

**ABOUT THE TEST** FoundationOne®Liquid CDx is a next generation sequencing (NGS) assay that identifies clinically relevant genomic alterations in circulating cell-free DNA.

<b>PATIENT</b>	<b>DISEASE</b> Lung cancer (NOS)	<b>PHYSICIAN</b>	<b>ORDERING PHYSICIAN</b> Yeh, Yi-Chen	<b>SPECIMEN</b>	<b>SPECIMEN ID</b> Y-H.C. 03/30/1956
	<b>NAME</b> Chen, Yueh-Hsiang		<b>MEDICAL FACILITY</b> Taipei Veterans General Hospital		<b>SPECIMEN TYPE</b> Blood
	<b>DATE OF BIRTH</b> 30 March 1956		<b>ADDITIONAL RECIPIENT</b> None		<b>DATE OF COLLECTION</b> 27 July 2022
	<b>SEX</b> Female		<b>MEDICAL FACILITY ID</b> 205872		<b>SPECIMEN RECEIVED</b> 29 July 2022
	<b>MEDICAL RECORD #</b> 46526130		<b>PATHOLOGIST</b> Not Provided		

## Biomarker Findings

**Blood Tumor Mutational Burden** - 4 Muts/Mb

**Microsatellite status** - MSI-High Not Detected

**Tumor Fraction** - Elevated Tumor Fraction Not Detected

## Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

**EGFR** L858R

**RAD54L** C391fs\*1

**ASXL1** L386\*

**TET2** H1761fs\*5, R1261H, H222fs\*3

**TP53** A138V

## Report Highlights

- Targeted therapies with **NCCN categories of evidence** in this tumor type: Afatinib (p. 11), Dacomitinib (p. 12), Erlotinib (p. 12), Gefitinib (p. 13), Osimertinib (p. 13)
- Evidence-matched **clinical trial options** based on this patient's genomic findings: (p. 15)
- Variants that may represent **clonal hematopoiesis** and may originate from non-tumor sources: **ASXL1** L386\* (p. 8), **TET2** H1761fs\*5, H222fs\*3, R1261H (p. 9)

### BIOMARKER FINDINGS

#### Blood Tumor Mutational Burden

- 4 Muts/Mb

#### Microsatellite status

- MSI-High Not Detected

#### Tumor Fraction

- Elevated Tumor Fraction Not Detected

### THERAPY AND CLINICAL TRIAL IMPLICATIONS

**No therapies or clinical trials. See Biomarker Findings section**

**MSI-High not detected. No evidence of microsatellite instability in this sample (see Appendix section).**

Tumor fraction is considered elevated when ctDNA levels are high enough that aneuploidy can be detected. The fact that elevated tumor fraction was not detected in this specimen indicates the possibility of lower levels of ctDNA but does not compromise confidence in any reported alterations. However, in the setting of a negative liquid biopsy result, orthogonal testing of a tissue specimen should be considered if clinically indicated (see Biomarker Findings section).

### GENOMIC FINDINGS

### VAF %

**EGFR** - L858R 5.2%

10 Trials see p. 15

### THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)

Afatinib	1
Dacomitinib	1
Erlotinib	1
Gefitinib	1
Osimertinib	1

### THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)

None

☐ NCCN category

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GENOMIC FINDINGS	VAF %	THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)
<b>RAD54L - C391fs*1</b>	31.2%	None	None
10 Trials see p. 17			

☐ NCCN category

**VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS (CH)**

Genomic findings below may include nontumor somatic alterations, such as CH. The efficacy of targeting such nontumor somatic alterations is unknown. This content should be interpreted based on clinical context. Refer to appendix for additional information on CH.

**ASXL1 - L386\*** ..... p. 8 **TET2 - H1761fs\*5, R1261H, H222fs\*3** ..... p. 9

**GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS**

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

**ASXL1 - L386\*** ..... p. 8 **TP53 - A138V** ..... p. 10  
**TET2 - H1761fs\*5, R1261H, H222fs\*3** ..... p. 9

**NOTE** Genomic alterations detected may be associated with activity of certain approved therapies; however, the therapies listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and/or exhaustive. Neither the therapies nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies. Therapies contained in this report may have been approved by the US FDA or other national authorities; however, they might not have been approved in your respective country. In the appropriate clinical context, germline testing of APC, ATM, BAP1, BRCA1, BRCA2, BRIP1, CHEK2, FH, FLCN, MEN1, MLH1, MSH2, MSH6, MUTYH, NF1, NF2, PALB2, PMS2, POLE, PTEN, RAD51C, RAD51D, RB1, RET, SDHA, SDHB, SDHC, SDHD, SMAD4, STK11, TGFBR2, TP53, TSC1, TSC2, VHL, and WT1 is recommended.

Variant Allele Frequency is not applicable for copy number alterations.

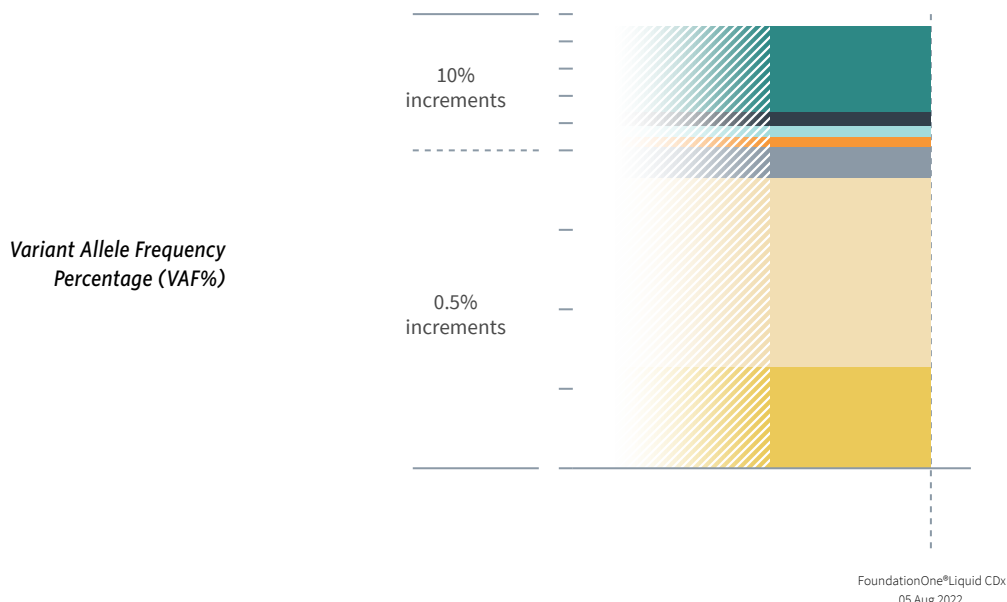
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ORDERED TEST # ORD-1423659-01



#### HISTORIC PATIENT FINDINGS

ORD-1423659-01  
VAF%

#### Blood Tumor Mutational Burden

4 Muts/Mb

#### Microsatellite status

MSI-High Not Detected

#### Tumor Fraction

Elevated Tumor Fraction Not Detected

<b>EGFR</b>	● L858R	5.2%
<b>RAD54L</b>	● C391fs*1	31.2%
<b>ASXL1</b>	● L386*	0.64%
<b>TET2</b>	● H222fs*3	4.2%
	● R1261H	1.4%
	● H1761fs*5	3.8%
<b>TP53</b>	● A138V	1.2%

**NOTE** This comparison table refers only to genes and biomarkers assayed by prior FoundationOne®Liquid CDx, FoundationOne®Liquid, FoundationOne®, or FoundationOne®CDx tests. Up to five previous tests may be shown.

For some genes in FoundationOne Liquid CDx, only select exons are assayed. Therefore, an alteration found by a previous test may not have been confirmed despite overlapping gene lists. Please refer to the Appendix for the complete list of genes and exons assayed. The gene and biomarker list will be updated periodically to reflect new knowledge about cancer biology.

As new scientific information becomes available, alterations that had previously been listed as Variants of Unknown Significance (VUS) may become reportable.

Tissue Tumor Mutational Burden (TMB) and blood TMB (bTMB) are estimated from the number of synonymous and non-synonymous single-nucleotide variants (SNVs) and insertions and deletions (indels) per area of coding genome sampled, after the removal of known and likely oncogenic driver events and germline SNPs. Tissue TMB is calculated based on variants with an allele frequency of  $\geq 5\%$ , and bTMB is calculated based on variants with an allele frequency of  $\geq 0.5\%$ .

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Not Tested = not baited, not reported on test, or test preceded addition of biomarker or gene

Not Detected = baited but not detected on test

Detected = present (VAF% is not applicable)

VAF% = variant allele frequency percentage

Cannot Be Determined = Sample is not of sufficient data quality to confidently determine biomarker status

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BIOMARKER FINDINGS

BIOMARKER

# Blood Tumor Mutational Burden

RESULT

4 Muts/Mb

## POTENTIAL TREATMENT STRATEGIES

### — Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased blood tumor mutational burden (bTMB) may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1<sup>1-3</sup>, anti-PD-1<sup>3-4</sup>, and anti-PD-1/CTLA4 therapies<sup>5-6</sup>. A Phase 2 multi-solid-tumor trial showed that bTMB  $\geq 16$  Muts/Mb (as measured by this assay) was associated with improved survival from treatment with a PD-1 inhibitor alone or in combination with a CTLA-4 inhibitor<sup>5</sup>. In non-small cell lung cancer (NSCLC), multiple clinical trials have shown patients with higher bTMB derive clinical benefit from immune checkpoint inhibitors following single-agent or combination treatments with either CTLA4 inhibitors or chemotherapy, with reported high bTMB cutpoints ranging from 6 Muts/Mb-16 Muts/Mb<sup>1</sup>. In head and neck squamous cell carcinoma (HNSCC), a Phase 3 trial showed that bTMB  $\geq 16$  Muts/Mb (approximate equivalency  $\geq 8$  Muts/Mb as measured by this assay) was associated with

improved survival from treatment with a PD-L1 inhibitor alone or in combination with a CTLA-4 inhibitor<sup>7</sup>.

## FREQUENCY & PROGNOSIS

NSCLC harbors a median bTMB of 16.8 Muts/Mb (range 1.9-52.5 Muts/Mb)<sup>4</sup>. Retrospective analysis of the Phase 3 OAK and Phase 2 POPLAR trials for patients with advanced or metastatic non-small cell lung cancer (NSCLC) reported that bTMB  $\geq 7$  Muts/Mb was associated with shorter PFS (2.8 vs. 4.2 months) and OS (7.4 vs. 11.9 months) compared with bTMB  $< 7$  Muts/Mb for patients treated with docetaxel<sup>8</sup>. In one study of advanced NSCLC in China, bTMB  $\geq 6$  Muts/Mb was associated with decreased PFS (10 vs. 18 months) and OS (11 vs. 25 months) compared with bTMB  $< 6$  Muts/Mb for patients treated with platinum-based chemotherapy<sup>9</sup>. A meta-analysis of 19 studies of immune checkpoint inhibitor-treated NSCLC (n = 2,315 patients) demonstrated that high TMB predicted a significantly longer OS than low TMB (HR = 0.70), and within the high TMB group, immunotherapy was associated with an improved PFS (HR = 0.62,  $P < 0.001$ ), OS (HR = 0.67,  $P < 0.001$ ) and a higher response rate (OR = 2.35,  $P < 0.001$ ) compared to chemotherapy<sup>10</sup>. In contrast, a large study of Chinese patients with untreated lung adenocarcinoma reported a shorter median OS for tumors with a higher number of mutations in a limited gene set compared with a lower mutation number (48.4 vs. 61.0 months)<sup>11</sup>. Another study of patients with NSCLC treated

with EGFR inhibitors or platinum doublet chemotherapy found elevated TMB to be correlated with poorer prognosis, as well as finding lower TMB in combination with PD-L1 negative status to be significantly associated with longer median survival in patients with lung adenocarcinoma<sup>12</sup>. However, no significant prognostic association of TMB and/or PD-L1 status with survival has been reported in patients with lung SCC<sup>12-13</sup>.

