

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT	DISEASE Brain anaplastic astrocytoma	PHYSICIAN	ORDERING PHYSICIAN Yeh, Yi-Chen	SPECIMEN	SPECIMEN SITE Brain
	NAME Wang, Lun-I		MEDICAL FACILITY Taipei Veterans General Hospital		SPECIMEN ID S111-08919A (PF22039)
	DATE OF BIRTH 26 September 1960		ADDITIONAL RECIPIENT None		SPECIMEN TYPE Slide Deck
	SEX Male		MEDICAL FACILITY ID 205872		DATE OF COLLECTION 04 March 2022
	MEDICAL RECORD # 48245789		PATHOLOGIST Not Provided		SPECIMEN RECEIVED 21 March 2022

Biomarker Findings

Microsatellite status - Cannot Be Determined^α

Tumor Mutational Burden - 4 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

FGFR1 K656E

PIK3CA E545K

CDK4 amplification

ATRX splice site 5787-2A>G

CDKN2A/B CDKN2A loss, CDKN2B loss

MTAP loss

^α Patients with Microsatellite status of Cannot Be Determined should be re-tested with an orthogonal (alternative) method.

Report Highlights

- Targeted therapies with potential clinical benefit **approved in another tumor type**: Everolimus (p. 8), Infigratinib (p. 8), Temsirolimus (p. 9)
- Evidence-matched **clinical trial options** based on this patient's genomic findings: (p. 10)

BIOMARKER FINDINGS

Microsatellite status - Cannot Be Determined

Tumor Mutational Burden - 4 Muts/Mb

GENOMIC FINDINGS

FGFR1 - K656E

10 Trials *see p. 12*

PIK3CA - E545K

10 Trials *see p. 14*

CDK4 - amplification

10 Trials *see p. 10*

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. *see Biomarker Findings section*

No therapies or clinical trials. *see Biomarker Findings section*

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)
none	Infigratinib
none	Everolimus
none	Temsirolimus
none	none

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GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

ATRX - splice site 5787-2A>G p. 5 **MTAP - loss** p. 7
CDKN2A/B - CDKN2A loss, CDKN2B loss p. 6

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

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ORDERED TEST # ORD-1326253-01

BIOMARKER FINDINGS

BIOMARKER

Microsatellite status

RESULT

Cannot Be Determined

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of prospective clinical evidence in multiple solid tumor types, microsatellite instability (MSI) and associated increased tumor mutational burden (TMB)¹⁻² may predict sensitivity to immune checkpoint inhibitors, including the approved PD-1-targeting agents cemiplimab, dostarlimab, nivolumab (alone or in combination with ipilimumab), and

pembrolizumab³⁻⁸ and PD-L1-targeting agents atezolizumab, avelumab, and durvalumab⁹⁻¹¹. As the MSI status of this tumor is unknown, the relevance of these therapeutic approaches is unclear.

FREQUENCY & PROGNOSIS

MSI-High has been reported in 3-8% of adult or pediatric astrocytomas and was generally not associated with Lynch syndrome¹²⁻¹⁴. Low-level MSI has been reported in 5-9% of glioblastoma (GBM) samples¹⁵⁻¹⁷. A large-scale study did not find high-level microsatellite instability (MSI-H) in any of 129 GBM samples¹⁵, although a small-scale study reported MSI-H in 4 of 15 pediatric GBMs and 1 of 12 adult GBMs¹⁸. The frequency of MSI has been reported to be increased in relapsed compared to primary GBM¹⁵, in GBMs with a

previous lower grade astrocytoma¹⁶, and in giant cell GBM compared to classic GBM¹⁷.

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor¹⁹. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2¹⁹⁻²¹. The level of MSI in this sample could not be determined with confidence. Depending on the clinical context, MSI testing of an alternate sample or by another methodology could be considered.

BIOMARKER

Tumor Mutational Burden

RESULT

4 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1²²⁻²⁴, anti-PD-1 therapies²²⁻²⁵, and combination nivolumab and ipilimumab²⁶⁻³¹. In glioma, a lack of association between TMB and clinical benefit from immune checkpoint inhibitors has been reported^{22,32-33}. However, multiple case studies have reported that patients with ultramutated gliomas driven by POLE

mutations have benefited from treatment with anti-PD-1³⁴⁻³⁵ or anti-PD-L1³⁶ therapies. Therefore, although increased TMB alone may not be a strong biomarker for PD-1 or PD-L1 inhibitors in this cancer type, these agents may have efficacy for patients with glioma harboring both high TMB and POLE mutation.

FREQUENCY & PROGNOSIS

Anaplastic astrocytoma harbors a median TMB of 1.8 mutations per megabase (mut/Mb), and 2% of cases have high TMB (>20 mut/Mb)³⁷. For pediatric patients, high TMB has been reported in a subset of high-grade gliomas, frequently in association with mutations in mismatch repair or proofreading genes and in TP53, whereas BRAF alterations or other oncogene fusions were observed more frequently in brain tumors harboring low TMB³⁸⁻³⁹. Increased TMB has been reported to correlate with higher tumor grade in glioma⁴⁰ and glioblastoma (GBM) tissue samples with biallelic mismatch repair deficiency

(bMMRD)³⁴, as well as with shorter OS of patients with diffuse glioma⁴¹.

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma⁴²⁻⁴³ and cigarette smoke in lung cancer^{7,44}, treatment with temozolomide-based chemotherapy in glioma⁴⁵⁻⁴⁶, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes⁴⁷⁻⁵¹, and microsatellite instability (MSI)^{47,50-51}. This sample harbors a TMB below levels that would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents^{22,32-36}.

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GENOMIC FINDINGS

GENE

FGFR1

ALTERATION

K656E

TRANSCRIPT ID

NM_023110

CODING SEQUENCE EFFECT

1966A>G

VARIANT ALLELE FREQUENCY (% VAF)

37.7%

reported in patients with primary brain tumors^{59,61} and lung squamous cell carcinoma⁶⁷ treated with FGFR inhibitors. In a phase 1 study of futibatinib, 2 patients with FGFR1-mutated primary brain tumors exhibited PRs⁶¹. A patient with FGFR1-mutated glioblastoma exhibited a PR when treated with infigratinib⁶⁸. For pediatric patients with FGFR1-mutated gliomas, a case series reported 1 sustained PR for a patient with high grade glioma, and a sustained SD and 1 PD for patients with low grade gliomas following treatment with Debio 1347⁵⁹.

astrocytomas⁷². Mutations in the FGFR1 kinase domain have been reported in both lower-grade gliomas and glioblastomas; one of these mutations has been described as an oncogenic mutation that disrupted autophosphorylation⁷³⁻⁷⁵. FGFR fusions were identified in 3/85 IDH1 and IDH2 wild-type gliomas, but were not found in any of 126 IDH1- or IDH2-mutant gliomas⁷⁶. Patients with FGFR1-altered pilocytic astrocytomas have been associated with poor prognosis⁷⁷⁻⁷⁸, although published data investigating the prognostic implications of FGFR1 alterations independent to co-occurring alterations in gliomas are limited (PubMed, Mar 2022)^{71,79}.

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Alterations that activate FGFR1 may predict sensitivity to selective FGFR inhibitors including erdafitinib⁵²⁻⁵⁴, pemigatinib⁵⁵, infigratinib⁵⁶⁻⁵⁷, rogaratinib⁵⁸, Debio 1347⁵⁹⁻⁶⁰, futibatinib⁶¹, and derazantinib⁶², or multikinase inhibitors such as pazopanib⁶³ and ponatinib⁶⁴⁻⁶⁶. In the context of FGFR1 mutation, clinical responses have been

FREQUENCY & PROGNOSIS

In the Brain Lower Grade Glioma TCGA dataset and the Glioblastoma Multiforme TCGA dataset, mutation of FGFR1 has been found in less than 1% of cases⁶⁹⁻⁷⁰. In pediatric patients, FGFR1 alterations have been identified in 18% of low-grade gliomas³⁹, including 5/9 pilomyxoid astrocytomas, 8% of high-grade gliomas³⁹, and in 6% (4/64) of thalamic gliomas⁷¹. FGFR1 mutation has also been reported in 5% (5/96) of pilocytic

FINDING SUMMARY

FGFR1 encodes the protein fibroblast growth factor receptor 1, which plays key roles in regulation of the cell cycle and angiogenesis and is an upstream regulator of the RAS, MAPK, and AKT signaling pathways⁸⁰. The FGFR1 alteration observed here has been characterized as activating and is predicted to be oncogenic^{74,81-83}.

