



Sample Information

Patient Name: 曾蕙滿
Gender: Female
ID No.: D200058217
History No.: 24776407
Age: 73

Ordering Doctor: DOC4205A 柯博仲
Ordering REQ.: 0CDTDCM
Signing in Date: 2022/12/21

Path No.: M111-00031
MP No.: MY22038
Assay: Oncomine Myeloid Assay
Sample Type: Bone Marrow
Bone Marrow Aspirating Date: 2022/12/15

Reporting Doctor: DOC5466K 葉奕成 (Phone: 8#5466)

Note:

Sample Cancer Type: Myelodysplastic Syndrome

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Report Highlights

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Relevant Myelodysplastic Syndrome Variants

Gene	Finding	Gene	Finding
ASXL1	None detected	NPM1	None detected
BCOR	None detected	NRAS	None detected
CBL	None detected	NUP214	None detected
CREBBP	None detected	RUNX1	RUNX1 p.(S141*) c.422C>A
ETV6	None detected	SF3B1	None detected
EZH2	None detected	SRSF2	SRSF2 p.(P95H) c.284C>A
FLT3	None detected	STAG2	STAG2 p.(D683Rfs*2) c.2046_2047insAGAT
GATA2	None detected	TP53	None detected
IDH2	None detected	U2AF1	None detected
KIT	None detected	WT1	None detected
KMT2A	None detected	ZRSR2	None detected
MECOM	None detected		

Relevant Biomarkers

Tier	Genomic Alteration	Relevant Therapies (In this cancer type)	Relevant Therapies (In other cancer type)	Clinical Trials
IA	<i>RUNX1 p.(S141*) c.422C>A</i> RUNX family transcription factor 1 Allele Frequency: 22.20% Prognostic significance: NCCN: Poor	None	allogeneic stem cells azacitidine cytarabine cytarabine + daunorubicin cytarabine + daunorubicin + etoposide cytarabine + etoposide + idarubicin cytarabine + fludarabine + idarubicin + filgrastim cytarabine + idarubicin cytarabine + mitoxantrone decitabine gemtuzumab ozogamicin + chemotherapy venetoclax + chemotherapy	0
IA	<i>SRSF2 p.(P95H) c.284C>A</i> serine and arginine rich splicing factor 2 Allele Frequency: 38.40% Prognostic significance: NCCN: Poor	None	None	0
IA	<i>STAG2 p.(D683Rfs*2) c.2046_2047insAGAT</i> stromal antigen 2 Allele Frequency: 19.90% Prognostic significance: NCCN: Poor	None	None	0

Public data sources included in relevant therapies: FDA1, NCCN, EMA2, ESMO

Public data sources included in prognostic and diagnostic significance: NCCN, ESMO

Variants (Exclude variant in Taiwan BioBank with >1% allele frequency)

DNA Sequence Variants

Gene	Amino Acid Change	Coding	Variant ID	Locus	Allele Frequency	Transcript	Variant Effect	Coverage
SRSF2	p.(P95H)	c.284C>A	COSM211504	chr17:74732959	38.40%	NM_003016.4	missense	1940
RUNX1	p.(S141*)	c.422C>A	COSM25125	chr21:36252940	22.20%	NM_001754.4	nonsense	1991
STAG2	p.(D683Rfs*2)	c.2046_2047insAGAT	.	chrX:123199742	19.90%	NM_001042749.2	frameshift Insertion	1995
TET2	p.(S1853=)	c.5559C>T	.	chr4:106197226	47.95%	NM_001127208.2	synonymous	1996
CEBPA	p.(H195_P196dup)	c.589_590insACCCG C	.	chr19:33792731	27.30%	NM_004364.4	nonframeshift Insertion	

Biomarker Descriptions

RUNX1 (RUNX family transcription factor 1)

Background: The RUNX1 gene encodes the runt-related transcription factor (RUNX) 1, part of the RUNX family of transcription factors which also includes RUNX2 and RUNX3¹. All RUNX proteins share several conserved regions with similar functionality including a highly conserved N-terminal 'runt' domain responsible for binding DNA, a C-terminal region composed of an activation domain, inhibitory domain, protein interacting motifs, and a nuclear matrix targeting signal². Each of these proteins are capable of interacting with core binding factor beta (CBFβ) to form the core binding factor (CBF) complex. Consequently, RUNX1, RUNX2, and RUNX3 are collectively known as core binding factor alpha (CBFα) since they can each function as the alpha subunit of CBF. Specifically, CBFβ binds to the 'runt' domain of RUNX1 leading to RUNX1 stabilization and increased affinity of the CBF complex for promoters involved in hematopoietic differentiation and cell cycle regulation^{3,4}. RUNX1 is frequently mutated in various hematological malignancies⁴. Germline mutations in RUNX1 result in a rare autosomal dominant condition known as familial platelet disorder, with predisposition to acute myeloid leukemia (FPD/AML)^{5,6}. Somatic mutations and chromosomal translocations in RUNX1 are often observed in myelodysplastic syndrome (MDS), acute myeloid leukemia (AML), acute lymphoblastic leukemia (ALL), and chronic myelomonocytic leukemia (CMML)⁴.