## FINDING SUMMARY

Blood tumor mutational burden (bTMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations from circulating tumor DNA in blood. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma<sup>14-15</sup> and cigarette smoke in lung cancer<sup>16-17</sup>, treatment with temozolomide-based chemotherapy in glioma<sup>18-19</sup>, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes<sup>20-24</sup>, and microsatellite instability (MSI)<sup>20,23-24</sup>. High bTMB levels were not detected in this sample. It is unclear whether the bTMB levels in this sample would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents<sup>1-2,4</sup>. Depending on the clinical context, TMB testing of an alternate sample or by another methodology could be considered.

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**BIOMARKER FINDINGS**
**BIOMARKER**

# Tumor Fraction

**RESULT**

Elevated Tumor Fraction Not Detected

**POTENTIAL TREATMENT STRATEGIES**
**— Targeted Therapies —**

Specimens with elevated tumor fraction values have high circulating-tumor DNA (ctDNA) content, and thus high sensitivity for identifying genomic alterations. Such specimens are at low risk of false negative results. However, if elevated tumor fraction is not detected, it does not exclude the presence of disease burden or compromise the confidence of reported alterations. Tumor fraction levels currently have limited implications for diagnosis, surveillance, or therapy and should not

be overinterpreted or compared from one blood draw to another. There are currently no targeted approaches to address specific tumor fraction levels. In the research setting, changes in tumor fraction estimates have been associated with treatment duration and clinical response and may be a useful indicator for future cancer management<sup>25-30</sup>.

**FREQUENCY & PROGNOSIS**

Detectable ctDNA levels have been reported in a variety of tumor types, with higher tumor fraction levels reported for patients with metastatic (Stage 4) tumors compared with patients with localized disease (Stages 1 to 3)<sup>31</sup>. Elevated tumor fraction levels have been reported to be associated with worse prognosis in a variety of cancer types, including pancreatic cancer<sup>32</sup>, Ewing sarcoma and osteosarcoma<sup>33</sup>, prostate cancer<sup>28</sup>, breast cancer<sup>34</sup>, leiomyosarcoma<sup>35</sup>, esophageal cancer<sup>36</sup>, colorectal

cancer<sup>37</sup>, and gastrointestinal cancer<sup>38</sup>.

**FINDING SUMMARY**

Tumor fraction provides an estimate of the percentage of ctDNA present in a cell-free DNA (cfDNA) sample. The tumor fraction estimate for this sample is based on the observed level of aneuploid instability. The tumor fraction algorithm utilized for FoundationOne Liquid CDx uses the allele frequencies of approximately 1,000 single-nucleotide polymorphism (SNP) sites across the genome. Unlike the maximum somatic allele frequency (MSAF) method of estimating ctDNA content<sup>39</sup>, the tumor fraction metric does not take into account the allele frequency of individual variants but rather produces a more holistic estimate of ctDNA content using data from across the genome. The amount of ctDNA detected may correlate with disease burden and response to therapy<sup>40-41</sup>.

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GENOMIC FINDINGS

GENE

**EGFR**

ALTERATION

L858R

TRANSCRIPT ID

NM\_005228

CODING SEQUENCE EFFECT

2573T>G

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

For patients with non-small cell lung cancer (NSCLC), EGFR activating mutations may predict sensitivity to EGFR-TKIs, including erlotinib<sup>42</sup>, gefitinib<sup>43-46</sup>, afatinib<sup>47-50</sup>, dacomitinib<sup>51</sup>, and osimertinib<sup>48,52</sup>; however, the data for patients with other tumor types are limited<sup>53-58</sup>. The Phase 1 CHRYSALIS study of amivantamab monotherapy or in combination with lazertinib for the treatment of EGFR-mutated non-small cell lung cancer (NSCLC) has produced encouraging preliminary results for treatment-naïve patients and patients who relapsed after treatment with osimertinib with and without chemotherapy, including osimertinib-relapsed patients with biomarkers indicating EGFR/MET-based osimertinib resistance<sup>59-62</sup>. In a Phase 1 trial, the HER3-targeted antibody patritumab deruxtecan elicited an ORR of 39% (22/57, 1 CR) and a median PFS of 8.2 months for patients with non-small cell lung cancer previously treated with an EGFR TKI, many of whom displayed TKI resistance

alterations<sup>63</sup>. A Phase 1 trial evaluating the EGFR inhibitor AZD3759 reported a reduction in the volume of brain metastases in 40% (8/20) of patients with previously treated non-small cell lung cancer (NSCLC) harboring either the EGFR L858R alteration or EGFR exon 19 deletion, including 3 confirmed PRs and 3 unconfirmed PRs<sup>64-65</sup>. In a Phase 1/2 trial for advanced NSCLC, the brain-penetrant third-generation EGFR TKI lazertinib enabled ORRs of 54% (69/127) for all evaluable patients and 44% (8/18, intracranial) for patients with brain metastases<sup>66</sup>. A Phase 1 trial evaluating the irreversible pan-HER inhibitor FCN-411 for NSCLC patients who had EGFR mutations and experienced disease progression on standard treatments reported an ORR of 15% with 10/67 patients achieving PR, and a DCR of 73% with 39 additional patients achieving SD<sup>67</sup>. OR was observed in a numerically higher proportion of patients with the EGFR T790M mutation than those without this mutation<sup>67</sup>.

— Nontargeted Approaches —

Patients with EGFR-mutated non-squamous metastatic non-small cell lung cancer previously treated with EGFR TKI have benefited from immune checkpoint inhibitors combined with anti-angiogenic and chemotherapy, particularly atezolizumab plus bevacizumab plus carboplatin and paclitaxel (OS HR 0.61 compared with bevacizumab/chemotherapy)<sup>68-70</sup> or sintilimab plus bevacizumab biosimilar plus cisplatin and pemetrexed (PFS HR 0.46 compared with chemotherapy alone)<sup>71</sup>. In retrospective analyses, patients with EGFR-mutated non-small cell lung

cancer that transformed to small cell lung cancer demonstrated response rates of 50% to taxane and 54% (with a median PFS of 3.4 months) to platinum-etoposide<sup>72-73</sup>.

FREQUENCY & PROGNOSIS

EGFR mutation has been reported in 12-36% of lung adenocarcinomas<sup>74-76</sup> and in 4% of lung squamous cell carcinomas<sup>77</sup>. EGFR protein expression/overexpression has been reported in up to 70% of NSCLC cases<sup>78-83</sup>. In addition, expression of EGFR protein has been shown to be higher in lung squamous cell carcinoma samples as compared to lung adenocarcinoma<sup>84-85</sup>. In patients with lung adenocarcinoma, EGFR mutation was a predictor of poor overall survival<sup>86-87</sup>. However, EGFR mutations have been reported to predict improved survival in patients with resected Stage 1-3 lung adenocarcinoma<sup>88</sup> or resected Stage 1 NSCLC<sup>89</sup>.

FINDING SUMMARY

EGFR encodes the epidermal growth factor receptor, which belongs to a class of proteins called receptor tyrosine kinases. In response to signals from the environment, EGFR passes biochemical messages to the cell that stimulate it to grow and divide<sup>90</sup>. EGFR L858 is located in the kinase domain and is encoded by exon 21. EGFR L858R has been characterized as activating<sup>91-93</sup> and patients with the L858R mutation have been shown to be sensitive to EGFR tyrosine kinase inhibitors, such as erlotinib, gefitinib<sup>91-93</sup>, and afatinib<sup>94</sup>.

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GENOMIC FINDINGS

GENE

# RAD54L

ALTERATION

C391fs\*1

TRANSCRIPT ID

NM\_003579

CODING SEQUENCE EFFECT

1092\_1093insCGAGACGCTGCTGCTAGTGAGGCAGACAGGC  
AGCTAGGAGAGAGCGGCTGCGGGAGCTCACCAGCATTGT  
GAATAGGTAATGACCTTAAGC

cancer<sup>95</sup> and prostate cancer<sup>96</sup> indicates that RAD54L inactivation may confer sensitivity to PARP inhibitors.

## FREQUENCY & PROGNOSIS

RAD54L alterations are rare in cancer<sup>97</sup>. Loss of heterozygosity (LOH) at chromosomal region 1p32-34, in which RAD54L resides, has been reported as a frequent event in breast cancer<sup>98</sup>, oligodendroglioma<sup>99</sup>, nontypical meningioma<sup>100-103</sup>, and parathyroid adenoma<sup>104</sup>, but it is not clear whether RAD54L loss of function is pathogenic in these cases. Increased RAD54L expression was reported in NSCLC samples in response to increased mutation rate<sup>105</sup> and also in castration-resistant prostate cancer (CRPC) cells<sup>106</sup>. RAD54L polymorphisms have been associated with increased risk of developing

meningioma<sup>107</sup>, glioma<sup>108</sup>, and decreased overall survival ( $P < 0.004$ ) in patients with potentially resectable pancreatic adenocarcinoma<sup>109</sup>. Germline mutations of RAD54L has been associated with increased risk of gastric cancer<sup>110</sup> but not lymphoid malignancies<sup>111</sup>.

## FINDING SUMMARY

RAD54L encodes a member of the SNF2/SWI2 superfamily and forms part of the RAD52 complex involved in recombination and DNA repair in response to ionizing radiation<sup>112-115</sup>. Alterations leading to disruption of critical domains with RAD54L are predicted to enhance genomic instability<sup>116</sup>. Alterations such as seen here may disrupt RAD54L function or expression<sup>116-121</sup>.

## POTENTIAL TREATMENT STRATEGIES

### — Targeted Therapies —

There are no therapies available that directly target RAD54L. Limited clinical evidence in ovarian

GENE

# ASXL1

ALTERATION

L386\*

TRANSCRIPT ID

NM\_015338

CODING SEQUENCE EFFECT

1157T>A

various solid tumor types<sup>97</sup> and are not known to act as drivers in any specific solid cancer type<sup>122</sup>. Published data investigating the prognostic implications of ASXL1 alterations in solid tumors are limited (PubMed, May 2022). In the context of clonal hematopoiesis, ASXL1 mutations are significantly enriched in current or former smokers<sup>123</sup>.

## FINDING SUMMARY

ASXL1 regulates epigenetic marks and transcription through interaction with polycomb complex proteins and various transcription activators and repressors<sup>124-126</sup>. Alterations such as seen here may disrupt ASXL1 function or expression<sup>127-129</sup>.

## POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion<sup>130-135</sup>. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy<sup>130-131</sup>. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease<sup>136</sup>. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH<sup>134,137-138</sup>. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

## POTENTIAL TREATMENT STRATEGIES

### — Targeted Therapies —

There are no targeted therapies available to address genomic alterations in ASXL1.

