

**ABOUT THE TEST** FoundationOne®Liquid CDx is a next generation sequencing (NGS) assay that identifies clinically relevant genomic alterations in circulating cell-free DNA.

<b>PATIENT</b>	<b>DISEASE</b> Unknown primary undifferentiated small cell carcinoma	<b>PHYSICIAN</b>	<b>ORDERING PHYSICIAN</b> Yeh, Yi-Chen	<b>SPECIMEN</b>	<b>SPECIMEN ID</b> CCH 12/15/1941
	<b>NAME</b> Hsu, Cho-Chiao		<b>MEDICAL FACILITY</b> Taipei Veterans General Hospital		<b>SPECIMEN TYPE</b> Blood
	<b>DATE OF BIRTH</b> 15 December 1941		<b>ADDITIONAL RECIPIENT</b> None		<b>DATE OF COLLECTION</b> 19 January 2022
	<b>SEX</b> Female		<b>MEDICAL FACILITY ID</b> 205872		<b>SPECIMEN RECEIVED</b> 21 January 2022
	<b>MEDICAL RECORD #</b> 12406450		<b>PATHOLOGIST</b> Not Provided		

## Biomarker Findings

**Blood Tumor Mutational Burden** - 21 Muts/Mb  
**Microsatellite status** - MSI-High Not Detected  
**Tumor Fraction** - Elevated Tumor Fraction

## Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

**FBXW7** K374\*

**MET** amplification - equivocal†

**AKT2** amplification

**CCNE1** amplification - equivocal†

**RAF1** amplification, RAF1-PHACTR1 non-canonical fusion

**CDKN2A/B** p16INK4a S12\*

**ERBB3** amplification - equivocal†

**MLL2** P562fs\*6, S3614\*

**TP53** E198\*

† See About the Test in appendix for details.

## Report Highlights

- Variants with **diagnostic implications** that may indicate a specific cancer type: **TP53 E198\*** (p. 12)
- Targeted therapies with potential clinical benefit **approved in another tumor type**: Cabozantinib (p. 13), Capmatinib (p. 13), Crizotinib (p. 14), Everolimus (p. 15), Temsirolimus (p. 15), Tepotinib (p. 16)
- Evidence-matched clinical trial options based on this patient's genomic findings: (p. 17)
- Variants that may represent clonal hematopoiesis and may originate from non-tumor sources: **MLL2 P562fs\*6, S3614\*** (p. 11)

### BIOMARKER FINDINGS

**Blood Tumor Mutational Burden** - 21 Muts/Mb

10 Trials see p. 17

**Microsatellite status** - MSI-High Not Detected

**Tumor Fraction** - Elevated Tumor Fraction

### GENOMIC FINDINGS

VAF %

**FBXW7** - K374\*

16.6%

9 Trials see p. 22

#### THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)

None

#### THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)

None

**MSI-High not detected. No evidence of microsatellite instability in this sample (see Appendix section).**

**Tumor fraction is considered elevated when ctDNA levels are high enough that aneuploidy can be detected. There is higher sensitivity for identifying genomic alterations and a lower risk of false negative results in specimens with elevated tumor fraction; the positive percent agreement observed between liquid and tissue for defined short variants is  $\geq 90\%$  (Li et al., 2021; AACR Abstract 2231) (see Biomarker Findings section).**

#### THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)

None

#### THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)

Everolimus

Temsirolimus

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GENOMIC FINDINGS		VAF %	THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)
<b>MET</b> -	amplification - equivocal	-	None	Cabozantinib
				Capmatinib
				Crizotinib
				Tepotinib
10 Trials see p. 24				
<b>AKT2</b> -	amplification	-	None	None
10 Trials see p. 19				
<b>CCNE1</b> -	amplification - equivocal	-	None	None
1 Trial see p. 21				
<b>RAF1</b> -	amplification	-	None	None
	RAF1-PHACTR1 non-canonical fusion	0.15%		
8 Trials see p. 26				

#### VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS (CH)

Genomic findings below may include nontumor somatic alterations, such as CH. The efficacy of targeting such nontumor somatic alterations is unknown. This content should be interpreted based on clinical context. Refer to appendix for additional information on CH.

**MLL2** - P562fs\*6, S3614\* ..... p. 11

#### GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

**CDKN2A/B** - p16INK4a S12\* ..... p. 10      **MLL2** - P562fs\*6, S3614\* ..... p. 11  
**ERBB3** - amplification - equivocal ..... p. 11      **TP53** - E198\* ..... p. 12

**NOTE** Genomic alterations detected may be associated with activity of certain approved therapies; however, the therapies listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and/or exhaustive. Neither the therapies nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies. Therapies contained in this report may have been approved by the US FDA or other national authorities; however, they might not have been approved in your respective country. In the appropriate clinical context, germline testing of APC, ATM, BAP1, BRCA1, BRCA2, BRIP1, CHEK2, FH, FLCN, MEN1, MLH1, MSH2, MSH6, MUTHY, NF1, NF2, PALB2, PMS2, POLE, PTEN, RAD51C, RAD51D, RB1, RET, SDHA, SDHB, SDHC, SDHD, SMAD4, STK11, TGFBR2, TP53, TSC1, TSC2, VHL, and WT1 is recommended.

Variant Allele Frequency is not applicable for copy number alterations.

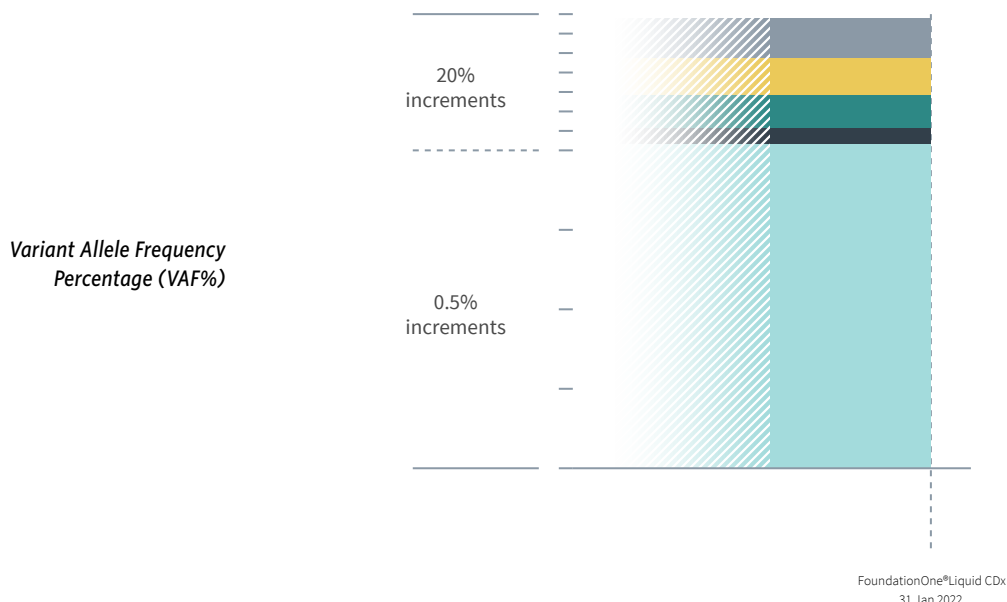
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ORDERED TEST # ORD-1285685-01



#### HISTORIC PATIENT FINDINGS

ORD-1285685-01  
VAF%

#### Blood Tumor Mutational Burden

21 Muts/Mb

#### Microsatellite status

MSI-High Not Detected

#### Tumor Fraction

44%

#### FBXW7

● K374\*

16.6%

#### MET

amplification

Detected

#### AKT2

amplification

Detected

#### CCNE1

amplification

Detected

#### RAF1

amplification

Detected

RAF1-PHACTR1  
non-canonical  
fusion

0.15%

#### CDKN2A/B

● p16INK4a S12\*

33.6%

#### ERBB3

amplification

Detected

#### MLL2

● S3614\*

37.8%

● P562fs\*6

8.7%

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HISTORIC PATIENT FINDINGS	ORD-1285685-01 VAF%
<b>TP53</b>	● E198* 41.0%

**NOTE** This comparison table refers only to genes and biomarkers assayed by prior FoundationOne®Liquid CDx, FoundationOne®Liquid, FoundationOne®, or FoundationOne®CDx tests. Up to five previous tests may be shown.

For some genes in FoundationOne Liquid CDx, only select exons are assayed. Therefore, an alteration found by a previous test may not have been confirmed despite overlapping gene lists. Please refer to the Appendix for the complete list of genes and exons assayed. The gene and biomarker list will be updated periodically to reflect new knowledge about cancer biology.

As new scientific information becomes available, alterations that had previously been listed as Variants of Unknown Significance (VUS) may become reportable.

Tissue Tumor Mutational Burden (TMB) and blood TMB (bTMB) are estimated from the number of synonymous and non-synonymous single-nucleotide variants (SNVs) and insertions and deletions (indels) per area of coding genome sampled, after the removal of known and likely oncogenic driver events and germline SNPs. Tissue TMB is calculated based on variants with an allele frequency of  $\geq 5\%$ , and bTMB is calculated based on variants with an allele frequency of  $\geq 0.5\%$ .

