



Diversifying ~~NOAH~~ Supersequences with New HSQC-Based Modules

Jonathan R. J. Yong,¹ Ēriks Kupče,² Tim D. W. Claridge^{1,*}

¹ *Chemistry Research Laboratory, Department of Chemistry, University of Oxford, Mansfield Road, Oxford, OX1 3TA, U.K.*

² *Bruker UK Ltd., Banner Lane, Coventry, CV4 9GH, U.K.*


* `tim.claridge@chem.ox.ac.uk`



Abstract

(Could we sell this a bit more...?) The sensitivity-enhanced HSQC, as well as HSQC-TOCSY, experiments can be incorporated into NOAH (NMR by Ordered Acquisition using ¹H detection) supersequences. Importantly, these heteronuclear modules preserve the magnetisation required for subsequent acquisition of other homonuclear modules in the supersequence. With these new modules, we reach a total of over 600 practically applicable NOAH supersequences which yield high-quality 2D spectra with greatly reduced experiment durations.

Introduction

In recent years, there has been significant interest in techniques which accelerate the acquisition of NMR data, especially for multidimensional spectra.^{1–3} One of the most versatile approaches is to utilise different “pools” of magnetisation for the sequential collection of different spectra without an intervening recovery delay, as illustrated by the NOAH (NMR by Ordered Acquisition using ¹H detection) technique.  Virtually all of the most common 2D experiments, such as HMBC, HSQC, COSY, NOESY, and TOCSY, can be concatenated in a modular fashion to form NOAH supersequences, providing time savings of up to $\sim 4\times$ as compared to individual acquisition of each spectrum.

One-bond heteronuclear correlation experiments, namely HMQC and HSQC, play a central role in

the structural elucidation of small organic molecules and biomolecules. (cite) These experiments are also a core component of many NOAH experiments, since the magnetisation they use (protons directly coupled to dilute nuclei, i.e. ^{13}C and ^{15}N) can be efficiently differentiated from the “bulk” magnetisation of uncoupled protons.^{4d,5} At the same time, due to the low natural abundance of these heteronuclei, these spectra are typically less sensitive than the homonuclear spectra that follow. Consequently, for dilute samples, the minimum experimental time is generally dictated by these heteronuclear experiments. Any improvements in experiment sensitivity can be translated into greater time savings.

In the 1990s, Cavanagh, Rance, and Kay introduced the sensitivity-enhanced HSQC (seHSQC) experiment, which improves on the sensitivity of an ordinary echo-antiecho HSQC by up to a factor of 2.⁶ This is accomplished by converting magnetisation that is both cosine- and sine-modulated in t_1 to observable magnetisation prior to detection, in the so-called preservation of equal pathways (PEP) scheme. Here, we show how the original seHSQC sequence can be modified such that it can be used as a NOAH module. We also introduce a HSQC-TOCSY module, derived from the ASAP-HSQC-TOCSY,⁷ that is also compatible with the NOAH strategy. Both of these modules can be inserted either independently or together into NOAH supersequences, allowing large amounts of chemical information to be acquired in short times.

^{13}C seHSQC

NOAH supersequences, such as the MSCN experiment in Figure 1b, rely on the fact that the output of any one module contains all the necessary magnetisation components required for downstream modules. The standard NOAH HSQC module (Figure 1c), derived from the symmetrised ASAP-HSQC,⁹ obeys this principle: it returns the “bulk” magnetisation belonging to uncoupled protons back to its equilibrium position ($+z$). Introducing the sensitivity enhancement scheme, however, requires some extra care. Using the original Cavanagh–Rance–Kay (CRK) seHSQC (Figure 1d) as part of a seHSQC/COSY NOAH supersequence, with the delay Δ' set to $1/(8 \cdot J_{\text{CH}})$, affords sensitivity gains which are most significant for CH peaks.¹⁰ However, the CRK seHSQC also causes

