embryonic cells grow slower than euploid cells⁵⁹. Similarly,
48 in unicellular eukaryotes growing in benign conditions, aneuploidy usually leads to slower growth and
decreased overall fitness, in part due to proteotoxic stress due to increased expression, gene dosage
50 imbalance, and hypo-osmotic-like stress^{34,57,36,48,43,24,64,58,60}.

However, aneuploidy can be beneficial under stressful conditions due to the wide range of phenotypes
52 it can produce, some of which are advantageous^{36,60}. Indeed, in a survey of 1,011 yeast strains,
aneuploidy has been detected in about 19%³⁷. Thus, aneuploidy can lead to rapid adaptation in
54 unicellular eukaryotes^{15,56,19,39}, as well as to rapid growth of somatic tumour cells^{44,50}. For example,
aneuploidy in *Saccharomyces cerevisiae* facilitates adaptation to a variety of stressful conditions
56 like heat and pH⁶², copper^{7,15}, salt¹⁰, and nutrient limitation^{11,17,1}, with similar results in *Candida*
*albicans*⁶⁰. Importantly, aneuploidy can also lead to drug resistance in pathogenic fungi such as
58 *C. albicans*^{47,46,14} and *Cryptococcus neoformans*⁵¹, which cause candidiasis and meningoencephalitis,
respectively.

60 Yona et al.⁶² demonstrated experimentally the importance of aneuploidy in adaptive evolution. They
evolved populations of *S. cerevisiae* under strong heat stress. The populations adapted to the heat stress
62 within 450 generations, and this adaptation was determined to be due a duplication of chromosome III.
Later on, after more than 1,500 generations, the populations reverted back to an euploid state, while
64 remaining adapted to the heat stress. Aneuploidy was therefore suggested to be a *transient adaptive*
solution, because it can rapidly appear and fixate in the population under stressful conditions, and can
66 then be rapidly lost when the cost of aneuploidy outweighs its benefit—after the stress is removed,
or after "refined" beneficial mutations appear and fixate⁶². Furthermore, it has been suggested that
68 aneuploidy is an evolutionary "stepping stone" that facilitates future adaptation by genetic mutations,

which require more time to evolve^{62,61}.

70 Here, we test the hypothesis that aneuploidy is a *an evolutionary stepping stone* that facilitates adaptive evolution by genetic mutations. First, we sequenced the genomes of evolved populations reported in⁶²
72 and analyzed their mutant allele frequencies to assess if the evolved euploid cells are descended from
aneuploid cells. Second, we develop an evolutionary genetic model and fit it to the experimental
74 results of Yona et al.⁶² in order to predict the genotype frequency dynamics in the experimental
populations, thereby estimating the frequency of evolved euploid cells that descended from aneuploid
76 cells. Our results show that aneuploidy reached high frequencies in the experimental populations,
but nevertheless, the majority of cells in the evolved euploid population likely did not descend from
78 aneuploid cells, but rather directly from wild-type euploid cells. These results suggest that at the
lineage level, aneuploidy may be an evolutionary detour, rather than a stepping stone, on the path to
80 adaptation.

Results

82 In the heat-stress experiment of Yona et al.⁶², four populations of *S. cerevisiae* evolved under 39 °C.
Aneuploidy fixed in all four experimental repetitions in the first 450 generations. Two of the repetitions,
84 marked *H2* and *H4*, carried no large-scale duplications other than a chromosome III trisomy. These
two repetitions continued to evolve under the same conditions, wherein aneuploidy was eliminated by
86 generation 1,700 and 2,350 in *H4* and *H2*, respectively.

Empirical frequencies of mutant alleles. For each of two evolved populations (*H2* and *H4*) we
88 sequenced the ancestral diploid population (generation 0), the aneuploid population (generation 450),
and the evolved euploid population (generation 1,700 or 2,350) to estimate the mutant allele frequencies
90 (Tables S1 and S2). Overall, between 100 and 173 mutant alleles were detected with at least a single
read in the six populations that were sampled. Disregarding 45 and 40 alleles that were present in
92 the ancestral populations at a frequency >10%, the aneuploid and euploid populations carried a large
number of mutant alleles: 82 and 95, respectively, in repetition *H2*, and 60 and 66 in repetition
94 *H4*.

Surprisingly, out of all these mutant alleles, none was present at a frequency >20% in both the
96 aneuploid and the evolved euploid populations. More importantly, a high mutant allele frequency
in the aneuploid population was associated with a low frequency in the evolved euploid population,
98 and vice-versa (Spearman's correlation coefficient $\rho = -0.64$ and -0.66 in the two experimental

repetitions; Figure 1), suggesting that mutant alleles frequent in the aneuploid populations decreased 100 in frequency when aneuploidy was lost. These results suggest evolved euploid cells are unlikely to have descended from aneuploid cells.

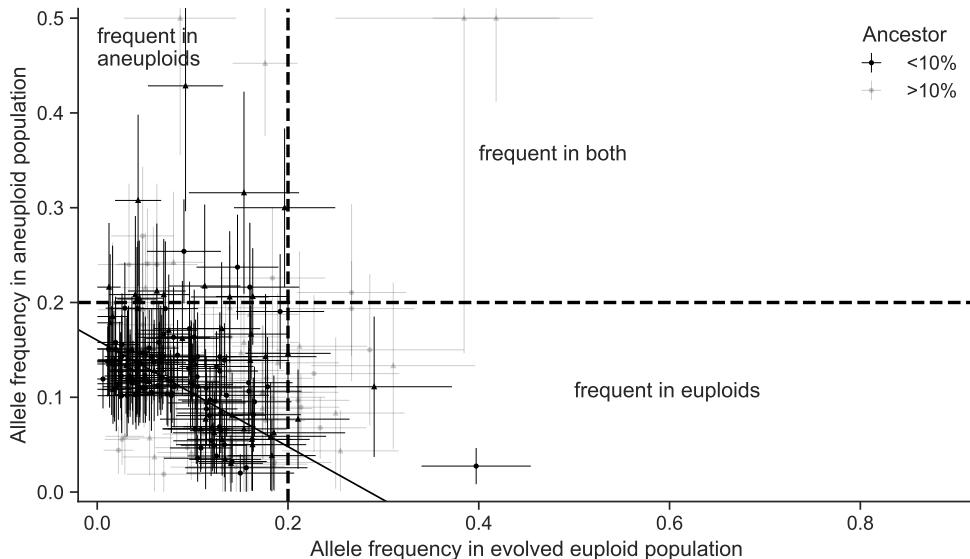


Figure 1: Frequencies of mutant alleles in the experimental populations are negatively correlated. Frequencies of mutant alleles when trisomy was widespread in the population (y-axis) and after it was eliminated (x-axis) in two experimental repetitions (circles for *H2* and triangles for *H4*) from Yona et al.⁶². Mutant alleles with >20% in the aneuploid population were <20% in the euploid population, and vice versa (the upper-right quadrant is empty), suggesting that the majority of evolved euploid cells did not descend from the most common aneuploid genotypes. Alleles with frequency below and above 10% in the ancestral populations are in black and gray, respectively. Solid black line is a linear orthogonal distance regression line (slope=−0.559, intercept=0.164; a regression through alleles that reach at least 20% in one of the populations has slope=−0.645 and intercept=0.297). Dashed vertical and horizontal lines show allele frequencies of 20%. Error bars show SEM (standard error of the mean) assuming the number of reads in Binomially distributed; the SEM may be large when the total number of reads is small. For the 18 mutant alleles with high frequency in the aneuploid populations (>20%), the highest frequencies in the euploid populations were 15.4%, 16%, 16.3% and 19.6% (the rest were below 15%). Similarly, for the 48 mutant alleles with high frequency in the evolved euploid populations, the highest frequencies in the aneuploid populations were 2.7%, 7.7%, and 11.1% (the rest were below 1%).

102 **Evolutionary genetic model.** To explore the dynamics during the evolutionary experiments, we developed an evolutionary genetic model, fitted the model to empirical data, and used it to predict the 104 genotype frequency dynamics, or specifically, the fraction of the evolved euploid population descended

from aneuploid cells.

106 The model includes the effects of natural selection, genetic drift, aneuploidy, and mutation, and follows
107 a population of cells characterized by their genotype: euploid wild-type, $2n$, is the ancestral diploid
108 genotype; euploid mutant, $2n^*$, has a diploid karyotype and a single beneficial mutation; aneuploid
109 wild-type, $2n+1$, has an extra chromosome due to a chromosome duplication event; and aneuploid
110 mutant, $2n+1^*$, has an extra chromosome (like $2n+1$) and a beneficial mutation (like $2n^*$). Fitness
111 values of the different genotypes are denoted by w_{2n} , w_{2n^*} , w_{2n+1} , and w_{2n+1^*} , and the rate of mutation
112 and aneuploidy are denoted by μ and δ , respectively. See Figure 2 for an illustration of the model.

We fitted this model to the experimental results⁶² – time for fixation (>95%) and for loss (<5%) of
114 aneuploidy – using approximate Bayesian computation with sequential Monte Carlo (ABC-SMC)⁵²,
thereby inferring the model parameters: rates aneuploidy and mutation and the fitness of all genotypes.
116 We then sampled posterior predictions for the genotype frequency dynamics using the estimated
parameter values and compared different versions of the model to test additional hypotheses about the
118 evolutionary process.

Estimated rates and fitness effects of aneuploidy and mutation. We inferred the posterior distribution of model parameters (Figure 3). We report parameter estimates using the MAP (maximum a posteriori) and providing the 50% HDI (highest density interval) in square brackets. See Supplementary Material for sensitivity analysis.

The estimated beneficial mutation rate is $\mu = 2.965 \cdot 10^{-6}$ [$2.718 \cdot 10^{-7} - 3.589 \cdot 10^{-6}$]. From the
124 literature, the mutation rate per base pair is roughly $2 - 3 \cdot 10^{-10}$ (refs.^{66,30}), but it may be higher under
heat stress, as several stresses¹⁸, including heat²⁰, may cause hypermutation in yeast. If we assume a
126 10-fold increase over the mutation rate reported in the literature, then the estimated beneficial mutation
rate can be explained by a genomic target size of 1,000 base pairs (i.e., 1,000 base pairs across the
128 genome in which a mutation would provide a fitness benefit). Supporting this, Flynn et al.¹² used a
deep mutational scan of a single protein, Hsp90, to find 465 amino-acid variants that increased growth
130 rate in 37 °C. Furthermore, Yona et al.⁶² found at least 10 genes on chromosome III that increased
heat tolerance when over-expressed. Assuming that other chromosomes also have a similar number of
132 heat-tolerance genes (and even more, as chromosome III is one of the smallest chromosomes¹⁶), we
get a total of 160 heat-tolerance genes in the genome. Indeed, mutations were found in 97 genes in an
134 evolutionary experiment with yeast under heat stress²⁰. Thus, to get a genomic target size of 1,000, it is
enough that the average gene target size is 6.25 base pairs. For example, Kohn and Anderson²⁷ found

