

**ABOUT THE TEST** FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT	DISEASE	Brain glioblastoma (GBM)	PHYSICIAN	ORDERING PHYSICIAN	Yeh, Yi-Chen	SPECIMEN	SPECIMEN SITE	Brain
	NAME	Tang, Li-Na		MEDICAL FACILITY	Taipei Veterans General Hospital		SPECIMEN ID	S111-037168 (PF22017)
	DATE OF BIRTH	30 May 1951		ADDITIONAL RECIPIENT	None		SPECIMEN TYPE	Slide Deck
	SEX	Female		MEDICAL FACILITY ID	205872		DATE OF COLLECTION	24 January 2022
	MEDICAL RECORD #	48114303		PATHOLOGIST	Not Provided		SPECIMEN RECEIVED	15 February 2022

## Biomarker Findings

**Tumor Mutational Burden** - 11 Muts/Mb  
**Microsatellite status** - MS-Stable

## Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

**FGFR1** K656E

**CCND2** amplification - equivocal<sup>†</sup>

**CDK4** amplification

**MDM2** amplification

**3 Disease relevant genes with no reportable alterations: EGFR, IDH1, PDGFRA**

<sup>†</sup> See About the Test in appendix for details.

## Report Highlights

- Targeted therapies with potential clinical benefit **approved in another tumor type: Infigratinib** (p. 5)
- Evidence-matched **clinical trial options** based on this patient's genomic findings: (p. 6)

### BIOMARKER FINDINGS

**Tumor Mutational Burden** - 11 Muts/Mb

2 Trials see p. 6

**Microsatellite status** - MS-Stable

### GENOMIC FINDINGS

**FGFR1** - K656E

10 Trials see p. 11

**CCND2** - amplification - equivocal

10 Trials see p. 7

**CDK4** - amplification

10 Trials see p. 9

**MDM2** - amplification

6 Trials see p. 13

#### THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)

none

#### THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)

none

No therapies or clinical trials. see Biomarker Findings section

#### THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)

none

#### THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)

Infigratinib

none

none

none

none

none

none

**NOTE** Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Matthew Hiemenz, M.D. | 22 February 2022  
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1301523-01

## BIOMARKER FINDINGS

## BIOMARKER

# Tumor Mutational Burden

## RESULT

11 Muts/Mb

## POTENTIAL TREATMENT STRATEGIES

## — Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1<sup>1-3</sup>, anti-PD-1 therapies<sup>1-4</sup>, and combination nivolumab and ipilimumab<sup>5-10</sup>. In glioma, a lack of association between TMB and clinical benefit from immune checkpoint inhibitors has been reported<sup>11-12</sup>. However, multiple case studies have reported that patients with ultramutated gliomas driven by POLE mutations have benefited from treatment with anti-PD-1<sup>13-14</sup> or anti-PD-L1<sup>15</sup> therapies. Therefore, although increased TMB alone may not be a

strong biomarker for PD-1 or PD-L1 inhibitors in this cancer type, these agents may have efficacy for patients with glioma harboring both high TMB and POLE mutation.

## FREQUENCY & PROGNOSIS

Glioblastoma (GBM) harbors a median TMB of 2.7 mutations per megabase (mut/Mb), and 4.2% of cases have high TMB (>20 mut/Mb)<sup>16</sup>. For pediatric patients, high TMB has been reported in a subset of high-grade gliomas, frequently in association with mutations in mismatch repair or proofreading genes and in TP53, whereas BRAF alterations or other oncogene fusions were observed more frequently in brain tumors harboring low TMB<sup>17-18</sup>. Increased TMB has been reported to correlate with higher tumor grade in glioma<sup>19</sup> and glioblastoma (GBM) tissue samples with biallelic mismatch repair deficiency (bMMRD)<sup>13</sup>, as well as with shorter OS of patients with diffuse glioma<sup>20</sup>.

## FINDING SUMMARY

Tumor mutation burden (TMB, also known as

mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma<sup>21-22</sup> and cigarette smoke in lung cancer<sup>23-24</sup>, treatment with temozolomide-based chemotherapy in glioma<sup>25-26</sup>, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes<sup>27-31</sup>, and microsatellite instability (MSI)<sup>27,30-31</sup>. This sample harbors a TMB below levels that would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents, for patients with glioma<sup>11-15</sup>. Although efficacy of immune checkpoint inhibitors has been observed for patients with other solid tumor types harboring TMB levels such as seen here<sup>1-4,32</sup>, an association between TMB and clinical benefit has generally not been observed for patients with glioma<sup>11-12</sup>, except for those with ultramutated glioma with POLE mutation<sup>13-15</sup>.

## BIOMARKER

# Microsatellite status

## RESULT

MS-Stable

## POTENTIAL TREATMENT STRATEGIES

## — Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors<sup>33-35</sup>, including approved therapies nivolumab and pembrolizumab<sup>36</sup>. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were

MSI-H and experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%,  $p=0.001$ )<sup>37</sup>.

## FREQUENCY & PROGNOSIS

Low-level MSI has been reported in 5-9% of glioblastoma (GBM) samples<sup>38-40</sup>. A large-scale study did not find high-level microsatellite instability (MSI-H) in any of 129 GBM samples<sup>38</sup>, although a small-scale study reported MSI-H in 4 of 15 pediatric GBMs and 1 of 12 adult GBMs<sup>41</sup>. The frequency of MSI has been reported to be increased in relapsed compared to primary GBM<sup>38</sup>, in GBMs with a previous lower grade astrocytoma<sup>39</sup>, and in giant cell GBM compared to classic GBM<sup>40</sup>.

## FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor<sup>42</sup>. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2<sup>42-44</sup>. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers<sup>45-47</sup>. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins<sup>42,44,46-47</sup>.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Matthew Hiemenz, M.D. | 22 February 2022  
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1301523-01

## GENOMIC FINDINGS

## GENE

# FGFR1

## ALTERATION

K656E

## TRANSCRIPT ID

NM\_023110

## CODING SEQUENCE EFFECT

1966A&gt;G

## VARIANT ALLELE FREQUENCY (% VAF)

90.1%

reported in patients with primary brain tumors<sup>55,57</sup> and lung squamous cell carcinoma<sup>63</sup> treated with FGFR inhibitors. In a phase 1 study of futibatinib, 2 patients with FGFR1-mutated primary brain tumors exhibited PRs<sup>57</sup>. A patient with FGFR1-mutated glioblastoma exhibited a PR when treated with infigratinib<sup>64</sup>. For pediatric patients with FGFR1-mutated gliomas, a case series reported 1 sustained PR for a patient with high grade glioma, and a sustained SD and 1 PD for patients with low grade gliomas following treatment with Debio 1347<sup>55</sup>.

astrocytomas<sup>68</sup>. Mutations in the FGFR1 kinase domain have been reported in both lower-grade gliomas and glioblastomas; one of these mutations has been described as an oncogenic mutation that disrupted autophosphorylation<sup>69-71</sup>. FGFR fusions were identified in 3/85 IDH1 and IDH2 wild-type gliomas, but were not found in any of 126 IDH1- or IDH2-mutant gliomas<sup>72</sup>. Three patients with FGFR1-mutated pilocytic astrocytomas experienced relatively short survival in one study<sup>73</sup>, although published data investigating the prognostic implications of FGFR1 alterations in gliomas are limited (PubMed, Mar 2021)<sup>67</sup>.

## POTENTIAL TREATMENT STRATEGIES

### — Targeted Therapies —

Alterations that activate FGFR1 may predict sensitivity to selective FGFR inhibitors including erdafitinib<sup>48-50</sup>, pemigatinib<sup>51</sup>, infigratinib<sup>52-53</sup>, rogaratinib<sup>54</sup>, Debio 1347<sup>55-56</sup>, futibatinib<sup>57</sup>, and derazantinib<sup>58</sup>, or multikinase inhibitors such as pazopanib<sup>59</sup> and ponatinib<sup>60-62</sup>. In the context of FGFR1 mutation, clinical responses have been

## FREQUENCY & PROGNOSIS

In the Brain Lower Grade Glioma TCGA dataset and the Glioblastoma Multiforme TCGA dataset, mutation of FGFR1 has been found in less than 1% of cases<sup>65-66</sup>. In pediatric patients, FGFR1 alterations have been identified in 18% of low-grade gliomas<sup>18</sup>, including 5/9 pilomyxoid astrocytomas, 8% of high-grade gliomas<sup>18</sup>, and in 6% (4/64) of thalamic gliomas<sup>67</sup>. FGFR1 mutation has also been reported in 5% (5/96) of pilocytic

## FINDING SUMMARY

FGFR1 encodes the protein fibroblast growth factor receptor 1, which plays key roles in regulation of the cell cycle and angiogenesis and is an upstream regulator of the RAS, MAPK, and AKT signaling pathways<sup>74</sup>. The FGFR1 alteration observed here has been characterized as activating and is predicted to be oncogenic<sup>70,75-77</sup>.

## GENE

# CCND2

## ALTERATION

amplification - equivocal

cancer<sup>80-81</sup>.

## FREQUENCY & PROGNOSIS

In the TCGA dataset, CCND2 amplification was observed in 3% of glioblastoma cases<sup>65</sup> and 7% of lower grade glioma cases<sup>66</sup>. CCND2 amplification has been reported in 3% of primary malignant gliomas in one study, with amplification occurring in one anaplastic astrocytoma and two glioblastoma cases<sup>82</sup>. CCND2 mRNA expression has been reported to be increased in higher grade (3 and 4) astrocytoma tumors as compared to lower grade tumors<sup>83</sup>. Cyclin D2 has been reported to be the main cyclin expressed in glioblastoma stem cells (GSCs) but was barely detectable in differentiated glioblastoma cells<sup>84</sup>. Cyclin D2, in complex with CDK4/6, has been

reported to be involved in the cell cycle progression of undifferentiated GSCs, but not differentiated GSCs, and to be involved in their tumorigenicity<sup>84</sup>. High CCND2 nuclear expression at the time of initial surgery for patients with glioblastoma was reported to significantly associate with early mortality in a multivariate analysis of 72 patients<sup>85</sup>.

## FINDING SUMMARY

CCND2 encodes the protein cyclin D2, which binds and regulates the cyclin-dependent kinases that control cell cycle progression, and is a downstream target of cancer signaling pathways including hedgehog and PI3K<sup>86-87</sup>. CCND2 has been reported to be amplified in cancer<sup>88</sup>, and may be biologically relevant in this context<sup>89-90</sup>.

## POTENTIAL TREATMENT STRATEGIES

### — Targeted Therapies —

Although preclinical studies suggest that cyclin D2 activates CDK4/6<sup>78-79</sup>, it is unknown whether CCND2 amplification or activating mutation predicts response to CDK4/6 inhibitors such as abemaciclib, palbociclib, and ribociclib. Clinical studies of CDK4/6 inhibitors have shown the most promise for estrogen receptor-positive breast

ORDERED TEST # ORD-1301523-01

## GENOMIC FINDINGS

## GENE

## CDK4

## ALTERATION

amplification

### POTENTIAL TREATMENT STRATEGIES

#### — Targeted Therapies —

CDK4 amplification or activation may predict sensitivity to CDK4/6 inhibitors such as abemaciclib, palbociclib, and ribociclib<sup>91-94</sup>. Clinical benefit has been reported for limited tumor types including patients with

CDK4-amplified liposarcoma and sarcoma in response to treatment with abemaciclib<sup>95</sup>, palbociclib<sup>91,96</sup>, and ribociclib<sup>97</sup>.