## FREQUENCY & PROGNOSIS

ASXL1 alterations occur infrequently across



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**GENOMIC FINDINGS**
**GENE**

# TET2

**ALTERATION**

H1761fs\*5, R1261H, H222fs\*3

**TRANSCRIPT ID**

NM\_001127208, NM\_001127208, NM\_001127208

**CODING SEQUENCE EFFECT**

5282\_5283insA, 3782G&gt;A, 663\_664insA

**POTENTIAL TREATMENT STRATEGIES**
**— Targeted Therapies —**

There are no targeted therapies available to address genomic alterations in TET2 in solid tumors.

**FREQUENCY & PROGNOSIS**

TET2 alterations have been reported at relatively

low frequencies in solid tumors and are more prevalent in hematological malignancies (cBioPortal, Jan 2022)<sup>139-140</sup>. Published data investigating the prognostic implications of TET2 alterations in solid tumors are limited (PubMed, Jan 2022).

**FINDING SUMMARY**

TET2 encodes a tumor suppressor involved in reversing DNA methylation marks, a process critical for proper gene regulation<sup>141-142</sup>. Alterations such as seen here may disrupt TET2 function or expression<sup>143-147</sup>.

**POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS**

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire

somatic mutations that allow for clonal expansion<sup>130-135</sup>. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy<sup>130-131</sup>. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease<sup>136</sup>. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH<sup>134,137-138</sup>. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

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ORDERED TEST # ORD-1423659-01

GENOMIC FINDINGS

GENE

TP53

ALTERATION

A138V

TRANSCRIPT ID

NM\_000546

CODING SEQUENCE EFFECT

413C>T

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor adavosertib<sup>148-151</sup>, or p53 gene therapy and immunotherapeutics such as SGT-53<sup>152-156</sup> and ALT-801<sup>157</sup>. In a Phase 1 study, adavosertib in combination with gemcitabine, cisplatin, or carboplatin elicited PRs in 9.7% and SDs in 53% of patients with solid tumors; the response rate was 21% (4/19) for patients with TP53 mutations versus 12% (4/33) for patients who were TP53 wildtype<sup>158</sup>. A Phase 2 trial of adavosertib in combination with chemotherapy (gemcitabine, carboplatin, paclitaxel, or doxorubicin) reported a 32% (30/94, 3 CR) ORR and a 73% (69/94) DCR for patients with platinum-refractory TP53-mutated ovarian, Fallopian tube, or peritoneal cancer<sup>159</sup>. A smaller Phase 2 trial of adavosertib in combination with carboplatin achieved a 43% (9/21, 1 CR) ORR and a 76% (16/21) DCR for patients with platinum-refractory TP53-mutated ovarian cancer<sup>160</sup>. The combination of adavosertib with paclitaxel and carboplatin for patients with TP53-mutated ovarian cancer also significantly increased PFS compared with paclitaxel and carboplatin alone<sup>161</sup>. In the Phase 2 VIKTORY trial, patients with TP53-mutated metastatic and/or recurrent gastric cancer experienced a 24% (6/25) ORR with adavosertib combined with paclitaxel<sup>162</sup>. A Phase 1 trial of neoadjuvant adavosertib in combination with cisplatin and docetaxel for head and neck squamous cell carcinoma (HNSCC) elicited a 71%

(5/7) response rate for patients with TP53 alterations<sup>163</sup>. The Phase 2 FOCUS4-C trial for patients with TP53- and RAS-mutated colorectal cancer reported improvement in PFS (3.61 vs. 1.87 months, HR=0.35, p=0.0022), but not OS (14.0 vs 12.8 months, p=0.93), following adavosertib treatment compared with active monitoring<sup>164</sup>. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel for patients with solid tumors, 75% (9/12) of evaluable patients experienced clinical benefit, including 2 confirmed and 1 unconfirmed PRs and 2 instances of SD with significant tumor shrinkage<sup>156</sup>. Missense mutations leading to TP53 inactivation may also be sensitive to therapies that reactivate mutated p53 such as APR-246<sup>165-167</sup>. In a Phase 1b trial for patients with p53-positive high-grade serous ovarian cancer, APR-246 combined with carboplatin and pegylated liposomal doxorubicin achieved a 52% (11/21) response rate and 100% DCR<sup>168</sup>. ATR inhibitor treatment of chronic lymphocytic leukemia (CLL) cells with biallelic inactivation of TP53 suppressed cell viability, promoted DNA damage, and attenuated xenograft growth in preclinical studies<sup>169-170</sup>; however, ATR inhibitors as monotherapy had little effect on these parameters in solid tumor models in other preclinical studies<sup>171-172</sup>. Therefore, it is unclear whether TP53 inactivation predicts sensitivity to ATR inhibition.

FREQUENCY & PROGNOSIS

TP53 is one of the most commonly mutated genes in lung cancer; mutations have been reported in 43-80% of non-small cell lung cancers (NSCLCs)<sup>76-77,173-178</sup>, including 42-52% of lung adenocarcinomas and 58-83% of lung squamous cell carcinomas (cBioPortal, COSMIC, Feb 2022)<sup>75-77,179</sup>. TP53 homozygous deletion has been observed in 1.4% of lung adenocarcinoma and <1% of lung squamous cell carcinoma cases (cBioPortal, Feb 2022)<sup>139-140</sup>. In one study of 55 patients with lung adenocarcinoma, TP53 alterations correlated with immunogenic features including PD-L1 expression, tumor mutation burden and neoantigen presentation; likely as a consequence of this association TP53 mutations correlated with improved clinical outcomes to PD-1 inhibitors

pembrolizumab and nivolumab in this study<sup>180</sup>. Mutations in TP53 have been associated with lymph node metastasis in patients with lung adenocarcinoma<sup>181</sup>.

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers<sup>182</sup>. Alterations such as seen here may disrupt TP53 function or expression<sup>183-187</sup>.

POTENTIAL GERMLINE IMPLICATIONS

Germline mutations in TP53 are associated with the very rare autosomal dominant disorder Li-Fraumeni syndrome and the early onset of many cancers<sup>188-190</sup>, including sarcomas<sup>191-192</sup>. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000<sup>193</sup> to 1:20,000<sup>192</sup>. For pathogenic TP53 mutations identified during tumor sequencing, the rate of germline mutations was 1% in the overall population and 6% in tumors arising before age 30<sup>194</sup>. In the appropriate clinical context, germline testing of TP53 is recommended.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion<sup>130-135</sup>. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy<sup>130-131</sup>. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease<sup>136</sup>. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH<sup>134,137-138</sup>. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

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THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

## Afatinib

Assay findings association

EGFR  
L858R

### AREAS OF THERAPEUTIC USE

Afatinib is an irreversible kinase inhibitor that targets the kinase domains of EGFR, ERBB2/HER2, and ERBB4. It is FDA approved for the first-line treatment of patients with metastatic non-small cell lung cancer (NSCLC) and nonresistant EGFR mutations and for the treatment of patients with metastatic, squamous NSCLC after progression on platinum-based chemotherapy. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

EGFR activating mutations may indicate sensitivity to afatinib or dacomitinib for patients with non-small cell lung cancer<sup>47,51,195-196</sup>, whereas data for patients with other tumor types are limited<sup>53-58,197</sup>.

### SUPPORTING DATA

Afatinib has shown significant clinical activity for patients with NSCLC and the EGFR common sensitizing mutations L858R or exon 19 deletions, based on extensive clinical evidence<sup>47,195,198-201</sup>. Two randomized Phase 3 trials reported significantly improved median PFS from afatinib compared with chemotherapy for patients with EGFR common sensitizing mutations (LUX-Lung 3, 13.6 vs. 6.9 months, HR 0.47,  $p < 0.001$ ; LUX-Lung 6, 11.0 vs. 5.6 months, HR 0.28,  $p < 0.0001$ )<sup>47,195</sup>. However, while afatinib significantly increased OS relative to chemotherapy for patients with EGFR exon 19 alterations in these two trials (LUX-Lung 3, 33.3 vs. 21.1 months, HR=0.54; LUX-Lung 6, 31.4 vs. 18.4 months, HR=0.64), no significant OS differences were observed in treatment for patients with L858R mutation<sup>94</sup>. A similar alteration-specific difference was observed for EGFR-mutated treatment-naïve NSCLC in a retrospective analysis, which reported numerically longer median OS from second- versus first-generation EGFR TKIs (48.8 vs. 26.4 months, HR=0.59) for patients with exon 19 deletions, but no substantial difference for patients with L858R (25.4 vs. 20.6 months, HR=0.90)<sup>198</sup>. A Phase 2b study of first-line afatinib compared with gefitinib, also for NSCLC with exon 19 deletions or L858R, reported similar median OS for the two therapies (27.9 vs. 24.5 months, HR=0.86) but significantly longer time-to-treatment-failure (13.7 vs. 11.5 months, HR=0.75) and higher ORR (73% vs. 56%,  $p = 0.0018$ ) with afatinib<sup>199</sup>.

Patients with metastatic NSCLC and common EGFR mutations who progressed on prior chemotherapy experienced an ORR of 50.0% (30/60) from afatinib in a Phase 4 trial<sup>200</sup>. As first-line therapy for NSCLC with EGFR exon 19 deletions or L858R, prospective or randomized Phase 2 trials have reported a median PFS of 10.2 months and OS of 24.8 months for patients unfit for chemotherapy<sup>201</sup> and an ORR of 72.5% ( $n = 40$ , 1 CR), DCR of 100% (40/40), and median PFS and OS of 15.2 and 30.0 months, respectively, for elderly patients  $\geq 70$  years old<sup>202</sup>. A retrospective study of afatinib administered to Asian patients with NSCLC, 99% of whom were previously treated with erlotinib and/or gefitinib, reported an ORR of 27.4% (63/230) for patients with common sensitizing EGFR mutations and an ORR of 24.4% (105/431) for the entire cohort<sup>203</sup>. In a case report, a patient with NSCLC with exon 19 deletion and leptomeningeal metastases experienced an ongoing 16-month PR from afatinib in extracranial, brain, and leptomeningeal lesions<sup>204</sup>. For patients with erlotinib- or gefitinib-resistant NSCLC and EGFR mutations, Phase 2/3 studies of afatinib treatment have generally reported ORRs of only 7 to 9%<sup>205-210</sup>; however, DCRs of more than 50% have been observed<sup>209</sup>. In a Phase 1b or observational study, patients with EGFR-mutated NSCLC who progressed on afatinib experienced further clinical benefit from subsequent treatment with afatinib and cetuximab<sup>211</sup> or osimertinib<sup>212</sup>, respectively. Extensive clinical data have demonstrated that afatinib is effective for patients with EGFR-mutated advanced NSCLC, including exon 19 deletions and L858 mutations, as well as uncommon sensitizing mutations in exons 18 or 20<sup>47,94,195,199,201,203,213</sup>. Afatinib has also shown activity for patients with advanced NSCLC and ERBB2 mutations, most of which were exon 20 insertions<sup>209,214-224</sup>. The randomized Phase 3 LUX-Lung 8 trial comparing afatinib with erlotinib as second-line therapy for advanced lung squamous cell carcinoma (SCC) reported significantly longer median OS (7.9 vs. 6.8 months, HR=0.81), significantly longer median PFS (2.6 vs. 1.9 months, HR=0.81), and higher DCR (51% vs. 40%,  $p = 0.002$ ) for patients treated with afatinib<sup>213</sup>. For patients who progressed on afatinib monotherapy, additional clinical benefit has been reported from afatinib combined with paclitaxel<sup>225</sup>.

