

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT	DISEASE Brain glioblastoma (GBM)	PHYSICIAN	ORDERING PHYSICIAN Yeh, Yi-Chen	SPECIMEN	SPECIMEN SITE Brain
	NAME Lin, Yu Fu		MEDICAL FACILITY Taipei Veterans General Hospital		SPECIMEN ID S111-537191
	DATE OF BIRTH 10 September 1946		ADDITIONAL RECIPIENT None		SPECIMEN TYPE Slide Deck
	SEX Male		MEDICAL FACILITY ID 205872		DATE OF COLLECTION 21 December 2022
	MEDICAL RECORD # 48288502		PATHOLOGIST Not Provided		SPECIMEN RECEIVED 05 January 2023

Biomarker Findings

Microsatellite status - MS-Stable
Tumor Mutational Burden - 8 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

EGFR P596L
KIT amplification
PDGFRA amplification
MTAP loss
CDKN2A/B CDKN2A loss, CDKN2B loss
TERT promoter -124C>T

1 Disease relevant genes with no reportable alterations: **IDH1**

Report Highlights

- Variants with **diagnostic implications** that may indicate a specific cancer type: **TERT** promoter -124C>T (p. 7)
- Targeted therapies with potential clinical benefit **approved in another tumor type**: Erlotinib (p. 8), Gefitinib (p. 8), Imatinib (p. 9), Nilotinib (p. 9), Osimertinib (p. 10), Sorafenib (p. 11), Sunitinib (p. 11)
- Evidence-matched **clinical trial options** based on this patient's genomic findings: (p. 12)
- Variants with **prognostic implications** for this tumor type that may impact treatment decisions: **TERT** promoter -124C>T (p. 7)

BIOMARKER FINDINGS

Microsatellite status - MS-Stable

Tumor Mutational Burden - 8 Muts/Mb

GENOMIC FINDINGS

EGFR - P596L

7 Trials see p. 12

KIT - amplification

10 Trials see p. 14

PDGFRA - amplification

1 Trial see p. 17

MTAP - loss

1 Trial see p. 16

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. See Biomarker Findings section

No therapies or clinical trials. See Biomarker Findings section

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)
none	Erlotinib
	Gefitinib
	Osimertinib
none	Imatinib
	Nilotinib
	Sorafenib
	Sunitinib
none	Imatinib
none	none

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

CDKN2A/B - CDKN2A loss, CDKN2B loss p. [6](#) **TERT** - promoter -124C>T p. [7](#)

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

BIOMARKER FINDINGS

BIOMARKER

Microsatellite status

RESULT

MS-Stable

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁻³, including approved therapies nivolumab and pembrolizumab⁴. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and

experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, $p=0.001$)⁵.

FREQUENCY & PROGNOSIS

Low-level MSI has been reported in 5-9% of glioblastoma (GBM) samples⁶⁻⁸. A large-scale study did not find high-level microsatellite instability (MSI-H) in any of 129 GBM samples⁶, although a small-scale study reported MSI-H in 4 of 15 pediatric GBMs and 1 of 12 adult GBMs⁹. The frequency of MSI has been reported to be increased in relapsed compared to primary GBM⁶, in GBMs with a previous lower grade astrocytoma⁷, and in giant cell GBM compared to classic GBM⁸.

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor¹⁰. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2¹⁰⁻¹². This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers¹³⁻¹⁵. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{10,12,14-15}.

BIOMARKER

Tumor Mutational Burden

RESULT

8 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1¹⁶⁻¹⁸, anti-PD-1 therapies¹⁶⁻¹⁹, and combination nivolumab and ipilimumab²⁰⁻²⁵. In glioma, a lack of association between TMB and clinical benefit from immune checkpoint inhibitors has been reported^{16,26-27}. However, multiple case studies have reported that patients with ultramutated gliomas driven by POLE mutations

have benefited from treatment with anti-PD-1²⁸⁻²⁹ or anti-PD-L1³⁰ therapies. Therefore, although increased TMB alone may not be a strong biomarker for PD-1 or PD-L1 inhibitors in this cancer type, these agents may have efficacy for patients with glioma harboring both high TMB and POLE mutation.

FREQUENCY & PROGNOSIS

Glioblastoma (GBM) harbors a median TMB of 2.7 mutations per megabase (mut/Mb), and 4.2% of cases have high TMB (>20 muts/Mb)³¹. For pediatric patients, high TMB has been reported in a subset of high-grade gliomas, frequently in association with mutations in mismatch repair or proofreading genes and in TP53, whereas BRAF alterations or other oncogene fusions were observed more frequently in brain tumors harboring low TMB³²⁻³³. Increased TMB has been reported to correlate with higher tumor grade in glioma³⁴ and glioblastoma (GBM) tissue samples with biallelic mismatch repair deficiency

(bMMRD)²⁸, as well as with shorter OS of patients with diffuse glioma³⁵.

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma³⁶⁻³⁷ and cigarette smoke in lung cancer³⁸⁻³⁹, treatment with temozolomide-based chemotherapy in glioma⁴⁰⁻⁴¹, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes⁴²⁻⁴⁶, and microsatellite instability (MSI)^{42,45-46}. This sample harbors a TMB below levels that would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents^{16,26-30}.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

GENOMIC FINDINGS

GENE

EGFR

ALTERATION

P596L

TRANSCRIPT ID

NM_005228.3

CODING SEQUENCE EFFECT

1787C>T

VARIANT CHROMOSOMAL POSITION

chr7:55233037

VARIANT ALLELE FREQUENCY (% VAF)

8.2%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

For patients with non-small cell lung cancer (NSCLC), EGFR activating mutations may predict sensitivity to EGFR-TKIs, including erlotinib⁴⁷, gefitinib⁴⁸⁻⁵¹, afatinib⁵²⁻⁵⁵, dacomitinib⁵⁶, and

osimertinib^{53,57}; however, the data for patients with other tumor types are limited⁵⁸⁻⁶³. Patients with EGFR-mutated bithalamic glioma have reported responses to osimertinib⁶⁴⁻⁶⁵. In a case series of 11 patients with bithalamic gliomas with EGFR mutations, EGFR inhibitors, including osimertinib, showed improved survival; however, it showed a lack of significant clinical responses⁶¹. On the basis of preclinical data, EGFR mutations confer sensitivity to EGFR inhibitors, including osimertinib⁶¹.

FREQUENCY & PROGNOSIS

EGFR alterations have been reported in 13.2% of anaplastic astrocytomas, 5.3-15.9% of glioblastoma multiformes (GBMs), and 0% of pilocytic astrocytomas in several genomic studies of CNS tumors⁶⁶⁻⁶⁹. In GBMs, Missense mutations in the EGFR extracellular domain have been found in 10-15% of cases and approximately half have a low-level amplification of the mutated allele⁷⁰⁻⁷¹. In a study of IDH-wildtype GBM samples, EGFR

alterations were detected in 50% (117/232) of IDH-wildtype GBM samples analyzed, including 41% (95/232) with a co-occurring EGFR amplification and mutation, 26% (61/232) with an EGFR domain truncation event, such as EGFRvIII, and 2.2% (5/232) with an EGFR fusion event⁷². No definitive correlation has been identified between EGFR amplification and length of survival in patients with GBM⁷³⁻⁷⁴; however, EGFR amplification has been associated with prolonged survival in patients over the age of 60 with GBM⁷⁵.

FINDING SUMMARY

EGFR encodes the epidermal growth factor receptor, which belongs to a class of proteins called receptor tyrosine kinases. In response to signals from the environment, EGFR passes biochemical messages to the cell that stimulate it to grow and divide⁷⁶. EGFR mutations that have been characterized in biochemical assays to be activating, as observed here, are predicted to confer sensitivity to EGFR-targeted therapies^{70,77-93}.

GENE

KIT

ALTERATION

amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence, primarily in gastrointestinal stromal tumor (GIST), melanoma, AML, and systemic mastocytosis, KIT activating alterations are associated with sensitivity to TKIs including imatinib, sunitinib, sorafenib, dasatinib, nilotinib, pazopanib, regorafenib, ponatinib, midostaurin, apatinib, avapritinib, and ripretinib⁹⁴⁻¹⁰³. The use of mTOR inhibitors as an alternative therapeutic strategy has demonstrated limited success in KIT-mutated, imatinib-resistant melanoma, with 1 PR and 3 SD observed for 4 patients treated with everolimus¹⁰⁴. However, no

responses were observed for 10 patients with mastocytosis following everolimus monotherapy, with 8/10 patients harboring the KIT D816V mutation¹⁰⁵. The role of KIT amplification as a biomarker for response to mTOR inhibitors has not been investigated (PubMed, Mar 2022). Clinical benefit has been observed for patients with KIT amplified or overexpressing tumors following treatment with imatinib¹⁰⁶⁻¹¹⁶, nilotinib¹¹⁷, sorafenib¹¹⁸⁻¹²¹, and sunitinib¹²²⁻¹²³, suggesting that KIT amplification may be sensitive to these inhibitors. However, evidence demonstrating clinical benefit for regorafenib, dasatinib, pazopanib, or ponatinib in the context of KIT amplified or overexpressing tumors is limited.

FREQUENCY & PROGNOSIS

In the TCGA datasets, KIT amplification has been reported in 2.5% of lower grade gliomas (grades 2 and 3)¹²⁴ and 9.2% of glioblastomas (Grade 4 astrocytoma)⁶⁷. KIT amplification has been variously reported in 4-47% of glioblastomas in the

scientific literature¹²⁵⁻¹²⁷. Amplification of KIT has been strongly correlated with the presence of KDR and/or PDGFRA amplification in glioblastoma^{126,128-129}. One study found no correlation between KIT amplification and overall survival in patients with glioblastoma, while a separate study reported that overexpression of KIT was associated with tumor grade and shorter survival in patients with malignant glioma^{125,130}.

