## **UR PSSM and Logo**

## Information Content and Sequence Logo Functions

The implementation below is based on the paper "Information Content of Binding Sites of Nucleotide Sequences".

We compute the information content (entropy) of each nucleotide position, i.e., how different they are from "random". For each nucleotide location along the motif i = 1, ..., n, let

$$R_i := H_g + \sum_{b=A}^{T} p_{b,i} \log 2(p_{b,i}),$$

denote the decrease in entropy from "random" ( $H_g := 2$ ) at location i, where  $p_{b,i}$  is the probability of base b at position i, which is estimated using the PPM entries (normalized nucleotide counts in each position). One can then compute the total decrease (total information gained) by  $R = \sum_{i=1}^n R_i$ . A sampling error correction can be applied to correct for biases that arises from using PPM and not the actual probabilities (see the Appendix in the paper).

## In [9]:

```
from Bio import motifs, SeqIO
from Bio.Seq import Seq
import numpy as np
import math
import matplotlib as mpl
from matplotlib.text import TextPath
from matplotlib.patches import PathPatch
from matplotlib.font_manager import FontProperties
import matplotlib.pyplot as plt
def compute pssm(lseq, pscount={'A':0.25,'C':0.25,'G':0.25,'T':0.25}, \
                 background={'A':0.25,'C':0.25,'G':0.25,'T':0.25}):
    '''Computes the PFM, PWM, and PSSM of a list of nucleotide sequences'''
   m = motifs.create(list(map(Seq, lseq)))
    pfm = m.counts
    # pwm with no pscount is a stochastic matrix
    pwm = m.counts.normalize(pseudocounts=pscount)
    pssm = pwm.log odds(background)
    return pfm, pwm, pssm, m, m.consensus
# The following code is based on:
# https://github.com/saketkc/motif-logos-matplotlib/blob/master/Sequence%20log
os%20in%20Python.ipynb
# However, the code there has few errors. The code below follows the paper "In
formation Content
# of Binding Sites of Nucleotide Sequences"
```

```
def calc_IC_approx_err(motif):
    '''Approximate calculate of small-sample correction error'''
    print('Computing approximate correction error...')
    bases = list(motif.pwm.keys())
    n = len(motif.counts[bases[0]]) # sequence length
    return (len(bases)-1)/(2 * n * np.log(2))
def calc IC exact err(motif):
    '''Exact computation of small-sample correction error'''
    \#\# O(n^3)
    print('Computing exact correction error...')
    pwm = motif.pwm
    bases = list(pwm.keys())
    n = na = len(motif.counts['A']) # sequence length
    nc = ng = nt = exact error = 0
    done = False
    while not done:
        #print (na,nc,ng,nt)
        pp = (0.25**na)*(0.25**nc)*(0.25**ng)*(0.25**nt)
        frac = pp*math.factorial(na+nc+ng+nt)/(math.factorial(na)*math.factori
al(nc)*
                                                 math.factorial(ng)*math.factori
al(nt))
        exact error += frac*sum([-p*np.nan to num(np.log2(p)) for p in \
                                  [na/n, nc/n, ng/n, nt/n]])
        if nt<=0:
            ## iterate inner loop
            if ng > 0:
                \#\# \ q => t
                ng = ng - 1
                nt = nt + 1
            elif nc > 0:
                \#\# \ c \ -> \ q
                nc = nc - 1;
                ng = ng + 1;
            else:
                ## a->c
                na = na - 1
                nc = nc + 1
        else:
            if ng > 0:
                \#\# \ q => t
                ng = ng - 1
                nt = nt + 1
            elif nc>0:
                \#\# c => g; all t -> g
                nc = nc - 1
                ng = nt + 1
                nt = 0
            elif na>0:
                ## a => c; all g,t -> c
                nc = nt + 1
                na = na - 1
                nt = 0
            else:
                done = True
    return exact error
```

```
def calc info content(motif, corr type = 'no'):
    '''Calculate information content with small sample correction.
   Note that for both corr_type=='approx' (should be used for
    sequences larger than 50 nt) and for corr type=='exact' (should
   be used for sequences smaller than 50 nt), the output can attain
   both negative and positive values. See the paper "Information Content
   of Binding Sites of Nucleotide Sequences". Thus, for sequence logos, use
    the default (i.e. corr type = 'no)
   pwm = motif.pwm # should not use relative information
   bases = list(pwm.keys())
   if corr type=='no':
       Hg = np.log2(len(bases))
   elif corr_type=='approx':
       Hg = np.log2(len(bases)) - calc IC approx err(motif)
   else: # exact
       Hg = calc IC exact err(motif)
    #print('Hg = {}'.format(Hg))
    return [Hg+sum([pwm[b][1]*np.nan_to_num(np.log2(pwm[b][1])) for b in bases
])\
            for 1 in range(0, len(motif))]
def calc rel info(motif, corr type = 'no'):
    '''Calculate relative information, i.e. the information content
    of each base along the sequence '''
   info cont = calc info content(motif, corr type)
   return {b: [np.nan_to_num(p*i) for p, i in zip(motif.pwm[b], info_cont)] \
            for b in list(pwm.keys())}
def gen nt sequence logo(rel info, fig size=(5,2)):
    '''Generates nucleotide sequence logo.
    rel info is computed by: rel info=calc rel info(motif, 'no')'''
    fp = FontProperties(family="Arial", weight="bold")
   globscale = 1.35 # this, and the values here below were set for family="A
rial"
   LETTERS = { T': TextPath((-0.305, 0), T', size=1, prop=fp),
                'G' : TextPath((-0.384, 0), "G", size=1, prop=fp),
                'A' : TextPath((-0.35, 0), "A", size=1, prop=fp),
                'C' : TextPath((-0.366, 0), "C", size=1, prop=fp) }
   COLOR SCHEME = {'G': 'orange',
                    'A': 'red',
                    'C': 'blue',
                    'T': 'darkgreen'}
   def letterAt(letter, x, y, yscale=1, ax=None):
        '''Plots a letter at a given position with a given scale.'''
       text = LETTERS[letter]
       t = mpl.transforms.Affine2D().scale(1*globscale, yscale*globscale) + \
            mpl.transforms.Affine2D().translate(x,y) + ax.transData
       p = PathPatch(text, lw=0, fc=COLOR_SCHEME[letter], transform=t)
        if ax != None: ax.add_artist(p)
        return p
    fig ax = nlt subplots(figsize=fig size)
```

```
maxi = 0
for i in range(0, len(list(rel_info.values())[0])):
    scores = [(b,rel_info[b][i]) for b in COLOR_SCHEME.keys()]
    scores.sort(key=lambda t: t[1])
    y = 0
    for base, score in scores:
        letterAt(base, x, y, score, ax)
        y += score
    x += 1
    maxi = max(maxi, y)
plt.xticks(range(1,x))
plt.xlim((0, x))
plt.ylim((0, maxi))
return fig, ax
```

