

**ABOUT THE TEST** FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

<b>PATIENT</b>	<b>DISEASE</b> Unknown primary squamous cell carcinoma (SCC)	<b>PHYSICIAN</b>	<b>ORDERING PHYSICIAN</b> Yeh, Yi-Chen	<b>SPECIMEN</b>	<b>SPECIMEN SITE</b> Lymph Node
	<b>NAME</b> Chou, Chia-Yun		<b>MEDICAL FACILITY</b> Taipei Veterans General Hospital		<b>SPECIMEN ID</b> S111-03291A (PF22098)
	<b>DATE OF BIRTH</b> 26 December 1960		<b>ADDITIONAL RECIPIENT</b> None		<b>SPECIMEN TYPE</b> Slide Deck
	<b>SEX</b> Female		<b>MEDICAL FACILITY ID</b> 205872		<b>DATE OF COLLECTION</b> 20 January 2022
	<b>MEDICAL RECORD #</b> 46530170		<b>PATHOLOGIST</b> Not Provided		<b>SPECIMEN RECEIVED</b> 31 August 2022

## Biomarker Findings

**Microsatellite status** - Cannot Be Determined <sup>a</sup>  
**Tumor Mutational Burden** - 8 Muts/Mb

## Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

**PIK3CA** E81K

**PTEN** R130\*, K223\*

**TET2** E1151\*

<sup>a</sup> Patients with Microsatellite status of Cannot Be Determined should be re-tested with an orthogonal (alternative) method.

## Report Highlights

- Targeted therapies with potential clinical benefit **approved in another tumor type**: Everolimus (p. [7](#)), Temsirolimus (p. [7](#))
- Evidence-matched **clinical trial options** based on this patient's genomic findings: (p. [8](#))
- Variants that may represent **clonal hematopoiesis** and may originate from non-tumor sources: **TET2 E1151\*** (p. [6](#))

## PATHOLOGIST COMMENTS

Erik Williams, M.D. 9-Sep-2022

This assay is not validated to identify HPV viral reads. However, manual review of the sequencing data reveals high-level read support for HPV-16 infection, consistent with the reported positivity for p16 by immunohistochemistry. Ancillary orthogonal HPV testing could be performed to confirm this finding, if clinically indicated. In the appropriate clinicopathologic context, this supports a diagnosis of an HPV-related squamous cell carcinoma.

### BIOMARKER FINDINGS

**Microsatellite status** -  
 Cannot Be Determined

**Tumor Mutational Burden** - 8 Muts/Mb

### GENOMIC FINDINGS

**PIK3CA** - E81K

10 Trials [see p. 8](#)

**PTEN** - R130\*, K223\*

10 Trials [see p. 10](#)

### THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. See Biomarker Findings section

No therapies or clinical trials. See Biomarker Findings section

#### THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)

none

none

#### THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)

Everolimus

Temsirolimus

none

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 09 September 2022  
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

**VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS (CH)**

Genomic findings below may include nontumor somatic alterations, such as CH. The efficacy of targeting such nontumor somatic alterations is unknown. This content should be interpreted based on clinical context. Refer to appendix for additional information on CH.

**TET2 - E1151\*** ..... [p. 6](#)

**GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS**

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

**TET2 - E1151\*** ..... [p. 6](#)

**NOTE** Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 09 September 2022  
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531  
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531  
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1446725-01

## BIOMARKER FINDINGS

## BIOMARKER

# Microsatellite status

## RESULT

Cannot Be Determined

## POTENTIAL TREATMENT STRATEGIES

### — Targeted Therapies —

On the basis of prospective clinical evidence in multiple solid tumor types, microsatellite instability (MSI) and associated increased tumor mutational burden (TMB)<sup>1-2</sup> may predict sensitivity to immune checkpoint inhibitors, including the approved PD-1-targeting agents cemiplimab, dostarlimab, nivolumab (alone or in combination with ipilimumab), and pembrolizumab<sup>3-8</sup> and PD-L1-targeting agents atezolizumab, avelumab, and durvalumab<sup>9-11</sup>. As the MSI status of this tumor is

unknown, the relevance of these therapeutic approaches is unclear.

## FREQUENCY & PROGNOSIS

MSI-high (MSI-H) has been observed at high frequency in endometrial cancers (14-33%)<sup>12-19</sup>, colorectal cancers (CRCs; 10-15%)<sup>2,20-23</sup>, and gastric cancers (12-35%)<sup>24-27</sup> and at lower frequencies in many other tumor types, including esophageal<sup>28</sup>, small bowel<sup>29-33</sup>, hepatobiliary<sup>34-40</sup>, prostate<sup>41-43</sup>, and urinary tract carcinomas<sup>44-46</sup>. In one study, MSI-H status was associated with a positive prognostic effect in patients with gastric cancer treated with surgery alone and a negative predictive effect in patients treated with chemotherapy<sup>47</sup>. Data regarding the role of MSI-H on prognosis and survival in endometrial cancer are conflicting<sup>12,15-16,18,48-50</sup>. However, studies specifically analyzing early stage endometrial cancer have reported a correlation between MSI-H

and decreased survival<sup>14,17,19,49</sup>, thereby suggesting that MSI-H predicts for poor prognosis in this subset of endometrial tumors.

## FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor<sup>22</sup>. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2<sup>22,51-52</sup>. The level of MSI in this sample could not be determined with confidence. Depending on the clinical context, MSI testing of an alternate sample or by another methodology could be considered.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 09 September 2022  
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1446725-01

## BIOMARKER FINDINGS

## BIOMARKER

# Tumor Mutational Burden

## RESULT

8 Muts/Mb

## POTENTIAL TREATMENT STRATEGIES

### — Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1<sup>53-55</sup>, anti-PD-1 therapies<sup>53-56</sup>, and combination nivolumab and ipilimumab<sup>57-62</sup>. In multiple pan-tumor studies, increased tissue tumor mutational burden (TMB) was associated with sensitivity to immune checkpoint inhibitors<sup>53-56,63-67</sup>. In the KEYNOTE 158 trial of pembrolizumab monotherapy for patients with solid tumors, significant improvement in ORR was observed for patients with TMB  $\geq 10$  Muts/Mb (as measured by this assay) compared with those with TMB  $< 10$  Muts/Mb in a large cohort that included multiple tumor types<sup>63</sup>; similar findings were observed in the KEYNOTE 028 and 012 trials<sup>56</sup>. At the same TMB cutpoint, retrospective analysis of patients with solid tumors treated with any checkpoint inhibitor identified that tissue TMB scores  $\geq 10$  Muts/Mb were associated with prolonged time to treatment failure compared with scores  $< 10$  muts/Mb (HR=0.68)<sup>67</sup>. For patients with solid tumors treated with nivolumab plus ipilimumab in the CheckMate 848 trial, improved

