

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT	DISEASE Salivary gland duct carcinoma	PHYSICIAN	ORDERING PHYSICIAN Yeh, Yi-Chen	SPECIMEN	SPECIMEN SITE Lung
	NAME Hsu, Hsin Yi		MEDICAL FACILITY Taipei Veterans General Hospital		SPECIMEN ID S111-44771 B (PF22129)
	DATE OF BIRTH 14 August 1942		ADDITIONAL RECIPIENT None		SPECIMEN TYPE Slide Deck
	SEX Male		MEDICAL FACILITY ID 205872		DATE OF COLLECTION 01 November 2022
	MEDICAL RECORD # 31701163		PATHOLOGIST Not Provided		SPECIMEN RECEIVED 18 November 2022

Biomarker Findings

Microsatellite status - MS-Stable
Tumor Mutational Burden - 2 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

HRAS Q61R
MYC amplification
PIK3CA E545K, P539R
LYN amplification
RAD21 amplification

Report Highlights

- Targeted therapies with potential clinical benefit **approved in another tumor type**: Cobimetinib (p. [7](#)), Selumetinib (p. [7](#)), Trametinib (p. [8](#))
- Evidence-matched clinical trial options based on this patient's genomic findings: (p. [9](#))

BIOMARKER FINDINGS

Microsatellite status - MS-Stable

Tumor Mutational Burden - 2 Muts/Mb

GENOMIC FINDINGS

HRAS - Q61R

10 Trials [see p. 9](#)

MYC - amplification

6 Trials [see p. 11](#)

PIK3CA - E545K, P539R

10 Trials [see p. 12](#)

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. See Biomarker Findings section

No therapies or clinical trials. See Biomarker Findings section

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)
none	Cobimetinib
	Selumetinib
	Trametinib
none	none
none	none

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GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

LYN - amplification [p. 5](#) **RAD21 - amplification** [p. 6](#)

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

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BIOMARKER FINDINGS

BIOMARKER

Microsatellite status

RESULT

MS-Stable

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁻³, including approved therapies nivolumab and pembrolizumab⁴. In a retrospective analysis of 361 patients with solid tumors treated

with pembrolizumab, 3% were MSI-H and experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, $p=0.001$)⁵.

FREQUENCY & PROGNOSIS

Rare instances of MSI have been reported in salivary gland carcinomas⁶⁻⁷; however, two larger studies reported no MSI in 58 total salivary gland tumors⁸⁻⁹. Published data investigating the prognostic implications of MSI in salivary gland carcinoma are limited (PubMed, Feb 2022).

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive

amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor¹⁰. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2¹⁰⁻¹². This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers¹³⁻¹⁵. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{10,12,14-15}.

BIOMARKER

Tumor Mutational Burden

RESULT

2 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1¹⁶⁻¹⁸, anti-PD-1 therapies¹⁶⁻¹⁹, and combination nivolumab and ipilimumab²⁰⁻²⁵. In multiple pan-tumor studies, increased tissue tumor mutational burden (TMB) was associated with sensitivity to immune checkpoint inhibitors^{16-19,26-30}. In the KEYNOTE 158 trial of pembrolizumab monotherapy for patients with solid tumors, significant improvement in ORR was observed for patients with TMB ≥ 10 Muts/Mb (as measured by this assay) compared with those with TMB < 10 Muts/Mb in a large cohort that included multiple tumor types²⁶; similar findings were observed in the KEYNOTE 028 and 012 trials¹⁹. At the same TMB cutpoint, retrospective analysis of

patients with solid tumors treated with any checkpoint inhibitor identified that tissue TMB scores ≥ 10 Muts/Mb were associated with prolonged time to treatment failure compared with scores < 10 muts/Mb (HR=0.68)³⁰. For patients with solid tumors treated with nivolumab plus ipilimumab in the CheckMate 848 trial, improved responses were observed in patients with a tissue TMB ≥ 10 Muts/Mb independent of blood TMB at any cutpoint in matched samples³¹. However, support for higher TMB thresholds and efficacy was observed in the prospective Phase 2 MyPathway trial of atezolizumab for patients with pan-solid tumors, where improved ORR and DCR was seen in patients with TMB ≥ 16 Muts/Mb than those with TMB ≥ 10 and < 16 Muts/Mb²⁹. Similarly, analyses across several solid tumor types reported that patients with higher TMB (defined as $\geq 16-20$ Muts/Mb) achieved greater clinical benefit from PD-1 or PD-L1-targeting monotherapy compared with patients with higher TMB treated with chemotherapy³² or those with lower TMB treated with PD-1 or PD-L1-targeting agents¹⁷.

FREQUENCY & PROGNOSIS

Studies of salivary gland carcinomas reported a median tumor mutational burden (TMB) of 1.7-3.6 Muts/Mb for salivary gland carcinomas, with 13% of salivary gland carcinoma cases harboring a TMB

of ≥ 10 Muts/Mb³³⁻³⁴. For patients with salivary gland carcinoma not treated with immunotherapy, no significant association between high levels of tissue tumor mutational burden (TMB) (≥ 10 mut/Mb) and OS was reported in one study, although OS numerically differed between patients with high and lower TMB (4.5 vs. 15.5 months, adjusted HR=1.20)³⁴.

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma³⁵⁻³⁶ and cigarette smoke in lung cancer³⁷⁻³⁸, treatment with temozolomide-based chemotherapy in glioma³⁹⁻⁴⁰, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes⁴¹⁻⁴⁵, and microsatellite instability (MSI)^{41,44-45}. This sample harbors a TMB level associated with lower rates of clinical benefit from treatment with PD-1- or PD-L1-targeting immune checkpoint inhibitors compared with patients with tumors harboring higher TMB levels, based on several studies in multiple solid tumor types^{17-18,26}.

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ORDERED TEST # ORD-1506062-01

GENOMIC FINDINGS

GENE

HRAS

ALTERATION

Q61R

TRANSCRIPT ID

NM_005343.2

CODING SEQUENCE EFFECT

182A>G

VARIANT CHROMOSOMAL POSITION

chr11:533874

VARIANT ALLELE FREQUENCY (% VAF)

12.5%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of significant clinical benefit for 1 patient each with cholangiocarcinoma⁴⁶ and salivary gland carcinoma⁴⁷ treated with trametinib, as well as strong preclinical data⁴⁸⁻⁵², HRAS activating alterations may predict sensitivity to MEK inhibitors, such as binimetinib, cobimetinib,

trametinib, and selumetinib. The reovirus Reolysin targets cells with activated RAS signaling⁵³⁻⁵⁵ and has demonstrated mixed clinical efficacy, with the highest rate of response reported for head and neck cancer⁵⁶⁻⁶⁴. HRAS activating mutations may also predict sensitivity to farnesyl transferase inhibitors based on Phase 2 studies of tipifarnib in head and neck squamous cell carcinoma (HNSCC) with HRAS-mutated allele frequency $\geq 20\%$ (ORR of 50.0% [9/18], mDOR of 14.7 months, mPFS of 5.9 months, and mOS of 15.4 months), HRAS-mutated salivary gland cancer (8.3% [1/12] PRs, 58.3% [7/12] SDs, mPFS of 7.0 months), and HRAS-mutated metastatic urothelial carcinoma (ORR of 41.7% [5/12], mPFS of 5.1 months)⁶⁵, as well as preclinical evidence in various cancer types⁶⁶⁻⁶⁸. HRAS mutations have been associated with secondary tumors, particularly cutaneous SCCs, occurring after treatment of primary tumors with RAF inhibitors⁶⁹⁻⁷¹. Preclinical studies have also reported that activating HRAS mutations are associated with resistance to EGFR inhibitors⁷²⁻⁷⁴.