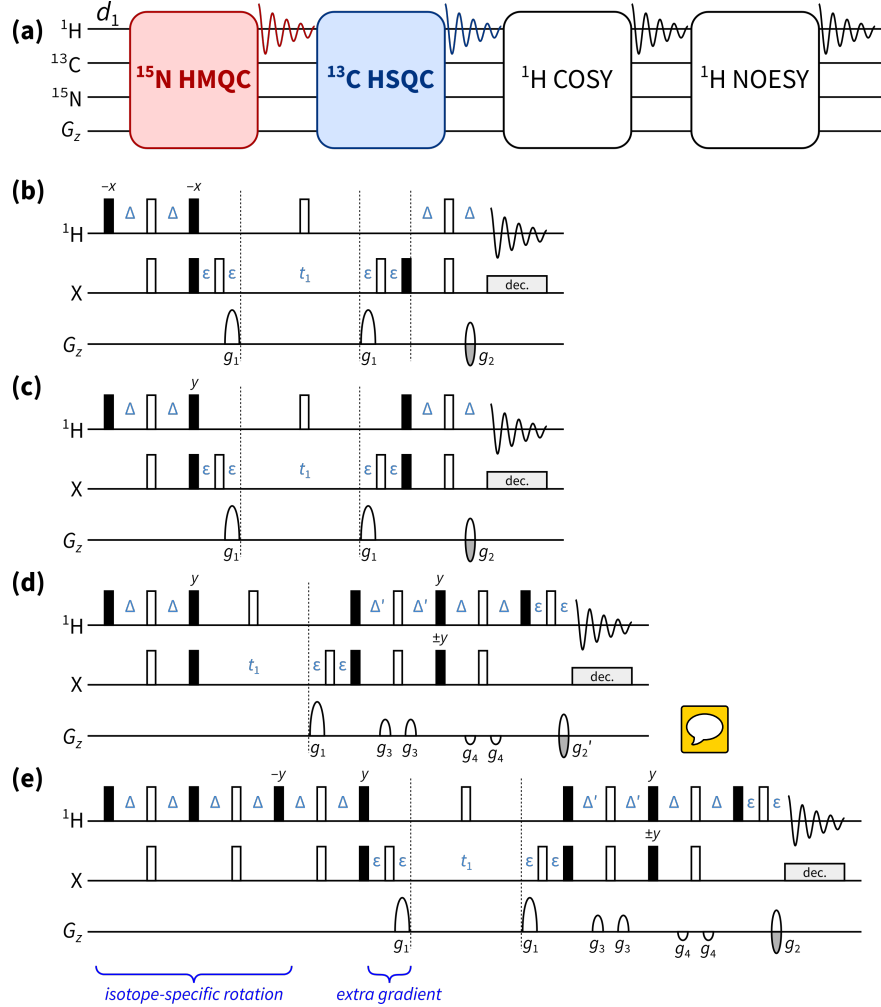


Figure 1: (a) Overview of a typical NOAH supersequence (MSCN, using the single-letter abbreviations previously defined^{4a}). The ^{15}N - ^1H HMQC and ^{13}C - ^1H HSQC modules are highlighted: these may be replaced with the new seHSQC module proposed in this work. (b) Original NOAH HMQC module,^{4a,8} abbreviated as “M”. (c) Original NOAH HSQC module without sensitivity enhancement,^{4a,9b} abbreviated as “S”. (d) Cavanagh-Rance-Kay (CRK) seHSQC.⁶ (e) NOAH seHSQC module, abbreviated as “Sp” (this work). Filled and unfilled bars represent 90° and 180° pulses respectively; all 180° pulses on ^{13}C are adiabatic (swept-frequency) pulses. All pulses are applied along $+x$ unless otherwise noted. The delays are chosen as follows: $\Delta = 1/(4 \cdot ^1J_{\text{XH}})$, $\Delta' = 1/(8 \cdot ^1J_{\text{CH}})$ or $1/(4 \cdot ^1J_{\text{NH}})$, and ε is the minimum time needed for a gradient and subsequent recovery. All gradients are 1 ms long, except for g_1 and g_2 in ^{15}N experiments which are 2.5 ms long. Gradient amplitudes, as percentages of maximum gradient strength, are as follows: $g_1 = 80\%$; $g_2 = \pm 40.2\%$ (^{13}C) or $\pm 16.2\%$ (^{15}N); $g_2' = g_2/2$; $g_3 = 11\%$; $g_4 = -5\%$. The signs of g_2 and g_2' are alternated in each t_1 increment to provide echo-antiecho selection.

bulk magnetisation to be dephased by coherence transfer pathway (CTP) gradients. Consequently, the downstream COSY can only utilise any bulk magnetisation that relaxes during the HSQC FID acquisition, leading to drastic intensity losses (Figure 2a).

