

ABOUT THE TEST FoundationOne® Heme is a comprehensive genomic profiling test designed to identify genomic alterations within hundreds of cancer-related genes in hematologic malignancies and sarcomas.

PATIENT	DISEASE Soft tissue sarcoma (NOS)	PHYSICIAN	ORDERING PHYSICIAN Yeh, Yi-Chen	SPECIMEN	SPECIMEN SITE Lung
	NAME Li, Min-Ling		MEDICAL FACILITY Taipei Veterans General Hospital		SPECIMEN ID S110-773268 (PF22021)
	DATE OF BIRTH 28 April 1967		ADDITIONAL RECIPIENT None		SPECIMEN TYPE Slide Deck
	SEX Female		MEDICAL FACILITY ID 205872		DATE OF COLLECTION 08 September 2021
	MEDICAL RECORD # 45579666		PATHOLOGIST Not Provided		SPECIMEN RECEIVED 23 February 2022

Biomarker Findings

Microsatellite status - MS-Stable
Tumor Mutational Burden - 3 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

TSC2 deletion exons 4-7
CHD2 rearrangement intron 38
GTSE1 R569C
RB1 loss exons 18-27
TP53 C176fs*71

† See About the Test in appendix for details.

Report Highlights

- Targeted therapies with potential clinical benefit **approved in another tumor type**: Nab-sirolimus (p. 6)
- Evidence-matched **clinical trial options** based on this patient's genomic findings: (p. 7)

BIOMARKER FINDINGS

Microsatellite status - MS-Stable

Tumor Mutational Burden - 3 Muts/Mb

GENOMIC FINDINGS

TSC2 - deletion exons 4-7

10 Trials see p. 7

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. see Biomarker Findings section

No therapies or clinical trials. see Biomarker Findings section

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)

none

THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)

Nab-sirolimus

GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

CHD2 - rearrangement intron 38	p. 3	RB1 - loss exons 18-27	p. 4
GTSE1 - R569C	p. 3	TP53 - C176fs*71	p. 5

NOTE Genomic alterations detected may be associated with activity of certain FDA-approved drugs; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type.

Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Lena Stuart, M.D. | 09 March 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1308441-01

BIOMARKER FINDINGS
BIOMARKER

Microsatellite status

RESULT

MS-Stable

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁻³, including approved therapies nivolumab and pembrolizumab⁴. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, $p=0.001$)⁵.

FREQUENCY & PROGNOSIS

Reports of MSI in sarcomas in the literature are conflicting and varied due to substantial heterogeneity, lack of consensus on the markers and methods used for MSI assessment, and small sample size in most studies⁶. In a computational analysis of paired tumor and normal sarcomas in the TCGA dataset, of which 40% were leiomyosarcomas and 25% were liposarcomas, only 0.8% (2/255) of samples were MSI-high (MSI-H)⁷. In smaller studies of soft tissue sarcoma, reports of MSI at any level have been rare, with the highest incidences between 11% (2/18) to 25% (10/40) of cases⁸⁻¹³. In one study, MSI was reported to occur more frequently in high-grade soft tissue sarcomas compared with lower grade¹⁴. However, published data investigating the prognostic implications of MSI in sarcoma are limited (PubMed, Jan 2022).

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor¹⁵. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2¹⁵⁻¹⁷. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers¹⁸⁻²⁰. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{15,17,19-20}.

BIOMARKER

Tumor Mutational Burden

RESULT

3 Muts/Mb

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1²¹⁻²³, anti-PD-1 therapies²¹⁻²⁴, and combination nivolumab and ipilimumab²⁵⁻³⁰. In multiple pan-tumor studies, higher TMB has been reported to be associated with increased ORR and OS from treatment with immune checkpoint inhibitors^{21-24,31}. Higher TMB was found to be significantly associated with improved OS upon immune checkpoint inhibitor treatment for patients with 9 types of advanced tumors²¹. Analyses across several solid tumor types reported that patients with higher TMB (defined as ≥ 16 -20 Muts/Mb) achieved greater clinical benefit from

PD-1 or PD-L1-targeting monotherapy, compared with patients with higher TMB treated with chemotherapy³² or those with lower TMB treated with PD-1 or PD-L1-targeting agents²². However, the KEYNOTE 158 trial of pembrolizumab monotherapy for patients with solid tumors found significant improvement in ORR for patients with TMB ≥ 10 Muts/Mb (based on this assay or others) compared to those with TMB < 10 Muts/Mb, in a large cohort that included multiple tumor types; similar findings were observed in the KEYNOTE 028 and 012 trials^{24,31}. Together, these studies suggest that patients with TMB ≥ 10 Muts/Mb may derive clinical benefit from PD-1 or PD-L1 inhibitors.

FREQUENCY & PROGNOSIS

Soft tissue sarcomas harbor a median TMB of 2.5 Muts/Mb, with angiosarcoma (13.4%) and malignant peripheral nerve sheath tumor (MPNST) (8.2%) having the highest percentage of cases with high TMB (> 20 Muts/Mb)³³. Increased mutation burden has been reported in undifferentiated pleomorphic sarcomas as compared to Ewing sarcomas or rhabdomyosarcomas³⁴⁻³⁶. Published data investigating the prognostic implications of TMB

in sarcoma are limited (PubMed, Feb 2022). High TMB was associated with improved PFS and metastasis-free survival in a study of undifferentiated sarcomas³⁷ and with reduced survival in a study of patients with rhabdomyosarcoma³⁸.

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma³⁹⁻⁴⁰ and cigarette smoke in lung cancer⁴¹⁻⁴², treatment with temozolomide-based chemotherapy in glioma⁴³⁻⁴⁴, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes⁴⁵⁻⁴⁹, and microsatellite instability (MSI)^{45,48-49}. This sample harbors a TMB level associated with lower rates of clinical benefit from treatment with PD-1- or PD-L1-targeting immune checkpoint inhibitors compared with patients with tumors harboring higher TMB levels, based on several studies in multiple solid tumor types^{22-23,31}.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Lena Stuart, M.D. | 09 March 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 • CLIA: 34D2044309
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1308441-01