FREQUENCY & PROGNOSIS

HRAS mutation has been implicated in salivary gland tumorigenesis and reported in adenocarcinomas as well as other histological subtypes of salivary gland carcinoma⁷⁵⁻⁸². In salivary gland adenocarcinomas, HRAS mutations have been reported in 3/13 cases (23%) examined in one study, but in none of nine cases in another^{79,81}. In one study of salivary gland adenocarcinomas, no statistically significant differences in recurrence or overall survival were detected regardless of HRAS mutational status⁷⁹.

FINDING SUMMARY

HRAS encodes a member of the RAS family of membrane proteins that bind GDP/GTP and possess GTPase activity. Activating mutations in RAS genes can cause uncontrolled cell proliferation and tumor formation⁸³. HRAS alterations affecting amino acids G12, G13, Q61 and K117, as well as the mutations A59T, A146T, A146V, and in-frame duplications in the HRAS switch II region, have been characterized to be activating and oncogenic⁸³⁻⁹⁶.

GENE

MYC

ALTERATION

amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Preclinical data indicate MYC overexpression may predict sensitivity to investigational agents targeting CDK1⁹⁷⁻⁹⁸, CDK2⁹⁹, Aurora kinase A¹⁰⁰⁻¹⁰⁷, Aurora kinase B¹⁰⁸⁻¹¹¹, glutaminase¹¹²⁻¹¹⁵, or BET bromodomain-containing proteins¹¹⁶⁻¹¹⁹, as well as agents targeting both HDAC and PI3K¹²⁰⁻¹²². Exploratory biomarker analysis in a Phase 2 study reported a PFS benefit associated with a combination of the Aurora A kinase inhibitor alisertib and paclitaxel as second-line therapy for patients with MYC-overexpressed small cell lung cancer but not for patients without MYC

overexpression¹²³. A PR was reported for a patient with MYC-amplified invasive ductal breast carcinoma treated with an unspecified Aurora kinase inhibitor and taxol¹²⁴.

— Nontargeted Approaches —

MYC amplification has also been suggested to predict response to chemotherapy in patients with breast cancer in some studies¹²⁵⁻¹²⁶. Preclinical evidence suggests that colon cancer cells with MYC amplification may be more sensitive to 5-fluorouracil and paclitaxel¹²⁷⁻¹²⁸.

FREQUENCY & PROGNOSIS

MYC amplification has been reported in various solid tumors including breast (9.6%), ovarian (6.7%), melanoma (5.8%), endometrial (5.5%), non-small cell lung (5.5%), prostate (4.7%), esophagogastric (4.4%), and colorectal (3.9%) cancer¹²⁹. An early study reported MYC protein expression in 42% of salivary gland pleomorphic adenomas and 56% of

carcinomas in pleomorphic adenoma of the salivary gland⁷⁸. MYC protein expression has also been detected in 86% (12/14) of adenoid cystic carcinomas in one study¹³⁰. Published data investigating the prognostic implications of MYC alterations in salivary gland carcinomas are limited (PubMed, Oct 2022). In 1 study, MYC overexpression was associated with fusions in MYB and shorter disease-free survival for patients with salivary gland ACC (n=33)¹³¹.

FINDING SUMMARY

MYC (c-MYC) encodes a transcription factor that regulates many genes related to cell cycle regulation and cell growth. It is an oncogene and may be activated in as many as 20% of cancers¹³². MYC dysregulation (amplification, overexpression, translocation) has been identified in a number of different cancer types¹³³. MYC amplification has been significantly linked with increased mRNA and protein levels and results in the dysregulation of a large number of target genes^{132,134-135}.

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GENOMIC FINDINGS

GENE

PIK3CA

ALTERATION

E545K, P539R

TRANSCRIPT ID

NM_006218.2, NM_006218.2

CODING SEQUENCE EFFECT

1633G>A, 1616C>G

VARIANT CHROMOSOMAL POSITION

chr3:178936091, chr3:178936074

VARIANT ALLELE FREQUENCY (% VAF)

12.4%, 12.7%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Clinical and preclinical data in various tumor types indicate that PIK3CA activating alterations may predict sensitivity to therapies targeting PI3K¹³⁶⁻¹⁴³, AKT¹⁴⁴⁻¹⁴⁵, or mTOR¹⁴⁶⁻¹⁵³. The Phase 2 NCI-MATCH study of copanlisib for patients with refractory solid tumors harboring PIK3CA mutations with or without PTEN loss met its primary endpoint with an ORR of 16% (4/25 PRs);

responses (PR or SD >6 months) were seen in patients with ameloblastoma, liposarcoma, and carcinomas of the endometrium, ovary, esophagus, lung, and prostate¹⁴³. However, the Phase 2 study of copanlisib for patients with endometrial carcinoma harboring PIK3CA hotspot mutations failed to report any objective responses (n=11)¹⁴². Two other studies of copanlisib for patients with genomically unselected tumors reported 1 CR and 2 PRs (1 unconfirmed) among 16 total patients with PIK3CA-mutated solid tumors with or without PTEN alterations¹⁴⁰⁻¹⁴¹. In the Phase 2 MATCH trial for patients with PIK3CA-mutated solid tumors, 28% (18/65) of patients experienced PFS lasting at least 6 months after treatment with taselisib; however, no ORs were observed in this study¹⁵⁴. A separate Phase 1b study of taselisib in combination with the CDK4/6 inhibitor palbociclib for patients with PIK3CA-mutated solid tumors reported an ORR of 0% (n=12) and a DCR of 17% (2/12)¹⁵⁵. In a Phase 1 trial of the dual PI3K/mTOR kinase inhibitor apitolisib, 79% (11/14) of patients with PIK3CA-mutated advanced solid tumors experienced disease control (3 PRs, 8 SDs)¹⁵⁶. The PI3K inhibitor alpelisib is approved as a single agent for the treatment of patients with PIK3CA-related overgrowth spectrum (PROS)¹⁵⁷, but has

shown limited activity as monotherapy for PIK3CA-mutated solid tumors with a Phase 1a study reporting an ORR of 6.0% (8/134) and a DCR of 58% (78/134)¹⁵⁸.