### FREQUENCY & PROGNOSIS

Across TCGA and MKSCC studies, CDK4 amplification has been reported in 4.0-9.4% of glioma cases and 14% of glioblastoma multiforme cases (cBioPortal, Sep 2021)<sup>65,88,98-100</sup>. A study has reported amplification of the 12q14-15 region, where CDK4 and MDM2 reside, in 5% (2/42) of glioblastomas<sup>101</sup>. Amplification of CDK4 and corresponding increased CDK4 protein expression has been reported to be associated with a poorer patient outcome in anaplastic astrocytoma and

glioblastoma<sup>102-105</sup>.

### FINDING SUMMARY

CDK4 encodes the cyclin-dependent kinase 4, which regulates the cell cycle, senescence, and apoptosis<sup>106</sup>. CDK4 and its functional homolog CDK6 are activated by D-type cyclins and promote cell cycle progression by inactivating the tumor suppressor Rb<sup>107-108</sup>. Amplification of the chromosomal region that includes CDK4 has been reported in multiple cancer types, including lung cancer, glioblastoma, and liposarcoma, and has been associated with overexpression of CDK4 protein<sup>91,109-115</sup>.

## GENE

## MDM2

## ALTERATION

amplification

### POTENTIAL TREATMENT STRATEGIES

#### — Targeted Therapies —

MDM2 antagonists disrupt the MDM2-p53 interaction, thereby stabilizing p53<sup>116</sup>. Preclinical studies have suggested that the amplification of MDM2, in the absence of concurrent TP53 mutations, may increase sensitivity to these agents<sup>117-118</sup>. Preliminary Phase 1 studies of the MDM2-p53 antagonist alrizomadlin (APG-115) reported a PR in a patient with liposarcoma harboring an MDM2 amplification and wildtype for TP53 and SD in 21%-38% (6/28 and 5/13, respectively) of patients in genomically unselected solid tumors<sup>119-120</sup>. A Phase 2 trial of alrizomadlin in combination with pembrolizumab reported a PR in 1 of 3 patients with malignant peripheral nerve sheath tumor that had failed standard therapy, as well as PRs in patients with multiple

types of solid tumors that had failed immunotherapy, including 1 out of 14 patients with non-small cell lung cancer; 1 out of 5 patients with urothelial carcinoma; and 2 out of 5, 1 out of 5, and 1 out of 11 patients with mucosal, uveal, and cutaneous melanoma, respectively<sup>121</sup>. Phase 1b studies of the MDM2 inhibitor idasanutlin for refractory AML in combination with cytarabine or venetoclax reported anti-leukemic response rates of 33% (25/75) and 37% (11/30), respectively<sup>122-123</sup>; clinical benefit (58% ORR, 7/12) with idasanutlin monotherapy has been reported for patients with polycythemia vera<sup>124</sup>. The dual MDM2/MDM4 inhibitor ALRN-6924 led to an ORR of 27% (4/15) for patients with TP53 wildtype peripheral T-cell lymphoma in a Phase 2 study<sup>125</sup>; responses have also been observed in TP53 wildtype AML, MDS, Merkel cell carcinoma, colorectal cancer, and liposarcoma<sup>126-127</sup>.

### FREQUENCY & PROGNOSIS

In the Glioblastoma Multiforme (GBM) TCGA dataset, amplification of MDM2 has been found in 8% of cases<sup>65</sup>. A study has reported amplification of the 12q14-15 region, where MDM2 and CDK4 reside, in 5% (2/42) of GBMs<sup>101</sup>. Amplification of

MDM2 has been associated with poor survival in patients with glioblastoma<sup>101,128</sup>.

### FINDING SUMMARY

MDM2 encodes an E3 ubiquitin protein ligase, which mediates the ubiquitination and subsequent degradation of p53, Rb1, and other proteins<sup>129-131</sup>. MDM2 acts to prevent the activity of the tumor suppressor p53; therefore, overexpression or amplification of MDM2 may be oncogenic<sup>132-133</sup>. Overexpression or amplification of MDM2 is frequent in cancer<sup>90</sup>. Although two retrospective clinical studies suggest that MDM2 amplification may predict a short time-to-treatment failure on anti-PD-1/PD-L1 immune checkpoint inhibitors, with 4/5 patients with MDM2 amplification<sup>134</sup> and 2/3 patients with MDM2 or MDM4 amplification<sup>135</sup> experiencing tumor hyperprogression, amplification of MDM2 or MDM4 was not associated with shorter progression-free survival (PFS) in a retrospective analysis of non-small cell lung cancer (NSCLC) outcomes with immune checkpoint inhibitors (hazard ratio of 1.4, p=0.44)<sup>136</sup>. The latter study reported PFS of >2 months for 5/8 patients with MDM2/MDM4 amplification<sup>136</sup>.

ORDERED TEST # ORD-1301523-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

## Infigratinib

Assay findings association

**FGFR1**  
K656E

### AREAS OF THERAPEUTIC USE

Infigratinib is a TKI that inhibits FGFR1, FGFR2, and FGFR3. It is FDA approved for the treatment of patients with unresectable locally advanced or metastatic cholangiocarcinoma who have FGFR2 rearrangements or fusions and have progressed after prior therapy. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

Based on individual responses in patients with FGFR1-mutated glioblastoma<sup>64</sup> and lung squamous cell carcinoma<sup>63</sup>, FGFR1 mutation may predict sensitivity to

infigratinib.

### SUPPORTING DATA

A Phase 2 study of infigratinib for patients with recurrent high-grade gliomas harboring FGFR alterations, reported a 9.5% (2/21) ORR, 1.7 month median PFS, and 6.7 month median OS<sup>64</sup>. Disease control greater than one year was observed in 4 patients, including a PR in a patient with FGFR1-mutated glioma, and SD in patients with glioma harboring FGFR1 mutation, FGFR3 mutation, or FGFR3-TACC3 fusion<sup>64</sup>.