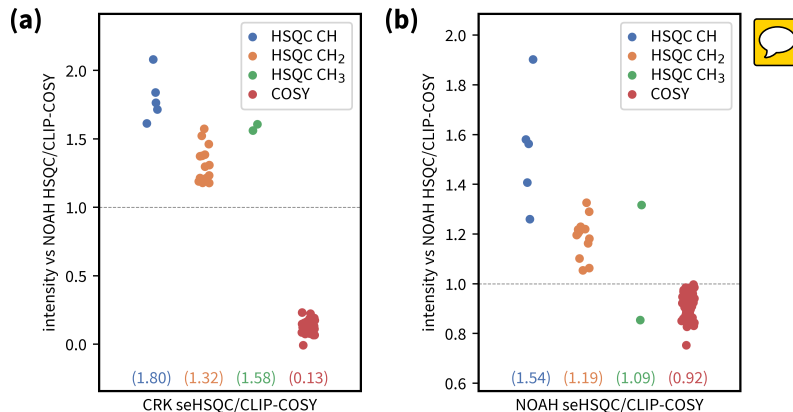


Figure 2: Sensitivity comparisons for seHSQC/CLIP-COSY¹¹ supersequences, using the CRK (Figure 1d) and NOAH (Figure 1e) seHSQC implementations. Spectra were obtained on a 700 MHz Bruker AV III equipped with a TCI H/C/N cryoprobe; the sample used was 40 mM andrographolide in DMSO-*d*₆. All intensities are normalised against a standard HSQC/CLIP-COSY supersequence (without sensitivity enhancement, i.e. Figure 1c). HSQC intensities are further split by multiplicity. Numbers in parentheses indicate averages over all peaks of a given type. (a) Using the original CRK seHSQC. The CRK seHSQC does not preserve the bulk magnetisation, leading to severely reduced COSY intensities. (b) Using the NOAH seHSQC.

Our solution is based on the simple observation that the bulk magnetisation in the seHSQC will be returned to $+z$ if the phase of the initial ^1H 90° pulse is changed to $+y$. To generate the HSQC signal, however, the same pulse needs to be applied along $+x$. Overall, what we need is therefore a pulse sequence element which simultaneously acts as a 90°_x pulse on protons coupled to ^{13}C , and as a 90°_y pulse on uncoupled protons. We accomplish this by prepending two spin-echoes, each of duration $2\Delta = 1/(2 \cdot ^1J_{\text{CH}})$, to the pulse sequence. We refer to this element as an *isotope-specific rotation* (ISR). It is similar to the zz -filter used previously in the NOAH HMBC module^{4b,4d} but has different pulse phases and thus leads to a different overall outcome. The BIG-BIRD element developed by Briand and Sørensen is also capable of effecting this;¹² however, we find that the ISR provides better performance (SI).

Apart from the ISR, the NOAH seHSQC module also contains an additional CTP gradient prior to the t_1 period (highlighted in Figure 1e). This gradient is not necessary for the seHSQC module itself. Instead, its purpose is to suppress artefacts in downstream modules, which arise from bulk magnetisation that evolves during either half of the HSQC t_1 period. This then evolves again in the t_1 period of another homonuclear module (e.g. COSY). Therefore, each COSY peak with $\Omega_1 = \Omega_{\text{H}}$

is accompanied by a pair of “wing” artefacts at $\Omega_1 = \Omega_H \pm (\Omega_H \cdot SW_C)/(2 \cdot SW_H)$, which can reach $\sim 5\%$ of the intensity of their parent peaks. Importantly, the artefacts arising from diagonal peaks can have intensities that are comparable to genuine crosspeaks (SI), which highlights the importance of suppressing these artefacts.

With these modifications, the NOAH seHSQC module provides clear sensitivity gains over the NOAH HSQC module, while preserving essentially the same amount of magnetisation for downstream modules (Figure 2b). The modifications present in the NOAH seHSQC, particularly the ISR, mean that the sensitivity improvements are slightly lower as compared to the original CRK implementation. On average, CH and CH₂ peaks have 54% and 19% increased sensitivity respectively. However, a dramatic improvement is seen in the COSY module which follows. In contrast to the CRK seHSQC, which completely destroys the requisite bulk magnetisation, the NOAH seHSQC preserves the majority of it, performing 92% as well as the original HSQC module.