FINDING SUMMARY

KIT (also called c-KIT) encodes a cell surface tyrosine kinase receptor that, upon ligand binding and dimerization, activates the PI3K-AKT and RAS-MAPK signaling pathways¹³¹. KIT aberrations, including point mutations, translocations, amplification, and overexpression, have been associated with various malignancies, and KIT is considered an oncoprotein¹³². KIT has been reported to be amplified in cancer¹³³ and may be biologically relevant in this context¹³⁴⁻¹³⁵.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

GENOMIC FINDINGS

GENE PDGFRA

ALTERATION
amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of extensive clinical evidence in solid tumors and hematologic cancers, PDGFRA activating alterations are associated with sensitivity to imatinib¹³⁶⁻¹⁷³. Sorafenib has shown clinical and preclinical activity against the FIP1L1-PDGFRA fusion in chronic eosinophilic leukemia (CEL) and mutations associated with clinical resistance to imatinib and sunitinib in both CEL and gastrointestinal stromal tumor (GIST)¹⁷⁴⁻¹⁷⁹. Complete responses to nilotinib have been reported in patients with CEL or hypereosinophilic syndrome with FIP1L1-PDGFRA or activating mutations^{152,180-181}; preclinical evidence has reported efficacy of nilotinib in the context of PDGFRA mutations associated with GIST¹⁸²⁻¹⁸³. Patients with GIST harboring PDGFRA activating

mutations have been reported to derive clinical benefit from treatment with sunitinib¹⁸⁴ or regorafenib¹⁸⁵⁻¹⁸⁶. Preclinical studies have reported sensitivity of activating PDGFRA mutations and FIP1L1-PDGFRA fusion to dasatinib^{176,182}. PDGFRA D842 mutations were reported to be sensitive to avapritinib in clinical⁹⁴ and preclinical⁹⁴ studies of GIST, and demonstrated sensitivity to ripretinib for 1 patient¹⁸⁷.

FREQUENCY & PROGNOSIS

PDGFRA amplification has been suggested to be more common in higher grade astrocytomas than in lower grade astrocytomas; studies have reported PDGFRA amplification in 16.3% (27/166) of Grade 2 astrocytomas and in 23.6% (91/386) of Grade 3 and 4 astrocytomas analyzed^{128,188-189}. PDGFRA amplification has been reported in 5.2-33% of glioblastoma cases^{67,125-126,188,190-191}. A retrospective analysis of TCGA glioma samples reported elevated expression of ERBB3 correlated with PDGFRA expression and co-expression of these genes was an indicator of poor prognosis in a GBM patient cohort¹⁹². Amplification of PDGFRA has been associated with tumor grade and poor progression-free and overall survival in patients with

glioblastoma^{188,190-191}. In addition, PDGFRA amplification has been reported to occur in conjunction with IDH1 mutation in glioblastoma, and both alterations in the same tumor have been associated with poor patient prognosis¹⁸⁸. Amplification of PDGFRA has also been strongly correlated with the presence of KDR and/or KIT amplification in glioblastomas, as well as with EGFR amplification^{126,128-129,193}.

FINDING SUMMARY

PDGFRA encodes platelet-derived growth factor receptor alpha (PDGFR-alpha), a tyrosine kinase receptor that, upon binding of cognate ligands (PDGFA or PDGFB), activates several signaling pathways, including PI3K and MAPK¹⁹⁴. PDGFR aberrations, including point mutations, translocations, amplification, and/or overexpression, have been associated with various malignancies¹³². Amplification of PDGFRA, frequently occurring with amplification of the genes KDR and KIT, has been associated with increased PDGFRA expression^{127,195-197} and poor prognosis^{127,188,198-199} in some subtypes of glioma.

GENE MTAP

ALTERATION
loss

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

MTAP inactivation produces specific metabolic vulnerabilities that may be sensitive to MAT2A²⁰⁰⁻²⁰¹ or PRMT5 inhibition²⁰¹⁻²⁰³. A Phase 1 trial of MAT2A inhibitor AG-270 reported 1 PR and 2 SDs lasting longer than 6 months for patients with advanced solid tumors displaying MTAP loss²⁰⁴. Preclinical data suggest that MTAP loss sensitizes cells to S-adenosyl-L-methionine (SAM)-competitive PRMT5 inhibitors²⁰⁵, dual PRMT1 and PRMT5 inhibitors²⁰⁶⁻²⁰⁸, and PRMT5 inhibitors that selectively bind the PRMT5 when complexed with S-methyl-5'-thioadenosine (MTA), such as MRTX1719, TNG908, and AMG193²⁰⁹. In preclinical models, MTAP inactivation showed

increased sensitivity to inhibitors of purine synthesis or purine analogs, especially upon addition of exogenous MTA²¹⁰⁻²²⁰. A Phase 2 study of L-alanosine, an inhibitor of adenine synthesis, as a monotherapy for 65 patients with MTAP-deficient cancers reported no responses and SD for 24% (13/55) of patients²²¹. Preclinical and limited clinical evidence suggest MTAP deficiency may confer sensitivity to pemetrexed²²².

FREQUENCY & PROGNOSIS

MTAP loss/homozygous deletion as well as loss of expression has been reported in a wide variety of solid tumors and hematologic cancers²²³⁻²²⁴; such events have been correlated with poor prognosis in a variety of cancer types, including hepatocellular carcinoma²²⁵, gastrointestinal stromal tumors²²⁶, mantle cell lymphoma (MCL)²²⁷, melanoma²²⁸⁻²²⁹, gastric cancer²³⁰, myxofibrosarcoma²³¹, nasopharyngeal carcinoma²³², ovarian carcinoma²²³ and non-small cell lung cancer²³³. MTAP loss was not prognostic in pediatric B-cell acute lymphocytic leukemia²³⁴ or in astrocytoma²³⁵. However, MTAP has also been reported to be

overexpressed in colorectal cancer (CRC) samples²³⁶, and MTAP retention is thought to be important for prostate cancer growth due to continuous supply of SAM²³⁷. Germline SNPs in MTAP have been correlated with the development of cutaneous melanoma²³⁸⁻²³⁹, esophageal cancer²⁴⁰⁻²⁴¹, osteosarcoma²⁴², and CRC²⁴³.

FINDING SUMMARY

MTAP encodes S-methyl-5'-thioadenosine (MTA) phosphorylase, a tumor suppressor involved in polyamine metabolism and methionine synthesis, although its enzymatic function is dispensable for its tumor suppressor activity²⁴⁴⁻²⁴⁵. Decreased expression of MTAP leads to MTA accumulation within tumor cells and their microenvironment^{225,246-247}, thereby reducing intracellular arginine methylation²⁰¹⁻²⁰³ and altering cell signaling²⁴⁷⁻²⁴⁸. MTAP is located at 9p21, adjacent to CDKN2A and CDKN2B, with which it is frequently co-deleted in various cancers. Other alterations in MTAP are rare and have not been extensively characterized.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

GENOMIC FINDINGS

GENE

CDKN2A/B

ALTERATION

CDKN2A loss, CDKN2B loss

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Preclinical data suggest that tumors with loss of p16INK4a function may be sensitive to CDK4/6 inhibitors, such as abemaciclib, ribociclib, and palbociclib²⁴⁹⁻²⁵². Clinical data in mesothelioma, breast cancer, and uterine leiomyosarcoma indicate that CDKN2A loss may predict sensitivity to abemaciclib²⁵³ and palbociclib treatment²⁵⁴⁻²⁵⁵. However, multiple other clinical studies have shown no significant correlation between p16INK4a loss or inactivation and therapeutic benefit of these agents²⁵⁶⁻²⁶²; it is not known whether CDK4/6 inhibitors would be beneficial in this case. Although preclinical studies have suggested that loss of p14ARF function may be associated with reduced sensitivity to MDM2 inhibitors²⁶³⁻²⁶⁴, the clinical relevance of p14ARF as a predictive biomarker is not clear. The p15INK4b protein encoded by CDKN2B is known to inhibit CDK4, and although concomitant loss of CDKN2A and CDKN2B may predict sensitivity to CDK4/6 inhibitors, such as ribociclib, abemaciclib, and palbociclib^{259-260,265-266}, direct supporting data for CDKN2B alteration as a predictive biomarker for these therapies are limited²⁶⁷⁻²⁶⁸.

FREQUENCY & PROGNOSIS

Concurrent putative homozygous deletion of CDKN2A and CDKN2B has been reported in 35% of patients with gliomas⁶⁸ and detected more frequently in patients with glioblastoma multiforme (GBM; 58%)⁶⁷ than in those with lower grade gliomas (6%)⁶⁶. In other studies, loss of CDKN2A/B by deletion has been reported in up to 78% of astrocytomas (including anaplastic astrocytomas and GBM)^{190,197,269}. A study found homozygous deletion of both p16INK4a and p14ARF in 26% (13/50) of glioblastomas (GBMs); 18% (9/50) of cases showed homozygous deletion of the p14ARF-encoding locus alone²⁷⁰. One study detected CDKN2A/B loss in 69% (161/232) and mutation in 2.6% (6/232) of IDH-wildtype GBM samples analyzed⁷². Decreased p14ARF and p16INK4a expression levels were found to be tightly associated in a study of glioma samples²⁷¹. Homozygous deletion of the genomic region including CDKN2A and CDKN2B has been found to be associated with poor prognosis in GBM and likely serves as an early event in GBM progression^{190,272}. In addition, expression of p16INK4a has been found to be lower in patients with high grade malignant gliomas compared to patients with low grade gliomas, and loss of p16INK4a expression has been associated with shorter overall survival in pilocytic astrocytomas²⁷³⁻²⁷⁴.

FINDING SUMMARY

CDKN2A encodes two different, unrelated tumor suppressor proteins, p16INK4a and p14ARF, whereas CDKN2B encodes the tumor suppressor

p15INK4b²⁷⁵⁻²⁷⁶. Both p15INK4b and p16INK4a bind to and inhibit CDK4 and CDK6, thereby maintaining the growth-suppressive activity of the Rb tumor suppressor; loss or inactivation of either p15INK4b or p16INK4a contributes to dysregulation of the CDK4/6-cyclin-Rb pathway and loss of cell cycle control²⁷⁷⁻²⁷⁸. The tumor suppressive functions of p14ARF involve stabilization and activation of p53, via a mechanism of MDM2 inhibition²⁷⁹⁻²⁸⁰. One or more alterations observed here are predicted to result in p16INK4a loss of function²⁸¹⁻³⁰². One or more alterations seen here are predicted to result in p14ARF loss of function^{285,302-305}. CDKN2B alterations such as seen here are predicted to inactivate p15INK4b³⁰⁶.