Not Tested = not baited, not reported on test, or test preceded addition of biomarker or gene

Not Detected = baited but not detected on test

Detected = present (VAF% is not applicable)

VAF% = variant allele frequency percentage

Cannot Be Determined = Sample is not of sufficient data quality to confidently determine biomarker status

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**BIOMARKER FINDINGS**
**BIOMARKER**

## Blood Tumor Mutational Burden

**RESULT**

21 Muts/Mb

**POTENTIAL TREATMENT STRATEGIES**
**— Targeted Therapies —**

On the basis of clinical evidence in NSCLC and HNSCC, increased bTMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1<sup>1-2</sup> and anti-PD-1<sup>3</sup> therapies. In NSCLC, multiple clinical trials have shown patients with higher bTMB derive clinical benefit from immune checkpoint inhibitors following single agent or combination treatments with either CTLA4 inhibitors or chemotherapy, with reported high bTMB cutpoints ranging from 6 to 16 Muts/Mb<sup>1</sup>. In HNSCC, a Phase 3 trial showed that bTMB  $\geq 16$  Muts/Mb (approximate equivalency  $\geq 8$  Muts/Mb as measured by this assay) was associated with improved survival from treatment with a PD-L1 inhibitor alone or in

combination with a CTLA-4 inhibitor<sup>4</sup>.

**FREQUENCY & PROGNOSIS**

Average bTMB levels in solid tumors other than NSCLC have not been evaluated (cBioPortal, COSMIC, PubMed, Mar 2021)<sup>5-7</sup>. In 1 retrospective study of patients with advanced neuroendocrine tumors not treated with immunotherapy, tumor mutational burden (TMB)-high ( $\geq 10$  Muts/Mb) was not correlated with any significant difference in OS compared with TMB-low ( $\leq 10$  Muts/Mb) measured in tissue samples (10.4 vs. 6.4 months, adjusted HR = 0.83)<sup>8</sup>. The impact of TMB on the prognosis and clinicopathological features of lung neuroendocrine cancers is unclear; large cell neuroendocrine carcinoma (LCNEC) cases with small cell lung cancer-like molecular features were reported to have significantly higher proliferative activity, as well as a trend toward better clinical benefit from treatment with chemotherapy, than non-small cell lung cancer-like tumors, but the average TMB was not significantly different between the two subsets of LCNEC<sup>9</sup>. MCPyV-negative Merkel cell carcinoma (MCC), associated with higher TMB, has been reported to have a higher number of predicted tumor neoantigens and a significantly higher UV mutation signature

than MCPyV-positive MCC<sup>10-11</sup>. Within MCPyV-negative MCC tumors, the mutational burden has been reported to be significantly higher in PD-L1-positive tumors (more than 1% positive tumor and macrophage cells by immunohistochemistry) than in PD-L1-negative tumors<sup>12</sup>.

**FINDING SUMMARY**

Blood tumor mutational burden (bTMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations from circulating tumor DNA in blood. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma<sup>13-14</sup> and cigarette smoke in lung cancer<sup>15-16</sup>, treatment with temozolomide-based chemotherapy in glioma<sup>17-18</sup>, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes<sup>19-23</sup>, and microsatellite instability (MSI)<sup>19,22-23</sup>. This sample harbors a bTMB level that may be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents<sup>1-3</sup>.

**BIOMARKER**

## Tumor Fraction

**RESULT**

Elevated Tumor Fraction

**POTENTIAL TREATMENT STRATEGIES**
**— Targeted Therapies —**

Specimens with elevated tumor fraction have high circulating-tumor DNA (ctDNA) content, and thus high sensitivity for identifying genomic alterations. Such specimens are at low risk of false negative results. Tumor fraction levels currently have limited implications for diagnosis, surveillance, or therapy and should not be overinterpreted or compared from one blood draw to another. There are currently no targeted

approaches to address specific tumor fraction levels. In the research setting, changes in tumor fraction estimates have been associated with treatment duration and clinical response and may be a useful indicator for future cancer management<sup>24-29</sup>.

**FREQUENCY & PROGNOSIS**

Detectable ctDNA levels have been reported in a variety of tumor types, with higher tumor fraction levels reported for patients with metastatic (Stage 4) tumors compared with patients with localized disease (Stages 1 to 3)<sup>30</sup>. Elevated tumor fraction levels have been reported to be associated with worse prognosis in a variety of cancer types, including pancreatic cancer<sup>31</sup>, Ewing sarcoma and osteosarcoma<sup>32</sup>, prostate cancer<sup>27</sup>, breast cancer<sup>33</sup>, leiomyosarcoma<sup>34</sup>, esophageal cancer<sup>35</sup>, colorectal cancer<sup>36</sup>, and gastrointestinal cancer<sup>37</sup>.

**FINDING SUMMARY**

Tumor fraction provides an estimate of the percentage of ctDNA present in a cell-free DNA (cfDNA) sample. The tumor fraction estimate for this sample is based on the observed level of aneuploid instability. The tumor fraction algorithm utilized for FoundationOne Liquid CDx uses the allele frequencies of approximately 1,000 single-nucleotide polymorphism (SNP) sites across the genome. Unlike the maximum somatic allele frequency (MSAF) method of estimating ctDNA content<sup>38</sup>, the tumor fraction metric does not take into account the allele frequency of individual variants but rather produces a more holistic estimate of ctDNA content using data from across the genome. The amount of ctDNA detected may correlate with disease burden and response to therapy<sup>39-40</sup>.

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ORDERED TEST # ORD-1285685-01

GENOMIC FINDINGS

GENE

**FBXW7**

ALTERATION

K374\*

TRANSCRIPT ID

NM\_033632

CODING SEQUENCE EFFECT

1120A>T

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

FBXW7 inactivating alterations may indicate sensitivity to mTOR inhibitors<sup>41-42</sup>. Several case studies reported clinical benefit for patients with FBXW7-mutated cancers, including lung adenocarcinoma<sup>43</sup>, renal cell carcinoma<sup>44</sup>, and cervical squamous cell carcinoma<sup>45</sup>.

— Nontargeted Approaches —

FBXW7 inactivation may also result in resistance to anti-tubulin chemotherapies based on results from preclinical studies<sup>46</sup>.

FREQUENCY & PROGNOSIS

Across publicly available datasets, FBXW7 mutation has been reported in 0% of pancreatic neuroendocrine tumors (NETs), 2.3% (1/44) of prostate neuroendocrine carcinomas, and 3.2% of small cell lung cancer samples (cBioPortal, Oct 2021)<sup>5-6</sup>. A sequencing study of gastrointestinal NETs reported FBXW7 mutations exclusively in rectal NETs (5/69, 7.2%)<sup>47</sup>. Another study of small intestinal NETs reported both FBXW7 mutation (1/52, 1.9%) and FBXW7 loss of heterozygosity (1/52, 1.9%)<sup>48</sup>. The frequency of rearrangement in neuroendocrine carcinoma has not been evaluated (cBioPortal, COSMIC, PubMed, Oct 2021)<sup>5-7</sup>. Published data investigating the prognostic implications of FBXW7 alteration in

neuroendocrine carcinoma are limited (PubMed, Oct 2021). Reduced FBXW7 expression has been associated with poor prognosis in some cancers such as colorectal cancer, gastric cancer, esophageal SCC, cervical SCC, melanoma, and non-small cell lung carcinoma<sup>49-56</sup>.

FINDING SUMMARY

FBXW7 encodes the F-box protein subunit of the SCF ubiquitin ligase complex, which targets proteins for degradation<sup>57</sup>. FBXW7 inactivation is associated with chromosomal instability and with stabilization of proto-oncogenes, such as mTOR, MYC, cyclin E, NOTCH, and JUN; FBXW7 is therefore considered a tumor suppressor<sup>57-58</sup>. Alterations such as seen here may disrupt FBXW7 function or expression<sup>58-65</sup>.

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GENOMIC FINDINGS

GENE

**MET**

ALTERATION

amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of extensive clinical evidence, MET amplification or activating mutations may predict sensitivity to MET-targeting therapies such as kinase inhibitors crizotinib, capmatinib, tepotinib, and cabozantinib. Crizotinib has benefited patients with MET-amplified non-small cell lung cancer (NSCLC) of varied histologies<sup>66-69</sup>, gastroesophageal cancer<sup>70</sup>, glioblastoma<sup>71</sup>, and carcinoma of unknown primary<sup>72</sup>. Capmatinib has demonstrated clinical efficacy for patients with MET-amplified NSCLC both as a monotherapy<sup>73-74</sup> and in combination with an EGFR-TKI for patients with concurrent activating EGFR mutations<sup>75-77</sup>. Tepotinib has demonstrated efficacy for patients with MET-amplified hepatocellular carcinoma<sup>78</sup> and NSCLC<sup>79</sup> as a monotherapy, as well as in combination with gefitinib for patients with MET-amplified and EGFR-mutated NSCLC<sup>80-82</sup>. Savolitinib elicited responses in patients with MET-amplified papillary renal cell carcinoma<sup>83</sup> and gastric cancer either alone or in combination with docetaxel<sup>84-85</sup>.

