

# Class 9: Structural Bioinformatics

Youn Soo Na (PID: A17014731)

The main database for structural data is called the PDB (Protein Data Bank). Let's see what it contains!

Data from: <https://www.rcsb.org/stats> Or from alternate link: <https://tinyurl.com/pdbstats24>

```
pdb24 <- read.csv("pdb_stats.csv", row.names=1)
head(pdb24)
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other
Protein (only)	167,192	15,572	12,529	208	77	32
Protein/Oligosaccharide	9,639	2,635	34	8	2	0
Protein/NA	8,730	4,697	286	7	0	0
Nucleic acid (only)	2,869	137	1,507	14	3	1
Other	170	10	33	0	0	0
Oligosaccharide (only)	11	0	6	1	0	4
Total						
Protein (only)	195,610					
Protein/Oligosaccharide	12,318					
Protein/NA	13,720					
Nucleic acid (only)	4,531					
Other	213					
Oligosaccharide (only)	22					

```
# Some of the "numeric" values are actually characters
# We need to change them to numeric values.
# as.numeric( sub(",", "", pdb24$Total))
# I could run this into a function to fix the whole table or any future table
# I read like this:
# x <- pdb24$Total
# as.numeric( sub(",", "", x) )
```

```

comma2numeric <- function(x) {
  as.numeric( sub(",", "", x) )
}

# Test it.
# comma2numeric(pdb24$X.ray)
# head(pdb24)

pdb24test <- apply(pdb24, 2, comma2numeric)
head(pdb24test)

```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other	Total
[1,]	167192	15572	12529	208	77	32	195610
[2,]	9639	2635	34	8	2	0	12318
[3,]	8730	4697	286	7	0	0	13720
[4,]	2869	137	1507	14	3	1	4531
[5,]	170	10	33	0	0	0	213
[6,]	11	0	6	1	0	4	22

```

## try a different read/import function:
library(readr)
pdbdb <- read_csv("pdb_stats.csv")

```

Rows: 6 Columns: 8

-- Column specification -----

Delimiter: ","

chr (1): Molecular Type

dbl (3): Multiple methods, Neutron, Other

num (4): X-ray, EM, NMR, Total

i Use `spec()` to retrieve the full column specification for this data.

i Specify the column types or set `show\_col\_types = FALSE` to quiet this message.

```
sum(pdbdb$Total)
```

```
[1] 226414
```

And answer the following questions:

Q1. What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy?

```
(sum(pdbdb$`X-ray`)/sum(pdbdb$Total) * 100)+(sum(pdbdb$EM)/sum(pdbdb$Total) *100)
```

```
[1] 93.4845
```

Q2. What proportion of structures in the PDB are protein?

```
# library(dplyr)
# pdbdb %>%
#   filter(rowSums(sapply(., function(x) grepl("protein", x, ignore.case = TRUE))) > 0)
pdbdb$Total[1]/sum(pdbdb$Total) * 100
```