GENOMIC FINDINGS
GENE

TSC2

ALTERATION

deletion exons 4-7

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

Loss or inactivation of TSC2 can activate mTOR signaling⁵⁰. mTOR inhibitors such as everolimus, temsirolimus, and sirolimus have shown activity against tumors associated with the genetic disease tuberous sclerosis complex (TSC), including subependymal giant cell astrocytomas and renal angiomyolipomas⁵¹⁻⁵⁶. Sirolimus and nab-sirolimus have shown activity for patients with TSC2-altered malignant perivascular epithelioid cell tumors (PEComas)⁵⁷⁻⁵⁹. Nab-sirolimus has also shown limited activity for patients with TSC2-mutated sarcomas⁶⁰. In the context of TSC2-altered malignancies unrelated to TSC, everolimus and temsirolimus activity has been limited⁶¹⁻⁶³ with the exception of anecdotal reports

across various solid tumors, including anaplastic thyroid cancer⁶⁴, renal cell carcinoma (RCC)⁶⁵⁻⁶⁶, glioblastoma⁶⁷, and CNS embryonal tumor⁶⁸, as well as a case of Hodgkin lymphoma⁶⁹. In the prospective NCI-MATCH study, only 6.7% (1/15) of patients with TSC2-mutated solid tumors responded to everolimus, with the single response reported for 1 patient with uterine leiomyosarcoma⁶¹. Retrospective analyses of the RECORD-3, GRANITE-1, and EVOLVE-1 studies of everolimus to treat patients with RCC, gastric cancer, or hepatocellular carcinoma, respectively, showed no significant association between alterations in mTOR, TSC1, or TSC2 and median PFS⁷⁰.

FREQUENCY & PROGNOSIS

TSC2 mutation or homozygous deletion was not observed in any of 207 sarcoma cases in one genomic study⁷¹. TSC2 mutations have been described in chordomas, perivascular epithelioid cell tumors, uterine angiosarcoma, and rhabdoid tumors, among others^{58,72-74}. The mTOR pathway is frequently activated in various sarcomas⁷⁵⁻⁷⁹. Published data investigating the prognostic

implications of TSC2 alterations in sarcomas are limited (PubMed, Jan 2022).

FINDING SUMMARY

The tumor suppressor protein Tuberin (TSC2) binds with Hamartin (TSC1) to inhibit mTOR signaling and cell growth^{50,80}. Alterations such as seen here may disrupt TSC2 function or expression⁸¹⁻⁸³.

POTENTIAL GERMLINE IMPLICATIONS

Inactivating germline mutations in TSC2 are associated with the autosomal dominant disorder tuberous sclerosis complex, which results in the development of hamartomas in multiple organ systems and an increased risk of developing renal cell carcinoma (RCC)⁸⁴⁻⁸⁶. TSC2 mutations account for approximately 75 to 80% of reported sporadic cases⁸⁷. Prevalence for this disorder in the general population is estimated to be 1/6,000 from birth and 1/12,000 to 1/14,000 in children under 10 years of age⁸⁷. In the appropriate clinical context, germline testing of TSC2 is recommended.

GENE

CHD2

ALTERATION

rearrangement intron 38

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

There are no therapies available to directly address genomic alterations in CHD2.

FREQUENCY & PROGNOSIS

Somatic mutations in CHD2 have been reported in 3% of all cancers in COSMIC, most frequently in skin (10.1%), esophagus (9.9%), pancreas (9.9%), liver (9.0%), gastric (8.7%), and prostate (7.7%) cancers (COSMIC, Jan 2022)⁸⁸. Loss-of-function mutations in CHD2 have been observed in nearly 50% of MSI-high colorectal and gastric cancers⁸⁹, and differential expression of CHD2 was reported to be associated with colon cancer progression⁹⁰. Deletion of this gene has also been observed in a Hodgkin lymphoma cell line⁹¹. In agreement with these findings, preclinical research has suggested

that CHD2 is a tumor suppressor that plays roles in the DNA damage response⁹².

FINDING SUMMARY

CHD2 encodes chromodomain helicase DNA binding protein 2, an ATPase/helicase that alters gene expression by modifying chromatin structure. Germline deletions and mutations in CHD2 are associated with several epilepsy syndromes, including Dravet syndrome and Lennox-Gastaut syndrome⁹³⁻⁹⁵.

GENE

GTSE1

ALTERATION

R569C

TRANSCRIPT ID

NM_016426

CODING SEQUENCE EFFECT

1705C>T

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

There are no therapies available to target alterations in GTSE1.

FREQUENCY & PROGNOSIS

Mutations in this gene have been observed in 1-4% of various solid tumor types (COSMIC, Jun 2021)⁸⁸. GTSE1 expression correlates with cell migration, invasive potential, and poor prognosis

in breast cancer⁹⁶⁻⁹⁷, bladder cancer⁹⁸, and hepatocellular carcinoma⁹⁹⁻¹⁰⁰. However, no correlations were found between GTSE1 expression levels and clinical data in lung cancer¹⁰¹.

FINDING SUMMARY

GTSE1 encodes a microtubule-interacting protein that interacts with p53 in response to DNA damage. Studies investigating the role of GTSE1 in cell invasiveness have generated mixed results.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Lena Stuart, M.D. | 09 March 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1308441-01

GENOMIC FINDINGS

GENE

RB1

ALTERATION

loss exons 18-27

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of limited clinical data¹⁰² and strong preclinical data¹⁰³⁻¹⁰⁵, RB1 inactivation may be associated with sensitivity to inhibitors of Aurora kinase A, particularly in small cell lung cancer. It should be noted that a trial of the Aurora kinase A inhibitor alisertib in advanced prostate cancer did not find an association between RB1 deletion and clinical benefit¹⁰⁶. Other approaches to target RB1 inactivation under investigation in preclinical studies include inhibitors of BCL-2 family members¹⁰⁷ and activation of the NOTCH pathway¹⁰⁸.

— Potential Resistance —

Rb inactivation may predict resistance to CDK4/6 inhibitors such as palbociclib, abemaciclib, and ribociclib, which act upstream of Rb¹⁰⁹⁻¹¹⁸.

— Nontargeted Approaches —

Loss of Rb function has been associated with increased sensitivity to cytotoxic agents and chemotherapeutics in both preclinical studies and in patients with bladder or breast cancer¹¹⁹⁻¹²⁰.

FREQUENCY & PROGNOSIS

RB1 mutations have been reported in 4.2% of soft tissue tumors analyzed in the COSMIC database (Jul 2021)⁸⁸. Putative homozygous deletion of RB1 was reported in 7.7% of soft tissue sarcomas, most frequently in pleomorphic liposarcoma (25%, 6/24), myxofibrosarcoma (18%, 7/38), and leiomyosarcoma (11%, 3/27)⁷¹. One study reported homozygous deletion of RB1 in 2/36 (5.5%) of undifferentiated pleomorphic sarcomas (previously called malignant fibrous histiocytoma), but loss of RB1 expression in 30/35 (86%), suggesting that loss of RB1 plays a pivotal role in

the pathogenesis of this group of soft tissue sarcomas¹²¹. In one study, decreased Rb protein expression was associated with improved overall survival in patients with soft tissue sarcoma¹²².