GENE

PIK3CA

ALTERATION

E545K

TRANSCRIPT ID

NM_006218

CODING SEQUENCE EFFECT

1633G>A

VARIANT ALLELE FREQUENCY (% VAF)

39.5%

combination with the CDK4/6 inhibitor palbociclib for patients with PIK3CA-mutated solid tumors reported an ORR of 0% (n=12) and a DCR of 17% (2/12)⁹⁸. In a Phase 1 trial of the dual PI3K/mTOR kinase inhibitor apitolisib, 79% (11/14) of patients with PIK3CA-mutated advanced solid tumors experienced disease control (3 PRs, 8 SDs)⁹⁹. The PI3K inhibitor alpelisib demonstrated an ORR of 6.0% (8/134) and a DCR of 58% (78/134) in a study for patients with PIK3CA-mutated solid tumors¹⁰⁰. However, the PI3K inhibitor copanlisib exhibited limited efficacy in PIK3CA-mutated tumors¹⁰¹⁻¹⁰².

HR=2.89, p=0.01) and in the TCGA cohort (6.1 vs. 9 months, p=0.008), but was not consistently associated with median OS¹⁰⁸. In a study of IDH-wildtype GBM, patients with alterations in PI3K class I genes (PIK3CA, PIK3R1, PIK3CG, and PIK3R2) had significantly longer OS (20.0 months altered vs. 16.9 months wildtype, HR=0.62, p=0.002) and PFS (11.0 months altered vs. 7.4 months wildtype, p=0.0043); patients with PIK3CA alterations experienced an improved OS but this association was not highly significant (20.0 months altered vs. 18.1 months wildtype, p=0.0407)¹⁰⁹.

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Clinical and preclinical data in various tumor types indicate that PIK3CA activating alterations may predict sensitivity to therapies targeting PI3K⁸⁴⁻⁸⁶, AKT⁸⁷⁻⁸⁸, or mTOR⁸⁹⁻⁹⁶. In the Phase 2 MATCH trial for patients with PIK3CA-mutated solid tumors, 28% (18/65) of patients experienced PFS lasting at least 6 months after treatment with taselisib; however, no ORs were observed in this study⁹⁷. A separate Phase 1b study of taselisib in

FREQUENCY & PROGNOSIS

PIK3CA mutations have been reported in 5-23% of high-grade gliomas (including glioblastomas, anaplastic astrocytomas, and anaplastic oligodendrogliomas)¹⁰³⁻¹⁰⁷. While another study did not observe PIK3CA mutations in low-grade astrocytomas or in anaplastic astrocytomas, it did report high ERK and AKT activity¹⁰³. One study found that PIK3CA mutation in glioblastoma (GBM) was associated with shorter median PFS in both a discovery cohort (6.9 vs. 12.4 months,

FINDING SUMMARY

PIK3CA encodes p110-α, which is the catalytic subunit of phosphatidylinositol 3-kinase (PI3K). The PI3K pathway is involved in cell signaling that regulates a number of critical cellular functions, including cell growth, proliferation, differentiation, motility, and survival¹¹⁰⁻¹¹¹. PIK3CA alterations that have been characterized as activating, such as observed here, are predicted to be oncogenic¹¹²⁻¹³³.

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GENOMIC FINDINGS
GENE
CDK4
ALTERATION
amplification

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

CDK4 amplification or activation may predict sensitivity to CDK4/6 inhibitors such as abemaciclib, palbociclib, and ribociclib¹³⁴⁻¹³⁷. Clinical benefit has been reported for limited tumor types including patients with

CDK4-amplified liposarcoma and sarcoma in response to treatment with abemaciclib¹³⁸, palbociclib^{134,139}, and ribociclib¹⁴⁰.

FREQUENCY & PROGNOSIS

Across TCGA and MKSCC studies, CDK4 amplification has been reported in 4.0-9.4% of glioma cases and 14% of glioblastoma multiforme cases (cBioPortal, Sep 2021)^{69,141-144}. A study has reported amplification of the 12q14-15 region, where CDK4 and MDM2 reside, in 5% (2/42) of glioblastomas¹⁴⁵. Amplification of CDK4 and corresponding increased CDK4 protein expression has been reported to be associated with a poorer patient outcome in anaplastic astrocytoma and

glioblastoma¹⁴⁶⁻¹⁴⁹.

FINDING SUMMARY

CDK4 encodes the cyclin-dependent kinase 4, which regulates the cell cycle, senescence, and apoptosis¹⁵⁰. CDK4 and its functional homolog CDK6 are activated by D-type cyclins and promote cell cycle progression by inactivating the tumor suppressor Rb¹⁵¹⁻¹⁵². Amplification of the chromosomal region that includes CDK4 has been reported in multiple cancer types, including lung cancer, glioblastoma, and liposarcoma, and has been associated with overexpression of CDK4 protein^{134,153-159}.

GENE
ATRX
ALTERATION
splice site 5787-2A>G

TRANSCRIPT ID
NM_000489

CODING SEQUENCE EFFECT
5787-2A>G

VARIANT ALLELE FREQUENCY (% VAF)
67.0%

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

No targeted therapies are available to directly address ATRX inactivation. Based on preclinical¹⁶⁰⁻¹⁶¹ and limited clinical data¹⁶², ATRX alterations may confer sensitivity to combination strategies involving WEE1 inhibition. In a Phase 2 study evaluating the WEE1 inhibitor adavosertib plus irinotecan for the treatment of pediatric patients with neuroblastoma, prolonged SD was reported for 44% (4/9) of patients with ATRX-deficient tumors and responses were seen in two tumors that had evidence of ALT¹⁶². Preclinical evidence also suggests that ATRX deficiency may impart sensitivity to synthetic lethal approaches

involving PARP inhibition and irinotecan¹⁶³, combined PARP and ATR inhibition¹⁶¹, or double-strand break-induction with agents such as doxorubicin, irinotecan, and topotecan¹⁶⁴; however, these approaches have not been demonstrated clinically.

FREQUENCY & PROGNOSIS

Somatic mutation of ATRX has been reported in a number of solid tumor types, often associated with ALT¹⁶⁵. ATRX mutation correlating with ALT has been reported in 10-20% of pancreatic neuroendocrine tumors (PNETs)¹⁶⁵⁻¹⁶⁷, 12.6% of pheochromocytomas and paragangliomas¹⁶⁸, and 48% of adolescent and young adult (AYA) patients with glioblastoma (GBM) or neuroblastoma¹⁶⁹⁻¹⁷³. ATRX loss in PNET^{166,174} and melanoma¹⁷⁵ and mutation in other neuroendocrine tumors¹⁶⁸ is associated with poor prognosis. Pediatric patients with high-grade glioma and ATRX mutation were shown to have more aggressive disease but are more responsive to treatment with double-strand break therapy¹⁶⁴. ATRX mutation or loss of expression is more frequent in Grade 2/3 astrocytoma and secondary GBM than primary GBM, oligodendroglioma, and oligoastrocytoma¹⁷⁶⁻¹⁷⁹ and has been proposed as a distinguishing biomarker¹⁷⁷⁻¹⁷⁹. ATRX mutation has not been detected in concurrence with MYCN

amplification in glioma and neuroblastoma¹⁷⁰⁻¹⁷³. Low-grade gliomas with both IDH1/2 mutation and ATRX mutation are associated with worse prognosis than those with IDH1/2 mutation but no ATRX mutation¹⁷⁷. Loss of ATRX protein expression has been reported in 33-39% of incidences of leiomyosarcoma (LMS) associating with ALT, a poor prognostic factor across all LMS subtypes, and with poor prognosis in extrauterine LMS but not in uterine LMS¹⁸⁰⁻¹⁸¹.

