Project ID: C23-M001-01503 Report No.: AA-23-03155_ONC Date Reported: May 30, 2023

ACTOnco® + Report

| PATIENT | | |
|--|----------------------|--|
| Identifier: 黃瓘絜 | Patient ID: 47801401 | |
| Date of Birth: May 08, 1998 | Gender: Female | |
| Diagnosis: Round cell sarcoma | | |
| ORDERING PHYSICIAN | | |
| Name: 顏秀如醫師 | Tel: 886-228712121 | |
| Facility: 臺北榮總 | | |
| Address: 臺北市北投區石牌路二段 201 號 | | |
| SPECIMEN | | |
| Specimen ID: S11217744A Collection site: L-spine | Type: FFPE tissue | |
| Date received: May 18, 2023 Lab ID: AA-23-03155 | D/ID: NA | |

ABOUT ACTORCO®+

The test is a next-generation sequencing (NGS)-based assay developed for efficient and comprehensive genomic profiling of cancers. This test interrogates coding regions of 440 genes associated with cancer treatment, prognosis and diagnosis. Genetic mutations detected by this test include small-scale mutations like single nucleotide variants (SNVs), small insertions and deletions (InDels) (≤ 15 nucleotides) and large-scale genomic alterations like copy number alterations (CNAs). The test also includes an RNA test, detecting fusion transcripts of 13 genes.

SUMMARY FOR ACTIONABLE VARIANTS

VARIANTS/BIOMARKERS WITH EVIDENCE OF CLINICAL SIGNIFICANCE

| Genomic | Probable Effects in F | Probable Sensitive in Other | |
|------------------------|-----------------------|-----------------------------|--|
| Alterations/Biomarkers | Sensitive | Cancer Types | |
| Not detected | | | |

VARIANTS/BIOMARKERS WITH POTENTIAL CLINICAL SIGNIFICANCE

| Genomic Alterations/Biomarkers | Possibly Sensitive | Possibly Resistant |
|--------------------------------|--------------------|--------------------------------------|
| RB1 Splice acceptor | - | Abemaciclib, Palbociclib, Ribociclib |

Note:

- The above summary tables present genomic variants and biomarkers based on the three-tiered approach proposed by US FDA for reporting tumor profiling NGS testing. "Variants/biomarkers with evidence of clinical significance" refers to mutations that are widely recognized as standard-of-care biomarkers (FDA level 2/AMP tier 1). "Variants/biomarkers with potential clinical significance" refers to mutations that are not included in the standard of care but are informational for clinicians, which are commonly biomarkers used as inclusion criterial for clinical trials (FDA level 3/AMP tier 2).
- The therapeutic agents and possible effects to a given drug are based on mapping the variants/biomarkers with ACT Genomics clinical knowledge database. The mapping results only provide information for reference, but not medical recommendation.
- Please refer to corresponding sections for more detailed information about genomic alteration and clinical relevance listed above.





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TESTING RESULTS

VARIANT(S) WITH CLINICAL RELEVANCE

- Single Nucleotide and Small InDel Variants

| Gene | Amino Acid Change | Allele Frequency |
|------|-------------------|------------------|
| RB1 | Splice acceptor | 47.1% |
| TP53 | G245V | 68.7% |

- Copy Number Alterations

| Chromosome Gene | | Variation | Copy Number |
|-----------------|------------|-----------------------|-------------|
| Chr17 | FLCN, TP53 | Heterozygous deletion | 1 |
| Chr17 | RPTOR | Amplification | 6 |

- Fusions

| Fusion Gene & Exon | Transcript ID |
|--------------------|-----------------------------------|
| No fu: | sion gene detected in this sample |

- Immune Checkpoint Inhibitor (ICI) Related Biomarkers

| Biomarker | Results |
|----------------------------------|-----------------------------|
| Tumor Mutational Burden (TMB) | 0.7 muts/Mb |
| Microsatellite Instability (MSI) | Microsatellite stable (MSS) |

Note:

- Variant(s) enlisted in the SNV table may currently exhibit no relevance to treatment response prediction. Please refer to INTERPRETATION for more biological information and/or potential clinical impacts of the variants.
- Loss of heterozygosity (LOH) information was used to infer tumor cellularity. Copy number alteration in the tumor was determined based on 40% tumor purity.
- For more therapeutic agents which are possibly respond to heterozygous deletion of genes listed above, please refer to APPENDIX for more information.
- TMB was calculated by using the sequenced regions of ACTOnco®+ to estimate the number of somatic nonsynonymous mutations per megabase of all protein-coding genes (whole exome). The threshold for high mutation load is set at ≥ 7.5 mutations per megabase. TMB, microsatellite status and gene copy number deletion cannot be determined if calculated tumor purity is < 30%.