FREQUENCY & PROGNOSIS

PIK3CA mutations have been reported in various malignancies, with the highest incidences in carcinomas of the uterus (51%)⁴¹, breast (36%)¹⁵⁹⁻¹⁶¹, bladder (23%)¹⁶²⁻¹⁶⁵, head and neck (15%)¹⁶⁶, and stomach (18%)¹⁶⁷. Published data investigating the prognostic implications of PIK3CA alterations in salivary gland carcinomas are limited (PubMed, Nov 2022).

FINDING SUMMARY

PIK3CA encodes p110-α, which is the catalytic subunit of phosphatidylinositol 3-kinase (PI3K). The PI3K pathway is involved in cell signaling that regulates a number of critical cellular functions, including cell growth, proliferation, differentiation, motility, and survival¹⁶⁸⁻¹⁶⁹. PIK3CA alterations that have been characterized as activating, such as observed here, are predicted to be oncogenic¹⁷⁰⁻¹⁹¹.

GENE

LYN

ALTERATION

amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Dasatinib is a kinase inhibitor that targets the BCR-ABL fusion protein, SRC family kinases including LYN (specifically at low nanomolar concentration)¹⁹²⁻¹⁹³, and other kinases, and has been approved to treat chronic myelocytic leukemia (CML) and acute lymphoblastic leukemia (ALL). A pediatric patient with relapsed B-cell acute lymphoblastic leukemia and an NCOR1-LYN fusion

achieved complete remission after 2 weeks of treatment with dasatinib¹⁹⁴. Similarly, a preclinical study showed that treatment with dasatinib significantly increased survival in a xenograft model of leukemic blast cells harboring NCOR1-LYN¹⁹⁵. In preclinical studies of LYN-expressing breast and prostate cancer, dasatinib has been reported to inhibit cell migration and invasion^{192,196}. However, amplification or other genomic alterations in LYN in solid tumors, and their potential predictive value for sensitivity of these tumors to dasatinib and other kinase inhibitors, remains poorly understood.

FREQUENCY & PROGNOSIS

LYN alterations are rare in solid tumors^{129,197}. However, LYN amplification has been reported more frequently, including in ovarian (3.1%),

melanoma (2.3%), prostate (2.2%), breast (1.9%), and endometrial (1.6%) cancers^{129,197}. LYN expression and activation have also been reported in several types of solid tumors, including glioblastoma¹⁹⁸, prostate cancer¹⁹⁹, head and neck squamous cell carcinoma (HNSCC)²⁰⁰, Ewing sarcoma²⁰¹, and breast cancer¹⁹⁶. High LYN expression was associated with lower survival rates for patients with breast, colorectal, and renal cancers^{196,202-203}.

FINDING SUMMARY

LYN encodes a SRC family intracellular membrane-associated tyrosine protein kinase. LYN is expressed predominantly in hematopoietic cells and conveys signals from the B-cell receptor (BCR) and other receptors to activate the PI3K, STAT, and other signaling pathways²⁰⁴⁻²⁰⁵.

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GENOMIC FINDINGS
GENE
RAD21
ALTERATION
amplification

tumors, including breast cancers (7%), melanoma (5.4%), and prostate (2.4%) cancers¹²⁹. RAD21 overexpression has been correlated with poor prognosis in endometrial cancer²⁰⁶, breast cancer²⁰⁷⁻²⁰⁸, Ewing sarcoma²⁰⁹, and colorectal cancer (CRC), especially in KRAS-mutant CRC²¹⁰.

an increase in deletions, insertions, and other rearrangements²¹⁶. High RAD21 expression has also been associated with increased genomic instability²¹⁷. Cohesin complex also organizes chromatin domains and regulates gene expression²¹⁸⁻²¹⁹. Both overexpression and reduction of expression of RAD21 has been reported to alter gene expression²²⁰. RAD21 amplification has been correlated with increased expression in breast^{207,217,221} and endometrial²⁰⁶ cancers. Other RAD21 alterations, including truncating and point mutations, have been reported in the context of cancer, but the majority have not been characterized.

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

There are no therapies to target alterations in this gene.

FREQUENCY & PROGNOSIS

RAD21 amplifications have been reported in solid

FINDING SUMMARY

RAD21 encodes a protein involved in DNA double-strand break repair and sister chromatid cohesion as a part of the cohesin complex²¹¹⁻²¹⁴. In preclinical studies, downregulation of RAD21 or other cohesin components leads to loss of expression from amplified genes, as well as amplifications themselves upon cell passaging²¹⁵, but also leads to

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THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Cobimetinib

Assay findings association
HRAS
Q61R

AREAS OF THERAPEUTIC USE

Cobimetinib is a MEK inhibitor that is FDA approved to treat patients with histiocytic neoplasms. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of limited clinical data⁴⁶⁻⁴⁷ and strong preclinical data⁴⁸⁻⁵², HRAS activating alterations may predict sensitivity to MEK inhibitors.

SUPPORTING DATA

Clinical data on the efficacy of cobimetinib for the treatment of salivary gland carcinoma are limited

(PubMed, Nov 2022). Single-agent cobimetinib has shown clinical activity in the context of histiocytic neoplasms, including Langerhans cell histiocytosis, Erdheim-Chester disease, Rosai-Dorfman disease, nodular histiocytosis, and mixed histiocytosis²²²⁻²²⁹. A Phase 1 study of cobimetinib monotherapy in solid tumors reported CRs for 1.0% (1/97) of patients and PRs for 6.2% (6/97) of patients, all of whom had melanoma²³⁰. Clinical benefit following treatment with cobimetinib has been seen for patients with non-small cell lung cancer (NSCLC)²³⁰⁻²³¹, adenoid cystic carcinoma²³⁰, and anaplastic pleomorphic xanthoastrocytoma²³².

Selumetinib

Assay findings association
HRAS
Q61R

AREAS OF THERAPEUTIC USE

Selumetinib is a MEK inhibitor that is FDA approved to treat pediatric patients with neurofibromatosis type 1 (NF1)-associated plexiform neurofibromas (PNs). Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of limited clinical data⁴⁶⁻⁴⁷ and strong preclinical data⁴⁸⁻⁵², HRAS activating alterations may predict sensitivity to MEK inhibitors.

SUPPORTING DATA

Clinical data on the efficacy of selumetinib for the treatment of salivary gland and mucoepidermoid carcinoma are limited (PubMed, Nov 2022). Selumetinib has demonstrated efficacy in NF1-associated neurofibroma in Phase 2 studies²³³⁻²³⁵ and a Phase 1 study²³⁶. Phase 2 studies reported clinical responses in low-grade

glioma²³⁷⁻²³⁸, melanoma²³⁹⁻²⁴³, and in lung²⁴⁴⁻²⁴⁶ and endometrial cancer²⁴⁷. A Phase 2 study of selumetinib for patients with activating alterations in the MAPK pathway reported a DCR of 15% (3/20), with no objective responses observed²⁴⁸. Phase 1 studies of selumetinib to treat patients with solid tumors reported 1/15 PR for a patient with colorectal cancer (CRC) and 5/15 SDs for patients with tonsil squamous cell carcinoma (SCC), non-small cell lung cancer (NSCLC), and CRC²⁴⁹; 2/39 PRs for patients with CRC and 18/39 SDs were achieved when selumetinib was administered in combination with cyclosporin A²⁵⁰. Multiple Phase 1 studies combining selumetinib with erlotinib or temsirolimus²⁵¹, docetaxel or dacarbazine²⁵², AKT inhibitors²⁵³, or cixutumumab (an anti-IGF-1R antibody)²⁵⁴ reported clinical responses for patients with advanced solid tumors including NSCLC, thyroid carcinoma, tongue SCC, and ovarian cancer.