responses were observed in patients with a tissue TMB  $\geq 10$  Muts/Mb independent of blood TMB at any cutpoint in matched samples<sup>68</sup>. However, support for higher TMB thresholds and efficacy was observed in the prospective Phase 2 MyPathway trial of atezolizumab for patients with pan-solid tumors, where improved ORR and DCR was seen in patients with TMB  $\geq 16$  Muts/Mb than those with TMB  $\geq 10$  and  $< 16$  Muts/Mb<sup>66</sup>. Similarly, analyses across several solid tumor types reported that patients with higher TMB (defined as  $\geq 16$ -20 Muts/Mb) achieved greater clinical benefit from PD-1 or PD-L1-targeting monotherapy compared with patients with higher TMB treated with chemotherapy<sup>69</sup> or those with lower TMB treated with PD-1 or PD-L1-targeting agents<sup>54</sup>.

## FREQUENCY & PROGNOSIS

Squamous cell carcinoma (SCC) of unknown primary harbors a median tumor mutational burden (TMB) of 7.6 Muts/Mb, and 22% of cases have high TMB ( $> 20$  Muts/Mb)<sup>70</sup>. High TMB has been reported frequently in skin SCC (67% of cases)<sup>70-71</sup>; in 10% of lung SCC<sup>71</sup>; 8-13% of head and neck SCC cases and non-small cell lung carcinoma, including lung SCC cases<sup>70</sup>; and in additional SCC cases, including urothelial (12%), cervical (6.5%), anal (3.8%), and esophageal (2.1%)<sup>71</sup>. For patients with squamous cell carcinoma (SCC) treated with PD-L1/PD-1 inhibitors, a Kaplan-Meier analysis showed a significant association for patients with high tumor mutational burden (TMB) with longer time to treatment failure (9.9 vs. 4.4 months)<sup>71</sup>. In the majority of cutaneous SCC cases, high mutational burden has been attributed to UV exposure rather than defective DNA mismatch

repair or polymerase activity<sup>72-73</sup>, although one study reported a small number of cutaneous SCC cases (4/39) harboring a mutation signature similar to that of human papillomavirus-positive head and neck SCC<sup>73</sup>. In patients with non-small cell lung cancer (NSCLC), TMB is similar between cases with squamous and non-squamous histology<sup>74</sup>, and increased TMB is associated with higher tumor grade and poor prognosis<sup>75</sup>, as well as with a decreased frequency of known driver mutations in EGFR, ALK, ROS1, or MET (1% of high-TMB samples each) but not BRAF (10%) or KRAS (9.4%)<sup>74</sup>. Although some studies have reported a lack of association between smoking and increased TMB in NSCLC<sup>75-76</sup>, several other large studies did find a strong prognostic association<sup>77-80</sup>.

## FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma<sup>81-82</sup> and cigarette smoke in lung cancer<sup>78,83</sup>, treatment with temozolomide-based chemotherapy in glioma<sup>84-85</sup>, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes<sup>23,86-89</sup>, and microsatellite instability (MSI)<sup>23,86,89</sup>. This sample harbors a TMB level associated with lower rates of clinical benefit from treatment with PD-1- or PD-L1-targeting immune checkpoint inhibitors compared with patients with tumors harboring higher TMB levels, based on several studies in multiple solid tumor types<sup>54-55,63</sup>.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 09 September 2022  
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1446725-01

GENOMIC FINDINGS

GENE

**PIK3CA**

ALTERATION  
E81K

TRANSCRIPT ID  
NM\_006218

CODING SEQUENCE EFFECT  
241G>A

VARIANT ALLELE FREQUENCY (% VAF)  
26.3%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Clinical and preclinical data in various tumor types indicate that PIK3CA activating alterations may predict sensitivity to therapies targeting PI3K<sup>90-97</sup>, AKT<sup>98-99</sup>, or mTOR<sup>100-107</sup>. The Phase 2 NCI-MATCH study of copanlisib for patients with refractory solid tumors harboring PIK3CA mutations with or without PTEN loss met its primary endpoint with an ORR of 16% (4/25 PRs); responses (PR or SD >6 months) were seen in patients with ameloblastoma, liposarcoma, and carcinomas of the endometrium, ovary, esophagus, lung, and prostate<sup>97</sup>. However, the Phase 2 study of copanlisib for patients with endometrial carcinoma harboring PIK3CA hotspot

mutations failed to report any objective responses (n=11)<sup>96</sup>. Two other studies of copanlisib for patients with genomically unselected tumors reported 1 CR and 2 PRs (1 unconfirmed) among 16 total patients with PIK3CA-mutated solid tumors with or without PTEN alterations<sup>94-95</sup>. In the Phase 2 MATCH trial for patients with PIK3CA-mutated solid tumors, 28% (18/65) of patients experienced PFS lasting at least 6 months after treatment with taselisib; however, no ORs were observed in this study<sup>108</sup>. A separate Phase 1b study of taselisib in combination with the CDK4/6 inhibitor palbociclib for patients with PIK3CA-mutated solid tumors reported an ORR of 0% (n=12) and a DCR of 17% (2/12)<sup>109</sup>. In a Phase 1 trial of the dual PI3K/mTOR kinase inhibitor apitolisib, 79% (11/14) of patients with PIK3CA-mutated advanced solid tumors experienced disease control (3 PRs, 8 SDs)<sup>110</sup>. The PI3K inhibitor alpelisib is approved as a single agent for the treatment of patients with PIK3CA-related overgrowth spectrum (PROS)<sup>111</sup>, but has shown limited activity as monotherapy for PIK3CA-mutated solid tumors with a Phase 1a study reporting an ORR of 6.0% (8/134) and a DCR of 58% (78/134)<sup>112</sup>. It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

FREQUENCY & PROGNOSIS

PIK3CA mutations have been reported in 15% of head and neck squamous cell carcinomas (SCCs), and in 13% (2/15) of vulva SCC, 7.9% of lung SCC, and 7.9% of esophageal SCC cases<sup>113-116</sup>. One study reported that PIK3CA mutations were associated with poor prognosis in patients with lung adenocarcinoma<sup>117</sup>. Expression of PIK3CA mRNA or p110-alpha protein in patients with esophageal SCC has been correlated with lymph node metastasis and poor prognosis, while PIK3CA mutation has been associated with longer survival<sup>118-120</sup>. In one study, PI3K mutations were prevalent in advanced Stage 4 HNSCC tumors and associated with tumor progression<sup>121</sup>.