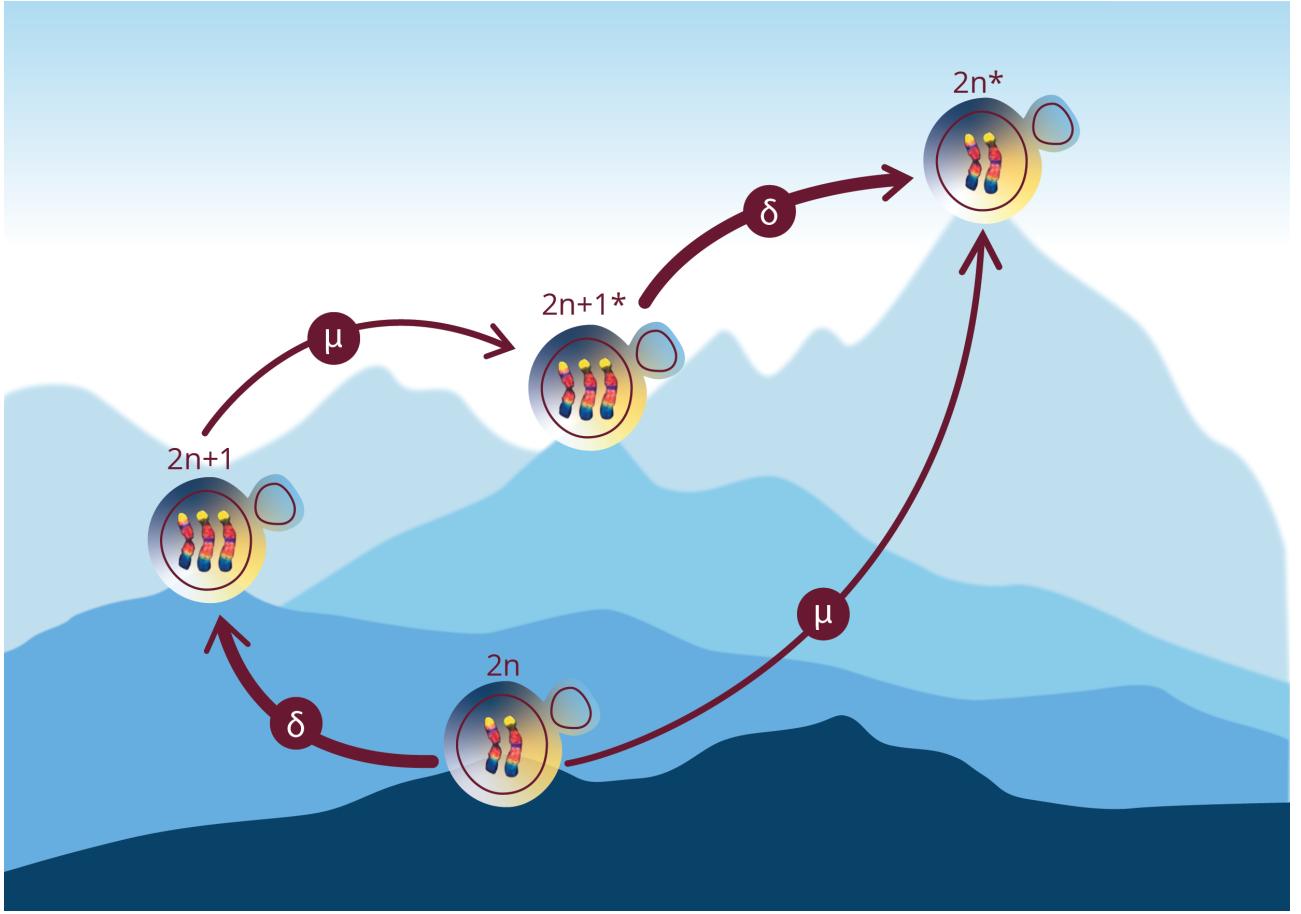


Figure 2: Model Illustration. There are four genotypes in our model: euploid wild-type, $2n$; euploid mutant, $2n^*$; aneuploid wild-type, $2n+1$; and aneuploid mutant, $2n+1^*$. Overall there are two possible trajectories from $2n$ to $2n^*$. Arrows denote transitions between genotypes, with transition rates μ for the beneficial mutation rate and δ for the aneuploidy rate.

136 a target size of 11 in a proton exporter gene (*PMA1*) that contributes to high-salt adaptation.

The estimated aneuploidy rate, $\delta = 1.72 \cdot 10^{-3}$ [$1.47 \cdot 10^{-3} - 2.786 \cdot 10^{-3}$] is higher than in previous
 138 studies: for chromosome III in diploid *S. cerevisiae*, Zhu et al.⁶⁶ estimated $6.7 \cdot 10^{-6}$ chromosome
 140 gain events per generation, and Kumaran et al.²⁹ estimate $3.0 - 4.3 \cdot 10^{-5}$ chromosome loss events
 142 per generation (95% confidence interval). However, this difference may be partly explained by an
 increased aneuploidy rate during heat stress: heat shock can increase the rate of chromosome fragment
 loss by 2-3 orders of magnitude⁵.

The estimated fitness values are $w_{2n+1} = 1.022$ [$1.021 - 1.023$], $w_{2n+1^*} = 1.025$ [$1.024 - 1.026$],
 144 $w_{2n^*} = 1.028$ [$1.026 - 1.029$], all relative to the fitness of $2n$, which is set to $w_{2n} = 1$. Thus, we
 146 can infer that the cost of chromosome III trisomy is $c = w_{2n^*} - w_{2n+1^*} = 0.003$ (or 0.3%) and the
 benefit of trisomy is $w_{2n+1} - 1 - c = 0.019$ (1.9%), whereas the benefit of the beneficial mutation is
 $w_{2n^*} - 1 = 0.028$ (2.8%).

148 If we allow for transitions (mutation, chromosome loss and gain) to less-fit genotypes (e.g., $2n^*$ to $2n+1^*$), then we infer similar but slightly different values, see Supplementary Material.

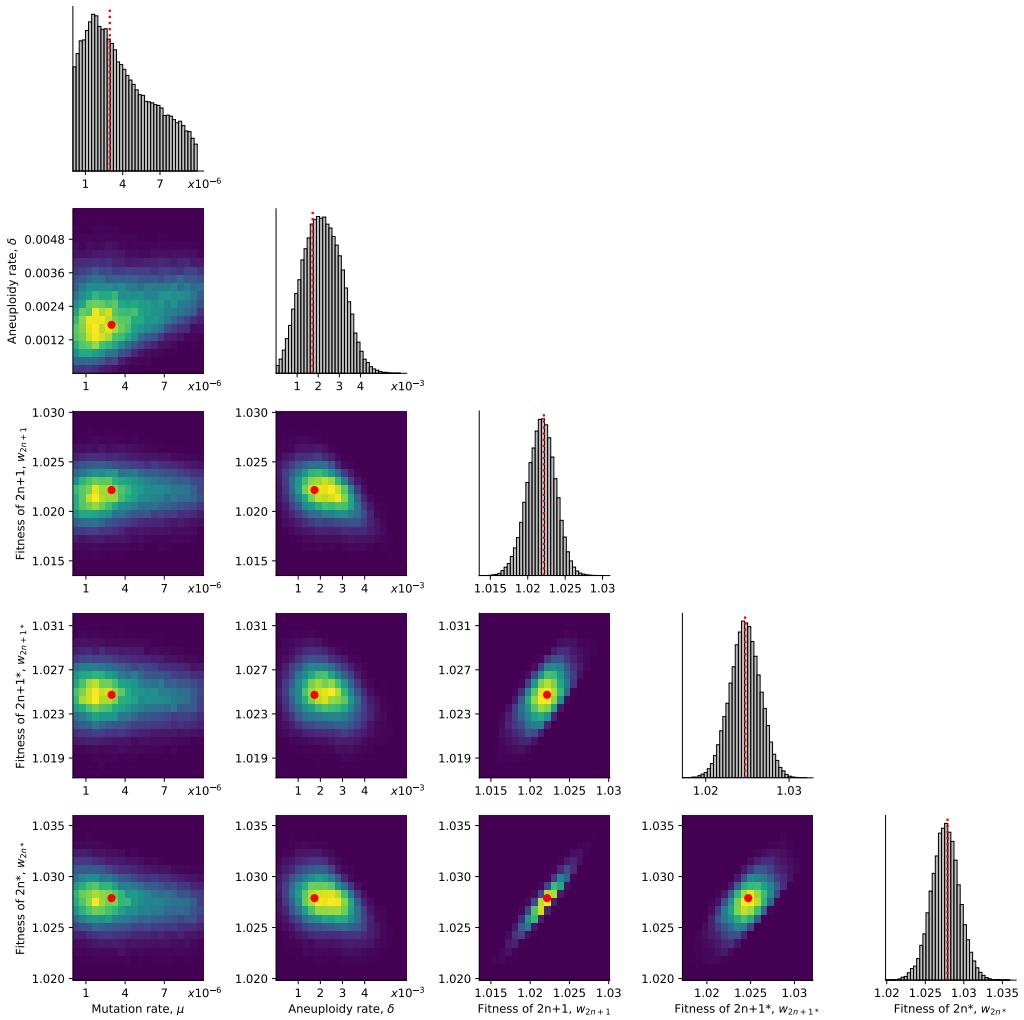


Figure 3: Posterior distribution of model parameters. On the diagonal, the marginal posterior distribution of each model parameter. Below the diagonal, the joint posterior distribution of pairs of model parameters (dark purple and bright yellow for low and high density, respectively). Red markers and orange lines for the joint MAP estimate (which may differ from the marginal MAP, as the marginal distribution integrates over all other parameters).

150 **Model comparison and goodness-of-fit.** To assess the fit of our model to the data, we use posterior predictive checks, in which we simulate the frequency dynamics using MAP parameter estimates and
152 compare them to the data. Our model fits the data well: $2n^*$ fixed in 61% of simulations by generation 1,700 and in 100% of simulations by generation 2,350 (Figure 4).

154 However, a model without aneuploidy (where the aneuploidy rate is fixed at zero, $\delta = 0$), fails to
156 explain the experimental observations (Figure 4). The estimated mutation rate without aneuploidy is $\mu = 7.98 \cdot 10^{-9}$ [$7.906 \cdot 10^{-9} - 8.138 \cdot 10^{-9}$], much lower compared to a model with aneuploidy. The

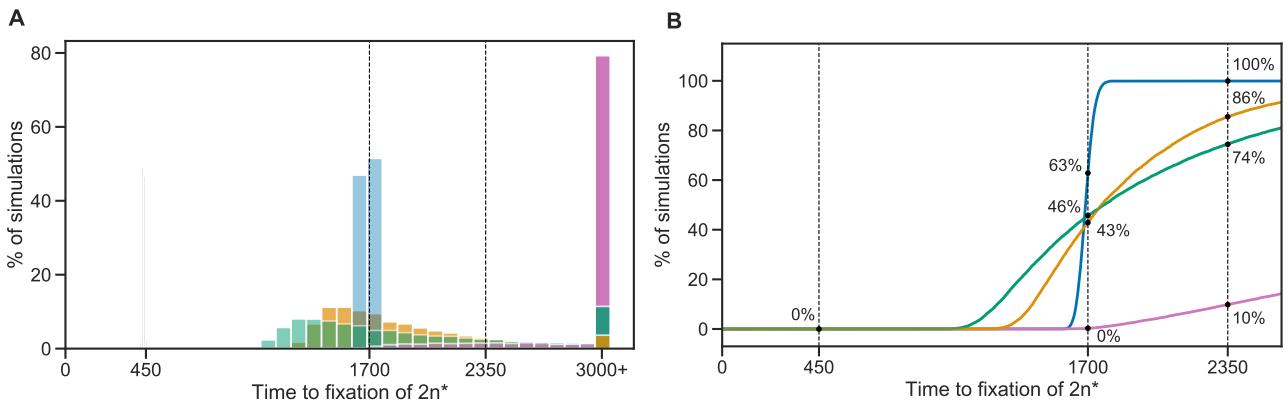


Figure 4: Model fit with and without aneuploidy. The distribution of time to fixation of $2n^*$ (i.e., adaptation time) in 10,000 simulations using MAP parameters of the model with beneficial aneuploidy (blue; $\delta > 0$, $w_{2n} < w_{2n+1} < w_{2n+1}^* < w_{2n}^*$) compared to alternative models: a model with the same parameter values but without aneuploidy (gray, $\delta = 0$, concentrated at $t = 450$); a model fitted to the data assuming no aneuploidy (green, $\delta = 0$); a model fitted to the data assuming neutral aneuploidy (yellow, $\delta > 0$, $w_{2n+1} = w_{2n}$, $w_{2n+1}^* = w_{2n}^*$); and a model with beneficial aneuploidy and an extended prior distribution (pink). In the experiment by Yona et al.⁶², one population lost aneuploidy by generation 1,700 and another by generation 2,350 (dashed lines) but not before generation 450. Thus, the blue distribution has a better fit compared to the other distributions (the gray distribution has a particularly poor fit). The MAP likelihood (eq. (4)) is 0.84, 0.78, 0.67, and 0.14 for the models represented by blue, yellow, green, and pink distributions, respectively. **(A)** Histogram of the time to fixation of $2n^*$. The last bin contains all values equal or greater than 3,000. **(B)** Cumulative distribution of the time to fixation.

fitness of the mutant is also much lower at $w_{2n}^* = 1.013$ [1.012 – 1.013]. This is because, without
158 aneuploidy, a high mutation rate or fitness effect will lead to faster appearance and fixation of $2n^*$ than
in the experimental observations.