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THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Trametinib

Assay findings association
HRAS
Q61R

AREAS OF THERAPEUTIC USE

Trametinib is a MEK inhibitor that is FDA approved as a monotherapy to treat patients with melanoma with BRAF V600E or V600K mutations. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of limited clinical data⁴⁶⁻⁴⁷ and strong preclinical data⁴⁸⁻⁵², HRAS activating alterations may predict sensitivity to MEK inhibitors.

SUPPORTING DATA

A Phase 1 trial of trametinib in 206 patients with solid tumors reported 21 (10%) objective responses²⁵⁵. Phase 1 monotherapy trials of RO4987655, another MEK inhibitor, have shown significant response rates in patients with melanoma, including those with BRAF and NRAS mutations, but very low response rates in patients with other solid tumors, including those with KRAS mutations²⁵⁶⁻²⁵⁷. A Phase 1b trial of trametinib in combination with gemcitabine in patients with solid tumors showed a complete response in a patient with breast cancer, as well as partial responses in pancreatic

and salivary gland cancer²⁵⁸. A Phase 1b trial of combination treatment with the MEK inhibitor binimetinib and the PI3K-alpha inhibitor alpelisib reported disease control (partial responses or stable disease) in 47% (21/45) of patients, including partial responses in 2 of 3 patients with KRAS-mutant ovarian cancer and 1 of 3 patients with NRAS-mutant melanoma; a 43% rate of stable disease was observed in patients with KRAS-mutant colorectal cancer, with responses independent of PIK3CA mutation status²⁵⁹. In Phase 1b trials of trametinib as a single agent or in combination with gemcitabine, 2 PRs were reported for patients with salivary gland cancer^{258,260}. A patient with HRAS Q61R-positive epithelial-myoepithelial salivary gland carcinoma experienced a PR for 5 months to treatment with trametinib⁴⁷. Whereas frequent adverse events precluded a recommended Phase 2 dose and schedule for the combination of trametinib and everolimus in a Phase 1b trial for solid tumors²⁶¹, a retrospective study for heavily pretreated patients with solid tumors reported tolerable regimens of the combination for 23/31 patients, with 16 patients treated >3 months and evaluable patients achieving a median PFS of 6.5 months²⁶².

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

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Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

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CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually

updated and should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that

may require medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://www.clinicaltrials.gov). Or, visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

GENE
HRAS
ALTERATION
Q61R
RATIONALE

HRAS activating alterations may predict sensitivity to MEK or farnesyl transferase

inhibitors.

NCT04985604
PHASE 1/2

DAY101 Monotherapy or in Combination With Other Therapies for Patients With Solid Tumors

TARGETS
BRAF, MEK

LOCATIONS: Busan (Korea, Republic of), Seoul (Korea, Republic of), Oregon, Barcelona (Spain), Madrid (Spain), California, Colorado, Toronto (Canada), Indiana

NCT04803318
PHASE 2

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

TARGETS
mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK

LOCATIONS: Guangzhou (China)

NCT03284502
PHASE 1

Cobimetinib and HM95573 in Patients With Locally Advanced or Metastatic Solid Tumors

TARGETS
MEK, RAFs, NRAS

LOCATIONS: Hwasun (Korea, Republic of), Pusan (Korea, Republic of), Seongnam (Korea, Republic of), Seoul (Korea, Republic of), Goyang-si (Korea, Republic of)

NCT04801966
PHASE NULL

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

TARGETS
CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF

LOCATIONS: Melbourne (Australia)

NCT04720976
PHASE 1/2

JAB-3312 Activity in Adult Patients With Advanced Solid Tumors

TARGETS
MEK, SHP2, PD-1, EGFR, KRAS

LOCATIONS: Utah, California, Arizona, Minnesota, Illinois, Michigan, Oklahoma, Missouri, Indiana, Connecticut

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CLINICAL TRIALS
NCT04965818
PHASE 1/2

Phase 1b/2 Study of Futibatinib in Combination With Binimetinib in Patients With Advanced KRAS Mutant Cancer

TARGETS
MEK, FGFRs

LOCATIONS: California, Indiana, Texas

NCT03905148
PHASE 1/2

Study of the Safety and Pharmacokinetics of BGB-283 and PD-0325901 in Patients With Advanced or Refractory Solid Tumors

TARGETS
RAFs, EGFR, MEK

LOCATIONS: Nedlands (Australia), Blacktown (Australia), Randwick (Australia), Melbourne (Australia), California, Texas

NCT05159245
PHASE 2

The Finnish National Study to Facilitate Patient Access to Targeted Anti-cancer Drugs

TARGETS
BRAF, VEGFRs, RET, KIT, ERBB2, TRKB, ALK, TRKC, ROS1, TRKA, SMO, PD-L1, MEK, CDK4, CDK6

LOCATIONS: Kuopio (Finland), Helsinki (Finland), Tampere (Finland), Turku (Finland)

NCT04551521
PHASE 2

CRAFT: The NCT-PMO-1602 Phase II Trial

TARGETS
PD-L1, AKTs, MEK, BRAF, ALK, RET, ERBB2

LOCATIONS: Würzburg (Germany), Mainz (Germany), Heidelberg (Germany), Tübingen (Germany)

NCT04892017
PHASE 1/2

A Safety, Tolerability and PK Study of DCC-3116 in Patients With RAS or RAF Mutant Advanced or Metastatic Solid Tumors.

TARGETS
ULK1, ULK2, MEK

LOCATIONS: Massachusetts, Texas, Pennsylvania

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CLINICAL TRIALS
GENE
MYC
ALTERATION
amplification

RATIONALE

MYC overexpression may predict sensitivity to inhibition of CDKs, especially CDK1 and CDK2, of Aurora kinases, including Aurora kinase A and B,

and of BET domain proteins, which are reported to downregulate MYC expression and MYC-dependent transcriptional programs.