Alterations and prevalence: RUNX1 is frequently rearranged in hematological malignancies with over 50 different observed translocations⁷. The most recurrent translocation, t(12;21)(q34;q11), results in ETV6-RUNX1 fusion and is observed in 20-25% of childhood ALL^{8,9,10}. This translocation is also observed in adult ALL at a lower frequency (2%)^{9,10}. Another recurrent translocation, t(8;21)(q22;q22), results in RUNX1-RUNX1T1 fusion and is observed in 5-10% of AML¹¹. The RUNX1-RUNX1T1 fusion, consists of the RHD domain of RUNX1 and the majority of RUNX1T1, which promotes oncogenesis by altering transcriptional regulation of RUNX1 target genes^{4,11}. Somatic mutations in RUNX1 include missense, nonsense, and frameshift mutations resulting in loss of function or dominant negative effects⁴. RUNX1 mutations are reported in approximately 10% of de novo AML as well as 10-15% of MDS^{4,12,13,14}.

Potential relevance: The t(8;21)(q22;q22.1)/RUNX1-RUNX1T1 translocation is recognized as a distinct AML disease category by the World Health Organization (WHO)¹⁵. Additionally, AML with RUNX1 mutations is a provisional entity in the WHO¹⁵. Translocations involving RUNX1, specifically t(8;21)(q22;q22)/RUNX1-RUNX1T1 in AML and t(12;21)(q34;q11)/ETV6-RUNX1 in ALL, are associated with favorable risk^{12,16}. On the other hand, mutations in RUNX1 confer poor prognosis in AML, MDS, and systemic mastocytosis (SM)^{12,13,17}.

SRSF2 (serine and arginine rich splicing factor 2)

Background: The SRSF2 gene encodes the serine/arginine (SR)-rich splicing factor 2, a member of the SR-rich family of pre-mRNA splicing factors which make up part of the spliceosome. SRSF2 contains an RNA recognition motif (RRM) that recognizes and binds exonic splicing enhancers (ESE) in a sequence-specific manner¹⁸. SR proteins are essential regulators of alternative RNA splicing due to their ability to bind RNA and interact with other splicing factors. These proteins can influence the exclusion of cassette exons, a form of alternative splicing also known as exon skipping, which allows for the production of different protein isoforms^{18,19}. SRSF2 is the target of somatic missense mutations and in-frame deletions in hematological malignancies, particularly myelodysplastic syndromes (MDS), chronic myelomonocytic leukemia (CMML), and myeloproliferative neoplasms (MPN)^{20,21,22}. Such mutations in SRSF2 result in a differential gain of function which influences cassette exon exclusion, thereby supporting an oncogenic role in cancer²³.

Alterations and prevalence: Mutations in SRSF2 are observed in approximately 10% of MDS cases and 30-40% of CMML^{21,24,25}. Missense mutations at P95 are most recurrent, which leads to an amino acid change from proline to histidine (H), leucine (L), or arginine (R)²⁵. Specifically, the P95H substitution alters SRSF2 affinity for ESEs and drives preferential recognition of cassette exons containing C- versus G-rich ESEs^{22,23}. Although less prevalent, recurrent in-frame deletions (P95H_R102del) are observed in primary myelofibrosis (PMF)²⁶. This mutation results in the deletion of 8 amino acids which has been shown to exhibit greater variation of splicing events relative to the P95 missense mutation alone²⁷.

Potential relevance: In CMML, SRSF2 mutations are often enriched and can be used to support diagnosis^{13,15}. SRSF2 mutations confer poor prognosis in MDS and systemic mastocytosis (SM) and are associated with decreased overall survival (OS)^{13,17,28}. In MPN, SRSF2 mutations are considered high-risk mutations and are independently associated with inferior OS as well as leukemia-free survival^{29,30}. Additionally, SRSF2 mutations are predictive of leukemic transformation in patients with PMF²⁹.

