

A short data exploration of the Fungi Kingdom in BugSigDB

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Overview

The fungi kingdom is an underrepresented taxon in microbiome studies (Perez and Johnson, 2013). Technical difficulties in sampling and identification, as well as a lack of quality controlled reference databases (Pérez, 2021) are some of the obstacles in including fungi in microbiome studies. Most studies have focused on fungal outgrowth when the host is compromised, with little known about the dynamics of the mycobiome during health (Huffnagle & Noverr, 2013).

This exploratory analysis seeks to assess how fungi is represented in the BugSigDB database. Reporting on the most common taxa, distribution across body sites, increasing and decreasing signatures, and diversity based on presence/absence data across experiments.

Setup and Data Import

Call libraries and import or load the BugSigDB database.

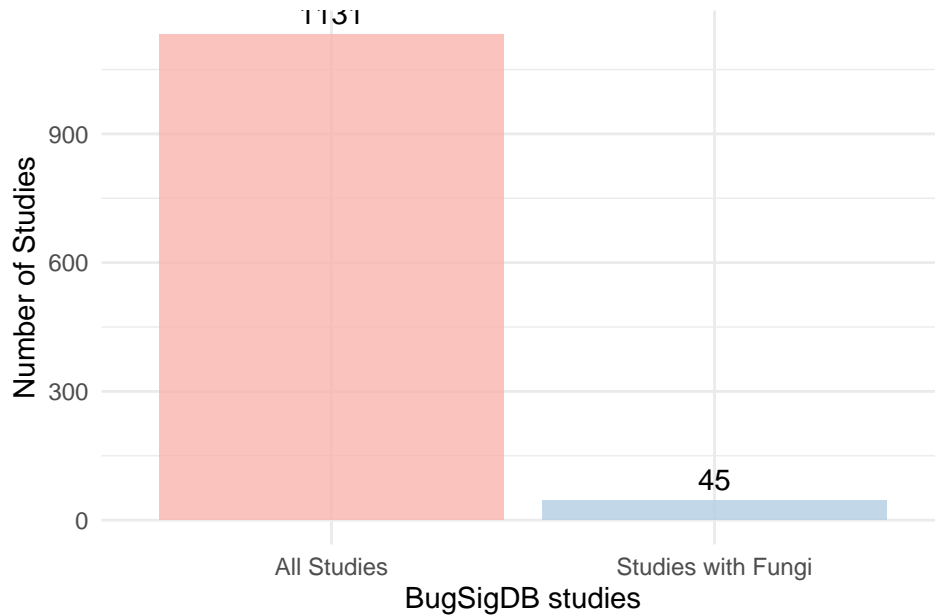
```
library(bugsigdbr)
library(BugSigDBStats)
library(tidyverse)
library(RColorBrewer)
# Import the full BugSigDB database
# bsdb <- importBugSigDB(cache = FALSE) # from BugSigDB
bsdb <- readRDS("bsdb.rds") # load from saved file
```

Identifying Studies and Experiments with Fungi Taxa

To identify the experiments that presented fungi taxa, values belonging to the Fungi kingdom were extracted from the `MetaPhlAn taxon names` column of the “bsdb” dataframe.

```
# Get experiments that have fungi
fungi <- grepl("k__Fungi", bsdb[["MetaPhlAn taxon names"]])
# Subset
fungi <- bsdb[fungi, ]
```

How many studies in BugSigDB have included fungi taxa in their analysis?



```
## The official BugSigDB release has a total of 1131 studies
## with 5520 experiments. 45 of these studies, with a total
## of 111 experiments included the Fungi kingdom in their analysis.
```

What sequencing type and method was used?

Fungi do not have the 16Sr RNA gene, instead a specific DNA region is targeted.