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THERAPEUTIC IMPLICATIONS

TARGETED THERAPIES

| Genomic Alterations | Therapies | Effect |
|---------------------|--------------------------------------|-----------|
| Level 4 | | |
| RB1 Splice acceptor | Abemaciclib, Palbociclib, Ribociclib | resistant |

Therapies associated with benefit or lack of benefit are based on biomarkers detected in this tumor and published evidence in professional guidelines or peer-reviewed journals.

| Level | Description |
|-------|--|
| 1 | FDA-recognized biomarkers predictive of response or resistance to FDA approved drugs in this indication |
| 2 | Standard care biomarkers (recommended by the NCCN guideline) predictive of response or resistance to FDA approved drugs in this indication |
| зА | Biomarkers predictive of response or resistance to therapies approved by the FDA or NCCN guideline in a different cancer type |
| 3B | Biomarkers that serve as inclusion criteria for clinical trials (minimal supportive data required) |
| 4 | Biomarkers that show plausible therapeutic significance based on small studies, few case reports, or preclinical studies |





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IMMUNE CHECKPOINT INHIBITORS (ICIs)

No genomic alterations detected to confer sensitivity or lack of benefit to immune checkpoint therapies.

- Other Biomarkers with Potential Clinical Effects for ICIs

| Genomic Alterations | Potential Clinical Effects |
|---------------------|----------------------------|
| | Not detected |

Note: Tumor non-genomic factors, such as patient germline genetics, PDL1 expression, tumor microenvironment, epigenetic alterations or other factors not provided by this test may affect ICI response.

CHEMOTHERAPIES

| Genomic Alterations | Therapies | Effect | Level of Evidence | Cancer Type |
|---------------------|--------------------|-----------|-------------------|-------------------|
| | Cisplatin | Sensitive | Clinical | Bladder carcinoma |
| RB1 | FAC | | | |
| Splice acceptor | T/FAC | Sensitive | Clinical | Breast cancer |
| | taxane/doxorubicin | | | |

HORMONAL THERAPIES

| Genomic Alterations | Therapies | Effect | Level of Evidence | Cancer Type |
|---------------------|-----------|-----------|-------------------|---------------|
| RB1 | Tomovifon | Decistant | Clinical | Drocat concer |
| Splice acceptor | Tamoxifen | Resistant | Clinical | Breast cancer |

OTHERS

No genomic alterations detected in this tumor predicted to confer sensitivity or lack of benefit to other therapies.

Note:

Therapeutic implications provided in the test are based solely on the panel of 440 genes sequenced. Therefore, alterations in genes not covered in this panel, epigenetic and post-transcriptional and post-translational factors may also determine a patient's response to therapies. In addition, several other patient-associated clinical factors, including but not limited to, prior lines of therapies received, dosage and combinations with other therapeutic agents, patient's cancer types, sub-types, and/or stages, may also determine the patient's clinical response to therapies.





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VARIANT INTERPRETATION

RB1 Splice acceptor

Biological Impact

The Retinoblastoma (RB1) gene encodes a tumor suppressor that negatively regulates the cell cycle, cell division, and DNA replication[1]. Loss-of-function RB1 could lead to unregulated cell division and growth, abrogation of multiple mechanisms that safeguard against cellular transformation, and tumorigenesis[2]. RB1 has also been implicated as a haploinsufficient tumor suppressor with one copy loss may lead to weak protein expression and is insufficient to execute its original physiological functions [3][4][5]. Deletion or inactivating mutation of RB1 is found in a number of tumors, including lung, prostate, bladder, breast cancers and sarcomas. RB1 mutations are found in approximately half of all retinoblastoma cases[6].

RB1 c.540-1G>A is a variant located at the splice acceptor region, which may result in the exon skipping.

Therapeutic and prognostic relevance

A deleterious mutation in one or more of the three DNA repair genes ATM, RB1, and FANCC predicted pathologic response and better overall survival to cisplatin-based chemotherapy for muscle-invasive bladder cancer patients[7]. High RB loss was found to be associated with improved pathologic clinical response in breast cancer patients treated with 5-fluorouracil/adriamycin/cytoxan (FAC), T/FAC, and Taxane/Adriamycin neoadjuvant therapy^[8].