Multiplicity editing can be easily incorporated into the NOAH seHSQC sequence (SI), leading to similar sensitivity gains relative to the HSQC module. It is noteworthy that the original HSQC places the bulk magnetisation in the xy -plane during the t_1 and editing periods, whereas the newly proposed NOAH seHSQC places the bulk along $\pm z$. In the former, the bulk magnetisation is therefore subject to homonuclear coupling (H_{HH}) evolution, leading to a decrease in downstream sensitivity when multiplicity editing is introduced. However, there is no such penalty in the NOAH seHSQC. In fact, the edited NOAH seHSQC slightly outperforms the edited HSQC in terms of preserving bulk magnetisation (SI).

Assorted thoughts on this section:

1. Internally we’ve called it the zz -filter, but I’m hesitant to do this in the paper, as it is not really the same thing as what we have in the HMBC (the pulse phases are different). The original zz -filter keeps ^{13}C - ^1H along $+z$ and excites the bulk. This one excites both but with different phases.
2. Although we don’t have the space to go into detail, we could have a very short paragraph mentioning that the sequence can be used in an ASAP context, and that it provides S/N



improvement over original ASAP-HSQC. Would that be useful? I don't think it's publishable on its own, unless we were to go into great depth looking at factors such as effect of relaxation vs isotropic mixing.

^{15}N seHSQC

The proposed seHSQC module can be further extended to ^{15}N experiments. Currently, in NOAH supersequences, ^{15}N - ^1H correlations are primarily obtained using the HMQC module.^{4a,8} Compared to this, the new seHSQC module can provide up to $5\times$ greater sensitivity (Figure 3). This arises partly because of the use of the PEP sensitivity enhancement scheme, in which the delays Δ' can be set to $1/(4 - J_{\text{HH}})$ (i.e. optimised for NH peaks). However, there is also a significant improvement due to the fact that peaks in the seHSQC are not broadened in the indirect dimension by J_{HH} , unlike in the HMQC.

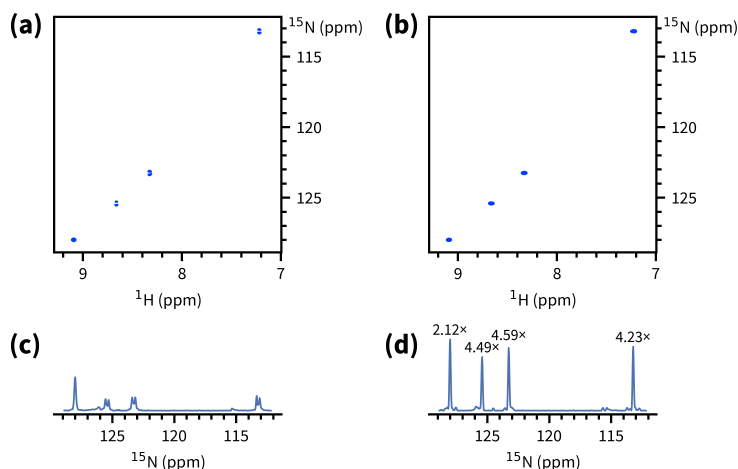


Figure 3: Comparison of the new ^{15}N - ^1H seHSQC with the standard NOAH HMQC module. Spectra were obtained on a 700 MHz Bruker AV III equipped with a TCI H/C/N cryoprobe; the sample used was 40 mM gramicidin in DMSO- d_6 . (a) HMQC spectrum. (b) seHSQC spectrum. (c) Projection of HMQC onto the f_1 axis. Splitting due to J_{HH} is clearly visible for three of the four peaks. (d) Projection of seHSQC onto the f_1 axis. Signal-to-noise improvements relative to the HMQC spectrum are indicated over each peak. The largest gains are observed for peaks where the multiplet structure is collapsed; however, even in the absence of that, a $\sim 2\times$ gain is still obtained.