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THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

## Dacomitinib

Assay findings association

**EGFR**  
L858R

### AREAS OF THERAPEUTIC USE

Dacomitinib is a second generation irreversible tyrosine kinase inhibitor that targets the kinase domains of EGFR, ERBB2/HER2, and ERBB4/HER4. It is FDA approved for the first-line treatment of patients with metastatic non-small cell lung cancer (NSCLC) with EGFR exon 19 deletion or exon 21 L858R substitution mutations. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

EGFR activating mutations may indicate sensitivity to afatinib or dacomitinib for patients with non-small cell lung cancer<sup>47,51,195-196</sup>, whereas data for patients with other tumor types are limited<sup>53-58,197</sup>. Patients with untreated advanced NSCLC and EGFR L858R mutations achieved an ORR of 73% (68/93)<sup>226</sup> and a median OS of 32.5 months with dacomitinib<sup>51</sup>.

### SUPPORTING DATA

A randomized Phase 3 trial for patients with non-small cell lung cancer (NSCLC) harboring activating EGFR mutations (primarily L858R or exon 19 deletions) reported improved clinical benefit with first-line dacomitinib compared with gefitinib (median OS [mOS] of 34.1 vs. 26.8 months, HR=0.760; median PFS [mPFS] of 14.7 vs. 9.2 months, HR=0.59)<sup>226-227</sup>; mOS was 34.1 to 36.7

months and ORR was 75% to 79%, depending on the dosing regimen<sup>228</sup>. A pooled subgroup analysis for patients with NSCLC harboring activating EGFR mutations reported improved clinical efficacy with dacomitinib treatment compared with erlotinib (mPFS of 14.6 vs. 9.6 months, HR=0.717; mOS of 26.6 vs. 23.2 months, HR=0.737)<sup>229</sup>. An analysis of dacomitinib in NSCLC comparing common activating EGFR alterations alone with co-occurring common and uncommon EGFR mutations showed no statistically significant difference in total ORR (33% vs. 40%, p=0.636) or DCR (77% vs. 73%, p=0.089); however, multivariate analysis revealed compound mutation status as an independent predictor of worse OS (HR=5.405)<sup>230</sup>. A Phase 1 trial of combination dacomitinib and a MEK1/2 inhibitor for patients with KRAS-mutated CRC, NSCLC, or pancreatic cancer reported 20/36 SDs and 16 PDs, however toxicity from this combination prevented long-term treatment in this patient population<sup>231</sup>. Phase 1/2 studies of dacomitinib for patients with advanced KRAS-wildtype non-small cell lung cancer (NSCLC) who had previously progressed on chemotherapy and erlotinib or gefitinib and were not selected for EGFR mutations reported an ORR of 5%-17% (3/66-9/53), median PFS of 3-4 months, and median OS of 8.5-11 months<sup>232-233</sup>.

## Erlotinib

Assay findings association

**EGFR**  
L858R

### AREAS OF THERAPEUTIC USE

Erlotinib is a small-molecule inhibitor of EGFR. It is FDA approved as a monotherapy or in combination with ramucirumab for patients with metastatic non-small cell lung cancer (NSCLC) harboring EGFR exon 19 deletions or exon 21 (L858R) mutations. Erlotinib is also FDA approved in combination with gemcitabine as a first-line treatment for advanced pancreatic cancer. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

Amplification or activation of EGFR may predict sensitivity to therapies such as erlotinib. For patients with activating mutations in EGFR, treatment with erlotinib has been associated with improved response and lengthened time to progression<sup>42,234-236</sup>.

### SUPPORTING DATA

For patients with EGFR-mutated non-small cell lung cancer (NSCLC), the Phase 3 EORTC trial improved PFS with first-line erlotinib relative to platinum-based chemotherapy (9.7 vs. 5.2 months, HR=0.37), though OS was not prolonged (22.9 vs 19.6 months, HR=0.92)<sup>42,237</sup>. This study and meta-analyses attribute the lack of OS

benefit to the effectiveness of post-progression salvage therapy in the control arm<sup>238</sup>. A Phase 3 study reported similar efficacy of erlotinib and gefitinib for patients with EGFR-mutated NSCLC<sup>239</sup>. Patients with EGFR-mutated NSCLC have experienced PFS benefit with the addition of bevacizumab to erlotinib in the first-line setting in Phase 3 trials including the ARTEMIS-CTONG1509 trial for Chinese patients (17.9 vs. 11.2 months, HR=0.55)<sup>240</sup>, the NEJ026 trial for Japanese patients (16.9 vs. 13.3 months, HR=0.605)<sup>241-242</sup>, and the international BEVERLY trial (15.4 vs. 9.7 months, HR=0.60)<sup>243</sup>. OS benefit has not been observed across these studies. In the maintenance setting, Phase 3 trials have reported significantly improved PFS with maintenance erlotinib following first-line platinum-based chemotherapy, with the largest benefit for patients with EGFR mutations<sup>234,244</sup>. In the neoadjuvant setting, a Phase 2 trial reported a numerically improved ORR and significantly longer PFS with erlotinib compared with chemotherapy for patients with EGFR-mutated advanced NSCLC<sup>235</sup>. In the placebo-controlled Phase 3 RELAY trial, the addition of ramucirumab to erlotinib improved PFS for previously untreated patients with NSCLC harboring EGFR L858R or exon 19 deletion (19.4 vs. 12.4 months, HR=0.59)<sup>245</sup>.

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**THERAPIES WITH CLINICAL BENEFIT**
**IN PATIENT'S TUMOR TYPE**

## Gefitinib

*Assay findings association*
**EGFR**  
L858R

### AREAS OF THERAPEUTIC USE

Gefitinib targets the tyrosine kinase EGFR and is FDA approved to treat non-small cell lung cancer (NSCLC) harboring exon 19 deletions or exon 21 (L858R) substitution mutations in EGFR. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

Activation of EGFR may predict sensitivity to therapies such as gefitinib. Clinical studies have consistently shown significant improvement in response rates and PFS for patients with EGFR-mutated non-small cell lung cancer (NSCLC) treated with gefitinib compared with chemotherapy<sup>236,246-251</sup>, and responses have been reported for patients with EGFR-rearranged NSCLC<sup>252-253</sup>.

### SUPPORTING DATA

Gefitinib achieved an ORR of 69.8% and OS of 19.2 months as first-line treatment for Caucasian patients with non-small cell lung cancer (NSCLC) and EGFR sensitizing mutations<sup>43</sup>. Phase 3 studies for Japanese patients<sup>248,254</sup>

and East Asian patients<sup>249,255</sup> with EGFR-mutated NSCLC reported longer PFS but not longer OS on first-line gefitinib compared with cisplatin and docetaxel or carboplatin and paclitaxel. Retrospective analysis of East Asian patients receiving first-line gefitinib reported greatest PFS benefit among patients with EGFR exon 19 insertions or deletions and shortest PFS for those with exon 20 insertions (1.2 months)<sup>256</sup>. Two Phase 3 trials of the combination gefitinib plus pemetrexed and carboplatin compared with gefitinib alone for patients with advanced NSCLC harboring EGFR activating mutations reported significantly higher ORRs (75.3% and 84% vs. 62.5% and 67%), longer median PFS (16 and 20.9 months vs. 8 and 11.9 months), and longer median OS (50.9 months and not reached vs. 17 and 38.8 months) with combination treatment; however, combination treatment was associated with increased Grade 3 or higher adverse events<sup>257-258</sup>. In a Phase 1 study for treatment-naïve patients with NSCLC, 63% (19/30) of patients experienced PR from the combination of gefitinib and the PD-L1 inhibitor durvalumab<sup>259</sup>.

## Osimertinib

*Assay findings association*
**EGFR**  
L858R

### AREAS OF THERAPEUTIC USE

Osimertinib is an irreversible EGFR TKI that is selective for EGFR TKI-sensitizing mutations and the EGFR T790M mutation. It is FDA approved in various treatment settings for patients with non-small cell lung cancer (NSCLC) whose tumors have EGFR exon 19 deletions, exon 21 L858R mutations, or T790M mutations. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

EGFR TKI-sensitizing mutations or rearrangements and/or the EGFR T790M mutation may predict sensitivity to osimertinib in non-small cell lung cancer<sup>52,252,260-262</sup>. Patients with untreated advanced NSCLC and EGFR exon 19 deletions or L858R mutations achieved an ORR of 80% and a median PFS of 21.4 and 14.4 months, respectively<sup>260</sup>.

### SUPPORTING DATA

The Phase 3 FLAURA study reported that, relative to erlotinib or gefitinib, first-line osimertinib significantly increased both median PFS (18.9 vs. 10.2 months, HR=0.46) and median OS (38.6 vs. 31.8 months; HR=0.80) for patients with advanced NSCLC and activating, sensitizing EGFR mutations (specifically, exon 19 deletion or L858R)<sup>260,263</sup>. In the Phase 3 ADAURA study, patients with early Stage (IB/II/IIIA) EGFR-mutated NSCLC experienced longer PFSs on osimertinib compared to placebo in the adjuvant setting (not reached vs. 28.1 months; HR=0.21)<sup>264</sup>. A Phase 1 study reported that

T790M-negative patients with acquired EGFR TKI resistance experienced an ORR of 21% and a median PFS of 2.8 months<sup>52</sup>. A Phase 1b/2 study evaluating osimertinib in combination with the CD73 inhibitor oleclumab for patients with advanced EGFR-mutated, T790M-negative NSCLC reported an ORR of 19% (4/19), a DCR of 81%, and mPFS of 11 months (Kim et al., 2021 AACR Abstract CT163). A Phase 2 trial of osimertinib in combination with bevacizumab versus osimertinib monotherapy for patients with untreated advanced non-small cell lung cancer (NSCLC) harboring EGFR del19 or L858R reported no difference in ORR (82% vs 86%) and median PFS (22.1 vs 20.2 months, HR 0.862 p=0.213)<sup>265</sup>. The Phase 2 BOOSTER study of osimertinib in combination with bevacizumab versus osimertinib monotherapy for patients with advanced NSCLC with EGFR-sensitizing mutations (exon 19 del or L858R) and L790M at progression on prior EGFR TKI reported no difference in ORR (55% vs 55%), median OS (24.0 vs 24.3 months, HR 1.03 p=0.91), or median PFS (15.4 vs 12.3 months, HR 0.96 p=0.83), although improved PFS was observed for the combination in the subgroup of current or former smokers (16.5 vs 8.4, HR 0.52) while nonsmokers had no benefit (HR 1.47)<sup>266</sup>. The Phase 1b TATTON study of osimertinib in combination with selumetinib, savolitinib, or durvalumab for patients with previously treated EGFR-mutated NSCLC reported ORRs of 42% (15/36), 44% (8/18), and 44% (10/23), respectively<sup>267</sup>.