```
[1] 86.39483
```

Q3. Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

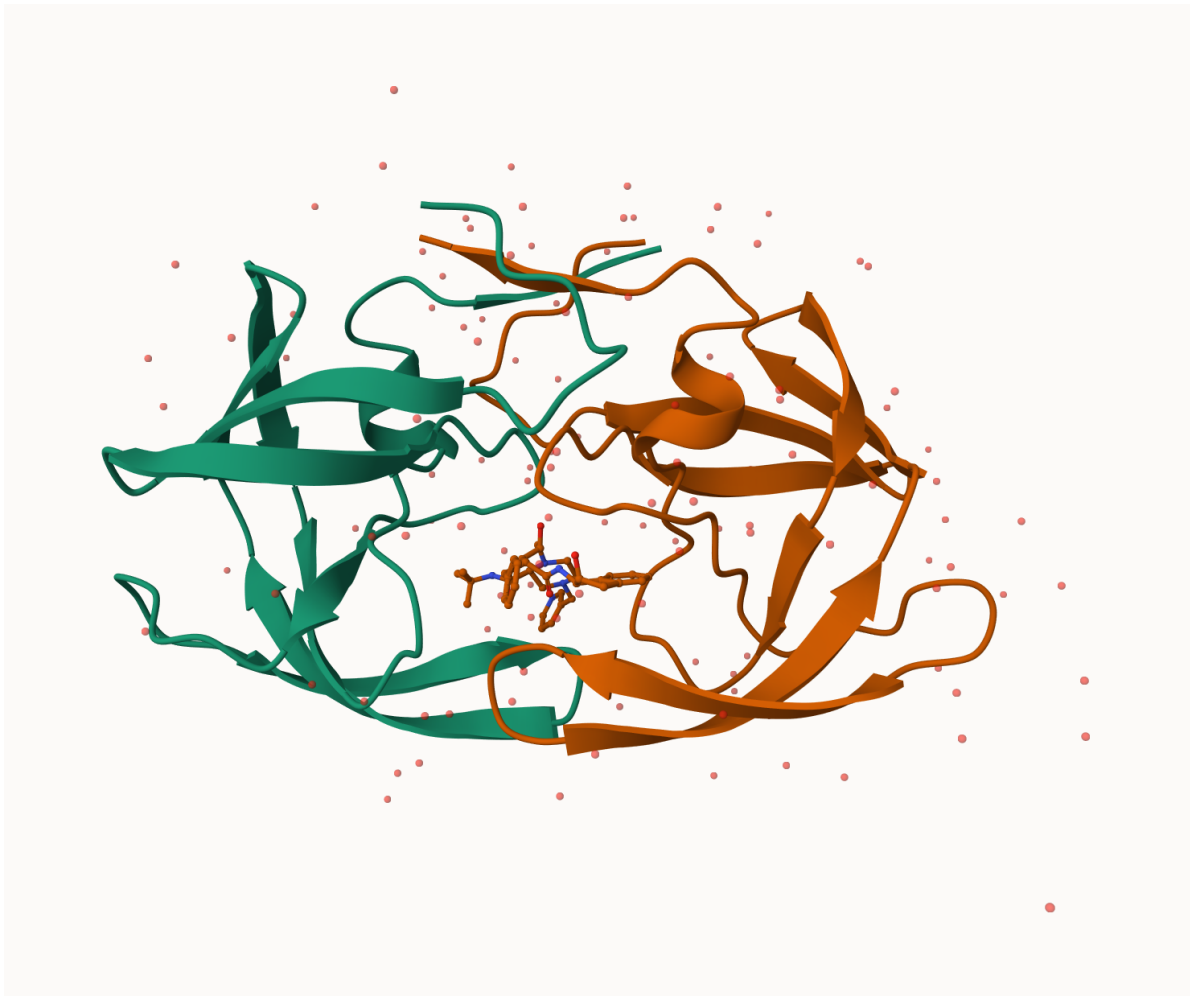
```
4553
```

## Mol\*

Mol\* (pronounced “molstar”) is a new web-based molecular viewer that we will need to learn the basics of here.

<https://molstar.org/viewer/>

