

DKOsimR Tutorial

Tutorial: How to generate synthetic CRISPR data using DKOsimR?

Abbreviations: SKO, single knockout; DKO, double knockout; %, percentage; GI, genetic interaction; std. dev., standard deviation.

1. Introduction

This tutorial introduces DKOsimR package for generating synthetic CRISPR double knockout data. It mainly includes:

- Full list of all tunable parameters to initialize the simulation. (Section 3)
- Descriptions on how users may generate synthetic CRISPR data in two modes:
 - default simulation, with 4 gene class: negative, wild-type, non-targeting control, positive (Section 4)
 - simulation applicable to approximate real lab data, with 3 gene class: negative (essential), unknown, non-targeting control (Section 5)
- Examples of applying GI detection method on simulated datasets (Section 6)
- Guidance on picking suitable parameters to simulate screens in practice (Section 7)

2. Install and Load DKOsimR

To start using the package, simply download and install R/RStudio as the first step. You may then install and load the package into your R session with following commands:

```
if(!requireNamespace("devtools", quietly = TRUE))
  install.packages("devtools")
devtools::install_github("yuegu-phd/DKOsimR", quiet = TRUE)
#> Installing 1 packages: data.table
library(DKOsimR)
```

Make sure all required dependencies are installed using `devtools::install(dependencies = TRUE)`

3. List of Tunable Parameters:

Let's go through the designed parameters in simulation framework. Below list out all tunable parameters and how it is represented in DKOsimR.

- Initialized Library Parameters:
 - `sample_name`: name of the simulation run
 - `coverage`: cell representation per guide
 - `n`: number of unique single gene
 - `n_guide_g`: number of guide per gene
 - `moi`: multiplicity of infection - % of cells that are transfected by any virus
 - `sd_freq0`: dispersion of initial counts distribution
- GI Parameters:
 - `p_gi`: % of GI presence
 - `sd_gi`: std. dev. of re-sampled phenotype with GI presence

- Gene Class Parameters:
 1. *% of theoretical phenotype to each gene class - make sure they add up to 1*
 - `pt_neg`: % negative
 - `pt_pos`: % positive
 - `pt_wt`: % wild-type
 - `pt_ctrl`: % non-targeting control
 2. *Mean and std. dev. of theoretical phenotype*
 - `mu_neg`: mean of negative genes
 - `sd_neg`: std. dev. of negative genes
 - `mu_pos`: mean of positive genes
 - `sd_pos`: std. dev. of positive genes
 - `sd_wt`: std. dev. of wild-type genes
- Guide Parameters:
 1. *high-efficacy guides proportion and CRISPR mode*
 - `p_high`: % of high-efficacy guides
 - `mode`: CRISPR mode:
 - * use `CRISPRn-100%Eff` if need 100% efficient guides without randomization
 - * use `CRISPRn` if need high-efficient guides drawn from distribution
 2. *Mean and std. dev. of guide-efficacy*
 - `mu_high`: mean of high-efficacy guides
 - `sd_high`: std. dev of high-efficacy guides
 - `mu_low`: mean of low-efficacy guides
 - `sd_low`: std. dev of high-efficacy guides
- Bottleneck Parameters:
 - `size.bottleneck`: bottleneck size - threshold indicating the ceiling of cell growth
 - `n.bottlenecks`: number of bottleneck encounters - how many times do we encounter bottlenecks?
 - `n.iterations`: number of maximum doubling cycles, by default, we assume a maximum of 30 doublings if we didn't encounter bottleneck
- Randomization Parameter:
 - `rseed`: values used for random number generator - use same number to control same sets of genes having GI
- Miscellaneous:
 - `path`: path to directory to save outputs of data and logs from simulation
 - `cores_free`: number of cores that are left to be free in parallel computing

4. Running Simulation

To generate synthetic double knockout data, **by default, the simulated datasets would be stored under data/ in the current directory, use `getwd()` to navigate your current working directory.** The default values for parameters of simulated CRISPR screens are set based on empirical assumptions as follows:

- Initialized Library Parameters:
 - `coverage`: 100
 - `n_guide_g`: 3
 - `moi`: 0.3