AMG 337 elicited an ORR of 50% (5/10), including 1 CR, for patients with MET-amplified gastric, esophageal, or gastroesophageal junction cancer<sup>86</sup>. Patients with MET-amplified NSCLC<sup>87</sup> or MET-amplified gastric cancer<sup>88</sup> treated with the MET-targeting antibody onartuzumab (MetMab) achieved clinical responses. In addition, high MET expression has been suggested to predict patient response to therapies such as the monoclonal HGF-targeting antibody rilotumumab, as well as the combination of ramucirumab and the monoclonal MET-targeting antibody emibetuzumab<sup>89</sup>. A first-in-human Phase 1 trial of telisotuzumab vedotin (teliso-V), a MET antibody-drug conjugate, reported activity in a subset of patients with MET-positive NSCLC, with an ORR of 19% (3/16) and a DCR of 56% (9/16); no responses were observed in any other patients<sup>90</sup>. A subsequent Phase 2 trial of teliso-V in patients with MET-positive NSCLC reported a 35% (13/37) ORR in patients with non-squamous, EGFR-wildtype tumors, which met the prespecified criteria for transition to the next stage; lower ORRs were observed in patients with squamous (14%; 3/21) or non-squamous EGFR-mutated (13%; 4/30) tumors<sup>91</sup>.

FREQUENCY & PROGNOSIS

MET alterations have not been reported in any of 15 ovarian or 21 prostate small cell carcinomas analyzed in the COSMIC dataset (Mar 2021)<sup>7</sup>. MET alterations have not been reported in any of

the 8 cervical small cell carcinomas analyzed in the COSMIC dataset (Mar 2021)<sup>7</sup>. MET expression has been reported frequently (in >25% of cases) in pulmonary neuroendocrine cancers, including large cell neuroendocrine carcinomas, SCLCs, and both typical and atypical carcinoids<sup>92-94</sup>. The frequency of MET amplification in neuroendocrine carcinoma has not been evaluated (cBioPortal, PubMed, Jun 2021)<sup>5-6</sup>. A study of 83 SCLC cases reported a trend toward correlation between MET expression and shorter OS, which reached significance in patients with limited disease-stage tumors<sup>94</sup>. Published data investigating the prognostic implications of MET alterations in neuroendocrine carcinoma are limited (PubMed, Jun 2021).

FINDING SUMMARY

MET encodes a receptor tyrosine kinase, also known as c-MET or hepatocyte growth factor receptor (HGFR), that is activated by the ligand HGF; MET activation results in signaling mediated partly by the RAS-RAF-MAPK and PI3K pathways to promote proliferation<sup>95-96</sup>. MET has been reported to be amplified in cancer<sup>5</sup>, with amplification positively correlating with protein expression in some cancer types<sup>97-101</sup> and associating with therapeutic response to MET inhibitors in a variety of cancer types<sup>66-68,70-72,102-103</sup>.

GENE

**AKT2**

ALTERATION

amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Amplification or activation of AKT2 may promote AKT-mTOR pathway activation and may predict sensitivity to inhibitors of the AKT and downstream mTOR pathways. Clinical benefit has been achieved in 1 patient with AKT2 amplification treated with an mTOR inhibitor<sup>104</sup>.

In preclinical studies, the AKT inhibitor MK-2206 showed evidence of enhancing anti-tumor activity of other chemotherapeutic agents in lung and ovarian tumor cells<sup>105</sup>.

FREQUENCY & PROGNOSIS

The incidence of AKT2 amplification has not been extensively assessed in neuroendocrine carcinoma subtypes (cBioPortal, Dec 2021)<sup>5-6</sup>. One study reported amplification of AKT2 in 5/48 (10.4%) of small intestine neuroendocrine tumors<sup>106</sup>. Published data investigating the prognostic implications of AKT2 alterations in neuroendocrine tumors are limited (PubMed, Dec 2021).

FINDING SUMMARY

AKT2 encodes an intracellular serine/threonine kinase that is also known as PKB-beta. AKT2 is one of three members of the AKT gene family, and activation of AKT2 has been implicated in multiple malignancies<sup>107-108</sup>. AKT isoforms appear to have different roles in tumorigenesis; AKT1 appears to contribute to the initiation of tumors, whereas AKT2 promotes invasion and metastasis in breast tumors<sup>109</sup>. Although AKT2 amplification has been reported to associate with AKT2 overexpression<sup>110-112</sup>, studies in various cancers suggest that AKT2 phosphorylation may have greater clinical relevance than AKT2 amplification or mRNA overexpression<sup>113-114</sup>.

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**GENOMIC FINDINGS**
**GENE**
**CCNE1**
**ALTERATION**

amplification - equivocal

**POTENTIAL TREATMENT STRATEGIES**
**— Targeted Therapies —**

There are no approved therapies that directly target CCNE1 alterations. Because amplification or overexpression of CCNE1 leads to increased genomic instability through the ATR-CHK1-WEE1 pathway<sup>115-116</sup> and cyclin E1 promotes cell cycle progression in a complex with CDK2<sup>117</sup>, clinical and preclinical studies have investigated inhibitors of CHK1, ATR, CDK2, and WEE1 as potential therapeutic approaches for tumors with CCNE1 activation. Clinical benefit has been reported for patients with recurrent high-grade serous ovarian carcinoma (HGSOC) with CCNE1 amplification or expression in response to treatment with the CHK1 inhibitor prexasertib<sup>118</sup>. Studies of the WEE1 inhibitor adavosertib observed PRs in patients with CCNE1-amplified HGSOC and ovarian cancer<sup>119-120</sup>. Similarly, in a Phase 2 study of patients with CCNE1-amplified solid tumors, adavosertib elicited an ORR of 26% with PRs

reported for patients with ovarian cancer, urothelial carcinoma, or melanoma<sup>121</sup>. Preclinical studies have demonstrated that cell lines with CCNE1 amplification or overexpression were sensitive to inhibitors of ATR<sup>122-123</sup>, CDK2<sup>124</sup>, or WEE1<sup>116,125</sup>. However, other studies have shown that sensitivity of various cell lines to CDK2 inhibitors, including SNS-032, dinaciclib, and seliciclib, at clinically achievable doses, is largely independent of CCNE1 copy number or expression<sup>126-129</sup>. One study has reported a reduction in tumor CCNE1 levels in 4/6 lung and esophageal cancer cases following treatment with the HDAC inhibitor vorinostat<sup>130</sup>.

**FREQUENCY & PROGNOSIS**

CCNE1 alterations were reported in 9% (28/320) of neuroendocrine tumors in one genomic study<sup>131</sup>. In one study of lung neuroendocrine tumors, including small cell lung cancer (SCLC) and large cell neuroendocrine carcinoma, cyclin E was overexpressed in 21% (4/19) large cell neuroendocrine carcinoma samples and 71% (25/35) of SCLC samples; overexpression of cyclin E was associated with advanced tumor stage and nodal metastasis<sup>132</sup>. In a study of gastroenteropancreatic neuroendocrine tumors, high cyclin E expression was not a significant independent prognostic marker, however, patients

with low p27 combined with high cyclin E expression had significantly worse overall survival than patients with high p27 combined with low cyclin E expression<sup>132</sup>. Another study examining neuroendocrine carcinoma of the lung reported that cyclin E1 expression was associated with Skp2 expression, observed in 86% of high grade neuroendocrine carcinomas<sup>133</sup>. High cyclin E expression alone, or in combination with decreased p27 expression, has been associated with shorter survival of gastroenteropancreatic neuroendocrine tumor patients<sup>132,134</sup>.

**FINDING SUMMARY**

CCNE1 encodes the protein cyclin E1, which plays a role in the regulated transition from the G1 to S phase by binding to and activating cyclin-dependent protein kinase 2 (CDK2). It also has a direct role in initiation of replication and the maintenance of genomic stability<sup>117</sup>. Amplification of chromosomal region 19q12-q13 has been demonstrated in many types of cancer, and CCNE1 is a well-studied gene within this amplicon<sup>135-136</sup>. Increased copy number of CCNE1 is highly associated with overexpression of the cyclin E1 protein<sup>137-138</sup>. Cyclin E1 overexpression can lead to cell transformation as a result of an increase in cyclin E1 activity<sup>117,139</sup>.