FINDING SUMMARY

PIK3CA encodes p110-alpha, which is the catalytic subunit of phosphatidylinositol 3-kinase (PI3K). The PI3K pathway is involved in cell signaling that regulates a number of critical cellular functions, including cell growth, proliferation, differentiation, motility, and survival<sup>122-123</sup>. PIK3CA mutations E81K and C378Y have been reported in cancer as well as in overgrowth syndromes characterized by activating mutations in the PI3K-AKT pathway, suggesting that these mutations may be activating, although this has not been directly demonstrated<sup>124-129</sup>.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 09 September 2022  
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1446725-01

GENOMIC FINDINGS

GENE

**PTEN**

ALTERATION

R130\*, K223\*

TRANSCRIPT ID

NM\_000314, NM\_000314

CODING SEQUENCE EFFECT

388C>T, 667A>T

VARIANT ALLELE FREQUENCY (% VAF)

13.7%, 15.8%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

PTEN loss or mutation leads to activation of the PI3K-AKT-mTOR pathway and may predict sensitivity to inhibitors of this pathway<sup>95,130-132</sup>. Across various tissues, most clinical studies have not observed an association between PTEN deficiency and response to inhibitors of the PI3K-AKT-mTOR pathway. However, limited studies in prostate cancer<sup>133-136</sup>, renal cell carcinoma<sup>137</sup>, breast cancer<sup>138-139</sup>, and colorectal cancer<sup>140</sup> have reported an association between PTEN deficiency and response to inhibitors targeting the PI3K-AKT-

mTOR pathway. Preclinical data indicate that PTEN loss or inactivation may predict sensitivity to PARP inhibitors<sup>141-145</sup>, and clinical benefit has been observed for patients with PTEN-altered breast cancer including triple negative breast cancer<sup>146</sup>, ovarian cancer<sup>147</sup>, uterine leiomyosarcoma<sup>148</sup>, and endometrial cancer<sup>145</sup> treated with PARP inhibitors. However, some studies have reported a lack of association between PTEN mutation and PARP inhibitor sensitivity<sup>149-150</sup>.

FREQUENCY & PROGNOSIS

PTEN mutations have been reported in 7% of lung squamous cell carcinomas (SCCs)<sup>114</sup>, 7% (2/29) of cutaneous SCCs cBio-Li et al., 2015; 25589618), 1-5% of esophageal SCCs<sup>115,151</sup>, and 0-5% of head and neck SCCs (HNSCC)<sup>113,152-153</sup>. In a study of oral SCC, loss of PTEN expression correlated with metastasis and higher stage tumors<sup>154</sup>.

FINDING SUMMARY

PTEN encodes an inositol phosphatase that functions as a tumor suppressor by negatively regulating the PI3K-AKT-mTOR pathway; loss of PTEN can lead to uncontrolled cell growth and suppression of apoptosis<sup>131</sup>. Alterations such as seen here may disrupt PTEN function or

expression<sup>155-196</sup>.

POTENTIAL GERMLINE IMPLICATIONS

One or more of the PTEN variants observed here has been described in the ClinVar database as a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with hamartoma tumor syndrome (ClinVar, Mar 2022)<sup>197</sup>. Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. PTEN mutations underlie several inherited disorders, collectively termed PTEN hamartoma tumor syndrome (PHTS), which include Cowden syndrome (CS) and its variant Lhermitte-Duclos disease (LD), Bannayan-Riley-Ruvalcaba syndrome (BRRS), PTEN-related Proteus syndrome (PS), and Proteus-like syndrome<sup>198-199</sup>. The mutation rate for PTEN in these disorders ranges from 20 to 85% of patients<sup>198,200</sup>. The estimated incidence of Cowden syndrome is 1/200,000, which may be an underestimate due to the high variability of this disorder<sup>198</sup>. Given the association between PTEN and these inherited syndromes, in the appropriate clinical context, germline testing for mutations affecting PTEN is recommended.

GENE

**TET2**

ALTERATION

E1151\*

TRANSCRIPT ID

NM\_017628

CODING SEQUENCE EFFECT

3451G>T

VARIANT ALLELE FREQUENCY (% VAF)

67.2%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no targeted therapies available to address genomic alterations in TET2 in solid tumors.

FREQUENCY & PROGNOSIS

TET2 alterations have been reported at relatively low frequencies in solid tumors and are more prevalent in hematological malignancies (cBioPortal, Jan 2022)<sup>201-202</sup>. Published data investigating the prognostic implications of TET2 alterations in solid tumors are limited (PubMed, Jan 2022).

FINDING SUMMARY

TET2 encodes a tumor suppressor involved in reversing DNA methylation marks, a process critical for proper gene regulation<sup>203-204</sup>. Alterations such as seen here may disrupt TET2 function or expression<sup>205-209</sup>.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to

occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion<sup>210-215</sup>. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy<sup>210-211</sup>. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease<sup>216</sup>. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH<sup>214,217-218</sup>. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 09 September 2022  
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531  
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531  
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531



**ORDERED TEST #** ORD-1446725-01

**THERAPIES WITH CLINICAL BENEFIT**
**IN OTHER TUMOR TYPE**

## Everolimus

*Assay findings association*
**PIK3CA**  
E81K

### AREAS OF THERAPEUTIC USE

Everolimus is an orally available mTOR inhibitor that is FDA approved to treat renal cell carcinoma (RCC) following antiangiogenic therapy; pancreatic neuroendocrine tumors; and well-differentiated non-functional neuroendocrine tumors of the lung or gastrointestinal tract. Everolimus is also approved to treat either renal angiomyolipoma or subependymal giant cell astrocytoma in association with tuberous sclerosis complex (TSC). Please see the drug label for full prescribing information.

### GENE ASSOCIATION

On the basis of clinical evidence<sup>100-107</sup>, PIK3CA activating mutations may predict sensitivity to mTOR inhibitors such as everolimus and temsirolimus. Studies have reported modest activity of these therapies as single agents (ORRs of 0-4%), but improved activity has been observed when they are combined with other agents such as bevacizumab and doxorubicin (ORRs of 25-44%), for patients with PIK3CA-mutated solid tumors<sup>104-107,219-223</sup>. It is not known whether this therapeutic approach would be relevant in the context of alterations that have not been

fully characterized, as seen here.