FINDING SUMMARY

RB1 encodes the retinoblastoma protein (Rb), a tumor suppressor and negative regulator of the cell cycle^{120,123}. Alterations such as seen here may disrupt RB1 function or expression¹²⁴⁻¹³⁰.

POTENTIAL GERMLINE IMPLICATIONS

Mutations in RB1 underlie the development of retinoblastoma (RB), a rare tumor that arises at a rate of approximately 1:20,000 live births, with nearly 5,000 new cases worldwide per year¹³¹. Germline mutations in RB1 account for approximately 40% of RB tumors¹³² and are associated with an increased risk of developing secondary malignancies that include soft tissue and bone sarcoma and malignant melanoma¹³³⁻¹³⁴. In the appropriate clinical context, germline testing of RB1 is recommended.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Lena Stuart, M.D. | 09 March 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1308441-01

GENOMIC FINDINGS
GENE
TP53
ALTERATION

C176fs*71

TRANSCRIPT ID

NM_000546

CODING SEQUENCE EFFECT

526delT

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor adavosertib¹³⁵⁻¹³⁸, or p53 gene therapy and immunotherapeutics such as SGT-53¹³⁹⁻¹⁴³ and ALT-801¹⁴⁴. In a Phase 1 study, adavosertib in combination with gemcitabine, cisplatin, or carboplatin elicited PRs in 9.7% and SDs in 53% of patients with solid tumors; the response rate was 21% (4/19) for patients with TP53 mutations versus 12% (4/33) for patients who were TP53 wildtype¹⁴⁵. A Phase 2 trial of adavosertib in combination with chemotherapy (gemcitabine, carboplatin, paclitaxel, or doxorubicin) reported a 32% (30/94, 3 CR) ORR and a 73% (69/94) DCR for patients with platinum-refractory TP53-mutated ovarian, Fallopian tube, or peritoneal cancer¹⁴⁶. A smaller Phase 2 trial of adavosertib in combination with carboplatin achieved a 43% (9/21, 1 CR) ORR and a 76% (16/21) DCR for patients with platinum-refractory TP53-mutated ovarian cancer¹⁴⁷. The combination of adavosertib with paclitaxel and carboplatin for patients with TP53-mutated ovarian cancer also significantly increased PFS compared with paclitaxel and carboplatin alone¹⁴⁸. In the Phase 2 VIKTORY trial, patients with

TP53-mutated metastatic and/or recurrent gastric cancer experienced a 24% (6/25) ORR with adavosertib combined with paclitaxel¹⁴⁹. A Phase 1 trial of neoadjuvant adavosertib in combination with cisplatin and docetaxel for head and neck squamous cell carcinoma (HNSCC) elicited a 71% (5/7) response rate for patients with TP53 alterations¹⁵⁰. The Phase 2 FOCUS4-C trial for patients with TP53- and RAS-mutated colorectal cancer reported improvement in PFS (3.61 vs. 1.87 months, HR=0.35, p=0.0022), but not OS (14.0 vs 12.8 months, p=0.93), following adavosertib treatment compared with active monitoring¹⁵¹. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel for patients with solid tumors, 75% (9/12) of evaluable patients experienced clinical benefit, including 2 confirmed and 1 unconfirmed PRs and 2 instances of SD with significant tumor shrinkage¹⁴³. ATR inhibitor treatment of chronic lymphocytic leukemia (CLL) cells with biallelic inactivation of TP53 suppressed cell viability, promoted DNA damage, and attenuated xenograft growth in preclinical studies¹⁵²⁻¹⁵³; however, ATR inhibitors as monotherapy had little effect on these parameters in solid tumor models in other preclinical studies¹⁵⁴⁻¹⁵⁵. Therefore, it is unclear whether TP53 inactivation predicts sensitivity to ATR inhibition.

FREQUENCY & PROGNOSIS

TP53 mutations and homozygous deletion have been observed in 33% and 10% of sarcoma samples in the TCGA dataset, respectively (cBioPortal, Feb 2022)¹⁵⁶⁻¹⁵⁷. TP53 alterations appear to lead to chromosomal instability and drive oncogenesis in soft tissue sarcomas¹⁵⁸. One study of soft tissue sarcomas reported that TP53 non-frameshift mutations correlated with poor prognosis, including lymph node metastasis, increased rate of relapse, and decreased overall survival¹⁵⁹.

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers¹⁶⁰. Alterations such as seen here may disrupt TP53 function or expression¹⁶¹⁻¹⁶⁵.

POTENTIAL GERMLINE IMPLICATIONS

Germline mutations in TP53 are associated with the very rare autosomal dominant disorder Li-Fraumeni syndrome and the early onset of many cancers¹⁶⁶⁻¹⁶⁸, including sarcomas¹⁶⁹⁻¹⁷⁰. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000¹⁷¹ to 1:20,000¹⁷⁰. For pathogenic TP53 mutations identified during tumor sequencing, the rate of germline mutations was 1% in the overall population and 6% in tumors arising before age 30¹⁷². In the appropriate clinical context, germline testing of TP53 is recommended.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion¹⁷³⁻¹⁷⁸. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy¹⁷³⁻¹⁷⁴. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease¹⁷⁹. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH^{177,180-181}. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Lena Stuart, M.D. | 09 March 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1308441-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Nab-sirolimus

Assay findings association
TSC2
deletion exons 4-7

AREAS OF THERAPEUTIC USE

Nab-sirolimus is an intravenous nanoparticle albumin-bound mTOR inhibitor that is FDA approved to treat adult patients with advanced or metastatic perivascular epithelioid cell tumors (PEComas). Please see the drug label for full prescribing information.

GENE ASSOCIATION

Based on a prospective study that showed a higher ORR for patients with TSC2-altered PEComas treated with nab-sirolimus than for those with neither TSC1- nor TSC2-altered disease⁵⁹ and additional responses reported for individual patients with TSC2-altered endometrial stromal sarcoma, leiomyosarcoma, high grade sarcoma (NOS), or lymphangioleiomyomatosis (LAM)⁶⁰, TSC2 mutation or loss may predict sensitivity to nab-sirolimus.

SUPPORTING DATA

Nab-sirolimus has been primarily investigated for the treatment of malignant PEComa but is also being explored to treat TSC2- or TSC1-altered solid tumors. The Phase 2 AMPECT trial of nab-sirolimus for patients with malignant PEComa prospectively showed a significantly higher ORR for patients with TSC2-altered PEComa relative to those lacking TSC2 alterations (89%; 8/9 vs. 13%; 2/16, $p < 0.001$)⁵⁹. In an expanded access program, 50% (3/6) of patients with TSC2-mutated malignant PEComa that had previously progressed on rapalogs, including everolimus, temsirolimus, and sirolimus, exhibited a PR when treated with nab-sirolimus¹⁸². Outside of malignant PEComa, PRs were reported for individual patients with TSC2-altered endometrial stromal sarcoma, leiomyosarcoma, high-grade sarcoma, or lymphangioleiomyomatosis treated with nab-sirolimus⁶⁰.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Lena Stuart, M.D. | 09 March 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1308441-01

CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov. Or visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

GENE
TSC2
ALTERATION

deletion exons 4-7

RATIONALE

Inactivating TSC2 alterations may lead to increased mTOR activation and predict sensitivity

to mTOR inhibitors.