- `sd_freq0`: 1/3.29 (chosen by setting a 10-fold difference between 95th and 5th percentiles of SKO counts distribution)
- GI Parameters:
 - `p_gi`: 0.03
 - `sd_gi`: 1.5
- Gene Class Parameters:
 1. *% of theoretical phenotype to each gene class - make sure they add up to 1*
 - `pt_neg`: 0.15
 - `pt_pos`: 0.05
 - `pt_wt`: 0.75
 - `pt_ctrl`: 0.05
 2. *Mean and std. dev. of theoretical phenotype*
 - `mu_neg`: -0.75
 - `sd_neg`: 0.1
 - `mu_pos`: 0.75
 - `sd_pos`: 0.1
 - `sd_wt`: 0.25
- Guide Parameters:
 1. *high-efficacy guides proportion and CRISPR mode*
 - `p_high`: 1
 - `mode`: CRISPR mode: CRISPRn-100%Eff
 2. *Mean and std. dev. of guide-efficacy*
 - `mu_high`: 0.9
 - `sd_high`: 0.1
 - `mu_low`: 0.05
 - `sd_low`: 0.07
- Bottleneck Parameters:
 - `size.bottleneck`: 2
 - `n.bottlenecks`: 1
 - `n.iterations`: 30
- Randomization Parameter:
 - `rseed`: NULL
- Miscellaneous:
 - `path`: current working directory
 - `cores_free`: 1

To run simulation by default, simply name your simulation by `sample_name` and specify the number of single genes by `n`. **Be cautious that number of genes in each gene class should be an integer to optimize simulation run.** A quick Simulation Settings Summary would be returned for each run. Additionally, number of cores used for parallel computing, Run Time in unit of hours would be collected after one successful run. An example running code is

```
dkosim(sample_name="test", n=40)
#>
#> # -----
#> # Simulation Settings Summary:
```

```

#> # -----
#> # Sample Name: test
#> # Number of Genes: 40
#> # Cell Library Size (Initial): 714000
#> # Coverage: 100 x
#> # Number of Single Knockout(SKO): 40
#> # Number of Double Knockout(DKO): 780
#> # Number of Guides per Gene: 3
#> # Number of Constructs: 7140
#> # Variance of Initialized Counts: 0.09
#>
#> # Genetic Interactions (GI):
#> ## Proportion of GI(%): 3
#> ## Number of Interacting Gene Pairs: 23
#> ## Variance of re-sampled phenotypes w/ GI: 2.25
#>
#> # Proportion of Each Initialized Gene Class (by theoretical phenotypes):
#> ## Negative(%): 15 ~ TN( -0.75 , 0.01 , -1, -0.025)
#> ## Positive(%): 5 ~ TN( 0.75 , 0.01 , 0.025, 1)
#> ## Wild-Type(%): 75 ~ TN(0, 0.0625 , -0.025, 0.025)
#> ## Non-Targeting Control(%): 5 ~ Delta(0)
#>
#> # Proportion of Guides (by efficacy):
#> ## High-efficacy(%): 100 ~ 1
#> ## Low-efficacy(%): 0 ~ TN( 0.05 , 0.0049 , 0, 1)
#>
#> # Multiplicity of Infection (MOI): 0.3
#> # Percentage of viral particles delivered in cells during transfection(%): 22.22 ~ Poisson( 0.3 )
#> # Resampling Size based on MOI (Passage Size): 317367
#> # Bottleneck Size ( 2 x Initial Guide-Level Library Size): 1428000
#> # Number of Bottleneck Encounters (Number of Passages): 1
#> # Total Available Doublings: 30
#> # Number of Replicates: 2
#> # Pseudo-count: 5e-07
#>
#> # -----
#>
#> Attaching package: 'dplyr'
#> The following objects are masked from 'package:stats':
#>
#>     filter, lag
#> The following objects are masked from 'package:base':
#>
#>     intersect, setdiff, setequal, union
#> Loading required package: iterators
#> Loading required package: parallel
#> Warning: package 'data.table' was built under R version 4.3.3
#>
#> Attaching package: 'data.table'
#> The following objects are masked from 'package:dplyr':
#>
#>     between, first, last
#> [1] "repA completed" "repB completed"

```

```
#> [1] "number of cores used: 13"
#> [1] "Run Time (hrs): 0.003516111111111111"
```

Alternatively, you may adjust values to any tunable parameters as desired, but please make sure your input on percentage of each gene class add up to 1 for all classes, and each initialized number of genes is an integer. **You may also change the output directory using path in the function; by default, the output simulated data and log is under the same directory of current project workspace.** The randomization seed can also be specified by `rseed` to ensure same subsets of gene-pairs has GI in multiple run.