**NOTE** Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

**ORDERED TEST #** ORD-1301523-01

**CLINICAL TRIALS**

**NOTE** Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://www.foundationmedicine.com/genomic-testing#support-services). Or visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

**BIOMARKER**

## Tumor Mutational Burden

**RESULT**

11 Muts/Mb

**RATIONALE**

Increased tumor mutational burden may predict response to anti-PD-1 (alone or in combination

with anti-CTLA-4) or anti-PD-L1 immune checkpoint inhibitors.

**NCT04977453**
**PHASE 1/2**

GI-101 as a Single Agent or in Combination With Pembrolizumab, Lenvatinib or Local Radiotherapy in Advanced Solid Tumors

**TARGETS**

FGFRs, RET, PDGFRA, VEGFRs, KIT, PD-1, CTLA-4

**LOCATIONS:** Suwon-si (Korea, Republic of), Seoul (Korea, Republic of)

**NCT04773951**
**PHASE 1**

A Phase I Study to Evaluate the Safety, Tolerability and Pharmacokinetics of JS004 in Advanced Solid Tumors

**TARGETS**

BTLA, PD-1

**LOCATIONS:** Beijing (China)

ORDERED TEST # ORD-1301523-01

CLINICAL TRIALS

 GENE  
**CCND2**

 RATIONALE  
CCND2 amplification or activation may predict sensitivity to CDK4/6 inhibitors.

 ALTERATION  
amplification - equivocal

**NCT04282031**

PHASE 1/2

A Study of BPI-1178 in Patients With Advanced Solid Tumor and HR+/HER2- Breast Cancer

 TARGETS  
CDK6, CDK4, ER, Aromatase

LOCATIONS: Shanghai (China)

**NCT04594005**

PHASE 1/2

CDK4/6 Tumor, Abemaciclib, Paclitaxel

 TARGETS  
CDK4, CDK6

LOCATIONS: Seoul (Korea, Republic of)

**NCT04391595**

PHASE NULL

LY3214996 Plus Abemaciclib in Recurrent Glioblastoma Patients

 TARGETS  
CDK4, CDK6, ERK1, ERK2

LOCATIONS: Arizona

**NCT03834740**

PHASE NULL

Ph0/2 Ribociclib &amp; Everolimus

 TARGETS  
CDK6, CDK4, mTOR

LOCATIONS: Arizona

**NCT02933736**

PHASE NULL

Ribociclib (LEE011) in Preoperative Glioma and Meningioma Patients

 TARGETS  
CDK6, CDK4

LOCATIONS: Arizona

**NCT04801966**

PHASE NULL

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

 TARGETS  
CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF

LOCATIONS: Melbourne (Australia)



ORDERED TEST # ORD-1301523-01

CLINICAL TRIALS

**NCT03994796**
**PHASE 2**

Genetic Testing in Guiding Treatment for Patients With Brain Metastases

**TARGETS**

TRKB, ALK, TRKC, ROS1, TRKA, CDK4, CDK6, PI3K, mTOR

**LOCATIONS:** Washington, Oregon, Idaho, Montana

**NCT03310879**
**PHASE 2**

Study of the CDK4/6 Inhibitor Abemaciclib in Solid Tumors Harboring Genetic Alterations in Genes Encoding D-type Cyclins or Amplification of CDK4 or CDK6

**TARGETS**

CDK4, CDK6

**LOCATIONS:** Massachusetts

**NCT03158389**
**PHASE 1/2**

 NCT Neuro Master Match - N<sup>2</sup>M<sup>2</sup> (NOA-20)

**TARGETS**

ALK, RET, CDK4, CDK6, mTOR, MDM2, PD-L1, SMO

**LOCATIONS:** Berlin (Germany), Dresden (Germany), Regensburg (Germany), Bochum (Germany), Frankfurt am Main (Germany), Essen (Germany), Mainz (Germany), Heidelberg (Germany), Cologne (Germany), Mannheim (Germany)

**NCT02981940**
**PHASE 2**

A Study of Abemaciclib in Recurrent Glioblastoma

**TARGETS**

CDK4, CDK6

**LOCATIONS:** Utah, California, Massachusetts



ORDERED TEST # ORD-1301523-01

CLINICAL TRIALS

**GENE**  
**CDK4**
**RATIONALE**  
CDK4 amplification may predict sensitivity to CDK4/6 inhibitors.

**ALTERATION**  
amplification

**NCT04282031**
**PHASE 1/2**

A Study of BPI-1178 in Patients With Advanced Solid Tumor and HR+/HER2- Breast Cancer

**TARGETS**  
CDK6, CDK4, ER, Aromatase

**LOCATIONS:** Shanghai (China)

**NCT03239015**
**PHASE 2**

Efficacy and Safety of Targeted Precision Therapy in Refractory Tumor With Druggable Molecular Event