NCT03239015
PHASE 2

Efficacy and Safety of Targeted Precision Therapy in Refractory Tumor With Druggable Molecular Event

TARGETS

EGFR, ERBB4, ERBB2, PARP, mTOR, MET, ROS1, RET, VEGFRs, BRAF, CDK4, CDK6

LOCATIONS: Shanghai (China)

NCT04337463
PHASE NULL

ATG-008 Combined With Toripalimab in Advanced Solid Tumors

TARGETS

mTORC1, mTORC2, PD-1

LOCATIONS: Chongqing (China), Chengdu (China)

NCT04803318
PHASE 2

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

TARGETS

mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK

LOCATIONS: Guangzhou (China)

NCT03297606
PHASE 2

Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)

TARGETS

VEGFRs, ABL, SRC, ALK, ROS1, AXL, TRKA, MET, TRKC, DDR2, KIT, EGFR, PD-1, CTLA-4, PARP, CDK4, CDK6, FLT3, CSF1R, RET, mTOR, ERBB2, MEK, BRAF, SMO

LOCATIONS: Vancouver (Canada), Edmonton (Canada), Saskatoon (Canada), Regina (Canada), Ottawa (Canada), Montreal (Canada), Toronto (Canada), Kingston (Canada), London (Canada)

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Lena Stuart, M.D. | 09 March 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1308441-01

CLINICAL TRIALS
NCT03660930
PHASE 1/2

Nanoparticle Albumin-Bound Rapamycin and Pazopanib Hydrochloride in Patients With Nonadipocytic Soft Tissue Sarcomas

TARGETS
 mTOR, FGFR3, KIT, FGFR1, VEGFRs, FGFR2

LOCATIONS: Washington

NCT02693535
PHASE 2

TAPUR: Testing the Use of Food and Drug Administration (FDA) Approved Drugs That Target a Specific Abnormality in a Tumor Gene in People With Advanced Stage Cancer

TARGETS
 VEGFRs, ABL, SRC, ALK, ROS1, AXL, TRKA, MET, TRKC, CDK4, CDK6, FLT3, CSF1R, KIT, RET, mTOR, EGFR, ERBB2, MEK, BRAF, SMO, DDR2, PARP, PD-1, CTLA-4, ERBB4

LOCATIONS: Hawaii, Washington, Oregon, California

NCT04185831
PHASE 2

A Molecularly Guided Anti-Cancer Drug Off-Label Trial

TARGETS
 PD-L1, MEK, mTOR

LOCATIONS: Uppsala (Sweden), Gothenburg (Sweden)

NCT03778996
PHASE 2

SM-88 as Maintenance Therapy for Advanced Ewing's Sarcoma Patients and as Salvage Therapy for Sarcoma Patients

TARGETS
 mTOR

LOCATIONS: California

NCT02584647
PHASE 1/2

PLX3397 Plus Sirolimus in Unresectable Sarcoma and Malignant Peripheral Nerve Sheath Tumors

TARGETS
 mTOR, CSF1R, KIT, FLT3

LOCATIONS: Iowa, Michigan, Missouri, New York

NCT03065062
PHASE 1

Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head & Neck and Other Solid Tumors

TARGETS
 PI3K-alpha, PI3K-gamma, mTORC1, mTORC2, CDK4, CDK6

LOCATIONS: Massachusetts

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

 Electronically signed by Lena Stuart, M.D. | 09 March 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

 Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1308441-01

APPENDIX
Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future. Please note that some VUS rearrangements between targeted genes and unknown fusion partners or intergenic regions detected by RNA sequencing may not be reported.

BRCA2
D293E

CARD11
T532M

DAXX
E457del

FANCA
K1297N

FBXO11
Q29_P32del

GPR124
K1288del

JAK3
R222H

P2RY8
M16K

PCLO
V71A

SETBP1
Y994*

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Lena Stuart, M.D. | 09 March 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1308441-01

APPENDIX
Genes Assayed in FoundationOne®Heme

FoundationOne Heme is designed to include genes known to be somatically altered in human hematologic malignancies and sarcomas that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay utilizes DNA sequencing to interrogate 406 genes as well as selected introns of 31 genes involved in rearrangements, in addition to RNA sequencing of 265 genes. The assay will be updated periodically to reflect new knowledge about cancer biology.