An example running code is

```
dkosim(sample_name="test",
        coverage=10,
        n=60,
        n_guide_g=2,
        sd_freq0 = 1/3.29,
        moi = 0.3,
        p_gi=0.03,
        sd_gi=1.5,
        pt_neg=0.15,
        pt_pos=0.05,
        pt_wt=0.75,
        pt_ctrl=0.05,
        mu_neg=-0.75,
        sd_neg=0.1,
        mu_pos=0.75,
        sd_pos=0.1,
        sd_wt=0.25,
        p_high=0.8,
        mode="CRISPRn",
        mu_high=0.8,
        sd_high=0.2,
        mu_low=0.1,
        sd_low=0.08,
        size.bottleneck = 3,
        n.bottlenecks= 2,
        n.iterations = 30,
        rseed = 111,
        path = ".",
        cores_free = 2)

#>
#> # -----
#> # Simulation Settings Summary:
#> # -----
#> # Sample Name: test
#> # Number of Genes: 60
#> # Cell Library Size (Initial): 72000
#> # Coverage: 10 x
#> # Number of Single Knockout(SKO): 60
#> # Number of Double Knockout(DKO): 1770
#> # Number of Guides per Gene: 2
#> # Number of Constructs: 7200
#> # Variance of Initialized Counts: 0.09
#>
```

```

#> # Genetic Interactions (GI):
#> ## Proportion of GI(%): 3
#> ## Number of Interacting Gene Pairs: 53
#> ## Variance of re-sampled phenotypes w/ GI: 2.25
#>
#> # Proportion of Each Initialized Gene Class (by theoretical phenotypes):
#> ## Negative(%): 15 ~ TN( -0.75 , 0.01 , -1, -0.025)
#> ## Positive(%): 5 ~ TN( 0.75 , 0.01 , 0.025, 1)
#> ## Wild-Type(%): 75 ~ TN(0, 0.0625 , -0.025, 0.025)
#> ## Non-Targeting Control(%): 5 ~ Delta(0)
#>
#> # Proportion of Guides (by efficacy):
#> ## High-efficacy(%): 80 ~ TN( 0.8 , 0.04 , 0.6, 1)
#> ## Low-efficacy(%): 20 ~ TN( 0.1 , 0.0064 , 0, 0.6)
#>
#> # Multiplicity of Infection (MOI): 0.3
#> # Percentage of viral particles delivered in cells during transfection(%): 22.22 ~ Poisson( 0.3 )
#> # Resampling Size based on MOI (Passage Size): 48005
#> # Bottleneck Size ( 3 x Initial Guide-Level Library Size): 216000
#> # Number of Bottleneck Encounters (Number of Passages): 2
#> # Total Available Doublings: 30
#> # Number of Replicates: 2
#> # Pseudo-count: 5e-06
#>
#> # -----
#>
#> [1] "repA completed" "repB completed"
#> [1] "number of cores used: 12"
#> [1] "Run Time (hrs): 0.00193416666666667"

```

5. Laboratory Data Pattern Approximation

To utilize the simulation framework in approximating actual laboratory data pattern, `dkosim_lab` was designed to include three initialized gene class. You may specify the percentage of essential genes by `pt_neg`, unknown by `pt_unknown`, and non-targeting controls by `pt_ctrl` accordingly. In our designed framework, essential genes is defined by negative genes; unknown genes might compose of genes with theoretical phenotype in either negative, positive, wild-type, or non-targeting control gene class.