## **An Example - UR PSSM**

```
In [23]:
```

```
import pandas as pd
import os
import numpy as np
from Bio import motifs, SeqIO
from Bio.Seq import Seq
import re
from collections import deque
from termcolor import colored
from pprint import pprint
mlen = 5
spec = 'dsDNA Bacteria'
top num = 12 #9
lfile = 'seq lfile.png'
dtype rnd = {'signf sorted':str, 'tot num Fall sort':int}
UR_rnd_xlsx_file = '/Users/yoramzarai/work/school/Simulation/Viruses/Data stat
s/UR rnd m'\
+str(mlen)+'_'+spec+'.xlsx'
df rnd = pd.read excel(UR rnd xlsx file, header=0, dtype=dtype rnd)
columns names = list(df rnd.columns)
# print('Found {} columns:'.format(len(columns names)))
# for i, s in enumerate(columns names, 1): print(i,s,sep='. ')
top UR = {df rnd.loc[s][columns names[0]] : df rnd.loc[s][columns names[1]] fo
r s in range(top num)}
eql nt = {s for s in top UR.keys() if len(set(s))==1}
diff_nt = set(top_UR.keys()) - eql_nt
print('eql:', eql nt)
print('diff:', diff_nt )
# compute motif count matrices
backg={'A':0.25,'C':0.25,'G':0.25,'T':0.25}
pfm, pwm, pssm, m, cons = compute pssm(diff nt, backg, backg)
#print(pfm, pwm, pssm, sep='\n')
# generate sequence logo
rel info = calc rel info(m, 'no')
fig, ax = gen nt sequence logo(rel info)
ax.set ylabel('Bits')
plt.axis([0.5, mlen+0.5, 0, ax.get_ylim()[1]])
plt.tight_layout()
if lfile!='':
   plt.savefig(lfile, dpi=200)
   print('Figure saved in {}'.format(lfile))
```

```
eql: {'AAAAA', 'CCCCC', 'TTTTT', 'GGGGG'}
diff: {'AATTT', 'GATCA', 'AGATC', 'TTTTC', 'GATCT', 'GGATC', 'AAAA
T', 'CCTGG'}
Figure saved in seq_lfile.png
```

/opt/local/Library/Frameworks/Python.framework/Versions/3.7/lib/py thon3.7/site-packages/ipykernel\_launcher.py:102: RuntimeWarning: d ivide by zero encountered in log2