HEMATOLOGICAL MALIGNANCY DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACTB	AKT1	AKT2	AKT3	ALK	AMER1 (FAM123B or WTX)	APC
APH1A	AR	ARAF	ARFRP1	ARHGAP26 (GRAF)		ARID1A	ARID2
ASXL1	ATM	ATR	ATRX	AURKA	AURKB	AXIN1	AXL
BAP1	BARD1	BCL10	BCL11B	BCL2	BCL2L2	BCL6	BCL7A
BCORL1	BIRC3	BLM	BRAF	BRCA1	BRCA2	BRD4	BRIP1
BTG2	BTK	BTLA	C11orf30 (EMSY)	CAD	CALR*	CARD11	CBFB
CCND1	CCND2	CCND3	CCNE1	CCT6B	CD22	CD274 (PD-L1)	CD36
CD70	CD79A	CD79B	CDC73	CDH1	CDK12	CDK4	CDK6
CDKN1B	CDKN2A	CDKN2B	CDKN2C	CEBPA	CHD2	CHEK1	CHEK2
CIITA	CKS1B	CPS1	CREBBP	CRKL	CRLF2	CSF1R	CSF3R
CTNNA1	CTNNB1	CUX1	CXCR4	DAXX	DDR2	DDX3X	DNM2
DOT1L	DTX1	DUSP2	DUSP9	EBF1	ECT2L	EED	EGFR
EP300	EPHA3	EPHA5	EPHA7	EPHB1	ERBB2	ERBB3	ERBB4
ESR1	ETS1	ETV6	EXOSC6	EZH2	FAF1	FAM46C	FANCA
FANCD2	FANCE	FANCF	FANCG	FANCL	FAS (TNFRSF6)	FBXO11	FBXO31
FGF10	FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1
FGFR3	FGFR4	FHIT	FLCN	FLT1	FLT3	FLT4	FLYWCH1
FOXO1	FOXO3	FOXP1	FRS2	GADD45B	GATA1	GATA2	GATA3
GNA11	GNA12	GNA13	GNAQ	GNAS	GPR124	GRIN2A	GSK3B
HDAC1	HDAC4	HDAC7	HGF	HIST1H1C	HIST1H1D	HIST1H1E	HIST1H2AC
HIST1H2AL	HIST1H2AM	HIST1H2BC	HIST1H2BJ	HIST1H2BK	HIST1H2BO	HIST1H3B	HNF1A
HSP90AA1	ICK	ID3	IDH1	IDH2	IGF1R	IKBKE	IKZF1
IKZF3	IL7R	INHBA	INPP4B	INPP5D (SHIP)	IRF1	IRF4	IRF8
JAK1	JAK2	JAK3	JARID2	JUN	KAT6A (MYST3)	KDM2B	KDM4C
KDM5C	KDM6A	KDR	KEAP1	KIT	KLHL6	KMT2A (MLL)	KMT2C (MLL3)
KRAS	LEF1	LRP1B	LRRK2	MAF	MAFB	MAGED1	MALT1
MAP2K2	MAP2K4	MAP3K1	MAP3K14	MAP3K6	MAP3K7	MAPK1	MCL1
MDM4	MED12	MEF2B	MEF2C	MEN1	MET	MIB1	MITF
MLH1	MPL	MRE11A	MSH2	MSH3	MSH6	MTOR	MUTYH
MYCL (MYCL1)	MYCN	MYD88	MYO18A	NCOR2	NCSTN	NF1	NF2
NFKBIA	NKX2-1	NOD1	NOTCH1	NOTCH2	NPM1	NRAS	NTSC2
NTRK2	NTRK3	NUP93	NUP98	P2RY8	PAG1	PAK3	PALB2
PAX5	PBRM1	PC	PCBP1	PCLO	PDCD1	PDCD11	PDCD1LG2 (PD-L2)
PDGFRB	PDK1	PHF6	PIK3CA	PIK3CG	PIK3R1	PIK3R2	PIM1
POT1	PPP2R1A	PRDM1	PRKAR1A	PRKDC	PRSS8	PTCH1	PTEN
PTPN2	PTPN6 (SHP-1)	PTPRO	RAD21	RAD50	RAD51	RAF1	RARA
RB1	RELN	RET	RHOA	RICTOR	RNF43	ROS1	RPTOR
SIPR2	SDHA	SDHB	SDHC	SDHD	SERP2	SETBP1	SETD2
SGK1	SMAD2	SMAD4	SMARCA1	SMARCA4	SMARCB1	SMC1A	SMC3
SOC3	SOC3	SOC3	SOX10	SOX2	SPEN	SPOP	SRC
STAG2	STAT3	STAT4	STAT5A	STAT5B	STAT6	STK11	SUFU
TAF1	TBL1XR1	TCF3 (E2A)	TCL1A (TCL1)	TET2	TGFB2	TLL2	TMEM30A
TMSB4XP8 (TMSL3)	TNFAIP3	TNFAIP3	TNFRSF11A	TNFRSF14	TNFRSF17	TOP1	TP53
TRAF2	TRAF3	TRAF5	TSC1	TSC2	TSHR	TUSC3	TYK2
U2AF2	VHL	WDR90	WHSC1 (MMSET or NSD2)		WISP3	WT1	XBP1
YY1AP1	ZMYM3	ZNF217	ZNF24 (ZSCAN3)	ZNF703	ZRSR2		XPO1

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Lena Stuart, M.D. | 09 March 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1308441-01

APPENDIX

Genes Assayed in FoundationOne®Heme

*Note: the assay was updated on 11/8/2016 to include the detection of alterations in CALR

HEMATOLOGICAL MALIGNANCY DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

ALK	BCL2	BCL6	BCR	BRAF	CCND1	CRLF2	EGFR	EPOR
ETV1	ETV4	ETV5	ETV6	EWSR1	FGFR2	IGH	IGK	IGL
JAK1	JAK2	KMT2A (MLL)	MYC	NTRK1	PDGFRA	PDGFRB	RAF1	RARA
RET	ROS1	TMPRSS2	TRG					

HEMATOLOGICAL MALIGNANCY RNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS*

ABL1	ABL1	ABL2	ACSL6	AFF1	AFF4	ALK	ARHGAP26 (GRAF)	
ARHGEF12	ARID1A	ARNT	ASXL1	ATF1	ATG5	AT1C	BCL10	BCL11A
BCL11B	BCL2	BCL3	BCL6	BCL7A	BCL9	BCOR	BCR	BIRC3
BRAF	BTG1	CAMTA1	CARS	CBFA2T3	CBFB	CBL	CND1	CCND2
CCND3	CD274 (PD-L1)	CDK6	CDX2	CHIC2	CHN1	CIC	CIITA	CLP1
CLTC	CLTCL1	CNTRL (CEP110)	COL1A1	CREB3L1	CREB3L2	CREBBP	CRLF2	CSF1
CTNNB1	DDIT3	DDX10	DDX6	DEK	DUSP22	EGFR	EIF4A2	ELF4
ELL	ELN	EML4	EP300	EPOR	EPS15	ERBB2	ERG	ETS1
ETV1	ETV4	ETV5	ETV6	EWSR1	FCGR2B	FCRL4	FEV	FGFR1
FGFR1OP	FGFR2	FGFR3	FLI1	FNBP1	FOXO1	FOXO3	FOXO4	FOXP1
FSTL3	FUS	GAS7	GLI1	GMPS	GPHN	HERPUD1	HEY1	HIP1
HIST1H4I	HLF	HMGA1	HMGA2	HOXA11	HOXA13	HOXA3	HOXA9	HOXC11
HOXC13	HOXD11	HOXD13	HSP90AA1	HSP90AB1	IGH	IGK	IGL	IKZF1
IL21R	IL3	IRF4	ITK	JAK1	JAK2	JAK3	JAZF1	KAT6A (MYST3)
KDSR	KIF5B	KMT2A (MLL)	LASP1	LCP1	LMO1	LMO2	LPP	LYL1
MAF	MAFB	MALT1	MDS2	MECOM	MKL1	MLF1	MLLT1 (ENL)	MLLT10 (AF10)
MLLT3	MLLT4 (AF6)	MLLT6	MN1	MXN1	MSI2	MSN	MUC1	MYB
MYC	MYH11	MYH9	NACA	NBEAP1 (BCL8)	NCOA2	NDRG1	NF1	NF2
NFKB2	NIN	NOTCH1	NPM1	NR4A3	NSD1	NTRK1	NTRK2	NTRK3
NUMA1	NUP214	NUP98	NUTM2A	OMD	P2RY8	PAFAH1B2	PAX3	PAX5
PAX7	PBX1	PCM1	PCSK7	PDCD1LG2 (PD-L2)	PDE4DIP	PDGFB	PDGFRA	PDGFRB
PER1	PHF1	PICALM	PIM1	PLAG1	PML	POU2AF1	PPP1CB	PRDM1
PRDM16	PRRX1	PSIP1	PTCH1	PTK7	RABEP1	RAF1	RALGDS	RAP1GDS1
RARA	RBM15	RET	RHOH	RNF213	ROS1	RPL22	RPN1	RUNX1
RUNX1T1 (ETO)	RUNX2	SEC31A	SEPT5	SEPT6	SEPT9	SET	SH3GL1	SLC1A2
SNX29 (RUNDC2A)	SRSF3	SS18	SSX1	SSX2	SSX4	STAT6	STL	SYK
TAF15	TAL1	TAL2	TBL1XR1	TCF3 (E2A)	TCL1A (TCL1)	TEC	TET1	TFE3
TFG	TFPT	TFRC	TLX1	TLX3	TMPRSS2	TNFRSF11A	TOP1	TP63
TPM3	TPM4	TRIM24	TRIP11	TTL	TYK2	USP6	WHSC1 (MMSET or NSD2)	
WHSC1L1	YPEL5	ZBTB16	ZMYM2	ZNF384	ZNF521			