By default, `dkosim_lab` run simulation with following parameter

- Initialized Library Parameters
 - coverage: 100
 - n_guide_g: 3
 - moi: 0.3
 - sd_freq0: 1/2.56 (chosen by setting a 10-fold difference between 90th and 10th percentiles of SKO counts distribution)
- GI Parameters:
 - p_gi: 0.03
 - sd_gi: 1.5
- Gene Class Parameters:
 1. % of theoretical phenotype to each gene class - make sure they add up to 1

- pt_neg: 0.15
 - pt_unknown: 0.80
 - pt_ctrl: 0.05
- 2. *Mean and std. dev. of theoretical phenotype*
 - mu_neg: -0.03
 - sd_neg: 0.25
 - sd_unknown: 0.25
- Guide Parameters:
 1. *high-efficacy guides proportion and CRISPR mode*
 - p_high: 0.75
 - mode: CRISPR mode: CRISPRn
 2. *Mean and std. dev. of guide-efficacy*
 - mu_high: 0.9
 - sd_high: 0.1
 - mu_low: 0.05
 - sd_low: 0.07
- Bottleneck Parameters:
 - size.bottleneck: 2
 - n.bottlenecks: 1
 - n.iterations: 30
- Randomization Parameter:
 - rseed: NULL
- Miscellaneous:
 - path: current working directory
 - cores_free: 1

An example running code is

```
dkosim_lab(sample_name="test_lab", n=20)
#>
#> # -----
#> # Simulation Settings Summary:
#> # -----
#> # Sample Name: test_lab
#> # Number of Genes: 20
#> # Cell Library Size (Initial): 177000
#> # Coverage: 100 x
#> # Number of Single Knockout(SKO): 20
#> # Number of Double Knockout(DKO): 190
#> # Number of Guides per Gene: 3
#> # Number of Constructs: 1770
#> # Variance of Initialized Counts: 0.15
#>
#> # Genetic Interactions (GI):
#> ## Proportion of GI(%): 3
#> ## Number of Interacting Gene Pairs: 6
#> ## Variance of re-sampled phenotypes w/ GI: 2.25
#>
```

```

#> # Proportion of Each Initialized Gene Class (by theoretical phenotypes):
#> ## Negative/Essential(%): 15 ~ TN( -0.03 , 0.0625 , -1, -0.025)
#> ## Unknown(%): 80 ~ TN(0, 0.0625 , -0.5, 0.5)
#> ## Non-Targeting Control(%): 5 ~ Delta(0)
#>
#> # Proportion of Guides (by efficacy):
#> ## High-efficacy(%): 75 ~ TN( 0.9 , 0.01 , 0.6, 1)
#> ## Low-efficacy(%): 25 ~ TN( 0.05 , 0.0049 , 0, 0.6)
#>
#> # Multiplicity of Infection (MOI): 0.3
#> # Percentage of viral particles delivered in cells during transfection(%): 22.22 ~ Poisson( 0.3 )
#> # Resampling Size based on MOI (Passage Size): 78675
#> # Bottleneck Size ( 2 x Initial Guide-Level Library Size): 354000
#> # Number of Bottleneck Encounters (Number of Passages): 1
#> # Total Available Doublings: 30
#> # Number of Replicates: 2
#> # Pseudo-count: 5e-06
#>
#> # -----
#>
#> [1] "repA completed" "repB completed"
#> [1] "number of cores used: 14"
#> [1] "Run Time (hrs): 0.000689166666666667"

```

Alternatively, you may adjust values to any tunable parameters as desired, or using the parameters described in the full article to approximate actual lab data pattern in double CRISPR screens.

6. Applying GI Detection Methods on Simulated Data

Here, we show how we may apply dLFC (Dede et al., 2020) to calculate GI on simulated data, as an example to analyze the simulation output.

```

knitr::opts_knit$set(root.dir = getwd())
# read example simulated data
sample_name = "example_data"
pseudo_counts = 5e-07
repA_path <- here::here("data", paste0(sample_name, "_repA.csv"))
repB_path <- here::here("data", paste0(sample_name, "_repB.csv"))

# each simulation is containing two replicates A and B, and we take average of the LFC for analysis
cell_lib_guide2_A = read.csv(repA_path) %>%
  dplyr::select(-X)

cell_lib_guide2_B = read.csv(repB_path) %>%
  dplyr::select(-X) %>%
  dplyr::select(guide_id, counts_guide_t0,
               counts_guide_t1, counts_guide_t2,
               rel_freq_guide_t0, rel_freq_guide_t2, LFC)

cell_lib_guide2 = merge(cell_lib_guide2_A, cell_lib_guide2_B, by = "guide_id") %>%
  dplyr::rename(counts_guide_t0_1 = counts_guide_t0.x,
               counts_guide_t0_2 = counts_guide_t0.y,
               counts_guide_t1_1 = counts_guide_t1.x,
               counts_guide_t1_2 = counts_guide_t1.y,