FINDING SUMMARY

ATRX encodes a SWI/SNF chromatin remodeling protein implicated in histone variant H3.3 deposition, transcriptional regulation, and telomere maintenance¹⁸²⁻¹⁸³. ATRX inactivation or loss of expression is associated with alternative lengthening of telomeres (ALT)^{165,181,184-185}. Alterations that disrupt the ADD domain (aa 167-270) or helicase domain (aa 2010-2280) of ATRX are predicted to result in loss of ATRX function¹⁸⁶⁻¹⁸⁸; however, the loss of ATRX function is not sufficient to induce ALT, which requires other undetermined factors^{182,189}. Germline mutations in ATRX give rise to alpha-thalassemia X-linked intellectual disability syndrome (ATR-X syndrome)¹⁹⁰.

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GENOMIC FINDINGS

GENE

CDKN2A/B

ALTERATION

CDKN2A loss, CDKN2B loss

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Preclinical data suggest that tumors with loss of p16INK4a function may be sensitive to CDK4/6 inhibitors, such as abemaciclib, ribociclib, and palbociclib¹⁹¹⁻¹⁹⁴. Although case studies have reported that patients with breast cancer or uterine leiomyosarcoma harboring CDKN2A loss responded to palbociclib treatment¹⁹⁵⁻¹⁹⁶, multiple other clinical studies have shown no significant correlation between p16INK4a loss or inactivation and therapeutic benefit of these agents^{137,140,197-201}; it is not known whether CDK4/6 inhibitors would be beneficial in this case. Although preclinical studies have suggested that loss of p14ARF function may be associated with reduced sensitivity to MDM2 inhibitors²⁰²⁻²⁰³, the clinical relevance of p14ARF as a predictive biomarker is not clear. There are no drugs that directly target the mutation or loss of CDKN2B in cancer. Because the p15INK4b protein encoded by CDKN2B is known to inhibit CDK4, tumors with CDKN2B mutation or loss may predict sensitivity to CDK4/6 inhibitors, such as ribociclib, abemaciclib, and palbociclib^{134-135,140,199-200,204}.

FREQUENCY & PROGNOSIS

Concurrent putative homozygous deletion of CDKN2A and CDKN2B has been reported in 35% of patients with gliomas¹⁴⁴ and detected more frequently in patients with glioblastoma multiforme (GBM; 58%)⁶⁹ than in those with lower grade gliomas (13%) (cBioPortal, Sep 2021)¹⁴¹⁻¹⁴². In other studies, loss of CDKN2A/B by deletion has been reported in up to 78% of astrocytomas (including anaplastic astrocytomas and GBM)²⁰⁵⁻²⁰⁷. Decreased p14ARF and p16INK4a expression levels were found to be tightly associated in a study of glioma samples²⁰⁸. Homozygous deletion of the genomic region including CDKN2A and CDKN2B has been found to be associated with poor prognosis in GBM and likely serves as an early event in GBM progression^{206,209}. In addition, expression of p16INK4a has been found to be lower in patients with high grade malignant gliomas compared to patients with low grade gliomas, and loss of p16INK4a expression has been associated with shorter overall survival in pilocytic astrocytomas²¹⁰⁻²¹¹.

FINDING SUMMARY

CDKN2A encodes two different, unrelated tumor suppressor proteins, p16INK4a and p14ARF, whereas CDKN2B encodes the tumor suppressor p15INK4b²¹²⁻²¹³. Both p15INK4b and p16INK4a bind to and inhibit CDK4 and CDK6, thereby maintaining the growth-suppressive activity of the Rb tumor suppressor; loss or inactivation of

either p15INK4b or p16INK4a contributes to dysregulation of the CDK4/6-cyclin-Rb pathway and loss of cell cycle control²¹⁴⁻²¹⁵. The tumor suppressive functions of p14ARF involve stabilization and activation of p53, via a mechanism of MDM2 inhibition²¹⁶⁻²¹⁷. One or more alterations observed here are predicted to result in p16INK4a loss of function²¹⁸⁻²³⁹. One or more alterations seen here are predicted to result in p14ARF loss of function^{222,239-242}. CDKN2B alterations such as seen here are predicted to inactivate p15INK4b²⁴³.

POTENTIAL GERMLINE IMPLICATIONS

Germline CDKN2A mutation is associated with melanoma-pancreatic cancer syndrome, a condition marked by increased risk of developing malignant melanoma and/or pancreatic cancer²⁴⁴. Mutation carriers within families may develop either or both types of cancer, and melanoma cases may be referred to as familial or hereditary melanoma²⁴⁵⁻²⁴⁶. CDKN2A is the most implicated gene in familial melanoma, with germline mutations present in 16% to 20% of familial melanoma cases²⁴⁷⁻²⁴⁹. CDKN2A alteration has also been implicated in familial melanoma-astrocytoma syndrome, an extremely rare tumor association characterized by dual predisposition to melanoma and nervous system tumors²⁵⁰⁻²⁵². In the appropriate clinical context, germline testing of CDKN2A is recommended.

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GENOMIC FINDINGS

GENE

MTAP

ALTERATION

loss

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Preclinical and limited clinical evidence indicate that MTAP inactivation produces specific metabolic vulnerabilities. MTAP inactivation may confer sensitivity to MAT2A inhibitors²⁵³. A Phase 1 trial of MAT2A inhibitor AG-270 reported 1 PR and 2 SDs lasting longer than 6 months for patients with advanced solid tumors displaying MTAP loss²⁵⁴. Although preclinical data have suggested that MTAP loss sensitizes cells to PRMT5 inhibition^{253,255-256}, MTAP loss may not be a biomarker of response to previously developed small-molecule SAM-uncompetitive PRMT5 inhibitors²⁵⁷; dual PRMT1 and PRMT5 inhibition may be more effective²⁵⁸⁻²⁶⁰. In preclinical cancer models, MTAP inactivation showed increased

sensitivity to inhibitors of purine synthesis or purine analogs, especially upon addition of exogenous MTA, which is converted to adenine in normal cells, thereby providing competition to purine poisons lacking in MTAP-deficient cells²⁶¹⁻²⁷¹. A Phase 2 study of L-alanosine, an inhibitor of adenine synthesis, as a monotherapy for 65 patients with MTAP-deficient cancers reported no responses and stable disease in 23.6% (13/55) of patients²⁷².

FREQUENCY & PROGNOSIS

MTAP loss/homozygous deletion as well as loss of expression has been reported in a wide variety of solid tumors and hematologic cancers²⁷³⁻²⁷⁴; such events have been correlated with poor prognosis in a variety of cancer types, including hepatocellular carcinoma²⁷⁵, gastrointestinal stromal tumors²⁷⁶, mantle cell lymphoma (MCL)²⁷⁷, melanoma²⁷⁸⁻²⁷⁹, gastric cancer²⁸⁰, myxofibrosarcoma²⁸¹, nasopharyngeal carcinoma²⁸², ovarian carcinoma²⁷³ and non-small cell lung cancer²⁸³. MTAP loss was not prognostic in pediatric B-cell acute lymphocytic leukemia²⁸⁴ or in astrocytoma²⁸⁵. However, MTAP has also

been reported to be overexpressed in colorectal cancer (CRC) samples²⁸⁶, and MTAP retention is thought to be important for prostate cancer growth due to continuous supply of SAM²⁸⁷. Germline SNPs in MTAP have been correlated with the development of cutaneous melanoma²⁸⁸⁻²⁸⁹, esophageal cancer²⁹⁰⁻²⁹¹, osteosarcoma²⁹², and CRC²⁹³.

FINDING SUMMARY

MTAP encodes S-methyl-5'-thioadenosine (MTA) phosphorylase, a tumor suppressor involved in polyamine metabolism and methionine synthesis, although its enzymatic function is dispensable for its tumor suppressor activity²⁹⁴⁻²⁹⁵. Decreased expression of MTAP leads to MTA accumulation within tumor cells and their microenvironment^{275,296-297}, thereby reducing intracellular arginine methylation^{253,255,298} and altering cell signaling^{297,299}. MTAP is located at 9p21, adjacent to CDKN2A and CDKN2B, with which it is frequently co-deleted in various cancers. Other alterations in MTAP are rare and have not been extensively characterized.