While a ^{15}N HSQC module (without sensitivity enhancement) would still reap the latter benefit,

it comes with other severe drawbacks. As previously discussed, the HSQC module places bulk magnetisation in the xy -plane during the t_1 period. Consequently, ~~due to J_{HH} evolution~~, the amount of bulk magnetisation that is retained decreases as t_1 is lengthened, leading to line broadening in the indirect dimensions of all downstream modules (SI). This is not a problem with the ^{13}C HSQC, since typical ^{13}C indirect dimension acquisition times are relatively short. ~~However, with the smaller spectral widths in ^{15}N experiments, downstream modules can suffer substantial losses in sensitivity and resolution.~~


The ^{15}N seHSQC module ~~has one major~~ change with respect to its ^{13}C counterpart, which is that the CTP gradients g_1 and g_2 (Figure 1e) are all lengthened to 2.5 ms. This is to effectively dephase any bulk magnetisation that is transverse just prior to detection, which can give rise to significant levels of artefacts in the seHSQC module itself. The ^{13}C seHSQC does not need this because of the larger amplitude of g_2 ; however, the corresponding ^{15}N gradient is weaker by a factor of $\gamma_{\text{C}}/\gamma_{\text{N}} \approx 2.5$, thus requiring a longer duration in order to produce the same attenuation. In practice, we find that gradient durations of 2 to 2.5 ms provide good artefact suppression. ~~These lengthened gradients do not cause any appreciable difference in the intensity of actual signals.~~ (SI).

In scenarios where high resolution in the ^{15}N dimension is not required, it can prove useful to reduce the number of t_1 increments and in its place increase the number of transients acquired.^{2b,13} In new versions of the NOAH pulse programmes (including those provided in the *Supporting Information*), this feature can be enabled by specifying a factor k by which to perform this scaling. Note that the scaling is only applied to the ^{15}N module; all other modules are left untouched. In our hands, setting $k = 2$ or 4 for the ^{15}N HMQC can lead to significant sensitivity gains (how much?), since J_{HH} splitting in the indirect dimension can no longer be resolved (SI). This point is not relevant to the seHSQC, and here k -scaling on its own has only a tiny effect on peak height; any sensitivity gained from the extra transients is offset by the broadening. However, the later t_1 increments which were not acquired can be reconstructed using linear projection (cite). The resulting spectra display sensitivity gains of up to a factor of k , although the fidelity of the reconstruction suffers for $k > 4$ (SI).

Assorted thoughts:

1. The S/N gains from k -scaling + LP have only been demonstrated on a high S/N sample. In such cases it's possible to argue that the extra S/N is not even necessary. Can we make the same claims for low S/N samples where the reconstruction is likely to not be so easy?
2. In fact, we could probably encode ^{15}N -only NUS in the pulse programme, using a different acquisition flag (let's say `-DNUS_NONLY`). Similar to k -scaling  we would still only sample 25% (for example) of points in the ^{15}N dimension 4 times as often. However, the sampling scheme would be different. I have one dataset in which I compared the two approaches (there, I constructed the NUS pulse programme manually). The naive k -scaling had marginally better S/N (probably because it samples earlier points, which have larger signal) and the reconstructions were comparable in quality. However, again, this wasn't a particularly challenging sample, and therefore arguably not the best to demonstrate the technique on. Do you think that there is anything serious to be gained through use of NUS, versus the existing k -scaling + LP? If yes, then we could consider doing more systematic investigations in future.

Double HSQC and HSQC-TOCSY

Next, we note that the HSQC module (though not the new seHSQC module) allows an arbitrary amount of ^{13}C - ^1H magnetisation to be excited, with the remainder returned to $+z$. In order to excite a proportion f of ^{13}C - ^1H magnetisation ($0 < f \leq 1$), the initial INEPT delay must be shortened by a factor of $\sin^{-1} f$. The remaining $(1 - f)$ of the magnetisation, plus any that recovers during the first HSQC FID, can then be used for a second HSQC module in the same supersequence. The collection of multiple HSQC spectra  in one multi-FID acquisition (MFA) experiment has previously been accomplished by **separating** the two CTPs in the CRK seHSQC, with the cosine- and sine-modulated CTPs each contributing to one spectrum.¹⁴ However, with a value of $f = 0.7$, the NOAH strategy already provides slightly higher sensitivity for both spectra. Furthermore, the sensitivity of the second HSQC can be further boosted by using the new seHSQC module in place of it **(SI)**.