NCT04983810
PHASE 1/2

A Study to Investigate Fadraciclub (CYC065), in Subjects With Advanced Solid Tumors and Lymphoma

TARGETS
CDK2, CDK9

LOCATIONS: Seoul (Korea, Republic of), Barcelona (Spain), California, Texas

NCT05252390
PHASE 1/2

NUV-868 as Monotherapy and in Combination With Olaparib or Enzalutamide in Adult Patients With Advanced Solid Tumors

TARGETS
BRD4, PARP, AR

LOCATIONS: Michigan, Texas, Tennessee, Maryland, Virginia, North Carolina

NCT04742959
PHASE 1/2

Crossover Relative Bioavailability and Dose Escalation Study of TT-00420 Tablet in Patients With Advanced Solid Tumors

TARGETS
Aurora kinase A, Aurora kinase B

LOCATIONS: California, Illinois, Ohio, Texas, New Jersey

NCT04840589
PHASE 1

Testing the Combination of ZEN003694 and Nivolumab With or Without Ipilimumab in Solid Tumors

TARGETS
PD-1, CTLA-4, BRD4, BRDT, BRD2, BRD3

LOCATIONS: Ohio, Pennsylvania, New York, Maryland

NCT04555837
PHASE 1/2

Alisertib and Pembrolizumab for the Treatment of Patients With Rb-deficient Head and Neck Squamous Cell Cancer

TARGETS
Aurora kinase A, PD-1

LOCATIONS: Texas

NCT01434316
PHASE 1

Veliparib and Dinaciclib in Treating Patients With Advanced Solid Tumors

TARGETS
PARP, CDK1, CDK9, CDK5, CDK2

LOCATIONS: Massachusetts

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CLINICAL TRIALS
GENE
PIK3CA
ALTERATION
E545K, P539R
RATIONALE

PIK3CA activating mutations may lead to activation of the PI3K-AKT-mTOR pathway and may therefore indicate sensitivity to inhibitors of

this pathway. Strong clinical data support sensitivity of PIK3CA-mutated solid tumors to the PI3K-alpha inhibitor alpelisib.

NCT04589845
PHASE 2

Tumor-Agnostic Precision Immuno-Oncology and Somatic Targeting Rational for You (TAPISTRY) Platform Study

TARGETS

TRKB, ALK, TRKC, ROS1, TRKA, RET, PD-L1, AKTs, ERBB2, MDM2, PI3K-alpha, RAFs, NRAS

LOCATIONS: Taipei City (Taiwan), Taoyuan County (Taiwan), Tainan (Taiwan), Shanghai City (China), Shanghai (China), Shatin (Hong Kong), Hong Kong (Hong Kong), Seoul (Korea, Republic of), Xi'an (China), Tianjin (China)

NCT04341259
PHASE 1

A Study Of The Pharmacokinetics And Safety Of Ipatasertib In Chinese Participants With Locally Advanced Or Metastatic Solid Tumors.

TARGETS

AKTs

LOCATIONS: Shanghai City (China)

NCT04803318
PHASE 2

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

TARGETS

mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK

LOCATIONS: Guangzhou (China)

NCT04526470
PHASE 1/2

Alpelisib and Paclitaxel in PIK3CA-altered Gastric Cancer

TARGETS

PI3K-alpha

LOCATIONS: Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of)

NCT03772561
PHASE 1

Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies

TARGETS

PARP, AKTs, PD-L1

LOCATIONS: Singapore (Singapore)

NCT04801966
PHASE NULL

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

TARGETS

CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF

LOCATIONS: Melbourne (Australia)

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CLINICAL TRIALS
NCT03297606
PHASE 2

Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)

TARGETS

VEGFRs, ABL, SRC, ALK, ROS1, AXL, TRKA, MET, TRKC, DDR2, KIT, EGFR, PD-1, CTLA-4, PARP, CDK4, CDK6, FLT3, CSF1R, RET, mTOR, ERBB2, MEK, BRAF, SMO

LOCATIONS: Vancouver (Canada), Edmonton (Canada), Saskatoon (Canada), Regina (Canada), Ottawa (Canada), Montreal (Canada), Toronto (Canada), Kingston (Canada), London (Canada)

NCT04551521
PHASE 2

CRAFT: The NCT-PMO-1602 Phase II Trial

TARGETS

PD-L1, AKTs, MEK, BRAF, ALK, RET, ERBB2

LOCATIONS: Würzburg (Germany), Mainz (Germany), Heidelberg (Germany), Tübingen (Germany)

NCT03994796
PHASE 2

Genetic Testing in Guiding Treatment for Patients With Brain Metastases

TARGETS

TRKB, ALK, TRKC, ROS1, TRKA, CDK4, CDK6, PI3K, mTOR

LOCATIONS: Washington, Oregon, Idaho, Montana

NCT03006172
PHASE 1

To Evaluate the Safety, Tolerability, and Pharmacokinetics of GDC-0077 Single Agent in Participants With Solid Tumors and in Combination With Endocrine and Targeted Therapies in Participants With Breast Cancer

TARGETS

PI3K-alpha, Aromatase, ER, CDK6, CDK4

LOCATIONS: London (United Kingdom), Surrey (United Kingdom), Bordeaux (France), Barcelona (Spain), Valencia (Spain), Toronto (Canada), Massachusetts, New York, Tennessee

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APPENDIX

Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

BRCA1
P346S

CASP8
Q46H

CDH1
L21V

CREBBP
M1691L

GATA6
A179_A183del

NBN
amplification

NTRK2
G195V

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APPENDIX

Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B or WTX)	
APC	AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX
AURKA	AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2
BCL6	BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1
BTG2	BTK	CALR	CARD11	CASP8	CBFB	CBL	CCND1	CCND2
CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73	CDH1
CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B	CDKN2C
CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R	CTCF
CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1	DDR2
DIS3	DNMT3A	DOT1L	EED	EGFR	EMSY (C11orf30)	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12	FGF14
FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3	FGFR4
FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3	GATA4
GATA6	GID4 (C17orf39)	GNA11	GNA13	GNAS	GNAS	GRM3	GSK3B	H3-3A (H3F3A)
HDAC1	HGF	HNFA1	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDM5A	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11 (MRE11A)	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD2 (WHSC1 or MMSET)	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3	NTRK3
P2RY8	PALB2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCC1 (PD-1)	PDCC1LG2 (PD-L2)
PDGFRA	PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1
PMS2	POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI
PRKN (PARK2)	PTCH1	PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51
RAD51B	RAD51C	RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10
REL	RET	RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC
SDHD	SETD2	SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO
SNCAIP	SOC1	SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3
STK11	SUFU	SYK	TBX3	TEK	TENT5C (FAM46C)	TET2	TET2	TGFB2
TIPARP	TNFAIP3	TNFRSF14	TP53	TSC1	TSC2	TYRO3	U2AF1	VEGFA
VHL	WT1	XPO1	XRCC2	ZNF217	ZNF703			

DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSP02	SDC4	SLC34A2	TERC*	TERT**	TPRSS2

*TERC is an NCRNA

**Promoter region of TERT is interrogated

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Homologous Recombination status
Loss of Heterozygosity (LOH) score
Microsatellite (MS) status
Tumor Mutational Burden (TMB)

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APPENDIX

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a., Ciplstraat 3, 2440 Geel, Belgium. 