##	PMID	Platform	Region	Type
## 1	27007700	RT-qPCR	23	16S
## 2	30508628	RT-qPCR	345	16S
## 3	29031597	Illumina	4	16S
## 4	29783199	Roche454	NA	16S
## 5	30405584	Illumina	12	16S
## 6	31834952	Illumina	34	16S
## 7	33806027	Illumina	NA	16S
## 8	32934117	Illumina	34	16S
## 9	22949626	Roche454	4	16S
## 10	30576008	Illumina	34	16S
## 11	29276170	Illumina	4	16S
## 12	31247001	Illumina	NA	16S
## 13	32024712	Illumina	34	16S
## 14	NA	Illumina	NA	16S
## 15	31672155	Illumina	4	16S
## 16	30631088	Roche454	567	16S
## 17	33565055	Illumina	45	16S
## 18	38589368	Illumina	34	16S
## 19	37158877	Illumina	34	16S
## 20	34376634	Ion Torrent	34	16S

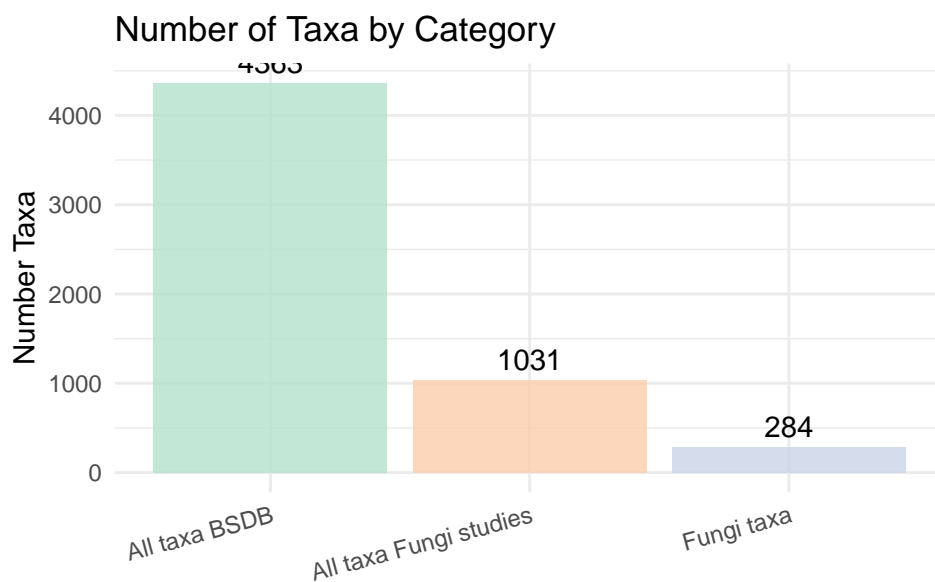
There are 20 studies that should be reviewed since 16S is listed as part of the sequencing protocol. In many cases there was more than one type of library used and the results for both were grouped together. BugSigDB does not allow for more than one type to be inputted and this might be the issue.

What taxa do we find in the experiments that have fungi?

Identify the taxonomic signatures in the experiments that contain fungi.

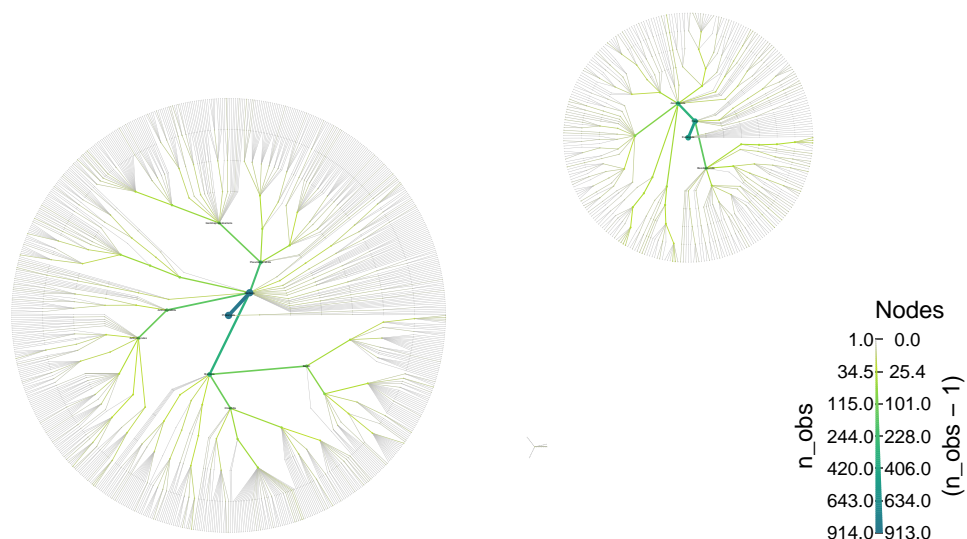
```
## [1] "Malassezia" "Malassezia globosa"
## [3] "Malassezia" "Malassezia globosa"
## [5] "Malassezia globosa" "Saccharomyces sp."
## [7] "[Candida] sp. (uncertain placement)" "Saccharomyces sp."
## [9] "[Candida] sp. (uncertain placement)" "Candida albicans"
```