By adding a period of isotropic mixing prior to detection, the NOAH HSQC module may be converted to a HSQC-TOCSY module. This is similar to the previously reported ASAP-HSQC-TOCSY,⁷ the key difference being that in the present NOAH context, unused ^{13}C - ^1H and bulk magnetisation is preserved for use in other modules, instead of later t_1 increments as in the ASAP experiment. Compared to the existing MFA HSQC-TOCSY/HSQC experiment,^{14a} our approach displays greater flexibility in three regards. Firstly, the vast majority of bulk magnetisation is preserved, allowing for homonuclear module(s) to be appended in a NOAH supersequence (in practice, losses of ca. 10% are observed due to pulse imperfections). ~~On the other hand,~~ the MFA sequence, much like the original CRK seHSQC which it is based on, dephases bulk magnetisation and causes a 80–90% sensitivity loss in downstream spectra. Secondly, the sensitivity of both spectra can be optimised through the value of f ; this allows a larger amount of ^{13}C - ^1H magnetisation to be used for the inherently less sensitive HSQC-TOCSY (in our experience, setting $f = 0.9$ provides a good balance). ~~In contrast, isotropic mixing in the MFA sequence is applied to the less sensitive sine-modulated component, leading to spectra with imbalanced sensitivity.~~ Lastly, since each NOAH module is independently executed, the NOAH approach allows multiplicity editing to be enabled for only the HSQC and not the HSQC-TOCSY, where accidental overlap may lead to crosspeaks being lost unexpectedly.

~~Despite these benefits, it should be noted that~~ the NOAH HSQC-TOCSY module will still have lower overall sensitivity than a conventional seHSQC-TOCSY, which can make use of the PEP scheme. ~~It is not possible to simply insert a TOCSY mixing block into the seHSQC module presented here, as that will lead to the bulk magnetisation being dephased.~~ As usual, the benefits

of fast acquisition schemes such as NOAH are most obviously realised in sufficiently concentrated samples, where NMR data acquisition times are chiefly limited by the requisite number of t_1 increments. In such settings, the same amount of data can be collected in much shorter times, without worrying about the slight loss in sensitivity inherent to all fast acquisition schemes. On the other hand, for dilute samples where this sensitivity loss is less easily tolerated, the *sensitivity per unit time* of each supersequence must then be taken into consideration. As long as the time savings outweigh any sensitivity losses, use of the NOAH supersequence will then prove to be a net benefit.

Systematic investigations in this area will be detailed elsewhere.

Assorted thoughts:

1. Do we need a figure illustrating the reduced delay and the HSQC-TOCSY pulse sequence?
Should it be merged with the existing Fig 1, or Fig 4?
2. I hope I have not been too critical of Parella's work. Everything I wrote *is* true, and I want to say something positive about our work, but when that's the only real basis for comparison I feel like I might be being a bit harsh!