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ORDERED TEST # ORD-1285685-01

GENOMIC FINDINGS

GENE

**RAF1**

ALTERATION

amplification, RAF1-PHACTR1 non-canonical fusion

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

RAF1 amplification may predict sensitivity to pan-RAF inhibitors. The addition of sorafenib to chemotherapy improved PFS for patients with melanoma and RAF1 copy number gains (HR=0.37, p=0.025) in a retrospective analysis<sup>140</sup>. A retrospective study reported RAF1 expression as a predictor of improved OS (HR=1.84) and tumor-free survival (HR=1.32) for patients with hepatocellular carcinoma treated with adjuvant sorafenib in multivariate analyses<sup>141</sup>. RAF1 activating rearrangements may predict sensitivity to pan-RAF and MEK inhibitors. A patient with RAF1-rearranged pancreatic cancer achieved a PR to sorafenib in combination with the glutamate antagonist riluzole in a case report<sup>142</sup>. Case studies

of patients with RAF1-rearranged tumors have reported clinical responses to MEK inhibitors, including a PR for a patient with a RAF1 fusion-positive melanoma and tumor regression for patients with RAF1-rearranged and fusion-positive melanoma<sup>143-145</sup>, complete cytological response for a patient with anaplastic pleomorphic xanthoastrocytoma<sup>146</sup>, and ongoing SD for a patient with pilocytic astrocytoma who had progressed on prior treatments<sup>147</sup>.

FREQUENCY & PROGNOSIS

In small cell lung carcinomas, RAF1 amplification was not observed across 142 samples<sup>148</sup>; RAF1 amplifications in neuroendocrine tumors have not been evaluated in the TCGA datasets (cBioPortal, Aug 2021)<sup>5-6</sup>. The frequency of RAF1 fusions or rearrangements has not been extensively evaluated in neuroendocrine carcinomas (PubMed, Mar 2021). RAF1 mutations have been observed in 0-1% of small cell lung cancer (SCLC) samples<sup>148-150</sup>, 1.9% of pancreatic carcinoid-endocrine tumors, 0% of small intestine carcinoid-endocrine tumors, and 0% (0/9) of stomach carcinoid-endocrine tumors (COSMIC, Mar 2021)<sup>7</sup>.

Published data investigating the prognostic implications of RAF1 alterations in neuroendocrine cancers are limited (PubMed, Mar 2021).

FINDING SUMMARY

RAF1 encodes c-RAF, a member of the RAF family of signaling kinases<sup>151</sup>. These kinases are downstream of RAS and activate the MEK-ERK signaling pathway that promotes cell proliferation and survival<sup>152</sup>. RAF1 has been reported to be amplified in cancer<sup>6</sup> and may be biologically relevant in this context<sup>153-154</sup>. Variants that express the c-RAF kinase domain in the absence of the N-terminal autoinhibitory domain, whether with or without a fusion partner, have been reported to be constitutively active and shown to drive hyperactivation of the MAPK pathway, thereby exhibiting transforming activity<sup>155-158</sup>. One or more of the rearrangements observed here were detected as a reciprocal fusion, are not clearly in-frame, or lack a fusion partner with an oligomerization domain, and it is unclear whether such events would lead to a production of an oncogenic variant.

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ORDERED TEST # ORD-1285685-01

GENOMIC FINDINGS

GENE

**CDKN2A/B**

ALTERATION

p16INK4a S12\*

TRANSCRIPT ID

NM\_000077

CODING SEQUENCE EFFECT

35C>A

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Preclinical data suggest that tumors with loss of p16INK4a function may be sensitive to CDK4/6 inhibitors, such as abemaciclib, ribociclib, and palbociclib<sup>159-162</sup>. Although case studies have reported that patients with breast cancer or uterine leiomyosarcoma harboring CDKN2A loss responded to palbociclib treatment<sup>163-164</sup>, multiple other clinical studies have shown no significant correlation between p16INK4a loss or inactivation and therapeutic benefit of these agents<sup>165-171</sup>; it is not known whether CDK4/6 inhibitors would be beneficial in this case.

FREQUENCY & PROGNOSIS

Loss of CDKN2A and/or CDKN2B gene expression, either due to deletion or promoter methylation, has been reported in a variety of neuroendocrine (NE) tumors in the scientific literature, including pancreatic endocrine tumors (9%, CDKN2A), NE lung tumors (15%, CDKN2B),

and various NE gastroenteropancreatic tumors (up to 44%, CDKN2A and CDKN2B)<sup>172-174</sup>. Loss of heterozygosity (LOH) at the chromosomal region 9p21, which contains CDKN2A and CDKN2B, has been reported to be common in small cell NE carcinomas (SCNCs), with incidences of 50% (8/16), 60% (3/5), and 40% (2/5) in SCNCs of the lung, head and neck, and gastrointestinal tract, respectively; however, a lower incidence of LOH (37%) at this region has also been reported in small cell lung cancer<sup>175-176</sup>. The p15.5 isoform of p15INK4b has been reported to be involved in NE lung tumor carcinogenesis independent of p16INK4a or p14ARF<sup>173</sup>. Loss of p14ARF has been reported in a variety of NE tumors, including 40-55% of NE lung tumors, 6% of typical carcinoids, 43% of atypical carcinoids, 50% of large cell NE carcinomas, and 73% of small cell carcinomas<sup>173,177</sup>. Promoter methylation of CDKN2A and CDKN2B has been reported in various NE tumors<sup>172-173,178-180</sup>. Studies in Merkel cell carcinoma have suggested that methylation of the p14ARF promoter may be a greater contributor to p14ARF protein loss than mutation; one study reported p14ARF methylation in 42% (8/19) of Merkel cell carcinoma cases, but no CDKN2A mutation<sup>181-183</sup>. CDKN2A promoter methylation (p16INK4a and p14ARF-encoding loci) in NE tumors has been associated with advanced tumor stage and poor patient outcome<sup>178,183-184</sup>.

FINDING SUMMARY

CDKN2A encodes two different, unrelated tumor

suppressor proteins, p16INK4a and p14ARF, whereas CDKN2B encodes the tumor suppressor p15INK4b<sup>185-186</sup>. Both p15INK4b and p16INK4a bind to and inhibit CDK4 and CDK6, thereby maintaining the growth-suppressive activity of the Rb tumor suppressor; loss or inactivation of either p15INK4b or p16INK4a contributes to dysregulation of the CDK4/6-cyclin-Rb pathway and loss of cell cycle control<sup>187-188</sup>. The tumor suppressive functions of p14ARF involve stabilization and activation of p53, via a mechanism of MDM2 inhibition<sup>189-190</sup>. One or more alterations observed here are predicted to result in p16INK4a loss of function<sup>191-212</sup>.

POTENTIAL GERMLINE IMPLICATIONS

Germline CDKN2A mutation is associated with melanoma-pancreatic cancer syndrome, a condition marked by increased risk of developing malignant melanoma and/or pancreatic cancer<sup>213</sup>. Mutation carriers within families may develop either or both types of cancer, and melanoma cases may be referred to as familial or hereditary melanoma<sup>214-215</sup>. CDKN2A is the most implicated gene in familial melanoma, with germline mutations present in 16% to 20% of familial melanoma cases<sup>216-218</sup>. CDKN2A alteration has also been implicated in familial melanoma-astrocytoma syndrome, an extremely rare tumor association characterized by dual predisposition to melanoma and nervous system tumors<sup>219-221</sup>. In the appropriate clinical context, germline testing of CDKN2A is recommended.

ORDERED TEST # ORD-1285685-01

GENOMIC FINDINGS

GENE

**ERBB3**

ALTERATION

amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

ERBB3 cooperates with other ERBB family members, in particular ERBB2, for efficient signaling<sup>222-225</sup>. Therefore, ERBB3 amplification or activating mutation may predict sensitivity to therapies targeting ERBB2, including antibodies such as trastuzumab, pertuzumab, and ado-trastuzumab emtansine (T-DM1), and dual EGFR/HER2 TKIs such as lapatinib and afatinib. Clinical and preclinical data support sensitivity of ERBB3

activating mutations to various anti-ERBB2 agents<sup>224,226-230</sup>, but data are generally limited for ERBB3 amplification. Biomarker analyses of several Phase 3 trials have not identified an association of ERBB3 expression levels with benefit from trastuzumab-, pertuzumab-, or T-DM1-containing regimens in HER2-positive breast cancer<sup>231-234</sup>, T-DM1 in HER2-positive gastric and gastroesophageal junction (GEJ) cancer<sup>235</sup>, pertuzumab combined with chemotherapy in ovarian cancer<sup>236</sup>, or afatinib in HNSCC<sup>237</sup>. Similarly, ERBB3 expression levels were not associated with PFS or OS from lapatinib plus capecitabine in a Phase 2 study of gastric/GEJ cancer<sup>238</sup> or in retrospective studies of HER2-positive breast cancer<sup>239-241</sup>.

FREQUENCY & PROGNOSIS

In neuroendocrine cancers, ERBB3 mutation has

been observed in 0-0.9% of small cell lung cancers<sup>148-150</sup> and was not seen in pancreatic or prostate neuroendocrine tumors<sup>242-244</sup>. The frequency of ERBB3 amplification in neuroendocrine cancers has not been assessed (cBioPortal, COSMIC, PubMed, Oct 2021)<sup>5-7</sup>. Published data investigating the prognostic implications of ERBB3 alterations in neuroendocrine cancers are limited (PubMed, May 2021).

FINDING SUMMARY

ERBB3 (also known as HER3) encodes a member of the epidermal growth factor receptor (EGFR) family<sup>245</sup>. One study has demonstrated a weak but significant association between ERBB3 gene amplification and ERBB3 protein expression in breast cancer tissue<sup>246</sup>.