POTENTIAL GERMLINE IMPLICATIONS

Germline CDKN2A mutation is associated with melanoma-pancreatic cancer syndrome, a condition marked by increased risk of developing malignant melanoma and/or pancreatic cancer³⁰⁷. Mutation carriers within families may develop either or both types of cancer, and melanoma cases may be referred to as familial or hereditary melanoma³⁰⁸⁻³⁰⁹. CDKN2A is the most implicated gene in familial melanoma, with germline mutations present in 16% to 20% of familial melanoma cases³¹⁰⁻³¹². CDKN2A alteration has also been implicated in familial melanoma-astrocytoma syndrome, an extremely rare tumor association characterized by dual predisposition to melanoma and nervous system tumors³¹³⁻³¹⁵. In the appropriate clinical context, germline testing of CDKN2A is recommended.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

GENOMIC FINDINGS

GENE

TERT

ALTERATION

promoter -124C>T

TRANSCRIPT ID

NM_198253.2

CODING SEQUENCE EFFECT

-124C>T

VARIANT CHROMOSOMAL POSITION

chr5:1295228

VARIANT ALLELE FREQUENCY (% VAF)

20.9%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Therapeutic options for targeting tumors with TERT mutations are limited, although a variety of approaches have been investigated, including immunotherapies using TERT as a tumor-associated antigen and antisense oligonucleotide- or peptide-based therapies. TERT peptide vaccines showed limited anticancer efficacy in clinical trials³¹⁶; however, in one preclinical study, the combination of a TERT peptide vaccine and anti-CTLA-4 therapy suppressed tumor growth³¹⁷. A Phase 2 study of the TERT inhibitor imetelstat for

patients with advanced non-small cell lung cancer reported no improvement in PFS or OS³¹⁸.

FREQUENCY & PROGNOSIS

TERT promoter mutations have been reported in 51-59% of gliomas³¹⁹⁻³²⁰, most frequently in glioblastoma (GBM, 54-84%), gliosarcoma (81%), oligodendroglioma (78%), and historically in oligoastrocytomas (25-31%) but less frequently in lower grade astrocytomas (10-18%) and in only 1% of ependymomas³¹⁹⁻³²³. In patients with glioblastoma (GBM), the prevalence of TERT promoter mutation is lower in pediatric primary GBM (11%) and adult secondary GBM (28%) compared with adult primary GBM (58-83%)^{319,321}. One study detected TERT promoter mutations in 78% (181/232) of IDH-wildtype GBM samples analyzed⁷². TERT promoter mutation has been shown to be significantly associated with increased TERT gene expression in astrocytoma, oligodendroglioma, and GBM³²⁴. TERT promoter mutations significantly associate with poor prognosis in patients with GBM, although this correlation may be due to the association with primary GBM as opposed to IDH-positive secondary GBM^{319,321,324-325}. In the context of IDH-wildtype glioma, TERT mutations are associated with reduced OS (NCCN CNS Cancers Guidelines, v1.2022).

FINDING SUMMARY

Telomerase reverse transcriptase (TERT, or hTERT) is a catalytic subunit of the telomerase complex, which is required to maintain appropriate chromosomal length³²⁶. Activation of TERT is a hallmark of cancer, being detected in up to 80-90% of malignancies and absent in quiescent cells³²⁷⁻³²⁹. Mutations within the promoter region of TERT that confer enhanced TERT promoter activity have been reported in two hotspots, located at -124 bp and -146 bp upstream of the transcriptional start site (also termed C228T and C250T, respectively)³³⁰⁻³³², as well as tandem mutations at positions -124/-125 bp and -138/-139 bp³³⁰.

POTENTIAL DIAGNOSTIC IMPLICATIONS

TERT mutations are associated with 1p/19q co-deletion in oligodendrogliomas, and are highly recurrent in IDH/ATRX-wildtype glioblastoma (GBM) (NCCN CNS Cancers Guidelines, v1.2022)³³³. The presence of EGFR gene amplification or TERT promoter mutations are indicative of diffuse astrocytic glioma with molecular features of glioblastoma, WHO grade 4 in IDH1/2-wildtype tumors (NCCN CNS Cancers Guidelines, v1.2022)³³⁴.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Erlotinib

Assay findings association
EGFR
P596L

AREAS OF THERAPEUTIC USE

Erlotinib is a small-molecule inhibitor of EGFR. It is FDA approved as a monotherapy or in combination with ramucirumab for patients with metastatic non-small cell lung cancer (NSCLC) harboring EGFR exon 19 deletions or exon 21 (L858R) mutations. Erlotinib is also FDA approved in combination with gemcitabine as a first-line treatment for advanced pancreatic cancer. Please see the drug label for full prescribing information.

GENE ASSOCIATION

Amplification or activation of EGFR may predict sensitivity to therapies such as erlotinib. For patients with activating mutations in EGFR, treatment with erlotinib has been associated with improved response and lengthened time to progression^{47,335-337}.

SUPPORTING DATA

In the MyPathway Phase 2a basket study for advanced solid tumors, 1 of 9 patients with EGFR activation mutations responded to erlotinib monotherapy; the responding patient had urethral adenocarcinoma³³⁸. A

patient with EGFR-mutated metastatic lacrimal gland adenoid cystic carcinoma experienced clinical benefit from erlotinib treatment that was ongoing at 14 months³³⁹. A clinical study of patients with glioblastoma (GBM) treated with gefitinib or erlotinib found that 9/49 (18%) had tumor shrinkage of 25% or more; in this study, the extracellular domain EGFRvIII mutation was correlated with response³⁴⁰. In a Phase 2 study of 65 patients with GBM or gliosarcoma, treatment with erlotinib, temozolomide, and radiotherapy resulted in longer progression-free survival relative to a historical control study utilizing a regimen of temozolomide and radiotherapy alone (19.3 months vs. 14.1 months)³⁴¹. However, in a Phase 1/2 trial of erlotinib monotherapy in 11 patients with relapsed or refractory GBM or anaplastic astrocytoma, all patients showed disease progression and the drug showed significant toxicity³⁴². In addition, a Phase 2 trial of patients with recurrent or progressive GBM treated with erlotinib and sorafenib did not meet its objective of a 30% increase in overall survival time compared with historical controls; sorafenib was found to increase erlotinib clearance³⁴³.

Gefitinib

Assay findings association
EGFR
P596L

AREAS OF THERAPEUTIC USE

Gefitinib targets the tyrosine kinase EGFR and is FDA approved to treat non-small cell lung cancer (NSCLC) harboring exon 19 deletions or exon 21 (L858R) substitution mutations in EGFR. Please see the drug label for full prescribing information.

GENE ASSOCIATION

Activation of EGFR may predict sensitivity to therapies such as gefitinib. Clinical studies have consistently shown significant improvement in response rates and PFS for patients with EGFR-mutated non-small cell lung cancer (NSCLC) treated with gefitinib compared with chemotherapy^{337,344-349}, and responses have been reported for patients with EGFR-rearranged NSCLC³⁵⁰⁻³⁵¹.

SUPPORTING DATA

A clinical study of patients with glioblastoma (GBM)

treated with gefitinib or erlotinib found that 9/49 (18%) had tumor shrinkage of 25% or more; in this study, the extracellular domain EGFRvIII mutation was correlated with response³⁴⁰. A Phase 2 clinical study of gefitinib in patients with high-grade glioma (including GBM, anaplastic astrocytoma, and oligodendroglioma) reported 18% (5/28) disease stabilization; efficacy was not correlated with EGFR expression³⁵². However, a Phase 1/2 clinical trial of gefitinib combined with radiotherapy in 178 patients with GBM reported no overall survival benefit of added gefitinib, and EGFR expression was found to be of no prognostic value for patients treated with gefitinib plus radiotherapy³⁵³. A Phase 2 trial of preoperative gefitinib treatment in patients with recurrent GBM reported that although EGFR phosphorylation was decreased in treated patients as compared to the control group, measurement of 12 downstream molecules revealed no significant changes³⁵⁴.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Imatinib

Assay findings association
KIT
amplification

PDGFRA
amplification

AREAS OF THERAPEUTIC USE

Imatinib targets the BCR-ABL fusion protein, PDGFR, and KIT. It is FDA approved for the treatment of KIT-positive gastrointestinal stromal tumors (GIST), Ph+ chronic myeloid leukemia (CML) and acute lymphoblastic leukemia (ALL), myelodysplastic syndrome/myeloproliferative syndrome (MDS/MPS), aggressive systemic mastocytosis without a D816V KIT mutation, hypereosinophilic syndrome and/or chronic eosinophilic leukemia, and dermatofibrosarcoma protuberans. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical and preclinical data in KIT-mutated^{107-108,148,355}, KIT-amplified¹⁰⁶⁻¹⁰⁹, or KIT-expressing tumors^{111-116,356-357}, KIT activating alterations may confer sensitivity to imatinib. PDGFRA amplification

may predict sensitivity to tyrosine kinase inhibitors such as imatinib; a patient with Merkel cell carcinoma expressing PDGFRA achieved a complete response to imatinib¹⁴⁶.

SUPPORTING DATA

In a clinical study where patients with recurrent glioblastoma were given imatinib, 2/24 patients achieved a PR, 10 patients reported SD, and median OS and PFS was observed to be 6.2 and 3 months, respectively³⁵⁸. However, other Phase 2 clinical trials of imatinib have reported no anti-tumor activity, with a study of 231 patients with glioblastoma reporting a radiographic response rate of only 3.4%^{89,116}. In another Phase 2 study, imatinib plus hydroxyurea was shown to be well tolerated among patients with recurrent/progressive low-grade glioma, but had negligible antitumor activity³⁵⁹.

Nilotinib

Assay findings association
KIT
amplification

AREAS OF THERAPEUTIC USE

Nilotinib targets tyrosine kinases such as ABL (including BCR-ABL), PDGFRs, KIT, CSF1R, DDR1, and DDR2. It is FDA approved to treat newly diagnosed pediatric or adult patients with Philadelphia chromosome-positive (Ph+) chronic myeloid leukemia (CML) in chronic phase, adults with Ph+ CML in chronic or accelerated phase with resistance or intolerance to prior therapy including imatinib, and pediatric patients with Ph+ CML in chronic phase with resistance or intolerance to prior tyrosine-kinase inhibitor therapy. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical and preclinical data in KIT-mutated^{117,360-363}, KIT-amplified¹¹⁷, or KIT-expressing tumors³⁶⁴, KIT activating alterations may confer sensitivity to nilotinib.