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Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1326253-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Everolimus

Assay findings association

PIK3CA
E545K

AREAS OF THERAPEUTIC USE

Everolimus is an orally available mTOR inhibitor that is FDA approved to treat renal cell carcinoma (RCC) following antiangiogenic therapy; pancreatic neuroendocrine tumors; and well-differentiated non-functional neuroendocrine tumors of the lung or gastrointestinal tract. Everolimus is also approved to treat either renal angiomyolipoma or subependymal giant cell astrocytoma in association with tuberous sclerosis complex (TSC). Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical evidence⁸⁹⁻⁹⁶, PIK3CA activating mutations may predict sensitivity to mTOR inhibitors such as everolimus and temsirolimus. Studies have reported modest activity of these therapies as single agents (ORRs of 0-4%), but improved activity has been observed when they are combined with other agents such as bevacizumab and doxorubicin (ORRs of 25-44%), for patients with PIK3CA-mutated solid tumors^{93-96,300-304}.

SUPPORTING DATA

Case reports have described 2 children with PIK3CA-mutated diffuse glioma or glioneuronal tumor who benefited from treatment with everolimus alone or in combination with temozolomide³⁰⁵⁻³⁰⁶, and 1 adult with glioblastoma (GBM) harboring PIK3CA mutation and KRAS amplification who experienced disease progression

with single-agent everolimus³⁰⁷. A Phase 2 trial of radiotherapy (RT), temozolomide (TMZ), and bevacizumab followed by everolimus and bevacizumab reported that 61% (31/51) of patients with newly diagnosed glioblastoma had objective responses with a median progression-free survival (PFS) of 11.3 months and median overall survival (OS) of 13.9 months³⁰⁸. A Phase 2 study of everolimus combined with TMZ and RT for the treatment of newly diagnosed glioblastoma reported a median PFS of 6.4 months and median OS of 15.8 months³⁰⁹. A Phase 1 trial of everolimus plus TMZ for patients with newly diagnosed or progressive glioblastoma reported partial responses (PR) in 11% (3/28) and stable disease (SD) in 57% (16/28) of cases³¹⁰. A pilot study of everolimus with gefitinib in patients with recurrent glioblastoma reported 14% (3/22) PRs, 36% (8/22) SDs, and median PFS and OS of 2.6 months and 5.8 months, respectively³¹¹. Everolimus treatment achieved SD in 45% (5/11) of pediatric patients with heavily pretreated low-grade CNS tumors; median PFS of these responses was 14 months³¹². Whereas frequent adverse events precluded a recommended Phase 2 dose and schedule for the combination of trametinib and everolimus in a Phase 1b trial for solid tumors³¹³, a retrospective study for heavily pretreated patients with solid tumors reported tolerable regimens of the combination for 23/31 patients, with 16 patients treated >3 months and evaluable patients achieving a median PFS of 6.5 months³¹⁴.

Infigratinib

Assay findings association

FGFR1
K656E

AREAS OF THERAPEUTIC USE

Infigratinib is a TKI that inhibits FGFR1, FGFR2, and FGFR3. It is FDA approved for the treatment of patients with unresectable locally advanced or metastatic cholangiocarcinoma who have FGFR2 rearrangements or fusions and have progressed after prior therapy. Please see the drug label for full prescribing information.

GENE ASSOCIATION

Based on individual responses in patients with FGFR1-mutated glioblastoma⁶⁸ and lung squamous cell carcinoma⁶⁷, FGFR1 mutation may predict sensitivity to

infigratinib.

SUPPORTING DATA

A Phase 2 study of infigratinib for patients with recurrent high-grade gliomas harboring FGFR alterations, reported a 9.5% (2/21) ORR, 1.7 month median PFS, and 6.7 month median OS⁶⁸. Disease control greater than one year was observed in 4 patients, including a PR in a patient with FGFR1-mutated glioma, and SD in patients with glioma harboring FGFR1 mutation, FGFR3 mutation, or FGFR3-TACC3 fusion⁶⁸.

ORDERED TEST # ORD-1326253-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Temsirolimus

Assay findings association
PIK3CA
E545K

AREAS OF THERAPEUTIC USE

Temsirolimus is an intravenous mTOR inhibitor that is FDA approved for the treatment of advanced renal cell carcinoma. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical evidence⁸⁹⁻⁹⁶, PIK3CA activating mutations may predict sensitivity to mTOR inhibitors such as everolimus and temsirolimus. Studies have reported modest activity of these therapies as single agents (ORRs of 0-4%), but improved activity has been observed when they are combined with other agents such as bevacizumab and doxorubicin (ORRs of 25-44%), for patients with PIK3CA-mutated solid tumors^{93-96,300-304}.

SUPPORTING DATA

A Phase 1, dose-escalation trial combining temsirolimus and radiation/temozolomide therapy, with or without adjuvant temozolomide monotherapy, in patients with

newly diagnosed glioblastoma reported no clinical responses but 24/25 patients experienced a period of stable disease; increased infection rates were noted with this regimen³¹⁵. A Phase 1/2 trial of temsirolimus in combination with sorafenib in glioblastoma was terminated at the Phase 2 interim analysis after patients failed to meet the primary endpoint of 6 month progression-free survival; significant toxicity was also observed in the combination therapy, even at low doses of temsirolimus³¹⁶. A Phase 2 study showed that addition of temsirolimus to bevacizumab therapy in patients with recurrent glioblastoma did not add clinical benefit³¹⁷. A Phase 2 clinical trial of temsirolimus in pediatric glioma reported disease stabilization in 7/17 patients including one patient with anaplastic astrocytoma³¹⁸. A Phase 1/2 study of temsirolimus in combination with erlotinib reported 6% (1/16) complete responses, 6% (1/16) partial responses, and 12.5% (2/16) instances of stable disease in patients with anaplastic glioma³¹⁹.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

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ORDERED TEST # ORD-1326253-01

CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov. Or visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

GENE
CDK4
RATIONALE
CDK4 amplification may predict sensitivity to

CDK4/6 inhibitors.

ALTERATION
amplification

NCT03239015
PHASE 2

Efficacy and Safety of Targeted Precision Therapy in Refractory Tumor With Druggable Molecular Event

TARGETS
EGFR, ERBB4, ERBB2, PARP, mTOR, MET, ROS1, RET, VEGFRs, BRAF, CDK4, CDK6

LOCATIONS: Shanghai (China)

NCT04282031
PHASE 1/2

A Study of BPI-1178 in Patients With Advanced Solid Tumor and HR+/HER2- Breast Cancer

TARGETS
CDK6, CDK4, ER, Aromatase

LOCATIONS: Shanghai (China)

NCT04594005
PHASE 1/2

CDK4/6 Tumor, Abemaciclib, Paclitaxel

TARGETS
CDK4, CDK6

LOCATIONS: Seoul (Korea, Republic of)

NCT02933736
PHASE NULL

Ribociclib (LEE011) in Preoperative Glioma and Meningioma Patients

TARGETS
CDK6, CDK4

LOCATIONS: Arizona

NCT04801966
PHASE NULL

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

TARGETS
CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF

LOCATIONS: Melbourne (Australia)

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CLINICAL TRIALS
NCT03994796
PHASE 2

Genetic Testing in Guiding Treatment for Patients With Brain Metastases

TARGETS

TRKB, ALK, TRKC, ROS1, TRKA, CDK4, CDK6, PI3K, mTOR

LOCATIONS: Washington, Oregon, Idaho, Montana

NCT04116541
PHASE 2

A Study Evaluating the Activity of Anti-cancer Treatments Targeting Tumor Molecular Alterations/ Characteristics in Advanced / Metastatic Tumors.

TARGETS

CDK6, CDK4, MDM2, MET, ROS1, RET, VEGFRs

LOCATIONS: Nice (France), Lyon (France), Marseille (France), Toulouse (France), Bordeaux (France)

NCT02981940
PHASE 2

A Study of Abemaciclib in Recurrent Glioblastoma

TARGETS

CDK4, CDK6

LOCATIONS: Utah, California, Massachusetts

NCT02896335
PHASE 2

Palbociclib In Progressive Brain Metastases

TARGETS

CDK4, CDK6

LOCATIONS: Massachusetts

NCT03310879
PHASE 2

Study of the CDK4/6 Inhibitor Abemaciclib in Solid Tumors Harboring Genetic Alterations in Genes Encoding D-type Cyclins or Amplification of CDK4 or CDK6

TARGETS

CDK4, CDK6

LOCATIONS: Massachusetts

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CLINICAL TRIALS
GENE
FGFR1
RATIONALE

FGFR inhibitors may be relevant in tumors with alterations that activate FGFR1.