Clinical and experimental data suggested that a non-functional retinoblastoma pathway is associated with resistance to tamoxifen in breast cancer^{[9][10]}.

Acquired RB1 mutations were found in hormone receptor positive breast cancer patients who developed resistance to palbociclib or ribociclib treatment[11]. Preclinical data also showed that knockdown of RB1 would impair antitumor activity of CDK4/6 inhibitor, abemaciclib[12].

Two large-scale genome-sequencing projects have identified a high prevalence of mutations in TP53 and RB1 in small cell lung cancer (SCLC)^{[13][14]}. Analyses of repeat biopsy samples from patients with EGFR-mutant adenocarcinoma that had transformed to the SCLC subtype have revealed that 100% of these patients have loss of RB1 and may be the alteration that induces this non-small-cell to small-cell transformation^{[10][15]}.

TP53 G245V, Heterozygous deletion

Biological Impact

TP53 encodes the p53 protein, a crucial tumor suppressor that orchestrates essential cellular processes including cell cycle arrest, senescence and apoptosis[16]. TP53 is a proto-typical haploinsufficient gene, such that loss of a single copy of TP53 can result in tumor formation[17].

TP53 G245V is a missense mutation locates within the DNA-binding domain (DBD) of the p53 protein (UniProtKB). TP53 G245V was demonstrated as a loss-of-function mutation with decreased transactivation activity and suppress cell growth when compared with wild type TP53^[18].

Loss of the second wild-type allele resulted in the biallelic inactivation of the gene.

Therapeutic and prognostic relevance

Despite having a high mutation rate in cancers, there are currently no approved targeted therapies for TP53 mutations. A phase II trial demonstrated that Wee1 inhibitor (AZD1775) in combination with carboplatin was well tolerated and showed promising anti-tumor activity in TP53-mutated ovarian cancer refractory or resistant (< 3 months) to standard first-line therapy (NCT01164995)[19].





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In a retrospective study (n=19), advanced sarcoma patients with TP53 loss-of-function mutations displayed improved progression-free survival (208 days versus 136 days) relative to patients with wild-type TP53 when treated with pazopanib^[20]. Results from another Phase I trial of advanced solid tumors (n=78) demonstrated that TP53 hotspot mutations are associated with better clinical response to the combination of pazopanib and vorinostat^[21].

Advanced solid tumor and colorectal cancer patients harboring a TP53 mutation have been shown to be more sensitive to bevacizumab when compared with patients harboring wild-type TP53^{[22][23][24]}. In a pilot trial (n=21), TP53-negative breast cancer patients demonstrated increased survival following treatment with bevacizumab in combination with chemotherapy agents, Adriamycin (doxorubicin) and Taxotere (docetaxel)^[25]. TP53 mutations were correlated with poor survival of advanced breast cancer patients receiving tamoxifen or primary chemotherapy^{[26][27]}. In a retrospective study of non-small cell lung cancer (NSCLC), TP53 mutations were associated with high expression of VEGF-A, the primary target of bevacizumab, offering a mechanistic explanation for why patients exhibit improved outcomes after bevacizumab treatment when their tumors harbor mutant TP53 versus wild-type TP53^[28].

FLCN Heterozygous deletion

Biological Impact

The FLCN gene encodes the tumor suppressor, Folliculin, a GTPase activating protein (GAP) for RagC/D GTPase proteins involved in amino acid sensing and signaling to mTORC1^[29]. FLCN has been implicated as a haploinsufficient gene with one copy loss may lead to weak protein expression and is insufficient to execute its original physiological functions^{[30][31]}. Inactivation of the FLCN gene by mutation or deletion results in the activation of the mTOR pathway and AKT signaling^{[32][33]}. Germline mutation of the FLCN gene causes the Birt-Hogg-Dubé syndrome, a rare disorder that is characterized by benign hamartomatous skin lesions and an increased risk of pneumothorax and renal tumors^[34].

Therapeutic and prognostic relevance

In a prospective Phase 2 study, four anaplastic thyroid cancer (ATC)/ poorly differentiated thyroid cancer (PDTC) patients who had PI3K/mTOR/AKT alterations, including TSC2, FLCN or NF1, showed impressive progression-free survival (PFS) of 15.2 months after receiving everolimus^[35]. mTOR inhibition via rapamycin also demonstrated potential in inhibiting the growth of renal cells deficient in FLCN in the preclinical setting^[36].