**TARGETS**  
EGFR, ERBB4, ERBB2, PARP, mTOR, MET, ROS1, RET, VEGFRs, BRAF, CDK4, CDK6

**LOCATIONS:** Shanghai (China)

**NCT04594005**
**PHASE 1/2**

CDK4/6 Tumor, Abemaciclib, Paclitaxel

**TARGETS**  
CDK4, CDK6

**LOCATIONS:** Seoul (Korea, Republic of)

**NCT04391595**
**PHASE NULL**

LY3214996 Plus Abemaciclib in Recurrent Glioblastoma Patients

**TARGETS**  
CDK4, CDK6, ERK1, ERK2

**LOCATIONS:** Arizona

**NCT03834740**
**PHASE NULL**

Ph0/2 Ribociclib &amp; Everolimus

**TARGETS**  
CDK6, CDK4, mTOR

**LOCATIONS:** Arizona

**NCT02933736**
**PHASE NULL**

Ribociclib (LEE011) in Preoperative Glioma and Meningioma Patients

**TARGETS**  
CDK6, CDK4

**LOCATIONS:** Arizona

ORDERED TEST # ORD-1301523-01

CLINICAL TRIALS

**NCT04801966**
**PHASE NULL**

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

**TARGETS**  
CDK4, CDK6, PI3K-alpha, PD-L1, MEK,  
PARP, PD-1, BRAF

**LOCATIONS:** Melbourne (Australia)

**NCT03994796**
**PHASE 2**

Genetic Testing in Guiding Treatment for Patients With Brain Metastases

**TARGETS**  
TRKB, ALK, TRKC, ROS1, TRKA, CDK4,  
CDK6, PI3K, mTOR

**LOCATIONS:** Washington, Oregon, Idaho, Montana

**NCT03310879**
**PHASE 2**

Study of the CDK4/6 Inhibitor Abemaciclib in Solid Tumors Harboring Genetic Alterations in Genes Encoding D-type Cyclins or Amplification of CDK4 or CDK6

**TARGETS**  
CDK4, CDK6

**LOCATIONS:** Massachusetts

**NCT03158389**
**PHASE 1/2**

 NCT Neuro Master Match - N<sup>2</sup>M<sup>2</sup> (NOA-20)

**TARGETS**  
ALK, RET, CDK4, CDK6, mTOR,  
MDM2, PD-L1, SMO

**LOCATIONS:** Berlin (Germany), Dresden (Germany), Regensburg (Germany), Bochum (Germany), Frankfurt am Main (Germany), Essen (Germany), Mainz (Germany), Heidelberg (Germany), Cologne (Germany), Mannheim (Germany)

ORDERED TEST # ORD-1301523-01

CLINICAL TRIALS

**GENE**  
**FGFR1**
**RATIONALE**  
FGFR inhibitors may be relevant in tumors with alterations that activate FGFR1.

**ALTERATION**  
K656E

**NCT04169672**
**PHASE 2**

Study of Surufatinib Combined With Toripalimab in Patients With Advanced Solid Tumors

**TARGETS**  
FGFR1, CSF1R, VEGFRs, PD-1

**LOCATIONS:** Shanghai (China), Beijing (China)

**NCT04803318**
**PHASE 2**

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

**TARGETS**  
mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK

**LOCATIONS:** Guangzhou (China)

**NCT04977453**
**PHASE 1/2**

GI-101 as a Single Agent or in Combination With Pembrolizumab, Lenvatinib or Local Radiotherapy in Advanced Solid Tumors

**TARGETS**  
FGFRs, RET, PDGFRA, VEGFRs, KIT, PD-1, CTLA-4

**LOCATIONS:** Suwon-si (Korea, Republic of), Seoul (Korea, Republic of)

**NCT03564691**
**PHASE 1**

Study of MK-4830 as Monotherapy and in Combination With Pembrolizumab (MK-3475) in Participants With Advanced Solid Tumors (MK-4830-001)

**TARGETS**  
ITL4, FGFRs, RET, PDGFRA, VEGFRs, KIT, PD-1

**LOCATIONS:** Seoul (Korea, Republic of), Haifa (Israel), Petah Tikva (Israel), Ramat Gan (Israel), Tel Aviv (Israel), Warszawa (Poland), Gdansk (Poland), Heraklion (Greece), Washington, Hospitalet de Llobregat (Spain)

**NCT03547037**
**PHASE 1**

A Study to Evaluate the Safety, Pharmacokinetics, Pharmacodynamics, and Immunogenicity of JNJ-63723283, an Anti-Programmed Cell Death (PD)-1 Monoclonal Antibody, as Monotherapy or in Combination With Erdafitinib in Japanese Participants With Advanced Solid Cancers

**TARGETS**  
PD-1, FGFRs

**LOCATIONS:** Chuo-Ku (Japan), Kashiwa (Japan)

**NCT04008797**
**PHASE 1**

A Study of E7386 in Combination With Other Anticancer Drug in Participants With Solid Tumor

**TARGETS**  
CBP, Beta-catenin, FGFRs, RET, PDGFRA, VEGFRs, KIT

**LOCATIONS:** Osakasayama (Japan), Chuo-Ku (Japan), Kashiwa (Japan)

© 2022 Foundation Medicine, Inc. All rights reserved.

 Electronically signed by Matthew Hiemenz, M.D. | 22 February 2022  
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

 Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1301523-01

CLINICAL TRIALS

**NCT04424966**

PHASE NULL

Infigratinib in Recurrent Glioblastoma Patients

**TARGETS**  
FGFR3, FGFR1, FGFR2

LOCATIONS: Arizona

**NCT04565275**

PHASE 1/2

A Study of ICP-192 in Patients With Advanced Solid Tumors

**TARGETS**  
FGFR2, FGFR1, FGFR3, FGFR4

LOCATIONS: Macquarie Park (Australia), Melbourne (Australia), Clayton (Australia), Frankston (Australia), Colorado, Minnesota, Arizona, Ohio, Florida

**NCT02549937**

PHASE 1/2

A Multi-Center, Open-Label Study of Sulfatinib(HMPL-012) in Patients With Advanced Solid Tumors

**TARGETS**  
FGFR1, CSF1R, VEGFRs

LOCATIONS: Milano (Italy), California, Colorado, Texas, New York, Tennessee, Virginia, Florida

**NCT04729348**

PHASE 2

Pembrolizumab And Lenvatinib In Leptomeningeal Metastases

**TARGETS**  
PD-1, KIT, VEGFRs, FGFRs, PDGFRA, RET

LOCATIONS: Massachusetts

ORDERED TEST # ORD-1301523-01

CLINICAL TRIALS

**GENE**  
**MDM2**

**RATIONALE**  
Inhibitors of the MDM2-p53 interaction are being tested in clinical trials. Overexpression or amplification of MDM2 may increase sensitivity to these agents, but more data are required.