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**THERAPIES WITH CLINICAL BENEFIT**
**IN PATIENT'S TUMOR TYPE**

**NOTE** Genomic alterations detected may be associated with activity of certain US FDA or other specific country approved therapies; however, the therapies listed in this report may have varied evidence in the patient's tumor type. The listed therapies are not ranked in order of potential or predicted efficacy for this patient or in order of level of evidence for this patient's tumor type. The therapies listed in this report may not be complete and/or exhaustive. Furthermore, the listed therapies are limited to US FDA approved pharmaceutical drug products that are linked to a specific genomic alteration. There may also be US FDA approved pharmaceutical drug products that are not linked to a genomic alteration. Further there may also exist pharmaceutical drug products that are not approved by the US FDA or other national authorities. There may also be other treatment modalities available than pharmaceutical drug products.

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**CLINICAL TRIALS**

**IMPORTANT** Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and should be investigated by the physician or

research staff. This is not a comprehensive list of all available clinical trials. There may also be compassionate use or early access programs available, which are not listed in this report. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials are not ranked in order of potential or predicted efficacy for this patient or

in order of level of evidence for this patient's tumor type. Clinical trials listed here may have additional enrollment criteria that may require medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://clinicaltrials.gov). However, [clinicaltrials.gov](https://clinicaltrials.gov) does not list all clinical trials that might be available.

**GENE**  
**EGFR**
**ALTERATION**  
 L858R

**RATIONALE**  
 EGFR activating mutations, rearrangements, or amplification may predict sensitivity to EGFR-targeted therapies. Strategies to overcome

resistance to current agents include next-generation EGFR inhibitors and combination therapies.

**NCT02609776**
**PHASE 1**

A Dose Escalation Study of JNJ-61186372 in Participants With Advanced Non-Small Cell Lung Cancer

**TARGETS**  
 EGFR, MET

**LOCATIONS:** Taipei (Taiwan), Taipei City (Taiwan), Taichung (Taiwan), Kaohsiung (Taiwan), Hangzhou (China), Nanchang (China), Nanjing (China), Hefei (China), Guangzhou (China), Changsha (China)

**NCT03114319**
**PHASE 1**

Dose Finding Study of TNO155 in Adult Patients With Advanced Solid Tumors

**TARGETS**  
 SHP2, EGFR

**LOCATIONS:** Taipei (Taiwan), Seoul (Korea, Republic of), Kobe-shi (Japan), Singapore (Singapore), Amsterdam (Netherlands), Rotterdam (Netherlands), Barcelona (Spain), Hospitalet de Llobregat (Spain), Toronto (Canada), Massachusetts

**NCT04077463**
**PHASE 1**

A Study of Lazertinib as Monotherapy or in Combination With JNJ-61186372 in Japanese Participants With Advanced Non-small Cell Lung Cancer

**TARGETS**  
 EGFR, MET

**LOCATIONS:** Taipei City (Taiwan), Taichung (Taiwan), Tainan (Taiwan), Kaohsiung (Taiwan), Hang Zhou (China), Shanghai (China), Guangzhou (China), Changsha (China), Wuhan (China), Jinan (China)

**NCT02099058**
**PHASE 1**

A Phase 1/1b Study With ABBV-399, an Antibody Drug Conjugate, in Subjects With Advanced Solid Cancer Tumors

**TARGETS**  
 MET, EGFR, PD-1

**LOCATIONS:** Taipei City (Taiwan), Tainan (Taiwan), Suwon (Korea, Republic of), Seoul (Korea, Republic of), Chuo-ku (Japan), Kashiwa-shi (Japan), Marseille CEDEX 05 (France), California, Colorado

**NCT04721015**
**PHASE 1**

Study of Intravenous (IV) ABBV-637 Alone or in Combination With IV Docetaxel/Osimertinib to Assess Adverse Events and Change in Disease Activity in Adult Participants With Relapsed/Refractory (R/R) Solid Tumors

**TARGETS**  
 EGFR

**LOCATIONS:** Taoyuan City (Taiwan), Tainan (Taiwan), Kaohsiung (Taiwan), Fukuoka-shi (Japan), Seoul (Korea, Republic of), Matsuyama-shi (Japan), Goyang (Korea, Republic of), Nagoya-shi (Japan), Chuo-ku (Japan), Kashiwa-shi (Japan)

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**CLINICAL TRIALS**
**NCT03720873**
**PHASE 2**

EGFR-TKIs Combine With Anlotinib as First-line Treatment for Patients With Advanced EGFR Mutation-positive NSCLC

**TARGETS**  
 EGFR, FGFRs, KIT, VEGFRs

**LOCATIONS:** Fuzhou (China)

**NCT04619004**
**PHASE 2**

HERTHENA-Lung01: Patritumab Deruxtecan in Subjects With Metastatic or Locally Advanced EGFR-mutated Non-Small Cell Lung Cancer

**TARGETS**  
 ERBB3

**LOCATIONS:** Hangzhou (China), Shanghai (China), Nanjing (China), Guangzhou (China), Wuhan (China), Zhengzhou (China), Beijing (China), Chengdu (China), Chang chun (China), Harbin (China)

**NCT04058704**
**PHASE 3**

A Study to Determine the Efficiency For Brain Metastasis NSCLC Patients Treated With Icotinib Alone or Combined With Radiation Therapy

**TARGETS**  
 EGFR

**LOCATIONS:** Hangzhou (China)

**NCT05015608**
**PHASE 3**

Study on Savolitinib Combined With Osimertinib in Treatment of Advanced NSCLC With MET Amplification

**TARGETS**  
 MET, EGFR

**LOCATIONS:** Shanghai (China)

**NCT05007938**
**PHASE 2**

Befotertinib and Icotinib in Treatment-naïve Patients With Advanced EGFR-Mutant Lung Cancer

**TARGETS**  
 EGFR

**LOCATIONS:** Xiamen (China)

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**CLINICAL TRIALS**
**GENE**
**RAD54L**
**RATIONALE**

RAD54L inactivation may predict sensitivity to PARP inhibitors.

**ALTERATION**

C391fs\*1

**NCT04380636**
**PHASE 3**

Phase 3 Study of Pembrolizumab With Concurrent Chemoradiation Therapy Followed by Pembrolizumab With or Without Olaparib in Stage III Non-Small Cell Lung Cancer (NSCLC) (MK-7339-012/KEYLYNK-012)

**TARGETS**

PD-L1, PARP, PD-1

**LOCATIONS:** Fuzhou (China), Xiamen (China), Hangzhou (China), Shanghai (China), Shanghai (China), Nanchang (China), Shenzhen (China), Nanjing (China), Changsha (China), Wuhan (China)

**NCT04624204**
**PHASE 3**

Placebo-controlled, Study of Concurrent Chemoradiation Therapy With Pembrolizumab Followed by Pembrolizumab and Olaparib in Newly Diagnosed Treatment-Naïve Limited-Stage Small Cell Lung Cancer (LS-SCLC) (MK 7339-013/KEYLYNK-013)

**TARGETS**

PARP, PD-1, TOP2

**LOCATIONS:** Fuzhou (China), Xiamen (China), Hangzhou (China), Hangzhou (China), Shanghai (China), Nanchang (China), Shanghai (China), Shenzhen (China), Nanjing (China), Changsha (China)

**NCT04644068**
**PHASE 1/2**

Study of AZD5305 as Monotherapy and in Combination With Anti-cancer Agents in Patients With Advanced Solid Malignancies

**TARGETS**

ERBB2, TROP2, PARP

**LOCATIONS:** Shanghai (China), Seoul (Korea, Republic of), Chuo-ku (Japan), Koto-ku (Japan), Melbourne (Australia), Warszawa (Poland), Gdynia (Poland), Grzeznica (Poland), Budapest (Hungary), Brno (Czechia)

**NCT04123366**
**PHASE 2**

Study of Olaparib (MK-7339) in Combination With Pembrolizumab (MK-3475) in the Treatment of Homologous Recombination Repair Mutation (HRRm) and/or Homologous Recombination Deficiency (HRD)-Positive Advanced Cancer (MK-7339-007/KEYLYNK-007)

**TARGETS**

PARP, PD-1

**LOCATIONS:** Fukuoka (Japan), Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Okayama (Japan), Nagoya (Japan), Tokyo (Japan), Kashiwa (Japan), Sapporo (Japan), Nedlands (Australia), Southport (Australia)

**NCT03742895**
**PHASE 2**

Efficacy and Safety of Olaparib (MK-7339) in Participants With Previously Treated, Homologous Recombination Repair Mutation (HRRm) or Homologous Recombination Deficiency (HRD) Positive Advanced Cancer (MK-7339-002 / LYNK-002)

**TARGETS**

PARP

**LOCATIONS:** Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Nedlands (Australia), Port Macquarie (Australia), Darlinghurst (Australia), Adana (Turkey), Ankara (Turkey), Jerusalem (Israel), Konya (Turkey), Ramat Gan (Israel)

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**CLINICAL TRIALS**
**NCT02264678**
**PHASE 1/2**

Ascending Doses of AZD6738 in Combination With Chemotherapy and/or Novel Anti Cancer Agents

**TARGETS**  
ATR, PARP, PD-L1

**LOCATIONS:** Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Goyang-si (Korea, Republic of), Cambridge (United Kingdom), Withington (United Kingdom), Manchester (United Kingdom), London (United Kingdom), Coventry (United Kingdom), Sutton (United Kingdom), Oxford (United Kingdom)

**NCT04939662**
**PHASE 2**

Olaparib and Bevacizumab in Relapsed Small Cell Lung Cancer Subjects

**TARGETS**  
PARP, VEGFA

**LOCATIONS:** Seoul (Korea, Republic of)

**NCT04659785**
**PHASE 1/2**

A Study of Fluzoparib Combined With Apatinib as Second-Line Treatment of Patients With Extensive Stage Small Cell Lung Cancer?FA-ES-SCLC?

**TARGETS**  
PARP, RET, VEGFR2

**LOCATIONS:** Tianjin (China)

**NCT05035745**
**PHASE 1/2**

Selinexor &amp; Talazoparib in Advanced Refractory Solid Tumors; Advanced/Metastatic Triple Negative Breast Cancer (START)

**TARGETS**  
XPO1, PARP

**LOCATIONS:** Singapore (Singapore)

**NCT03772561**
**PHASE 1**

Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies

**TARGETS**  
PARP, AKTs, PD-L1

**LOCATIONS:** Singapore (Singapore)

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**APPENDIX**
**Variants of Unknown Significance**

**NOTE** One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

**AXL**  
A181S

**FLT1**  
T453S

**JAK3**  
rearrangement

**LTK**  
R606Q

**MRE11A**  
R388W

**MTOR**  
R1987Q

**NOTCH3**  
C568Y

**P2RY8**  
E323G

**PARP3**  
R379Q

**ZNF703**  
L63H

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**APPENDIX**
**Genes assayed in FoundationOne®Liquid CDx**

FoundationOne Liquid CDx interrogates 324 genes, including 309 genes with complete exonic (coding) coverage and 15 genes with only select non-coding coverage (indicated with an \*); 75 genes (indicated in bold) are captured with increased sensitivity and have complete exonic (coding) coverage unless otherwise noted.

