

ABOUT THE TEST FoundationOne®Liquid CDx is a next generation sequencing (NGS) assay that identifies clinically relevant genomic alterations in circulating cell-free DNA.

PATIENT	DISEASE Pancreas ductal adenocarcinoma	PHYSICIAN	ORDERING PHYSICIAN Yeh, Yi-Chen	SPECIMEN	SPECIMEN ID TYL 48782587
	NAME Li, Tsui-Yu		MEDICAL FACILITY Taipei Veterans General Hospital		SPECIMEN TYPE Blood
	DATE OF BIRTH 28 January 1949		ADDITIONAL RECIPIENT None		DATE OF COLLECTION 16 August 2022
	SEX Female		MEDICAL FACILITY ID 205872		SPECIMEN RECEIVED 18 August 2022
	MEDICAL RECORD # 48782587		PATHOLOGIST Not Provided		

Biomarker Findings

Blood Tumor Mutational Burden - 3 Muts/Mb
Microsatellite status - MSI-High Not Detected
Tumor Fraction - Elevated Tumor Fraction Not Detected

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

KRAS G12D
CBL C381Y
DNMT3A C494*, splice site 1667+1G>A
TP53 R175H

Report Highlights

- Evidence-matched clinical trial options based on this patient's genomic findings: (p. 9)
- Variants that may represent clonal hematopoiesis and may originate from non-tumor sources: **CBL** C381Y (p. 7), **DNMT3A** C494*, splice site 1667+1G>A (p. 7)

BIOMARKER FINDINGS

Blood Tumor Mutational Burden
 - 3 Muts/Mb

Microsatellite status
 - MSI-High Not Detected

Tumor Fraction
 - Elevated Tumor Fraction Not Detected

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. See Biomarker Findings section

MSI-High not detected. No evidence of microsatellite instability in this sample (see Appendix section).

Tumor fraction is considered elevated when ctDNA levels are high enough that aneuploidy can be detected. The fact that elevated tumor fraction was not detected in this specimen indicates the possibility of lower levels of ctDNA but does not compromise confidence in any reported alterations. However, in the setting of a negative liquid biopsy result, orthogonal testing of a tissue specimen should be considered if clinically indicated (see Biomarker Findings section).

GENOMIC FINDINGS

VAF %

KRAS - G12D 0.12%

3 Trials see p. 9

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)

None

THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)

None

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS (CH)

Genomic findings below may include nontumor somatic alterations, such as CH. The efficacy of targeting such nontumor somatic alterations is unknown. This content should be interpreted based on clinical context. Refer to appendix for additional information on CH.

CBL - C381Y p. 7 **DNMT3A** - C494*, splice site 1667+1G>A p. 7

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. · 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

CBL - C381Y **p. [7](#)** **TP53 - R175H** **p. [8](#)**
DNMT3A - C494*, splice site 1667+1G>A **p. [7](#)**

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the therapies listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and/or exhaustive. Neither the therapies nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies. Therapies contained in this report may have been approved by the US FDA or other national authorities; however, they might not have been approved in your respective country. In the appropriate clinical context, germline testing of *APC, ATM, BAP1, BRCA1, BRCA2, BRIP1, CHEK2, FH, FLCN, MEN1, MLH1, MSH2, MSH6, MUTYH, NF1, NF2, PALB2, PMS2, POLE, PTEN, RAD51C, RAD51D, RB1, RET, SDHA, SDHB, SDHC, SDHD, SMAD4, STK11, TGFBR2, TP53, TSC1, TSC2, VHL*, and *WT1* is recommended.

Variant Allele Frequency is not applicable for copy number alterations.

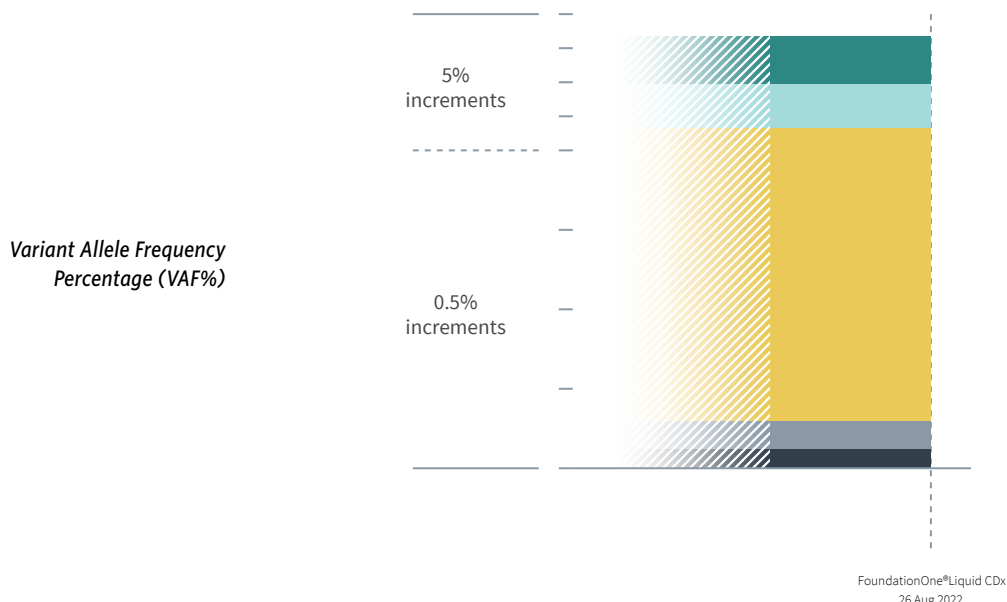
Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. • 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1437612-01



HISTORIC PATIENT FINDINGS

ORD-1437612-01
VAF%

Blood Tumor Mutational Burden

3 Muts/Mb

Microsatellite status

MSI-High Not Detected

Tumor Fraction

Elevated Tumor Fraction Not Detected

KRAS	● G12D	0.12%
CBL	● C381Y	7.0%
DNMT3A	● C494*	5.0%
	● splice site 1667+1G>A	6.4%
TP53	● R175H	0.18%

NOTE This comparison table refers only to genes and biomarkers assayed by prior FoundationOne®Liquid CDx, FoundationOne®Liquid, FoundationOne®, or FoundationOne®CDx tests. Up to five previous tests may be shown.

For some genes in FoundationOne Liquid CDx, only select exons are assayed. Therefore, an alteration found by a previous test may not have been confirmed despite overlapping gene lists. Please refer to the Appendix for the complete list of genes and exons assayed. The gene and biomarker list will be updated periodically to reflect new knowledge about cancer biology.

As new scientific information becomes available, alterations that had previously been listed as Variants of Unknown Significance (VUS) may become reportable.

Tissue Tumor Mutational Burden (TMB) and blood TMB (bTMB) are estimated from the number of synonymous and non-synonymous single-nucleotide variants (SNVs) and insertions and deletions (indels) per area of coding genome sampled, after the removal of known and likely oncogenic driver events and germline SNPs. Tissue TMB is calculated based on variants with an allele frequency of $\geq 5\%$, and bTMB is calculated based on variants with an allele frequency of $\geq 0.5\%$.