RPTOR Amplification

Biological Impact

RPTOR (regulatory-associated protein of mTOR, also known as RAPTOR) gene encodes a scaffold protein which is involved in assembly, localization and substrate binding of the mTORC1 complex and plays essential roles in mTOR signaling^{[37][38][39][40]}. Conditional disruption of RAPTOR has been reported to result in a development block of B cell^[41].

Therapeutic and prognostic relevance

RAPTOR overexpression has been reported to associate with acquired resistance to mTOR inhibition in vitro[42].

A study of breast cancer patients (n=150) indicated that higher expression of RAPTOR was associated with higher tumor grade^[43].

A study of hepatocellular carcinoma patients (n=62) indicated that high Rictor/Raptor ratio was associated with shorter disease-free survival^[44].





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US FDA-APPROVED DRUG(S)

Everolimus (AFINITOR)

Everolimus, a derivative of sirolimus, works as an inhibitor of mammalian target of rapamycin complex 1 (mTORC1) and blocks mTORC1-mediated downstream signals for cell growth, proliferation, and survival. Everolimus is developed and marketed by Novartis under the trade name AFINITOR.

- FDA Approval Summary of Everolimus (AFINITOR)

| RADIANT-4 ^[45] NCT01524783 | Lung or gastrointestinal neuroendocrine tumor (Approved on 2016/02/26) |
|---|---|
| | |
| NC101324763 | Everolimus vs. Placebo [PFS(M): 11 vs. 3.9] |
| BOLERO-2 ^[46] | Breast cancer (Approved on 2012/07/20) |
| NCT00863655 | ER+/HER2- |
| NC 100003033 | Everolimus + exemestane vs. Placebo + exemestane [PFS(M): 7.8 vs. 3.2] |
| | Tuberous sclerosis complex (tsc)-associated renal angiomyolipoma (Approved on |
| EXIST-2 NCT00790400 | 2012/04/26) |
| | |
| | Everolimus vs. Placebo [ORR(%): 41.8 vs. 0] |
| DADIANT 0[47] | Pancreatic neuroendocrine tumor (Approved on 2011/05/05) |
| RADIANT-3 ^[47] | |
| NCT00510068 | Everolimus vs. Placebo [PFS(M): 11 vs. 4.6] |
| ENGE 4[48] | Subependymal giant cell astrocytoma (Approved on 2010/10/29) |
| EXIST-1 ^[48] | |
| NCT00789828 | Everolimus vs. Placebo [ORR(%): 35.0] |
| DECODD 4[49] | Renal cell carcinoma (Approved on 2009/05/30) |
| RECORD-1 ^[49] | - |
| NCT00410124 | Everolimus vs. Placebo [PFS(M): 4.9 vs. 1.9] |

Temsirolimus (TORISEL)

Temsirolimus is a soluble ester of sirolimus (rapamycin, brand-name drug Rapamune) and functions as an inhibitor of mammalian target of rapamycin complex (mTORC). The inhibitory molecular mechanism is similar to Everolimus. Temsirolimus is developed by Wyeth Pharmaceuticals and marketed by Pfizer under the trade name TORISEL.

- FDA Approval Summary of Temsirolimus (TORISEL)

| [50] | Renal cell carcinoma (Approved on 2007/05/30) |
|-------------|---|
| NCT00065468 | - |
| NC10005468 | Temsirolimus vs. Ifn-α [OS(M): 10.9 vs. 7.3] |

D=day; W=week; M=month





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ONGOING CLINICAL TRIALS

Trials were searched by applying filters: study status, patient's diagnosis, intervention, location and/or biomarker(s). Please visit https://clinicaltrials.gov to search and view for a complete list of open available and updated matched trials.

No trial has been found.





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SUPPLEMENTARY INFORMATION OF TESTING RESULTS DETAILED INFORMATION OF VARIANTS WITH CLINICAL RELEVANCE

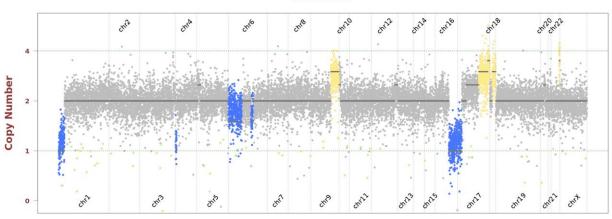
- Single Nucleotide and Small InDel Variants

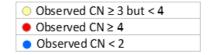
| Gene | Amino Acid Change | Exon | cDNA Change | Accession Number | COSMIC ID | Allele Frequency | Coverage |
|------|----------------------|------|----------------|---------------------|-------------|---------------------|----------|
| RB1 | Splice acceptor | - | c.540-1G>A | NM_000321 | COSM3469141 | 47.1% | 459 |
| TP53 | G245V | 7 | c.734G>T | NM_000546 | COSM11196 | 68.7% | 839 |

- Copy Number Alterations

Observed copy number (CN) for each evaluated position is shown on the y-axis. Regions referred to as amplification or deletion are shown in color. Regions without significant changes are represented in gray.