ABOUT FOUNDATIONONE CDx

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:
www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLE

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes

detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for *ERBB2* and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical

methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials

Ranking of Therapies in Summary Table

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

1. In the fraction-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by

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About FoundationOne®CDx

- analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.
- TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
 - Homologous Recombination status may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas (Coleman et al., 2017; 28916367). Samples with deleterious *BRCA1/2* alteration and/or Loss of Heterozygosity (LOH) score $\geq 16\%$ will be reported as "HRD Positive" and samples with absence of these findings will be reported as "HRD Not Detected," agnostic of potential secondary *BRCA1/2* reversion alterations. Certain potentially deleterious missense or small in-frame deletions in *BRCA1/2* may not be classified as deleterious and, in the absence of an elevated LOH profile, samples with such mutations may be classified as "HRD Not Detected." A result of "HRD Not Detected" does not rule out the presence of a *BRCA1/2* alteration or an elevated LOH profile outside the assay performance characteristic limitations.
 - The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and

extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.

- Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
- Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine whether the patient is a candidate for biopsy.
- Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of *HER2* overexpression as 20% in breast cancer. Based on the F1CDx *HER2* CDx concordance study, approximately 10% of *HER2* amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal

hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

*Interquartile Range = 1st Quartile to 3rd Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's

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APPENDIX

About FoundationOne®CDx

tumor sequencing is germline or somatic.
Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D (MLL2)*, *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking

into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

REFERENCE SEQUENCE INFORMATION

Sequence data is mapped to the human genome, Genome Reference Consortium Human Build 37 (GRCh37), also known as hg19.

MR Suite Version (RG) 7.3.0

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APPENDIX References

1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) PMID: 25392179
2. Kroemer G, et al. Oncoimmunology (2015) PMID: 26140250
3. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
4. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
5. Ayers et al., 2016; ASCO-SITC Abstract P60
6. Suzuki H, et al. Diagn. Mol. Pathol. (1998) PMID: 9917133
7. Moore A, et al. Head Neck (2020) PMID: 31762146
8. Ohki K, et al. Int J Oral Maxillofac Surg (2001) PMID: 11829237
9. Nakano T, et al. Oral Oncol. (2019) PMID: 30846173
10. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
11. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
12. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
13. Boland CR, et al. Cancer Res. (1998) PMID: 9823339
14. Pawlik TM, et al. Dis. Markers (2004) PMID: 15528785
15. Boland CR, et al. Gastroenterology (2010) PMID: 20420947
16. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
17. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
18. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
19. Cristescu R, et al. Science (2018) PMID: 30309915
20. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
21. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
22. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
23. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
24. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
25. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
26. Marabelle A, et al. Lancet Oncol. (2020) PMID: 32919526
27. Ott PA, et al. J. Clin. Oncol. (2019) PMID: 30557521
28. Cristescu R, et al. J Immunother Cancer (2022) PMID: 35101941
29. Friedman CF, et al. Cancer Discov (2022) PMID: 34876409
30. Sturgill EG, et al. Oncologist (2022) PMID: 35274716
31. Schenker at al., 2022; AACR Abstract 7845
32. Legrand et al., 2018; ASCO Abstract 12000
33. Chalmers ZR, et al. Genome Med (2017) PMID: 28420421
34. Shao C, et al. JAMA Netw Open (2020) PMID: 33119110
35. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
36. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
37. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
38. Rizvi NA, et al. Science (2015) PMID: 25765070
39. Johnson BE, et al. Science (2014) PMID: 24336570
40. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
41. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
42. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
43. Heitzner E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
44. Nature (2012) PMID: 22810696
45. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
46. Nadauld et al., 2016; ASCO Abstract e23162
47. Chintakuntlawar et al., 2015; ASCO Abstract e17053
48. Rosenberger G, et al. Hum. Mutat. (2009) PMID: 19035362
49. Rosseland CM, et al. J. Cell. Physiol. (2008) PMID: 18163378
50. Ricciardi MR, et al. J. Mol. Med. (2012) PMID: 22399013