Not Tested = not baited, not reported on test, or test preceded addition of biomarker or gene

Not Detected = baited but not detected on test

Detected = present (VAF% is not applicable)

VAF% = variant allele frequency percentage

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. · 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1437612-01

Cannot Be Determined = Sample is not of sufficient data quality to confidently determine biomarker status

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. · 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1437612-01

BIOMARKER FINDINGS

BIOMARKER

Blood Tumor Mutational Burden

RESULT

3 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased blood tumor mutational burden (bTMB) may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1¹⁻³, anti-PD-1³⁻⁴, and anti-PD-1/CTLA4 therapies⁵⁻⁶. A Phase 2 multi-solid-tumor trial showed that bTMB ≥ 16 Muts/Mb (as measured by this assay) was associated with improved survival from treatment with a PD-1 inhibitor alone or in combination with a CTLA-4 inhibitor⁵. In non-small cell lung cancer (NSCLC), multiple clinical trials have shown patients with higher bTMB derive clinical benefit from immune checkpoint

inhibitors following single-agent or combination treatments with either CTLA4 inhibitors or chemotherapy, with reported high bTMB cutpoints ranging from 6 Muts/Mb–16 Muts/Mb¹. In head and neck squamous cell carcinoma (HNSCC), a Phase 3 trial showed that bTMB ≥ 16 Muts/Mb (approximate equivalency ≥ 8 Muts/Mb as measured by this assay) was associated with improved survival from treatment with a PD-L1 inhibitor alone or in combination with a CTLA-4 inhibitor⁷.

FREQUENCY & PROGNOSIS

Average bTMB levels in solid tumors other than NSCLC have not been evaluated (cBioPortal, COSMIC, PubMed, Mar 2022)⁸⁻¹⁰. Published data investigating the prognostic implications of bTMB levels in pancreatic carcinoma are limited (PubMed, Jul 2022). A study of patients with pancreatic ductal adenocarcinoma harboring mismatch repair gene mutations reported improved prognosis for patients with high TMB measured in tissue samples (defined as >50 mutations; survival 69–314 months) compared to those with lower TMB (average of 5.7 mutations;

10–42 months)¹¹.

FINDING SUMMARY

Blood tumor mutational burden (bTMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations from circulating tumor DNA in blood. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma¹²⁻¹³ and cigarette smoke in lung cancer¹⁴⁻¹⁵, treatment with temozolomide-based chemotherapy in glioma¹⁶⁻¹⁷, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes¹⁸⁻²², and microsatellite instability (MSI)^{18,21-22}. High bTMB levels were not detected in this sample. It is unclear whether the bTMB levels in this sample would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents^{1-2,4}. Depending on the clinical context, TMB testing of an alternate sample or by another methodology could be considered.

BIOMARKER

Tumor Fraction

RESULT

Elevated Tumor Fraction Not Detected

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Specimens with elevated tumor fraction values have high circulating-tumor DNA (ctDNA) content, and thus high sensitivity for identifying genomic alterations. Such specimens are at low risk of false negative results. However, if elevated tumor fraction is not detected, it does not exclude the presence of disease burden or compromise the confidence of reported alterations. Tumor fraction levels currently have limited implications for diagnosis, surveillance, or therapy and should not

be overinterpreted or compared from one blood draw to another. There are currently no targeted approaches to address specific tumor fraction levels. In the research setting, changes in tumor fraction estimates have been associated with treatment duration and clinical response and may be a useful indicator for future cancer management²³⁻²⁸.

FREQUENCY & PROGNOSIS

Detectable ctDNA levels have been reported in a variety of tumor types, with higher tumor fraction levels reported for patients with metastatic (Stage 4) tumors compared with patients with localized disease (Stages 1 to 3)²⁹. Elevated tumor fraction levels have been reported to be associated with worse prognosis in a variety of cancer types, including pancreatic cancer³⁰, Ewing sarcoma and osteosarcoma³¹, prostate cancer²⁶, breast cancer³², leiomyosarcoma³³, esophageal cancer³⁴, colorectal

cancer³⁵, and gastrointestinal cancer³⁶.

FINDING SUMMARY

Tumor fraction provides an estimate of the percentage of ctDNA present in a cell-free DNA (cfDNA) sample. The tumor fraction estimate for this sample is based on the observed level of aneuploid instability. The tumor fraction algorithm utilized for FoundationOne Liquid CDx uses the allele frequencies of approximately 1,000 single-nucleotide polymorphism (SNP) sites across the genome. Unlike the maximum somatic allele frequency (MSAF) method of estimating ctDNA content³⁷, the tumor fraction metric does not take into account the allele frequency of individual variants but rather produces a more holistic estimate of ctDNA content using data from across the genome. The amount of ctDNA detected may correlate with disease burden and response to therapy³⁸⁻³⁹.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. · 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1437612-01

GENOMIC FINDINGS

GENE

KRAS

ALTERATION

G12D

TRANSCRIPT ID

NM_004985

CODING SEQUENCE EFFECT

35G>A

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Preclinical evidence suggests that KRAS activation may predict sensitivity to MEK inhibitors, such as trametinib, binimetinib, cobimetinib, and selumetinib⁴⁰⁻⁴⁵. For patients with pancreatic cancer, MEK inhibitor combinations are under investigation. A Phase 2 study of trametinib with pembrolizumab versus gemcitabine after stereotactic body radiotherapy (SBRT) reported increased median OS (mOS, 14.9 months vs. 12.8 months, HR=0.69) benefit for patients with KRAS-mutated, PD-L1 positive disease⁴⁶. Combination MEK/autophagy inhibitors are also under investigation based on preclinical evidence of increased autophagy downstream of KRAS-mutated pancreatic tumors⁴⁷⁻⁴⁸. A heavily pretreated patient with pancreatic cancer treated with trametinib plus hydroxychloroquine experienced a PR⁴⁷. A Phase 2 study of the reoviral agent pelareorep with gemcitabine for patients with pancreatic cancer reported 1 PR, 23

SDs, and 5 PDs for 34 patients with a favorable median OS of 10.2 months⁴⁹. A Phase 1b study of second-line pelareorep with pembrolizumab and chemotherapy reported 1 PR of 17.4 months and a DCR of 30% (3/10)⁵⁰; an earlier study reported no benefit from pelareorep in combination with paclitaxel/carboplatin⁵¹. Trials combining MEK inhibitors with other targeted therapies, such as EGFR inhibitors⁵² or PI3K-AKT pathway inhibitors⁵³⁻⁵⁴, reported no PRs and frequent adverse events for patients with KRAS-mutated pancreatic cancer. Clinical trials combining various MEK inhibitors with gemcitabine reported no additional benefit compared to gemcitabine alone irrespective of KRAS mutation status⁵⁵⁻⁵⁸, despite promising results in earlier trials of MEK inhibitor monotherapies⁵⁹⁻⁶⁴. In a Phase 1 study evaluating the MEK-pan-RAF dual inhibitor CH5126766, 6 patients harboring KRAS mutations experienced PRs, including 3 with non-small cell lung cancer (NSCLC), 1 with low-grade serous ovarian carcinoma (LGSOC), 1 with endometrial adenocarcinoma, and 1 with multiple myeloma⁶⁵. Combination of CH5126766 with the FAK inhibitor defactinib elicited PR rates of 50% (4/8) for patients with KRAS-mutated low-grade serous ovarian cancer and 12% (2/17) for patients with KRAS-mutated non-small cell lung cancer (NSCLC) in a Phase 1 study⁶⁶⁻⁶⁷. Preclinical and clinical data suggest that KRAS mutations may predict clinical benefit from SHP2 inhibitors⁶⁸⁻⁶⁹. A Phase 1 study of RMC-4630 for relapsed/refractory solid tumors reported a DCR of 58% (23/40) for patients with NSCLC and KRAS