### SUPPORTING DATA

Everolimus has been clinically tested in patients with several squamous cell carcinomas (SCC). A Phase 2 study of everolimus therapy has reported no objective responses in any of nine patients with refractory head and neck squamous cell carcinoma<sup>224</sup>. A Phase 1 trial in patients with advanced solid tumors reported that everolimus in combination with low dose weekly cisplatin showed activity in several tumor types, with 3 partial responses and prolonged stable disease observed in 5 patients out of 28 evaluable patients; one patient with oropharyngeal squamous cell carcinoma obtained stable disease after more than 6 cycles of treatments<sup>225</sup>. Whereas frequent adverse events precluded a recommended Phase 2 dose and schedule for the combination of trametinib and everolimus in a Phase 1b trial for solid tumors<sup>226</sup>, a retrospective study for heavily pretreated patients with solid tumors reported tolerable regimens of the combination for 23/31 patients, with 16 patients treated >3 months and evaluable patients achieving a median PFS of 6.5 months<sup>227</sup>.

## Temsirolimus

*Assay findings association*
**PIK3CA**  
E81K

### AREAS OF THERAPEUTIC USE

Temsirolimus is an intravenous mTOR inhibitor that is FDA approved for the treatment of advanced renal cell carcinoma. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

On the basis of clinical evidence<sup>100-107</sup>, PIK3CA activating mutations may predict sensitivity to mTOR inhibitors such as everolimus and temsirolimus. Studies have reported modest activity of these therapies as single agents (ORRs of 0-4%), but improved activity has been observed when they are combined with other agents such as bevacizumab and doxorubicin (ORRs of 25-44%), for patients with PIK3CA-mutated solid tumors<sup>104-107,219-223</sup>. It is not known whether this therapeutic approach would be relevant in the context of alterations that have not been fully characterized, as seen here.

### SUPPORTING DATA

In the context of squamous cell carcinoma (SCC), temsirolimus has been tested primarily for head and neck squamous cell carcinoma (HNSCC), and temsirolimus in

combination with the VEGF antibody bevacizumab has shown significant efficacy<sup>228</sup>. A study assessing temsirolimus in combination with metformin in patients with advanced solid tumors reported a partial response for one patient with HNSCC, despite disease progression after treatment with docetaxel and cisplatin and subsequent treatment with zalutumumab<sup>229</sup>. A Phase 1 study of temsirolimus in combination with carboplatin and paclitaxel in 18 patients with HNSCC reported a partial response rate of 22% and recommended Phase 2 testing<sup>230</sup>. However, a Phase 2 study of temsirolimus and erlotinib in patients with recurrent and/or metastatic, platinum-refractory HNSCC reported that this combination therapy was poorly tolerated, with the trial ending early after 50% (6/12) of patients withdrew from the study<sup>231</sup>. A Phase 2 study of temsirolimus in patients with recurrent or metastatic cervical carcinoma reported a partial response in 3% (1/33) of patients and stable disease in 57.6% (19/33) of patients, with mild to moderate adverse effects and no toxicities greater than grade 3<sup>232</sup>. In a Phase 1 study of temsirolimus and bevacizumab, 2/4 of patients with cervical squamous cell carcinoma experienced a partial response<sup>233</sup>.

**NOTE** Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 09 September 2022  
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

**ORDERED TEST #** ORD-1446725-01

**CLINICAL TRIALS**

**NOTE** Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually

updated and should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that

may require medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://www.foundationmedicine.com/genomic-testing#support-services). Or, visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

**GENE**
**PIK3CA**
**ALTERATION**
**E81K**
**RATIONALE**

PIK3CA activating mutations may lead to activation of the PI3K-AKT-mTOR pathway and may therefore indicate sensitivity to inhibitors of this pathway. Strong clinical data support sensitivity of PIK3CA-mutated solid tumors to the

PI3K-alpha inhibitor apelsib. It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

**NCT04589845**
**PHASE 2**

Tumor-Agnostic Precision Immuno-Oncology and Somatic Targeting Rational for You (TAPISTRY) Platform Study

**TARGETS**

TRKB, ALK, TRKC, ROS1, TRKA, RET, PD-L1, AKTs, ERBB2, MDM2, PI3K-alpha

**LOCATIONS:** Taipei City (Taiwan), Taoyuan County (Taiwan), Tainan (Taiwan), Shanghai City (China), Shanghai (China), Shatin (Hong Kong), Hong Kong (Hong Kong), Seoul (Korea, Republic of), Xi'an (China), Tianjin (China)

**NCT04341259**
**PHASE 1**

A Study Of The Pharmacokinetics And Safety Of Ipatasertib In Chinese Participants With Locally Advanced Or Metastatic Solid Tumors.

**TARGETS**

AKTs

**LOCATIONS:** Shanghai City (China)

**NCT03239015**
**PHASE 2**

Efficacy and Safety of Targeted Precision Therapy in Refractory Tumor With Druggable Molecular Event

**TARGETS**

EGFR, ERBB4, ERBB2, PARP, mTOR, MET, ROS1, RET, VEGFRs, BRAF, CDK4, CDK6

**LOCATIONS:** Shanghai (China)

**NCT04337463**
**PHASE NULL**

ATG-008 Combined With Toripalimab in Advanced Solid Tumors

**TARGETS**

mTORC1, mTORC2, PD-1

**LOCATIONS:** Chongqing (China), Chengdu (China)

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 09 September 2022  
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531



ORDERED TEST # ORD-1446725-01

**CLINICAL TRIALS**
**NCT04803318**
**PHASE 2**

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

**TARGETS**  
 mTOR, FGFRs, RET, PDGFRA, VEGFRs,  
 KIT, MEK

**LOCATIONS:** Guangzhou (China)

**NCT04526470**
**PHASE 1/2**

Alpelisib and Paclitaxel in PIK3CA-altered Gastric Cancer

**TARGETS**  
 PI3K-alpha

**LOCATIONS:** Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of)

**NCT05125523**
**PHASE 1**

A Study of Sirolimus for Injection (Albumin Bound) in Patients With Advanced Solid Tumors

**TARGETS**  
 mTOR

**LOCATIONS:** Tianjin (China)

**NCT03772561**
**PHASE 1**

Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies

**TARGETS**  
 PARP, AKTs, PD-L1

**LOCATIONS:** Singapore (Singapore)

**NCT04801966**
**PHASE NULL**

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

**TARGETS**  
 CDK4, CDK6, PI3K-alpha, PD-L1, MEK,  
 PARP, PD-1, BRAF

**LOCATIONS:** Melbourne (Australia)

**NCT04632992**
**PHASE 2**

A Study Evaluating Targeted Therapies in Participants Who Have Advanced Solid Tumors With Genomic Alterations or Protein Expression Patterns Predictive of Response

**TARGETS**  
 TRKB, ALK, TRKC, ROS1, TRKA, PD-L1,  
 ERBB2, PI3K-alpha, RET, AKTs

**LOCATIONS:** Alaska, Washington, California, Idaho

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

 Electronically signed by Erik Williams, M.D. | 09 September 2022  
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

 Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1446725-01

**CLINICAL TRIALS**
**GENE**  
**PTEN**
**ALTERATION**  
 R130\*, K223\*

**RATIONALE**  
 PTEN loss or inactivating mutations may lead to increased activation of the PI3K-AKT-mTOR pathway and may indicate sensitivity to inhibitors

of this pathway. PTEN loss or inactivation may also predict sensitivity to PARP inhibitors.