SUPPORTING DATA

Clinical data on the efficacy of nilotinib for the treatment of CNS tumors are limited (PubMed, Jul 2022). Nilotinib

has been primarily investigated as a therapeutic option for the treatment of CML or gastrointestinal stromal tumors (GIST). In the context of CML, a Phase 3 clinical trial of Ph+ patients treated with imatinib or nilotinib (300 mg or 400 mg) reported progression-free survival (PFS) rates of 93% and 97-98% and overall survival (OS) rates of 93% and 94-97%, respectively, at 4 years³⁶⁵. For imatinib-resistant Japanese patients with CML, a Phase 2 trial reported a 47.8% major medical response rate to treatment with nilotinib at 12 months³⁶⁶. A Phase 3 clinical trial of single-agent nilotinib in 240 patients with advanced GIST who failed prior treatment with imatinib or sunitinib reported no significant difference in progression-free survival between nilotinib and the best supportive care, but did report increased overall survival for nilotinib-treated patients³⁶⁷. A Phase 2 trial has shown that nilotinib was well tolerated and suggested it may be particularly useful for treating patients with GIST harboring mutations in KIT exon 17³⁶⁸. Preclinical, cell-based assays have reported efficacy for nilotinib alone and in combination with additional therapies in the context of leiomyosarcoma and synovial sarcoma³⁶⁹.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Osimertinib

Assay findings association
EGFR
P596L

AREAS OF THERAPEUTIC USE

Osimertinib is an irreversible EGFR TKI that is selective for EGFR TKI-sensitizing mutations and the EGFR T790M mutation. It is FDA approved in various treatment settings for patients with non-small cell lung cancer (NSCLC) whose tumors have EGFR exon 19 deletions, exon 21 L858R mutations, or T790M mutations. Please see the drug label for full prescribing information.

GENE ASSOCIATION

EGFR TKI-sensitizing mutations or rearrangements and/or the EGFR T790M mutation may predict sensitivity to osimertinib in non-small cell lung cancer^{57,350,370-372}.

EGFR mutations may confer sensitivity to osimertinib on the basis of clinical responses to the third-generation TKI osimertinib for patients with EGFR-mutated glioma⁶⁴⁻⁶⁵ and additional clinical studies suggesting clinical benefit for these patients^{61,373}.

SUPPORTING DATA

Clinical benefit from osimertinib has been observed for cases of pediatric and adult patients with EGFR-altered glioma^{61,64-65,373-374}. Osimertinib has been studied primarily for the treatment of EGFR-mutated NSCLC. The Phase 3 FLAURA study reported that, relative to erlotinib or gefitinib, first-line osimertinib significantly increased both median PFS (mPFS; 18.9 vs. 10.2 months, HR=0.46) and median OS (38.6 vs. 31.8 months; HR=0.80) for patients with advanced non-small cell lung cancer (NSCLC) and activating, sensitizing EGFR mutations (specifically, exon 19 deletion or L858R)^{370,375}. In the Phase 3 ADAURA study, patients with early-stage (IB/II/IIIA) EGFR-mutated NSCLC experienced longer disease-free survival on osimertinib compared with placebo in the adjuvant setting (65.8 vs. 28.1 months, HR=0.27)³⁷⁶. A Phase 1 study reported that T790M-negative patients with

acquired EGFR TKI resistance experienced an ORR of 21% and a median PFS of 2.8 months⁵⁷. A Phase 1b/2 study evaluating osimertinib in combination with the CD73 inhibitor oleclumab for patients with advanced EGFR-mutated, T790M-negative NSCLC reported an ORR of 19% (4/21), a DCR of 81%, and mPFS of 11 months (Kim et al., 2021 AACR Abstract CT163). A Phase 2 study of osimertinib for EGFR-TKI-naïve patients with metastatic or recurrent NSCLC and uncommon EGFR mutations reported a 50% (18/36) ORR and an 89% (32/36) DCR with a median PFS of 8.2 months and a median duration of response of 11.2 months; patients harboring L861Q, G719X, or S768I mutations had ORRs of 78% (7/9), 53% (10/19), and 38% (3/8), respectively³⁷⁷. A Phase 2 trial of osimertinib in combination with bevacizumab versus osimertinib monotherapy for patients with untreated advanced non-small cell lung cancer (NSCLC) harboring EGFR del19 or L858R reported no difference in ORR (82% vs 86%) and median PFS (22.1 vs 20.2 months, HR 0.862 p=0.213)³⁷⁸. The Phase 2 BOOSTER study of osimertinib in combination with bevacizumab versus osimertinib monotherapy for patients with advanced NSCLC with EGFR-sensitizing mutations (exon 19 del or L858R) and L790M at progression on prior EGFR TKI reported no difference in ORR (55% vs 55%), median OS (24.0 vs 24.3 months, HR 1.03 p=0.91), or median PFS (15.4 vs 12.3 months, HR 0.96 p=0.83), although improved PFS was observed for the combination in the subgroup of current or former smokers (16.5 vs 8.4, HR 0.52) while nonsmokers had no benefit (HR 1.47)³⁷⁹. The Phase 1b TATTON study of osimertinib in combination with selumetinib, savolitinib, or durvalumab for patients with previously treated EGFR-mutated NSCLC reported ORRs of 42% (15/36), 44% (8/18), and 44% (10/23), respectively³⁸⁰.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Sorafenib

Assay findings association
KIT
amplification

AREAS OF THERAPEUTIC USE

Sorafenib is a kinase inhibitor that targets the RAF kinases, KIT, FLT3, RET, VEGFRs, and PDGFRs. It is FDA approved for the treatment of unresectable hepatocellular carcinoma, advanced renal cell carcinoma, and recurrent or metastatic differentiated thyroid carcinoma. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical and preclinical data in KIT-mutated³⁸¹⁻³⁸⁸ or KIT-expressing tumors¹¹⁸⁻¹²¹, KIT activating alterations may predict sensitivity to sorafenib.

SUPPORTING DATA

Phase 2 studies of sorafenib plus temozolomide report limited activity in patients with relapsed glioblastoma multiforme (GBM)³⁸⁹. A Phase 1/2 trial of temsirolimus in

combination with sorafenib in patients with glioblastoma was terminated at the Phase 2 interim analysis after patients failed to meet the primary endpoint of 6 month progression-free survival³⁹⁰. A Phase 2 trial of sorafenib and erlotinib in glioblastoma also did not meet its primary endpoint, and erlotinib clearance was increased by the addition of sorafenib³⁴³. In a Phase 1 trial in patients with high-grade glioma, the combination of sorafenib with radiation therapy (RT) and temozolomide (TMZ) resulted in increased toxicity and did not result in significant improvement in clinical efficacy compared with RT and TMZ alone³⁹¹. In a clinical study of sorafenib in pediatric patients with low-grade astrocytoma, one patient achieved a partial response (PR), one had stable disease (SD), and 9 patients had progressive disease; this study was terminated early due to unexpectedly high disease progression rates³⁹².

Sunitinib

Assay findings association
KIT
amplification

AREAS OF THERAPEUTIC USE

Sunitinib is a small-molecule tyrosine kinase inhibitor that targets PDGFRs, VEGFRs, KIT, FLT3, CSF-1R, and RET. It is FDA approved for the treatment of advanced or metastatic pancreatic neuroendocrine tumors, gastrointestinal stromal tumors (GISTs) in patients who have progressed on or are intolerant to imatinib, and advanced renal cell carcinoma (RCC) as well as for the adjuvant treatment of patients at high risk of recurrent RCC after nephrectomy. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical and preclinical data in KIT-mutated^{122,393-397} or KIT-expressing tumors¹²²⁻¹²³, KIT activating alterations may predict sensitivity to sunitinib.

SUPPORTING DATA

Phase 2 clinical trials of sunitinib in glioblastoma have reported no significant improvement in clinical outcome³⁹⁸⁻³⁹⁹. A Phase 2 trial that examined sunitinib treatment followed by radiation therapy in patients with glioblastoma reported a median progression-free survival (PFS) of 7.7 weeks, and a median overall survival (OS) of 12.8 weeks; 83.3% (10/12) of patients experienced neurological deterioration prior to radiation therapy⁴⁰⁰. Another Phase 2 study that examined daily sunitinib treatment in patients with glioblastoma reported no objective response in any of the 40 patients, with a median PFS of 2.2 months and a median OS of 9.2 months; five patients in the study had stable disease for more than six months⁴⁰¹.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually

updated and should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that

may require medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://www.clinicaltrials.gov). Or, visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

GENE EGFR

ALTERATION
P596L

RATIONALE
EGFR activating mutations, rearrangements, or amplification may predict sensitivity to EGFR-targeted therapies. Strategies to overcome

resistance to current agents include next-generation EGFR inhibitors and combination therapies.

NCT03783403

PHASE 1

A Study of CC-95251, a Monoclonal Antibody Directed Against SIRP α , in Subjects With Advanced Solid and Hematologic Cancers

TARGETS
CD20, EGFR, SIRP-alpha

LOCATIONS: Seoul (Korea, Republic of), Heidelberg (Australia), Melbourne (Australia), Manchester (United Kingdom), Edmonton (Canada), Rouen (France), Oregon, Marseille (France), Creteil (France), Nantes Cedex 01 (France)

NCT04946968

PHASE 2

Phase-2 Dacomitinib Study on Patients With EGFR-Driven Advanced Solid Tumours With Low EGFR-AS1 lncRNA Expr or Other Novel Emerging Biomarkers

TARGETS
ERBB4, EGFR, ERBB2

LOCATIONS: Singapore (Singapore)

NCT04670679

PHASE 1

A Dose Escalation/Expansion Study of ERAS-601 in Patients With Advanced or Metastatic Solid Tumors

TARGETS
SHP2, EGFR

LOCATIONS: Perth (Australia), Melbourne (Australia), Nevada, California, Texas, Massachusetts, New York, Tennessee, Florida

NCT04616196

PHASE 1/2

Study of NKTR 255 in Combination With Cetuximab in Solid Tumors

TARGETS
EGFR

LOCATIONS: California, Montana, Arizona, Minnesota, Illinois, Michigan, Texas, New York

NCT04720976

PHASE 1/2

JAB-3312 Activity in Adult Patients With Advanced Solid Tumors

TARGETS
MEK, SHP2, PD-1, EGFR, KRAS

LOCATIONS: Utah, California, Arizona, Minnesota, Illinois, Michigan, Oklahoma, Missouri, Indiana, Connecticut

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

CLINICAL TRIALS
NCT02800486
PHASE 2

Super Selective Intra-arterial Repeated Infusion of Cetuximab (Erbix) With Reirradiation for Treatment of Relapsed/Refractory GBM, AA, and AOA

TARGETS
 EGFR

LOCATIONS: New York

NCT02861898
PHASE 1/2

Super-selective Intra-arterial Repeated Infusion of Cetuximab for the Treatment of Newly Diagnosed Glioblastoma

TARGETS
 EGFR

LOCATIONS: New York

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

CLINICAL TRIALS
GENE
KIT
ALTERATION
 amplification

RATIONALE

KIT amplification or activating mutations may predict sensitivity to small molecule tyrosine kinase inhibitors. Also, because KIT activation

leads to activation of the PI3K-AKT-mTOR pathway, PI3K and mTOR pathway inhibitors may be relevant in a tumor with KIT activation.