mutations and a DCR of 75% (12/16) for patients with NSCLC and KRAS G12C mutations⁷⁰. Interim results from a Phase 1/2 study of RMC-4630 plus cobimetinib reported tumor reduction in 3 of 8 patients with KRAS-mutated colorectal cancer⁷¹. Preclinical data suggest that KRAS mutation may confer sensitivity to SOS1 inhibitors⁷²⁻⁷³. Phase 1 studies of the SOS1 inhibitor BI 1701963 alone or in combination with MEK inhibitors, KRAS G12C inhibitors, or irinotecan are recruiting for patients with solid tumors harboring KRAS mutations⁷⁴⁻⁷⁵.

FREQUENCY & PROGNOSIS

KRAS mutations have been observed in 91-95% of pancreatic ductal adenocarcinoma cases⁷⁶⁻⁷⁷, with the majority of mutations found at codon 12⁷⁸⁻⁸¹. KRAS mutations, particularly G12D, have been associated with decreased median survival time in patients with pancreatic ductal adenocarcinoma⁷⁹.

FINDING SUMMARY

KRAS encodes a member of the RAS family of small GTPases. Activating mutations in RAS genes can cause uncontrolled cell proliferation and tumor formation^{41,82}. KRAS alterations affecting amino acids G12, G13, Q22, P34, A59, Q61, and A146, as well as mutations G10_A11insG, G10_A11insAG (also reported as G10_A11dup and G12_G13insAG), A18D, L19F, D33E, G60_A66dup/E62_A66dup, E62K, E63K, R68S, and K117N have been characterized as activating and oncogenic^{41,83-105}.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. · 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1437612-01

GENOMIC FINDINGS

GENE

CBL

ALTERATION

C381Y

TRANSCRIPT ID

NM_005188

CODING SEQUENCE EFFECT

1142G>A

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

CBL inactivation may lead to the hyperactivation of various receptor tyrosine kinases (RTKs), including MET¹⁰⁶, PDGFRA¹⁰⁷, KIT¹⁰⁸, VEGFR2¹⁰⁹, and the TAM (TYRO3, AXL, MER) RTKs¹¹⁰. These RTKs are targets of the multikinase inhibitor sitravatinib¹¹¹, which has shown activity in CBL-mutated advanced solid tumors¹¹². Among 8 patients with CBL inactivating alterations in a Phase 1b trial, sitravatinib produced 2 PRs (25% ORR), with 1 NSCLC and 1 melanoma responding for over 4 months, and 4 SD outcomes, with 3 prolonged SDs seen in a patient with NSCLC, a patient with esophageal cancer, and a patient with a pancreatic neuroendocrine tumor¹¹². CBL has

been shown to downregulate EGFR¹¹³⁻¹¹⁷ and FLT3¹¹⁸⁻¹²⁰. Preclinical models of myeloid malignancies have demonstrated that CBL inactivation confers sensitivity to the FLT3-targeting therapies sunitinib¹¹⁸, midostaurin¹²⁰, and quizartinib¹²¹, as well as to dasatinib¹²², although clinical evidence for this approach in solid tumors is lacking.

FREQUENCY & PROGNOSIS

CBL mutation has been observed in 0-0.3% of cases in several genomic studies of pancreatic adenocarcinoma^{76-77,123}. Reduced CBL expression has been associated with reduced disease-specific survival in one study of pancreatic cancer¹²⁴. CBL protein levels have been reported to be inversely correlated with activated EGFR in pancreatic cancer¹²⁵. One preclinical study reported that pancreatic cancer cells with low CBL expression were chemoresistant and that this resistance could be alleviated by combination of the EGFR inhibitor erlotinib with chemotherapy¹²⁵.

FINDING SUMMARY

CBL encodes an E3 ubiquitin protein ligase that is involved in cell signaling and ubiquitination, targeting proteins such as EGFR, FGFR1, FGFR2, PDGFR-alpha, PDGFR-beta, FLT3, and SRC for

degradation by the proteasome¹²⁶⁻¹³⁰. CBL alterations that result in loss or disruption of the tyrosine kinase binding domain, RING finger domain, and/or tail domain, as observed here, are predicted to be inactivating and to promote tumorigenesis¹³¹⁻¹⁴⁸.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion¹⁴⁹⁻¹⁵⁴. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy¹⁴⁹⁻¹⁵⁰. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease¹⁵⁵. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH^{153,156-157}. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

GENE

DNMT3A

ALTERATION

C494*, splice site 1667+1G>A

TRANSCRIPT ID

NM_022552, NM_022552

CODING SEQUENCE EFFECT

1482C>A, 1667+1G>A

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no targeted therapies available to address genomic alterations in DNMT3A in solid tumors.

FREQUENCY & PROGNOSIS

DNMT3A alterations have been reported at

relatively low frequencies in solid tumors and are more prevalent in hematological malignancies (cBioPortal, Feb 2022)⁸⁻⁹. Published data investigating the prognostic implications of DNMT3A alterations in solid tumors are limited (PubMed, Feb 2022).

FINDING SUMMARY

The DNMT3A gene encodes the protein DNA methyltransferase 3A, an enzyme that is involved in the methylation of newly synthesized DNA, a function critical for gene regulation¹⁵⁸⁻¹⁵⁹. The role of DNMT3A in cancer is uncertain, as some reports describe increased expression and contribution to tumor growth, whereas others propose a role for DNMT3A as a tumor suppressor¹⁶⁰⁻¹⁶⁵. Alterations such as seen here may disrupt DNMT3A function or expression¹⁶⁶⁻¹⁶⁹.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion¹⁴⁹⁻¹⁵⁴. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy¹⁴⁹⁻¹⁵⁰. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease¹⁵⁵. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH^{153,156-157}. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