**NCT04644068**
**PHASE 1/2**

Study of AZD5305 as Monotherapy and in Combination With Anti-cancer Agents in Patients With Advanced Solid Malignancies

**TARGETS**  
 ERBB2, TROP2, PARP

**LOCATIONS:** Shanghai (China), Seoul (Korea, Republic of), Chongqing (China), Chuo-ku (Japan), Koto-ku (Japan), Melbourne (Australia), Warszawa (Poland), Gdynia (Poland), Grzebnica (Poland), Budapest (Hungary)

**NCT04341259**
**PHASE 1**

A Study Of The Pharmacokinetics And Safety Of Ipatasertib In Chinese Participants With Locally Advanced Or Metastatic Solid Tumors.

**TARGETS**  
 AKTs

**LOCATIONS:** Shanghai City (China)

**NCT04337463**
**PHASE NULL**

ATG-008 Combined With Toripalimab in Advanced Solid Tumors

**TARGETS**  
 mTORC1, mTORC2, PD-1

**LOCATIONS:** Chongqing (China), Chengdu (China)

**NCT02264678**
**PHASE 1/2**

Ascending Doses of AZD6738 in Combination With Chemotherapy and/or Novel Anti Cancer Agents

**TARGETS**  
 ATR, PARP, PD-L1

**LOCATIONS:** Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Goyang-si (Korea, Republic of), Cambridge (United Kingdom), Withington (United Kingdom), Manchester (United Kingdom), London (United Kingdom), Coventry (United Kingdom), Sutton (United Kingdom), Oxford (United Kingdom)

**NCT04001569**
**PHASE 1/2**

AZD8186 and Paclitaxel in Advanced Gastric Cancer

**TARGETS**  
 PI3K-beta

**LOCATIONS:** Seongnam-si (Korea, Republic of)

**NCT05035745**
**PHASE 1/2**

Selinexor &amp; Talazoparib in Advanced Refractory Solid Tumors; Advanced/Metastatic Triple Negative Breast Cancer (START)

**TARGETS**  
 XPO1, PARP

**LOCATIONS:** Singapore (Singapore)

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

 Electronically signed by Erik Williams, M.D. | 09 September 2022  
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

 Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1446725-01

**CLINICAL TRIALS**
**NCT03772561**
**PHASE 1**

Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies

**TARGETS**  
 PARP, AKTs, PD-L1

**LOCATIONS:** Singapore (Singapore)

**NCT04801966**
**PHASE NULL**

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

**TARGETS**  
 CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF

**LOCATIONS:** Melbourne (Australia)

**NCT04632992**
**PHASE 2**

A Study Evaluating Targeted Therapies in Participants Who Have Advanced Solid Tumors With Genomic Alterations or Protein Expression Patterns Predictive of Response

**TARGETS**  
 TRKB, ALK, TRKC, ROS1, TRKA, PD-L1, ERBB2, PI3K-alpha, RET, AKTs

**LOCATIONS:** Alaska, Washington, California, Idaho

**NCT04991480**
**PHASE 1/2**

A Study of ART4215 for the Treatment of Advanced or Metastatic Solid Tumors

**TARGETS**  
 PARP, Pol theta

**LOCATIONS:** London (United Kingdom), Oklahoma, New York, Tennessee, Texas, Florida

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 09 September 2022  
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1446725-01

## APPENDIX

## Variants of Unknown Significance

**NOTE** One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

**CSF1R**  
P386L

**CYP17A1**  
G438V and Q424\*

**DDR1**  
R466H

**FGFR3**  
L164V

**FLCN**  
S407R

**NOTCH1**  
S1674Y

**PIK3C2G**  
E1307G

**PIK3CA**  
E78K

**PTPRO**  
K25E

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 09 September 2022  
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1446725-01

**APPENDIX**

Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

**DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS**

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B or WTX)	
APC	AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX
AURKA	AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2
BCL6	BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1
BTG2	BTK	CALR	CARD11	CASP8	CBFB	CBL	CCND1	CCND2
CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73	CDH1
CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B	CDKN2C
CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R	CTCF
CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1	DDR2
DIS3	DNMT3A	DOT1L	EED	EGFR	EMSY (C11orf30)	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12	FGF14
FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3	FGFR4
FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3	GATA4
GATA6	GID4 (C17orf39)	GNA11	GNA13	GNAS	GNAS	GRM3	GSK3B	H3-3A (H3F3A)
HDAC1	HGF	HNFA1	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDM5A	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11 (MRE11A)	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD2 (WHSC1 or MMSET)	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3	NTRK3
P2RY8	PALB2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)
PDGFRA	PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1
PMS2	POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI
PRKN (PARK2)	PTCH1	PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51
RAD51B	RAD51C	RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10
REL	RET	RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC
SDHD	SETD2	SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO
SNCAIP	SOC1	SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3
STK11	SUFU	SYK	TBX3	TEK	TENT5C (FAM46C)	TET2	TET2	TGFB2
TIPARP	TNFAIP3	TNFRSF14	TP53	TSC1	TSC2	TYRO3	U2AF1	VEGFA
VHL	WT1	XPO1	XRCC2	ZNF217	ZNF703			

**DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS**

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSP02	SDC4	SLC34A2	TERC*	TERT**	TPRSS2

\*TERC is an NCRNA

\*\*Promoter region of TERT is interrogated


**ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS**

Homologous Recombination status  
Loss of Heterozygosity (LOH) score  
Microsatellite (MS) status  
Tumor Mutational Burden (TMB)

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

ORDERED TEST # ORD-1446725-01

**APPENDIX**
**About FoundationOne®CDx**

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a., Ciplstraat 3, 2440 Geel, Belgium. 

**ABOUT FOUNDATIONONE CDx**

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:  
[www.rochefoundationmedicine.com/f1cdxtech](http://www.rochefoundationmedicine.com/f1cdxtech).

**INTENDED USE**

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

**TEST PRINCIPLE**

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes

detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

**THE REPORT**

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

**Diagnostic Significance**

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

**Qualified Alteration Calls (Equivocal and Subclonal)**

An alteration denoted as "amplification – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for *ERBB2* and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical

methodology has identified as being present in <10% of the assayed tumor DNA.