160 We also checked a model in which aneuploidy occurs but is adaptively neutral compared to the wild-type, that is, $w_{2n+1} = w_{2n}$ and $w_{2n+1}^* = w_{2n}^*$ but $\delta > 0$. This model fits the data better than the model
162 with no aneuploidy (in which $\delta = 0$), but worse than a model with positive selection for aneuploidy,
in which $w_{2n} < w_{2n+1} < w_{2n+1}^* < w_{2n}^*$ (Figure 4).

164 **Model predictions of genotype frequency dynamics.** We simulated 50 replicate genotype frequency dynamics using the MAP estimate parameters. Figure 5A shows the simulated frequencies of
166 the four genotypes ($2n$, $2n+1$, $2n+1^*$ and $2n^*$), as well as the frequencies of $2n^*$ cells that arose from either $2n+1$ cells via a sequences of mutation and chromosome loss events ($2n_A^*$), or directly from

168 $2n$ cells via a mutation event ($2n_M^*$). We find that $2n+1^*$ never reaches substantial frequency as it is
quickly replaced by $2n^*$ in a process similar to *stochastic tunneling*^{22,28}.

170 To test the hypothesis that aneuploidy facilitates adaptation, we estimated F_A , the expected frequency
of $2n^*$ that arose from $2n+1$, computed as the average frequency of such $2n_A^*$ cells at the end of
172 simulations using the MAP estimate parameters. Surprisingly, we observe that the majority of $2n^*$
cells are $2n_M^*$, a product of a direct mutation in $2n$ cells, rather than descending from $2n+1$ cells
174 ($F_A^{MAP} = 0.106$, average end point of 50 purple lines in Figure 5A). This is despite the fact that the
 $2n+1$ genotype reaches high frequencies in the population (at least 0.98, Figure 5A).

176 This result is not unique to the MAP parameter estimate. We simulated genotype frequency dynamics
using parameter samples from the posterior distribution, and computed the posterior distribution of F_A
178 (Figure 5B). The posterior mode F_A was just 0.147 [0.0154-0.370 95% CI] and only in 489 of 100,000
posterior samples (0.489%) F_A was larger than 0.5 (see Supporting Material for results when transitions
180 to less-fit genotypes are allowed, such as $2n^*$ to $2n+1^*$). Thus, if we sample a random cell from the
evolved $2n^*$ population, it is more likely to have descended directly from an euploid cell than from
182 an aneuploid cell. The probability of $2n^*$ descending from $2n+1$ (F_A) increases with the aneuploidy
rate, δ , and decreases with both the population size N and the mutation rate, μ (Figure 5C,D). In some
184 cases it can also be affected by the fitness parameters (Figure S10).

Genetic instability in aneuploid cells. It has been suggested that aneuploidy increases genetic
186 instability^{49,21}. Therefore, we inferred model parameters under the assumption that the mutation rate
increases in aneuploid cells by a factor $\tau = 1, 33/32$ (due to an additional chromosome), 2, 5, 10, or
188 100 (due to genetic instability). We found that the posterior distribution was similar for $\tau = 1, 33/32$,
2, and 5 (Figure S4). For each τ , we computed the WAIC, a criterion for model selection (lower is
190 better, see Methods), and found WAIC is lowest for $\tau = 33/32$ and $\tau = 1$ (Table S3).

Assuming a strong increase of the mutation rate in aneuploid cells, i.e. $\tau = 100$, the inferred a
192 mutation rate was $\mu = 4.094 \cdot 10^{-7}$ [$6.252 \cdot 10^{-8} - 6.046 \cdot 10^{-7}$], and the inferred aneuploidy rate
that was $\delta = 0.744 \cdot 10^{-3}$ [$0.506 \cdot 10^{-3} - 1.827 \cdot 10^{-3}$]. Compared to inference made assuming
194 no effect of aneuploidy on the mutation rate, these rates were about 7-8-fold lower about 2-3-fold
lower. Assuming $\tau = 10$, the inferred a mutation rate was only slightly lower compared to $\tau = 1$
196 ($\mu = 1.67 \cdot 10^{-6}$ [$2.836 \cdot 10^{-8} - 2.245 \cdot 10^{-6}$]).

Therefore, we do not find any evidence of an increase in mutation rate in aneuploid cells. This may
198 be because, unless the increase is strong ($\tau \geq 10$), it does not seem to affect our inference; or because

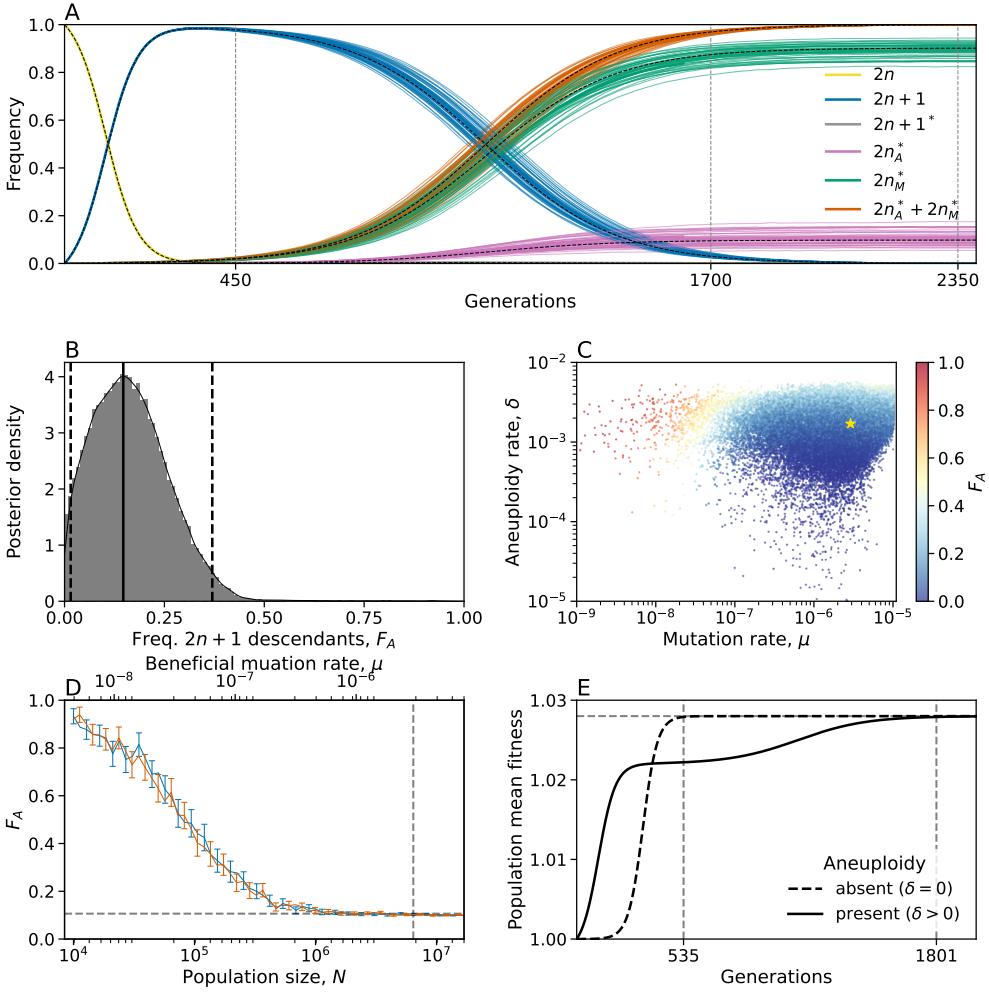


Figure 5: Predicted frequency of aneuploid-descended cells. (A) Posterior predicted genotype frequencies over time, including the source of $2n^*$: $2n_A^*$ arose from $2n+1$, whereas $2n_M^*$ arose directly from $2n$. Colored curves are 50 simulations using the MAP estimate parameters. Black dashed curves are the expected genotype frequencies without genetic drift (from a deterministic model). See Figure S9 for log-log scale, in which the sequence of events is easier to observe. (B) Posterior distribution of F_A , the expected frequency of $2n^*$ cells descended from $2n+1$ cells, computed as the average frequency at the end of 100 simulations for 100,000 samples from the parameter posterior distribution. Solid and dashed lines show the mode and 95% CI. (C) F_A values (color coded) from panel B, with their corresponding mutation rate μ on x-axis and aneuploidy rate δ on the y-axis. Yellow star shows the MAP estimate. See also Figure S10. (D) F_A as a function of the population size (N , bottom x-axis) and the beneficial mutation rate (μ , top x-axis) in posterior predictions with MAP parameters. Markers show F_A in 250 simulations per population size or mutation rate value. Error bars show mean F_A with 95% CI (bootstrap, $n = 10,000$). Blue and red bars for varying population size and mutation rate, respectively. Vertical dashed line for population size in the experiment, $6.425 \cdot 10^6$, and the MAP mutation rate, $2.965 \cdot 10^{-6}$. Horizontal line for $F_A^{MAP} = 0.106$. (E) Population mean fitness in a model without drift using MAP estimate parameters. Solid lines for mean fitness with aneuploidy ($\delta > 0$), where the population reaches adaptation (mean fitness at 99.99% of maximum value) at generation 1,802. Dashed lines for mean fitness without aneuploidy ($\delta = 0$), where the population adapts much earlier, at generation 535.

chromosome III is one of the smallest chromosomes¹⁶. We also checked the differences in genotype
200 frequency dynamics for different τ values. We observe $\tau = 100$ could be distinguished if accurate
data was available for the waiting time until the frequency of $2n$ to decrease below 95% (Figure S5A)
202 or for waiting time for the frequency of $2n+1$ to either reach or go below 95% (Figure S5B). We also
did not find evidence for an increase in the aneuploidy rate in aneuploid cells (data not shown).

204 Discussion

In a landmark study on the role of chromosome duplication in adaptive evolution, Yona et al.⁶²
206 found that a chromosome III trisomy was acquired by *S. cerevisiae* populations evolving under heat
stress, only to be later replaced by euploid mutant cells that carry "refined" solutions to the stress.
208 Additionally, such a replacement also occurred when they initiated evolutionary experiments with a
population in which all cells carry a chromosome III trisomy. They hypothesized that aneuploidy is
210 a "useful yet short-lived intermediate that facilitates further adaptation", suggesting that the euploid
mutant cells evolved by heat-resistance mutations in aneuploid cells followed by reversion of trisomy
212 due to a chromosome loss event.

If indeed the evolved euploid population is descended from the aneuploid population, then mutant
214 alleles that were common in the aneuploid populations should also be common in the evolved euploid
population. However, we found that this is not the case (Figure 1): mutant allele frequencies in the
216 aneuploid and euploid populations are negatively correlated, such that common alleles in the former
are rare in the latter. Furthermore, we developed an evolutionary genetic model of adaptive evolution
218 by aneuploidy and mutation (Figure 2), fitted it to the experimental results of Yona et al.⁶², and
used it to predict the genotype frequency dynamics. The model predicted that only about 10-15% of
220 the evolved euploid population descended from aneuploid cells—that is, the majority of the euploid
population are not descended from aneuploid cells, but rather are direct descendants of the ancestral
222 wild-type population (Figure 5).

This happens despite aneuploidy reaching a high frequency in the population (>95%). Conventional
224 wisdom might suggest that once the aneuploid genotype $2n+1$ reaches high frequency, it will have a
better chance at producing "refined" solutions via mutations, and its descendants will come to dominate
226 the population: the frequency of $2n_A^*$ (which arises from $2n+1^*$) will be higher than the frequency of
 $2n_M^*$ (which arises directly from $2n$).

228 So how does $2n_M^*$ prevail? Initially, the supply rates of $2n+1$ and $2n_M^*$ are $N\delta \approx 11,000$ and $N\mu \approx 19$,

respectively (assuming MAP parameter estimates). Therefore, both genotypes are expected to appear
230 immediately at the beginning of the experiment (Figure S9). However, $2n+1$ appears at a much higher frequency as $\delta \gg \mu$ by 2-3 orders of magnitude. After they first appear, $2n_M^*$ has higher fitness. But
232 as long as the frequency of $2n$ is high, the supply rate of $2n+1$ is higher than that of $2n_M^*$, again due to $\delta \gg \mu$. However, supply rates of both genotypes decreases with the frequency of $2n$. Therefore, when
234 the latter decreases, mainly due to the increase in the frequency of $2n+1$, both supply rates diminish. At this stage, the higher fitness of $2n_M^*$ comes into play and it starts to take over the population,
236 which is mainly composed of $2n+1$. For the aneuploid lineage to compete with the mutant lineage, it must produce $2n_A^*$ via a mutation followed by chromosome loss. Although this is a stochastic process
238 (due to drift), our results show that the time until $2n_A^*$ reaches a frequency of 0.1% is roughly 450 generations, without much variation (intersection of purple lines and vertical dashed line in Figure S9).
240 However, by that time $2n_M^*$ is already at a roughly 10-fold higher frequency (1.86%), and since both mutants have the same fitness, their relative frequency remains roughly the same until the end of the
242 experiment.