ORDERED TEST # ORD-1437612-01

GENOMIC FINDINGS

GENE

TP53

ALTERATION

R175H

TRANSCRIPT ID

NM_000546

CODING SEQUENCE EFFECT

524G>A

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor adavosertib¹⁷⁰⁻¹⁷³, or p53 gene therapy and immunotherapeutics such as SGT-53¹⁷⁴⁻¹⁷⁸ and ALT-801¹⁷⁹. In a Phase 1 study, adavosertib in combination with gemcitabine, cisplatin, or carboplatin elicited PRs in 9.7% and SDs in 53% of patients with solid tumors; the response rate was 21% (4/19) for patients with TP53 mutations versus 12% (4/33) for patients who were TP53 wildtype¹⁸⁰. A Phase 2 trial of adavosertib in combination with chemotherapy (gemcitabine, carboplatin, paclitaxel, or doxorubicin) reported a 32% (30/94, 3 CR) ORR and a 73% (69/94) DCR for patients with platinum-refractory TP53-mutated ovarian, Fallopian tube, or peritoneal cancer¹⁸¹. A smaller Phase 2 trial of adavosertib in combination with carboplatin achieved a 43% (9/21, 1 CR) ORR and a 76% (16/21) DCR for patients with platinum-refractory TP53-mutated ovarian cancer¹⁸². The combination of adavosertib with paclitaxel and carboplatin for patients with TP53-mutated ovarian cancer also significantly increased PFS compared with paclitaxel and carboplatin alone¹⁸³. In the Phase 2 VIKTORY trial, patients with TP53-mutated metastatic and/or recurrent gastric cancer experienced a 24% (6/25) ORR with adavosertib combined with paclitaxel¹⁸⁴. A Phase 1 trial of neoadjuvant adavosertib in combination with cisplatin and docetaxel for head and neck squamous cell carcinoma (HNSCC) elicited a 71% (5/7) response rate for patients with TP53 alterations¹⁸⁵. The Phase 2 FOCUS4-C trial for

patients with TP53- and RAS-mutated colorectal cancer reported improvement in PFS (3.61 vs. 1.87 months, HR=0.35, p=0.0022), but not OS (14.0 vs 12.8 months, p=0.93), following adavosertib treatment compared with active monitoring¹⁸⁶. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel for patients with solid tumors, 75% (9/12) of evaluable patients experienced clinical benefit, including 2 confirmed and 1 unconfirmed PRs and 2 instances of SD with significant tumor shrinkage¹⁷⁸. Missense mutations leading to TP53 inactivation may also be sensitive to therapies that reactivate mutated p53 such as APR-246¹⁸⁷⁻¹⁸⁹. In a Phase 1b trial for patients with p53-positive high-grade serous ovarian cancer, APR-246 combined with carboplatin and pegylated liposomal doxorubicin achieved a 52% (11/21) response rate and 100% DCR¹⁹⁰. ATR inhibitor treatment of chronic lymphocytic leukemia (CLL) cells with biallelic inactivation of TP53 suppressed cell viability, promoted DNA damage, and attenuated xenograft growth in preclinical studies¹⁹¹⁻¹⁹²; however, ATR inhibitors as monotherapy had little effect on these parameters in solid tumor models in other preclinical studies¹⁹³⁻¹⁹⁴. Therefore, it is unclear whether TP53 inactivation predicts sensitivity to ATR inhibition.

FREQUENCY & PROGNOSIS

TP53 mutations have been reported in 33-75% of pancreatic carcinomas, with the majority occurring as missense mutations, while deletion of TP53 has been found in 66% of pancreatic ductal adenocarcinoma cases^{76,195-197}. TP53 mutations are common in pancreatic ductal adenocarcinomas and are known to occur in the process of pancreatic carcinogenesis¹⁹⁸⁻¹⁹⁹. Additionally, aberrant expression of p53 has been found in 54-81% of pancreatic ductal adenocarcinoma cases^{196,200-202}. Studies have found inconsistent results regarding the prognostic significance of p53 expression in pancreatic ductal adenocarcinoma, although one study correlated low levels of TP53 mRNA with poor patient prognosis^{200,203-204}.

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in

aggressive advanced cancers²⁰⁵. Alterations such as seen here may disrupt TP53 function or expression²⁰⁶⁻²¹⁰.

POTENTIAL GERMLINE IMPLICATIONS

One or more of the TP53 variants observed here has been described in the ClinVar database as a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with Li-Fraumeni syndrome (ClinVar, Mar 2022)²¹¹. Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. Germline mutations in TP53 are associated with the very rare autosomal dominant disorder Li-Fraumeni syndrome and the early onset of many cancers²¹²⁻²¹⁴, including sarcomas²¹⁵⁻²¹⁶. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000²¹⁷ to 1:20,000²¹⁶. For pathogenic TP53 mutations identified during tumor sequencing, the rate of germline mutations was 1% in the overall population and 6% in tumors arising before age 30²¹⁸. In the appropriate clinical context, germline testing of TP53 is recommended.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion¹⁴⁹⁻¹⁵⁴. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy¹⁴⁹⁻¹⁵⁰. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease¹⁵⁵. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH^{153,156-157}. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

ORDERED TEST # ORD-1437612-01

CLINICAL TRIALS

IMPORTANT Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and should be investigated by the physician or

research staff. This is not a comprehensive list of all available clinical trials. There may also be compassionate use or early access programs available, which are not listed in this report. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials are not ranked in order of potential or predicted efficacy for this patient or

in order of level of evidence for this patient's tumor type. Clinical trials listed here may have additional enrollment criteria that may require medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov. However, clinicaltrials.gov does not list all clinical trials that might be available.

GENE
KRAS
ALTERATION
G12D
RATIONALE

Multiple clinical studies have reported lack of efficacy of MEK inhibitors as monotherapy for treatment of KRAS-mutant pancreatic cancer. Emerging data suggest patients with KRAS-mutant pancreatic cancer may be sensitive to

combination MEK/autophagy inhibitors. Limited clinical and preclinical studies indicate KRAS mutations may predict sensitivity to MEK-pan-RAF dual inhibitors.

NCT04892017
PHASE 1/2

A Safety, Tolerability and PK Study of DCC-3116 in Patients With RAS or RAF Mutant Advanced or Metastatic Solid Tumors.

TARGETS
 ULK1, ULK2, MEK

LOCATIONS: Massachusetts, Texas, Pennsylvania

NCT04132505
PHASE 1

Binimetinib and Hydroxychloroquine in Treating Patients With KRAS Mutant Metastatic Pancreatic Cancer

TARGETS
 MEK

LOCATIONS: Texas

NCT03825289
PHASE 1

Trametinib and Hydroxychloroquine in Treating Patients With Pancreatic Cancer

TARGETS
 MEK

LOCATIONS: Utah

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. · 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1437612-01

APPENDIX
Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

ALK
V163L

ATR
S476C

CDKN2A/B
rearrangement

RNF43
C119S

TET2
Q232R

VEGFA
T8fs*78

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. · 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1437612-01

APPENDIX
Genes assayed in FoundationOne®Liquid CDx

FoundationOne Liquid CDx interrogates 324 genes, including 309 genes with complete exonic (coding) coverage and 15 genes with only select non-coding coverage (indicated with an *); 75 genes (indicated in bold) are captured with increased sensitivity and have complete exonic (coding) coverage unless otherwise noted.