**Ranking of Therapies and Clinical Trials**
*Ranking of Therapies in Summary Table*

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

*Ranking of Clinical Trials*

Pediatric trial qualification → Geographical proximity → Later trial phase.

**NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION**

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) ([www.nccn.org](http://www.nccn.org)). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to [NCCN.org](http://NCCN.org). NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

**Limitations**

1. In the fractional-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 09 September 2022  
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531  
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531  
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531



ORDERED TEST # ORD-1446725-01

APPENDIX

About FoundationOne®CDx

- analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.
- TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation [https://www.accessdata.fda.gov/cdrh\\_docs/pdf17/P170019B.pdf](https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf). The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
  - Homologous Recombination status may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas (Coleman et al., 2017; 28916367). Samples with deleterious *BRCA1/2* alteration and/or Loss of Heterozygosity (LOH) score  $\geq 16\%$  will be reported as "HRD Positive" and samples with absence of these findings will be reported as "HRD Not Detected," agnostic of potential secondary *BRCA1/2* reversion alterations. Certain potentially deleterious missense or small in-frame deletions in *BRCA1/2* may not be classified as deleterious and, in the absence of an elevated LOH profile, samples with such mutations may be classified as "HRD Not Detected." A result of "HRD Not Detected" does not rule out the presence of a *BRCA1/2* alteration or an elevated LOH profile outside the assay performance characteristic limitations.
  - The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and

- extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
- Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
  - Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine whether the patient is a candidate for biopsy.
  - Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of *HER2* overexpression as 20% in breast cancer. Based on the F1CDx *HER2* CDx concordance study, approximately 10% of *HER2* amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

## REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal

hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

## VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

\*Interquartile Range = 1st Quartile to 3rd Quartile

## VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

ORDERED TEST # ORD-1446725-01

**APPENDIX**
**About FoundationOne®CDx**

tumor sequencing is germline or somatic.  
Interpretation should be based on clinical context.

### VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D (MLL2)*, *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

### LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

### NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

### NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

### TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking

into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

### SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

### REFERENCE SEQUENCE INFORMATION

Sequence data is mapped to the human genome, Genome Reference Consortium Human Build 37 (GRCh37), also known as hg19.

MR Suite Version (RG) 7.1.0

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of the data. Please refer to the original report for the suitability of use.