**ALTERATION**  
amplification

### NCT04589845

#### PHASE 2

Tumor-Agnostic Precision Immuno-Oncology and Somatic Targeting Rational for You (TAPISTRY) Platform Study

**TARGETS**  
TRKB, ALK, TRKC, ROS1, TRKA, RET, PD-L1, AKTs, ERBB2, MDM2, PI3K-alpha

**LOCATIONS:** Zhongzheng Dist. (Taiwan), Taipei City (Taiwan), Taoyuan County (Taiwan), Tainan (Taiwan), Hong Kong (Hong Kong), Seoul (Korea, Republic of), Xi'an (China), Tianjin (China), Beijing (China), Chengdu City (China)

### NCT03449381

#### PHASE 1

This Study Aims to Find the Best Dose of BI 907828 in Patients With Different Types of Advanced Cancer (Solid Tumors)

**TARGETS**  
MDM2

**LOCATIONS:** Tokyo, Chuo-ku (Japan), Berlin (Germany), Tübingen (Germany), Leuven (Belgium), Barcelona (Spain), Ottawa (Canada), Connecticut, New York, Tennessee, Florida

### NCT03611868

#### PHASE 1/2

A Study of APG-115 in Combination With Pembrolizumab in Patients With Metastatic Melanomas or Advanced Solid Tumors

**TARGETS**  
MDM2, PD-1

**LOCATIONS:** Brisbane (Australia), South Brisbane (Australia), Bedford Park (Australia), California, Arizona, Missouri, Arkansas, Pennsylvania, New York

### NCT03158389

#### PHASE 1/2

NCT Neuro Master Match - N<sup>2</sup>M<sup>2</sup> (NOA-20)

**TARGETS**  
ALK, RET, CDK4, CDK6, mTOR, MDM2, PD-L1, SMO

**LOCATIONS:** Berlin (Germany), Dresden (Germany), Regensburg (Germany), Bochum (Germany), Frankfurt am Main (Germany), Essen (Germany), Mainz (Germany), Heidelberg (Germany), Cologne (Germany), Mannheim (Germany)

### NCT03107780

#### PHASE 1

MDM2 Inhibitor AMG-232 in Treating Patients With Recurrent or Newly Diagnosed Glioblastoma

**TARGETS**  
MDM2

**LOCATIONS:** California, Michigan, Pennsylvania, Massachusetts, New York, Maryland, North Carolina, Alabama

### NCT03725436

#### PHASE 1

ALRN-6924 and Paclitaxel in Treating Patients With Advanced, Metastatic, or Unresectable Solid Tumors

**TARGETS**  
MDM2, MDM4

**LOCATIONS:** Texas

ORDERED TEST # ORD-1301523-01

**APPENDIX**
**Variants of Unknown Significance**

**NOTE** One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

**ATRX**  
E1464del

**AXL**  
R490C

**BRCA1**  
N909I

**BRCA2**  
N108S

**CSF1R**  
V229I

**EP300**  
M755I

**GATA3**  
S110F

**MAP3K13**  
A604T

**MLL2**  
N2965S

**MPL**  
P70L

**MST1R**  
V670G

**PARP1**  
S504P

**RAD52**  
amplification

**ROS1**  
A174E

**TBX3**  
E401K

**TEK**  
amplification

**TSC2**  
E1490G

ORDERED TEST # ORD-1301523-01

**APPENDIX**
**Genes Assayed in FoundationOne®CDx**

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

**DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS**

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTB	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL	CCND1
CCND2	CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2	DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FAM46C	FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3
FGFR4	FH	FLCN	FLT1	FLT3	FOXO2	FUBP1	GABRA6	GATA3
GATA4	GATA6	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3F3A
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDMSA	KDMS5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKB1A	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3	P2RY8	PALB2
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA
PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET
RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2
SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOC3
SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU
SYK	TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53
TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1	WT1	XPO1
XRCC2	ZNF217	ZNF703						

**DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS**

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSP02	SDC4	SLC34A2	TERC*	TERT**	TPRSS2

\*TERC is an NCRNA

\*\*Promoter region of TERT is interrogated

**ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS**

Loss of Heterozygosity (LOH) score

Microsatellite (MS) status

Tumor Mutational Burden (TMB)



ORDERED TEST # ORD-1301523-01

**APPENDIX**
**About FoundationOne®CDx**

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.


**ABOUT FOUNDATIONONE CDx**

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:  
[www.rochefoundationmedicine.com/f1cdxtech](http://www.rochefoundationmedicine.com/f1cdxtech).

**INTENDED USE**

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

**TEST PRINCIPLES**

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

**THE REPORT**

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

**Diagnostic Significance**

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

**Qualified Alteration Calls (Equivocal and Subclonal)**

An alteration denoted as "amplification – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

**Ranking of Therapies and Clinical Trials**
**Ranking of Therapies in Summary Table**

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

**Ranking of Clinical Trials**

Pediatric trial qualification → Geographical proximity → Later trial phase.

**NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION**

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) ([www.nccn.org](http://www.nccn.org)). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to [NCCN.org](http://NCCN.org). NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

**Limitations**

1. In the fractional-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Matthew Hiemenz, M.D. | 22 February 2022  
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1301523-01

## APPENDIX

## About FoundationOne®CDx

- Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.
2. TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation [https://www.accessdata.fda.gov/cdrh\\_docs/pdf17/P170019B.pdf](https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf). The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
  3. The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
  4. Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
  5. Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine

whether the patient is a candidate for biopsy.

6. Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of *HER2* overexpression as 20% in breast cancer. Based on the F1CDx *HER2* CDx concordance study, approximately 10% of *HER2* amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

### REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

### VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

### Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

\*Interquartile Range = 1<sup>st</sup> Quartile to 3<sup>rd</sup> Quartile

### VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

### VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear

ORDERED TEST # ORD-1301523-01

APPENDIX

About FoundationOne®CDx

cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

#### LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

#### NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

#### NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

#### TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

#### SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

MR Suite Version 6.0.0

The median exon coverage for this sample is 872x

ORDERED TEST # **ORD-1301523-01**
**APPENDIX**
**References**

1. Samstein RM, et al. Nat. Genet. (2019) pmid: 30643254
2. Goodman AM, et al. Mol. Cancer Ther. (2017) pmid: 28835386
3. Goodman AM, et al. Cancer Immunol Res (2019) pmid: 31405947
4. Cristescu R, et al. Science (2018) pmid: 30309915
5. Ready N, et al. J. Clin. Oncol. (2019) pmid: 30785829
6. Hellmann MD, et al. N. Engl. J. Med. (2018) pmid: 29658845
7. Hellmann MD, et al. Cancer Cell (2018) pmid: 29657128
8. Hellmann MD, et al. Cancer Cell (2018) pmid: 29731394
9. Rozeman EA, et al. Nat Med (2021) pmid: 33558721
10. Sharma P, et al. Cancer Cell (2020) pmid: 32916128
11. Zhao J, et al. Nat. Med. (2019) pmid: 30742119
12. Touat M, et al. Nature (2020) pmid: 32322066
13. Bouffet E, et al. J. Clin. Oncol. (2016) pmid: 27001570
14. Johanns TM, et al. Cancer Discov (2016) pmid: 27683556
15. Lukas RV, et al. J. Neurooncol. (2018) pmid: 30073642
16. Chalmers ZR, et al. Genome Med (2017) pmid: 28420421
17. Patel RR, et al. Pediatr Blood Cancer (2020) pmid: 32386112
18. Johnson A, et al. Oncologist (2017) pmid: 28912153
19. Draaisma K, et al. Acta Neuropathol Commun (2015) pmid: 26699864
20. Wang L, et al. BMC Cancer (2020) pmid: 32164609
21. Pfeifer GP, et al. Mutat. Res. (2005) pmid: 15748635
22. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) pmid: 23875803
23. Pfeifer GP, et al. Oncogene (2002) pmid: 12379884
24. Rizvi NA, et al. Science (2015) pmid: 25765070
25. Johnson BE, et al. Science (2014) pmid: 24336570
26. Choi S, et al. Neuro-oncology (2018) pmid: 29452419
27. Cancer Genome Atlas Research Network, et al. Nature (2013) pmid: 23636398
28. Briggs S, et al. J. Pathol. (2013) pmid: 23447401
29. Heitzner E, et al. Curr. Opin. Genet. Dev. (2014) pmid: 24583393
30. Nature (2012) pmid: 22810696
31. Roberts SA, et al. Nat. Rev. Cancer (2014) pmid: 25568919
32. Marabelle A, et al. Lancet Oncol. (2020) pmid: 32919526
33. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) pmid: 25392179
34. Kroemer G, et al. Oncoimmunology (2015) pmid: 26140250
35. Lal N, et al. Oncoimmunology (2015) pmid: 25949894
36. Le DT, et al. N. Engl. J. Med. (2015) pmid: 26028255
37. Ayers et al., 2016; ASCO-SITC Abstract P60
38. Martínez R, et al. Oncology (2004) pmid: 15331927
39. Martínez R, et al. J. Cancer Res. Clin. Oncol. (2005) pmid: 15672285
40. Martínez R, et al. Cancer Genet. Cytogenet. (2007) pmid: 17498554
41. Szybka M, et al. Clin. Neuropathol. () pmid: 12908754
42. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) pmid: 26337942
43. You JF, et al. Br. J. Cancer (2010) pmid: 21081928
44. Bairwa NK, et al. Methods Mol. Biol. (2014) pmid: 24623249
45. Boland CR, et al. Cancer Res. (1998) pmid: 9823339
46. Pawlik TM, et al. Dis. Markers (2004) pmid: 15528785
47. Boland CR, et al. Gastroenterology (2010) pmid: 20420947
48. Liorot Y, et al. N. Engl. J. Med. (2019) pmid: 31340094
49. Tabernero J, et al. J. Clin. Oncol. (2015) pmid: 26324363
50. Karkera JD, et al. Mol. Cancer Ther. (2017) pmid: 28416604
51. Necchi et al., 2018; ESMO Abstract 900P
52. Pal SK, et al. Cancer Discov (2018) pmid: 29848605
53. Pal SK, et al. Cancer (2020) pmid: 32208524
54. Schuler M, et al. Lancet Oncol. (2019) pmid: 31405822
55. Farouk Sait S, et al. JCO Precis Oncol (2021) pmid: 34250399
56. Voss MH, et al. Clin. Cancer Res. (2019) pmid: 30745300
57. Bahleda R, et al. Ann Oncol (2020) pmid: 32622884
58. Papadopoulos KP, et al. Br. J. Cancer (2017) pmid: 28972963