What percentage of taxa in BugSigDB belong to the fungi kingdom?



```
## In the 4363 signatures curated in BugSigDB, 4079 taxa are classified as Bacteria, Archaeobacteria,
## Eukaryota or Virus. Out of these, 284 ( 6.5 %) belong to the Fungi kingdom.
```

Heattree for Bacteria, Archea, Fungi and Viruses found in fungi experiments



Unexpected taxa found

##	superkingdom	kingdom	phyla	class
## 1	Eukaryota	Metazoa	Chordata	Actinopteri
## 2	Eukaryota	Metazoa	Arthropoda	Insecta
## 3	Eukaryota	Evosea	Mastigamoebida	Entamoebidae
## 4	Eukaryota	Metazoa	Arthropoda	Collembola
## 5	Eukaryota	Euglenozoa	Kinetoplastea	Trypanosomatida
## 6	Eukaryota	Metazoa	Nematoda	Chromadorea
## 7	Eukaryota	Apicomplexa	<NA>	<NA>
## 8	Eukaryota	Apicomplexa	Aconoidasida	Haemosporida
## 9	Eukaryota	Cercozoa	Cercozoa sp. B134	<NA>
## 10	Eukaryota	Viridiplantae	Streptophyta	Magnoliopsida
## 11	Eukaryota	Cercozoa	Cercomonadida	Cercomonadidae
## 12	Eukaryota	Synurophyceae	Ochromonadales	Ochromonadaceae
## 13	Eukaryota	Metazoa	Arthropoda	Arachnida
##	order		family	genus
## 1	Gerreiformes		Gerreidae	Gerres
## 2	Blattodea		Rhinotermitidae	Coptotermitinae
## 3	Entamoeba		<NA>	<NA>
## 4	Poduromorpha		Hypogastruridae	Hypogastrura
## 5	Trypanosomatidae		Leishmaniinae	Leishmania
## 6	Rhabditida		Hoplolaimidae	Rotylenchulinae
## 7	<NA>		<NA>	<NA>
## 8	Plasmodiidae		Plasmodium	Plasmodium ovale
## 9	<NA>		<NA>	<NA>
## 10	Malpighiales		Linaceae	Linum
## 11	Paracercomonas		Paracercomonas sp.	<NA>
## 12	Poterioochromonas	Poterioochromonas malhamensis		<NA>
## 13	Ricinulei		Ricinoididae	Cryptocellus