The median exon coverage for this sample is 451x

**ORDERED TEST #**    **ORD-1446725-01**
**APPENDIX    References**

1. Histopathology (2007) PMID: 17204026
2. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
3. Overman MJ, et al. Lancet Oncol. (2017) PMID: 28734759
4. Overman MJ, et al. J. Clin. Oncol. (2018) PMID: 29355075
5. Lipson EJ, et al. Clin. Cancer Res. (2013) PMID: 23169436
6. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
7. Rizvi NA, et al. Science (2015) PMID: 25765070
8. Oaknin A, et al. JAMA Oncol (2020) PMID: 33001143
9. Hochster et al., 2017; ASCO Abstract 673
10. Fleming et al., 2018; ASCO Abstract 5585
11. Bang et al., 2018; ASCO Abstract 92
12. Zigelboim I, et al. J. Clin. Oncol. (2007) PMID: 17513808
13. Hampel H, et al. Cancer Res. (2006) PMID: 16885385
14. Stelloo E, et al. Clin. Cancer Res. (2016) PMID: 27006490
15. Kanopienė D, et al. Medicina (Kaunas) (2014) PMID: 25458958
16. Black D, et al. J. Clin. Oncol. (2006) PMID: 16549821
17. Nout RA, et al. Gynecol. Oncol. (2012) PMID: 22609107
18. Steinbakk A, et al. Cell Oncol (Dordr) (2011) PMID: 21547578
19. Bilbao C, et al. Int. J. Radiat. Oncol. Biol. Phys. (2010) PMID: 20005452
20. Guastadisegni C, et al. Eur. J. Cancer (2010) PMID: 20627535
21. Pawlik TM, et al. Dis. Markers (2004) PMID: 15528785
22. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
23. Nature (2012) PMID: 22810696
24. Hiyama T, et al. J. Gastroenterol. Hepatol. (2004) PMID: 15209621
25. Wu MS, et al. Cancer Res. (1998) PMID: 9537253
26. dos Santos NR, et al. Gastroenterology (1996) PMID: 8536886
27. Fang WL, et al. Biomed Res Int (2013) PMID: 23555086
28. Farris AB, et al. Am. J. Surg. Pathol. (2011) PMID: 21422910
29. Agaram NP, et al. Am. J. Clin. Pathol. (2010) PMID: 20395525
30. Ruemmele P, et al. Am. J. Surg. Pathol. (2009) PMID: 19252434
31. Planck M, et al. Cancer (2003) PMID: 12627520
32. Hibi K, et al. Jpn. J. Cancer Res. (1995) PMID: 7775257
33. Muneyuki T, et al. Dig. Dis. Sci. (2000) PMID: 11117578
34. Zhang SH, et al. World J. Gastroenterol. (2005) PMID: 15918185
35. Chiappini F, et al. Carcinogenesis (2004) PMID: 14656944
36. Suto T, et al. J Surg Oncol (2001) PMID: 11223838
37. Momoi H, et al. J. Hepatol. (2001) PMID: 11580146
38. Liengswangwong U, et al. Int. J. Cancer (2003) PMID: 14506736
39. Moy AP, et al. Virchows Arch. (2015) PMID: 25680569
40. Yoshida T, et al. J. Gastroenterol. (2000) PMID: 11063221
41. Pritchard CC, et al. Nat Commun (2014) PMID: 25255306
42. Azzouzi AR, et al. BJU Int. (2007) PMID: 17233803
43. Burger M, et al. J. Mol. Med. (2006) PMID: 16924473
44. Bai S, et al. Am. J. Clin. Pathol. (2013) PMID: 23690119
45. Giedl J, et al. Am. J. Clin. Pathol. (2014) PMID: 25319978
46. Yamamoto Y, et al. Clin. Cancer Res. (2006) PMID: 16675567
47. Smyth et al., 2015; ASCO Gastrointestinal Cancers Symposium Abstract 62
48. Bilbao-Sieyro C, et al. Oncotarget (2014) PMID: 25026289
49. Mackay HJ, et al. Eur. J. Cancer (2010) PMID: 20304627
50. Arabi H, et al. Gynecol. Oncol. (2009) PMID: 19275958
51. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
52. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
53. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
54. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
55. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
56. Cristescu R, et al. Science (2018) PMID: 30309915
57. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
58. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
59. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
60. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
61. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
62. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
63. Marabelle A, et al. Lancet Oncol. (2020) PMID: 32919526
64. Ott PA, et al. J. Clin. Oncol. (2019) PMID: 30557521
65. Cristescu R, et al. J Immunother Cancer (2022) PMID: 35101941
66. Friedman CF, et al. Cancer Discov (2022) PMID: 34876409
67. Sturgill EG, et al. Oncologist (2022) PMID: 35274716
68. Schenker at al., 2022; AACR Abstract 7845
69. Legrand et al., 2018; ASCO Abstract 12000
70. Chalmers ZR, et al. Genome Med (2017) PMID: 28420421
71. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31003990
72. South AP, et al. J. Invest. Dermatol. (2014) PMID: 24662767
73. Pickering CR, et al. Clin. Cancer Res. (2014) PMID: 25303977
74. Spigel et al., 2016; ASCO Abstract 9017
75. Xiao D, et al. Oncotarget (2016) PMID: 27009843
76. Shim HS, et al. J Thorac Oncol (2015) PMID: 26200269
77. Govindan R, et al. Cell (2012) PMID: 22980976
78. Ding L, et al. Nature (2008) PMID: 18948947
79. Imielinski M, et al. Cell (2012) PMID: 22980975
80. Kim Y, et al. J. Clin. Oncol. (2014) PMID: 24323028
81. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
82. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
83. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
84. Johnson BE, et al. Science (2014) PMID: 24336570
85. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
86. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
87. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
88. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
89. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
90. Fritsch C, et al. Mol. Cancer Ther. (2014) PMID: 24608574
91. Juric D, et al. J. Clin. Oncol. (2018) PMID: 29401002
92. Gallant JN, et al. NPJ Precis Oncol (2019) PMID: 30793038
93. Delestre F, et al. Sci Transl Med (2021) PMID: 34613809
94. Morschhauser F, et al. Mol Cancer Ther (2020) PMID: 31619463
95. Patnaik A, et al. Ann. Oncol. (2016) PMID: 27672108
96. Santin AD, et al. Gynecol Oncol Rep (2020) PMID: 31934607
97. Damodaran S, et al. J Clin Oncol (2022) PMID: 35133871
98. André F, et al. N. Engl. J. Med. (2019) PMID: 31091374
99. Smyth LM, et al. NPJ Breast Cancer (2021) PMID: 33863913
100. Park HS, et al. PLoS ONE (2016) PMID: 27105424
101. Lim SM, et al. Oncotarget (2016) PMID: 26859683
102. Hou MM, et al. Oncotarget (2014) PMID: 25426553
103. Varnier R, et al. Eur J Cancer (2019) PMID: 31351267
104. Janku F, et al. Cell Rep (2014) PMID: 24440717
105. Moroney J, et al. Clin. Cancer Res. (2012) PMID: 22927482
106. Basho RK, et al. JAMA Oncol (2017) PMID: 27893038
107. Moroney JW, et al. Clin. Cancer Res. (2011) PMID: 21890452
108. Krop et al., 2018; ASCO Abstract 101
109. Pascual J, et al. Cancer Discov (2021) PMID: 32958578
110. Dolly SO, et al. Clin. Cancer Res. (2016) PMID: 26787751
111. Canaud et al., 2021; ESMO Abstract LBA23
112. Aust Fam Physician (1986) PMID: 2941002
113. Nature (2015) PMID: 25631445
114. Nature (2012) PMID: 22960745
115. Lin DC, et al. Nat. Genet. (2014) PMID: 24686850
116. Han MR, et al. Exp. Mol. Med. (2018) PMID: 29422544
117. Zhang L, et al. Onco Targets Ther (2013) PMID: 23674897
118. Wada S, et al. Ann. Surg. Oncol. (2006) PMID: 16788758
119. Shigaki H, et al. Clin. Cancer Res. (2013) PMID: 23528889
120. Akagi I, et al. Int. J. Oncol. (2009) PMID: 19212681
121. Lui VW, et al. Cancer Discov (2013) PMID: 23619167
122. Samuels Y, et al. Cancer Cell (2005) PMID: 15950905
123. Nat. Rev. Cancer (2009) PMID: 19629070
124. Rivière JB, et al. Nat. Genet. (2012) PMID: 22729224
125. Loconte DC, et al. PLoS ONE (2015) PMID: 25915946
126. Huchtagowder V, et al. Clin. Genet. (2017) PMID: 27307077
127. Kuentz P, et al. Genet. Med. (2017) PMID: 28151489
128. Mirzaa G, et al. JCI Insight (2016) PMID: 27631024
129. Mirzaa GM, et al. Am J Med Genet C Semin Med Genet (2013) PMID: 23592320
130. Courtney KD, et al. J. Clin. Oncol. (2010) PMID: 20085938
131. Simpson L, et al. Exp. Cell Res. (2001) PMID: 11237521
132. Milella M, et al. Sci Rep (2017) PMID: 28220839
133. Templeton AJ, et al. Eur. Urol. (2013) PMID: 23582881
134. Sweeney C, et al. Lancet (2021) PMID: 34246347
135. de Bono JS, et al. Clin. Cancer Res. (2019) PMID: 30037818
136. Saura C, et al. Cancer Discov (2017) PMID: 27872130
137. Voss MH, et al. Clin. Cancer Res. (2018) PMID: 30327302
138. André F, et al. J. Clin. Oncol. (2016) PMID: 27091708
139. Schmid P, et al. J. Clin. Oncol. (2019) PMID: 31841354
140. Weldon Gilcrease G, et al. Invest New Drugs (2019) PMID: 30302599
141. Mendes-Pereira AM, et al. EMBO Mol Med (2009) PMID: 20049735
142. Shen Y, et al. Clin. Cancer Res. (2013) PMID: 23881923
143. Chatterjee P, et al. PLoS ONE (2013) PMID: 23565244
144. McCormick A, et al. Int. J. Gynecol. Cancer (2016) PMID: 26905328
145. Forster MD, et al. Nat Rev Clin Oncol (2011) PMID: 21468130
146. Eikesdal HP, et al. Ann Oncol (2021) PMID: 33242536