Predictions for small populations and low mutation rates. We examined the effect of the population size, N , and the beneficial mutation rate, μ , on the frequency of $2n+1$ descendants in the evolved population, F_A . We found that F_A is expected to decrease as the population size or mutation rate
244 increase (Figure 5D), ranging from >90% when the population size is 10,000 or the mutation rate is $6 \cdot 10^{-9}$, to about 10% when the population size is above 1,000,000 (less than the experimental population size, which was 6,425,000) or the mutation rate is above $2 \cdot 10^{-6}$ (less than the inferred mutation
246 rate, which is $2.965 \cdot 10^{-6}$). Thus, our model provides a testable prediction: if the experiment was
248 repeated under a lower population size (via stronger daily dilutions or in a smaller volume) or a lower mutation rate (via a non-mutagenic stress or stress with a smaller target size such as drug resistance),
250 then the fraction of the population descending from aneuploid cells would be much higher.
252

Aneuploidy delays rather than facilitates adaptation. An additional interesting result of our
254 study is that aneuploidy increases, rather than decreases, the adaptation time (Figure 5E). This happens despite the fact that the mean fitness initially increases faster in the presence of aneuploidy
256 (Figure 5E). This is because once $2n+1$ is common, selection for the mutant strain ($2n+1^*$ or $2n^*$) is weaker compared to when $2n^*$ competes directly with $2n$.

258 **Rate and fitness effect of aneuploidy and mutation.** We inferred the rates of aneuploidy and
mutation and their effects on fitness. We estimate that the aneuploidy rate (i.e., number of chromosome
260 gains per generation) is $1.7 \cdot 10^{-3}$, higher than a previous estimate of $6.7 \cdot 10^{-6}$ (ref 65). This may be due
to genetic instability caused by heat stress⁵. In addition, we find no evidence for increased mutation
262 rates in aneuploid cells. Previous empirical studies have suggested that genetic instability (e.g.,
elevated mutation rates) in aneuploid cells is due to stress associated with the aneuploid state^{3,6,63,21}.
264 However, in the experiment of Yona et al.⁶², both the wild-type and the aneuploid were under heat
stress, which may explain why we did not find evidence for an increased mutation rate specifically in
266 aneuploid cells.

Conclusions. Here, we tested the hypothesis that aneuploidy cells are an evolutionary "stepping
268 stone", or adaptive intermediate, between wild-type euploid cells and mutant euploid cells. Our
results suggest that, although it seems the population goes from euploid to aneuploid and back, this is
270 not the case at the individual level. We estimate that only about 10-15% of the euploid cells descended
from aneuploid cells, whereas the rest are direct descendants of the wild-type euploid cells. Thus,
272 aneuploidy can delay, rather than accelerate, adaptation, and cells that become aneuploid may
leave less descendants than cells that remain euploid. This surprising result reinforces the importance
274 of models when making interpretations on evolutionary processes, and emphasizes the unintuitive
outcomes of clonal interference during adaptive evolution.

276 Models and Methods

DNA sequencing. Whole-genome sequencing of the ancestral diploid strain ($2n$) was performed
278 on a single colony of the ancestor. Whole-genome sequencing of the four evolving populations ($H2$
after 450 and 2,350 generations, and $H4$ after 450 and 1,700 generations) was performed on a sample
280 from these populations (rather than from single colonies) in order to maintain the population diversity.
Cells were grown in 5ml of YPD medium, either at 30 °C (ancestral diploid) or 39 °C (evolved
282 populations) in shaking conditions (200rpm) until reaching stationary phase. Following growth, 3ml
of each culture were centrifuge (14,000rpm) and cell pellets were used for DNA extraction. Genomic
284 DNA was extracted using $\text{A}^{\text{T}}\text{I}\text{J}\text{M}\text{a}\text{s}\text{t}\text{e}\text{r}\text{P}\text{u}\text{r}\text{e}$ Yeast DNA Purification Kit $\text{A}^{\text{T}}\text{I}$ (Lucigen) according to the
manufacture instructions. Following extraction, DNA concentrations were determined by Qubit assay
286 (Thermo Fisher) and 1 μ g DNA was used for library preparation using Illumina sample preparation
kit (Illumina). Samples were sequenced using a 100 bp pair end read output run using Illumina

Evolutionary genetic model. We model the evolution of a population of cells using a Wright-Fisher model³⁵, assuming a constant effective population size N , non-overlapping generations, and including the effects of natural selection, genetic drift, aneuploidy, and mutation. We focus on beneficial genetic modifications, neglecting the effects of deleterious and neutral mutations or karyotypic changes. The model allows for a single aneuploid karyotype (e.g., chromosome III duplication) and a single mutation to accumulate in the genotype. Thus, the model follows four genotypes (Figure 2): euploid wild-type, $2n$, the initial genotype; euploid mutant, $2n^*$, with the standard karyotype and a single beneficial mutation; aneuploid wild-type, $2n+1$, with an extra chromosome, i.e., following chromosome duplication; and aneuploid mutant, $2n+1^*$, with an extra chromosome and a beneficial mutation.

Transitions between the genotypes occur as follows (Figure 2): Beneficial mutations from $2n$ to $2n^*$ and from $2n+1$ to $2n+1^*$ occur with probability μ , the mutation rate. We neglect back-mutations (i.e., from $2n^*$ to $2n$ and from $2n+1^*$ to $2n+1$). Aneuploidy is formed by chromosome mis-segregation, so that cells transition from $2n$ to $2n+1$ and from $2n+1^*$ to $2n^*$ with probability δ , the aneuploidy rate. That is, we assume chromosomes are gained and lost at the same rate, and we neglect events that form a less-fit genotype (i.e., $2n+1$ to $2n$ and $2n^*$ to $2n+1^*$). A model that assumed increased aneuploidy rates in aneuploid cells did not perform well and was abandoned.

In the experiment by Yona et al.⁶², the population was grown every day from $1.6 \cdot 10^6$ cells until reaching stationary phase and then diluted 1:120. Thus, we set the population size to $N = 6.425 \cdot 10^6$, the harmonic mean of $\{2^k \cdot 1.6 \cdot 10^6\}_{k=0}^7$ ⁹. The initial population has N cells with genotype $2n$. The effect of natural selection on the frequency f_i of genotype $i = 2n, 2n + 1, 2n + 1^*,$ or $2n^*$ is given by

$$310 \quad f_i^s = \frac{f_i w_i}{\bar{w}}, \quad (1)$$

where w_i is the fitness of genotype i and $\bar{w} = \sum_j f_j w_j$ is the population mean fitness. The effect of mutation and aneuploidy on genotype frequencies is given by

$$\begin{aligned} f_{2n}^m &= (1 - \delta - \mu)f_{2n}^s, \\ f_{2n+1}^m &= \delta f_{2n}^s + (1 - \mu)f_{2n+1}^s, \\ f_{2n+1^*}^m &= \mu f_{2n+1}^s + (1 - \delta)f_{2n+1^*}^s, \\ f_{2n^*}^m &= \mu f_{2n}^s + \delta f_{2n+1}^s + f_{2n^*}^s. \end{aligned} \quad (2)$$

314 Finally, random genetic drift is modeled using a multinomial distribution³⁵,

$$\mathbf{f}' \sim \frac{1}{N} \cdot \text{Mult}(N, \mathbf{f}^m), \quad (3)$$

316 where $\mathbf{f}^m = (f_{2n}^m, f_{2n+1}^m, f_{2n+1^*}^m, f_{2n^*}^m)$ are the frequencies of the genotypes after mutation and
 318 aneuploidy, \mathbf{f}' are the genotype frequencies in the next generation, and $Mult(N, \mathbf{f})$ is a multinomial distribution parameterized by the population size N and the genotype frequencies \mathbf{f} . Overall, the change in genotype frequencies from one generation to the next is given by the transformation $f_i \rightarrow f'_i$.

320 **Empirical data for model inference.** We use the results of evolutionary experiments reported by Yona et al.⁶². In their heat-stress experiment, four populations of *S. cerevisiae* evolved under 39 °C.
 322 Aneuploidy fixed in all four population in the first 450 generations. Hereafter, fixation or elimination of a genotype *by generation t* means that more than 95% or less than 5% of the population carry the
 324 genotype at generation t , and possibly earlier. From re-analysis of data not published in the original paper, aneuploidy did not fix before at least 200 generations elapsed. The experiment continued with
 326 two populations, in which aneuploidy was eliminated by generation 1,700 and 2,350 while still under the same conditions of elevated heat (39 °C).

328 **Likelihood function.** Because our model, just like the Wright-Fisher model, is non-linear and stochastic, computing the distribution of fixation time $T(g)$ of genotype g for use in the likelihood
 330 function is intractable (it is even hard to use a diffusion-equation approximation due to the model having multiple genotypes, rather than just two). We overcome this problem by approximating the likelihood
 332 using simulations. We simulate 1,000 experiments per parameter vector $\theta = (\mu, \delta, s, b, c)$, resulting in a set of simulated observations $\tilde{\mathbf{X}} = \{\tilde{X}_i\}_{i=1}^{1000}$. We then compute the approximate likelihood,

$$\begin{aligned} \mathcal{L}(\theta) = P^4(200 \leq T(2n + 1) \leq 450) \cdot & \left[1 - \right. \\ & P_{\tilde{\mathbf{X}}}^4(\{T(2n^*) < 1700\} \mid 200 \leq T(2n + 1) \leq 450) - \\ & P_{\tilde{\mathbf{X}}}^4(\{1700 < T(2n^*) < 2350\} \mid 200 \leq T(2n + 1) \leq 450) + \\ & \left. P_{\tilde{\mathbf{X}}}^4(\{T(2n^*) < 1700\} \wedge \{1700 < T(2n^*) < 2350\} \mid 200 \leq T(2n + 1) \leq 450) \right], \end{aligned} \quad (4)$$

334 where $\{ \dots \}$ is the "logical not" operator, $P^4(\dots)$ is the 4th power of $P(\dots)$, and all probabilities
 336 $P_{\tilde{\mathbf{X}}}(\dots)$ are approximated from the results of the simulations $\tilde{\mathbf{X}}$. For example, $P_{\tilde{\mathbf{X}}}(\{T(2n^*) < 1700\} \mid$
 338 $200 \leq T(2n + 1) \leq 450)$ is approximated by taking simulations in which $2n+1$ fixed before generation
 340 450 but not before generation 200, and computing the fraction of such simulations in which $2n^*$ did not fix by generation 1,700, and hence aneuploidy did not extinct before generation 1,700. Figure S1 compares results with less and more simulated experiments, demonstrating that 1,000 simulations are likely sufficient.

342 For a model without aneuploidy (that is, when the aneuploidy rate is fixed at zero, $\delta = 0$), we disregard

the increased expression in chromosome III and the growth advantage measured in generation 450, and

344 focus on the growth advantage measured in later generations, presumably due to a beneficial mutation.

Therefore, the likelihood is approximated by

$$\begin{aligned} \mathcal{L}_!(\theta) = & 1 - P_{\tilde{\mathbf{X}}}^4(\{T(2n^*) < 1700\}) - \\ 346 \quad & P_{\tilde{\mathbf{X}}}^4(\{1700 < T(2n^*) < 2350\}) + \\ & P_{\tilde{\mathbf{X}}}^4(\{T(2n^*) < 1700\} \wedge \{1700 < T(2n^*) < 2350\}). \end{aligned} \quad (5)$$

Parameter inference. To infer model parameters, we use approximate Bayesian computation with

348 a sequential Monte-Carlo scheme, or ABC-SMC⁵², implemented in the `pyABC` Python package²⁶

pyabc.readthedocs.io. This approach uses numerical stochastic simulations of the model to infer

350 a posterior distribution over the model parameters. It is a method of likelihood-free, simulation-

based inference⁸, that is, for estimating a posterior distribution when a likelihood function cannot be

352 directly computed. It is therefore suitable in our case, in which the likelihood function can only be

approximated from simulations, and cannot be directly computed.