51. Leiser D, et al. Mol Oncol (2015) PMID: 25933688
52. Kiessling MK, et al. Oncotarget (2015) PMID: 26544513
53. Strong JE, et al. EMBO J. (1998) PMID: 9628872
54. Coffey MC, et al. Science (1998) PMID: 9812900
55. Gong J, et al. Front Oncol (2014) PMID: 25019061
56. Forsyth P, et al. Mol. Ther. (2008) PMID: 18253152
57. Vidal L, et al. Clin. Cancer Res. (2008) PMID: 18981012
58. Gollamudi R, et al. Invest New Drugs (2010) PMID: 19572105
59. Harrington KJ, et al. Clin. Cancer Res. (2010) PMID: 20484020
60. Comins C, et al. Clin. Cancer Res. (2010) PMID: 20926400
61. Lolkema MP, et al. Clin. Cancer Res. (2011) PMID: 21106728
62. Galanis E, et al. Mol. Ther. (2012) PMID: 22871663
63. Karapanagiotou EM, et al. Clin. Cancer Res. (2012) PMID: 22316603
64. Morris DG, et al. Invest New Drugs (2013) PMID: 22886613
65. Ho et al., 2020; ASCO Abstract 6504
66. End DW, et al. Cancer Res. (2001) PMID: 11196150
67. Chen X, et al. Oncogene (2014) PMID: 24240680
68. Cohen-Jonathan E, et al. Radiat. Res. (2000) PMID: 10297863
69. Oberholzer PA, et al. J. Clin. Oncol. (2012) PMID: 22067401
70. Su F, et al. N. Engl. J. Med. (2012) PMID: 22256804
71. Lacouture ME, et al. Oncologist (2013) PMID: 23457002
72. Hah JH, et al. Head Neck (2014) PMID: 24123531
73. Frasca F, et al. J. Clin. Endocrinol. Metab. (2013) PMID: 23559083
74. Kasper S, et al. Oncogene (2013) PMID: 22797062
75. Oral Dis (2009) PMID: 19317835
76. Vander Poorten V, et al. Head Neck (2012) PMID: 21618326
77. Milasin J, et al. Int J Oral Maxillofac Surg (1993) PMID: 8106812
78. Deguchi H, et al. Acta Pathol. Jpn. (1) PMID: 8396843
79. van Halteren HK, et al. Int. J. Cancer (1994) PMID: 8168996
80. Yoo J, et al. Cancer (2000) PMID: 10649241
81. Yoo J, et al. Arch. Pathol. Lab. Med. (2000) PMID: 10835516
82. Cros J, et al. Ann. Oncol. (2013) PMID: 23933559
83. Pylyayeva-Gupta Y, et al. Nat. Rev. Cancer (2011) PMID: 21993244
84. Baker R, et al. J. Biol. Chem. (2013) PMID: 24247240
85. Buhrman G, et al. Proc. Natl. Acad. Sci. U.S.A. (2010) PMID: 20194776
86. Denayer E, et al. Hum. Mutat. (2008) PMID: 17979197
87. Fasano O, et al. Proc. Natl. Acad. Sci. U.S.A. (1984) PMID: 6330729
88. Feig LA, et al. Mol. Cell. Biol. (1988) PMID: 3043178
89. Janakiramam M, et al. Cancer Res. (2010) PMID: 20570890
90. Privé GG, et al. Proc. Natl. Acad. Sci. U.S.A. (1992) PMID: 1565661
91. Scheffzek K, et al. Science (1997) PMID: 9219684
92. Stephen AG, et al. Cancer Cell (2014) PMID: 24651010
93. Niihori T, et al. J. Hum. Genet. (2011) PMID: 21850009
94. Wey M, et al. Biochemistry (2013) PMID: 24224811
95. Eijkelenboom A, et al. Sci Rep (2019) PMID: 31160609
96. Lorenz S, et al. Hum Mol Genet (2013) PMID: 23335589
97. Horiuchi D, et al. J. Exp. Med. (2012) PMID: 22430491
98. Goga A, et al. Nat. Med. (2007) PMID: 17589519
99. Molenaar JJ, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) PMID: 19525400
100. Dammert MA, et al. Nat Commun (2019) PMID: 31375684
101. Mollaoglu G, et al. Cancer Cell (2017) PMID: 28089889
102. Cardnell RJ, et al. Oncotarget (2017) PMID: 29088717
103. Wang L, et al. Mol Oncol (2017) PMID: 28417568
104. Takahashi Y, et al. Ann. Oncol. (2015) PMID: 25632068
105. Li Y, et al. Thyroid (2018) PMID: 30226440
106. Mahadevan D, et al. PLoS ONE (2014) PMID: 24893165
107. Park SI, et al. Target Oncol (2019) PMID: 31429028
108. Helfrich BA, et al. Mol. Cancer Ther. (2016) PMID: 27496133
109. Hook KE, et al. Mol. Cancer Ther. (2012) PMID: 22222631
110. Yang D, et al. Proc. Natl. Acad. Sci. U.S.A. (2010) PMID: 20643922
111. He J, et al. Anticancer Drugs (2019) PMID: 30540594
112. Shroff EH, et al. Proc. Natl. Acad. Sci. U.S.A. (2015) PMID: 25964345
113. Effenberger M, et al. Oncotarget (2017) PMID: 29156762
114. Qu X, et al. Biochem. Biophys. Res. Commun. (2018) PMID: 30103944
115. Xiang Y, et al. J. Clin. Invest. (2015) PMID: 25915584
116. Delmore JE, et al. Cell (2011) PMID: 21889194
117. Bandopadhyay P, et al. Clin. Cancer Res. (2014) PMID: 24297863
118. Lovén J, et al. Cell (2013) PMID: 23582323
119. Otto C, et al. Neoplasia (2019) PMID: 31734632
120. Dong LH, et al. J Hematol Oncol (2013) PMID: 23866964
121. Pei Y, et al. Cancer Cell (2016) PMID: 26977882
122. Fu XH, et al. Acta Pharmacol. Sin. (2019) PMID: 30224636
123. Owonikoko TK, et al. J Thorac Oncol (2020) PMID: 31655296
124. Ganesan P, et al. Mol. Cancer Ther. (2014) PMID: 25253784
125. Pereira CB, et al. PLoS ONE (2013) PMID: 23555992
126. Yasojima H, et al. Eur. J. Cancer (2011) PMID: 21741827
127. Arango D, et al. Cancer Res. (2001) PMID: 11406570
128. Bottone MG, et al. Exp. Cell Res. (2003) PMID: 14516787
129. Zehir A, et al. Nat. Med. (2017) PMID: 28481359
130. von Holstein SL, et al. Ophthalmology (2013) PMID: 23725736
131. Fujii K, et al. Histopathology (2017) PMID: 28594149
132. Dang CV, et al. Semin. Cancer Biol. (2006) PMID: 16904903
133. Nesbit CE, et al. Oncogene (1999) PMID: 10378696
134. Blacato J, et al. Br. J. Cancer (2004) PMID: 15083194
135. Fromont G, et al. Hum. Pathol. (2013) PMID: 23574779
136. Fritsch C, et al. Mol. Cancer Ther. (2014) PMID: 24608574
137. Juric D, et al. J. Clin. Oncol. (2018) PMID: 29401002
138. Gallant JN, et al. NPJ Precis Oncol (2019) PMID: 30793038
139. Delestre F, et al. Sci Transl Med (2021) PMID: 34613809
140. Morschhauser F, et al. Mol Cancer Ther (2020) PMID: 31619463
141. Patnaik A, et al. Ann. Oncol. (2016) PMID: 27672108
142. Santin AD, et al. Gynecol Oncol Rep (2020) PMID: 31934607
143. Damodaran S, et al. J Clin Oncol (2022) PMID: 35133871
144. André F, et al. N. Engl. J. Med. (2019) PMID: 31091374
145. Smyth LM, et al. NPJ Breast Cancer (2021) PMID: 33863913
146. Varnier R, et al. Eur J Cancer (2019) PMID: 31351267