59. Cheng FT, et al. J Natl Compr Canc Netw (2017) pmid: 29223982
60. Khodadoust MS, et al. Leukemia (2016) pmid: 26055304
61. Tanasi I, et al. Blood (2019) pmid: 31434701
62. Strati P, et al. Leuk. Lymphoma (2018) pmid: 29119847
63. Slosberg ED, et al. Oncotarget (2018) pmid: 29765547
64. Lassman et al., 2019; SNO Abstract ACTR-33
65. Brennan CW, et al. Cell (2013) pmid: 24120142
66. Cancer Genome Atlas Research Network, et al. N. Engl. J. Med. (2015) pmid: 26061751
67. Ryall S, et al. Acta Neuropathol Commun (2016) pmid: 27577993
68. Jones DT, et al. Nat. Genet. (2013) pmid: 23817572
69. Rand V, et al. Proc. Natl. Acad. Sci. U.S.A. (2005) pmid: 16186508
70. Lew ED, et al. Sci Signal (2009) pmid: 19224897
71. Zhang J, et al. Nat. Genet. (2013) pmid: 23583981
72. Di Stefano AL, et al. Clin. Cancer Res. (2015) pmid: 25609060
73. Becker AP, et al. J. Neuropathol. Exp. Neurol. (2015) pmid: 26083571
74. Turner N, et al. Nat. Rev. Cancer (2010) pmid: 20094046
75. Liu A, et al. Development (2003) pmid: 14602678
76. Petiot A, et al. Dev. Dyn. (2002) pmid: 12112473
77. Hart KC, et al. Oncogene (2000) pmid: 10918587
78. Busk PK, et al. Exp. Cell Res. (2005) pmid: 15707582
79. Busk PK, et al. Cell Cycle () pmid: 12695654
80. Finn RS, et al. Lancet Oncol. (2015) pmid: 25524798
81. DeMichele A, et al. Clin. Cancer Res. (2015) pmid: 25501126
82. Büschges R, et al. Brain Pathol. (1999) pmid: 10416984
83. Kheirollahi M, et al. Med. Oncol. (2011) pmid: 20077038
84. Koyama-Nasu R, et al. Oncogene (2013) pmid: 22964630
85. Bouchart C, et al. Cancer Med (2019) pmid: 31568682
86. Katoh Y, et al. Curr. Mol. Med. (2009) pmid: 19860666
87. White PC, et al. Oncogene (2006) pmid: 16301994
88. Gao J, et al. Sci Signal (2013) pmid: 23550210
89. Zack TI, et al. Nat. Genet. (2013) pmid: 24071852
90. Beroukhir R, et al. Nature (2010) pmid: 20164920
91. Dickson MA, et al. J. Clin. Oncol. (2013) pmid: 23569312
92. Flaherty KT, et al. Clin. Cancer Res. (2012) pmid: 22090362
93. Patnaik A, et al. Cancer Discov (2016) pmid: 27217383
94. Infante JR, et al. Clin. Cancer Res. (2016) pmid: 27542767
95. Dickson et al., 2019; ASCO Abstract 11004
96. Dickson MA, et al. JAMA Oncol (2016) pmid: 27124835
97. Peguero et al., 2016; ASCO Abstract 2528
98. Cerami E, et al. Cancer Discov (2012) pmid: 22588877
99. Jonsson P, et al. Clin. Cancer Res. (2019) pmid: 31263031
100. Ceccarelli M, et al. Cell (2016) pmid: 26824661
101. Zheng S, et al. Genes Dev. (2013) pmid: 23796897
102. Kim H, et al. Proc. Natl. Acad. Sci. U.S.A. (2010) pmid: 20080666
103. Ruano Y, et al. Am. J. Clin. Pathol. (2009) pmid: 19141386
104. Fischer U, et al. Mol. Cancer Res. (2008) pmid: 18403636
105. Bäcklund LM, et al. Br. J. Cancer (2005) pmid: 15970925
106. Choi YJ, et al. Oncogene (2014) pmid: 23644662
107. Cell (1995) pmid: 7736585
108. Musgrave EA, et al. Nat. Rev. Cancer (2011) pmid: 21734724
109. Wikman H, et al. Genes Chromosomes Cancer (2005) pmid: 15543620
110. Rao SK, et al. J. Neurooncol. (2010) pmid: 19609742
111. Chung L, et al. Am. J. Surg. Pathol. (2009) pmid: 19574885
112. Ragazzini P, et al. Histol. Histopathol. (2004) pmid: 15024701
113. Dujardin F, et al. Mod. Pathol. (2011) pmid: 21336260
114. Zhang K, et al. Cancer Res. (2013) pmid: 23393200
115. Horvai AE, et al. Mod. Pathol. (2009) pmid: 19734852
116. Cheok CF, et al. Nat Rev Clin Oncol (2011) pmid: 20975744
117. Ohnstad HO, et al. Cancer (2013) pmid: 23165797
118. Gamble LD, et al. Oncogene (2012) pmid: 21725357
119. Zhang et al., 2019; ASCO Abstract 3124
120. Rasco et al., 2019; ASCO Abstract 3126
121. Tolcher et al., 2021; ASCO Abstract 2506
122. Martinelli et al., 2016; EHA21 Abstract S504
123. Daver et al., 2018; ASH Abstract 767
124. Mascarenhas et al., 2019; ASH Abstract 134
125. Shustov et al., 2018; ASH Abstract 1623
126. Sallman et al., 2018; ASH Abstract 4066
127. Meric-Bernstam et al., 2017; ASCO Abstract 2505
128. Fischer U, et al. Int. J. Cancer (2010) pmid: 19839052
129. Sdek P, et al. Mol. Cell (2005) pmid: 16337594
130. Brady M, et al. Mol. Cell. Biol. (2005) pmid: 15632057
131. Li M, et al. Mol. Cell (2004) pmid: 15053880
132. Brown CJ, et al. Nat. Rev. Cancer (2009) pmid: 19935675
133. Cordon-Cardo C, et al. Cancer Res. (1994) pmid: 8306343
134. Kato S, et al. Clin. Cancer Res. (2017) pmid: 28351930
135. Singavi et al., 2017; ESMO Abstract 1140PD
136. Rizvi H, et al. J. Clin. Oncol. (2018) pmid: 29337640

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Matthew Hiemenz, M.D. | 22 February 2022  
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531