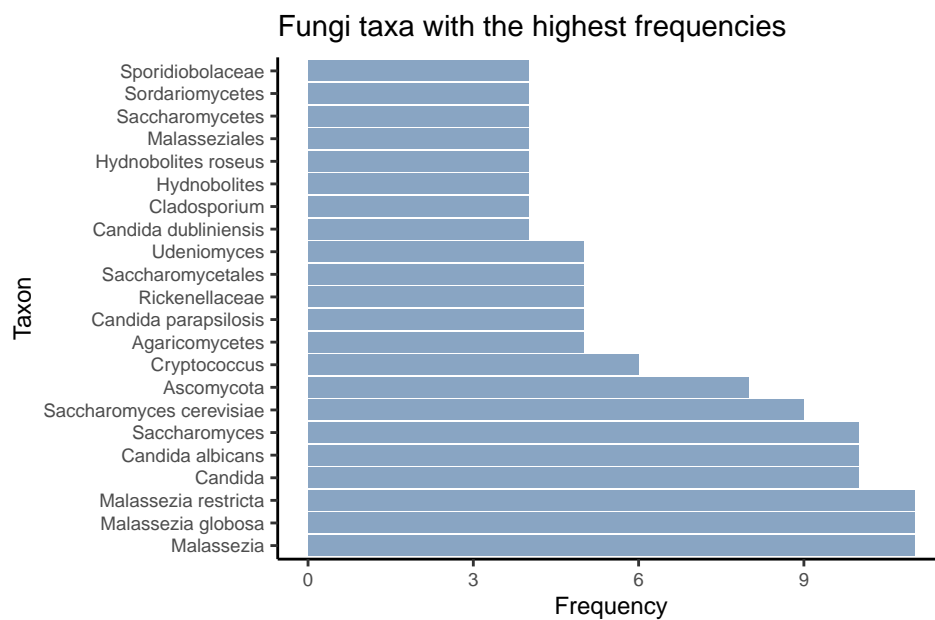
```

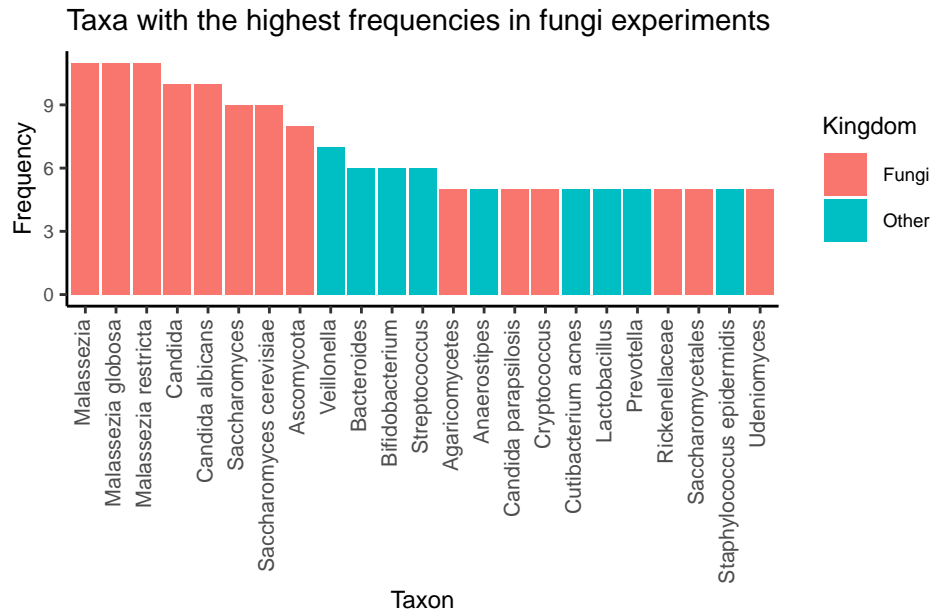
##               species
## 1      Gerres decacanthus
## 2      Coptotermes
## 3      <NA>
## 4 Hypogastrura sp. BOLD:AAI2333
## 5      <NA>
## 6      Rotylenchus
## 7      <NA>
## 8      <NA>
## 9      <NA>
## 10     Linum usitatissimum
## 11     <NA>
## 12     <NA>
## 13     Cryptocellus magnus