```



```

        counts_guide_t2_1 = counts_guide_t2.x,
        counts_guide_t2_2 = counts_guide_t2.y,
        rel_freq_guide_t0_1 = rel_freq_guide_t0.x,
        rel_freq_guide_t0_2 = rel_freq_guide_t0.y,
        rel_freq_guide_t2_1 = rel_freq_guide_t2.x,
        rel_freq_guide_t2_2 = rel_freq_guide_t2.y,
        LFC_t2_1 = LFC.x,
        LFC_t2_2 = LFC.y) %>%
mutate(guide1_type = ifelse(guide1_type == 1, "high", "low"),
       guide2_type = ifelse(guide2_type == 1, "high", "low"),
       construct_type = ifelse(is.na(gene2_behavior), gene1_behavior,
                               paste0(gene1_behavior, " + ", gene2_behavior)),
       LFC_t2_1 = log2(((rel_freq_guide_t2_1 + pseudo_counts) /
                        (rel_freq_guide_t0_1 + pseudo_counts))),
       LFC_t2_2 = log2(((rel_freq_guide_t2_2 + pseudo_counts) /
                        (rel_freq_guide_t0_2 + pseudo_counts)))) %>%
mutate(LFC = (LFC_t2_1 + LFC_t2_2)/2) # aggregate the LFC between 2 replicates by averaging

# read the simulated data and prepare sample name
sim_data = cell_lib_guide2 %>%
  mutate(construct_type = ifelse(KO_type == "SKO", "SKO",
                                ifelse(gene1_behavior == "Non-Targeting Control"
                                       | gene2_behavior == "Non-Targeting Control", "SKO", "DKO" )))

nc_gene = unique(as.character(filter(sim_data, gene1_behavior == "Non-Targeting Control")$gene1))

# calculate single mutant/knockout fitness
gene_SMF = sim_data %>%
  filter(construct_type == "SKO", gene1_behavior != "Non-Targeting Control") %>%
  group_by(gene1) %>%
  dplyr::summarise(SMF = mean(LFC))
# calculate mean LFC for gene pairs that don't contain control
gene_LFC = sim_data %>%
  filter(construct_type == "DKO") %>%
  group_by(gene_pair_id) %>%
  dplyr::summarise(LFC_mean = mean(LFC))
# calculate dLFC = LFC_mean - (SMF1 + SMF2)
sim_data_dLFC = left_join(sim_data, gene_SMF, by = "gene1") %>%
  dplyr::rename(gene1_SMF = SMF) %>%
  left_join(gene_SMF, by = c("gene2"="gene1")) %>%
  dplyr::rename(gene2_SMF = SMF) %>%
  left_join(gene_LFC, by = "gene_pair_id") %>%
  mutate(gene1_SMF = ifelse(is.na(gene1_SMF), 0, gene1_SMF),
         gene2_SMF = ifelse(is.na(gene2_SMF), 0, gene2_SMF),
         LFC_mean = ifelse(construct_type == "SKO",
                           ifelse(gene1_SMF == 0, gene2_SMF, gene1_SMF), LFC_mean)) %>%
  mutate(dLFC = LFC_mean - (gene1_SMF+gene2_SMF))

# add data frame for the non-control group
control = sim_data_dLFC %>% filter(gene1 %in% nc_gene | gene2 %in% nc_gene)
sim_dLFC_noncontrol = sim_data_dLFC %>% anti_join(control, by = "gene1_gene2_id")
# add data frame for the non-interacting group