GENE

**MLL2**

ALTERATION

P562fs\*6, S3614\*

TRANSCRIPT ID

NM\_003482, NM\_003482

CODING SEQUENCE EFFECT

1685\_1686delCC, 10841C>G

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no targeted therapies available to address genomic alterations in MLL2.

FREQUENCY & PROGNOSIS

MLL2 alterations are observed in a number of solid tumor contexts (COSMIC, Jan 2022)<sup>7</sup>, and are

especially prevalent in lung squamous cell carcinoma (SCC)<sup>247</sup> and small cell lung carcinoma (SCLC)<sup>248</sup>. MLL2 mutation was found to be an independent prognostic factor of poor PFS and OS in non-small cell lung cancer, but not in SCLC<sup>249</sup>. One study reported that MLL2 truncating mutations were more common in recurrent ovary granulosa cell tumors (GCT) compared with primary GCTs (24% [10/42] vs. 3.0% [1/32])<sup>250</sup>. In a study of esophageal SCC, high MLL2 expression positively correlated with tumor stage, differentiation, and size, and negatively correlated with OS<sup>251</sup>.

FINDING SUMMARY

MLL2 encodes an H3K4-specific histone methyltransferase that is involved in the transcriptional response to progesterone signaling<sup>252</sup>. Germline de novo mutations of MLL2 are responsible for the majority of cases of Kabuki

syndrome, a complex and phenotypically distinctive developmental disorder<sup>253</sup>. A significant number of inactivating MLL2 alterations have been observed in multiple tumor types, suggesting a tumor suppressor role<sup>254</sup>.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion<sup>255-260</sup>. Comprehensive genomic profiling of solid tumors may detect nontumor alterations that are due to CH<sup>259,261-262</sup>. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

ORDERED TEST # ORD-1285685-01

GENOMIC FINDINGS

GENE

TP53

ALTERATION  
E198\*

TRANSCRIPT ID  
NM\_000546

CODING SEQUENCE EFFECT  
592G>T

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor adavosertib<sup>263-266</sup>, or p53 gene therapy and immunotherapeutics such as SGT-53<sup>267-271</sup> and ALT-801<sup>272</sup>. In a Phase 1 study, adavosertib in combination with gemcitabine, cisplatin, or carboplatin elicited PRs in 9.7% and SDs in 53% of patients with solid tumors; the response rate was 21% (4/19) for patients with TP53 mutations versus 12% (4/33) for patients who were TP53 wildtype<sup>273</sup>. A Phase 2 trial of adavosertib in combination with chemotherapy (gemcitabine, carboplatin, paclitaxel, or doxorubicin) reported a 32% (30/94, 3 CR) ORR and a 73% (69/94) DCR for patients with platinum-refractory TP53-mutated ovarian, Fallopian tube, or peritoneal cancer<sup>274</sup>. A smaller Phase 2 trial of adavosertib in combination with carboplatin achieved a 43% (9/21, 1 CR) ORR and a 76% (16/21) DCR for patients with platinum-refractory TP53-mutated ovarian cancer<sup>275</sup>. The combination of adavosertib with paclitaxel and carboplatin for patients with TP53-mutated ovarian cancer also significantly increased PFS compared with paclitaxel and carboplatin alone<sup>276</sup>. In the Phase 2 VIKTORY trial, patients with TP53-mutated metastatic and/or recurrent gastric cancer experienced a 24% (6/25) ORR with adavosertib combined with paclitaxel<sup>84</sup>. A Phase 1 trial of neoadjuvant adavosertib in combination with cisplatin and docetaxel for head and neck squamous cell carcinoma (HNSCC) elicited a 71% (5/7) response rate for patients with TP53

alterations<sup>277</sup>. The Phase 2 FOCUS4-C trial for patients with TP53- and RAS-mutated colorectal cancer reported improvement in PFS (3.61 vs. 1.87 months, HR=0.35, p=0.0022), but not OS (14.0 vs 12.8 months, p=0.93), following adavosertib treatment compared with active monitoring<sup>278</sup>. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel for patients with solid tumors, 75% (9/12) of evaluable patients experienced clinical benefit, including 2 confirmed and 1 unconfirmed PRs and 2 instances of SD with significant tumor shrinkage<sup>271</sup>. ATR inhibitor treatment of chronic lymphocytic leukemia (CLL) cells with biallelic inactivation of TP53 suppressed cell viability, promoted DNA damage, and attenuated xenograft growth in preclinical studies<sup>279-280</sup>; however, ATR inhibitors as monotherapy had little effect on these parameters in solid tumor models in other preclinical studies<sup>281-282</sup>. Therefore, it is unclear whether TP53 inactivation predicts sensitivity to ATR inhibition.

FREQUENCY & PROGNOSIS

TP53 mutation has been reported in a number of carcinoid-endocrine cases, including 48% (15/31) of the stomach, 9.1% of large intestine, 6.8% of pancreatic, 5.8% of lung, and 4.8% of small intestine origin; TP53 mutations were also observed in 19% of Merkel cell carcinomas, 41% (9/22) of prostate small cell carcinomas, 14% (10/72) of cervical endocrine tumors, and 63% of small cell lung cancer samples (COSMIC, Jan 2022)<sup>7,283-290</sup>. The frequency of TP53 mutation or loss and altered p53 levels in neuroendocrine lung tumors has been correlated with the degree of malignancy, as TP53 alterations are more frequent in the most malignant tumor types, including SCLC and large cell neuroendocrine carcinoma<sup>291-293</sup>. Within neuroendocrine tumors, expression of p53 has been associated with aggressive and poorly differentiated gastroenteropancreatic neuroendocrine tumors and with shorter survival in patients with high-grade neuroendocrine carcinomas<sup>134,294-296</sup>.

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in

aggressive advanced cancers<sup>297</sup>. Alterations such as seen here may disrupt TP53 function or expression<sup>298-302</sup>.

POTENTIAL DIAGNOSTIC IMPLICATIONS

Mutations in TP53 or RB1 are characteristic of poorly differentiated neuroendocrine carcinomas (NECs) (NCCN Neuroendocrine and Adrenal Tumors, v1.2021)<sup>303-306</sup>.

POTENTIAL GERMLINE IMPLICATIONS

Germline mutations in TP53 are associated with the very rare autosomal dominant disorder Li-Fraumeni syndrome and the early onset of many cancers<sup>307-309</sup>, including sarcomas<sup>310-311</sup>. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000<sup>312</sup> to 1:20,000<sup>311</sup>. For pathogenic TP53 mutations identified during tumor sequencing, the rate of germline mutations was 1% in the overall population and 6% in tumors arising before age 30<sup>313</sup>. In the appropriate clinical context, germline testing of TP53 is recommended.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion<sup>255-260</sup>. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy<sup>255-256</sup>. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease<sup>314</sup>. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH<sup>259,261-262</sup>. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

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THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

## Cabozantinib

Assay findings association

**MET**  
amplification - equivocal

### AREAS OF THERAPEUTIC USE

Cabozantinib inhibits multiple tyrosine kinases, including MET, RET, VEGFRs, and ROS1. It is FDA approved as monotherapy to treat patients with renal cell carcinoma (RCC), hepatocellular carcinoma (HCC), medullary thyroid cancer (MTC), and differentiated thyroid cancer (DTC). It is also approved in combination with nivolumab to treat RCC. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

Sensitivity of MET alterations to cabozantinib is suggested by clinical responses in patients with non-small cell lung cancer (NSCLC) harboring MET mutations associated with MET exon 14 skipping, with or without concurrent MET amplification<sup>315-316</sup>, as well as by extensive preclinical data<sup>317-323</sup>.

### SUPPORTING DATA

A randomized Phase 2 discontinuation study of cabozantinib in 9 solid tumor types reported ORRs of 0% to 22% and response durations of 3.3 to 11.2 months across cohorts with ORRs of 10% or greater observed for patients with ovarian cancer (22% [15/69, 1 CR]), metastatic breast cancer (14% [6/44]), and non-small cell lung cancer (NSCLC) (10% [6/60])<sup>324-325</sup>. A Phase 1 study of cabozantinib for advanced solid tumors reported a 17%

(4/23) ORR in the dose escalation cohort and an ORR of 20% (4/20) and a DCR of 100% (20/20) in the expansion cohort for Japanese patients with NSCLC<sup>326</sup>. In the context of studies for specific solid tumors, the randomized Phase 3 EXAM study for patients with advanced medullary thyroid cancer reported an association of cabozantinib with improved PFS compared with placebo (11.2 vs. 4.0 months, HR=0.28) and a higher ORR (28% vs. 0%), with PFS improvement observed regardless of RET mutation status<sup>327</sup>. The randomized Phase 3 CELESTIAL study for patients with advanced hepatocellular carcinoma (HCC) previously treated with sorafenib reported significantly longer OS (10.2 vs. 8.0 months, HR=0.76) and PFS (5.2 vs. 1.9 months, HR=0.44) as well as an increased ORR (3.8% vs. 0.4%) with cabozantinib when compared to placebo<sup>328</sup>. The Phase 2 CABOSUN trial of first line cabozantinib versus sunitinib for patients with intermediate- or poor-risk advanced clear cell renal cell carcinoma demonstrated significantly improved median PFS (8.2 vs. 5.6 months, HR=0.66), prolonged median OS (30.3 vs. 21.8 months), and higher ORR (33% [26/79] vs. 12% [9/78]) with cabozantinib compared with sunitinib<sup>329</sup>. The Phase 2 CABONE study of cabozantinib reported ORRs of 26% (10/39) for patients with advanced Ewing sarcoma and 12% (5/42) for patients with advanced osteosarcoma<sup>330</sup>.