```

Exploring Frequencies

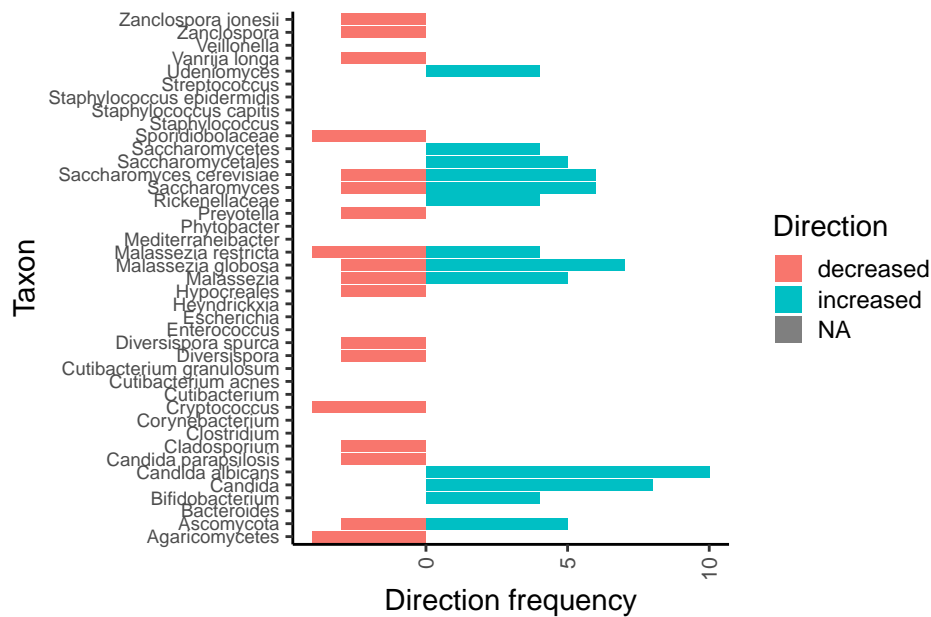
Which are the most frequent taxa in BugSigDB fungi experiments?





Increasing vs Decreasing signatures

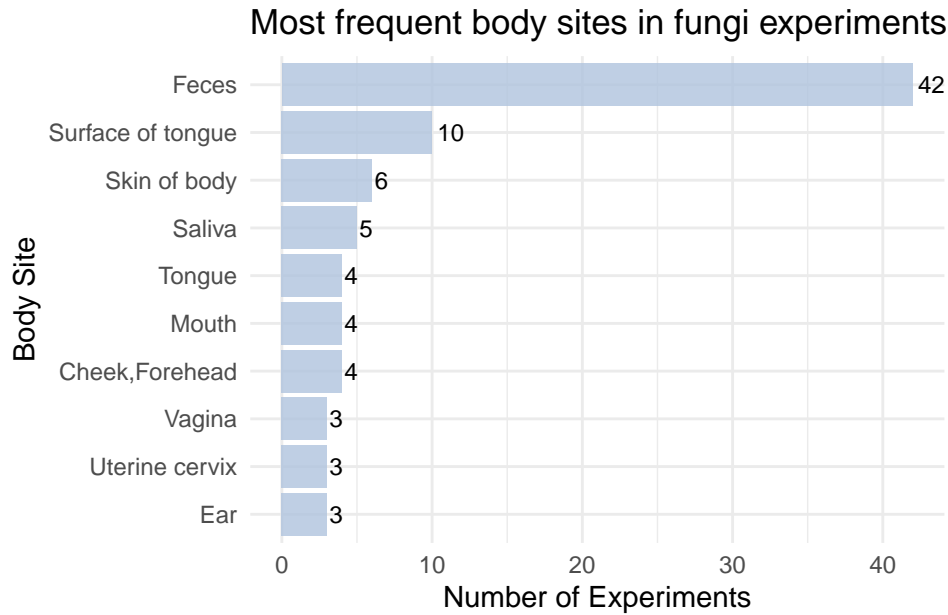
What taxa usually increases or decreases for the condition being tested?



Body Site Analysis

What body sites were tested for fungi? ### Body sites for the experiments that contain fungal taxa