NCT05024214
PHASE 1/2

Phase Ib/II Trial of Envafolelimab Plus Lenvatinib for Subjects With Solid Tumors

TARGETS

PD-L1, FGFRs, RET, PDGFRA, VEGFRs, KIT, FLT3, CSF1R

LOCATIONS: Hangzhou (China), Shanghai (China), Dongguan (China), Guangzhou (China), Zhuhai (China), Benbu (China), Zhengzhou (China), Jinan (China), Dalian (China), Tianjin (China)

NCT05098847
PHASE 2

Cryoablation Combined With Sintilimab Plus Lenvatinib In Previously Treated Unresectable Liver Metastasis From Solid Tumors

TARGETS

FGFRs, RET, PDGFRA, VEGFRs, KIT, PD-1

LOCATIONS: Shanghai (China)

NCT03564691
PHASE 1

Study of MK-4830 as Monotherapy and in Combination With Pembrolizumab (MK-3475) in Participants With Advanced Solid Tumors (MK-4830-001)

TARGETS

ITL4, FGFRs, RET, PDGFRA, VEGFRs, KIT, PD-1

LOCATIONS: Shanghai (China), Seoul (Korea, Republic of), Brisbane (Australia), Liverpool (Australia), Petah Tikva (Israel), Ramat Gan (Israel), Tel Aviv (Israel), Haifa (Israel), Warszawa (Poland), Gdansk (Poland)

NCT04803318
PHASE 2

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

TARGETS

mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK

LOCATIONS: Guangzhou (China)

NCT04008797
PHASE 1

A Study of E7386 in Combination With Other Anticancer Drug in Participants With Solid Tumor

TARGETS

CBP, Beta-catenin, FGFRs, RET, PDGFRA, VEGFRs, KIT

LOCATIONS: Kurume (Japan), Matsuyama (Japan), Seodaemun (Korea, Republic of), Osakasayama (Japan), Nagoya (Japan), Chuo-Ku (Japan), Koto-ku (Japan), Chiba (Japan), Kashiwa (Japan), Hidaka (Japan)

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

 Electronically signed by Erik Williams, M.D. | 11 January 2023
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | www.rochefoundationmedicine.com

 Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

CLINICAL TRIALS
NCT04977453
PHASE 1/2

GI-101 as a Single Agent or in Combination With Pembrolizumab, Lenvatinib or Local Radiotherapy in Advanced Solid Tumors

TARGETS
 FGFRs, RET, PDGFRA, VEGFRs, KIT, PD-1, CTLA-4

LOCATIONS: Daejeon (Korea, Republic of), Suwon-si (Korea, Republic of), Seoul (Korea, Republic of)

NCT03025893
PHASE 2/3

A Phase II/III Study of High-dose, Intermittent Sunitinib in Patients With Recurrent Glioblastoma Multiforme

TARGETS
 FLT3, VEGFRs, CSF1R, KIT, RET

LOCATIONS: Groningen (Netherlands), Nijmegen (Netherlands), Amsterdam (Netherlands)

NCT04729348
PHASE 2

Pembrolizumab And Lenvatinib In Leptomeningeal Metastases

TARGETS
 PD-1, KIT, VEGFRs, FGFRs, PDGFRA, RET

LOCATIONS: Massachusetts

NCT02379416
PHASE 1

Combination Nilotinib and Paclitaxel in Adults With Relapsed Solid Tumors

TARGETS
 ABL, KIT

LOCATIONS: Maryland

NCT05064280
PHASE 2

Phase II Study of Pembrolizumab in Combination With Lenvatinib in Patients With TNBC, NSCLC, and Other Tumor Types and Brain Metastases

TARGETS
 PD-1, KIT, VEGFRs, FGFRs, PDGFRA, RET

LOCATIONS: Texas

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

CLINICAL TRIALS
GENE
MTAP
RATIONALE

MTAP loss may predict sensitivity to MAT2A inhibitors, or to inhibitors that target PRMT5

when in complex with MTA.

ALTERATION

loss

NCT05245500
PHASE 1/2

Phase 1/2 Study of MRTX1719 in Solid Tumors With MTAP Deletion

TARGETS

PRMT5-MTA

LOCATIONS: Colorado, Massachusetts, New York, Tennessee, Texas

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

CLINICAL TRIALS

GENE

PDGFRA

RATIONALE

PDGFRA amplification may predict sensitivity to imatinib and to anti-PDGFRA antibodies.

ALTERATION

amplification

NCT01738139**PHASE 1**

Ipilimumab and Imatinib Mesylate in Advanced Cancer

TARGETS

KIT, ABL, CTLA-4

LOCATIONS: Texas

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

APPENDIX
Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

BCL6
P472R

BRIP1
S1001N

CCNE1
R27H

GNAS
R16C

MAPK1
N238K

MRE11A
G409del

PIK3C2B
S20_R21del

PTEN
L152Q

ROS1
D2213E

SGK1
H72R

TGFBR2
rearrangement

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

APPENDIX

Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B or WTX)	
APC	AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX
AURKA	AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2
BCL6	BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1
BTG2	BTK	CALR	CARD11	CASP8	CBFB	CBL	CCND1	CCND2
CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73	CDH1
CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B	CDKN2C
CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R	CTCF
CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1	DDR2
DIS3	DNMT3A	DOT1L	EED	EGFR	EMSY (C11orf30)	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12	FGF14
FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3	FGFR4
FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3	GATA4
GATA6	GID4 (C17orf39)	GNA11	GNA13	GNAS	GNAS	GRM3	GSK3B	H3-3A (H3F3A)
HDAC1	HGF	HNFI1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDM5A	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11 (MRE11A)	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD2 (WHSC1 or MMSET)	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3	NTRK3
P2RY8	PALB2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCCD1 (PD-1)	PDCCD1LG2 (PD-L2)
PDGFRA	PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1
PMS2	POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI
PRKN (PARK2)	PTCH1	PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51
RAD51B	RAD51C	RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10
REL	RET	RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC
SDHD	SETD2	SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO
SNCAIP	SOC1	SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3
STK11	SUFU	SYK	TBX3	TEK	TENT5C (FAM46C)	TET2	TET2	TGFB2
TIPARP	TNFAIP3	TNFRSF14	TP53	TSC1	TSC2	TYRO3	U2AF1	VEGFA
VHL	WT1	XPO1	XRCC2	ZNF217	ZNF703			

DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSP02	SDC4	SLC34A2	TERC*	TERT**	TPRSS2

*TERC is an NCRNA

**Promoter region of TERT is interrogated

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS


Homologous Recombination status
Loss of Heterozygosity (LOH) score
Microsatellite (MS) status
Tumor Mutational Burden (TMB)

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

ORDERED TEST # ORD-1537947-01

APPENDIX

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Ciplstraat 3, 2440 Geel, Belgium. 

ABOUT FOUNDATIONONE CDx

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:
www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLE

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes

detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for *ERBB2* and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical

methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials

Ranking of Therapies in Summary Table

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

1. In the fraction-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

APPENDIX

About FoundationOne®CDx

- analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.
- TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
 - Homologous Recombination status may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas (Coleman et al., 2017; 28916367). Samples with deleterious *BRCA1/2* alteration and/or Loss of Heterozygosity (LOH) score $\geq 16\%$ will be reported as "HRD Positive" and samples with absence of these findings will be reported as "HRD Not Detected," agnostic of potential secondary *BRCA1/2* reversion alterations. Certain potentially deleterious missense or small in-frame deletions in *BRCA1/2* may not be classified as deleterious and, in the absence of an elevated LOH profile, samples with such mutations may be classified as "HRD Not Detected." A result of "HRD Not Detected" does not rule out the presence of a *BRCA1/2* alteration or an elevated LOH profile outside the assay performance characteristic limitations.
 - The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and

- extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
- Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
 - Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine whether the patient is a candidate for biopsy.
 - Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of *HER2* overexpression as 20% in breast cancer. Based on the F1CDx *HER2* CDx concordance study, approximately 10% of *HER2* amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal

hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

*Interquartile Range = 1st Quartile to 3rd Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1537947-01

APPENDIX

About FoundationOne®CDx

tumor sequencing is germline or somatic.
 Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D (MLL2)*, *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking

into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

REFERENCE SEQUENCE INFORMATION

Sequence data is mapped to the human genome, Genome Reference Consortium Human Build 37 (GRCh37), also known as hg19.

MR Suite Version (RG) 7.4.0

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of the data. The median exon coverage for this sample is 560x. The suitability of use.