344 The ABC-SMC algorithm employs sequential importance sampling over multiple iterations^{55,25,53}. In

iteration t of the algorithm, a set of parameter vectors, $\{\theta_{i,t}\}_{i=1}^{n_t}$, also called *particles*, are constructed

356 in the following way. A proposal particle, θ^* , is sampled from a proposal distribution, and is either

accepted or rejected, until n_t particles are accepted. The number of particles, n_t , is adapted at every

358 iteration t using the adaptive population strategy²⁶ pyabc.readthedocs.io. For $t = 0$, the proposal

particle is sampled from the prior distribution, $p(\theta)$. For $t > 0$, the proposal particle is sampled from

360 the particles accepted in the previous iteration, $\{\theta_{i,t-1}\}_{i=1}^{n_{t-1}}$, each with a probability relative to its weight

$W_{t-1}(\theta_{i,t-1})$ (see below). The proposal particle is then perturbed using a kernel perturbation kernel,

362 $K_t(\theta^* | \theta)$ where θ is the sample from the previous iteration. Then, a set of synthetic observations

$\tilde{\mathbf{X}}^*$ is simulated, and the proposal particle θ^* is accepted if its approximate likelihood (eq. (4)) is high

364 enough, $\mathcal{L}(\theta^*) > 1 - \epsilon_t$ (or more commonly, if $1 - \mathcal{L}(\theta^*) < \epsilon_t$), where $\epsilon_t > 0$ is the *acceptance*

threshold, as higher values of ϵ_t allow more particles to be accepted. The acceptance threshold ϵ_t

366 is chosen as the median of the $1 - \mathcal{L}(\theta)$ of the particles accepted in the previous iteration, $t - 1$,

and $\epsilon_0 = 0.01$. For each accepted particle $\theta_{i,t}$ a weight $W_t(\theta_{i,t})$ is assigned: for $t = 0$, $W_0(\theta_{i,0}) = 1$,

368 and for $t > 0$, $W_t(\theta_{i,t}) = p(\theta_{i,t}) / \sum_{i=1}^{n_{t-1}} W_{t-1}(\theta_{i,t-1}) K_t(\theta_{i,t}, \theta_{i,t-1})$, where $p(\theta)$ is the prior density of θ

and $K_t(\theta', \theta)$ is the probability of a perturbation from θ to θ' . $K_t(\theta' | \theta)$ is a multivariate normal

370 distribution, fitted at iteration t to the particles from the previous iteration, $\{\theta_{i,t-1}\}_{i=1}^{n_{t-1}}$, and their

weights, $\{W(\theta_{i,t-1})\}_{i=1}^{n_{t-1}}$.

372 Acceptance is determined according to the approximate likelihood (eq. (4)), which has a maximum

value of $\mathcal{L}_{max} = 0.875$ (giving a minimal value of $\epsilon_{min} = 0.125$). We terminated the inference
 374 iterations when the change in ϵ value from one iteration to the next was small. With our standard prior
 and model, we reached $\epsilon = 0.13$ (or $\mathcal{L} = 0.87$) after six iterations, with $n_6 = 982$ accepted parameter
 376 vectors and effective sample size ESS=651 (Figure S2). Running the inference algorithm with different
 initialization seeds and less or more simulations for approximating the likelihood produced similar
 378 posterior distributions (Figure S1).

After producing a set of weighted particles from the the posterior distribution using the above ABC-
 380 SMC algorithm, we approximate the posterior using kernel density estimation (KDE) with Gaussian
 kernels. We truncate the estimated posterior to avoid positive posterior density for values with zero
 382 prior density. The MAP (maximum a posteriori) estimate is computed as the the maximum of the
 estimated joint posterior density. We then draw 5,000,000 samples from the posterior distribution
 384 to compute the HDI (highest density interval) and draw 50,000 samples to visualize the posterior
 distribution with histograms.

386 **Model comparison.** We examine several versions of our evolutionary models, e.g. without aneuploidy or with increased mutation rate in aneuploid cells, as well as several different prior distributions
 388 (see below). To compare these, we plot posterior predictions: for each model we execute 10,000
 simulations using the MAP parameter estimates and plot the distributions of time to fixation of $2n^*$,
 390 one of key properties of the model likelihood. These plots visualize the fit of each model to the
 data. Also, for similar models we plot the marginal and joint posterior distributions of the parameters;
 392 if these are similar, we consider the models interchangeable. We validate this by comparing HDI
 (highest density interval) of posterior distributions.
 394 Where posterior plots are very similar and the number of parameters is the same, we use WAIC, or
 the widely applicable information criterion ¹³, defined as

$$396 \quad WAIC(\theta) = -2 \log \mathbb{E}[\mathcal{L}(\theta)] + 2\mathbb{V}[\log \mathcal{L}(\theta)] \quad (6)$$

where θ is a parameter vector, and $\mathbb{E}[\cdot]$ and $\mathbb{V}[\cdot]$ are the expectation and variance taken over the
 398 posterior distribution, which in practice are approximated using 50,000 samples from the posterior
 KDE. We validated that upon resampling WAIC values do not significantly change and that differences
 400 in WAIC between models are preserved. WAIC values are scaled as a deviance measure: lower values
 imply higher predictive accuracy²³.

402 **Prior distributions.** We used informative prior distributions for $w_{2n+1} = 1 - c + b$, $w_{2n+1^*} = (1+s)(1-c)+b$ and $w_{2n^*} = 1+s$, which we estimated from growth curves data from mono-culture growth
404 experiments previously reported by Yona et al.⁶², Figs. 3C, 4A, and S2. We used Curveball, a method
for predicting results of competition experiments from growth curve data³⁸ curveball.yoavram.com.
406 Briefly, Curveball takes growth curves of two strains growing separately in mono-culture and predicts
how they would grow in a mixed culture, that is, it predicts the results of a competition assay. From these
408 predictions, relative fitness values can be computed. Because Curveball uses a maximum-likelihood
approach to estimate model parameters, we were able to estimate a distribution of relative fitness
410 values to be used as a prior distribution by sampling 10,000 samples from a truncated multivariate
normal distribution defined by the maximum-likelihood covariance matrix (Figure S3).

412 We used growth curves of $2n$ and $2n+1$ in 39 °C to estimate an informative prior distribution for
 w_{2n+1} (Figure S3-D, assuming $w_{2n} = 1$). In this prior distribution, we used the same prior for w_{2n+1^*}
414 and w_{2n^*} . To increase computational efficiency, we also assumed $w_{2n^*} > w_{2n+1^*} > w_{2n+1} > w_{2n}$;
running the inference without this assumption produced similar results. See *supporting material* for
416 an extended informative prior distribution that uses growth curves of $2n^*$ and $2n+1$ growing in 39 °C;
this prior distribution proved to be less useful.

418 As a control, we tested an uninformative uniform prior with $U(1, 6)$, for (i) all w_{2n+1} , w_{2n+1^*} , w_{2n^*} , or
(ii) only for w_{2n+1^*} , w_{2n^*} , using the above informative prior for w_{2n+1} . In these cases the inference
420 algorithm failed to converge.

For the mutation rate, μ , and aneuploidy rate, δ , we used uninformative uniform priors, $\mu \sim$
422 $U(10^{-9}, 10^{-5})$ and $\delta \sim U(10^{-6}, 10^{-2})$. A wider mutation rate prior, $\mu \sim U(10^{-9}, 10^{-3})$, produced
similar results.

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Supplementary Material

596 Supplementary Analysis

Sensitivity analysis. Changing a single parameter while keeping the rest fixed at the MAP estimate produces a worse fit to the data (Figure S6). Furthermore, we fitted models with a mutation rate fixed at $\mu = 10^{-5}$, 10^{-6} and 10^{-7} . We inferred similar parameters estimates for the model with $\mu = 10^{-6}$ compared to the model with a free μ parameter, in which the inferred mutation rate is $\mu \approx 3 \cdot 10^{-6}$. Inference assuming $\mu = 10^{-5}$ or $\mu = 10^{-7}$ produced similar estimates except that the estimated aneuploidy rate, δ , was higher, and assuming $\mu = 10^{-7}$, the estimated fitness of $2n+1$ was lower (Figure S7).

604 **Extended informative prior distribution.** In an extended informative prior distribution, we used additional growth curves of $2n^*$ (*refined* strain from Yona et al.⁶²) and $2n+1$ in 39 °C to estimate w_{2n^*}/w_{2n+1} (Figure S3L). The same distribution was used for w_{2n^*}/w_{2n+1*} . Thus, our main informative prior uses a single prior distribution for fitness values of $2n+1$, $2n+1^*$, and $2n^*$, whereas the extended informative prior uses one distribution for $2n+1$, and another distribution for both $2n+1^*$ and $2n^*$.

610 We estimated the parameters under this extended informative prior. Inference took much longer to run but the posterior distribution seemed to converge, as it did not change much in the final iterations. The posterior predictive plot shows that inference with this extended prior produces a posterior distribution that fails to explain the empirical observations (pink in Figure 4). However, 614 the inferred posterior distribution is considerably narrower (compare Figures 3 and S8) and therefore parameter estimates are less variable. The estimated mutation rate was much lower compared to 616 the main informative prior, with $\mu = 2.474 \cdot 10^{-9}$ [$2.423 \cdot 10^{-9} - 2.612 \cdot 10^{-9}$]. Other parameter estimates are: $\delta = 2.705 \cdot 10^{-3}$ [$2.094 \cdot 10^{-3} - 3.094 \cdot 10^{-3}$], $w_{2n+1} = 1.022$ [$1.021 - 1.024$], 618 $w_{2n+1*} = 1.052$ [$1.05 - 1.054$], $w_{2n^*} = 1.053$ [$1.051 - 1.055$], the latter two being much higher compare to the main informative prior. Notably, the mode of the posterior ratio $w_{2n^*}/w_{2n+1} = 1.0009$ 620 is much lower than the mode of the prior ratio of 1.033 (Figure S3H) and closer to the ratio of 1 that we assume in the main informative prior. Together with the posterior predictive results, we conclude 622 that the main informative prior is preferable over the extended informative prior.

Model with transitions to less-fit genotypes We also estimated the parameters of a version of the

624 model that includes transitions (mutation, chromosome loss and gain) to less-fit genotypes (e.g., $2n^*$ to $2n+1^*$),

$$\begin{aligned}
 f_{2n}^m &= (1 - \delta - \mu)f_{2n}^s + \delta f_{2n+1}^s + \mu f_{2n+1}^s, \\
 f_{2n+1}^m &= \delta f_{2n}^s + (1 - \delta - \mu)f_{2n+1}^s + \mu f_{2n+1}^s, \\
 f_{2n+1}^m &= \mu f_{2n+1}^s + (1 - \delta - \mu)f_{2n+1}^s + \delta f_{2n}^s, \\
 f_{2n}^m &= \mu f_{2n}^s + \delta f_{2n+1}^s + (1 - \delta - \mu)f_{2n}^s.
 \end{aligned} \tag{7}$$

The inferred values are slightly different. The estimated mutation rate, $\mu = 1.036 \cdot 10^{-7}$ [8.01 ·

628 $10^{-8} - 1.339 \cdot 10^{-7}$], corresponds to a mutation target size of $\sim 300 - 500$, assuming the mutation
rate per base pair is roughly $2 \cdot 10^{-10}$ (ref.⁶⁶) or $3.3 \cdot 10^{-10}$ (ref.³⁰). The estimated aneuploidy

630 rate, $\delta = 2.358 \cdot 10^{-4}$ [$1.766 \cdot 10^{-4} - 2.837 \cdot 10^{-4}$] is 5-35-fold higher than in previous studies:
for chromosome III in diploid *S. cerevisiae*, Zhu et al.⁶⁶ estimated $6.7 \cdot 10^{-6}$ chromosome gain

632 events per generation, and Kumaran et al.²⁹ estimate $3.0 - 4.3 \cdot 10^{-5}$ chromosome loss events per
generation (95% confidence interval). The estimated fitness values are $w_{2n+1} = 1.024$ [1.023 – 1.025],

634 $w_{2n+1}^* = 1.025$ [1.024 – 1.026], $w_{2n} = 1.032$ [1.031 – 1.033], all relative to the fitness of $2n$, which
is set to $w_{2n} = 1$. Thus, we can infer that the cost of trisomy is $c = w_{2n} - w_{2n+1}^* = 0.007$ (or 0.7%)

636 and the benefit of trisomy is $w_{2n+1} - 1 - c = 0.017$ (1.7%), whereas the benefit of beneficial mutation
is $w_{2n} - 1 = 0.032$ (3.2%).