STAG2 (stromal antigen 2)

Background: The STAG2 gene encodes the stromal antigen 2 protein, one of the core proteins in the cohesin complex, which regulates the separation of sister chromatids during cell division^{31,32}. Components of the cohesion complex include SMC1A, SMC3, and RAD21, which bind to STAG1/STAG2 paralogs^{33,34}. Inactivating mutations in STAG2 contribute to X-linked neurodevelopmental disorders, aneuploidy, and chromosomal instability in cancer^{33,35}.

Alterations and prevalence: Somatic mutations in STAG2 include nonsense, frameshift, splice site variants¹³. Somatic mutations in STAG2 are observed in various solid tumors including 14% of bladder cancer, 10% of uterine cancer, 3% of stomach cancer, and 4%

Biomarker Descriptions (continued)

of lung adenocarcinoma³⁶. In addition, mutations in STAG2 are observed in 5-10% of myelodysplastic syndrome(MDS), 3% of acute myeloid leukemia, and 2% of diffuse large B-cell lymphoma^{13,36}.

Potential relevance: Nonsense, frameshift, and splice site STAG2 mutations are associated with poor prognosis in MDS¹³. Truncating mutations in STAG2 lead to a loss of function in bladder cancer and are often identified as an early event associated with low grade and stage tumors³⁷.

Relevant Therapy Summary

☒ In this cancer type
 ☐ In other cancer type
 ☒ In this cancer type and other cancer types
 ☒ No evidence

RUNX1 p.(S141*) c.422C>A

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
Allogeneic hematopoietic stem cell transplantation	×	○	×	×	×
azacitidine	×	○	×	×	×
cytarabine	×	○	×	×	×
cytarabine + daunorubicin	×	○	×	×	×
cytarabine + daunorubicin + etoposide	×	○	×	×	×
cytarabine + etoposide + idarubicin	×	○	×	×	×
cytarabine + fludarabine + idarubicin + filgrastim	×	○	×	×	×
cytarabine + idarubicin	×	○	×	×	×
cytarabine + mitoxantrone	×	○	×	×	×
decitabine	×	○	×	×	×
gemtuzumab ozogamicin + cytarabine + daunorubicin	×	○	×	×	×
venetoclax + azacitidine	×	○	×	×	×
venetoclax + cytarabine	×	○	×	×	×
venetoclax + decitabine	×	○	×	×	×

Relevant Therapy Details

Current NCCN Information

☒ In this cancer type ☐ In other cancer type ☒ In this cancer type and other cancer types

NCCN information is current as of 2022-10-03. For the most up-to-date information, search www.nccn.org.
For NCCN International Adaptations & Translations, search www.nccn.org/global/international_adaptations.aspx.

RUNX1 p.(S141*) c.422C>A

☐ cytarabine + daunorubicin

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 1

Population segment (Line of therapy):

- (Induction therapy); Other recommended intervention

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

☐ cytarabine + daunorubicin + etoposide

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 1

Population segment (Line of therapy):

- (Induction therapy); Other recommended intervention

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

☐ cytarabine + etoposide + idarubicin

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 1

Population segment (Line of therapy):

- (Induction therapy); Other recommended intervention

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

☐ cytarabine + idarubicin

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 1

Population segment (Line of therapy):

- (Induction therapy); Other recommended intervention

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

RUNX1 p.(S141*) c.422C>A (continued)**○ Allogeneic hematopoietic stem cell transplantation**

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2A

Population segment (Line of therapy):

- (Consolidation therapy)

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

○ cytarabine

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2A

Population segment (Line of therapy):

- (Consolidation therapy)

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

○ cytarabine + daunorubicin

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2A

Population segment (Line of therapy):

- (Consolidation therapy)

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

○ cytarabine + daunorubicin

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2A

Population segment (Line of therapy):

- (Induction therapy); Preferred intervention

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

○ cytarabine + idarubicin

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2A

Population segment (Line of therapy):

- (Induction therapy); Preferred intervention

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

RUNX1 p.(S141*) c.422C>A (continued)**○ cytarabine + mitoxantrone**

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2A

Population segment (Line of therapy):

- (Induction therapy); Preferred intervention

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

○ decitabine

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2A

Population segment (Line of therapy):

- (Induction therapy)

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

○ gemtuzumab ozogamicin + cytarabine + daunorubicin

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2A

Population segment (Line of therapy):

- (Induction therapy); Preferred intervention

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

○ venetoclax + azacitidine

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2A

Population segment (Line of therapy):

- (Induction therapy)

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

○ venetoclax + cytarabine

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2A

Population segment (Line of therapy):

- (Induction therapy)