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OTHER DETECTED VARIANTS

| Gene | Amino Acid Change | Exon | cDNA Change | Accession Number | COSMIC ID | Allele Frequency | Coverage |
|----------|----------------------|------|----------------|---------------------|-----------|---------------------|----------|
| ADAMTSL1 | A864T | 19 | c.2590G>A | NM_001040272 | - | 50.9% | 986 |
| HR | R98C | 2 | c.292C>T | NM_005144 | - | 49.0% | 449 |
| HSP90AA1 | K279M | 5 | c.836A>T | NM_005348 | - | 50.8% | 1598 |
| KMT2C | N4686S | 54 | c.14057A>G | NM_170606 | - | 50.0% | 1216 |
| LIG3 | R942Q | 20 | c.2825G>A | NM_013975 | - | 46.0% | 1949 |
| NBN | S341R | 9 | c.1023C>G | NM_002485 | - | 48.8% | 842 |
| PDIA3 | V125I | 4 | c.373G>A | NM_005313 | - | 54.5% | 576 |
| PIK3C2G | E1260D | 28 | c.3780G>T | NM_004570 | - | 46.3% | 1000 |
| POLE | K101E | 4 | c.301A>G | NM_006231 | - | 49.7% | 674 |
| RPTOR | R1196K | 30 | c.3587G>A | NM_020761 | - | 14.0% | 2379 |
| RXRA | M127T | 3 | c.380T>C | NM_002957 | - | 38.1% | 788 |
| SDHA | R554Q | 12 | c.1661G>A | NM_004168 | - | 47.5% | 1165 |
| SETD2 | P226S | 3 | c.676C>T | NM_014159 | - | 49.6% | 1418 |

Note:

- This table enlists variants detected by the panel other than those with clinical relevance (reported in Testing Result section).

The clinical impact of a genetic variant is determined according to ACT Genomics in-house clinical knowledge database. A negative result does not necessarily indicate absence of biological effect on the tumor. Some variants listed here may possibly have preclinical data or may show potential clinical relevance in the future.





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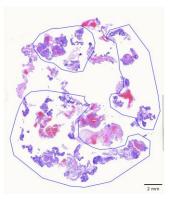
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TEST DETAILS SPECIMEN RECEIVED AND PATHOLOGY REVIEW





Collection date: Apr 21, 2023Facility retrieved: 臺北榮總

H&E-stained section No.: S11217744A

Collection site: L-spine

Examined by: Dr. Chien-Ta Chiang

- 1. The percentage of viable tumor cells in total cells in the whole slide (%): 25%
- 2. The percentage of viable tumor cells in total cells in the encircled areas in the whole slide (%): 40%
- 3. The percentage of necrotic cells (including necrotic tumor cells) in total cells in the whole slide (%): 5%
- The percentage of necrotic cells (including necrotic tumor cells) in total cells in the encircled areas in the whole slide (%): 5%
- 5. Additional comment: NA
- Manual macrodissection: Performed on the highlighted region
- The outline highlights the area of malignant neoplasm annotated by a pathologist.

RUN QC

Panel: ACTOnco®+

DNA test

- Mean Depth: 931x

- Target Base Coverage at 100x: 95%

RNA test

Average unique RNA Start Sites per control GSP2: 117

LIMITATIONS

- This test does not provide information of variant causality and does not detect variants in non-coding regions that could affect gene expression. This report does not report polymorphisms and we do not classify whether a mutation is germline or somatic.
 Variants identified by this assay were not subject to validation by Sanger or other technologies.
- 2. The possibility cannot be excluded that certain pathogenic variants detected by other sequencing tools may not be reported in the test because of technical limitation of bioinformatics algorithm or the NGS sequencing platform, e.g. low coverage.
- 3. This test has been designed to detect fusions in 13 genes sequenced. Therefore, fusion in genes not covered by this test would not be reported. For novel fusions detected in this test, Sanger sequencing confirmation is recommended if residue specimen is available.