ORDERED TEST # **ORD-1537947-01**
APPENDIX
References

1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) PMID: 25392179
2. Kroemer G, et al. Oncoimmunology (2015) PMID: 26140250
3. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
4. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
5. Ayers et al., 2016; ASCO-SITC Abstract P60
6. Martinez R, et al. Oncology (2004) PMID: 15331927
7. Martinez R, et al. J. Cancer Res. Clin. Oncol. (2005) PMID: 15672285
8. Martinez R, et al. Cancer Genet. Cytogenet. (2007) PMID: 17498554
9. Szybka M, et al. Clin. Neuropathol. (2010) PMID: 12908754
10. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
11. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
12. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
13. Boland CR, et al. Cancer Res. (1998) PMID: 9823339
14. Pawlik TM, et al. Dis. Markers (2004) PMID: 15528785
15. Boland CR, et al. Gastroenterology (2010) PMID: 20420947
16. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
17. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
18. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
19. Cristescu R, et al. Science (2018) PMID: 30309915
20. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
21. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
22. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
23. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
24. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
25. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
26. Zhao J, et al. Nat. Med. (2019) PMID: 30742119
27. Touat M, et al. Nature (2020) PMID: 32322066
28. Bouffett E, et al. J. Clin. Oncol. (2016) PMID: 27001570
29. Johanns TM, et al. Cancer Discov (2016) PMID: 27258356
30. Lukas RV, et al. J. Neurooncol. (2018) PMID: 30073642
31. Chalmers ZR, et al. Genome Med (2017) PMID: 28420421
32. Patel RR, et al. Pediatr Blood Cancer (2020) PMID: 32386112
33. Johnson A, et al. Oncologist (2017) PMID: 28912153
34. Draaisma K, et al. Acta Neuropathol Commun (2015) PMID: 26699864
35. Wang L, et al. BMC Cancer (2020) PMID: 32164609
36. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
37. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
38. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
39. Rizvi NA, et al. Science (2015) PMID: 25765070
40. Johnson BE, et al. Science (2014) PMID: 24336570
41. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
42. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
43. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
44. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
45. Nature (2012) PMID: 22810696
46. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
47. Rosell R, et al. Lancet Oncol. (2012) PMID: 22285168
48. Douillard JY, et al. Br. J. Cancer (2014) PMID: 24263064
49. Hayashi T, et al. Hum Pathol (2020) PMID: 32673682
50. Cao L, et al. Onco Targets Ther (2018) PMID: 29780256
51. Yang TY, et al. J. Clin. Oncol. (2011) PMID: 21422421
52. Sequist LV, et al. J. Clin. Oncol. (2013) PMID: 23816960
53. Qin BD, et al. Onco Targets Ther (2018) PMID: 30127622
54. Frega S, et al. J Thorac Oncol (2016) PMID: 27131295
55. Long X, et al. Onco Targets Ther (2020) PMID: 33116645
56. Mok TS, et al. J. Clin. Oncol. (2018) PMID: 29864379
57. Jänne PA, et al. N. Engl. J. Med. (2015) PMID: 25923549
58. Hong MH, et al. Cancer (2020) PMID: 32749686
59. Kim HS, et al. Oncotarget (2015) PMID: 26462025
60. Kim HS, et al. Clin. Cancer Res. (2015) PMID: 25424851
61. Mondal G, et al. Acta Neuropathol (2020) PMID: 32303840
62. Cavalieri S, et al. Eur. J. Cancer (2018) PMID: 29734047
63. Chi AS, et al. JCO Precis Oncol (2020) PMID: 32923886
64. Makhlin I, et al. CNS Oncol (2019) PMID: 31769726
65. Goyal A, et al. World Neurosurg (2021) PMID: 33940677
66. Jonsson P, et al. Clin. Cancer Res. (2019) PMID: 31263031
67. Brennan CW, et al. Cell (2013) PMID: 24120142
68. Ceccarelli M, et al. Cell (2016) PMID: 26824661
69. Thomas AA, et al. Neuro-oncology (2017) PMID: 28472509
70. Lee JC, et al. PLoS Med. (2006) PMID: 17177598
71. Vivanco I, et al. Cancer Discov (2012) PMID: 22588883
72. Yan et al. 2020; DOI:10.1200/PO.19.00385
73. Srividya MR, et al. J. Clin. Pathol. (2010) PMID: 20702468
74. Das P, et al. J Clin Neurosci (2011) PMID: 20888234
75. Smith JS, et al. J. Natl. Cancer Inst. (2001) PMID: 11504770
76. Ciardiello F, et al. N. Engl. J. Med. (2008) PMID: 18337605
77. Foster JM, et al. World J Surg Oncol (2010) PMID: 20942962
78. Cai CQ, et al. Oncogene (2008) PMID: 18193092
79. Stabile LP, et al. Cancer Res. (2005) PMID: 15735034
80. Zhang W, et al. J Thorac Oncol (2006) PMID: 17409930
81. Siegfried JM, et al. J Thorac Oncol (2012) PMID: 22258476
82. U M, et al. PLoS Comput. Biol. (2014) PMID: 24743239
83. Cho J, et al. Mol. Cancer (2014) PMID: 24894453
84. Hama T, et al. Oncologist (2009) PMID: 19726454
85. Tam IY, et al. Mol. Cancer Ther. (2009) PMID: 19671738
86. Kancha RK, et al. Clin. Cancer Res. (2009) PMID: 19147750
87. Chen YR, et al. Oncogene (2006) PMID: 16205628
88. Ymer SI, et al. Cancers (Basel) (2011) PMID: 24212795
89. Razis E, et al. Clin. Cancer Res. (2009) PMID: 19789313
90. Wang H, et al. Neoplasia (2011) PMID: 21532887
91. Kim N, et al. Int. J. Cancer (2019) PMID: 31290142
92. Sueangoen N, et al. Cell Biosci (2020) PMID: 32190291
93. Lundby A, et al. Cell (2019) PMID: 31585087
94. Evans EK, et al. Sci Transl Med (2017) PMID: 29093181
95. Abbaspour Babaei M, et al. Drug Des Devel Ther (2016) PMID: 27536065
96. Ramaswamy A, et al. J Gastrointest Oncol (2016) PMID: 27563456
97. Demetri GD, et al. Lancet (2013) PMID: 23177515
98. Gotlib J, et al. N. Engl. J. Med. (2016) PMID: 27355533
99. Jawhar M, et al. Blood (2017) PMID: 28424161
100. Xu X, et al. Int J Clin Exp Pathol (2014) PMID: 25031773
101. Gotlib J, et al. Blood (2005) PMID: 15972446
102. Luo C, et al. Onco Targets Ther (2017) PMID: 29066909
103. Janku F, et al. ESMO Open (2022) PMID: 35753087
104. Si L, et al. J. Clin. Oncol. (2012) PMID: 22162580
105. Parikh SA, et al. Leuk Lymphoma (2010) PMID: 20038218
106. Wei X, et al. Oncol. Res. (2019) PMID: 30075827
107. Hodi FS, et al. J. Clin. Oncol. (2013) PMID: 23775962
108. Carvajal RD, et al. JAMA (2011) PMID: 21642685
109. Guo J, et al. J. Clin. Oncol. (2011) PMID: 21690468
110. Debiec-Rychter M, et al. Gastroenterology (2005) PMID: 15685537
111. Dematteo RP, et al. Lancet (2009) PMID: 19303137
112. Faivre S, et al. J. Clin. Oncol. (2005) PMID: 16135502
113. Hotte SJ, et al. J. Clin. Oncol. (2005) PMID: 15659505
114. Alcedo JC, et al. Head Neck (2004) PMID: 15350030
115. Brandwein JM, et al. Leukemia (2011) PMID: 21403650
116. Reardon DA, et al. Br. J. Cancer (2009) PMID: 19904263
117. Lee SJ, et al. Oncologist (2015) PMID: 26424760
118. Llovet JM, et al. Clin. Cancer Res. (2012) PMID: 22374331
119. Zhang HL, et al. Clin Genitourin Cancer (2013) PMID: 23058498
120. Seino S, et al. Gastroenterology (2014) PMID: 25450081
121. Li XF, et al. Med. Oncol. (2009) PMID: 18846437
122. Minor DR, et al. Clin. Cancer Res. (2012) PMID: 22261812
123. Mahipal A, et al. Melanoma Res. (2012) PMID: 23114504
124. Cancer Genome Atlas Research Network, et al. N. Engl. J. Med. (2015) PMID: 26061751
125. Nobusawa S, et al. Neuropathology (2011) PMID: 21382095
126. Joensuu H, et al. J. Pathol. (2005) PMID: 16021678
127. Burford A, et al. PLoS ONE (2013) PMID: 23990986
128. Holtkamp N, et al. Neuro-oncology (2007) PMID: 17504929
129. Puputti M, et al. Mol. Cancer Res. (2006) PMID: 17189383
130. Skardelly M, et al. Transl Oncol (2009) PMID: 19701495
131. Int. J. Biochem. Cell Biol. (1999) PMID: 10582339
132. Semin. Oncol. (2004) PMID: 15175998
133. Gao J, et al. Sci Signal (2013) PMID: 23550210
134. Zack TI, et al. Nat. Genet. (2013) PMID: 24071852
135. Beroukhir R, et al. Nature (2010) PMID: 20164920
136. Arefi M, et al. Int. J. Hematol. (2012) PMID: 22806436
137. Baccarani M, et al. Haematologica (2007) PMID: 17666373
138. Cassier PA, et al. Clin. Cancer Res. (2012) PMID: 22718859
139. Chalmers ZR, et al. Blood Cancer J (2015) PMID: 25658984
140. Cools J, et al. N. Engl. J. Med. (2003) PMID: 12660384
141. Curtis CE, et al. Br. J. Haematol. (2007) PMID: 17555450
142. Debiec-Rychter M, et al. Eur. J. Cancer (2004) PMID: 15010069
143. Dileo P, et al. Int. J. Cancer (2011) PMID: 20473908
144. Fanta PT, et al. J. Clin. Oncol. (2015) PMID: 24638008
145. Florian S, et al. Leuk. Res. (2006) PMID: 16406018
146. Frenard C, et al. JAAD Case Rep (2016) PMID: 27051816
147. Griffin JH, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) PMID: 12808148
148. Heinrich MC, et al. J. Clin. Oncol. (2003) PMID: 14645423
149. Helbig G, et al. Br. J. Haematol. (2009) PMID: 19120352
150. Helbig G, et al. Am. J. Hematol. (2014) PMID: 24009127
151. Hus M, et al. Leuk. Res. (2011) PMID: 21093052
152. Ikezoe T, et al. Leuk. Res. (2010) PMID: 20303172
153. Intermesoli T, et al. Br. J. Haematol. (2009) PMID: 19735261
154. Jain N, et al. Leuk. Res. (2009) PMID: 19013640
155. Jovanovic JV, et al. Blood (2007) PMID: 17299092
156. Kang HJ, et al. Acta Oncol (2012) PMID: 22150077