*Note: some VUS rearrangements between targeted genes and unknown fusion partners or intergenic regions detected by RNA sequencing may not be reported.

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Microsatellite (MS) status

Tumor Mutational Burden (TMB)

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

 Electronically signed by Lena Stuart, M.D. | 09 March 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

 Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1308441-01

APPENDIX
Performance Specifications

The median exon coverage for this sample is 801x

ACCURACY

Sensitivity: Base Substitutions	At $\geq 5\%$ Minor Allele Frequency	>99.0%
Sensitivity: Insertions/Deletions (1-40bp)	At $\geq 10\%$ Minor Allele Frequency	98.0%
Sensitivity: Focal Copy Number Alterations (Homozygous Deletions or Amplifications)	At ≥ 8 copies	>95.0%
Sensitivity: Microsatellite Instability-High (MSI-H) status	Positive Predictive Agreement (PPA)	100.0% (87.54%-100.00%)*
Sensitivity: Microsatellite Stable (MSS) status	Positive Predictive Agreement (PPA)	89.66% (81.50%, 94.46%)*
Sensitivity: Known Gene Fusions	>95.0%	
Specificity: Base Substitutions, Insertions/Deletions, and Focal Copy Number Alterations	Positive Predictive Value (PPV)	>99.0%
Specificity: Known Gene Fusions	Positive Predictive Value (PPV)	>95.0%
Specificity: Microsatellite Instability-High (MSI-H) status	Negative Predictive Agreement (NPA)	97.44% (91.12%-99.29%)*
Specificity: Microsatellite Stable (MSS) status	Negative Predictive Agreement (NPA)	94.44% (86.57%, 97.82%)*
Accuracy: Tumor Mutation Burden	At $\geq 20\%$ tumor nuclei	>90.0%
Reproducibility (average concordance between replicates)	97.0% inter-batch precision 97.0% intra-batch precision 95.0% microsatellite status precision 96.0% tumor mutation burden precision	

*95% Confidence Interval

Assay specifications were determined for typical median exon coverage of approximately 500X. For additional information regarding the validation of FoundationOne®Heme, please refer to the article He, J. et al. Integrated genomic DNA/RNA profiling of hematologic malignancies in the clinical setting, Blood (2016 Jun. 16).

In the fractional-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the FoundationOne Heme assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by analytical concordance to a PCR comparator

assay using a pan-tumor sample set. Patients with results categorized as "MS-Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.

Tumor Mutational Burden (TMB) is determined by measuring the number of somatic mutations in sequenced genes on the FoundationOne Heme test and extrapolating to the genome as a whole. TMB is assayed for all FoundationOne Heme samples and is reported as the number of mutations per megabase (Muts/Mb). Tumor Mutational Burden is reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine Tumor Mutational Burden.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Lena Stuart, M.D. | 09 March 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 • CLIA: 34D2044309
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1308441-01

APPENDIX

About FoundationOne®Heme

ABOUT FOUNDATIONONE HEME

FoundationOne®Heme is a comprehensive genomic profiling test for hematologic malignancies and sarcomas. The test is designed to provide physicians with clinically actionable information to help with diagnostic sub-classification, prognosis assessment, and targeted therapeutic selection. Test results provide information about clinically significant alterations, potential targeted therapies, available clinical trials, and quantitative markers that may support immunotherapy clinical trial enrollment. FoundationOne Heme is analytically validated to detect all classes of genomic alterations in more than 400 cancer-related genes. In addition to DNA sequencing, FoundationOne Heme employs RNA sequencing across 265 genes to capture a broad range of gene fusions, common drivers of hematologic malignancies and sarcomas.

FoundationOne Heme was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne Heme has not been cleared or approved by the United States Food and Drug Administration (FDA). The FDA has determined that such clearance or approval is not necessary. FoundationOne Heme may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratory is certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA) as qualified to perform high-complexity clinical testing.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne Heme identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

Electronically signed by Lena Stuart, M.D. | 09 March 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

(Equivocal and Subclonal)

An alteration denoted as "amplification – equivocal" implies that FoundationOne Heme data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne Heme for identifying a copy number amplification is five (5) for *ERBB2* and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that FoundationOne Heme data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that FoundationOne Heme analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials

Ranking of Therapies in Summary Table

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source

or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne Heme.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. These include: subclonal alterations in heterogeneous samples, low sample quality or with homozygous losses of <3 exons; and deletions and insertions >40bp, or in repetitive/high homology sequences. FoundationOne Heme is performed using DNA and RNA derived from tumor, and as such germline events may not be reported.

The following targets typically have low coverage resulting in a reduction in sensitivity: SDHD exon 4, TNFRSF11A exon1, and TP53 exon 1.

FoundationOne Heme fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Ciplastraat 3, 2440 Geel, Belgium.

© 2022 Foundation Medicine, Inc. All rights reserved.

Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1308441-01

APPENDIX

About FoundationOne®Heme

CE

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*),

MPL, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
muts/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

MR Suite Version 6.0.0

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Lena Stuart, M.D. | 09 March 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # **ORD-1308441-01**
APPENDIX
References

1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) PMID: 25392179
2. Kroemer G, et al. Oncoimmunology (2015) PMID: 26140250
3. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
4. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
5. Ayers et al., 2016; ASCO-SITC Abstract P60
6. Monument MJ, et al. ISRN Oncol (2012) PMID: 23401795
7. Bonneville R, et al. JCO Precis Oncol (2017) PMID: 29850653
8. Wooster R, et al. Nat. Genet. (1994) PMID: 8162069
9. Kawaguchi K, et al. Oncol. Rep. (2005) PMID: 15643505
10. Saito T, et al. Hum. Pathol. (2003) PMID: 14562278
11. Suwa K, et al. J Orthop Sci (1999) PMID: 10370164
12. Garcia JJ, et al. Mod. Pathol. (2006) PMID: 16619000
13. Aue G, et al. Cancer Genet. Cytogenet. (1998) PMID: 9689926
14. Rucińska M, et al. Med. Sci. Monit. (2005) PMID: 15668629
15. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
16. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
17. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
18. Boland CR, et al. Cancer Res. (1998) PMID: 9823339
19. Pawlik TM, et al. Dis. Markers (2004) PMID: 15528785
20. Boland CR, et al. Gastroenterology (2010) PMID: 20420947
21. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
22. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
23. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
24. Cristescu R, et al. Science (2018) PMID: 30309915
25. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
26. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
27. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
28. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
29. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
30. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
31. Marabelle A, et al. Lancet Oncol. (2020) PMID: 32919526
32. Legrand et al., 2018; ASCO Abstract 12000
33. Chalmers ZR, et al. Genome Med (2017) PMID: 28420421
34. Lim J, et al. Clin. Cancer Res. (2015) PMID: 26330427
35. Brohl AS, et al. PLoS Genet. (2014) PMID: 25010205
36. Chen X, et al. Cancer Cell (2013) PMID: 24332040
37. Steele CD, et al. Cancer Cell (2019) PMID: 30889380
38. Casey DL, et al. Clin Cancer Res (2020) PMID: 31699828
39. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
40. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
41. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
42. Rizvi AS, et al. Science (2015) PMID: 25765070
43. Johnson BE, et al. Science (2014) PMID: 24336570
44. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
45. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
46. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
47. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
48. Nature (2012) PMID: 22810696
49. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
50. Tee AR, et al. Curr. Biol. (2003) PMID: 12906785
51. Kwiatkowski DJ, et al. Eur J Hum Genet (2015) PMID: 25782670
52. Luo C, et al. Orphanet J Rare Dis (2021) PMID: 34217357
53. Wang T, et al. Cancer Biol Ther (2020) PMID: 31597506
54. Guo G, et al. Front Oncol (2020) PMID: 33575217
55. Espinosa M, et al. BMC Cancer (2018) PMID: 29764404
56. Chuang CK, et al. Int Urol Nephrol (2017) PMID: 28547571
57. Wagner AJ, et al. J. Clin. Oncol. (2010) PMID: 20048174
58. Dickson MA, et al. Int. J. Cancer (2013) PMID: 22927055
59. Wagner AJ, et al. J Clin Oncol (2021) PMID: 34637337
60. Dickson et al., 2021; ASCO Abstract 3111
61. Adib E, et al. Clin Cancer Res (2021) PMID: 33727259
62. Nassar AH, et al. Mol Cancer Ther (2020) PMID: 31653662
63. De S, et al. Gegenbaurs Morphol Jahrb (1986) PMID: 3032730
64. Wagle N, et al. N. Engl. J. Med. (2014) PMID: 25295501
65. Tannir NM, et al. Eur. Urol. (2016) PMID: 26626617
66. Maroto P, et al. J Natl Compr Canc Netw (2018) PMID: 29632054
67. Zureick AH, et al. BMJ Case Rep (2019) PMID: 31154346
68. Hu W, et al. Front Oncol (2020) PMID: 33344249
69. Perini GF, et al. Blood Cancer J (2016) PMID: 27176796
70. Voss MH, et al. Clin. Cancer Res. (2018) PMID: 30327302
71. Barretina J, et al. Nat. Genet. (2010) PMID: 20601955
72. Lee-Jones L, et al. Genes Chromosomes Cancer (2004) PMID: 15236319
73. Hayashi T, et al. Hum. Pathol. (2012) PMID: 22748302
74. Lee RS, et al. J. Clin. Invest. (2012) PMID: 22797305
75. Ando K, et al. Cancers (Basel) (2013) PMID: 24216993
76. Zenali MJ, et al. Ann. Clin. Lab. Sci. (2009) PMID: 19429803
77. Zhang YX, et al. Clin. Cancer Res. (2013) PMID: 23714727
78. Brewer Savannah KJ, et al. Clin. Cancer Res. (2012) PMID: 22821997
79. Curr Oncol Rep (2013) PMID: 23605780
80. Inoki K, et al. Genes Dev. (2003) PMID: 12869586
81. Hodges AK, et al. Hum. Mol. Genet. (2001) PMID: 11741833
82. Int. J. Cancer (2006) PMID: 16206276
83. Li Y, et al. Mol. Cell. Biol. (2004) PMID: 15340059
84. Ann. N. Y. Acad. Sci. (1991) PMID: 2039135
85. Kandt RS, et al. Nat. Genet. (1992) PMID: 1303246
86. Cell (1993) PMID: 8269512
87. Curatolo P, et al. Lancet (2008) PMID: 18722871
88. Tate JG, et al. Nucleic Acids Res. (2019) PMID: 30371878
89. Kim MS, et al. Histopathology (2011) PMID: 21447119
90. Bandrés E, et al. Oncol. Rep. (2007) PMID: 17390049
91. Feys T, et al. Haematologica (2007) PMID: 17606441
92. Nagarajan P, et al. Oncogene (2009) PMID: 19137022
93. Capelli LP, et al. Eur J Med Genet (2012) PMID: 22178256
94. Suls A, et al. Am. J. Hum. Genet. (2013) PMID: 24207121
95. Lund C, et al. Epilepsy Behav (2014) PMID: 24614520
96. Scolz M, et al. PLoS ONE (2012) PMID: 23236459
97. Pérez-Peña J, et al. Oncotarget (2017) PMID: 28423514
98. Liu A, et al. Int. J. Biol. Macromol. (2019) PMID: 30414902
99. Wu X, et al. Sci Rep (2017) PMID: 28698581
100. Guo L, et al. Cell Biol. Toxicol. (2016) PMID: 27240802
101. Tian T, et al. Asian Pac. J. Cancer Prev. (2011) PMID: 22292647
102. Owonikoko et al., 2016; ESMO Abstract 14230
103. Hook KE, et al. Mol. Cancer Ther. (2012) PMID: 22222631
104. Gong X, et al. Cancer Discov (2019) PMID: 30373917
105. Oser MG, et al. Cancer Discov (2019) PMID: 30373918
106. Beltran H, et al. Clin. Cancer Res. (2019) PMID: 30232224
107. Allaman-Pillet N, et al. Ophthalmic Genet. (2019) PMID: 21955141
108. Viatour P, et al. J. Exp. Med. (2011) PMID: 21875955
109. Condorelli R, et al. Ann. Oncol. (2018) PMID: 29236940
110. Fry DW, et al. Mol. Cancer Ther. (2004) PMID: 15542782
111. Dean JL, et al. Oncogene (2010) PMID: 20473330
112. Dean JL, et al. Cell Cycle (2012) PMID: 22767154
113. Garnett MJ, et al. Nature (2012) PMID: 22460902
114. Roberts PJ, et al. J. Natl. Cancer Inst. (2012) PMID: 22302033
115. Patnaik A, et al. Cancer Discov (2016) PMID: 27217383
116. O'Leary B, et al. Cancer Discov (2018) PMID: 30206110
117. Costa C, et al. Cancer Discov (2019) PMID: 31594766
118. Chen SH, et al. Oncogene (2018) PMID: 29059158
119. Derenzini M, et al. Clin. Cancer Res. (2008) PMID: 18381962
120. Knudsen ES, et al. Nat. Rev. Cancer (2008) PMID: 19143056
121. Chibon F, et al. Cancer Res. (2000) PMID: 11103795
122. Shim BY, et al. Cancer Res Treat (2010) PMID: 20948919
123. Burkhart DL, et al. Nat. Rev. Cancer (2008) PMID: 18650841
124. Berge EO, et al. Mol. Cancer (2010) PMID: 20594292
125. Giacinti C, et al. Oncogene (2006) PMID: 16936740
126. Otterson GA, et al. Proc. Natl. Acad. Sci. U.S.A. (1997) PMID: 9342358
127. Otterson GA, et al. Am. J. Hum. Genet. (1999) PMID: 10486322
128. Qin XQ, et al. Genes Dev. (1992) PMID: 1534305
129. Rubin SM, et al. Cell (2005) PMID: 16360038
130. Sun H, et al. Mol. Cell. Biol. (2006) PMID: 16449662
131. Chen Z, et al. Hum. Mutat. (2014) PMID: 24282159
132. Yun J, et al. Int J Ophthalmol (2011) PMID: 22553621
133. Houston SK, et al. Int Ophthalmol Clin (2011) PMID: 21139478
134. Ng AK, et al. Semin Radiat Oncol (2010) PMID: 19959033
135. Hirai H, et al. Cancer Biol. Ther. (2010) PMID: 20107315
136. Bridges KA, et al. Clin. Cancer Res. (2011) PMID: 21799033
137. Rajeshkumar NV, et al. Clin. Cancer Res. (2011) PMID: 21389100
138. Osman AA, et al. Mol. Cancer Ther. (2015) PMID: 25504633
139. Xu L, et al. Mol. Cancer Ther. (2002) PMID: 12489850
140. Xu L, et al. Mol. Med. (2001) PMID: 11713371
141. Camp ER, et al. Cancer Gene Ther. (2013) PMID: 23470564
142. Kim SS, et al. Nanomedicine (2015) PMID: 25240597
143. Pirollo KF, et al. Mol. Ther. (2016) PMID: 27357628
144. Hajdenberg et al., 2012; ASCO Abstract e15010
145. Leijen S, et al. J. Clin. Oncol. (2016) PMID: 27601554
146. Moore et al., 2019; ASCO Abstract 5513
147. Leijen S, et al. J. Clin. Oncol. (2016) PMID: 27998224
148. Oza et al., 2015; ASCO Abstract 5506
149. Lee J, et al. Cancer Discov (2019) PMID: 31315834
150. Méndez E, et al. Clin. Cancer Res. (2018) PMID: 29535125
151. Seligmann JF, et al. J Clin Oncol (2021) PMID: 34538072
152. Kwok M, et al. Blood (2016) PMID: 26563132
153. Boudny M, et al. Haematologica (2019) PMID: 30975914
154. Dillon MT, et al. Mol. Cancer Ther. (2017) PMID: 28062704
155. Middleton FK, et al. Cancers (Basel) (2018) PMID: 30127241