## Capmatinib

Assay findings association

**MET**  
amplification - equivocal

### AREAS OF THERAPEUTIC USE

Capmatinib is a selective MET tyrosine kinase inhibitor that is FDA approved to treat patients with non-small cell lung cancer harboring MET exon 14 skipping-associated alterations. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

On the basis of clinical data in non-small cell lung cancer<sup>73,79-82,331</sup>, hepatocellular carcinoma<sup>78</sup>, renal cell carcinoma<sup>83</sup>, and gastric cancer<sup>84</sup>, MET amplification may predict sensitivity to selective MET inhibitors.

### SUPPORTING DATA

Clinical data on the efficacy of capmatinib for the

treatment of neuroendocrine tumors are limited (PubMed, Jun 2021). Capmatinib has been investigated primarily for the treatment of NSCLC, demonstrating efficacy as monotherapy for patients with MET amplification<sup>74,332-333</sup> or MET exon 14 skipping alterations<sup>333-334</sup> as well as in combination with EGFR inhibitors for patients with MET amplification<sup>75-77</sup>. Multiple Phase 1 and 2 clinical studies have reported limited efficacy for capmatinib monotherapy in non-NSCLC indications, with no responses observed for patients with MET-amplified glioblastoma (n=10)<sup>335</sup>, MET-overexpressing gastric cancer (n=9)<sup>336</sup>, or other advanced solid tumors with MET amplification or overexpression (n=11)<sup>336-337</sup>.

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THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

## Crizotinib

Assay findings association

**MET**  
amplification - equivocal

### AREAS OF THERAPEUTIC USE

Crizotinib is an inhibitor of the kinases MET, ALK, ROS1, and RON. It is FDA approved to treat patients with ALK rearrangement- or ROS1 rearrangement-positive non-small cell lung cancer (NSCLC), and to treat pediatric and young adult patients with ALK rearrangement-positive anaplastic large cell lymphoma (ALCL). Please see the drug label for full prescribing information.

### GENE ASSOCIATION

Sensitivity of MET alterations to crizotinib is suggested by extensive clinical data in patients with MET-amplified cancers, including non-small cell lung cancer (NSCLC)<sup>66-68,338-339</sup>, gastric cancer<sup>102</sup>, gastroesophageal cancer<sup>70</sup>, glioblastoma<sup>71</sup>, and carcinoma of unknown primary<sup>72</sup>, as well as in patients with MET-mutated cancers, including NSCLC<sup>315,340-344</sup>, renal cell carcinoma (RCC)<sup>345</sup>, and histiocytic sarcoma<sup>340</sup>. Crizotinib has also benefited patients with NSCLC or histiocytic sarcoma tumors harboring various alterations associated with MET exon 14 skipping<sup>315,340,342-344,346</sup>.

### SUPPORTING DATA

A case study reported a patient with ALK-rearranged lung large-cell neuroendocrine carcinoma experienced a PR and PFS of 5 months with crizotinib treatment<sup>347</sup>. In

another case, a patient with chemotherapy-pretreated ALK-rearranged lung atypical carcinoid tumor had tumor reduction after 3 months of crizotinib treatment<sup>348</sup>. Crizotinib has demonstrated efficacy in patients with NSCLC and ALK rearrangements<sup>349-353</sup>, ROS1 rearrangements<sup>354-358</sup>, an NTRK1 fusion<sup>359</sup>, or MET activation<sup>66-68,315,338-339,341-344,360-366</sup>. Crizotinib has also benefited patients with MET-mutated renal cell carcinoma<sup>367</sup> and patients with MET-amplified gastroesophageal cancer, glioblastoma, and carcinoma of unknown primary<sup>70-72</sup>. While a Phase 1b study evaluating crizotinib for the treatment of patients with ALK-positive malignancies, reported ORR of 52.9% (9/17) and 66.7% (6/9) in patients with lymphoma and inflammatory myofibroblastic tumors (IMT), respectively, an ORR of 11.8% (2/17) was reported for patients with other types of tumors<sup>368</sup>. Whereas median PFS and median OS were not reached for patients with lymphoma or IMT, median PFS was 1.3 months and median OS was 8.3 months for patients with other tumor types, and the median duration of treatment was ~1 month relative to 1-3 years for patients with lymphoma or IMT<sup>368</sup>. A Phase 1 clinical trial of crizotinib in pediatric solid tumors reported objective responses in 14/79 patients, including nine CRs and five PRs; response was enriched in patients with activating alterations in ALK<sup>369</sup>.

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Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1285685-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

## Everolimus

Assay findings association

FBXW7  
K374\*

### AREAS OF THERAPEUTIC USE

Everolimus is an orally available mTOR inhibitor that is FDA approved to treat renal cell carcinoma (RCC) following antiangiogenic therapy; pancreatic neuroendocrine tumors; and well-differentiated non-functional neuroendocrine tumors of the lung or gastrointestinal tract. Everolimus is also approved to treat either renal angiomyolipoma or subependymal giant cell astrocytoma in association with tuberous sclerosis complex (TSC). Please see the drug label for full prescribing information.

### GENE ASSOCIATION

On the basis of several case reports describing clinical benefit for patients with FBXW7-mutated cancers treated with mTOR inhibitors, including lung adenocarcinoma<sup>43</sup>, renal cell carcinoma<sup>44</sup>, and cervical squamous cell carcinoma<sup>370</sup>, FBXW7 loss or inactivation may predict sensitivity to everolimus or temsirolimus.

### SUPPORTING DATA

Phase 3 studies demonstrated that everolimus compared with placebo significantly increased median PFS for patients with pancreatic neuroendocrine tumors (NETs) (11.0 vs. 4.6 months)<sup>371</sup> or with well-differentiated, nonfunctional NET of the lung or gastrointestinal tract (11.0 vs. 3.9 months)<sup>372</sup>. The Phase 2 ITMO study of

everolimus in combination with the somatostatin analogue octreotide long-acting repeatable (LAR) in patients with advanced, previously untreated NETs of the gastroenteropancreatic tract and lung reported an ORR of 18% (9/50), and median PFS and OS of 33.6 months and 61.9 months, respectively<sup>373</sup>. In the RADIANT-2 Phase 3 study, for patients with a NET from various primary sites and with a history of carcinoid syndrome, addition of everolimus to the somatostatin analogue octreotide LAR resulted in a nonsignificant increase in PFS compared to the addition of placebo (16.4 vs. 11.3 months) and did not significantly change OS (29.2 vs. 35.2 months)<sup>374-375</sup>; however, a subgroup analysis of patients with lung NET revealed increased median PFS (13.6 vs. 5.6 months)<sup>376</sup>. A Phase 2 study of pancreatic NETs after failure on systematic chemotherapy reported 9.6% PR and 67.8% SD in response to everolimus and 4.4% PR and 80% SD in response to everolimus plus octreotide LAR<sup>377</sup>. Whereas frequent adverse events precluded a recommended Phase 2 dose and schedule for the combination of trametinib and everolimus in a Phase 1b trial for solid tumors<sup>378</sup>, a retrospective study for heavily pretreated patients with solid tumors reported tolerable regimens of the combination for 23/31 patients, with 16 patients treated >3 months and evaluable patients achieving a median PFS of 6.5 months<sup>379</sup>.

## Temsirolimus

Assay findings association

FBXW7  
K374\*

### AREAS OF THERAPEUTIC USE

Temsirolimus is an intravenous mTOR inhibitor that is FDA approved for the treatment of advanced renal cell carcinoma. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

On the basis of several case reports describing clinical benefit for patients with FBXW7-mutated cancers treated with mTOR inhibitors, including lung adenocarcinoma<sup>43</sup>, renal cell carcinoma<sup>44</sup>, and cervical squamous cell carcinoma<sup>370</sup>, FBXW7 loss or inactivation may predict sensitivity to everolimus or temsirolimus.

### SUPPORTING DATA

In a Phase 2 trial, treatment of patients with advanced neuroendocrine carcinoma with temsirolimus resulted in

a response rate of 5.6%; responses were correlated with baseline levels of phosphorylated mTOR<sup>380</sup>. In a Phase 2 study of 55 patients with advanced pancreatic neuroendocrine tumors, 37% showed an objective response to temsirolimus with bevacizumab and 49% of 49 patients evaluated for PFS showed no disease progression at 12 months<sup>381</sup>. A Phase 1 study of 40 evaluable patients treated with a combination therapy of lenalidomide and temsirolimus reported partial response in 2.5% (1/40) and stable disease in 48% (19/40) of cases; the median OS rate in this study was 7.8 months<sup>382</sup>. In a Phase 2 trial, combination of temsirolimus with bevacizumab resulted in a response rate of 41% in patients with pancreatic neuroendocrine tumors<sup>383</sup>. In a Phase 2 study, temsirolimus did not alter PFS in any of 95 patients with small cell lung cancer (SCLC)<sup>384</sup>.