ABL1 Exons 4-9	ACVR1B	AKT1 Exon 3	AKT2	AKT3	ALK Exons 20-29, Introns 18, 19	ALOX12B	AMER1 (FAM123B or WTX)	APC
AR	ARAF Exons 4, 5, 7, 11, 13, 15, 16	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BCR* Introns 8, 13, 14	BRAF Exons 11-18, Introns 7-10	BRCA1 Introns 2, 7, 8, 12, 16, 19, 20	BRCA2 Intron 2	BRD4	BRIP1	BTG1
BTG2	BTK Exons 2, 15	CALR	CARD11	CASP8	CBFB	CBL	CCND1	CCND2
CCND3	CCNE1	CD22	CD70	CD74* Introns 6-8	CD79A	CD79B	CD274 (PD-L1)	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1 Exon 3	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2 Exons 5, 17, 18	DIS3	DNMT3A	DOT1L	EED	EGFR Introns 7, 15, 24-27	EMSY (C11orf30)	EP300	EPHA3
EPHB1	EPHB4	ERBB2	ERBB3 Exons 3, 6, 7, 8, 10, 12, 20, 21, 23, 24, 25	ERBB4	ERCC4	ERG	ERRF1	ESR1 Exons 4-8
ETV4* Intron 8	ETV5* Introns 6, 7	ETV6* Introns 5, 6	EWSR1* Introns 7-13	EZH2 Exons 4, 16, 17, 18	EZR* Introns 9-11	FANCA	FANCC	FANCG
FANCL	FAS	FBXW7	FGF10	FGF12	FGF14	FGF19	FGF23	FGF3
FGF4	FGF6	FGFR1 Introns 1, 5, Intron 17	FGFR2 Intron 1, Intron 17	FGFR3 Exons 7, 9 (alternative designation exon 10), 14, 18, Intron 17	FGFR4	FH	FLCN	FLT1
FLT3 Exons 14, 15, 20	FOXL2	FUBP1	GABRA6	GATA3	GATA4	GATA6	GID4 (C17orf39)	GNA11 Exons 4, 5
GNA13	GNAQ Exons 4, 5	GNAS Exons 1, 8	GRM3	GSK3B	H3-3A (H3F3A)	HDAC1	HGF	HNFI1A
HRAS Exons 2, 3	HSD3B1	ID3	IDH1 Exon 4	IDH2 Exon 4	IGF1R	IKBKE	IKZF1	INPP4B
IRF2	IRF4	IRS2	JAK1	JAK2 Exon 14	JAK3 Exons 5, 11, 12, 13, 15, 16	JUN	KDMSA	KDMS5C
KDM6A	KDR	KEAP1	KEL	KIT Exons 8, 9, 11, 12, 13, 17, Intron 16	KLHL6	KMT2A (MLL) Introns 6, 8-11, Intron 7	KMT2D (MLL2)	

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. · 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1437612-01

APPENDIX

Genes assayed in FoundationOne®Liquid CDx

FoundationOne Liquid CDx interrogates 324 genes, including 309 genes with complete exonic (coding) coverage and 15 genes with only select non-coding coverage (indicated with an *); 75 genes (indicated in bold) are captured with increased sensitivity and have complete exonic (coding) coverage unless otherwise noted.

KRAS	LTK	LYN	MAF	MAP2K1 (MEK1) Exons 2, 3	MAP2K2 (MEK2) Exons 2-4, 6, 7	MAP2K4	MAP3K1	MAP3K13
MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1	MERTK	MET
MITF	MKNK1	MLH1	MPL Exon 10	MRE11 (MRE11A)	MSH2 Intron 5	MSH3	MSH6	MST1R
MTAP	MTOR Exons 19, 30, 39, 40, 43-45, 47, 48, 53, 56	MUTYH	MYB* Intron 14	MYC Intron 1	MYCL (MYCL1)	MYCN	MYD88 Exon 4	NBN
NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2 Intron 26	NOTCH3	NPM1 Exons 4-6, 8, 10
NRAS Exons 2, 3	NSD2 (WHSC1 or MMSET)	NSD3 (WHSC1L1)	NTSC2	NTRK1 Exons 14, 15, Introns 8-11	NTRK2 Intron 12	NTRK3 Exons 16, 17	NUTM1* Intron 1	P2RY8
PALB2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA Exons 12, 18, Introns 7, 9, 11
PDGFRB Exons 12-21, 23	PDK1	PIK3C2B	PIK3C2G	PIK3CA Exons 2, 3, 5-8, 10, 14, 19, 21 (Coding Exons 1, 2, 4-7, 9, 13, 18, 20) PPP2R2A	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PRDM1	PRKAR1A	PRKCI	PRKN (PARK2)	
PTCH1	PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B
RAD51C	RAD51D	RAD52	RAD54L	RAF1 Exons 3, 4, 6, 7, 10, 14, 15, 17, Introns 4-8	RARA Intron 2	RB1	RBM10	REL
RET Introns 7, 8, Exons 11, 13-16, Introns 9-11	RICTOR	RNF43	ROS1 Exons 31, 36-38, 40, Introns 31-35	RPTOR	RSPO2* Intron 1	SDC4* Intron 2	SDHA	SDHB
SDHC	SDHD	SETD2	SF3B1	SGK1	SLC34A2* Intron 4	SMAD2	SMAD4	SMARCA4
SMARCB1	SMO	SNCAIP	SOC3	SOX2	SOX9	SPEN	SPOP	SRC
STAG2	STAT3	STK11	SUFU	SYK	TBX3	TEK	TENT5C (FAM46C)	TERC* ncRNA
TERT* Promoter	TET2	TGFB2	TIPARP	TMPSR2* Introns 1-3	TNFAIP3	TNFRSF14	TP53	TSC1
TSC2	TYRO3	U2AF1	VEGFA	VHL	WT1	XPO1	XRCC2	ZNF217
ZNF703								

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Microsatellite (MS) status
Blood Tumor Mutational Burden (bTMB)
Tumor Fraction

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. · 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1437612-01

APPENDIX

About FoundationOne®Liquid CDx

FoundationOne Liquid CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium. The CE-IVD regulatory status of FoundationOne Liquid CDx is applicable in countries that accept and/or recognize the CE mark.



ABOUT FOUNDATIONONE LIQUID CDx

FoundationOne Liquid CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne Liquid CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details.

INTENDED USE

FoundationOne Liquid CDx is a next generation sequencing based *in vitro* diagnostic device that analyzes 324 genes. Substitutions and insertion and deletion alterations (indels) are reported in 311 genes, copy number alterations (CNAs) are reported in 310 genes, and gene rearrangements are reported in 324 genes. The test also detects the genomic signatures blood tumor mutational burden (bTMB), microsatellite instability (MSI), and tumor fraction. FoundationOne Liquid CDx utilizes circulating cell-free DNA (cfDNA) isolated from plasma derived from the anti-coagulated peripheral whole blood of cancer patients. The test is intended to be used as a companion diagnostic to identify patients who may benefit from treatment with targeted therapies in accordance with the approved therapeutic product labeling. Additionally, FoundationOne Liquid CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with malignant neoplasms.