ALTERATION
K656E
NCT04169672
PHASE 2

Study of Surufatinib Combined With Toripalimab in Patients With Advanced Solid Tumors

TARGETS

FGFR1, CSF1R, VEGFRs, PD-1

LOCATIONS: Shanghai (China), Beijing (China)

NCT04977453
PHASE 1/2

GI-101 as a Single Agent or in Combination With Pembrolizumab, Lenvatinib or Local Radiotherapy in Advanced Solid Tumors

TARGETS

FGFRs, RET, PDGFRA, VEGFRs, KIT, PD-1, CTLA-4

LOCATIONS: Suwon-si (Korea, Republic of), Seoul (Korea, Republic of)

NCT04803318
PHASE 2

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

TARGETS

mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK

LOCATIONS: Guangzhou (China)

NCT03564691
PHASE 1

Study of MK-4830 as Monotherapy and in Combination With Pembrolizumab (MK-3475) in Participants With Advanced Solid Tumors (MK-4830-001)

TARGETS

ITL4, FGFRs, RET, PDGFRA, VEGFRs, KIT, PD-1

LOCATIONS: Seoul (Korea, Republic of), Petah Tikva (Israel), Ramat Gan (Israel), Tel Aviv (Israel), Haifa (Israel), Warszawa (Poland), Gdansk (Poland), Heraklion (Greece), Washington, Hospitalet de Llobregat (Spain)

NCT03547037
PHASE 1

A Study to Evaluate the Safety, Pharmacokinetics, Pharmacodynamics, and Immunogenicity of JNJ-63723283, an Anti-Programmed Cell Death (PD)-1 Monoclonal Antibody, as Monotherapy or in Combination With Erdafitinib in Japanese Participants With Advanced Solid Cancers

TARGETS

PD-1, FGFRs

LOCATIONS: Chuo-Ku (Japan), Kashiwa (Japan)

NCT04008797
PHASE 1

A Study of E7386 in Combination With Other Anticancer Drug in Participants With Solid Tumor

TARGETS

CBP, Beta-catenin, FGFRs, RET, PDGFRA, VEGFRs, KIT

LOCATIONS: Osakasayama (Japan), Chuo-Ku (Japan), Kashiwa (Japan)

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ORDERED TEST # ORD-1326253-01

CLINICAL TRIALS

NCT04424966

PHASE NULL

Infigratinib in Recurrent Glioblastoma Patients

TARGETS
FGFR3, FGFR1, FGFR2

LOCATIONS: Arizona

NCT04565275

PHASE 1/2

A Study of ICP-192 in Patients With Advanced Solid Tumors

TARGETS
FGFR2, FGFR1, FGFR3, FGFR4

LOCATIONS: Macquarie Park (Australia), Melbourne (Australia), Clayton (Australia), Frankston (Australia), Colorado, Minnesota, Arizona, Ohio, Florida

NCT02549937

PHASE 1/2

A Multi-Center, Open-Label Study of Sulfatinib(HMPL-012) in Patients With Advanced Solid Tumors

TARGETS
FGFR1, CSF1R, VEGFRs

LOCATIONS: Milano (Italy), California, Colorado, Texas, New York, Tennessee, Virginia, Florida

NCT04965818

PHASE 1/2

Phase 1b/2 Study of Futibatinib in Combination With Binimetinib in Patients With Advanced KRAS Mutant Cancer

TARGETS
MEK, FGFRs

LOCATIONS: California, Indiana, Texas

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CLINICAL TRIALS
GENE
PIK3CA
ALTERATION
E545K
RATIONALE

PIK3CA activating mutations may lead to activation of the PI3K-AKT-mTOR pathway and may therefore indicate sensitivity to inhibitors of

this pathway. Strong clinical data support sensitivity of PIK3CA-mutated solid tumors to the PI3K-alpha inhibitor alpelisib.

NCT04589845
PHASE 2

Tumor-Agnostic Precision Immuno-Oncology and Somatic Targeting Rational for You (TAPISTRY) Platform Study

TARGETS

TRKB, ALK, TRKC, ROS1, TRKA, RET, PD-L1, AKTs, ERBB2, MDM2, PI3K-alpha

LOCATIONS: Zhongzheng Dist. (Taiwan), Taipei City (Taiwan), Taoyuan County (Taiwan), Tainan (Taiwan), Hong Kong (Hong Kong), Seoul (Korea, Republic of), Xi'an (China), Tianjin (China), Beijing (China), Chengdu City (China)

NCT03239015
PHASE 2

Efficacy and Safety of Targeted Precision Therapy in Refractory Tumor With Druggable Molecular Event

TARGETS

EGFR, ERBB4, ERBB2, PARP, mTOR, MET, ROS1, RET, VEGFRs, BRAF, CDK4, CDK6

LOCATIONS: Shanghai (China)

NCT04337463
PHASE NULL

ATG-008 Combined With Toripalimab in Advanced Solid Tumors

TARGETS

mTORC1, mTORC2, PD-1

LOCATIONS: Chongqing (China), Chengdu (China)

NCT04803318
PHASE 2

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

TARGETS

mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK

LOCATIONS: Guangzhou (China)

NCT03772561
PHASE 1

Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies

TARGETS

PARP, AKTs, PD-L1

LOCATIONS: Singapore (Singapore)

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CLINICAL TRIALS
NCT04801966
PHASE NULL

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

TARGETS

CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF

LOCATIONS: Melbourne (Australia)

NCT04632992
PHASE 2

A Study Evaluating Targeted Therapies in Participants Who Have Advanced Solid Tumors With Genomic Alterations or Protein Expression Patterns Predictive of Response

TARGETS

TRKB, ALK, TRKC, ROS1, TRKA, PD-L1, ERBB2, PI3K-alpha, RET, AKTs

LOCATIONS: Alaska, Washington, Oregon, California, Idaho

NCT03994796
PHASE 2

Genetic Testing in Guiding Treatment for Patients With Brain Metastases

TARGETS

TRKB, ALK, TRKC, ROS1, TRKA, CDK4, CDK6, PI3K, mTOR

LOCATIONS: Washington, Oregon, Idaho, Montana

NCT03711058
PHASE 1/2

Study of PI3Kinase Inhibition (Copanlisib) and Anti-PD-1 Antibody Nivolumab in Relapsed/Refractory Solid Tumors With Expansions in Mismatch-repair Proficient (MSS) Colorectal Cancer

TARGETS

PD-1, PI3K

LOCATIONS: Maryland

NCT03006172
PHASE 1

To Evaluate the Safety, Tolerability, and Pharmacokinetics of GDC-0077 Single Agent in Participants With Solid Tumors and in Combination With Endocrine and Targeted Therapies in Participants With Breast Cancer

TARGETS

PI3K-alpha, Aromatase, ER, CDK6, CDK4

LOCATIONS: London (United Kingdom), Surrey (United Kingdom), Bordeaux (France), Barcelona (Spain), Valencia (Spain), Toronto (Canada), Massachusetts, New York, Tennessee

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APPENDIX
Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

BCORL1
V872G

CARD11
S694L

CBL
H42_L43insH

DOT1L
G1087S

ERBB2
V541M

GNA13
rearrangement and
rearrangement

STK11
F354L

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APPENDIX
Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTK	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL	CCND1
CCND2	CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2	DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FAM46C	FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3
FGFR4	FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3
GATA4	GATA6	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3F3A
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDMSA	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKKN1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD3 (WHSC1L1)	NTSC2	NTRK1	NTRK2	NTRK3	P2RY8	PALB2
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA
PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET
RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2
SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOC3
SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU
SYK	TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53
TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1	WT1	XPO1
XRCC2	ZNF217	ZNF703						

DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV1
ETV4	ETVS	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT
KMT2A (MLL)	MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA
RAF1	RARA	RET	ROS1	RSPO2	SDC4	SLC34A2	TERC*	TERT**
TPRSS2								

*TERC is an NCRNA

**Promoter region of TERT is interrogated

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Loss of Heterozygosity (LOH) score

Microsatellite (MS) status

Tumor Mutational Burden (TMB)

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About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.



ABOUT FOUNDATIONONE CDx

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:
www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLES

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials

Ranking of Therapies in Summary Table

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

1. In the fractional-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-

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- Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.
- TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
 - The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
 - Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
 - Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine

whether the patient is a candidate for biopsy.

- Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of *HER2* overexpression as 20% in breast cancer. Based on the F1CDx *HER2* CDx concordance study, approximately 10% of *HER2* amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

*Interquartile Range = 1st Quartile to 3rd Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 291656669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear

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cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

MR Suite Version 6.1.0

The median exon coverage for this sample is 554x

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APPENDIX **References**

1. Histopathology (2007) PMID: 17204026
2. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
3. Overman MJ, et al. Lancet Oncol. (2017) PMID: 28734759
4. Overman MJ, et al. J. Clin. Oncol. (2018) PMID: 29355075
5. Lipson EJ, et al. Clin. Cancer Res. (2013) PMID: 23169436
6. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
7. Rizvi NA, et al. Science (2015) PMID: 25765070
8. Oaknin A, et al. JAMA Oncol (2020) PMID: 33001143
9. Hochster et al., 2017; ASCO Abstract 673
10. Fleming et al., 2018; ASCO Abstract 5585
11. Bang et al., 2018; ASCO Abstract 92
12. Alonso M, et al. Cancer Res. (2001) PMID: 11280776
13. Rodríguez-Hernández I, et al. PLoS ONE (2013) PMID: 24073290
14. Vladimirova V, et al. Neuropathol. Appl. Neurobiol. (2008) PMID: 18053027
15. Martinez R, et al. Oncology (2004) PMID: 15331927
16. Martinez R, et al. J. Cancer Res. Clin. Oncol. (2005) PMID: 15672285
17. Martinez R, et al. Cancer Genet. Cytogenet. (2007) PMID: 17498554
18. Szybka M, et al. Clin. Neuropathol. () PMID: 12908754
19. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
20. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
21. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
22. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
23. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
24. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
25. Cristescu R, et al. Science (2018) PMID: 30309915
26. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
27. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
28. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
29. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
30. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
31. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
32. Zhao J, et al. Nat. Med. (2019) PMID: 30742119
33. Touat M, et al. Nature (2020) PMID: 32322066
34. Bouffet E, et al. J. Clin. Oncol. (2016) PMID: 27001570
35. Johanns TM, et al. Cancer Discov (2016) PMID: 27683556
36. Lukas RV, et al. J. Neurooncol. (2018) PMID: 30073642
37. Chalmers ZR, et al. Genome Med (2017) PMID: 28420421
38. Patel RR, et al. Pediatr Blood Cancer (2020) PMID: 32386112
39. Johnson A, et al. Oncologist (2017) PMID: 28912153
40. Draaisma K, et al. Acta Neuropathol Commun (2015) PMID: 26699864
41. Wang L, et al. BMC Cancer (2020) PMID: 32164609
42. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
43. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
44. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
45. Johnson BE, et al. Science (2014) PMID: 24336570
46. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
47. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
48. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
49. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
50. Nature (2012) PMID: 22810696
51. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
52. Loriot Y, et al. N. Engl. J. Med. (2019) PMID: 31340094
53. Tabernero J, et al. J. Clin. Oncol. (2015) PMID: 26324363
54. Karkera JD, et al. Mol. Cancer Ther. (2017) PMID: 28416604
55. Necchi et al., 2018; ESMO Abstract 900P
56. Pal SK, et al. Cancer Discov (2018) PMID: 29848605
57. Pal SK, et al. Cancer (2020) PMID: 32208524
58. Schuler M, et al. Lancet Oncol. (2019) PMID: 31405822
59. Farouk Sait S, et al. JCO Precis Oncol (2021) PMID: 34250399
60. Voss MH, et al. Clin. Cancer Res. (2019) PMID: 30745300
61. Bahleda R, et al. Ann Oncol (2020) PMID: 32622884
62. Papadopoulos KP, et al. Br. J. Cancer (2017) PMID: 28972963
63. Cheng FT, et al. J Natl Compr Canc Netw (2017) PMID: 29223982
64. Khodadoust MS, et al. Leukemia (2016) PMID: 26055304
65. Tanasi I, et al. Blood (2019) PMID: 31434701
66. Strati P, et al. Leuk. Lymphoma (2018) PMID: 29119847
67. Slosberg ED, et al. Oncotarget (2018) PMID: 29765547
68. Lassman et al., 2019; SNO Abstract ACTR-33
69. Brennan CW, et al. Cell (2013) PMID: 24120142
70. Cancer Genome Atlas Research Network, et al. N. Engl. J. Med. (2015) PMID: 26061751
71. Ryall S, et al. Acta Neuropathol Commun (2016) PMID: 27572993
72. Jones DT, et al. Nat. Genet. (2013) PMID: 23817572
73. Rand V, et al. Proc. Natl. Acad. Sci. U.S.A. (2005) PMID: 16186508
74. Lew ED, et al. Sci Signal (2009) PMID: 19224897
75. Zhang J, et al. Nat. Genet. (2013) PMID: 23583981
76. Di Stefano AL, et al. Clin. Cancer Res. (2015) PMID: 25609060
77. Becker AP, et al. J. Neuropathol. Exp. Neurol. (2015) PMID: 26083571
78. Ahrendsen JT, et al. J Neuropathol Exp Neurol (2021) PMID: 34580728
79. Schüller U, et al. Acta Neuropathol (2021) PMID: 33433639
80. Turner N, et al. Nat. Rev. Cancer (2010) PMID: 20094046
81. Liu A, et al. Development (2003) PMID: 14602678
82. Petiot A, et al. Dev. Dyn. (2002) PMID: 12112473
83. Hart KC, et al. Oncogene (2000) PMID: 10918587
84. Fritsch C, et al. Mol. Cancer Ther. (2014) PMID: 24608574
85. Juric D, et al. J. Clin. Oncol. (2018) PMID: 29401002
86. Gallant JN, et al. NPJ Precis Oncol (2019) PMID: 30793038
87. André F, et al. N. Engl. J. Med. (2019) PMID: 31091374
88. Smyth LM, et al. NPJ Breast Cancer (2021) PMID: 33863913
89. Park HS, et al. PLoS ONE (2016) PMID: 27105424
90. Lim SM, et al. Oncotarget (2016) PMID: 26859683
91. Hou MM, et al. Oncotarget (2014) PMID: 25426553
92. Varnier R, et al. Eur J Cancer (2019) PMID: 31351267
93. Janku F, et al. Cell Rep (2014) PMID: 24440717
94. Moroney J, et al. Clin. Cancer Res. (2012) PMID: 22927482
95. Basho RK, et al. JAMA Oncol (2017) PMID: 27893038
96. Moroney JW, et al. Clin. Cancer Res. (2011) PMID: 21890452
97. Krop et al., 2018; ASCO Abstract 101
98. Pascual J, et al. Cancer Discov (2021) PMID: 32958578
99. Dolly SO, et al. Clin. Cancer Res. (2016) PMID: 26787751
100. Aust Fam Physician (1986) PMID: 2941002
101. Santin AD, et al. Gynecol Oncol Rep (2020) PMID: 31934607
102. Patnaik A, et al. Ann. Oncol. (2016) PMID: 27672108
103. El-Habr EA, et al. Clin. Neuropathol. () PMID: 20569675
104. Gallia GL, et al. Mol. Cancer Res. (2006) PMID: 17050665
105. Broderick DK, et al. Cancer Res. (2004) PMID: 15289301
106. Derakhshandeh-Peykar P, et al. J. Neurogenet. (2011) PMID: 22026810
107. Nature (2008) PMID: 18772890
108. Tanaka S, et al. Acta Neuropathol Commun (2019) PMID: 31036078
109. Yan et al. 2020; DOI:10.1200/P0.19.00385
110. Samuels Y, et al. Cancer Cell (2005) PMID: 15950905
111. Nat. Rev. Cancer (2009) PMID: 19629070
112. Kang S, et al. Proc. Natl. Acad. Sci. U.S.A. (2005) PMID: 15647370
113. Ikenoue T, et al. Cancer Res. (2005) PMID: 15930273
114. Gymnopoulos M, et al. Proc. Natl. Acad. Sci. U.S.A. (2007) PMID: 17376864
115. Horn S, et al. Oncogene (2008) PMID: 18317450
116. Rudd ML, et al. Clin. Cancer Res. (2011) PMID: 21266528
117. Hon WC, et al. Oncogene (2012) PMID: 22120714
118. Burke JE, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) PMID: 22949682
119. Wu H, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) PMID: 19915146
120. Laurenti R, et al. Rev Saude Publica (1990) PMID: 2103068
121. Dan S, et al. Cancer Res. (2010) PMID: 20530683
122. Oda K, et al. Cancer Res. (2008) PMID: 18829572
123. Zhao L, et al. Oncogene (2008) PMID: 18794883
124. Lui VW, et al. Cancer Discov (2013) PMID: 23619167
125. Ross RL, et al. Oncogene (2013) PMID: 22430209
126. Rivière JB, et al. Nat. Genet. (2012) PMID: 22729224
127. Shibata T, et al. Cancer Lett. (2009) PMID: 19394761
128. Dogruluk T, et al. Cancer Res. (2015) PMID: 26627007
129. Croessmann S, et al. Clin. Cancer Res. (2018) PMID: 29284706
130. Ng PK, et al. Cancer Cell (2018) PMID: 29533785
131. Spangle JM, et al. (2020) PMID: 32929011
132. Chen L, et al. Nat Commun (2018) PMID: 29636477
133. Jin N, et al. J Clin Invest (2021) PMID: 34779417
134. Dickson MA, et al. J. Clin. Oncol. (2013) PMID: 23569312
135. Flaherty KT, et al. Clin. Cancer Res. (2012) PMID: 22090362
136. Patnaik A, et al. Cancer Discov (2016) PMID: 27217383
137. Infante JR, et al. Clin. Cancer Res. (2016) PMID: 27542767
138. Dickson et al., 2019; ASCO Abstract 11004
139. Dickson MA, et al. JAMA Oncol (2016) PMID: 27124835
140. Peguero et al., 2016; ASCO Abstract 2528
141. Cerami E, et al. Cancer Discov (2012) PMID: 22588877
142. Gao J, et al. Sci Signal (2013) PMID: 23550210
143. Jonsson P, et al. Clin. Cancer Res. (2019) PMID: 31263031
144. Ceccarelli M, et al. Cell (2016) PMID: 26824661
145. Zheng S, et al. Genes Dev. (2013) PMID: 23796897
146. Kim H, et al. Proc. Natl. Acad. Sci. U.S.A. (2010) PMID: 20080666
147. Ruano Y, et al. Am. J. Clin. Pathol. (2009) PMID: 19141386
148. Fischer U, et al. Mol. Cancer Res. (2008) PMID: 18403636