```

```

control0 = sim_data_dLFC %>% filter(interaction_gene == 0)
sim_dLFC_noncontrol0 = sim_data_dLFC %>% anti_join(control0, by = "gene1_gene2_id")

# compute correlation and p-values
cor_test_all <- cor.test(sim_data_dLFC$interaction_gene,
                        sim_data_dLFC$dLFC, method = "pearson")
cor_test_noncontrol <- cor.test(sim_dLFC_noncontrol$interaction_gene,
                               sim_dLFC_noncontrol$dLFC, method = "pearson")
cor_test_noncontrol0 <- cor.test(sim_dLFC_noncontrol0$interaction_gene,
                                 sim_dLFC_noncontrol0$dLFC, method = "pearson")

# show correlation value
print(cor_test_all)
#>
#> Pearson's product-moment correlation
#>
#> data: sim_data_dLFC$interaction_gene and sim_data_dLFC$dLFC
#> t = 106.64, df = 16108, p-value < 2.2e-16
#> alternative hypothesis: true correlation is not equal to 0
#> 95 percent confidence interval:
#> 0.6341445 0.6522501
#> sample estimates:
#> cor
#> 0.6432872
print(cor_test_noncontrol)
#>
#> Pearson's product-moment correlation
#>
#> data: sim_dLFC_noncontrol$interaction_gene and sim_dLFC_noncontrol$dLFC
#> t = 102.06, df = 14533, p-value < 2.2e-16
#> alternative hypothesis: true correlation is not equal to 0
#> 95 percent confidence interval:
#> 0.6365640 0.6555061
#> sample estimates:
#> cor
#> 0.6461345
print(cor_test_noncontrol0)
#>
#> Pearson's product-moment correlation
#>
#> data: sim_dLFC_noncontrol0$interaction_gene and sim_dLFC_noncontrol0$dLFC
#> t = 53.569, df = 475, p-value < 2.2e-16
#> alternative hypothesis: true correlation is not equal to 0
#> 95 percent confidence interval:
#> 0.9123672 0.9380457
#> sample estimates:
#> cor
#> 0.9262742

```

Visualization on scatterplot to the calculated GI in this example

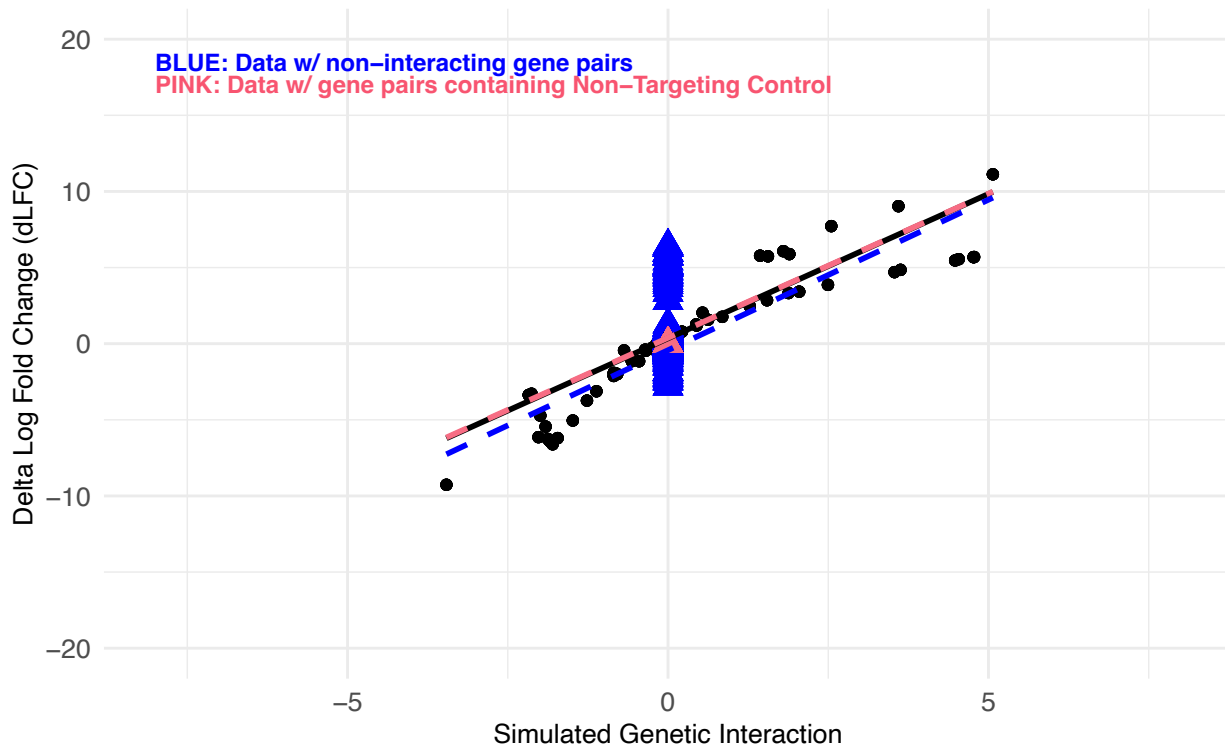
```

#> Ignoring unknown labels:
#> * colour : "Gene-gene Interaction Type"
#> `geom_smooth()` using formula = 'y ~ x'
#> `geom_smooth()` using formula = 'y ~ x'