TEST PRINCIPLES

The FoundationOne Liquid CDx assay is performed exclusively as a laboratory service using circulating cell-free DNA (cfDNA) isolated from plasma derived from anti-coagulated peripheral whole blood from patients with solid malignant neoplasms. The assay employs a single DNA extraction method to obtain cfDNA from plasma from whole blood. Extracted

cfDNA undergoes whole-genome shotgun library construction and hybridization-based capture of 324 cancer-related genes including coding exons and select introns of 309 genes, as well as only select intronic regions or non-coding regions of 15 genes. Hybrid-capture selected libraries are sequenced with deep coverage using the NovaSeq® 6000 platform. Sequence data are processed using a customized analysis pipeline designed to accurately detect genomic alterations, including base substitutions, indels, select copy number variants, and select genomic rearrangements. Substitutions and insertion and deletion alterations (indels) are reported in 311 genes, copy number alterations (CNAs) are reported in 310 genes, and gene rearrangements are reported in 324 genes. The assay also reports tumor fraction, and genomic signatures including MSI and bTMB. A subset of targeted regions in 75 genes is baited for increased sensitivity.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. *Note:* A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

QUALIFIED ALTERATION CALLS (EQUIVOCAL)

All equivocal calls, regardless of alteration type, imply that there is adequate evidence to call the alteration with confidence. However, the repeatability of equivocal calls may be lower than non-equivocal calls.

RANKING OF THERAPIES AND CLINICAL TRIALS

Ranking of Therapies in Summary Table

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

LIMITATIONS

1. For *in vitro* diagnostic use.
2. For prescription use only. This test must be ordered by a qualified medical professional in accordance with clinical laboratory regulations.
3. A negative result does not rule out the presence of a mutation below the limits of detection of the assay. Patients for whom no companion diagnostic alterations are detected should be considered for confirmation with an appropriately validated tumor tissue test, if available.
4. The FoundationOne Liquid CDx assay does not detect heterozygous deletions.
5. The test is not intended to provide information on cancer predisposition.
6. Performance has not been validated for cfDNA input below the specified minimum input.
7. Tissue TMB and blood TMB (bTMB) are estimated from the number of synonymous and nonsynonymous single-nucleotide variants (SNVs) and insertions and deletions (indels) per area of coding genome sampled, after the removal of known and likely oncogenic driver events and germline SNPs. Tissue TMB is calculated based on variants with an allele frequency of $\geq 5\%$, and bTMB is calculated based on variants with an allele frequency of $\geq 0.5\%$.
8. Tumor fraction is the percentage of circulating tumor DNA (ctDNA) present in a cell-free DNA (cfDNA) sample. The tumor fraction estimate is computationally derived from the observed level of aneuploidy in the sample. Tumor fraction is considered elevated when ctDNA levels are high enough that aneuploidy can be detected and is significantly distinct from that typically found in non-tumor samples.
9. Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the tumor genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor. The MSI algorithm is based on genome wide analysis of 1765 microsatellite loci and not based on the 5 or 7 MSI loci described in current clinical practice guidelines for solid tissue testing.
10. Genomic findings from circulating cell-free DNA (cfDNA) may originate from circulating tumor DNA fragments, germline alterations, or non-tumor somatic alterations, such as clonal hematopoiesis of indeterminate potential (CHIP). Genes with alterations that may be derived from CHIP include, but are not limited

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. · 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1437612-01

APPENDIX
About FoundationOne® Liquid CDx

to: *ASXL1*, *ATM*, *CBL*, *CHEK2*, *DNMT3A*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, *TP53*, and *U2AF1*.

11. Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. If a reported alteration is suspected to be germline, confirmatory testing should be considered in the appropriate clinical context.
12. The test is not intended to replace germline testing or to provide information about cancer predisposition.

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >30%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to

distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *ATM*, *CBL*, *CHEK2*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne Liquid CDx.

TREATMENT DECISIONS ARE THE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this test or the information contained in this report.

Certain sample of variant characteristics may result in reduced sensitivity. These include: low sample quality, deletions and insertions >40bp, or repetitive/high homology sequences. FoundationOne Liquid CDx is performed using cell-free DNA, and as such germline events may not be reported.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. • 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1437612-01

APPENDIX

About FoundationOne®Liquid CDx

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
Muts/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

REFERENCE SEQUENCE INFORMATION

Sequence data is mapped to the human genome, Genome Reference Consortium Human Build 37 (GRCh37), also known as hg19.

MR Suite Version 7.0.0

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. · 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1437612-01