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NEXT-GENERATION SEQUENCING (NGS) METHODS

DNA test

Extracted genomic DNA was amplified using primers targeting coding exons of analyzed genes and subjected to library construction. Barcoded libraries were subsequently conjugated with sequencing beads by emulsion PCR and enriched using Ion Chef system. Sequencing was performed according to Ion Proton or Ion S5 sequencer protocol (Thermo Fisher Scientific).

Raw reads generated by the sequencer were mapped to the hg19 reference genome using the Ion Torrent Suite. Coverage depth was calculated using Torrent Coverage Analysis plug-in. Single nucleotide variants (SNVs) and short insertions/deletions (InDels) were identified using the Torrent Variant Caller plug-in. VEP (Variant Effect Predictor) was used to annotate every variant using databases from Clinvar, COSMIC and Genome Aggregation database. Variants with coverage ≥ 20, allele frequency ≥ 5% and actionable variants with allele frequency ≥ 2% were retained. This test provides uniform coverage of the targeted regions, enabling target base coverage at 100x ≥ 85% with a mean coverage ≥ 500x.

Variants reported in Genome Aggregation database with > 1% minor allele frequency (MAF) were considered as polymorphisms. ACT Genomics in-house database was used to determine technical errors. Clinically actionable and biologically significant variants were determined based on the published medical literature.

The copy number alterations (CNAs) were predicted as described below:

Amplicons with read counts in the lowest 5th percentile of all detectable amplicons and amplicons with a coefficient of variation ≥ 0.3 were removed. The remaining amplicons were normalized to correct the pool design bias. ONCOCNV (an established method for calculating copy number aberrations in amplicon sequencing data by Boeva et al., 2014) was applied for the normalization of total amplicon number, amplicon GC content, amplicon length, and technology-related biases, followed by segmenting the sample with a gene-aware model. The method was used as well for establishing the baseline of copy number variations.

Tumor mutational burden (TMB) was calculated by using the sequenced regions of ACTOnco®+ to estimate the number of somatic nonsynonymous mutations per megabase of all protein-coding genes (whole exome). The TMB calculation predicted somatic variants and applied a machine learning model with a cancer hotspot correction. TMB may be reported as "TMB-High", "TMB-Low" or "Cannot Be Determined". TMB-High corresponds to ≥ 7.5 mutations per megabase (Muts/Mb); TMB-Low corresponds to < 7.5 Muts/Mb. TMB is reported as "Cannot Be Determined" if the tumor purity of the sample is < 30%.

Classification of microsatellite instability (MSI) status is determined by a machine learning prediction algorithm. The change of a number of repeats of different lengths from a pooled microsatellite stable (MSS) baseline in > 400 genomic loci are used as the features for the algorithm. The final output of the results is either microsatellite Stable (MSS) or microsatellite instability high (MSI-H).

Extracted RNA was reverse-transcribed and subjected to library construction. Sequencing was performed according to Ion Proton or Ion S5 sequencer protocol (Thermo Fisher Scientific). To ensure sequencing quality for fusion variant analysis, the average unique RNA Start Sites (SS) per control Gene Specific Primer 2 (GSP 2) should be ≥ 10.

The fusion analysis pipeline aligned sequenced reads to the human reference genome, identified regions that map to noncontiguous regions of the genome, applied filters to exclude probable false-positive events and, annotated previously characterized fusion events according to Quiver Gene Fusion Database, a curated database owned and maintained by ArcherDX. In general, samples with detectable fusions need to meet the following criteria: (1) Number of unique start sites (SS) for the GSP2 ≥ 3; (2) Number of supporting reads spanning the fusion junction ≥ 5; (3) Percentage of supporting reads spanning the fusion junction ≥ 10%; (4) Fusions annotated in Quiver Gene Fusion Database.





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DATABASE USED

- Reference genome: Human genome sequence hg19
- COSMIC v.92
- Genome Aggregation database r2.1.1
- ClinVar (version 20210404)
- ACT Genomics in-house database
- Quiver Gene Fusion Database version 5.1.18

Variant Analysis:

解剖病理專科醫師朱盈霞 Ying-Hsia Chu, M.D. 病解字第 000653 號 Sign Off

解剖病理專科醫師朱盈霞 Ying-Hsia Chu, M.D. 病解字第 000653 號 Jyth- am





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GENE LIST SNV & CNV

| ABCB1* | ABCC2* | ABCG2* | ABL1 | ABL2 | ADAMTS1 | ADAMTS13 | ADAMTS15 | ADAMTS16 | ADAMTS18 | ADAMTS6 | ADAMTS9 |
|----------|---------|---------|----------|----------|---------|-----------|-----------|----------|----------|----------|---------------|
| ADAMTSL1 | ADGRA2 | ADH1C* | AKT1 | AKT2 | AKT3 | ALDH1A1* | ALK | AMER1 | APC | AR | ARAF |
| ARID1A | ARID1B | ARID2 | ASXL1 | ATM | ATR | ATRX | AURKA | AURKB | AXIN1 | AXIN2 | AXL |
| B2M | BAP1 | BARD1 | BCL10 | BCL2* | BCL2L1 | BCL2L2* | BCL6 | BCL9 | BCOR | BIRC2 | BIRC3 |
| BLM | BMPR1A | BRAF | BRCA1 | BRCA2 | BRD4 | BRIP1 | BTG1 | BTG2* | ВТК | BUB1B | CALR |
| CANX | CARD11 | CASP8 | CBFB | CBL | CCNA1 | CCNA | CCNB1 | CCNB2 | CCNB3 | CCND1 | CCND2 |
| CCND3 | CCNE1 | CCNE2 | CCNH | CD19 | CD274 | CD58 | CD70* | CD79A | CD79B | CDC73 | CDH1 |
| CDK1 | CDK12 | CDK2 | CDK4 | CDK5 | CDK6 | CDK7 | CDK8 | CDK9 | CDKN1A | CDKN1B | CDKN2A |
| CDKN2B | CDKN2C | CEBPA* | CHEK1 | CHEK2 | CIC | CREBBP | CRKL | CRLF2 | CSF1R | CTCF | CTLA4 |
| CTNNA1 | CTNNB1 | CUL3 | CYLD | CYP1A1* | CYP2B6* | CYP2C19* | CYP2C8* | CYP2D6 | CYP2E1* | CYP3A4* | CYP3A5* |
| DAXX | DCUN1D1 | DDR2 | DICER1 | DNMT3A | DOT1L | DPYD | DTX1 | E2F3 | EGFR | EP300 | EPCAM |
| EPHA2 | ЕРНА3 | EPHA5 | ЕРНА7 | ЕРНВ1 | ERBB2 | ERBB3 | ERBB4 | ERCC1 | ERCC2 | ERCC3 | ERCC4 |
| ERCC5 | ERG | ESR1 | ESR2 | ETV1 | ETV4 | EZH2 | FAM46C | FANCA | FANCC | FANCD2 | FANCE |
| FANCF | FANCG | FANCL | FAS | FAT1 | FBXW7 | FCGR2B | FGF1* | FGF10 | FGF14 | FGF19* | FGF23 |
| FGF3 | FGF4* | FGF6 | FGFR1 | FGFR2 | FGFR3 | FGFR4 | FH | FLCN | FLT1 | FLT3 | FLT4 |
| FOXL2* | FOXP1 | FRG1 | FUBP1 | GATA1 | GATA2 | GATA3 | GNA11 | GNA13 | GNAQ | GNAS | GREM1 |
| GRIN2A | GSK3B | GSTP1* | GSTT1* | HGF | HIF1A | HIST1H1C* | HIST1H1E* | HNF1A | HR | HRAS* | HSP90AA |
| HSP90AB1 | HSPA4 | HSPA5 | IDH1 | IDH2 | IFNL3* | IGF1 | IGF1R | IGF2 | IKBKB | IKBKE | IKZF1 |
| IL6 | IL7R | INPP4B | INSR | IRF4 | IRS1 | IRS2* | JAK1 | JAK2 | JAK3 | JUN* | KAT6A |
| KDM5A | KDM5C | KDM6A | KDR | KEAP1 | KIT | KMT2A | КМТ2С | KMT2D | KRAS | LCK | LIG1 |
| LIG3 | LMO1 | LRP1B | LYN | MALT1 | MAP2K1 | MAP2K2 | MAP2K4 | MAP3K1 | MAP3K7 | MAPK1 | МАРК3 |
| MAX | MCL1 | MDM2 | MDM4 | MED12 | MEF2B | MEN1 | MET | MITF | MLH1 | MPL | MRE11 |
| MSH2 | MSH6 | MTHFR* | MTOR | MUC16 | MUC4 | MUC6 | митүн | MYC | MYCL | MYCN | MYD88 |
| NAT2* | NBN | NEFH | NF1 | NF2 | NFE2L2 | NFKB1 | NFKBIA | NKX2-1* | NOTCH1 | NOTCH2 | <i>NOTCH3</i> |
| NOTCH4 | NPM1 | NQ01* | NRAS | NSD1 | NTRK1 | NTRK2 | NTRK3 | PAK3 | PALB2 | PARP1 | PAX5 |
| PAX8 | PBRM1 | PDCD1 | PDCD1LG2 | PDGFRA | PDGFRB | PDIA3 | PGF | PHOX2B* | PIK3C2B | PIK3C2G | РІКЗСЗ |
| PIK3CA | РІКЗСВ | PIK3CD | PIK3CG | PIK3R1 | PIK3R2 | PIK3R3 | PIM1 | PMS1 | PMS2 | POLB | POLD1 |
| POLE | PPARG | PPP2R1A | PRDM1 | PRKAR1A | PRKCA | PRKCB | PRKCG | PRKCI | PRKCQ | PRKDC | PRKN |
| PSMB8 | PSMB9 | PSME1 | PSME2 | PSME3 | PTCH1 | PTEN | PTGS2 | PTPN11 | PTPRD | PTPRT | RAC1 |
| RAD50 | RAD51 | RAD51B | RAD51C | RAD51D | RAD52 | RAD54L | RAF1 | RARA | RB1 | RBM10 | RECQL4 |
| REL | RET | RHOA | RICTOR | RNF43 | ROS1 | RPPH1 | RPTOR | RUNX1 | RUNX1T1 | RXRA | SDHA |
| SDHB | SDHC | SDHD | SERPINB3 | SERPINB4 | SETD2 | SF3B1 | SGK1 | SH2D1A* | SLC19A1* | SLC22A2* | SLCO1B1 |
| SLCO1B3* | SMAD2 | SMAD3 | SMAD4 | SMARCA4 | SMARCB1 | SMO | SOCS1* | SOX2* | SOX9 | SPEN | SPOP |
| SRC | STAG2 | STAT3 | STK11 | SUFU | SYK | SYNE1 | TAF1 | TAP1 | TAP2 | TAPBP | ТВХ3 |
| TEK | TERT | TET1 | TET2 | TGFBR2 | TMSB4X* | TNF | TNFAIP3 | TNFRSF14 | TNFSF11 | TOP1 | TP53 |
| TPMT* | TSC1 | TSC2 | TSHR | TYMS | U2AF1 | UBE2A* | UBE2K | UBR5 | UGT1A1* | USH2A | VDR* |
| VEGFA | VEGFB | VHL | WT1 | XIAP | XPO1 | XRCC2 | ZNF217 | | | | |