```

```
#> `geom_smooth()` using formula = 'y ~ x'
```

Scatter Plot for dLFC vs. Simulated GI:
 Correlation $r = 0.643$, $p < 1e-16$
 Correlation r (w/o non-targeting controls) = **0.646**, $p < 1e-16$
 Correlation r (w/o non-interacting gene pairs) = **0.926**, $p < 1e-16$



7. Summary Guidance in Picking Suitable Parameters

In this section, from empirical DKO simulation runs, we provide a summary guidance on picking suitable parameters to run your simulated CRISPR screens in practice. **Please note that this section is mainly for a general suggestion to pick hyperparameters that could efficiently run simulations in practice, adjust the choice of parameters based on your desires and purpose when using DKOsimR, and check reference article for more detailed info.** Following the list of all tunable parameters:

- Initialized Library Parameters:
 - **sample_name**: name simulation with number of genes, number of guides per gene, number of initialized gene classes, and coverage, separate by “_”. For example, “DKOsimR_120x3x4_100x_run1” indicates the 1st run for DKO simulation run with 120 genes, 3 guides per genes, and all 4 initialized gene classes (negative, wild-type, positive, non-targeting control), with 100x coverage.
 - **coverage**: 100 would be sufficient for most cases, set lower to shorten running time but simulated data might have weaker reproducibility.
 - **n**: put the number of unique single gene by desire. 100 would be sufficient for most cases.
 - **n_guide_g**: set as 2 or 3.
 - **moi**: keep unchanged. Default value is 0.3.
 - **sd_freq0**: 1/3.29 by default, chosen by setting a 10-fold difference between 95th and 5th percentiles of SKO counts distribution. Set to 1/2.56, 1/2.07, 1/1.68, 1/1.35, 1/1.05, if you want a 10-fold difference between 90th and 10th, 85th and 15th, 80th and 20th, 75th and 25th, 70th and 30th percentiles, respectively.
- GI Parameters:

- `p_gi`: 0.03 by default, chosen by empirical assumption on the presence of GI in real lab that are close to 3%. Can be tuned higher if want higher presence of GIs.
- `sd_gi`: 1.5 by default and would be sufficient for most cases. This directly affects GI magnitude, and can be tuned lower for smaller GI magnitude.
- Gene Class Parameters:
 1. *% of theoretical phenotype to each gene class - make sure they add up to 1*
 - `pt_neg`: set to values from 0.15 to 0.3 as desired
 - `pt_pos`: set to values from 0 to 0.05 as desired.
 - `pt_wt`: set to values from 0.7 to 0.9 as desired.
 - `pt_ctrl`: set to values from 0.05 to 0.1 as desired.
 2. *Mean and std. dev. of theoretical phenotype*
 - `mu_neg`: set to values from -0.01 to -0.8 as desired
 - `sd_neg`: set to values from 0.1 to 0.5 as desired
 - `mu_pos`: set to values from 0.01 to 0.8 as desired
 - `sd_pos`: set to values from 0.1 to 0.5 as desired
 - `sd_wt`: set to values from 0.1 to 0.5 as desired
- Guide Parameters:
 1. *high-efficacy guides proportion and CRISPR mode*
 - `p_high`: 1 by default, set from 0.75 to 0.8 to resemble real lab data.
 - `mode`: CRISPR mode:
 - * use `CRISPRn-100%Eff` if need 100% efficient guides without randomization (theoretical case), ensure `p_high` is 1 when using this mode (default mode)
 - * use `CRISPRn` if need high-efficient guides drawn from distribution (practical case for actual lab setting)
 2. *Mean and std. dev. of guide-efficacy*
 - `mu_high`: keep unchanged with default value of 0.9.
 - `sd_high`: keep unchanged with default value of 0.1.
 - `mu_low`: keep unchanged with default value of 0.05.
 - `sd_low`: keep unchanged with default value 0.07.
- Bottleneck Parameters:
 - `size.bottleneck`: 2 would be sufficient for most cases. Set it with values of at least 2 to let the simulated cells have chance to grow before reaching the first bottleneck
 - `n.bottlenecks`: 1 by default would be sufficient for most cases. This value directly represents the number of passaging, considering use 2 or 3 if want more passages, but might dramatically decrease the cell population diversity as a result.
 - `n.iterations`: keep unchanged with default value of 30.
- Randomization Parameter:
 - `rseed`: use a same number to control same sets of genes having GI
- Miscellaneous:
 - `path`: keep default or change to path to your project folder
 - `cores_free`: keep default for shortest running time, but increase the number to avoid hard coding issues