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

RUNX1 p.(S141*) c.422C>A (continued)**○ venetoclax + decitabine**

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2A

Population segment (Line of therapy):

- (Induction therapy)

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

○ azacitidine

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2B

Population segment (Line of therapy):

- (Induction therapy)

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

○ cytarabine + daunorubicin + etoposide

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2B

Population segment (Line of therapy):

- (Induction therapy); Other recommended intervention

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

○ cytarabine + etoposide + idarubicin

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2B

Population segment (Line of therapy):

- (Induction therapy); Other recommended intervention

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

○ cytarabine + fludarabine + idarubicin + filgrastim

Cancer type: Acute Myeloid Leukemia

Variant class: RUNX1 mutation

NCCN Recommendation category: 2B

Population segment (Line of therapy):

- (Induction therapy); Other recommended intervention

Reference: NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]

Prognostic Details

Current NCCN Information

NCCN information is current as of 2022-10-03. For the most up-to-date information, search www.nccn.org.
For NCCN International Adaptations & Translations, search www.nccn.org/global/international_adaptations.aspx.

RUNX1 p.(S141*) c.422C>A

Prognostic significance: NCCN: Poor

Cancer type: Myelodysplastic Syndrome

Variant class: RUNX1 truncating mutation

NCCN Recommendation category: 2A

Summary:

- NCCN Guidelines® independently associate the biomarker with poor prognosis

Reference: NCCN Guidelines® - NCCN-Myelodysplastic Syndromes [Version 1.2023]

SRSF2 p.(P95H) c.284C>A

Prognostic significance: NCCN: Poor

Cancer type: Myelodysplastic Syndrome

Variant class: SRSF2 P95 mutation

NCCN Recommendation category: 2A

Summary:

- NCCN Guidelines® associate the biomarker with poor prognosis

Reference: NCCN Guidelines® - NCCN-Myelodysplastic Syndromes [Version 1.2023]

STAG2 p.(D683Rfs*2) c.2046_2047insAGAT

Prognostic significance: NCCN: Poor

Cancer type: Myelodysplastic Syndrome

Variant class: STAG2 truncating mutation

NCCN Recommendation category: 2A

Summary:

- NCCN Guidelines® associate the biomarker with poor prognosis

Reference: NCCN Guidelines® - NCCN-Myelodysplastic Syndromes [Version 1.2023]

Signatures

Testing Personnel:

Laboratory Supervisor:

Pathologist:

References

1. de et al. Runx transcription factors in the development and function of the definitive hematopoietic system. *Blood*. 2017 Apr 13;129(15):2061-2069. PMID: 28179276
2. Chuang et al. RUNX family: Regulation and diversification of roles through interacting proteins. *Int. J. Cancer*. 2013 Mar 15;132(6):1260-71. PMID: 23180629
3. Jung et al. Prognostic factor analysis in core-binding factor-positive acute myeloid leukemia. *Anticancer Res*. 2014 Feb;34(2):1037-45. PMID: 24511052
4. Sood et al. Role of RUNX1 in hematological malignancies. *Blood*. 2017 Apr 13;129(15):2070-2082. PMID: 28179279
5. Béri-Dexheimer et al. Clinical phenotype of germline RUNX1 haploinsufficiency: from point mutations to large genomic deletions. *Eur. J. Hum. Genet*. 2008 Aug;16(8):1014-8. PMID: 18478040
6. Hayashi et al. Myeloid neoplasms with germ line RUNX1 mutation. *Int. J. Hematol*. 2017 Aug;106(2):183-188. PMID: 28534116
7. De et al. RUNX1 translocations and fusion genes in malignant hemopathies. *Future Oncol*. 2011 Jan;7(1):77-91. PMID: 21174539
8. De et al. ETV6 fusion genes in hematological malignancies: a review. *Leuk. Res*. 2012 Aug;36(8):945-61. PMID: 22578774
9. Pui et al. Acute lymphoblastic leukemia. *N. Engl. J. Med*. 2004 Apr 8;350(15):1535-48. PMID: 15071128
10. NCCN Guidelines® - Acute Lymphoblastic Leukemia [Version 2.2019]. 2019 May 15
11. Huret et al. Atlas of genetics and cytogenetics in oncology and haematology in 2013. *Nucleic Acids Res*. 2013 Jan;41(Database issue):D920-4. PMID: 23161685
12. NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2022]
13. NCCN Guidelines® - NCCN-Myelodysplastic Syndromes [Version 1.2023]
14. Weinstein et al. The Cancer Genome Atlas Pan-Cancer analysis project. *Nat. Genet*. 2013 Oct;45(10):1113-20. PMID: 24071849
15. Arber et al. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. *Blood*. 2016 May 19;127(20):2391-405. PMID: 27069254
16. NCCN Guidelines® - NCCN-Acute Lymphoblastic Leukemia [Version 1.2022]
17. NCCN Guidelines® - NCCN-Systemic Mastocytosis [Version 1.2020]
18. Liang et al. SRSF2 mutations drive oncogenesis by activating a global program of aberrant alternative splicing in hematopoietic cells. *Leukemia*. 2018 Dec;32(12):2659-2671. PMID: 29858584
19. Cui et al. Comparative Analysis and Classification of Cassette Exons and Constitutive Exons. *Biomed Res Int*. 2017;2017:7323508. doi: 10.1155/2017/7323508. Epub 2017 Dec 4. PMID: 29349080
20. Meggendorfer et al. The mutational landscape of 18 investigated genes clearly separates four subtypes of myelodysplastic/myeloproliferative neoplasms. *Haematologica*. 2018 May;103(5):e192-e195. PMID: 29700173
21. Arbab et al. Prognostic significance of SRSF2 mutations in myelodysplastic syndromes and chronic myelomonocytic leukemia: a meta-analysis. *Hematology*. 2018 Dec;23(10):778-784. PMID: 29757120
22. Kim et al. SRSF2 Mutations Contribute to Myelodysplasia by Mutant-Specific Effects on Exon Recognition. *Cancer Cell*. 2015 May 11;27(5):617-30. PMID: 25965569
23. Zhang et al. Disease-associated mutation in SRSF2 misregulates splicing by altering RNA-binding affinities. *Proc. Natl. Acad. Sci. U.S.A.* 2015 Aug 25;112(34):E4726-34. PMID: 26261309
24. Ethan et al. AACR Project GENIE: Powering Precision Medicine through an International Consortium. *Cancer Discov*. 2017 Aug;7(8):818-831. PMID: 28572459
25. Thol et al. Frequency and prognostic impact of mutations in SRSF2, U2AF1, and ZRSR2 in patients with myelodysplastic syndromes. *Blood*. 2012 Apr 12;119(15):3578-84. PMID: 22389253
26. Lasho et al. SRSF2 mutations in primary myelofibrosis: significant clustering with IDH mutations and independent association with inferior overall and leukemia-free survival. *Blood*. 2012 Nov 15;120(20):4168-71. PMID: 22968464
27. Komeno et al. SRSF2 Is Essential for Hematopoiesis, and Its Myelodysplastic Syndrome-Related Mutations Dysregulate Alternative Pre-mRNA Splicing. *Mol. Cell. Biol*. 2015 Sep 1;35(17):3071-82. PMID: 26124281
28. Jawhar et al. Splenomegaly, elevated alkaline phosphatase and mutations in the SRSF2/ASXL1/RUNX1 gene panel are strong adverse prognostic markers in patients with systemic mastocytosis. *Leukemia*. 2016 Dec;30(12):2342-2350. PMID: 27416984
29. NCCN Guidelines® - NCCN-Myeloproliferative Neoplasms [Version 3.2022]
30. Vannucchi et al. Mutations and prognosis in primary myelofibrosis. *Leukemia*. 2013 Sep;27(9):1861-9. PMID: 23619563
31. Mehta et al. Cohesin: functions beyond sister chromatid cohesion. *FEBS Lett*. 2013 Aug 2;587(15):2299-312. PMID: 23831059
32. Aquila et al. The role of STAG2 in bladder cancer. *Pharmacol. Res*. 2018 May;131:143-149. PMID: 29501732

References (continued)

33. Mullegama et al. De novo loss-of-function variants in STAG2 are associated with developmental delay, microcephaly, and congenital anomalies. *Am. J. Med. Genet. A.* 2017 May;173(5):1319-1327. PMID: 28296084
34. van et al. Synthetic lethality between the cohesin subunits STAG1 and STAG2 in diverse cancer contexts. *Elife.* 2017 Jul 10;6. PMID: 28691904
35. Solomon et al. Mutational inactivation of STAG2 causes aneuploidy in human cancer. *Science.* 2011 Aug 19;333(6045):1039-43. PMID: 21852505
36. Cerami et al. The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. *Cancer Discov.* 2012 May;2(5):401-4. PMID: 22588877
37. Solomon et al. Frequent truncating mutations of STAG2 in bladder cancer. *Nat. Genet.* 2013 Dec;45(12):1428-30. PMID: 24121789