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

ORDERED TEST # ORD-1446725-01

**APPENDIX**
**References**

147. Dougherty et al., 2014; ASCO Abstract 5536
148. Pan M, et al. Perm J (2021) PMID: 33970096
149. Sandhu SK, et al. Lancet Oncol. (2013) PMID: 23810788
150. Romero I, et al. Gynecol Oncol (2020) PMID: 32988624
151. Song Y, et al. Nature (2014) PMID: 24670651
152. Stransky N, et al. Science (2011) PMID: 21798893
153. Agrawal N, et al. Science (2011) PMID: 21798897
154. Rahmani A, et al. Int J Clin Exp Pathol (2012) PMID: 23119114
155. Campbell RB, et al. J. Biol. Chem. (2003) PMID: 12857747
156. Rodríguez-Escudero I, et al. Hum. Mol. Genet. (2011) PMID: 21828076
157. He X, et al. Cancer Res. (2013) PMID: 23475934
158. Han SY, et al. Cancer Res. (2000) PMID: 10866302
159. Myers MP, et al. Proc. Natl. Acad. Sci. U.S.A. (1998) PMID: 9811831
160. Pradella LM, et al. BMC Cancer (2014) PMID: 24498881
161. Kim JS, et al. Mol. Cell. Biol. (2011) PMID: 21536651
162. Denning G, et al. Oncogene (2007) PMID: 17213812
163. Hlobilkova A, et al. Anticancer Res. ( ) PMID: 16619501
164. Redfern RE, et al. Protein Sci. (2010) PMID: 20718038
165. Shenoy S, et al. PLoS ONE (2012) PMID: 22505997
166. Wang Y, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) PMID: 19329485
167. Okumura K, et al. J. Biol. Chem. (2006) PMID: 16829519
168. Lee JO, et al. Cell (1999) PMID: 10555148
169. Maxwell GL, et al. Cancer Res. (1998) PMID: 9635567
170. Risinger JJ, et al. Clin. Cancer Res. (1998) PMID: 9865913
171. Kato H, et al. Clin. Cancer Res. (2000) PMID: 11051241
172. Fenton TR, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) PMID: 22891331
173. Ngeow J, et al. J. Clin. Endocrinol. Metab. (2012) PMID: 23066114
174. Lobo GP, et al. Hum. Mol. Genet. (2009) PMID: 19457929
175. Liu J, et al. Oncogene (2014) PMID: 23995781
176. Maehama T, et al. Annu. Rev. Biochem. (2001) PMID: 11395408
177. De Vivo I, et al. J. Med. Genet. (2000) PMID: 10807691
178. Ramaswamy S, et al. Proc. Natl. Acad. Sci. U.S.A. (1999) PMID: 10051603
179. Liu JL, et al. Mol. Cell. Biol. (2005) PMID: 15988030
180. Karoui M, et al. Br. J. Cancer (2004) PMID: 15026806
181. Gil A, et al. PLoS ONE (2015) PMID: 25875300
182. Furnari FB, et al. Cancer Res. (1998) PMID: 9823298
183. Spinelli L, et al. J. Med. Genet. (2015) PMID: 25527629
184. Mingo J, et al. Eur. J. Hum. Genet. (2018) PMID: 29706633
185. Wang Q, et al. J. Mol. Graph. Model. (2010) PMID: 20538496
186. Andrés-Pons A, et al. Cancer Res. (2007) PMID: 17942903
187. Butler MG, et al. J. Med. Genet. (2005) PMID: 15805158
188. Georgescu MM, et al. Proc. Natl. Acad. Sci. U.S.A. (1999) PMID: 10468583
189. Staal FJ, et al. Br. J. Cancer (2002) PMID: 12085208
190. Nguyen HN, et al. Oncogene (2014) PMID: 24292679
191. Rahdar M, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) PMID: 19114656
192. Das S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) PMID: 12808147
193. Wang X, et al. Biochem. J. (2008) PMID: 18498243
194. Valiente M, et al. J. Biol. Chem. (2005) PMID: 15951562
195. Nguyen HN, et al. Oncogene (2015) PMID: 25263454
196. Shan L, et al. Cell Discov (2020) PMID: 32704382
197. Landrum MJ, et al. Nucleic Acids Res. (2018) PMID: 29165669
198. Blumenthal GM, et al. Eur. J. Hum. Genet. (2008) PMID: 18781191
199. Orloff MS, et al. Oncogene (2008) PMID: 18794875
200. Zbuk KM, et al. Nat. Rev. Cancer (2007) PMID: 17167516
201. Cerami E, et al. Cancer Discov (2012) PMID: 22588877
202. Gao J, et al. Sci Signal (2013) PMID: 23550210
203. Ito S, et al. Nature (2010) PMID: 20639862
204. Guo JU, et al. Cell (2011) PMID: 21496894
205. Iyer LM, et al. Cell Cycle (2009) PMID: 19411852
206. Ko M, et al. Nature (2010) PMID: 21057493
207. Yang H, et al. Oncogene (2013) PMID: 22391558
208. Hu L, et al. Cell (2013) PMID: 24315485
209. Wang Y, et al. Mol. Cell (2015) PMID: 25601757
210. Jaiswal S, et al. N. Engl. J. Med. (2014) PMID: 25426837
211. Genovese G, et al. N. Engl. J. Med. (2014) PMID: 25426838
212. Xie M, et al. Nat. Med. (2014) PMID: 25326804
213. Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) PMID: 28669404
214. Severson EA, et al. Blood (2018) PMID: 29678827
215. Fuster JJ, et al. Circ. Res. (2018) PMID: 29420212
216. Hematology Am Soc Hematol Educ Program (2018) PMID: 30504320
217. Chabon JJ, et al. Nature (2020) PMID: 32269342
218. Razavi P, et al. Nat. Med. (2019) PMID: 31768066
219. Janku F, et al. Cancer Res. (2013) PMID: 23066039
220. Janku F, et al. J. Clin. Oncol. (2012) PMID: 22271473
221. Janku F, et al. Mol. Cancer Ther. (2011) PMID: 21216929
222. Moulder S, et al. Ann. Oncol. (2015) PMID: 25878190
223. Byeon et al., 2020; doi: 10.21037/tcr.2020.04.07
224. Varadarajan et al., 2012; ASCO Abstract 5541
225. Fury MG, et al. Cancer Chemother. Pharmacol. (2012) PMID: 21913034
226. Tolcher AW, et al. Ann. Oncol. (2015) PMID: 25344362
227. Patterson et al., 2018; AACR Abstract 3891
228. Trafalis DT, et al. Anticancer Drugs (2012) PMID: 22510794
229. MacKenzie MJ, et al. Invest New Drugs (2012) PMID: 20978924
230. Fury MG, et al. Cancer Chemother. Pharmacol. (2012) PMID: 22644799
231. Bauman JE, et al. Oral Oncol. (2013) PMID: 23384718
232. Tinker AV, et al. Gynecol. Oncol. (2013) PMID: 23672928
233. Piha-Paul SA, et al. Oncotarget (2014) PMID: 24742900

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 09 September 2022  
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531  
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531  
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531