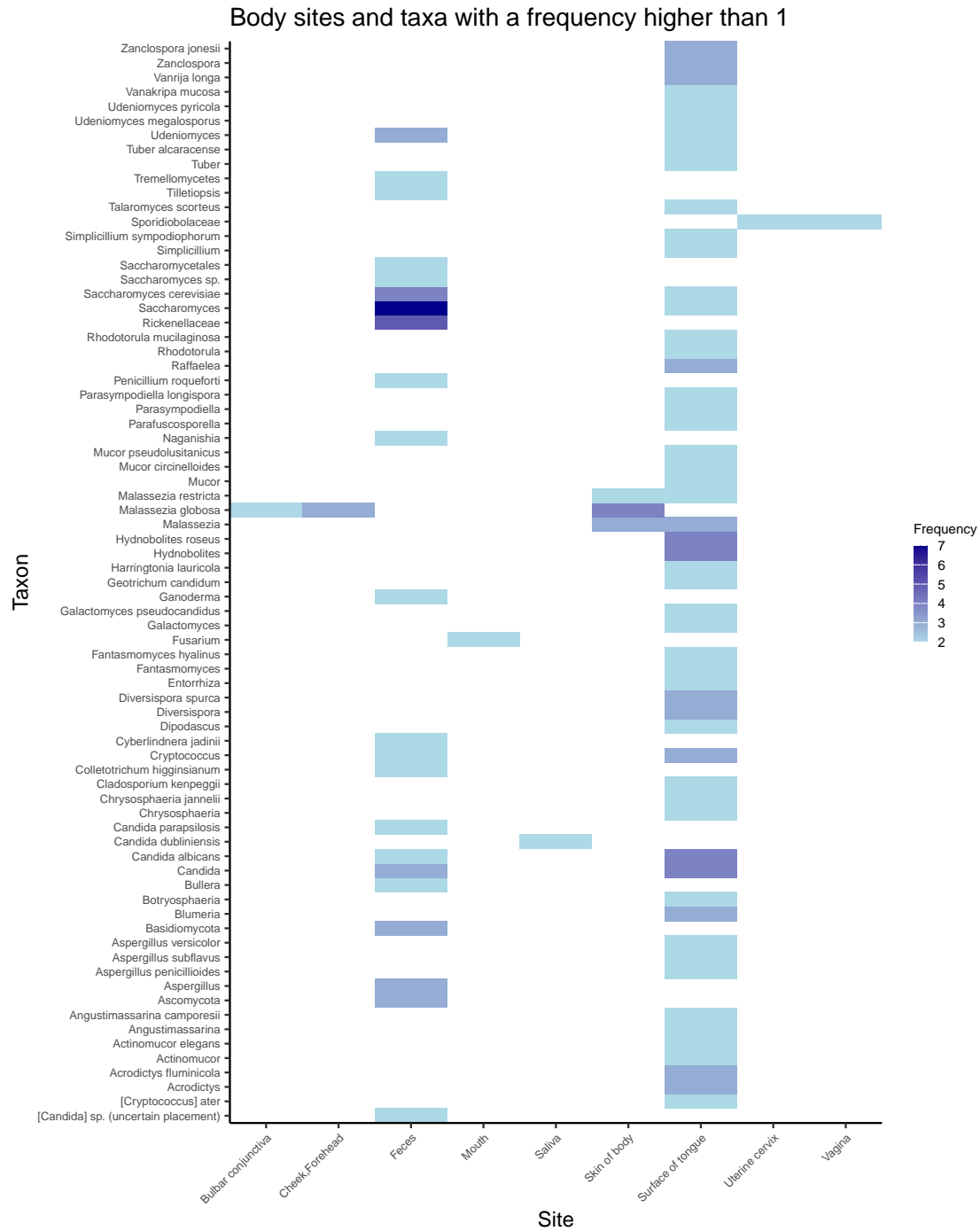
[1] "25 body sites were sampled in the fungi experiments."



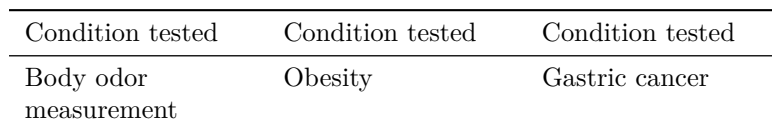
Significant fungal species often found in the human intestine (From Pérez, 2021).