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

ORDERED TEST # **ORD-1537947-01**
APPENDIX **References**

157. Klion AD, et al. Blood (2004) PMID: 14504092
158. Kobayashi M, et al. Respiratory (2009) PMID: 19192229
159. Kocáková I, et al. Klin Onkol (2014) PMID: 24635438
160. Metzgeroth G, et al. Br. J. Haematol. (2008) PMID: 18950453
161. Murayama Y, et al. World J Gastrointest Oncol (2012) PMID: 22645636
162. Ogbogu PU, et al. J. Allergy Clin. Immunol. (2009) PMID: 19910029
163. Ohnishi H, et al. Br. J. Haematol. (2006) PMID: 16856885
164. Pardanani A, et al. Blood (2003) PMID: 12842979
165. Pardanani A, et al. Blood (2004) PMID: 15284118
166. Qu SQ, et al. Oncotarget (2016) PMID: 27120808
167. Score J, et al. Leukemia (2006) PMID: 16498388
168. Shah S, et al. J Hematol Oncol (2014) PMID: 24669761
169. Sugimoto Y, et al. Cancer Genet (2015) PMID: 26319757
170. Volz HC, et al. Int. J. Cardiol. (2011) PMID: 20609486
171. von Bubnoff N, et al. Leukemia (2005) PMID: 15618966
172. Walz C, et al. Genes Chromosomes Cancer (2006) PMID: 16845659
173. Yoo C, et al. Cancer Res Treat (2016) PMID: 26130666
174. Al-Riyami AZ, et al. Leuk. Lymphoma (2013) PMID: 23157309
175. Lierman E, et al. Blood (2006) PMID: 16645167
176. Lierman E, et al. Leukemia (2009) PMID: 19212337
177. Metzgeroth G, et al. Leukemia (2012) PMID: 21818111
178. Roubaud G, et al. Ann. Oncol. (2012) PMID: 22294526
179. von Bubnoff N, et al. Oncogene (2011) PMID: 20972453
180. Hochhaus A, et al. J. Cancer Res. Clin. Oncol. (2013) PMID: 24057647
181. Tabouret E, et al. Leuk. Res. (2011) PMID: 20832858
182. Dewaele B, et al. Clin. Cancer Res. (2008) PMID: 18794084
183. Weisberg E, et al. Gastroenterology (2006) PMID: 17087936
184. Brohl AS, et al. Clin Sarcoma Res (2015) PMID: 26396737
185. Grellety T, et al. Future Sci OA (2015) PMID: 28031906
186. Kollár A, et al. Clin Sarcoma Res (2014) PMID: 25905001
187. Jaku et al., 2017; ASCO Abstract 2515
188. Phillips JJ, et al. Brain Pathol. (2013) PMID: 23438035
189. Motomura K, et al. J. Neuropathol. Exp. Neurol. (2013) PMID: 23242283
190. Sottoriva A, et al. Proc. Natl. Acad. Sci. U.S.A. (2013) PMID: 23412337
191. Alentorn A, et al. Neuro-oncology (2012) PMID: 23074200
192. Song K, et al. Am J Cancer Res (2018) PMID: 29888103
193. Szerlip NJ, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) PMID: 22323597
194. Andrae J, et al. Genes Dev. (2008) PMID: 18483217
195. Flavahan WA, et al. Nature (2016) PMID: 26700815
196. Roszik J, et al. Sci Rep (2016) PMID: 26787600
197. Verhaak RG, et al. Cancer Cell (2010) PMID: 20129251
198. Koschmann C, et al. Oncotarget (2016) PMID: 27582545
199. Puget S, et al. PLoS ONE (2012) PMID: 22389665
200. Kalev P, et al. Cancer Cell (2021) PMID: 33450196
201. Marjon K, et al. Cell Rep (2016) PMID: 27068473
202. Mavrakis KJ, et al. Science (2016) PMID: 26912361
203. Kryukov GV, et al. Science (2016) PMID: 26912360
204. Heist et al., 2019; AACR-NCI-EORTC Abstract B116
205. Guccione E, et al. Nat. Rev. Mol. Cell Biol. (2019) PMID: 31350521
206. Fedoriv A, et al. Cancer Cell (2019) PMID: 31257072
207. Srouf N, et al. Cancer Cell (2019) PMID: 31287990
208. Gao G, et al. Nucleic Acids Res. (2019) PMID: 30916320
209. Smith CR, et al. J Med Chem (2022) PMID: 35041419
210. Hansen LJ, et al. Cancer Res. (2019) PMID: 31040154
211. Tang B, et al. Cancer Res. (2018) PMID: 29844120
212. Munshi PN, et al. Oncologist (2014) PMID: 24928612
213. de Oliveira SF, et al. PLoS ONE (2016) PMID: 26751376
214. Lubin M, et al. PLoS ONE (2009) PMID: 19478948
215. Tang B, et al. Cancer Biol. Ther. (2012) PMID: 22825330
216. Collins CC, et al. Mol. Cancer Ther. (2012) PMID: 22252602
217. Bertino JR, et al. Cancer Biol. Ther. (2011) PMID: 21301207
218. Coulthard SA, et al. Mol. Cancer Ther. (2011) PMID: 21282358
219. Miyazaki S, et al. Int. J. Oncol. (2007) PMID: 17912432
220. Efferth T, et al. Blood Cells Mol. Dis. () PMID: 11987241
221. Kindler HL, et al. Invest New Drugs (2009) PMID: 18618081
222. Alhalabi O, et al. Nat Commun (2022) PMID: 35379845
223. Wei R, et al. Sci Rep (2016) PMID: 27929028
224. Zhao M, et al. BMC Genomics (2016) PMID: 27556634
225. Kirovski G, et al. Am. J. Pathol. (2011) PMID: 21356366
226. Huang HY, et al. Clin. Cancer Res. (2009) PMID: 19887491
227. Marcé S, et al. Clin. Cancer Res. (2006) PMID: 16778103
228. Meyer S, et al. Exp. Dermatol. (2010) PMID: 20500769
229. Wild PJ, et al. Arch Dermatol (2006) PMID: 16618867
230. Kim J, et al. Genes Chromosomes Cancer (2011) PMID: 21412930
231. Li CF, et al. Oncotarget (2014) PMID: 25426549
232. He HL, et al. Medicine (Baltimore) (2015) PMID: 26656376
233. Su CY, et al. Eur J Surg Oncol (2014) PMID: 24969958
234. Mirebeau D, et al. Haematologica (2006) PMID: 16818274
235. Becker AP, et al. Pathobiology (2015) PMID: 26088413
236. Snezhkina AV, et al. Oxid Med Cell Longev (2016) PMID: 27433286
237. Bistulfi G, et al. Oncotarget (2016) PMID: 26910893
238. Antonopoulou K, et al. J. Invest. Dermatol. (2015) PMID: 25407435
239. Maccioni L, et al. BMC Cancer (2013) PMID: 23816148
240. Hyland PL, et al. Int J Epidemiol (2016) PMID: 26635288
241. Lin X, et al. Cancer Sci. (2017) PMID: 27960044
242. Zhi L, et al. J Cancer (2016) PMID: 27994653
243. Gu F, et al. Br. J. Cancer (2013) PMID: 23361049
244. Limm K, et al. PLoS ONE (2016) PMID: 27479139
245. Tang B, et al. G3 (Bethesda) (2014) PMID: 25387827
246. Limm K, et al. Eur. J. Cancer (2013) PMID: 23265702
247. Stevens AP, et al. J. Cell. Biochem. (2009) PMID: 19097084
248. Limm K, et al. Eur. J. Cancer (2014) PMID: 25087184
249. Konecny GE, et al. Clin. Cancer Res. (2011) PMID: 21278246
250. Katsumi Y, et al. Biochem. Biophys. Res. Commun. (2011) PMID: 21871868
251. Cen L, et al. Neuro-oncology (2012) PMID: 22711607
252. Logan JE, et al. Anticancer Res. (2013) PMID: 23898052
253. Fennell DA, et al. Lancet Oncol (2022) PMID: 35157829
254. Elvin JA, et al. Oncologist (2017) PMID: 28283584
255. Gao J, et al. Curr Oncol (2015) PMID: 26715889
256. Gopalan et al., 2014; ASCO Abstract 8077
257. Peguero et al., 2016; ASCO Abstract 2528
258. Konecny et al., 2016; ASCO Abstract 5557
259. DeMichele A, et al. Clin. Cancer Res. (2015) PMID: 25501126
260. Finn RS, et al. Lancet Oncol. (2015) PMID: 25524798
261. Infante JR, et al. Clin. Cancer Res. (2016) PMID: 27542767
262. Johnson DB, et al. Oncologist (2014) PMID: 24797823
263. Van Maercken T, et al. Mol. Cancer Ther. (2011) PMID: 21460101
264. Gamble LD, et al. Oncogene (2012) PMID: 21725357
265. Flaherty KT, et al. Clin. Cancer Res. (2012) PMID: 22090362
266. Dickson MA, et al. J. Clin. Oncol. (2013) PMID: 23569312
267. Su D, et al. Nat Commun (2019) PMID: 31700061
268. Tramontana TF, et al. JCO Precis Oncol (2020) PMID: 32923894
269. Weber RG, et al. Oncogene (2007) PMID: 16909113
270. Nakamura M, et al. Brain Pathol. (2001) PMID: 11303791
271. Chakravarti A, et al. Clin. Cancer Res. (2001) PMID: 11489817
272. Feng J, et al. Cancer (2012) PMID: 21713760
273. Raabe EH, et al. Clin. Cancer Res. (2011) PMID: 21636552
274. Liu W, et al. J. Exp. Clin. Cancer Res. (2011) PMID: 21843312
275. Quelle DE, et al. Cell (1995) PMID: 8521522
276. Mutat. Res. (2005) PMID: 15878778
277. Gazzeri S, et al. Oncogene (1998) PMID: 9484839
278. Oncogene (1999) PMID: 10498883
279. Sherr CJ, et al. Cold Spring Harb. Symp. Quant. Biol. (2005) PMID: 16869746
280. Ozenne P, et al. Int. J. Cancer (2010) PMID: 20549699
281. Ruas M, et al. Oncogene (1999) PMID: 10498896
282. Jones R, et al. Cancer Res. (2007) PMID: 17909018
283. Haferkamp S, et al. Aging Cell (2008) PMID: 18843795
284. Huot TJ, et al. Mol. Cell. Biol. (2002) PMID: 12417717
285. Rizo H, et al. J. Biol. Chem. (2001) PMID: 11518711
286. Gombart AF, et al. Leukemia (1997) PMID: 9324288
287. Yang R, et al. Cancer Res. (1995) PMID: 7780957
288. Parry D, et al. Mol. Cell. Biol. (1996) PMID: 8668202
289. Greenblatt MS, et al. Oncogene (2003) PMID: 12606942
290. Yarbrough WG, et al. J. Natl. Cancer Inst. (1999) PMID: 10491434
291. Poi MJ, et al. Mol. Carcinog. (2001) PMID: 11255261
292. Byeon JJ, et al. Mol. Cell (1998) PMID: 9660926
293. Kannengiesser C, et al. Hum. Mutat. (2009) PMID: 19260062
294. Lal G, et al. Genes Chromosomes Cancer (2000) PMID: 10719365
295. Koh J, et al. Nature (1995) PMID: 7777061
296. McKenzie HA, et al. Hum. Mutat. (2010) PMID: 20340136
297. Miller PJ, et al. Hum. Mutat. (2011) PMID: 21462282
298. Kutscher CL, et al. Physiol. Behav. (1977) PMID: 905385
299. Scaini MC, et al. Hum. Mutat. (2014) PMID: 24659262
300. Jenkins NC, et al. J. Invest. Dermatol. (2013) PMID: 23190892
301. Walker GJ, et al. Int. J. Cancer (1999) PMID: 10389768
302. Rutter JL, et al. Oncogene (2003) PMID: 12853981
303. Itahana K, et al. Cancer Cell (2008) PMID: 18538737
304. Zhang Y, et al. Mol. Cell (1999) PMID: 10360174
305. Zhang Y, et al. Cell (1998) PMID: 9529249
306. Jafri M, et al. Cancer Discov (2015) PMID: 25873077
307. Whelan AJ, et al. N Engl J Med (1995) PMID: 7666917
308. Adv Exp Med Biol (2010) PMID: 20687502
309. Hogg D, et al. J Cutan Med Surg (1998) PMID: 9479083
310. De Unamuno B, et al. Melanoma Res (2018) PMID: 29543703
311. Soura E, et al. J Am Acad Dermatol (2016) PMID: 26892650
312. Huerta C, et al. Acta Derm Venereol (2018) PMID: 29543703