638 We simulated genotype frequency dynamics using parameter samples from the posterior distribution,
and computed the posterior distribution of F_A . The mean F_A in this case is just 0.0189 [0.0004 - 0.1214
640 95% CI], lower than without the transitions to less-fit genotypes. Here, F_A is the sum of frequencies
of both $2n_A^*$ and $2n + 1_A^*$, which reaches a frequency of 0.0007. Out of 100,000 posterior samples,
642 none had F_A above 0.05 (i.e., 5% of the population).

Supplementary Figures & Tables

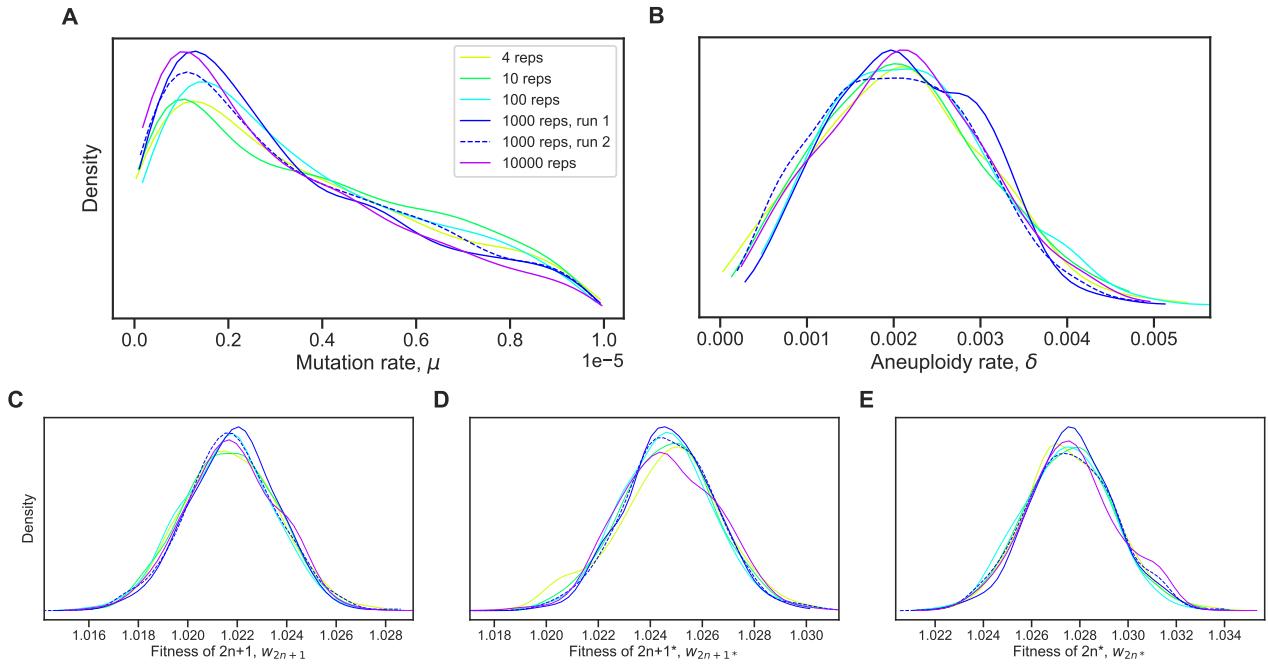


Figure S1: Posterior distribution validation. The posterior distribution of model parameters is roughly the same regardless of the number of simulations (4-10,000 replicates) used to approximate the likelihood (eq. (4)).

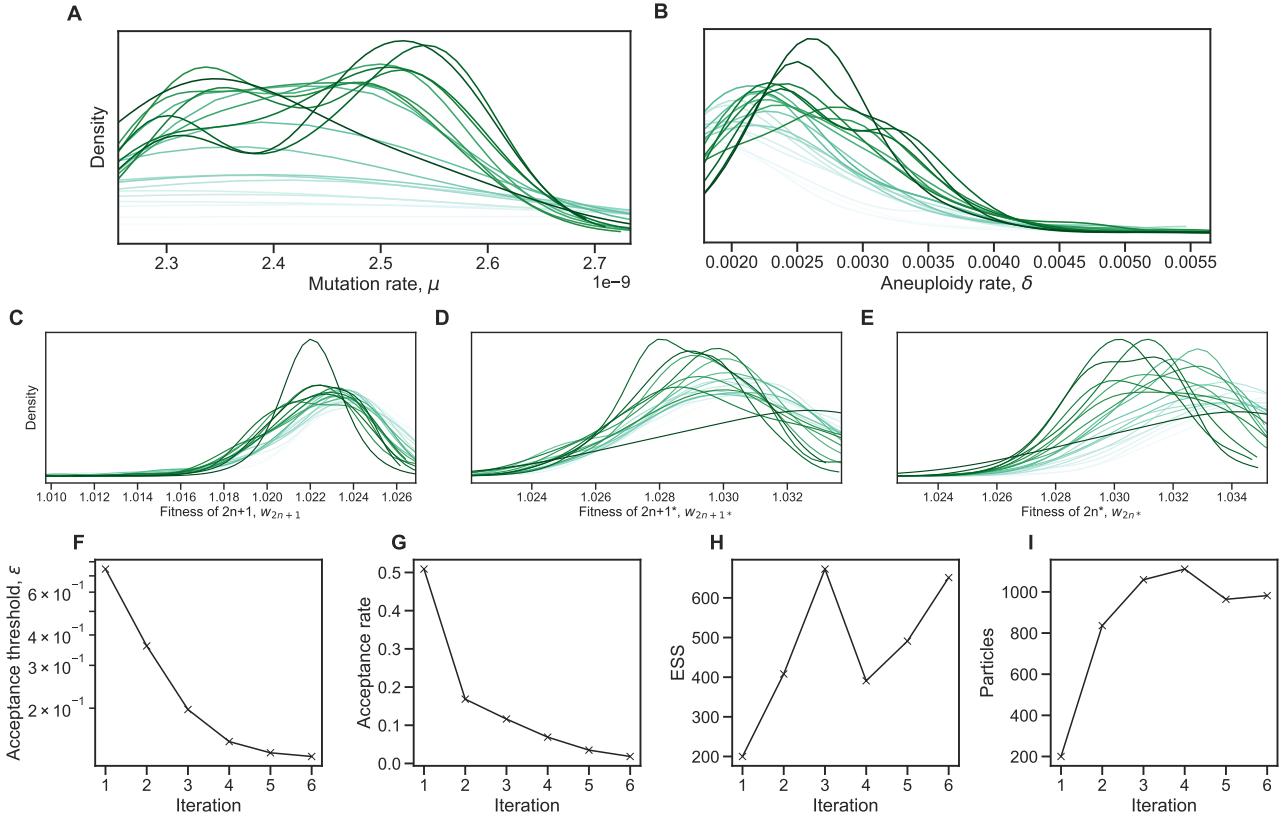


Figure S2: Inference convergence. The ABC-SMC algorithm was used to infer the model parameters. **(A-E)** The approximate posterior distributions of model parameters at each iteration of the ABC-SMC algorithm demonstrates convergence, as the posterior did not significantly change after the first iteration, $t = 1$. **(F-I)** ABC-SMC measures of convergence. After iteration number 6, the acceptance threshold was $\epsilon = 0.13$ (i.e., $\mathcal{L} = 0.87$, eq. (4)), the acceptance rate was 0.018, the number of particles was 982, and the effective sample size ESS=651.

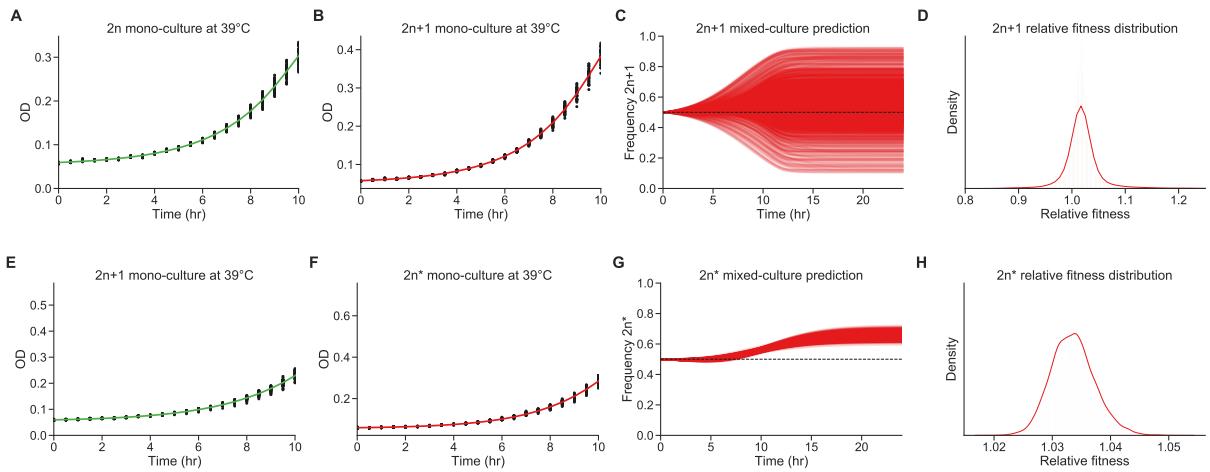


Figure S3: Fitness estimation from growth curves. **(A-D)** Fitness estimation from growth curves of $2n$ and $2n+1$ at 39°C . $w_{2n+1}/w_{2n}=1.024$ (95% CI: 0.959 - 1.115). **Curveball (E-H)** Fitness estimation from growth curves of $2n+1$ and $2n^*$ at 39°C . $w_{2n^*}/w_{2n+1}=1.033$ (95% CI: 1.027 - 1.041). Growth curves previously described in Yona et al.⁶², Figs. 3C, 4A, and S2. Fitness estimated from growth curves using Curveball, a method for predicting results of competition experiments from growth curve data³⁸ curveball.yoavram.com. See *Models and Methods, Prior distributions* for more details. **(A,B;E,F)** Mono-culture growth curve data (markers) and best-fit growth models (lines). **(C,G)** The mixed-culture prediction for the strains from A,B and E,F respectively, 6,375 generated curves. **(D,H)** The relative fitness distribution for $2n+1$ relative to $2n$ (panel D) and $2n^*$ relative to $2n+1$ (panel H). Figures generated by Curveball.