<b>ABL1</b> Exons 4-9	ACVR1B	<b>AKT1</b> Exon 3	AKT2	AKT3	<b>ALK</b> Exons 20-29, Introns 18, 19	ALOX12B	AMER1 (FAM123B or WTX)	<b>APC</b>
<b>AR</b>	<b>ARAF</b> Exons 4, 5, 7, 11, 13, 15, 16	ARFRP1	ARID1A	ASXL1	<b>ATM</b>	<b>ATR</b>	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BCR* Introns 8, 13, 14	<b>BRAF</b> Exons 11-18, Introns 7-10	<b>BRCA1</b> Introns 2, 7, 8, 12, 16, 19, 20	<b>BRCA2</b> Intron 2	BRD4	BRIP1	BTG1
BTG2	<b>BTK</b> Exons 2, 15	CALR	CARD11	CASP8	CBFB	CBL	<b>CCND1</b>	CCND2
CCND3	CCNE1	CD22	CD70	CD74* Introns 6-8	CD79A	CD79B	<b>CD274</b> (PD-L1)	CDC73
<b>CDH1</b>	<b>CDK12</b>	<b>CDK4</b>	<b>CDK6</b>	CDK8	CDKN1A	CDKN1B	<b>CDKN2A</b>	CDKN2B
CDKN2C	CEBPA	CHEK1	<b>CHEK2</b>	CIC	CREBBP	<b>CRKL</b>	CSF1R	CSF3R
CTCF	CTNNA1	<b>CTNNB1</b> Exon 3	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
<b>DDR2</b> Exons 5, 17, 18	DIS3	DNMT3A	DOT1L	EED	<b>EGFR</b> Introns 7, 15, 24-27	EMSY (C11orf30)	EP300	EPHA3
EPHB1	EPHB4	<b>ERBB2</b>	<b>ERBB3</b> Exons 3, 6, 7, 8, 10, 12, 20, 21, 23, 24, 25	ERBB4	ERCC4	ERG	<b>ERRFI1</b>	<b>ESR1</b> Exons 4-8
ETV4* Intron 8	ETV5* Introns 6, 7	<b>ETV6*</b> Introns 5, 6	EWSR1* Introns 7-13	<b>EZH2</b> Exons 4, 16, 17, 18	EZR* Introns 9-11	FANCA	FANCC	FANCG
FANCL	FAS	FBXW7	FGF10	FGF12	FGF14	FGF19	FGF23	FGF3
FGF4	FGF6	<b>FGFR1</b> Introns 1, 5, Intron 17	<b>FGFR2</b> Intron 1, Intron 17	<b>FGFR3</b> Exons 7, 9 (alternative designation exon 10), 14, 18, Intron 17	FGFR4	FH	FLCN	FLT1
<b>FLT3</b> Exons 14, 15, 20	<b>FOXL2</b>	FUBP1	GABRA6	GATA3	GATA4	GATA6	GID4 (C17orf39)	<b>GNA11</b> Exons 4, 5
GNA13	<b>GNAQ</b> Exons 4, 5	<b>GNAS</b> Exons 1, 8	GRM3	GSK3B	H3-3A (H3F3A)	HDAC1	HGF	HNFI1A
<b>HRAS</b> Exons 2, 3	HSD3B1	ID3	<b>IDH1</b> Exon 4	<b>IDH2</b> Exon 4	IGF1R	IKBKE	IKZF1	INPP4B
IRF2	IRF4	IRS2	JAK1	<b>JAK2</b> Exon 14	<b>JAK3</b> Exons 5, 11, 12, 13, 15, 16	JUN	KDMSA	KDMS5C
KDM6A	KDR	KEAP1	KEL	<b>KIT</b> Exons 8, 9, 11, 12, 13, 17, Intron 16	KLHL6	KMT2A (MLL) Introns 6, 8-11, Intron 7	KMT2D (MLL2)	

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FoundationOne Liquid CDx interrogates 324 genes, including 309 genes with complete exonic (coding) coverage and 15 genes with only select non-coding coverage (indicated with an \*); 75 genes (indicated in bold) are captured with increased sensitivity and have complete exonic (coding) coverage unless otherwise noted.

<b>KRAS</b>	LTK	LYN	MAF	<b>MAP2K1</b> (MEK1) Exons 2, 3	<b>MAP2K2</b> (MEK2) Exons 2-4, 6, 7	MAP2K4	MAP3K1	MAP3K13
MAPK1	MCL1	<b>MDM2</b>	MDM4	MED12	MEF2B	MEN1	MERTK	<b>MET</b>
MITF	MKNK1	MLH1	<b>MPL</b> Exon 10	MRE11 (MRE11A)	MSH2 Intron 5	MSH3	MSH6	MST1R
MTAP	<b>MTOR</b> Exons 19, 30, 39, 40, 43-45, 47, 48, 53, 56	MUTYH	MYB* Intron 14	<b>MYC</b> Intron 1	MYCL (MYCL1)	<b>MYCN</b>	<b>MYD88</b> Exon 4	NBN
<b>NF1</b>	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2 Intron 26	NOTCH3	<b>NPM1</b> Exons 4-6, 8, 10
<b>NRAS</b> Exons 2, 3	NSD2 (WHSC1 or MMSET)	NSD3 (WHSC1L1)	NTSC2	<b>NTRK1</b> Exons 14, 15, Introns 8-11	NTRK2 Intron 12	<b>NTRK3</b> Exons 16, 17	NUTM1* Intron 1	P2RY8
<b>PALB2</b>	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	<b>PDCD1LG2</b> (PD-L2)	<b>PDGFRA</b> Exons 12, 18, Introns 7, 9, 11
<b>PDGFRB</b> Exons 12-21, 23	PDK1	PIK3C2B	PIK3C2G	<b>PIK3CA</b> Exons 2, 3, 5-8, 10, 14, 19, 21 (Coding Exons 1, 2, 4-7, 9, 13, 18, 20) PPP2R2A	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PRDM1	PRKAR1A	PRKCI	PRKN (PARK2)	
PTCH1	<b>PTEN</b>	<b>PTPN11</b>	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B
RAD51C	RAD51D	RAD52	RAD54L	<b>RAF1</b> Exons 3, 4, 6, 7, 10, 14, 15, 17, Introns 4-8	RARA Intron 2	<b>RB1</b>	RBM10	REL
<b>RET</b> Introns 7, 8, Exons 11, 13-16, Introns 9-11	RICTOR	RNF43	<b>ROS1</b> Exons 31, 36-38, 40, Introns 31-35	RPTOR	RSPO2* Intron 1	SDC4* Intron 2	SDHA	SDHB
SDHC	SDHD	SETD2	SF3B1	SGK1	SLC34A2* Intron 4	SMAD2	SMAD4	SMARCA4
SMARCB1	<b>SMO</b>	SNCAIP	SOC51	SOX2	SOX9	SPEN	SPOP	SRC
STAG2	STAT3	<b>STK11</b>	SUFU	SYK	TBX3	TEK	TENT5C (FAM46C)	TERC* ncRNA
<b>TERT*</b> Promoter	TET2	TGFB2	TIPARP	TMPSR2* Introns 1-3	TNFAIP3	TNFRSF14	<b>TP53</b>	TSC1
TSC2	TYRO3	U2AF1	<b>VEGFA</b>	VHL	WT1	XPO1	XRCC2	ZNF217

ZNF703

**ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS**

Microsatellite (MS) status  
Blood Tumor Mutational Burden (bTMB)  
Tumor Fraction

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**APPENDIX**
**About FoundationOne® Liquid CDx**

FoundationOne Liquid CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Ciplastraat 3, 2440 Geel, Belgium. The CE-IVD regulatory status of FoundationOne Liquid CDx is applicable in countries that accept and/or recognize the CE mark.


**ABOUT FOUNDATIONONE LIQUID CDx**

FoundationOne Liquid CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne Liquid CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details.

**INTENDED USE**

FoundationOne Liquid CDx is a next generation sequencing based *in vitro* diagnostic device that analyzes 324 genes. Substitutions and insertion and deletion alterations (indels) are reported in 311 genes, copy number alterations (CNAs) are reported in 310 genes, and gene rearrangements are reported in 324 genes. The test also detects the genomic signatures blood tumor mutational burden (bTMB), microsatellite instability (MSI), and tumor fraction. FoundationOne Liquid CDx utilizes circulating cell-free DNA (cfDNA) isolated from plasma derived from the anti-coagulated peripheral whole blood of cancer patients. The test is intended to be used as a companion diagnostic to identify patients who may benefit from treatment with targeted therapies in accordance with the approved therapeutic product labeling. Additionally, FoundationOne Liquid CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with malignant neoplasms.

**TEST PRINCIPLES**

The FoundationOne Liquid CDx assay is performed exclusively as a laboratory service using circulating cell-free DNA (cfDNA) isolated from plasma derived from anti-coagulated peripheral whole blood from patients with solid malignant neoplasms. The assay employs a single DNA extraction method to obtain cfDNA from plasma from whole blood. Extracted

cfDNA undergoes whole-genome shotgun library construction and hybridization-based capture of 324 cancer-related genes including coding exons and select introns of 309 genes, as well as only select intronic regions or non-coding regions of 15 genes. Hybrid-capture selected libraries are sequenced with deep coverage using the NovaSeq® 6000 platform. Sequence data are processed using a customized analysis pipeline designed to accurately detect genomic alterations, including base substitutions, indels, select copy number variants, and select genomic rearrangements. Substitutions and insertion and deletion alterations (indels) are reported in 311 genes, copy number alterations (CNAs) are reported in 310 genes, and gene rearrangements are reported in 324 genes. The assay also reports tumor fraction, and genomic signatures including MSI and bTMB. A subset of targeted regions in 75 genes is baited for increased sensitivity.

**THE REPORT**

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. *Note:* A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

**QUALIFIED ALTERATION CALLS (EQUIVOCAL)**

All equivocal calls, regardless of alteration type, imply that there is adequate evidence to call the alteration with confidence. However, the repeatability of equivocal calls may be lower than non-equivocal calls.

**RANKING OF THERAPIES AND CLINICAL TRIALS**
*Ranking of Therapies in Summary Table*

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

*Ranking of Clinical Trials*

Pediatric trial qualification → Geographical proximity → Later trial phase.

**LIMITATIONS**

1. For *in vitro* diagnostic use.
2. For prescription use only. This test must be ordered by a qualified medical professional in accordance with clinical laboratory regulations.
3. A negative result does not rule out the presence of a mutation below the limits of detection of the assay. Patients for whom no companion diagnostic alterations are detected should be considered for confirmation with an appropriately validated tumor tissue test, if available.
4. The FoundationOne Liquid CDx assay does not detect heterozygous deletions.
5. The test is not intended to provide information on cancer predisposition.
6. Performance has not been validated for cfDNA input below the specified minimum input.
7. Tissue TMB and blood TMB (bTMB) are estimated from the number of synonymous and nonsynonymous single-nucleotide variants (SNVs) and insertions and deletions (indels) per area of coding genome sampled, after the removal of known and likely oncogenic driver events and germline SNPs. Tissue TMB is calculated based on variants with an allele frequency of  $\geq 5\%$ , and bTMB is calculated based on variants with an allele frequency of  $\geq 0.5\%$ .
8. Tumor fraction is the percentage of circulating tumor DNA (ctDNA) present in a cell-free DNA (cfDNA) sample. The tumor fraction estimate is computationally derived from the observed level of aneuploidy in the sample. Tumor fraction is considered elevated when ctDNA levels are high enough that aneuploidy can be detected and is significantly distinct from that typically found in non-tumor samples.
9. Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the tumor genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor. The MSI algorithm is based on genome wide analysis of 1765 microsatellite loci and not based on the 5 or 7 MSI loci described in current clinical practice guidelines for solid tissue testing.
10. Genomic findings from circulating cell-free DNA (cfDNA) may originate from circulating tumor DNA fragments, germline alterations, or non-tumor somatic alterations, such as clonal hematopoiesis of indeterminate potential (CHIP). Genes with alterations that may be derived from CHIP include, but are not limited

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**APPENDIX**
**About FoundationOne® Liquid CDx**

to: *ASXL1*, *ATM*, *CBL*, *CHEK2*, *DNMT3A*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, *TP53*, and *U2AF1*.

11. Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. If a reported alteration is suspected to be germline, confirmatory testing should be considered in the appropriate clinical context.
12. The test is not intended to replace germline testing or to provide information about cancer predisposition.

**REPORT HIGHLIGHTS**

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

**VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING**

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >30%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to

distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

**VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS**

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *ATM*, *CBL*, *CHEK2*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

**NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION**

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) ([www.nccn.org](http://www.nccn.org)). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to [NCCN.org](http://NCCN.org). NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

**LEVEL OF EVIDENCE NOT PROVIDED**

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

**NO GUARANTEE OF CLINICAL BENEFIT**

This report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

**NO GUARANTEE OF REIMBURSEMENT**

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne Liquid CDx.

**TREATMENT DECISIONS ARE THE RESPONSIBILITY OF PHYSICIAN**

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this test or the information contained in this report.

Certain sample of variant characteristics may result in reduced sensitivity. These include: low sample quality, deletions and insertions >40bp, or repetitive/high homology sequences. FoundationOne Liquid CDx is performed using cell-free DNA, and as such germline events may not be reported.

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APPENDIX

About FoundationOne®Liquid CDx

## SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
Muts/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

## REFERENCE SEQUENCE INFORMATION

Sequence data is mapped to the human genome, Genome Reference Consortium Human Build 37 (GRCh37), also known as hg19.

MR Suite Version 7.0.0

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**APPENDIX** **References**

1. Gandara DR, et al. Nat. Med. (2018) PMID: 30082870
2. Wang Z, et al. JAMA Oncol (2019) PMID: 30816954
3. Sturgill EG, et al. Oncologist (2022) PMID: 35274716
4. Aggarwal C, et al. Clin. Cancer Res. (2020) PMID: 32102950
5. Schenker et al., 2022; AACR Abstract CT022
6. Saori et al., 2021; ESMO Abstract 80P
7. Li et al., 2020; ASCO Abstract 6511
8. Nie W, et al. J Natl Compr Canc Netw (2020) PMID: 32380463
9. Ma Y, et al. Front Oncol (2021) PMID: 34055609
10. Meng G, et al. PLoS One (2022) PMID: 35113949
11. Xiao D, et al. Oncotarget (2016) PMID: 27009843
12. Chen Y, et al. J. Exp. Clin. Cancer Res. (2019) PMID: 31088500
13. Yu H, et al. J Thorac Oncol (2019) PMID: 30253973
14. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
15. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
16. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
17. Rizvi NA, et al. Science (2015) PMID: 25765070
18. Johnson BE, et al. Science (2014) PMID: 24336570
19. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
20. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
21. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
22. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
23. Nature (2012) PMID: 22810696
24. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
25. Bronkhorst AJ, et al. Biomol Detect Quantif (2019) PMID: 30923679
26. Raja R, et al. Clin. Cancer Res. (2018) PMID: 30093454
27. Hrebien S, et al. Ann. Oncol. (2019) PMID: 30860573
28. Choudhury AD, et al. JCI Insight (2018) PMID: 30385733
29. Goodall J, et al. Cancer Discov (2017) PMID: 28450425
30. Goldberg SB, et al. Clin. Cancer Res. (2018) PMID: 29330207
31. Bettgowda C, et al. Sci Transl Med (2014) PMID: 24553385
32. Lapin M, et al. J Transl Med (2018) PMID: 30400802
33. Shulman DS, et al. Br. J. Cancer (2018) PMID: 30131550
34. Stover DG, et al. J. Clin. Oncol. (2018) PMID: 29298117
35. Hemming ML, et al. JCO Precis Oncol (2019) PMID: 30793095
36. Egvud M, et al. Ann. Thorac. Surg. (2019) PMID: 31059681
37. Fan G, et al. PLoS ONE (2017) PMID: 28187169
38. Vu et al., 2020; DOI: 10.1200/PO.19.00204
39. Li G, et al. J Gastrointest Oncol (2019) PMID: 31602320
40. Zhang EW, et al. Cancer (2020) PMID: 32757294
41. Butler TM, et al. Cold Spring Harb Mol Case Stud (2019) PMID: 30833418
42. Rosell R, et al. Lancet Oncol. (2012) PMID: 22285168
43. Douillard JY, et al. Br. J. Cancer (2014) PMID: 24263064
44. Hayashi T, et al. Hum Pathol (2020) PMID: 32673682
45. Cao L, et al. Onco Targets Ther (2018) PMID: 29780256
46. Yang TY, et al. J. Clin. Oncol. (2011) PMID: 21422421
47. Sequist LV, et al. J. Clin. Oncol. (2013) PMID: 23816960
48. Qin BD, et al. Onco Targets Ther (2018) PMID: 30127622
49. Frega S, et al. J Thorac Oncol (2016) PMID: 27131295
50. Long X, et al. Onco Targets Ther (2020) PMID: 33116645
51. Mok TS, et al. J. Clin. Oncol. (2018) PMID: 29864379
52. Jänne PA, et al. N. Engl. J. Med. (2015) PMID: 25923549
53. Hong MH, et al. Cancer (2020) PMID: 32749686
54. Kim HS, et al. Oncotarget (2015) PMID: 26462025
55. Kim HS, et al. Clin. Cancer Res. (2015) PMID: 25424851
56. Mondal G, et al. Acta Neuropathol (2020) PMID: 32303840
57. Cavaliere S, et al. Eur. J. Cancer (2018) PMID: 29734047
58. Chi AS, et al. JCO Precis Oncol (2020) PMID: 32923886
59. Leighl et al., 2021; ESMO Abstract 1192MO
60. Cho et al., 2020; ESMO Abstract 1258O
61. Bauml et al., 2021; ASCO Abstract 9006
62. Shu et al., 2021; ESMO Abstract 1193MO
63. Jänne PA, et al. Cancer Discov (2021) PMID: 34548309
64. Ahn MJ, et al. Lancet Respir Med (2017) PMID: 29056570
65. Yang Z, et al. Sci Transl Med (2016) PMID: 27928026
66. Ahn MJ, et al. Lancet Oncol (2019) PMID: 31587882
67. Lin L, et al. Lung Cancer (2022) PMID: 35248866
68. Reck M, et al. Lancet Respir Med (2019) PMID: 30922878
69. Socinski MA, et al. J Thorac Oncol (2021) PMID: 34311108
70. Socinski MA, et al. N. Engl. J. Med. (2018) PMID: 29863955
71. Lu et al., 2021; ESMO Abstract VP9-2021
72. Dingemans AC, et al. Ann Oncol (2021) PMID: 33864941
73. Marcoux N, et al. J. Clin. Oncol. (2019) PMID: 30550363
74. Vallee A, et al. Int. J. Oncol. (2013) PMID: 23934203
75. Imielinski M, et al. Cell (2012) PMID: 22980975
76. Nature (2014) PMID: 25079552
77. Nature (2012) PMID: 22960745
78. Watzka SB, et al. Eur J Cardiothorac Surg (2010) PMID: 20353893
79. Liang Z, et al. BMC Cancer (2010) PMID: 20637128
80. Grob TJ, et al. Lung Cancer (2013) PMID: 23238037
81. Park S, et al. Histol. Histopathol. (2012) PMID: 22207554
82. Dobashi Y, et al. Hum. Pathol. (2011) PMID: 21040950
83. Ludovini V, et al. Cancer Chemother. Pharmacol. (2013) PMID: 23314677
84. Skrzypski M, et al. Clin Lung Cancer (2013) PMID: 23870818
85. Kim SH, et al. Histol. Histopathol. (2012) PMID: 22419022
86. Lee JS, et al. Ann. Surg. Oncol. (2013) PMID: 23525704
87. Oakley GJ, et al. J Thorac Oncol (2011) PMID: 21587084
88. Marks JL, et al. J Thorac Oncol (2008) PMID: 18303429
89. Izar B, et al. Ann. Thorac. Surg. (2013) PMID: 23932319
90. Ciardiello F, et al. N. Engl. J. Med. (2008) PMID: 18337605
91. Lynch TJ, et al. N. Engl. J. Med. (2004) PMID: 15118073
92. Paez JG, et al. Science (2004) PMID: 15118125
93. Pao W, et al. Proc. Natl. Acad. Sci. U.S.A. (2004) PMID: 15329413
94. Yang JC, et al. Lancet Oncol. (2015) PMID: 25589191
95. Swisher EM, et al. Lancet Oncol. (2017) PMID: 27908594
96. de Bono J, et al. N. Engl. J. Med. (2020) PMID: 32343890
97. Zehir A, et al. Nat. Med. (2017) PMID: 28481359
98. Rasio D, et al. Cancer Res. (1997) PMID: 9192813
99. Bello MJ, et al. Cancer Genet. Cytogenet. (2000) PMID: 10640146
100. Kim NR, et al. J. Neurooncol. (2009) PMID: 19347254
101. Leuraud P, et al. J. Neurooncol. (2000) PMID: 11263500
102. Lopez-Gines C, et al. Cancer Genet. Cytogenet. (2004) PMID: 14734222
103. Mendiola M, et al. Mol. Carcinog. (1999) PMID: 10326867
104. Carling T, et al. Int. J. Cancer (1999) PMID: 10449612
105. Vålk K, et al. Oncology (2010) PMID: 21412013
106. Li L, et al. Sci Signal (2017) PMID: 28536297
107. Leone PE, et al. BMC Cancer (2003) PMID: 12614485
108. Zhang H, et al. J. Neurooncol. (2016) PMID: 26514363
109. Li D, et al. J. Clin. Oncol. (2006) PMID: 16520463
110. Li WQ, et al. Carcinogenesis (2013) PMID: 23504502
111. Shinozuka K, et al. Biol. Blood Marrow Transplant. (2016) PMID: 26743341
112. Kanaar R, et al. Curr. Biol. (1996) PMID: 8805304
113. Sigurdsson S, et al. J. Biol. Chem. (2002) PMID: 12205100
114. Swagemakers SM, et al. J. Biol. Chem. (1998) PMID: 9774452
115. van Veelen LR, et al. Mutat. Res. (2005) PMID: 15914205
116. Smirnova M, et al. J. Biol. Chem. (2004) PMID: 15056673
117. Matsuda M, et al. Oncogene (1999) PMID: 10362365
118. Golub EI, et al. Nucleic Acids Res. (1997) PMID: 9321665
119. Spies J, et al. Mol. Cell (2016) PMID: 27264870
120. Goyal N, et al. Nat Commun (2018) PMID: 29295984
121. Lenger N, et al. Biophys. J. (2019) PMID: 30961891
122. Bailey MH, et al. Cell (2018) PMID: 29625053
123. Bolton KL, et al. Nat Genet (2020) PMID: 33106634
124. Scheuermann JC, et al. Nature (2010) PMID: 20436459
125. Cho YS, et al. J. Biol. Chem. (2006) PMID: 16606617
126. Park UH, et al. J. Biol. Chem. (2011) PMID: 21047783
127. Inoue D, et al. J. Clin. Invest. (2013) PMID: 24216483
128. Abdel-Wahab O, et al. Cancer Cell (2012) PMID: 22897849
129. Br. J. Cancer (2013) PMID: 23736028
130. Jaiswal S, et al. N. Engl. J. Med. (2014) PMID: 25426837
131. Genovese G, et al. N. Engl. J. Med. (2014) PMID: 25426838
132. Xie M, et al. Nat. Med. (2014) PMID: 25326804
133. Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) PMID: 28669404
134. Severson EA, et al. Blood (2018) PMID: 29678827
135. Fuster JJ, et al. Circ. Res. (2018) PMID: 29420212
136. Hematology Am Soc Hematol Educ Program (2018) PMID: 30504320
137. Chabon JJ, et al. Nature (2020) PMID: 32269342
138. Razavi P, et al. Nat. Med. (2019) PMID: 31768066
139. Cerami E, et al. Cancer Discov (2012) PMID: 22588877
140. Gao J, et al. Sci Signal (2013) PMID: 23550210
141. Ito S, et al. Nature (2010) PMID: 20639862
142. Guo JU, et al. Cell (2011) PMID: 21496894
143. Iyer LM, et al. Cell Cycle (2009) PMID: 19411852
144. Ko M, et al. Nature (2010) PMID: 21057493
145. Yang H, et al. Oncogene (2013) PMID: 22391558
146. Hu L, et al. Cell (2013) PMID: 24315485
147. Wang Y, et al. Mol. Cell (2015) PMID: 25601757
148. Hirai H, et al. Cancer Biol. Ther. (2010) PMID: 20107315
149. Bridges KA, et al. Clin. Cancer Res. (2011) PMID: 21799033
150. Rajeshkumar NV, et al. Clin. Cancer Res. (2011) PMID: 21389100
151. Osman AA, et al. Mol. Cancer Ther. (2015) PMID: 25504633
152. Xu L, et al. Mol. Cancer Ther. (2002) PMID: 12489850
153. Xu L, et al. Mol. Med. (2001) PMID: 11713371
154. Camp ER, et al. Cancer Gene Ther. (2013) PMID: 23470564
155. Kim SS, et al. Nanomedicine (2015) PMID: 25240597
156. Pirolo KF, et al. Mol. Ther. (2016) PMID: 27357628
157. Hajdenberg et al., 2012; ASCO Abstract e15010
158. Leijen S, et al. J. Clin. Oncol. (2016) PMID: 27601554
159. Moore et al., 2019; ASCO Abstract 5513
160. Leijen S, et al. J. Clin. Oncol. (2016) PMID: 27998224
161. Oza et al., 2015; ASCO Abstract 5506
162. Lee J, et al. Cancer Discov (2019) PMID: 31315834