APPENDIX
References

1. Gandara DR, et al. Nat. Med. (2018) PMID: 30082870
2. Wang Z, et al. JAMA Oncol (2019) PMID: 30816954
3. Sturgill EG, et al. Oncologist (2022) PMID: 35274716
4. Aggarwal C, et al. Clin. Cancer Res. (2020) PMID: 32102950
5. Schenker et al., 2022; AACR Abstract CT022
6. Saori et al., 2021; ESMO Abstract 80P
7. Li et al., 2020; ASCO Abstract 6511
8. Cerami E, et al. Cancer Discov (2012) PMID: 22588877
9. Gao J, et al. Sci Signal (2013) PMID: 23550210
10. Tate JG, et al. Nucleic Acids Res. (2019) PMID: 30371878
11. Hu et al., 2017; ASCO Abstract e15791
12. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
13. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
14. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
15. Rizvi NA, et al. Science (2015) PMID: 25765070
16. Johnson BE, et al. Science (2014) PMID: 24336570
17. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
18. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
19. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
20. Heitzner E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
21. Nature (2012) PMID: 22810696
22. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
23. Bronkhorst AJ, et al. Biomol Detect Quantif (2019) PMID: 30923679
24. Raja R, et al. Clin. Cancer Res. (2018) PMID: 30093454
25. Hrebien S, et al. Ann. Oncol. (2019) PMID: 30860573
26. Choudhury AD, et al. JCI Insight (2018) PMID: 30385733
27. Goodall J, et al. Cancer Discov (2017) PMID: 28450425
28. Goldberg SB, et al. Clin. Cancer Res. (2018) PMID: 29330207
29. Bettgowda C, et al. Sci Transl Med (2014) PMID: 24553385
30. Lapin M, et al. J Transl Med (2018) PMID: 30400802
31. Shulman DS, et al. Br. J. Cancer (2018) PMID: 30131550
32. Stover DG, et al. J. Clin. Oncol. (2018) PMID: 29298117
33. Hemming ML, et al. JCO Precis Oncol (2019) PMID: 30793095
34. Egyud M, et al. Ann. Thorac. Surg. (2019) PMID: 31059681
35. Fan G, et al. PLoS ONE (2017) PMID: 28187169
36. Vu et al., 2020; DOI: 10.1200/PO.19.00204
37. Li G, et al. J Gastrointest Oncol (2019) PMID: 31602320
38. Zhang EW, et al. Cancer (2020) PMID: 32757294
39. Butler TM, et al. Cold Spring Harb Mol Case Stud (2019) PMID: 30833418
40. Nakano H, et al. Proc. Natl. Acad. Sci. U.S.A. (1984) PMID: 6320174
41. Pylyayeva-Gupta Y, et al. Nat. Rev. Cancer (2011) PMID: 21993244
42. Yamaguchi T, et al. Int. J. Oncol. (2011) PMID: 21523318
43. Watanabe M, et al. Cancer Sci. (2013) PMID: 23438367
44. Gilmartin AG, et al. Clin. Cancer Res. (2011) PMID: 21245089
45. Yeh JJ, et al. Mol. Cancer Ther. (2009) PMID: 19372556
46. Zhu X, et al. Lancet Oncol (2022) PMID: 35240087
47. Kinsey CG, et al. Nat. Med. (2019) PMID: 30833748
48. Bryant KL, et al. Nat. Med. (2019) PMID: 30833752
49. Mahalingam D, et al. Cancers (Basel) (2018) PMID: 29799479
50. Mahalingam D, et al. Clin. Cancer Res. (2019) PMID: 31694832
51. Noonan AM, et al. Mol. Ther. (2016) PMID: 27039845
52. Ko AH, et al. Clin. Cancer Res. (2016) PMID: 26251290
53. Chung V, et al. JAMA Oncol (2017) PMID: 27978579
54. Bedard PL, et al. Clin. Cancer Res. (2015) PMID: 25500057
55. Van Laethem JL, et al. Target Oncol (2017) PMID: 27975152
56. Infante JR, et al. Eur. J. Cancer (2013) PMID: 23583440
57. Infante JR, et al. Eur. J. Cancer (2014) PMID: 24915778
58. Van Cutsem E, et al. Int. J. Cancer (2018) PMID: 29756206
59. Bodoky G, et al. Invest New Drugs (2012) PMID: 21594619
60. Rinehart J, et al. J. Clin. Oncol. (2004) PMID: 15483017
61. Lorusso PM, et al. J. Clin. Oncol. (2005) PMID: 16009947
62. Infante JR, et al. Lancet Oncol. (2012) PMID: 22805291
63. Weekes CD, et al. Clin. Cancer Res. (2013) PMID: 23434733
64. Garrido-Laguna I, et al. Oncoscience (2015) PMID: 25897431
65. Guo C, et al. Lancet Oncol (2020) PMID: 33128873
66. Krebs et al., 2021; AACR Abstract CT019
67. Shinde et al., 2020; AACR Abstract CT143
68. Lu H, et al. Mol Cancer Ther (2019) PMID: 31068384
69. Mainardi S, et al. Nat Med (2018) PMID: 29808006
70. Koczywas et al., 2021; AACR Abstract LB001
71. Bendell et al., 2020; EORTC-NCI-AACR Abstract 5
72. Hillig RC, et al. Proc Natl Acad Sci U S A (2019) PMID: 30683722
73. Hofmann MH, et al. Cancer Discov (2021) PMID: 32816843
74. Hofmann et al., 2021; AACR Abstract CT210
75. Gort et al., 2020; ASCO Abstract TPS3651
76. Biankin AV, et al. Nature (2012) PMID: 23103869
77. Witkiewicz AK, et al. Nat Commun (2015) PMID: 25855536
78. Feldmann G, et al. J Hepatobiliary Pancreat Surg (2007) PMID: 17520196
79. Rachakonda PS, et al. PLoS ONE (2013) PMID: 23565280
80. Hruban RH, et al. Am. J. Pathol. (1993) PMID: 8342602
81. Maitra A, et al. Best Pract Res Clin Gastroenterol (2006) PMID: 16549325
82. Kahn S, et al. Anticancer Res. () PMID: 3310850
83. Akagi K, et al. Biochem. Biophys. Res. Commun. (2007) PMID: 17150185
84. Bollag G, et al. J. Biol. Chem. (1996) PMID: 8955068
85. Buhrman G, et al. Proc. Natl. Acad. Sci. U.S.A. (2010) PMID: 20194776
86. Sci. STKE (2004) PMID: 15367757
87. Edkins S, et al. Cancer Biol. Ther. (2006) PMID: 16969076
88. Feig LA, et al. Mol. Cell. Biol. (1988) PMID: 3043178
89. Gremer L, et al. Hum. Mutat. (2011) PMID: 20949621
90. Janakiraman M, et al. Cancer Res. (2010) PMID: 20570890
91. Kim E, et al. Cancer Discov (2016) PMID: 27147599
92. Lukman S, et al. PLoS Comput. Biol. (2010) PMID: 20838576
93. Naguib A, et al. J Mol Signal (2011) PMID: 21371307
94. Prior IA, et al. Cancer Res. (2012) PMID: 22589270
95. Privé GG, et al. Proc. Natl. Acad. Sci. U.S.A. (1992) PMID: 1565661
96. Scheffzek K, et al. Science (1997) PMID: 9219684
97. Scholl C, et al. Cell (2009) PMID: 19490892
98. Smith G, et al. Br. J. Cancer (2010) PMID: 20147967
99. Tyner JW, et al. Blood (2009) PMID: 19075190
100. Valencia A, et al. Biochemistry (1991) PMID: 2029511
101. White Y, et al. Nat Commun (2016) PMID: 26854029
102. Wiest JS, et al. Oncogene (1994) PMID: 8058307
103. Angeles AKJ, et al. Oncol Lett (2019) PMID: 31289513
104. Tong JH, et al. Cancer Biol. Ther. (2014) PMID: 24642870
105. Loree JM, et al. Clin Cancer Res (2021) PMID: 34117033
106. Mancini A, et al. J. Biol. Chem. (2002) PMID: 11847211
107. Miyake S, et al. Proc. Natl. Acad. Sci. U.S.A. (1998) PMID: 9653117
108. Masson K, et al. Biochem. J. (2006) PMID: 16780420
109. Singh AJ, et al. Proc. Natl. Acad. Sci. U.S.A. (2007) PMID: 17372230
110. Paolino M, et al. Nature (2014) PMID: 24553136
111. Patwardhan PP, et al. Oncotarget (2016) PMID: 26675259
112. Bazhenova et al., 2018; ESMO Abstract 4080
113. Shtiegman K, et al. Oncogene (2007) PMID: 17486068
114. Padrón D, et al. Cancer Res. (2007) PMID: 17699773
115. Hosaka T, et al. Anticancer Res. () PMID: 17695511
116. Han W, et al. Cancer Biol. Ther. (2006) PMID: 16969069
117. Yang S, et al. Cancer Res. (2006) PMID: 16849543
118. Sargin B, et al. Blood (2007) PMID: 17446348
119. Oshikawa G, et al. J. Biol. Chem. (2011) PMID: 21768087
120. Reindl C, et al. Clin. Cancer Res. (2009) PMID: 19276253
121. Taylor SJ, et al. Blood (2012) PMID: 22990016
122. Makishima H, et al. Leukemia (2012) PMID: 22246246
123. Bailey P, et al. Nature (2016) PMID: 26909576
124. Donahue TR, et al. Clin. Cancer Res. (2012) PMID: 22261810
125. Kadera BE, et al. Clin. Cancer Res. (2015) PMID: 25348515
126. Bacher U, et al. Ann. Hematol. (2010) PMID: 20195608
127. Miyake S, et al. J. Biol. Chem. (1999) PMID: 10347229
128. Polzer H, et al. Exp. Hematol. (2013) PMID: 23127761
129. Levkowitz G, et al. Genes Dev. (1998) PMID: 9851973
130. Bunda S, et al. Cancer Res. (2013) PMID: 23400592
131. Andoniu CE, et al. EMBO J. (1994) PMID: 7925293
132. Aranaz P, et al. Haematologica (2012) PMID: 22315494
133. Fernandes MS, et al. J. Biol. Chem. (2010) PMID: 20622007
134. Grand FH, et al. Blood (2009) PMID: 19387008
135. Javadi M, et al. J. Biol. Chem. (2013) PMID: 23696637
136. Kassenbrock CK, et al. J. Biol. Chem. (2004) PMID: 15117950
137. Levkowitz G, et al. Mol. Cell (1999) PMID: 10635327
138. Loh ML, et al. Blood (2009) PMID: 19571318
139. Martinelli S, et al. Am. J. Hum. Genet. (2010) PMID: 20619386
140. Saito Y, et al. Leuk. Res. (2012) PMID: 22591685
141. Sanada M, et al. Nature (2009) PMID: 19620960
142. Score J, et al. Blood (2012) PMID: 22053108
143. Shiba N, et al. Leukemia (2011) PMID: 21494262
144. Standaert ML, et al. Biochemistry (2004) PMID: 15581361
145. Tan YH, et al. PLoS ONE (2010) PMID: 20126411
146. Thien CB, et al. Mol. Cell (2001) PMID: 11239464
147. Visser GD, et al. Exp. Cell Res. (2005) PMID: 16246327
148. Li M, et al. Cancer Res. (2016) PMID: 26676746
149. Jaiswal S, et al. N. Engl. J. Med. (2014) PMID: 25426837
150. Genovesse G, et al. N. Engl. J. Med. (2014) PMID: 25426838
151. Xie M, et al. Nat. Med. (2014) PMID: 25326804
152. Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) PMID: 28669404
153. Severson EA, et al. Blood (2018) PMID: 29678827
154. Fuster JJ, et al. Circ. Res. (2018) PMID: 29420212
155. Hematology Am Soc Hematol Educ Program (2018) PMID: 30504320