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Lena Stuart, M.D. | 09 March 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 | CLIA: 34D2044309
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 | CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 | CLIA: 22D2027531

ORDERED TEST # **ORD-1308441-01**
APPENDIX
References

156. Cerami E, et al. Cancer Discov (2012) PMID: 22588877
157. Gao J, et al. Sci Signal (2013) PMID: 23550210
158. Pérot G, et al. Am. J. Pathol. (2010) PMID: 20884963
159. Taubert H, et al. Cancer Res. (1996) PMID: 8797580
160. Brown CJ, et al. Nat. Rev. Cancer (2009) PMID: 19935675
161. Joerger AC, et al. Annu. Rev. Biochem. (2008) PMID: 18410249
162. Kato S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) PMID: 12826609
163. Kamada R, et al. J. Biol. Chem. (2011) PMID: 20978130
164. Zerdoumi Y, et al. Hum. Mol. Genet. (2017) PMID: 28472496
165. Yamada H, et al. Carcinogenesis (2007) PMID: 17690113
166. Bougeard G, et al. J. Clin. Oncol. (2015) PMID: 26014290
167. Sorrell AD, et al. Mol Diagn Ther (2013) PMID: 23355100
168. Nichols KE, et al. Cancer Epidemiol. Biomarkers Prev. (2001) PMID: 11219776
169. Kleihues P, et al. Am. J. Pathol. (1997) PMID: 9006316
170. Gonzalez KD, et al. J. Clin. Oncol. (2009) PMID: 19204208
171. Lalloo F, et al. Lancet (2003) PMID: 12672316
172. Mandelker D, et al. Ann. Oncol. (2019) PMID: 31050713
173. Jaiswal S, et al. N. Engl. J. Med. (2014) PMID: 25426837
174. Genovese G, et al. N. Engl. J. Med. (2014) PMID: 25426838
175. Xie M, et al. Nat. Med. (2014) PMID: 25326804
176. Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) PMID: 28669404
177. Severson EA, et al. Blood (2018) PMID: 29678827
178. Fuster JJ, et al. Circ. Res. (2018) PMID: 29420212
179. Hematology Am Soc Hematol Educ Program (2018) PMID: 30504320
180. Chabon JJ, et al. Nature (2020) PMID: 32269342
181. Razavi P, et al. Nat. Med. (2019) PMID: 31768066
182. Dickson et al., 2021; CTOS Abstract 1818755

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Lena Stuart, M.D. | 09 March 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor. Cambridge, MA 02141 · CLIA: 22D2027531