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**APPENDIX**
**References**

163. Méndez E, et al. Clin. Cancer Res. (2018) PMID: 29535125
164. Seligmann JF, et al. J Clin Oncol (2021) PMID: 34538072
165. Lehmann S, et al. J. Clin. Oncol. (2012) PMID: 22965953
166. Mohell N, et al. Cell Death Dis (2015) PMID: 26086967
167. Franssón Á, et al. J Ovarian Res (2016) PMID: 27179933
168. Gourley et al., 2016; ASCO Abstract 5571
169. Kwok M, et al. Blood (2016) PMID: 26563132
170. Boudny M, et al. Haematologica (2019) PMID: 30975914
171. Dillon MT, et al. Mol. Cancer Ther. (2017) PMID: 28062704
172. Middleton FK, et al. Cancers (Basel) (2018) PMID: 30127241
173. Mogi A, et al. J. Biomed. Biotechnol. (2011) PMID: 21331359
174. Tekpli X, et al. Int. J. Cancer (2013) PMID: 23011884
175. Vignot S, et al. J. Clin. Oncol. (2013) PMID: 23630207
176. Maeng CH, et al. Anticancer Res. (2013) PMID: 24222160
177. Cortot AB, et al. Clin Lung Cancer (2014) PMID: 24169260
178. Itakura M, et al. Br. J. Cancer (2013) PMID: 23922113
179. Kim Y, et al. J. Clin. Oncol. (2014) PMID: 24323028
180. Dong ZY, et al. Clin. Cancer Res. (2017) PMID: 28039262
181. Seo JS, et al. Genome Res. (2012) PMID: 22975805
182. Brown CJ, et al. Nat. Rev. Cancer (2009) PMID: 19935675
183. Joerger AC, et al. Annu. Rev. Biochem. (2008) PMID: 18410249
184. Kato S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) PMID: 12826609
185. Kamada R, et al. J. Biol. Chem. (2011) PMID: 20978130
186. Zerdoumi Y, et al. Hum. Mol. Genet. (2017) PMID: 28472496
187. Yamada H, et al. Carcinogenesis (2007) PMID: 17690113
188. Bougeard G, et al. J. Clin. Oncol. (2015) PMID: 26014290
189. Sorrell AD, et al. Mol Diagn Ther (2013) PMID: 23355100
190. Nichols KE, et al. Cancer Epidemiol. Biomarkers Prev. (2001) PMID: 11219776
191. Kleihues P, et al. Am. J. Pathol. (1997) PMID: 9006316
192. Gonzalez KD, et al. J. Clin. Oncol. (2009) PMID: 19204208
193. Laloo F, et al. Lancet (2003) PMID: 12672316
194. Mandelker D, et al. Ann. Oncol. (2019) PMID: 31050713
195. Wu YL, et al. Lancet Oncol. (2014) PMID: 24439929
196. Passaro et al., 2019; ELCC Abstract 1150
197. Audet et al., 2013; ASCO Abstract 6041
198. Lau SC, et al. Clin Lung Cancer (2019) PMID: 31178389
199. Paz-Ares L, et al. Ann. Oncol. (2017) PMID: 28426106
200. Thongprasert S, et al. Lung Cancer Manag (2019) PMID: 31807143
201. Januszewski et al., 2018; IASLC WCLC Abstract P1.13-17
202. Suzuki et al., 2018; IASLC WCLC Abstract P1.01-92
203. Chang et al., 2018; IASLC WCLC Abstract P1.01-11
204. Llinás-Quintero N, et al. Case Rep Oncol Med (2019) PMID: 31637072
205. Miller VA, et al. Lancet Oncol. (2012) PMID: 22452896
206. Chen X, et al. Lung Cancer (2013) PMID: 23664448
207. Katakami N, et al. J. Clin. Oncol. (2013) PMID: 23816963
208. Landi L, et al. Clin Lung Cancer (2014) PMID: 25242668
209. De Grève J, et al. Lung Cancer (2015) PMID: 25682316
210. Yang JC, et al. Lancet Oncol. (2015) PMID: 26051236
211. Horn L, et al. Lung Cancer (2017) PMID: 29110849
212. Yamamoto N, et al. Adv Ther (2020) PMID: 31863283
213. Soria JC, et al. Lancet Oncol. (2015) PMID: 26156651
214. Dziadziuszko R, et al. J Thorac Oncol (2019) PMID: 30825613
215. Lai WV, et al. Eur. J. Cancer (2019) PMID: 30685684
216. Greulich H, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) PMID: 22908275
217. Gow CH, et al. J Thorac Oncol (2015) PMID: 26134234
218. Mazières J, et al. Ann. Oncol. (2016) PMID: 26598547
219. Mazières J, et al. J. Clin. Oncol. (2013) PMID: 23610105
220. De Grève J, et al. Lung Cancer (2012) PMID: 22325357
221. Li BT, et al. Lung Cancer (2015) PMID: 26559459
222. Costa DB, et al. J Thorac Oncol (2016) PMID: 26964772
223. Yuan B, et al. Front Oncol (2020) PMID: 32477948
224. Fang W, et al. Oncologist (2019) PMID: 31748336
225. Schuler M, et al. Ann. Oncol. (2016) PMID: 26646759
226. Wu YL, et al. Lancet Oncol. (2017) PMID: 28958502
227. Opsomer RJ, et al. Acta Urol Belg (1985) PMID: 2986437
228. Wu et al., 2018; WCLC abstract MA26.11
229. Ramalingam SS, et al. Ann. Oncol. (2016) PMID: 26768165
230. Li HS, et al. J Thorac Dis (2022) PMID: 35693621
231. van Geel RMJM, et al. Br. J. Cancer (2020) PMID: 32147669
232. Ripellino JA, et al. J Cell Biol (1988) PMID: 2450100
233. Park K, et al. J Thorac Oncol (2014) PMID: 25521398
234. Cappuzzo F, et al. Lancet Oncol. (2010) PMID: 20493771
235. Zhong WZ, et al. J. Clin. Oncol. (2019) PMID: 31194613
236. Petrelli F, et al. Clin Lung Cancer (2012) PMID: 22056888
237. Leon et al., 2014; doi.org/10.1093/annonc/mdl349.52
238. Lee CK, et al. J. Natl. Cancer Inst. (2017) PMID: 28376144
239. Yang JJ, et al. Br. J. Cancer (2017) PMID: 28103612
240. Zhou Q, et al. Cancer Cell (2021) PMID: 34388377
241. Kawashima Y, et al. Lancet Respir Med (2022) PMID: 34454653
242. Saito H, et al. Lancet Oncol (2019) PMID: 30975627
243. Piccirillo et al., 2021; ESMO Abstract 12070
244. Faehling M, et al. J Cancer Res Clin Oncol (2018) PMID: 29687154
245. Nakagawa K, et al. Lancet Oncol. (2019) PMID: 31591063
246. Han JY, et al. J. Clin. Oncol. (2012) PMID: 22370314
247. Maemondo M, et al. N. Engl. J. Med. (2010) PMID: 20573926
248. Mitsudomi T, et al. Lancet Oncol. (2010) PMID: 20022809
249. Mok TS, et al. N. Engl. J. Med. (2009) PMID: 19692680
250. Qi WX, et al. Curr Med Res Opin (2015) PMID: 25329826
251. Zhao H, et al. J Thorac Oncol (2015) PMID: 25546556
252. Wang J, et al. Int. J. Cancer (2019) PMID: 30255937
253. Baik CS, et al. J Thorac Oncol (2015) PMID: 26398831
254. Yoshioka H, et al. Ann. Oncol. (2019) PMID: 31553438
255. Fukuoka M, et al. J. Clin. Oncol. (2011) PMID: 21670455
256. Sutiman N, et al. J Thorac Oncol (2017) PMID: 27908825
257. Noronha V, et al. J. Clin. Oncol. (2019) PMID: 31411950
258. Hosomi Y, et al. J. Clin. Oncol. (2020) PMID: 31682542
259. Creelan BC, et al. Br J Cancer (2021) PMID: 33012782
260. Soria JC, et al. N. Engl. J. Med. (2018) PMID: 29151359
261. Alanazi A, et al. Lung Cancer Manag (2020) PMID: 33318755
262. Kim et al., 2021; DOI: 10.1200/PO.20.00296
263. Ramalingam SS, et al. N. Engl. J. Med. (2019) PMID: 31751012
264. Herbst et al., 2020; ASCO Abstract LBA5
265. Kenmotsu et al., 2021; ESMO Abstract LBA44
266. Soo et al., 2021; ESMO Abstract VP3-2021
267. Oxnard GR, et al. Ann. Oncol. (2020) PMID: 32139298

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