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. · 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1437612-01

APPENDIX
References

156. Chabon JJ, et al. Nature (2020) pmid: 32269342
157. Razavi P, et al. Nat. Med. (2019) pmid: 31768066
158. Trowbridge JJ, et al. Nat. Genet. (2011) pmid: 22200773
159. Prog Mol Biol Transl Sci (2011) pmid: 21507354
160. Yang J, et al. Mol Med Rep () pmid: 21887466
161. Vallböhmer D, et al. Clin Lung Cancer (2006) pmid: 16870044
162. Daskalos A, et al. Cancer (2011) pmid: 21351083
163. Fabbri M, et al. Proc. Natl. Acad. Sci. U.S.A. (2007) pmid: 17890317
164. Gao Q, et al. Proc. Natl. Acad. Sci. U.S.A. (2011) pmid: 22011581
165. Kim MS, et al. APMIS (2013) pmid: 23031157
166. Chen ZX, et al. J. Cell. Biochem. (2005) pmid: 15861382
167. Guo X, et al. Nature (2015) pmid: 25383530
168. Sandoval JE, et al. J. Biol. Chem. (2019) pmid: 30705090
169. Zhang ZM, et al. Nature (2018) pmid: 29414941
170. Hirai H, et al. Cancer Biol. Ther. (2010) pmid: 20107315
171. Bridges KA, et al. Clin. Cancer Res. (2011) pmid: 21799033
172. Rajeshkumar NV, et al. Clin. Cancer Res. (2011) pmid: 21389100
173. Osman AA, et al. Mol. Cancer Ther. (2015) pmid: 25504633
174. Xu L, et al. Mol. Cancer Ther. (2002) pmid: 12489850
175. Xu L, et al. Mol. Med. (2001) pmid: 11713371
176. Camp ER, et al. Cancer Gene Ther. (2013) pmid: 23470564
177. Kim SS, et al. Nanomedicine (2015) pmid: 25240597
178. Pirollo KF, et al. Mol. Ther. (2016) pmid: 27357628
179. Hajdenberg et al., 2012; ASCO Abstract e15010
180. Leijen S, et al. J. Clin. Oncol. (2016) pmid: 27601554
181. Moore et al., 2019; ASCO Abstract 5513
182. Leijen S, et al. J. Clin. Oncol. (2016) pmid: 27998224
183. Oza et al., 2015; ASCO Abstract 5506
184. Lee J, et al. Cancer Discov (2019) pmid: 31315834
185. Méndez E, et al. Clin. Cancer Res. (2018) pmid: 29535125
186. Seligmann JF, et al. J Clin Oncol (2021) pmid: 34538072
187. Lehmann S, et al. J. Clin. Oncol. (2012) pmid: 22965953
188. Mohell N, et al. Cell Death Dis (2015) pmid: 26086967
189. Fransson Å, et al. J Ovarian Res (2016) pmid: 27179933
190. Gourley et al., 2016; ASCO Abstract 5571
191. Kwok M, et al. Blood (2016) pmid: 26563132
192. Boudny M, et al. Haematologica (2019) pmid: 30975914
193. Dillon MT, et al. Mol. Cancer Ther. (2017) pmid: 28062704
194. Middleton FK, et al. Cancers (Basel) (2018) pmid: 30127241
195. Morton JP, et al. Proc. Natl. Acad. Sci. U.S.A. (2010) pmid: 20018721
196. Scarpa A, et al. Am. J. Pathol. (1993) pmid: 8494051
197. Luo Y, et al. Pathol. Oncol. Res. (2013) pmid: 22782330
198. Iacobuzio-Donahue CA, et al. Clin. Cancer Res. (2012) pmid: 22896692
199. Macgregor-Das AM, et al. J Surg Oncol (2013) pmid: 22806689
200. Oshima M, et al. Ann. Surg. (2013) pmid: 23470568
201. Ottenhof NA, et al. Cell Oncol (Dordr) (2012) pmid: 22351431
202. Tsiambas E, et al. J BUON () pmid: 20414934
203. Ansari D, et al. Br J Surg (2011) pmid: 21644238
204. Grochola LF, et al. Pancreas (2011) pmid: 21404460
205. Brown CJ, et al. Nat. Rev. Cancer (2009) pmid: 19935675
206. Joerger AC, et al. Annu. Rev. Biochem. (2008) pmid: 18410249
207. Kato S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) pmid: 12826609
208. Kamada R, et al. J. Biol. Chem. (2011) pmid: 20978130
209. Zerdoumi Y, et al. Hum. Mol. Genet. (2017) pmid: 28472496
210. Yamada H, et al. Carcinogenesis (2007) pmid: 17690113
211. Landrum MJ, et al. Nucleic Acids Res. (2018) pmid: 29165669
212. Bougeard G, et al. J. Clin. Oncol. (2015) pmid: 26014290
213. Sorrell AD, et al. Mol Diagn Ther (2013) pmid: 23355100
214. Nichols KE, et al. Cancer Epidemiol. Biomarkers Prev. (2001) pmid: 11219776
215. Kleihues P, et al. Am. J. Pathol. (1997) pmid: 9006316
216. Gonzalez KD, et al. J. Clin. Oncol. (2009) pmid: 19204208
217. Lalloo F, et al. Lancet (2003) pmid: 12672316
218. Mandelker D, et al. Ann. Oncol. (2019) pmid: 31050713

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Giles Maule, M.D., Ph.D | 26 August 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. · 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531