Name	Observations/Comments	In BSDB
<i>Candida albicans</i>	Most prevalent fungus in the gastrointestinal tract of human adults. Major inducers of Th17 cells.	Is among top 20 frequent fungi species, not frequently repeated in body sites.
<i>Candida glabrata</i>	Opportunistic pathogen. Unknown role(s) in the gut.	Listed in NCBI as <i>Nakaseomyces glabratus</i> . Not found in BSDB
<i>Candida parapsilosis</i>	Opportunistic pathogen. Unknown role(s) in the gut.	Is among top 20 frequent fungi species, not frequently repeated in body sites.
<i>Rhodotorula mucilaginosa</i>	Higher abundance in neonates associated with childhood atopy (Fujimura et al., 2016).	Found in 1 study.
<i>Issatchenkia orientalis</i> (Candida krusei or Pichia kudriavzevii)	Increased abundance in infants associated with atopic wheeze (Arrieta et al., 2018).	<i>Pichia kudriavzevii</i> in NCBI. Not found in BSDB
<i>Malassezia restricta</i>	While often found in stool samples, this skin fungus may not be a true symbiont of the human gut (Fiers et al., 2019).	Frequent in BSDB, found only in feces in decreasing signatures.
<i>Saccharomyces</i> spp.	While they are often found in stool samples (likely because they are components of many beverages and foods), these yeasts are not considered natural residents of the human gut.	Frequent in BSDB, found in feces.

Fungi taxa frequency by body site



What conditions were tested for in the experiments that include fungi?



Asthma	Acute myeloid leukemia	Antimicrobial agent
Air pollution	Human papilloma virus infection,Cervical glandular intraepithelial neoplasia	Cervical glandular intraepithelial neoplasia,Human papilloma virus infection
Human papilloma virus infection	Type II diabetes mellitus	Type I diabetes mellitus
Diabetes mellitus	COVID-19	Parkinson's disease
High fat diet	Health study participation	HIV infection
Ethnic group	Age	Crohn's disease
Colorectal cancer	Root	Antimicrobial agent,Amoxicillin
Biological sex	Dry eye syndrome	Acne
Urinary tract infection	Allergic asthma	Behcet's syndrome
Gut microbiome measurement	Microbiome measurement	Polycystic ovary syndrome
Otitis media,Otitis externa	Otitis externa,Otitis media	Hypertension
Response to risperidone	Acute pancreatitis	Lower respiratory tract disease

What can presence/absence tell us about diversity?

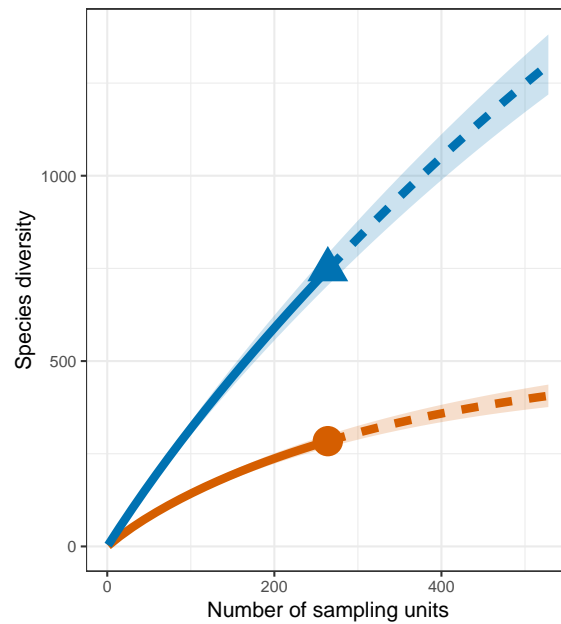
Diversity in Fungi taxa vs Other taxa

```
# reorganize for iNext
incidence <- freqKingdom %>%
  add_count(Kingdom, name = "Samples") %>%
  dplyr::select(Kingdom, Samples, Frequency, Taxon, everything())

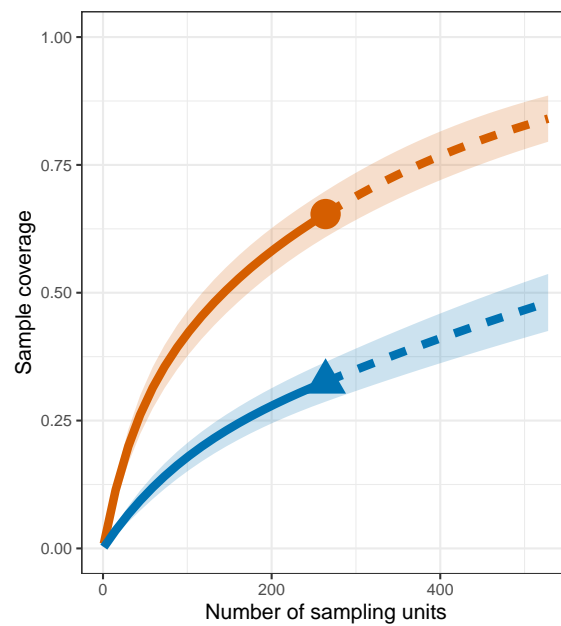
incidence <- reshape2::dcast(incidence, formula =
  Taxon~Kingdom, value.var = "Frequency", fill = 0)
## convert to list
incidence_list<-as.list(incidence)
# Remove zeros from each element of the list
incidence <- lapply(incidence_list, function(x) {
  x[x > 0]})
incidence<- incidence[-1]
incidence$Fungi<-c(264, incidence$Fungi)
incidence$Other<-c(264, incidence$Other)

library(iNEXT)
out.inc <- iNEXT(incidence, q=0, datatype="incidence_freq")
```