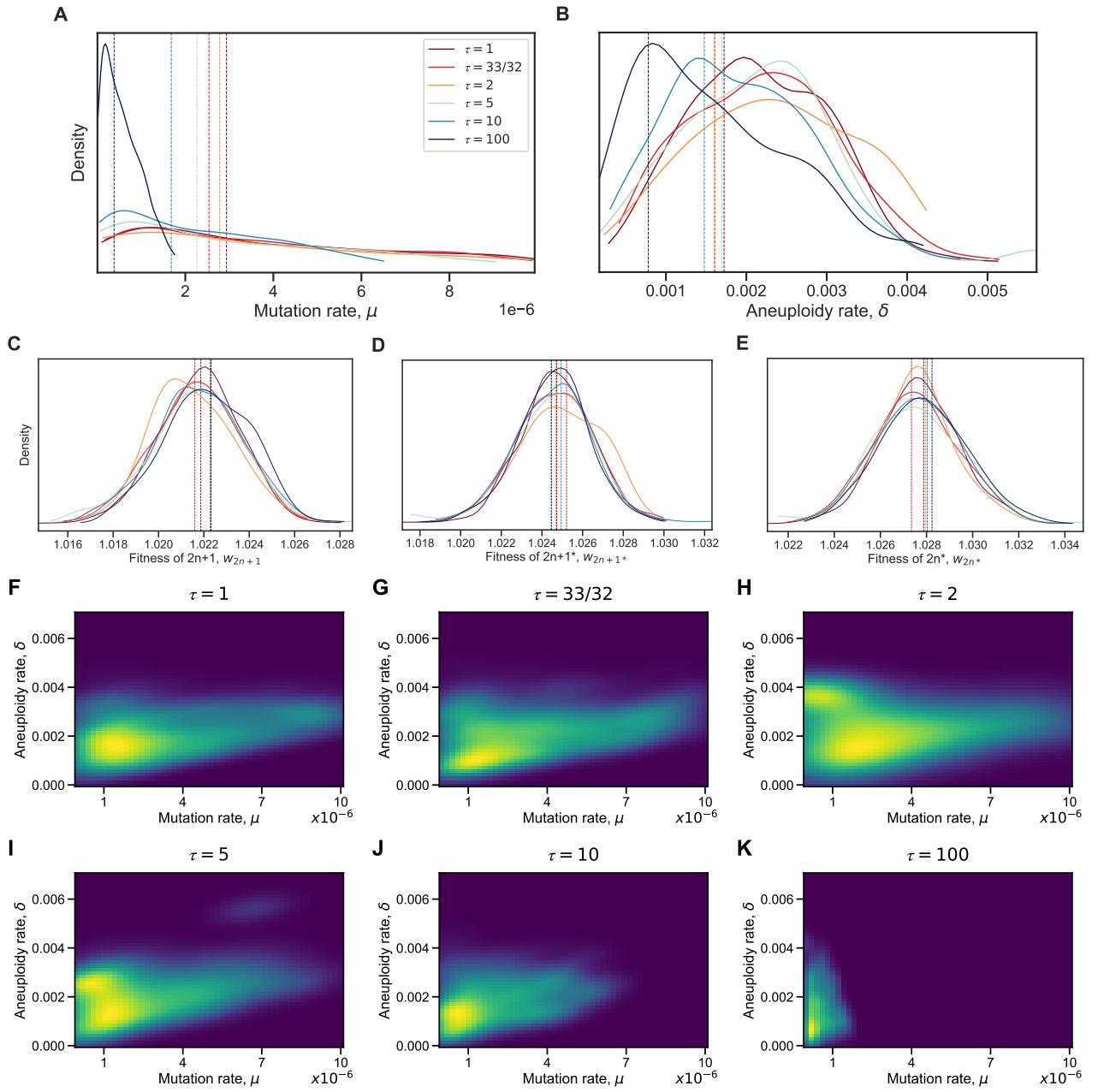


Figure S4: Model with elevated mutation rate in aneuploid cells. (A-E) The inferred posterior distributions for models with different values of τ , the fold-increase in mutation rate in aneuploid cells ($2n+1$ and $2n+1^*$). Vertical dashed lines represent the MAP (maximum a posteriori) of each distribution. When the increase in mutation rate is high, $\tau = 10$ and $\tau = 100$, the inferred mutation (A) and aneuploidy (B) rates tend to be lower. (F-K) The inferred joint posterior distribution of mutation rate (μ) and aneuploidy rate (δ) with different τ values (dark purple and bright yellow for low and high density, respectively).

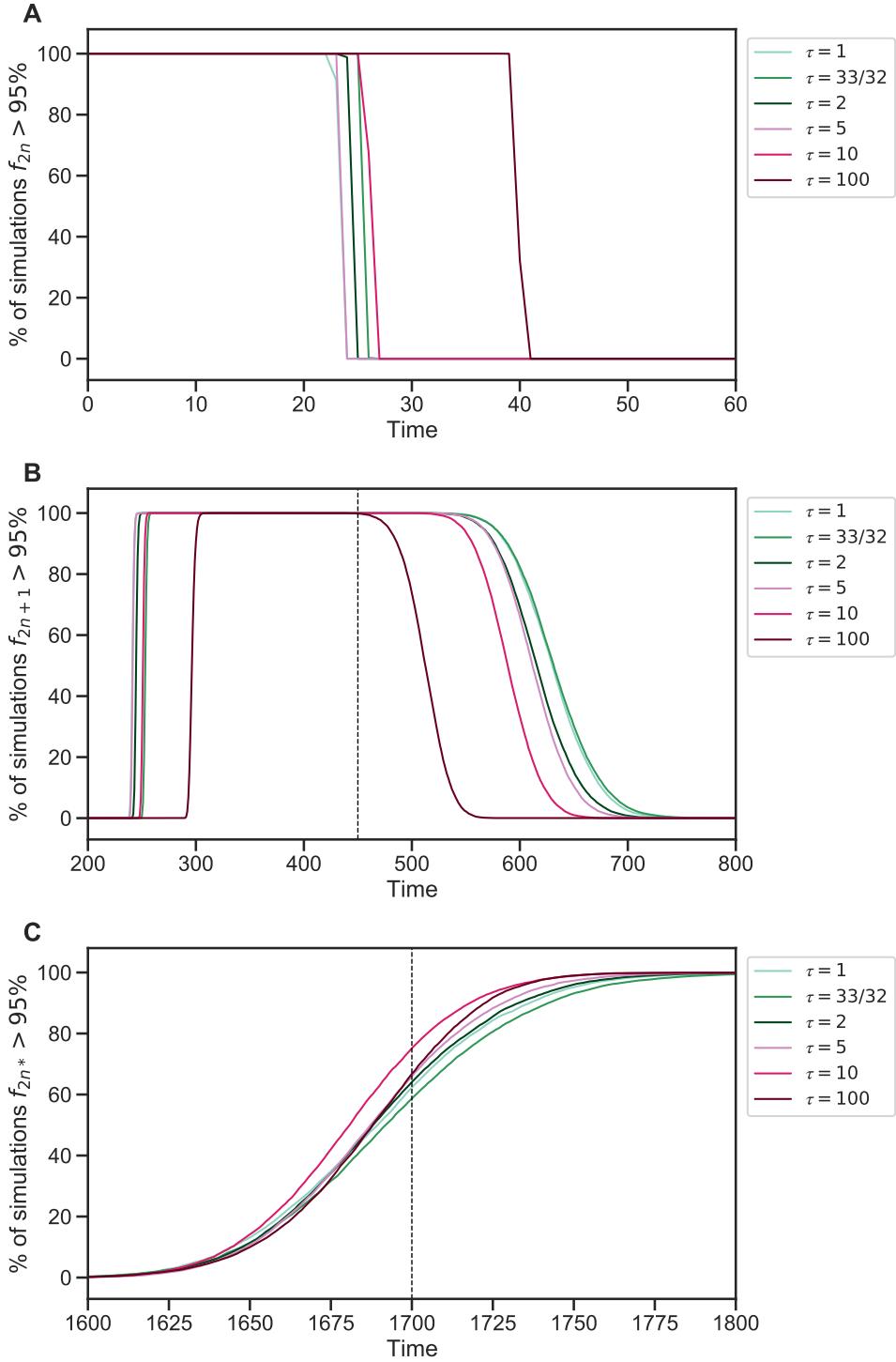


Figure S5: Genotype fixations for models with increased genetic instability. We estimated the parameters for different models, each assuming a different value of τ , the fold-increase in mutation rate in aneuploid cells. We then generated 10,000 simulations using the MAP estimate of each model and evaluated the fraction of simulations in which the frequency of genotype $2n$ (**A**), $2n+1$ (**B**), and $2n^*$ (**C**) is above 95% (y-axis) at each generation (x-axis). Note that $2n+1^*$ did not fix. We can see that $\tau = 100$ can be distinguished if the waiting time for $f_{2n} < 95\%$ is known (panel A) or if the waiting time for $f_{2n+1} > 95\%$ or $f_{2n+1} < 95\%$ is known (panel B). It is harder to distinguish between $1 \leq \tau \leq 10$.

Table S1: Mutant alleles in population $H2$.

Mutant alleles identified in the ancestor (generation 0), aneuploid (generation 450), and evolved (generation 2,350) of population $H2$. See supplementary file.

Table S2: Mutant alleles in population $H4$.

Mutant alleles identified in the ancestor (generation 0), aneuploid (generation 450), and evolved (generation 1,700) of population $H4$. See supplementary file.

Table S3: WAIC values for different τ values.

Model	WAIC
$\tau = 1$	-9
$\tau = 33/32$	-9
$\tau = 2$	-8
$\tau = 5$	-12
$\tau = 10$	-9
$\tau = 100$	-12

WAIC defined in eq. (6).

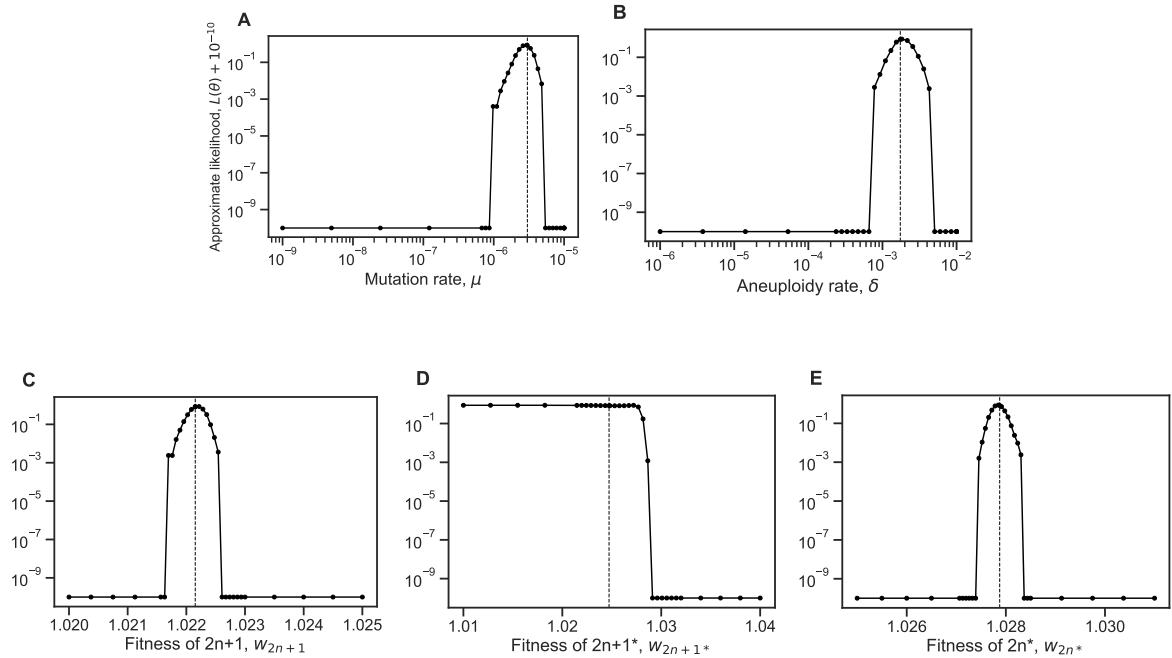


Figure S6: Likelihood profiles. Sensitivity of the model approximate likelihood, $\mathcal{L}(\theta)$, to changing a single parameter while the other parameters remain fixed at their MAP estimates. Dashed vertical line represents the MAP value. The prior distributions for the mutation rate and aneuploidy rate are $\mu \sim U(10^{-9}, 10^{-5})$ and $\delta \sim U(10^{-6}, 10^{-2})$, respectively.