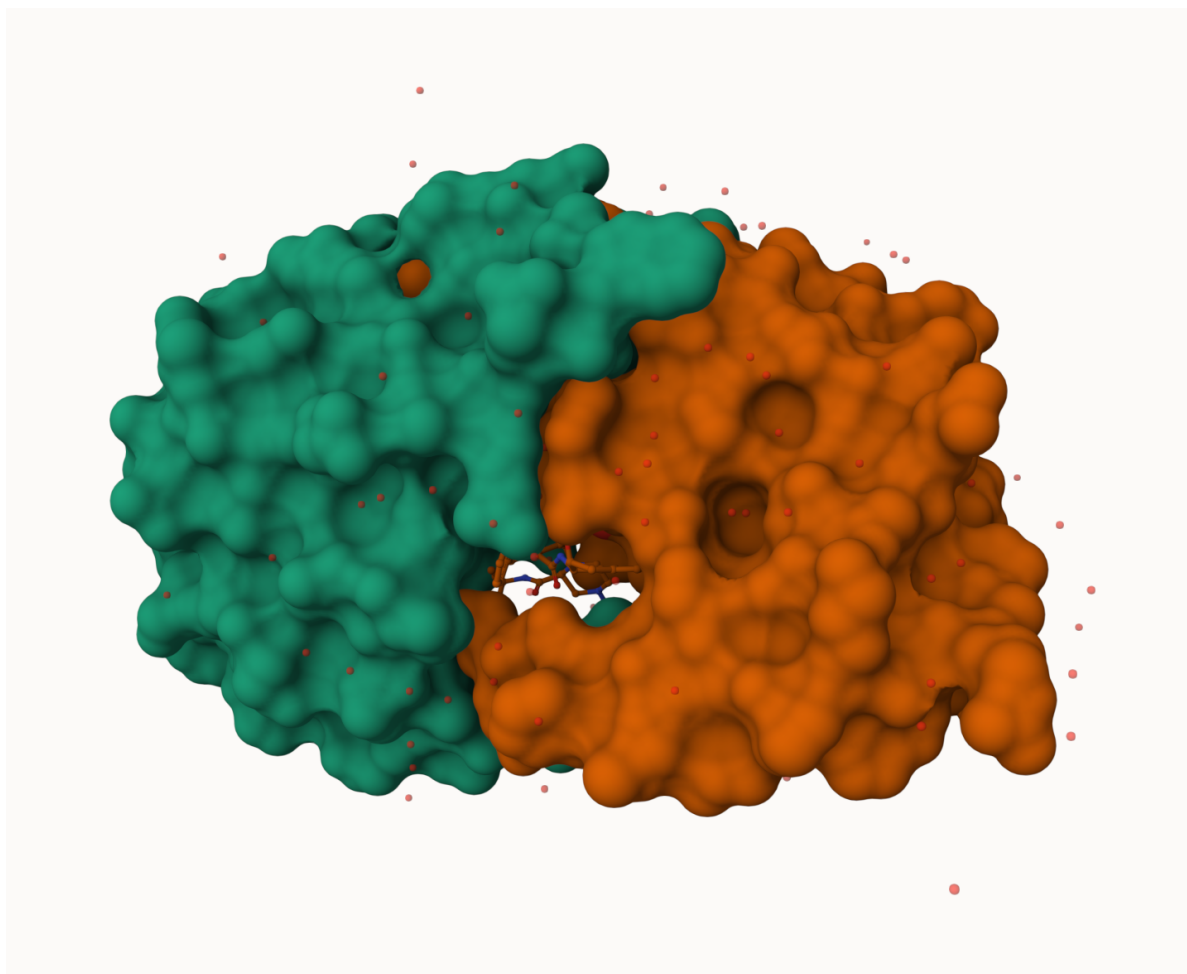
We will use PDB code: 1HSG

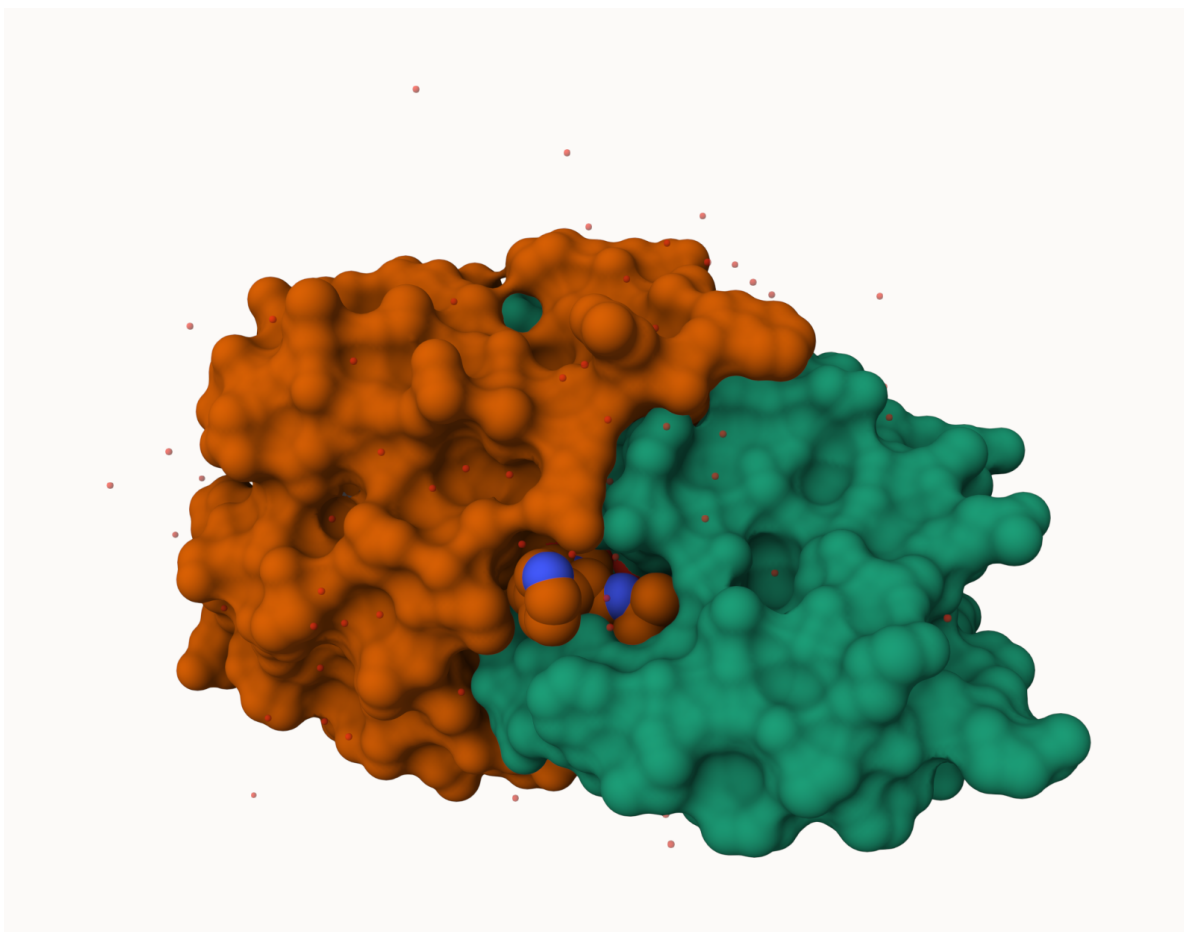


Note: This is an aspartic protease that uses 2 aspartic acid

Here are some more custome images:







Q4. Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