To approximate patterns from actual laboratory DKO screens. We suggest using `dkosim_lab()` by inputting the same number of genes in each gene class as the laboratory settings. A case example in mimicking Fong-2024 A549 (Fong et al., 2024) has been described in the second major results section of

DKOsim article. Here, we provide a list of input parameters for DKOsim (mimicking Fong-2024 A549). Empirically, it took around 2 hours to finish running this approximation with 14 cores enabled for parallel computing.

- Initialized Library Parameters:
 - `sample_name`: DKOsimR_246x3x3_1000x_mFong_run1
 - `coverage`: 1000
 - `n`: 246
 - `n_guide_g`: 3 (default)
 - `moi`: 0.3 (default)
 - `sd_freq0`: 1/2.56
- GI Parameters:
 - `p_gi`: 0.03 (default)
 - `sd_gi`: 1.5 (default)
- Gene Class Parameters:
 1. *% of theoretical phenotype to each gene class*
 - `pt_neg`: 64/246
 - `pt_unknown`: 178/246
 - `pt_ctrl`: 4/246
 2. *Mean and std. dev. of theoretical phenotype*
 - `mu_neg`: -0.03
 - `sd_neg`: 0.25
 - `sd_unknown`: 0.2
- Guide Parameters:
 1. *high-efficacy guides proportion and CRISPR mode*
 - `p_high`: 0.75 (default)
 - `mode`: CRISPRn (default)
 2. *Mean and std. dev. of guide-efficacy*
 - `mu_high`: 0.9 (default)
 - `sd_high`: 0.1 (default)
 - `mu_low`: 0.05 (default)
 - `sd_low`: 0.07 (default)
- Bottleneck Parameters:
 - `size.bottleneck`: 2 (default)
 - `n.bottlenecks`: 1 (default)
 - `n.iterations`: 30 (default)
- Randomization Parameter:
 - `rseed`: NULL (default)
- Miscellaneous:
 - `path`: NULL (default)
 - `cores_free`: NULL (default)

Codes for the mimicking are provided below, but not run in this tutorial, check full results in the reference article.

```
dkosim_lab(sample_name="DKOsimR_246x3x3_1000x_mFong_run1",
            coverage=1000,
            n=246,
            n_guide_g=3,
            sd_freq0 = 1/2.56,
            moi = 0.3,
            p_gi=0.03,
            sd_gi=1.5,
            pt_neg=64/246,
            pt_unknown=178/246,
            pt_ctrl=4/246,
            mu_neg=-0.03,
            sd_neg=0.25,
            sd_unknown=0.2,
            p_high=0.75,
            mode="CRISPRn",
            mu_high=0.9,
            sd_high=0.1,
            mu_low=0.05,
            sd_low=0.07,
            size.bottleneck = 2,
            n.bottlenecks= 1,
            n.iterations = 30)
```

Reference

- Gu, Y., Hart, T., Leon-Novelo, L., Shen, J.P.. Double-CRISPR Knockout Simulation (DKOsim): A Monte-Carlo Randomization System to Model Cell Growth Behavior and Infer the Optimal Library Design for Growth-Based Double Knockout Screens. *bioRxiv* 2025.09.11.675497. DOI: 10.1101/2025.09.11.675497.
- Dede, M., McLaughlin, M., Kim, E., & Hart, T. (2020). Multiplex enCas12a screens detect functional buffering among paralogs otherwise masked in monogenic Cas9 knockout screens. *Genome biology*, 21(1), 262. DOI: 10.1186/s13059-020-02173-2
- Fong, S.H., Kuenzi, B.M., Mattson, N.M. et al. A multilineage screen identifies actionable synthetic lethal interactions in human cancers. *Nat Genet* 57, 154–164 (2025). <https://doi.org/10.1038/s41588-024-01971-9>