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ORDERED TEST # ORD-1285685-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

## Tepotinib

*Assay findings association*
**MET**  
amplification - equivocal

### AREAS OF THERAPEUTIC USE

Tepotinib is a selective MET tyrosine kinase inhibitor that is FDA approved to treat patients with metastatic non-small cell lung cancer harboring MET exon 14 skipping alterations. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

On the basis of clinical data in non-small cell lung cancer<sup>73,79-82,331</sup>, hepatocellular carcinoma<sup>78</sup>, renal cell carcinoma<sup>83</sup>, and gastric cancer<sup>84</sup>, MET amplification may predict sensitivity to selective MET inhibitors.

### SUPPORTING DATA

Clinical data on the efficacy of tepotinib for the treatment of neuroendocrine tumors are limited (PubMed, Aug 2021). Tepotinib has primarily been investigated in non-small cell lung cancer and has demonstrated efficacy as a

single agent for patients with MET amplification<sup>79</sup> and MET exon 14-skipping alterations<sup>385-386</sup>. Tepotinib has also been shown to be efficacious in combination with gefitinib for patients with concurrent EGFR mutation and MET amplification or MET overexpression in Phase 2 studies<sup>81-82</sup>. In advanced hepatocellular carcinoma, Phase 2 studies of tepotinib reported improved ORR and PFS for both treatment-naïve and previously treated patients with MET protein overexpression<sup>78,387-389</sup>. In a Phase 1 study of advanced solid tumors, tepotinib monotherapy yielded an ORR of 1.3% and a DCR of 24%, with 2 confirmed PRs observed for patients with esophageal or lung cancer and 2 unconfirmed PRs for patients with colorectal or nasopharyngeal cancer<sup>390</sup>. In another Phase 1 study of solid tumors, tepotinib yielded a DCR of 17% (2/12), with 2 SD of ≥12 weeks observed in a patient with gastric cancer and another with urachal cancer<sup>391</sup>.

**NOTE** Genomic alterations detected may be associated with activity of certain US FDA or other specific country approved therapies; however, the therapies listed in this report may have varied evidence in the patient's tumor type. The listed therapies are not ranked in order of potential or predicted efficacy for this patient or in order of level of evidence for this patient's tumor type. The therapies listed in this report may not be complete and/or exhaustive. Furthermore, the listed therapies are limited to US FDA approved pharmaceutical drug products that are linked to a specific genomic alteration. There may also be US FDA approved pharmaceutical drug products that are not linked to a genomic alteration. Further there may also exist pharmaceutical drug products that are not approved by the US FDA or other national authorities. There may also be other treatment modalities available than pharmaceutical drug products.

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**CLINICAL TRIALS**

**IMPORTANT** Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and should be investigated by the physician or

research staff. This is not a comprehensive list of all available clinical trials. There may also be compassionate use or early access programs available, which are not listed in this report. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials are not ranked in order of potential or predicted efficacy for this patient or

in order of level of evidence for this patient's tumor type. Clinical trials listed here may have additional enrollment criteria that may require medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://clinicaltrials.gov). However, [clinicaltrials.gov](https://clinicaltrials.gov) does not list all clinical trials that might be available.

**BIOMARKER**

## Blood Tumor Mutational Burden

**RESULT**

21 Muts/Mb

**RATIONALE**

Increased tumor mutational burden may predict response to anti-PD-1 (alone or in combination

with anti-CTLA-4) or anti-PD-L1 immune checkpoint inhibitors.

**NCT04237649**
**PHASE NULL**

KAZ954 Alone and With PDR001, NZV930 and NIR178 in Advanced Solid Tumors

**TARGETS**  
ADORA2A, CD73, PD-1

**LOCATIONS:** Taipei (Taiwan), Sunto Gun (Japan), Singapore (Singapore), Milano (Italy), Barcelona (Spain), California, Illinois, Missouri, Connecticut, Texas

**NCT04181788**
**PHASE 1/2**

Sasanlimab (PF-06801591, PD-1 Inhibitor) in Participants With Advanced Malignancies

**TARGETS**  
PD-1

**LOCATIONS:** Taipei (Taiwan), Kaohsiung (Taiwan), Shanghai (China), Nanjing (China), Incheon (Korea, Republic of), Seoul (Korea, Republic of), Chongqing (China), Beijing (China), Chuo-ku (Japan), Kopeysk (Russian Federation)

**NCT02829723**
**PHASE 1/2**

Phase I/II Study of BLZ945 Single Agent or BLZ945 in Combination With PDR001 in Advanced Solid Tumors

**TARGETS**  
PD-1, CSF1R

**LOCATIONS:** Taipei (Taiwan), Tainan (Taiwan), Nagoya (Japan), Koto ku (Japan), Singapore (Singapore), Tel Aviv (Israel), Zurich (Switzerland), Rozzano (Italy), Barcelona (Spain), Hospitalet de Llobregat (Spain)

**NCT04521621**
**PHASE 1/2**

A Study of V937 in Combination With Pembrolizumab (MK-3475) in Participants With Advanced/ Metastatic Solid Tumors (V937-013)

**TARGETS**  
PD-1

**LOCATIONS:** Taipei (Taiwan), Taoyuan (Taiwan), Tainan (Taiwan), Kaohsiung (Taiwan), Kashiwa (Japan), Afula (Israel), Jerusalem (Israel), Tel Aviv (Israel), Warszawa (Poland), Oslo (Norway)

**NCT03530397**
**PHASE 1**

A Study to Evaluate MEDI5752 in Subjects With Advanced Solid Tumors

**TARGETS**  
PD-L1, PD-1, CTLA-4

**LOCATIONS:** Taipei (Taiwan), Cheongju-si (Korea, Republic of), Incheon (Korea, Republic of), Seoul (Korea, Republic of), Gyeonggi-do (Korea, Republic of), Amsterdam (Netherlands), Napoli (Italy), Roma (Italy), Villejuif Cedex (France), Barcelona (Spain)

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**CLINICAL TRIALS**
**NCT03192345**
**PHASE 1**

A First-in-human Study of the Safety, Pharmacokinetics, Pharmacodynamics and Anti-tumor Activity of SAR439459 Monotherapy and Combination of SAR439459 and Cemiplimab in Patients With Advanced Solid Tumors

**TARGETS**  
PD-1, TGF-beta

**LOCATIONS:** Taipei 100 (Taiwan), Tainan (Taiwan), Kaohsiung (Taiwan), Seoul (Korea, Republic of), Heidelberg West (Australia), Melbourne (Australia), Tallinn (Estonia), Hannover (Germany), Essen (Germany), Utrecht (Netherlands)

**NCT04261439**
**PHASE 1**

A Phase I/Ib Study of NIZ985 Alone and in Combination With Spartalizumab

**TARGETS**  
PD-1

**LOCATIONS:** Taipei (Taiwan), Chuo ku (Japan), Essen (Germany), Napoli (Italy), Leuven (Belgium), Barcelona (Spain), California, Texas

**NCT03565445**
**PHASE 1**

A Study of ASP1948, Targeting an Immune Modulatory Receptor, in Subjects With Advanced Solid Tumors

**TARGETS**  
PD-1, NRP1

**LOCATIONS:** Taipei (Taiwan), Seoul (Korea, Republic of), Tokyo (Japan), Chiba (Japan), Meldola (Italy), Modena (Italy), Newcastle upon Tyne (United Kingdom), Monza (Italy), Milano (Italy), Glasgow (United Kingdom)

**NCT03799003**
**PHASE 1**

A Study of ASP1951 in Subjects With Advanced Solid Tumors

**TARGETS**  
PD-1, TNFRSF18

**LOCATIONS:** Taipei (Taiwan), Taichung (Taiwan), Daegu (Korea, Republic of), Chungcheongbukdo (Korea, Republic of), Seoul (Korea, Republic of), Gyeonggi-do (Korea, Republic of), Washington, California, Nevada

**NCT03179436**
**PHASE 1/2**

Safety, Pharmacokinetics (PK), and Efficacy of MK-1308 in Combination With Pembrolizumab in Advanced Solid Tumors (MK-1308-001)

**TARGETS**  
CTLA-4, PD-1

**LOCATIONS:** Hangzhou (China), Chongqing (China), Beijing (China), Cairns (Australia), Brisbane (Australia), Kurralt Park (Australia), Waratah (Australia), Blacktown (Australia), Wollstonecraft (Australia), Melbourne (Australia)

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ORDERED TEST # ORD-1285685-01

**CLINICAL TRIALS**
**GENE**
**AKT2**
**RATIONALE**

AKT2 amplification or mutation may lead to AKT- sensitivity to inhibitors of this pathway.  
mTOR pathway activation and may predict