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APPENDIX
References

147. Basse C, et al. JCO Precis Oncol (2018) PMID: 32914004
148. Sultova E, et al. Arch Gynecol Obstet (2021) PMID: 33277683
149. Mackay HJ, et al. Cancer (2014) PMID: 24166148
150. Myers AP, et al. Gynecol. Oncol. (2016) PMID: 27016228
151. Dhami J, et al. Cold Spring Harb Mol Case Stud (2018) PMID: 29588307
152. Harris EJ, et al. Front Oncol (2019) PMID: 30863722
153. Hanna GJ, et al. Clin Cancer Res (2018) PMID: 29301825
154. Krop et al., 2018; ASCO Abstract 101
155. Pascual J, et al. Cancer Discov (2021) PMID: 32958578
156. Dolly SO, et al. Clin. Cancer Res. (2016) PMID: 26787751
157. Canaud et al., 2021; ESMO Abstract LBA23
158. Aust Fam Physician (1986) PMID: 2941002
159. Stephens PJ, et al. Nature (2012) PMID: 22722201
160. Banerji S, et al. Nature (2012) PMID: 22722202
161. Nature (2012) PMID: 23000897
162. Nature (2014) PMID: 24476821
163. Guo G, et al. Nat. Genet. (2013) PMID: 24121792
164. Iyer G, et al. J. Clin. Oncol. (2013) PMID: 23897969
165. Kim PH, et al. Eur. Urol. (2015) PMID: 25092538
166. Nature (2015) PMID: 25631445
167. Nature (2014) PMID: 25079317
168. Samuels Y, et al. Cancer Cell (2005) PMID: 15950905
169. Nat. Rev. Cancer (2009) PMID: 19629070
170. Kang S, et al. Proc. Natl. Acad. Sci. U.S.A. (2005) PMID: 15647370
171. Ikenoue T, et al. Cancer Res. (2005) PMID: 15930273
172. Gymnopoulos M, et al. Proc. Natl. Acad. Sci. U.S.A. (2007) PMID: 17376864
173. Horn S, et al. Oncogene (2008) PMID: 18317450
174. Rudd ML, et al. Clin. Cancer Res. (2011) PMID: 21266528
175. Hon WC, et al. Oncogene (2012) PMID: 22120714
176. Burke JE, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) PMID: 22949682
177. Wu H, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) PMID: 19915146
178. Laurenti R, et al. Rev Saude Publica (1990) PMID: 2103068
179. Dan S, et al. Cancer Res. (2010) PMID: 20530683
180. Oda K, et al. Cancer Res. (2008) PMID: 18829572
181. Zhao L, et al. Oncogene (2008) PMID: 18794883
182. Lui VW, et al. Cancer Discov (2013) PMID: 23619167
183. Ross RL, et al. Oncogene (2013) PMID: 22430209
184. Rivière JB, et al. Nat. Genet. (2012) PMID: 22729224
185. Shibata T, et al. Cancer Lett. (2009) PMID: 19394761
186. Dogruluk T, et al. Cancer Res. (2015) PMID: 26627007
187. Croessmann S, et al. Clin. Cancer Res. (2018) PMID: 29284706
188. Ng PK, et al. Cancer Cell (2018) PMID: 29533785
189. Spangle JM, et al. (2020) PMID: 32929011
190. Chen L, et al. Nat Commun (2018) PMID: 29636477
191. Jin N, et al. J Clin Invest (2021) PMID: 34779417
192. Nam S, et al. Cancer Res. (2005) PMID: 16230377
193. Williams NK, et al. J. Biol. Chem. (2009) PMID: 18984583
194. Dai HP, et al. Front Oncol (2020) PMID: 32266142
195. Tomii T, et al. Leukemia (2021) PMID: 33199837
196. Choi YL, et al. Cancer Res. (2010) PMID: 20215510
197. Nguyen B, et al. Cell (2022) PMID: 35120664
198. Stettner MR, et al. Cancer Res. (2005) PMID: 15994925
199. Goldenberg-Furmanov M, et al. Cancer Res. (2004) PMID: 14871838
200. Wheeler SE, et al. Clin. Cancer Res. (2012) PMID: 22490227
201. Guan H, et al. Mol. Cancer Ther. (2008) PMID: 18644993
202. Huang TH, et al. Cancer Cell (2013) PMID: 23764002
203. Roseweir AK, et al. BMC Cancer (2016) PMID: 26984511
204. Xu Y, et al. Immunity (2005) PMID: 15664155
205. Cell Commun. Signal (2012) PMID: 22805580
206. Supernat A, et al. Oncol Lett (2012) PMID: 23205091
207. Xu H, et al. Breast Cancer Res. (2011) PMID: 21255398
208. Sharaf R, et al. Genome Med (2022) PMID: 35227290
209. Su XA, et al. Genes Dev (2021) PMID: 33766983
210. Deb S, et al. Br. J. Cancer (2014) PMID: 24548858
211. Xu H, et al. Nat. Rev. Cancer (2011) PMID: 21326324
212. Hill VK, et al. Biochim. Biophys. Acta (2016) PMID: 27207471
213. Solomon DA, et al. BMB Rep (2014) PMID: 24856830
214. Bauerschmidt C, et al. Nucleic Acids Res. (2010) PMID: 19906707
215. Yun J, et al. Nucleic Acids Res. (2016) PMID: 26420833
216. Gelot C, et al. Nucleus (2016) PMID: 27326661
217. Yan M, et al. Breast Cancer Res. (2012) PMID: 22537934
218. Sofueva S, et al. EMBO J. (2013) PMID: 24185899
219. Deng Z, et al. EMBO J. (2012) PMID: 23010778
220. Yun J, et al. EMBO Rep. (2016) PMID: 27466323
221. Mahmood SF, et al. Carcinogenesis (2014) PMID: 24148822
222. Diamond EL, et al. Cancer Discov (2016) PMID: 26566875
223. Diamond EL, et al. Nature (2019) PMID: 30867592
224. Jacobsen E, et al. N. Engl. J. Med. (2017) PMID: 29236635
225. Berce PC, et al. JAAD Case Rep (2022) PMID: 35372654
226. Giuffrè C, et al. Indian J Ophthalmol (2020) PMID: 32823478
227. Razanamahery J, et al. Clin Exp Rheumatol () PMID: 31376269
228. Moyon Q, et al. Chest (2020) PMID: 31669429
229. Gupta RK, et al. Neurol Int (2022) PMID: 36135991
230. Rosen LS, et al. Invest New Drugs (2016) PMID: 27424159
231. Cho et al., 2020; AACR Abstract CT201
232. Touat et al., 2019; DOI: 10.1200/PO.18.00298
233. Schalkwijk S, et al. Cancer Chemother Pharmacol (2021) PMID: 33903938
234. Glassberg et al., 2020; ASPHO Abstract 2015
235. Coyne et al., 2020; ASCO Abstract 3612
236. Dombi E, et al. N. Engl. J. Med. (2016) PMID: 28029918
237. Fangusaro J, et al. Lancet Oncol. (2019) PMID: 31151904
238. Banerjee A, et al. Neuro-oncology (2017) PMID: 28339824
239. Gupta A, et al. Ann. Oncol. (2014) PMID: 24567366
240. Robert C, et al. Lancet Oncol. (2013) PMID: 23735514
241. Kirkwood JM, et al. Clin. Cancer Res. (2012) PMID: 22048237
242. Banerji U, et al. Clin. Cancer Res. (2010) PMID: 20179232
243. Boers-Sonderen MJ, et al. Anticancer Drugs (2012) PMID: 22293660
244. Lopez-Chavez A, et al. J. Clin. Oncol. (2015) PMID: 25667274
245. Hainsworth JD, et al. J Thorac Oncol (2010) PMID: 20802351
246. Middleton G, et al. Nature (2020) PMID: 32669708
247. Coleman RL, et al. Gynecol. Oncol. (2015) PMID: 25887099
248. Eckstein OS, et al. J Clin Oncol (2022) PMID: 35363510
249. Deming DA, et al. Invest New Drugs (2016) PMID: 26666244
250. Krishnamurthy A, et al. Cancer Res. (2018) PMID: 30042150
251. Infante JR, et al. Invest New Drugs (2017) PMID: 28424891
252. LoRusso PM, et al. BMC Cancer (2017) PMID: 28264648
253. Tolcher AW, et al. Clin. Cancer Res. (2015) PMID: 25516890
254. Wilky BA, et al. Br. J. Cancer (2015) PMID: 25268371
255. Infante JR, et al. Lancet Oncol. (2012) PMID: 22805291
256. Leijen S, et al. Clin. Cancer Res. (2012) PMID: 22767668
257. Zimmer L, et al. Clin. Cancer Res. (2014) PMID: 24947927
258. Infante JR, et al. Eur. J. Cancer (2013) PMID: 23583440
259. Juric et al., 2014; ASCO Abstract 9051
260. Kurata et al., 2013; ASCO Abstract e20004
261. Tolcher AW, et al. Ann. Oncol. (2015) PMID: 25344362
262. Patterson et al., 2018; AACR Abstract 3891

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