Q5. There is a critical “conserved” water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have?

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend “Ball & Stick” for these side-chains). Add this figure to your Quarto document.

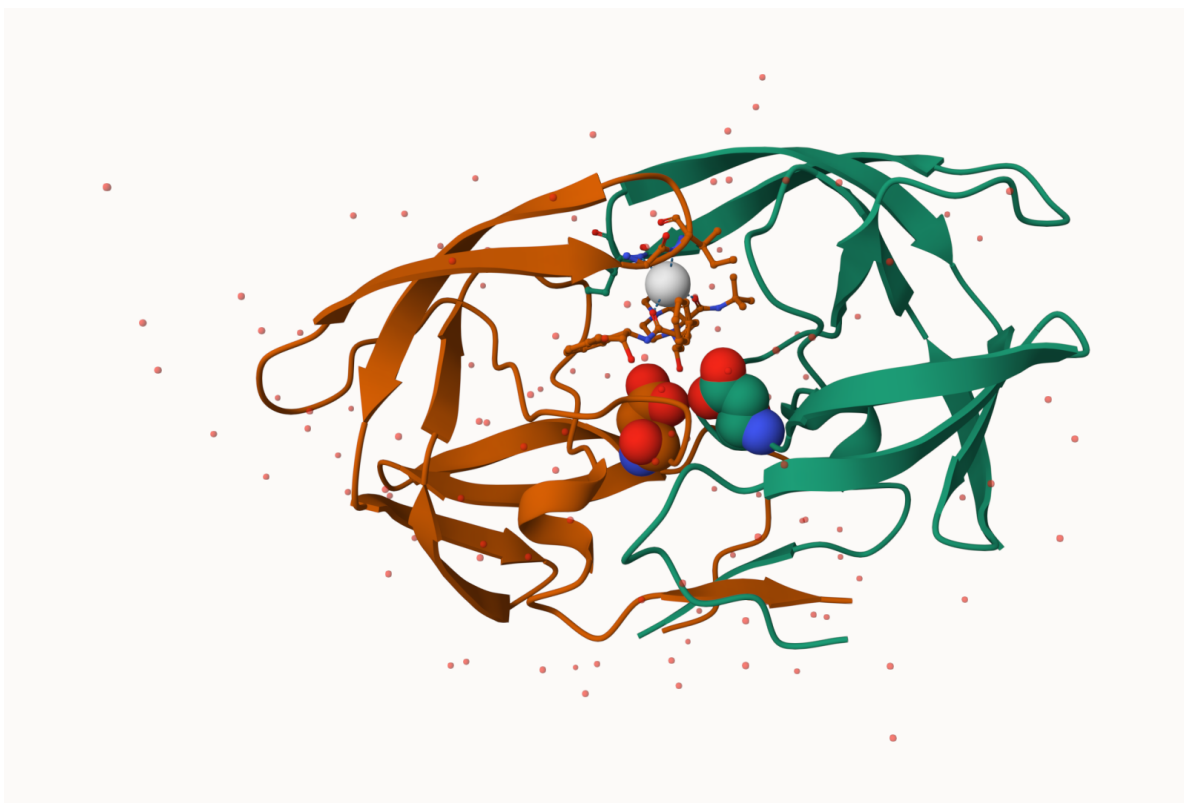


Figure 1: Water molecule is represented as a white ball