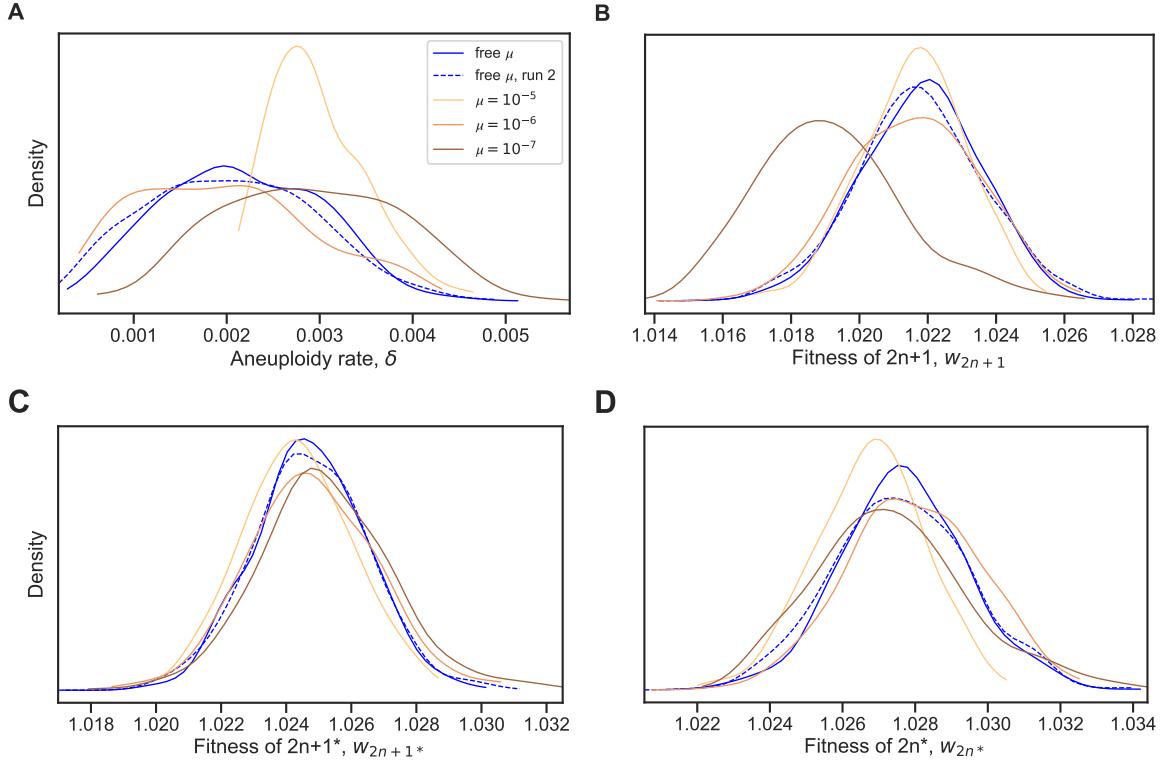


Figure S7: Model with fixed mutation rate. (A-D) The inferred posterior distributions for models with free and fixed mutation rate, μ . The MAP (maximum a posteriori) and 50% HDI (highest density interval) for each model are: **free μ , run 1:** $\delta = 1.720 \cdot 10^{-3}$ [$1.470 \cdot 10^{-3} - 2.786 \cdot 10^{-3}$], $w_{2n+1} = 1.022$ [1.021 – 1.023], $w_{2n+1^*} = 1.025$ [1.024 – 1.026], $w_{2n^*} = 1.028$ [1.026 – 1.029]; **free μ , run 2:** $\delta = 2.129 \cdot 10^{-3}$ [$1.334 \cdot 10^{-3} - 2.695 \cdot 10^{-3}$], $w_{2n+1} = 1.022$ [1.02 – 1.023], $w_{2n+1^*} = 1.025$ [1.023 – 1.026], $w_{2n^*} = 1.028$ [1.026 – 1.029]; **$\mu = 10^{-5}$:** $\delta = 2.903 \cdot 10^{-3}$ [$2.399 \cdot 10^{-3} - 3.156 \cdot 10^{-3}$], $w_{2n+1} = 1.022$ [1.021 – 1.023], $w_{2n+1^*} = 1.024$ [1.023 – 1.025], $w_{2n^*} = 1.027$ [1.026 – 1.028]; **$\mu = 10^{-6}$:** $\delta = 1.917 \cdot 10^{-3}$ [$9.624 \cdot 10^{-4} - 2.447 \cdot 10^{-3}$], $w_{2n+1} = 1.022$ [1.02 – 1.023], $w_{2n+1^*} = 1.025$ [1.023 – 1.026], $w_{2n^*} = 1.028$ [1.027 – 1.029]; **$\mu = 10^{-7}$:** $\delta = 2.901 \cdot 10^{-3}$ [$2.139 \cdot 10^{-3} - 3.671 \cdot 10^{-3}$], $w_{2n+1} = 1.019$ [1.017 – 1.02], $w_{2n+1^*} = 1.025$ [1.024 – 1.026], $w_{2n^*} = 1.027$ [1.026 – 1.029].

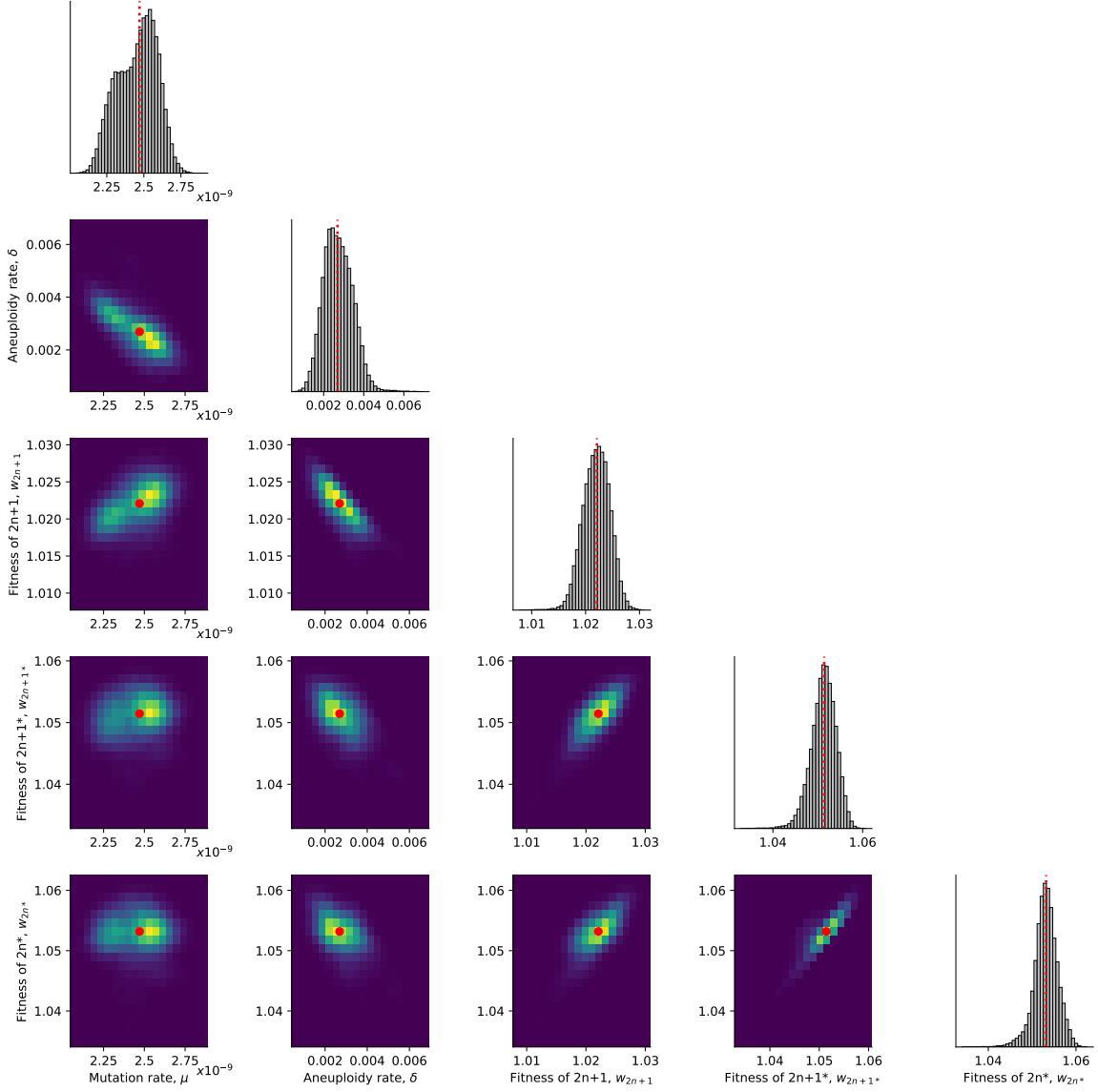


Figure S8: Posterior distribution of parameters inferred with the extended prior distribution. On the diagonal, the inferred posterior distribution of each model parameter. Below the diagonal, the inferred joint posterior distribution of pairs of model parameters (dark purple and bright yellow for low and high density, respectively). Red markers and orange lines for the joint MAP estimate (which may differ from the marginal MAP, as the marginal distribution integrates over all other parameters).

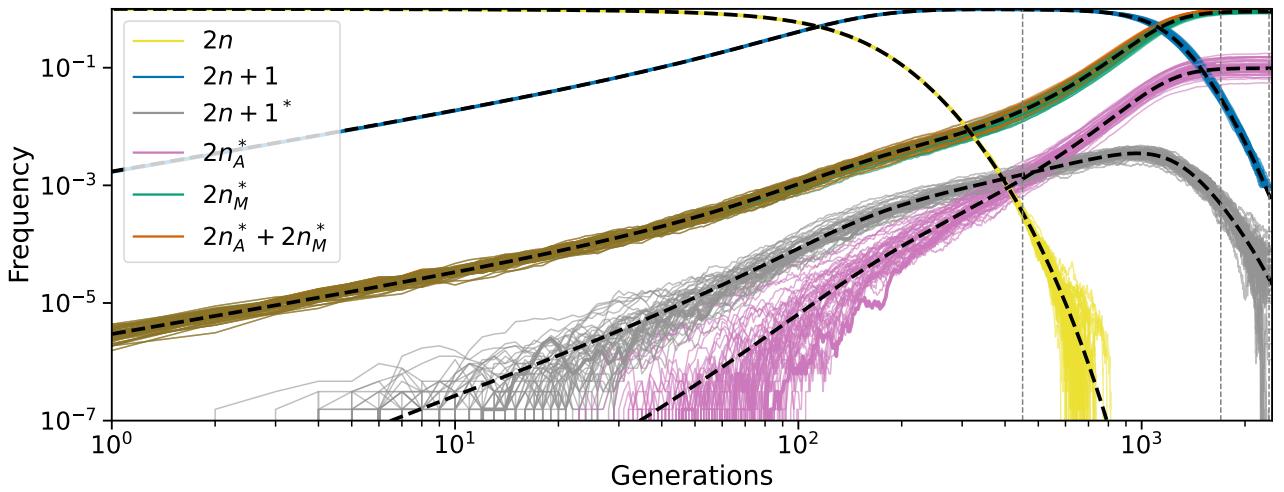


Figure S9: Posterior predicted genotype frequencies in log-log scale. Frequency dynamics of the different genotypes with MAP parameter estimates, same as Figure 5A, but in log-log scale. Black dashed curves for a deterministic model without genetic drift. Clearly, appearance of $2n+1$ and $2n_M^*$ is deterministic. Appearance of $2n+1^*$, and therefore $2n_A^*$, is stochastic, however, the frequency dynamics are deterministic above a frequency of roughly 0.001. Note that the $2n_M^*$ and the $2n_A^* + 2n_M^*$ lines are overlapping for much of their trajectories.

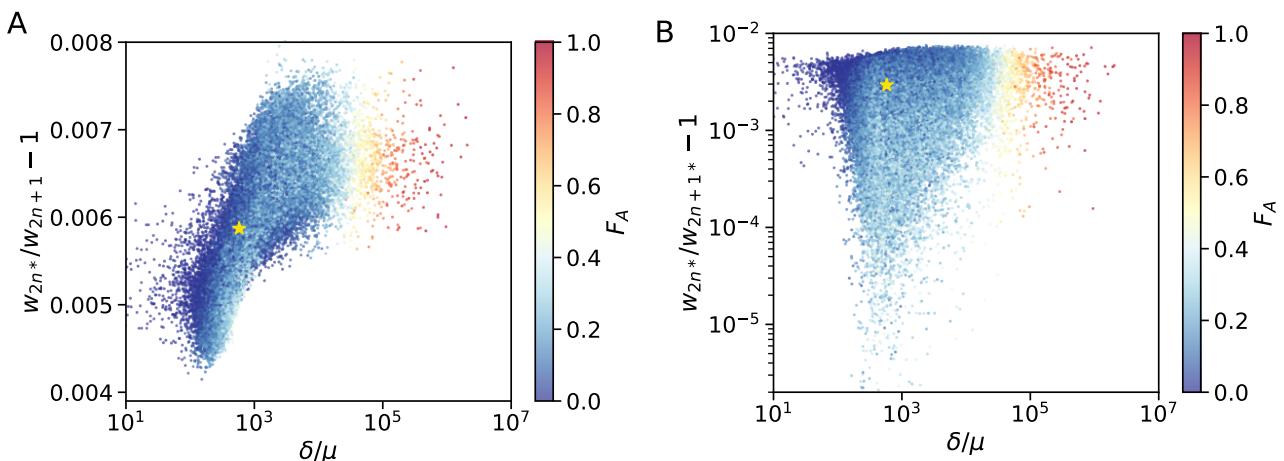


Figure S10: Posterior distribution of F_A . (A,B) F_A values (color coded) as in Figure 5 for different parameter choices on the x- and y-axes. White star denotes the MAP estimate.