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

ORDERED TEST # ORD-1537947-01

APPENDIX
References

- 29405243
313. Kaufman DK, et al. Neurology (1993) PMID: 8414022
314. Bahuau M, et al. Cancer Res (1998) PMID: 9622062
315. Chan AK, et al. Clin Neuropathol () PMID: 28699883
316. Nat Rev Clin Oncol (2017) PMID: 27245281
317. Duperret EK, et al. Mol Ther (2018) PMID: 29249395
318. Chiappori AA, et al. Ann Oncol (2015) PMID: 25467017
319. Killela PJ, et al. Proc. Natl. Acad. Sci. U.S.A. (2013) PMID: 23530248
320. Killela PJ, et al. Oncotarget (2014) PMID: 24722048
321. Nonoguchi N, et al. Acta Neuropathol. (2013) PMID: 23955565
322. Liu X, et al. Cell Cycle (2013) PMID: 23603989
323. Koelsche C, et al. Acta Neuropathol. (2013) PMID: 24154961
324. Arita H, et al. Acta Neuropathol. (2013) PMID: 23764841
325. Reitman ZJ, et al. Acta Neuropathol. (2013) PMID: 24217890
326. Shay JW, et al. Semin. Cancer Biol. (2011) PMID: 22015685
327. Shay JW, et al. Eur. J. Cancer (1997) PMID: 9282118
328. Kim NW, et al. Science (1994) PMID: 7605428
329. Hanahan D, et al. Cell (2000) PMID: 10647931
330. Horn S, et al. Science (2013) PMID: 23348503
331. Huang FW, et al. Science (2013) PMID: 23348506
332. Vinagre J, et al. Nat Commun (2013) PMID: 23887589
333. Weller M, et al. Nat Rev Clin Oncol (2021) PMID: 33293629
334. Louis DN, et al. Neuro Oncol (2021) PMID: 34185076
335. Cappuzzo F, et al. Lancet Oncol. (2010) PMID: 20493771
336. Zhong WZ, et al. J. Clin. Oncol. (2019) PMID: 31194613
337. Petrelli F, et al. Clin Lung Cancer (2012) PMID: 22056888
338. Hainsworth JD, et al. J. Clin. Oncol. (2018) PMID: 29320312
339. Nie KK, et al. Chin Med J (Engl) (2018) PMID: 29998897
340. Mellingerhoff IK, et al. N. Engl. J. Med. (2005) PMID: 16282176
341. Prados MD, et al. J. Clin. Oncol. (2009) PMID: 19075262
342. Kesavabhotla K, et al. J. Exp. Ther. Oncol. (2012) PMID: 22946346
343. Peereboom DM, et al. Neuro-oncology (2013) PMID: 23328813
344. Han JY, et al. J. Clin. Oncol. (2012) PMID: 22370314
345. Maemondo M, et al. N. Engl. J. Med. (2010) PMID: 20573926
346. Mitsudomi T, et al. Lancet Oncol. (2010) PMID: 20022809
347. Mok TS, et al. N. Engl. J. Med. (2009) PMID: 19692680
348. Qi WX, et al. Curr Med Res Opin (2015) PMID: 25329826
349. Zhao H, et al. J Thorac Oncol (2015) PMID: 25546556
350. Wang J, et al. Int. J. Cancer (2019) PMID: 30255937
351. Baik CS, et al. J Thorac Oncol (2015) PMID: 26398831
352. Franceschi E, et al. Br. J. Cancer (2007) PMID: 17353924
353. Chakravarti A, et al. Int. J. Radiat. Oncol. Biol. Phys. (2013) PMID: 23182702
354. Hegi ME, et al. Mol. Cancer Ther. (2011) PMID: 21471286
355. Debiec-Rychter M, et al. Eur. J. Cancer (2006) PMID: 16624552
356. Kamenz T, et al. World J. Gastroenterol. (2006) PMID: 16570351
357. Wang YY, et al. Proc. Natl. Acad. Sci. U.S.A. (2005) PMID: 15650049
358. Hassler MR, et al. Springerplus (2014) PMID: 25674429
359. Reardon DA, et al. Cancer (2012) PMID: 22371319
360. Carvajal RD, et al. Clin. Cancer Res. (2015) PMID: 25695690
361. Hochhaus A, et al. J. Cancer Res. Clin. Oncol. (2015) PMID: 26002753
362. Blay JY, et al. Lancet Oncol. (2015) PMID: 25882987
363. Kajimoto N, et al. Int J Clin Exp Pathol (2015) PMID: 26722383
364. Sako H, et al. PLoS ONE (2014) PMID: 25221952
365. Hughes TP, et al. Blood (2014) PMID: 24335106
366. Takahashi N, et al. Biomark Res (2014) PMID: 24650752
367. Reichardt P, et al. Ann. Oncol. (2012) PMID: 22357255
368. Cauchi C, et al. Cancer Chemother. Pharmacol. (2012) PMID: 22119758
369. Villar VH, et al. PLoS ONE (2012) PMID: 22662203
370. Soria JC, et al. N. Engl. J. Med. (2018) PMID: 29151359
371. Alanazi A, et al. Lung Cancer Manag (2020) PMID: 33318755
372. Kim et al., 2021; DOI: 10.1200/PO.20.00296
373. Abousand et al., 2021; DOI: 10.26502/jcsct.5079114
374. Cardona AF, et al. J Neurooncol (2021) PMID: 34498213
375. Ramalingam SS, et al. N. Engl. J. Med. (2019) PMID: 31751012
376. Tsuboi et al., 2022; ESMO Abstract LBA47
377. Cho JH, et al. J. Clin. Oncol. (2019) PMID: 31825714
378. Kenmotsu H, et al. J Thorac Oncol (2022) PMID: 35636696
379. Soo et al., 2021; ESMO Abstract VP3-2021
380. Oxnard GR, et al. Ann. Oncol. (2020) PMID: 32139298
381. Quintás-Cardama A, et al. Nat Clin Pract Oncol (2008) PMID: 18936790
382. Bisagni G, et al. J Thorac Oncol (2009) PMID: 19461405
383. Handolias D, et al. Br. J. Cancer (2010) PMID: 20372153
384. Dişel U, et al. Lung Cancer (2011) PMID: 20970876
385. Park SH, et al. Invest New Drugs (2012) PMID: 22270258
386. Catania C, et al. Onco Targets Ther (2014) PMID: 24855380
387. Guo T, et al. Clin. Cancer Res. (2007) PMID: 17699867
388. Hu S, et al. Mol. Cancer Ther. (2008) PMID: 18483300
389. Zustovich et al., 2013; 23898124; Reardon et al.
390. Lee EQ, et al. Neuro-oncology (2012) PMID: 23099651
391. Hottinger AF, et al. Br. J. Cancer (2014) PMID: 24786603
392. Karajannis MA, et al. Neuro-oncology (2014) PMID: 24803676
393. Heinrich MC, et al. J. Clin. Oncol. (2008) PMID: 18955458
394. Buchbinder EI, et al. Cancer (2015) PMID: 26264378
395. Reichardt P, et al. BMC Cancer (2016) PMID: 26772734
396. Hirai F, et al. Mol Clin Oncol (2016) PMID: 27073655
397. Goemans BF, et al. Leuk. Res. (2010) PMID: 20435347
398. Pan E, et al. J. Neurooncol. (2012) PMID: 22832897
399. Kreisl TN, et al. J. Neurooncol. (2013) PMID: 23086433
400. Balaña C, et al. Target Oncol (2014) PMID: 24424564
401. Hutterer M, et al. Neuro-oncology (2014) PMID: 24311637

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2023 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 11 January 2023
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531