Example spectra and conclusion

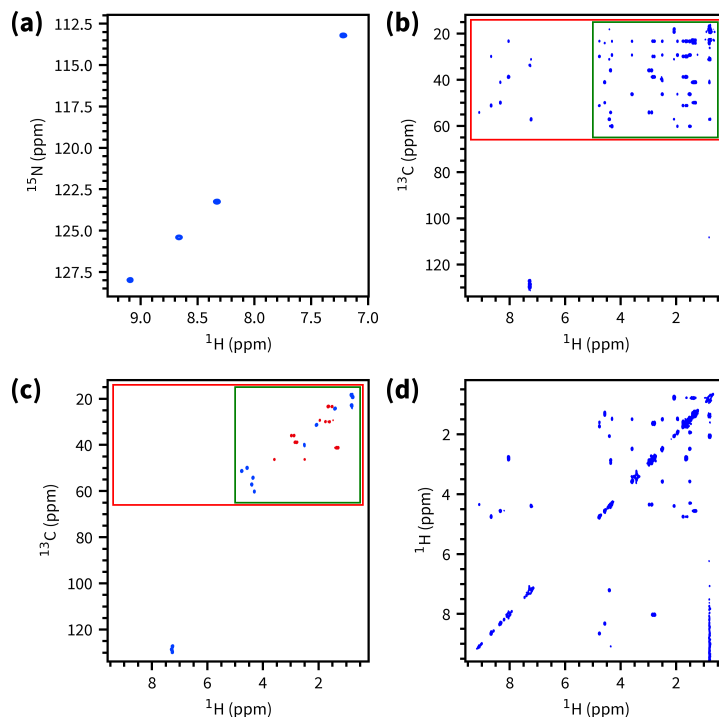




Figure 4: Example spectra obtained from the NOAH-4 SpStSpCc supersequence. Spectra were obtained on a 700 MHz Bruker AV III equipped with a TCI H/C/N cryoprobe; the sample used was 40 mM gramicidin in DMSO- d_6 . 256 t_1 increments were used, with 2 scans per increment. The total experiment time was 17 minutes and 35 seconds. (a) ^{15}N seHSQC. (b) ^{13}C HSQC-TOCSY ($f = 0.9$). (c) Multiplicity-edited ^{13}C seHSQC. Notice that having the edited seHSQC removes the need for the less desirable HSQC-TOCSY editing. (d) CLIP-COSY. Should we zoom in on the interesting bits for (b) and (c) – either the red or the green box? (Although I would prefer that we use the same box for both, i.e. not the red box for (b) and green box for (c).) Should we perform magnetisation on (d) (I notice that this was done for the ACIE paper)? I already hid negative contours for (b) and (d)!

The NOAH-4 SpStSpCc (^{15}N seHSQC, ^{13}C HSQC-TOCSY, ^{13}C seHSQC, and CLIP-COSY) supersequence is one of many ways in which the new modules discussed above can be included in practical experiments. The spectra thus obtained are shown in Figure 4. While individual collection of the four spectra above would require 57 minutes and 8 seconds, the NOAH-4 supersequence takes only 17 minutes and 35 seconds, which represents a $3.25\times$ speedup. One can also prepend the NOAH HMBC module;^{4d} this uses the semi-adiabatic zz -filter to preserve magnetisation of protons directly coupled to ^{13}C and ^{15}N heteronuclei, which is precisely the magnetisation required by the

HSQC-based modules presented here.  Examples of such spectra, with time savings of up to $3.8\times$, are shown in the *Supporting Information* (SI).

 The new seHSQC and HSQC-TOCSY implementations add to the preexisting diversity in NOAH modules, bringing the total number of plausible NOAH supersequences to over 600. The AU scripts needed for processing of these modules, as well as a number of the more commonly used pulse sequences, are provided in the *Supporting Information*; others are available upon request from the authors. However, a more user-friendly and customisable method for the generation of NOAH pulse sequences is clearly needed to handle the sheer variety currently available. Our work towards this will be reported in the near future.

Final assorted thoughts:

1. We should probably provide “new” versions of pulse programmes here. The only thing that is backwards-*incompatible* is the NUS implementation. Can we introduce that in the SI? [Perhaps just to avoid confusion, we should rename the new NUS script `noah_nus2.py`?]

Acknowledgements

J.R.J.Y. thanks the Clarendon Fund (University of Oxford) and the EPSRC Centre for Doctoral Training in Synthesis for Biology and Medicine (EP/L015838/1) for a studentship, generously supported by AstraZeneca, Diamond Light Source, Defence Science and Technology Laboratory, Evotec, GlaxoSmithKline, Janssen, Novartis, Pfizer, Syngenta, Takeda, UCB, and Vertex. **Any other acknowledgements?**

References

1. (a) Frydman, L.; Scherf, T.; Lupulescu, A. *Proc. Natl. Acad. Sci. U. S. A.* **2002**, *99*, 15858–15862; (b) Frydman, L.; Lupulescu, A.; Scherf, T. *J. Am. Chem. Soc.* **2003**, *125*, 9204–9217.
2. (a) Nolis, P.; Pérez-Trujillo, M.; Parella, T. *Angew. Chem. Int. Ed.* **2007**, *46*, 7495–7497; (b) Parella, T.; Nolis, P. *Concepts Magn. Reson.* **2010**, *36A*, 1–23.