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ORDERED TEST # **ORD-1326253-01**
APPENDIX **References**

149. Bäcklund LM, et al. Br. J. Cancer (2005) PMID: 15970925
150. Choi YJ, et al. Oncogene (2014) PMID: 23644662
151. Cell (1995) PMID: 7736585
152. Musgrove EA, et al. Nat. Rev. Cancer (2011) PMID: 21734724
153. Wikman H, et al. Genes Chromosomes Cancer (2005) PMID: 15543620
154. Rao SK, et al. J. Neurooncol. (2010) PMID: 19609742
155. Chung L, et al. Am. J. Surg. Pathol. (2009) PMID: 19574885
156. Ragazzini P, et al. Histol. Histopathol. (2004) PMID: 15024701
157. Dujardin F, et al. Mod. Pathol. (2011) PMID: 21336260
158. Zhang K, et al. Cancer Res. (2013) PMID: 23393200
159. Horvai AE, et al. Mod. Pathol. (2009) PMID: 19734852
160. Liang J, et al. Cancer Res. (2020) PMID: 31551363
161. Garbarino J, et al. Transl Oncol (2021) PMID: 34118569
162. Cole et al., 2021; AACR Abstract CT059
163. George SL, et al. EBioMedicine (2020) PMID: 32846370
164. Koschmann C, et al. Sci Transl Med (2016) PMID: 26936505
165. Heaphy CM, et al. Science (2011) PMID: 21719641
166. Singhi et al., 2015; USCAP Abstract 1797
167. Jiao Y, et al. Science (2011) PMID: 21252315
168. Fishbein L, et al. Nat Commun (2015) PMID: 25608029
169. Morosini et al., 2014; ASCO Abstract 11008
170. Cheung NK, et al. JAMA (2012) PMID: 22416102
171. Molenaar JJ, et al. Nature (2012) PMID: 22367537
172. Pugh TJ, et al. Nat. Genet. (2013) PMID: 23334666
173. Cheung NK, et al. Nat. Rev. Cancer (2013) PMID: 23702928
174. Marinoni I, et al. Gastroenterology (2014) PMID: 24148618
175. Qadeer ZA, et al. J. Invest. Dermatol. (2014) PMID: 24468746
176. Kannan K, et al. Oncotarget (2012) PMID: 23104868
177. Haberler C, et al. Clin. Neuropathol. (2015) PMID: 24559763
178. Reuss DE, et al. Acta Neuropathol. (2015) PMID: 25427834
179. Sahm F, et al. Acta Neuropathol. (2014) PMID: 25143301
180. Singhi et al., 2015; USCAP Abstract 93
181. Liao JY, et al. Am. J. Surg. Pathol. (2015) PMID: 25229770
182. Clynes D, et al. Trends Biochem. Sci. (2013) PMID: 23916100
183. Ratnakumar K, et al. Epigenetics (2013) PMID: 23249563
184. Lovejoy CA, et al. PLoS Genet. (2012) PMID: 22829774
185. Bower K, et al. PLoS ONE (2012) PMID: 23185534
186. Nan X, et al. Proc. Natl. Acad. Sci. U.S.A. (2007) PMID: 17296936
187. Garrick D, et al. Gene (2004) PMID: 14729260
188. Eustermann S, et al. Nat. Struct. Mol. Biol. (2011) PMID: 21666677
189. Flynn RL, et al. Science (2015) PMID: 25593184
190. Gibbons RJ, et al. Cell (1995) PMID: 7697714
191. Konecny GE, et al. Clin. Cancer Res. (2011) PMID: 21278246
192. Katsumi Y, et al. Biochem. Biophys. Res. Commun. (2011) PMID: 21871868
193. Cen L, et al. Neuro-oncology (2012) PMID: 22711607
194. Logan JE, et al. Anticancer Res. (2013) PMID: 23898052
195. Elvin JA, et al. Oncologist (2017) PMID: 28283584
196. Gao J, et al. Curr Oncol (2015) PMID: 26715889
197. Gopalan et al., 2014; ASCO Abstract 8077
198. Konecny et al., 2016; ASCO Abstract 5557
199. DeMichele A, et al. Clin. Cancer Res. (2015) PMID: 25501126
200. Finn RS, et al. Lancet Oncol. (2015) PMID: 25524798
201. Johnson DB, et al. Oncologist (2014) PMID: 24797823
202. Van Maerken T, et al. Mol. Cancer Ther. (2011) PMID: 21460101
203. Gamble LD, et al. Oncogene (2012) PMID: 21725357
204. Shapiro et al., 2013; ASCO Abstract 2500
205. Verhaak RG, et al. Cancer Cell (2010) PMID: 20129251
206. Sottoriva A, et al. Proc. Natl. Acad. Sci. U.S.A. (2013) PMID: 23412337
207. Weber RG, et al. Oncogene (2007) PMID: 16909113
208. Chakravarti A, et al. Clin. Cancer Res. (2001) PMID: 11489817
209. Feng J, et al. Cancer (2012) PMID: 21713760
210. Raabe EH, et al. Clin. Cancer Res. (2011) PMID: 21636552
211. Liu W, et al. J. Exp. Clin. Cancer Res. (2011) PMID: 21843312
212. Quelle DE, et al. Cell (1995) PMID: 8521522
213. Mutat. Res. (2005) PMID: 15878778
214. Gazzeri S, et al. Oncogene (1998) PMID: 9484839
215. Oncogene (1999) PMID: 10498883
216. Sherr CJ, et al. Cold Spring Harb. Symp. Quant. Biol. (2005) PMID: 16869746
217. Ozenne P, et al. Int. J. Cancer (2010) PMID: 20549699
218. Ruas M, et al. Oncogene (1999) PMID: 10498896
219. Jones R, et al. Cancer Res. (2007) PMID: 17909018
220. Haferkamp S, et al. Aging Cell (2008) PMID: 18843795
221. Huot TJ, et al. Mol. Cell. Biol. (2002) PMID: 12417717
222. Rizo H, et al. J. Biol. Chem. (2001) PMID: 11518711
223. Gombart AF, et al. Leukemia (1997) PMID: 9324288
224. Yang R, et al. Cancer Res. (1995) PMID: 7780957
225. Parry D, et al. Mol. Cell. Biol. (1996) PMID: 8668202
226. Greenblatt MS, et al. Oncogene (2003) PMID: 12606942
227. Yarbrough WG, et al. J. Natl. Cancer Inst. (1999) PMID: 10491434
228. Poi MJ, et al. Mol. Carcinog. (2001) PMID: 11255261
229. Byeon IJ, et al. Mol. Cell (1998) PMID: 9660926
230. Kannengiesser C, et al. Hum. Mutat. (2009) PMID: 19260062
231. Lal G, et al. Genes Chromosomes Cancer (2000) PMID: 10719365
232. Koh J, et al. Nature (1995) PMID: 7777061
233. McKenzie HA, et al. Hum. Mutat. (2010) PMID: 20340136
234. Miller PJ, et al. Hum. Mutat. (2011) PMID: 21462282
235. Kutscher CL, et al. Physiol. Behav. (1977) PMID: 905385
236. Scaini MC, et al. Hum. Mutat. (2014) PMID: 24659262