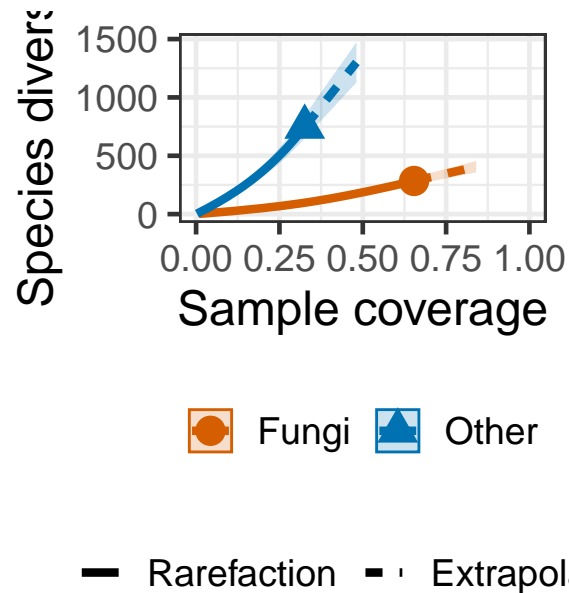
```
ggiNEXT(out.inc, type=1, color.var="Assemblage") +  
theme_bw(base_size = 8) +  
theme(legend.position="None")
```



```
ggiNEXT(out.inc, type=2, color.var="Assemblage") +  
ylim(c(0,1)) +  
theme_bw(base_size = 8) +  
theme(legend.position="None")
```



```
ggiNEXT(out.inc, type=3, color.var = "Assemblage") +
xlim(c(0,1)) +
theme_bw(base_size = 18) +
theme(legend.position="bottom",
legend.title=element_blank(),
text=element_text(size=18),
legend.box = "vertical")
```



Conclusions

This short exploratory analysis suggests there is an under-representation of fungi compared to bacteria in BugSigDB. This could just be the reflection of an existing bias in microbiome research toward bacterial studies.

At least 20 studies require a second revision to ensure the sequencing method is correctly input.

A taxonomic exploration revealed some unexpected taxa, like the insecta class and Malpighiales order.

The most frequent fungal taxa belong to the *Malassezia* genus, and include the phylum Ascomycota and genus *Saccharomyces*. The most frequent bacteria is *Staphylococcus epidermis*.

The increasing/decreasing abundance patterns may suggest taxa that don't coexist or are antagonistic to each other. This analysis also highlighted there are studies that are missing this input.

The studies took samples from several body sites and tested for different conditions. The frequencies for certain taxa seem to point these may be more prevalent in some body part than in others.

The diversity analysis suggests that fungal diversity is poorly captured with current sampling efforts. However this analysis should be carried out with the same taxa level and not with a mix of taxa levels.