3. (a) Kupče, Ě.; Freeman, R.; John, B. K. *J. Am. Chem. Soc.* **2006**, *128*, 9606–9607; (b) Kovacs, H.; Kupče, Ě. *Magn. Reson. Chem.* **2016**, *54*, 544–560.
4. (a) Kupče, Ě.; Claridge, T. D. W. *Angew. Chem. Int. Ed.* **2017**, *56*, 11779–11783; (b) Kupče, Ě.; Claridge, T. D. W. *Chem. Commun.* **2018**, *54*, 7139–7142; (c) Claridge, T. D. W.; Mayzel, M.; Kupče, Ě. *Magn. Reson. Chem.* **2019**, *57*, 946–952; (d) Kupče, Ě.; Claridge, T. D. W. *J. Magn. Reson.* **2019**, *307*, 106568.
5. Garbow, J. R.; Weitekamp, D. P.; Pines, A. *Chem. Phys. Lett.* **1982**, *93*, 504–509.
6. (a) Palmer, A. G.; Cavanagh, J.; Wright, P. E.; Rance, M. *J. Magn. Reson.* **1991**, *93*, 151–170; (b) Kay, L.; Keifer, P.; Saarinen, T. *J. Am. Chem. Soc.* **1992**, *114*, 10663–10665; (c) Cavanagh, J.; Rance, M. *Annu. Rep. NMR Spectrosc.* **1993**, *27*, 1–58.
7. Becker, J.; Koos, M. R. M.; Schulze-Sünninghausen, D.; Luy, B. *J. Magn. Reson.* **2019**, *300*, 76–83.
8. Kupče, Ě.; Freeman, R. *Magn. Reson. Chem.* **2007**, *45*, 2–4.
9. (a) Schulze-Sünninghausen, D.; Becker, J.; Luy, B. *J. Am. Chem. Soc.* **2014**, *136*, 1242–1245; (b) Schulze-Sünninghausen, D.; Becker, J.; Koos, M. R. M.; Luy, B. *J. Magn. Reson.* **2017**, *281*, 151–161.
10. (a) Schleucher, J.; Schwendinger, M.; Sattler, M.; Schmidt, P.; Schedletzky, O.; Glaser, S. J.; Sørensen, O. W.; Griesinger, C. *J. Biomol. NMR* **1994**, *4*, 301–306; (b) Kontaxis, G.; Stonehouse, J.; Laue, E. D.; Keeler, J. *J. Magn. Reson.* **1994**, *111*, 70–76.
11. Koos, M. R. M.; Kummerlöwe, G.; Kaltschnee, L.; Thiele, C. M.; Luy, B. *Angew. Chem. Int. Ed.* **2016**, *55*, 7655–7659.
12. Briand, J.; Sørensen, O. W. *J. Magn. Reson.* **1997**, *125*, 202–206.
13. Pérez-Trujillo, M.; Nolis, P.; Bermel, W.; Parella, T. *Magn. Reson. Chem.* **2007**, *45*, 325–329.
14. (a) Nolis, P.; Motiram-Corral, K.; Pérez-Trujillo, M.; Parella, T. *ChemPhysChem* **2019**, *20*, 356–360; (b) Nolis, P.; Motiram-Corral, K.; Pérez-Trujillo, M.; Parella, T. *J. Magn. Reson.* **2019**, *300*, 1–7.

Supporting Information

for

Diversifying NOAH Supersequences with New
HSQC-based Modules

Jonathan R. J. Yong,¹ Ēriks Kupĉe,² Tim D. W. Claridge^{1,*}

¹ *Chemistry Research Laboratory, Department of Chemistry, University of Oxford,
Mansfield Road, Oxford, OX1 3TA, U.K.*

² *Bruker UK Ltd., Banner Lane, Coventry, CV4 9GH, U.K.*

* `tim.claridge@chem.ox.ac.uk`

Contents

1	Comparison of BIG-BIRD and ISR elements	3
2	Origin and suppression of “wing” artefacts	4
3	Multiplicity editing in seHSQC	5
4	^{15}N HSQC and line broadening	6
5	Effect of lengthened gradients in ^{15}N experiments	7
6	Effect of k -scaling	8
7	HSQC-TOCSY SNR comparisons	9
8	Other example spectra	10
9	Pulse programmes	11
10	Processing scripts	12

1 Comparison of BIG-BIRD and ISR elements

Just pure S/N comparisons. 

2 Origin and suppression of “wing” artefacts

Gramicidin SpSpCc experiments

3 Multiplicity editing in seHSQC

Include S/N comparisons

4 ^{15}N HSQC and line broadening

...

5 Effect of lengthened gradients in ^{15}N experiments

...

6 Effect of k -scaling

... both signal and artefact intensity, plus example spectra

7 HSQC-TOCSY SNR comparisons

... including Parella work

8 Other example spectra

...

9 Pulse programmes

...

10 Processing scripts

...