237. Jenkins NC, et al. J. Invest. Dermatol. (2013) PMID: 23190892
238. Walker GJ, et al. Int. J. Cancer (1999) PMID: 10389768
239. Rutter JL, et al. Oncogene (2003) PMID: 12853981
240. Itahana K, et al. Cancer Cell (2008) PMID: 18538737
241. Zhang Y, et al. Mol. Cell (1999) PMID: 10360174
242. Zhang Y, et al. Cell (1998) PMID: 9529249
243. Jafri M, et al. Cancer Discov (2015) PMID: 25873077
244. Whelan AJ, et al. N Engl J Med (1995) PMID: 7666917
245. Adv Exp Med Biol (2010) PMID: 20687502
246. Hogg D, et al. J Cutan Med Surg (1998) PMID: 9479083
247. De Unamuno B, et al. Melanoma Res (2018) PMID: 29543703
248. Soura E, et al. J Am Acad Dermatol (2016) PMID: 26892650
249. Huerta C, et al. Acta Derm Venereol (2018) PMID: 29405243
250. Kaufman DK, et al. Neurology (1993) PMID: 8414022
251. Bahuau M, et al. Cancer Res (1998) PMID: 9622062
252. Chan AK, et al. Clin Neuropathol (2019) PMID: 28699883
253. Marjon K, et al. Cell Rep (2016) PMID: 27068473
254. Heist et al., 2019; AACR-NCI-EORTC Abstract B116
255. Mavrikis KJ, et al. Science (2016) PMID: 26912361
256. Endoscopy (1989) PMID: 2691236
257. Guccione E, et al. Nat. Rev. Mol. Cell Biol. (2019) PMID: 31350521
258. Fedorow A, et al. Cancer Cell (2019) PMID: 31257072
259. Srour N, et al. Cancer Cell (2019) PMID: 31287990
260. Gao G, et al. Nucleic Acids Res. (2019) PMID: 30916320
261. Hansen LJ, et al. Cancer Res. (2019) PMID: 31040154
262. Tang B, et al. Cancer Res. (2018) PMID: 29844120
263. Munshi PN, et al. Oncologist (2014) PMID: 24928612
264. de Oliveira SF, et al. PLoS ONE (2016) PMID: 26751376
265. Lubin M, et al. PLoS ONE (2009) PMID: 19478948
266. Tang B, et al. Cancer Biol. Ther. (2012) PMID: 22825330
267. Collins CC, et al. Mol. Cancer Ther. (2012) PMID: 22252602
268. Bertino JR, et al. Cancer Biol. Ther. (2011) PMID: 21301207
269. Coulthard SA, et al. Mol. Cancer Ther. (2011) PMID: 21282358
270. Miyazaki S, et al. Int. J. Oncol. (2007) PMID: 17912432
271. Efferth T, et al. Blood Cells Mol. Dis. (2019) PMID: 11987241
272. Kindler HL, et al. Invest New Drugs (2009) PMID: 18618081
273. Wei R, et al. Sci Rep (2016) PMID: 27929028
274. Zhao M, et al. BMC Genomics (2016) PMID: 27556634
275. Kirovski G, et al. Am. J. Pathol. (2011) PMID: 21356366
276. Huang HY, et al. Clin. Cancer Res. (2009) PMID: 19887491
277. Marcé S, et al. Clin. Cancer Res. (2006) PMID: 16778103
278. Meyer S, et al. Exp. Dermatol. (2010) PMID: 20500769
279. Wild PJ, et al. Arch Dermatol (2006) PMID: 16618867
280. Kim J, et al. Genes Chromosomes Cancer (2011) PMID: 21412930
281. Li CF, et al. Oncotarget (2014) PMID: 25426549
282. He HL, et al. Medicine (Baltimore) (2015) PMID: 26656376
283. Su CY, et al. Eur J Surg Oncol (2014) PMID: 24969958
284. Mirebeau D, et al. Haematologica (2006) PMID: 16818274
285. Becker AP, et al. Pathobiology (2015) PMID: 26088413
286. Snezhkina AV, et al. Oxid Med Cell Longev (2016) PMID: 27433286
287. Bistulfi G, et al. Oncotarget (2016) PMID: 26910893
288. Antonopoulou K, et al. J. Invest. Dermatol. (2015) PMID: 25407435
289. Maccioni L, et al. BMC Cancer (2013) PMID: 23816148
290. Hyland PL, et al. Int J Epidemiol (2016) PMID: 26635288
291. Lin X, et al. Cancer Sci. (2017) PMID: 27960044
292. Zhi L, et al. J Cancer (2016) PMID: 27994653
293. Gu F, et al. Br. J. Cancer (2013) PMID: 23361049
294. Limm K, et al. PLoS ONE (2016) PMID: 27479139
295. Tang B, et al. G3 (Bethesda) (2014) PMID: 25387827
296. Limm K, et al. Eur. J. Cancer (2013) PMID: 23265702
297. Stevens AP, et al. J. Cell. Biochem. (2009) PMID: 19097084
298. Kryukov GV, et al. Science (2016) PMID: 26912360
299. Limm K, et al. Eur. J. Cancer (2014) PMID: 25087184
300. Janku F, et al. Cancer Res. (2013) PMID: 23066039
301. Janku F, et al. J. Clin. Oncol. (2012) PMID: 22271473
302. Janku F, et al. Mol. Cancer Ther. (2011) PMID: 21216929
303. Moulder S, et al. Ann. Oncol. (2015) PMID: 25878190

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APPENDIX
References

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| <p>304. Byeon et al., 2020; doi: 10.21037/tcr.2020.04.07</p> <p>305. Gojo J, et al. Front Oncol (2019) PMID: 31998633</p> <p>306. McNall-Knapp et al., 2020; SNO Abstract LGG-47</p> <p>307. Blumenthal DT, et al. J. Neurooncol. (2016) PMID: 27531351</p> <p>308. Hainsworth JD, et al. Clin Adv Hematol Oncol (2012) PMID: 22706484</p> | <p>309. Ma DJ, et al. Neuro-oncology (2015) PMID: 25526733</p> <p>310. Mason WP, et al. Invest New Drugs (2012) PMID: 22160854</p> <p>311. Kreisl TN, et al. J. Neurooncol. (2009) PMID: 19018475</p> <p>312. Segal et al., 2016; ISPNO Abstract EPT-21</p> <p>313. Tolcher AW, et al. Ann. Oncol. (2015) PMID: 25344362</p> <p>314. Patterson et al., 2018; AACR Abstract 3891</p> | <p>315. Sarkaria JN, et al. Clin. Cancer Res. (2010) PMID: 20921209</p> <p>316. Lee EQ, et al. Neuro-oncology (2012) PMID: 23099651</p> <p>317. Lassen U, et al. Anticancer Res. (2013) PMID: 23564811</p> <p>318. Georger B, et al. Eur. J. Cancer (2012) PMID: 22033322</p> <p>319. Wen PY, et al. Neuro-oncology (2014) PMID: 24470557</p> |
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