^{*}Analysis of copy number alterations NOT available.

FUSION

| ALK | BRAF | LC LD | FGFR1 | FGFR2 | FGFR3 | MET | NRG1 | NTRK1 | NTRK2 | NTRK3 | RET | ROS1 |
|-----|------|-------|-------|-------|-------|-------|-------|--------|--------|--------|-----|------|
| ALN | DRAF | EGFK | FGFKI | FGFKZ | rurk3 | IVIEI | INKGI | INIKKT | INTRAZ | INTRAS | KEI | KOSI |





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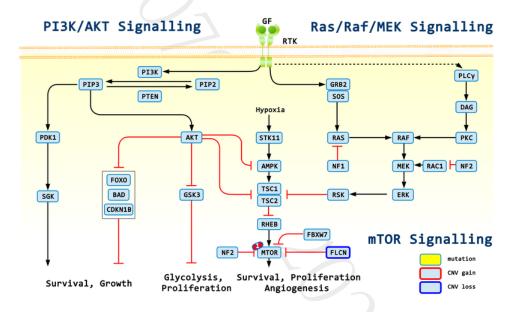
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APPENDIX

POSSIBLE THERAPEUTIC IMPLICATIONS FOR HETEROZYGOUS DELETION

| Gene | Therapies | Possible effect |
|------|--------------------------|-----------------|
| FLCN | Everolimus, Temsirolimus | sensitive |

SIGNALING PATHWAYS AND MOLECULAR-TARGETED AGENTS



1: Everolimus, Temsirolimus





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本報告中列出之生物標記變異與藥物資訊並非依照潛在治療有效性排序。

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藥物潛在臨床效益(或缺乏潛在臨床效益)的實證證據是依據至少一篇臨床療效個案報告或臨床前試驗做為評估。本公司盡力提供適時及 準確之資料,但由於醫學科技之發展日新月異,本公司不就本報告提供的資料是否為準確、適宜或最新作保證。

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