**ALTERATION**

amplification

**NCT04803318**
**PHASE 2**

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

**TARGETS**  
mTOR, FGFRs, RET, PDGFRA, VEGFRs,  
KIT, MEK

**LOCATIONS:** Guangzhou (China)

**NCT03772561**
**PHASE 1**

Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies

**TARGETS**  
PARP, AKTs, PD-L1

**LOCATIONS:** Singapore (Singapore)

**NCT03297606**
**PHASE 2**

Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)

**TARGETS**  
VEGFRs, ABL, SRC, ALK, ROS1, AXL,  
TRKA, MET, TRKC, DDR2, KIT, EGFR,  
PD-1, CTLA-4, PARP, CDK4, CDK6,  
FLT3, CSF1R, RET, mTOR, ERBB2, MEK,  
BRAF, SMO

**LOCATIONS:** Vancouver (Canada), Edmonton (Canada), Saskatoon (Canada), Regina (Canada), Ottawa (Canada), Montreal (Canada), Toronto (Canada), Kingston (Canada), London (Canada)

**NCT03673787**
**PHASE 1/2**

A Trial of Ipatasertib in Combination With Atezolizumab

**TARGETS**  
AKTs, PD-L1

**LOCATIONS:** Sutton (United Kingdom)

**NCT03217669**
**PHASE 1**

Epacadostat (INCB24360) in Combination With Sirolimus in Advanced Malignancy

**TARGETS**  
IDO1, mTOR

**LOCATIONS:** Kansas

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**CLINICAL TRIALS**
**NCT03065062**
**PHASE 1**

Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head & Neck and Other Solid Tumors

**TARGETS**  
PI3K-alpha, PI3K-gamma, mTORC1, mTORC2, CDK4, CDK6

**LOCATIONS:** Massachusetts

**NCT01582191**
**PHASE 1**

A Phase 1 Trial of Vandetanib (a Multi-kinase Inhibitor of EGFR, VEGFR and RET Inhibitor) in Combination With Everolimus (an mTOR Inhibitor) in Advanced Cancer

**TARGETS**  
mTOR, EGFR, SRC, RET, VEGFRs

**LOCATIONS:** Texas

**NCT02159989**
**PHASE 1**

Sapanisertib and Ziv-Aflibercept in Treating Patients With Recurrent Solid Tumors That Are Metastatic or Cannot Be Removed by Surgery

**TARGETS**  
VEGFA, VEGFB, PIGF, mTORC1, mTORC2

**LOCATIONS:** Texas

**NCT02321501**
**PHASE 1**

Phase I/Ib Dose Escalation & Biomarker Study of Ceritinib (LDK378) + Everolimus for Locally Advanced or Metastatic Solid Tumors With an Expansion in Non-Small Cell Lung Cancer (NSCLC) Characterized by Abnormalities in Anaplastic Lymphoma Kinase (ALK) Expression

**TARGETS**  
ROS1, ALK, mTOR

**LOCATIONS:** Texas

**NCT03017833**
**PHASE 1**

Sapanisertib and Metformin in Treating Patients With Advanced or Metastatic Relapsed or Refractory Cancers

**TARGETS**  
mTORC1, mTORC2

**LOCATIONS:** Texas

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**CLINICAL TRIALS**
**GENE**
**CCNE1**
**RATIONALE**

Strong preclinical and clinical data suggest that  
CCNE1 amplification may predict sensitivity to

WEE1 inhibitors.

**ALTERATION**

amplification - equivocal

**NCT03968653**
**PHASE 1**

Study of Oral Debio 0123 in Combination With Carboplatin in Participants With Advanced Solid  
Tumors

**TARGETS**  
**WEE1**
**LOCATIONS:** Groningen (Netherlands), Nijmegen (Netherlands), Leiden (Netherlands), Barcelona (Spain)

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**CLINICAL TRIALS**
**GENE**  
**FBXW7**
**RATIONALE**  
Loss or inactivation of FBXW7 may lead to  
increased mTOR activation and may predict

sensitivity to mTOR inhibitors.

**ALTERATION**  
K374\*

**NCT04803318**
**PHASE 2**

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

**TARGETS**  
mTOR, FGFRs, RET, PDGFRA, VEGFRs,  
KIT, MEK

**LOCATIONS:** Guangzhou (China)

**NCT03297606**
**PHASE 2**

Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)

**TARGETS**  
VEGFRs, ABL, SRC, ALK, ROS1, AXL,  
TRKA, MET, TRKC, DDR2, KIT, EGFR,  
PD-1, CTLA-4, PARP, CDK4, CDK6,  
FLT3, CSF1R, RET, mTOR, ERBB2, MEK,  
BRAF, SMO

**LOCATIONS:** Vancouver (Canada), Edmonton (Canada), Saskatoon (Canada), Regina (Canada), Ottawa (Canada), Montreal (Canada), Toronto (Canada), Kingston (Canada), London (Canada)

**NCT03217669**
**PHASE 1**

Epacadostat (INCB24360) in Combination With Sirolimus in Advanced Malignancy

**TARGETS**  
IDO1, mTOR

**LOCATIONS:** Kansas

**NCT03065062**
**PHASE 1**

Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head &amp; Neck and Other Solid Tumors

**TARGETS**  
PI3K-alpha, PI3K-gamma, mTORC1,  
mTORC2, CDK4, CDK6

**LOCATIONS:** Massachusetts

**NCT01582191**
**PHASE 1**

A Phase 1 Trial of Vandetanib (a Multi-kinase Inhibitor of EGFR, VEGFR and RET Inhibitor) in Combination With Everolimus (an mTOR Inhibitor) in Advanced Cancer

**TARGETS**  
mTOR, EGFR, SRC, RET, VEGFRs

**LOCATIONS:** Texas

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**CLINICAL TRIALS**
**NCT02159989**
**PHASE 1**

Sapanisertib and Ziv-Aflibercept in Treating Patients With Recurrent Solid Tumors That Are Metastatic or Cannot Be Removed by Surgery

**TARGETS**  
VEGFA, VEGFB, PIGF, mTORC1,  
mTORC2

**LOCATIONS:** Texas

**NCT02321501**
**PHASE 1**

Phase I/Ib Dose Escalation &amp; Biomarker Study of Ceritinib (LDK378) + Everolimus for Locally Advanced or Metastatic Solid Tumors With an Expansion in Non-Small Cell Lung Cancer (NSCLC) Characterized by Abnormalities in Anaplastic Lymphoma Kinase (ALK) Expression

**TARGETS**  
ROS1, ALK, mTOR

**LOCATIONS:** Texas

**NCT03017833**
**PHASE 1**

Sapanisertib and Metformin in Treating Patients With Advanced or Metastatic Relapsed or Refractory Cancers

**TARGETS**  
mTORC1, mTORC2

**LOCATIONS:** Texas

**NCT03430882**
**PHASE 1**

Sapanisertib, Carboplatin, and Paclitaxel in Treating Patients With Recurrent or Refractory Malignant Solid Tumors

**TARGETS**  
mTORC1, mTORC2

**LOCATIONS:** Texas

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**CLINICAL TRIALS**
**GENE**
**MET**
**RATIONALE**

Activation of MET may lead to increased MET expression and activation and may therefore

confer sensitivity to MET inhibitors.

**ALTERATION**

amplification - equivocal

**NCT03175224**
**PHASE 1/2**

CBT-101 Study for Advanced Solid Tumors and c-Met Dysregulation

**TARGETS**  
MET

**LOCATIONS:** Taipei City (Taiwan), Taipei (Taiwan), New Taipei City (Taiwan), Taoyuan City (Taiwan), Tainan (Taiwan), Singapore (Singapore), Nedlands (Australia), Saransk (Russian Federation), North Adelaide (Australia), Bedford Park (Australia)

**NCT04647838**
**PHASE 2**

Tepotinib in Solid Tumors Harboring MET Alterations

**TARGETS**  
MET

**LOCATIONS:** Cheonan (Korea, Republic of), Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of)

**NCT03375320**
**PHASE 3**

Cabozantinib S-malate in Treating Patients With Neuroendocrine Tumors Previously Treated With Everolimus That Are Locally Advanced, Metastatic, or Cannot Be Removed by Surgery

**TARGETS**  
MET, ROS1, RET, VEGFRs

**LOCATIONS:** Alaska, Hawaii, Oregon, Idaho, Montana

**NCT03297606**
**PHASE 2**

Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)

**TARGETS**  
VEGFRs, ABL, SRC, ALK, ROS1, AXL, TRKA, MET, TRKC, DDR2, KIT, EGFR, PD-1, CTLA-4, PARP, CDK4, CDK6, FLT3, CSF1R, RET, mTOR, ERBB2, MEK, BRAF, SMO

**LOCATIONS:** Vancouver (Canada), Edmonton (Canada), Saskatoon (Canada), Regina (Canada), Ottawa (Canada), Montreal (Canada), Toronto (Canada), Kingston (Canada), London (Canada)

**NCT04116541**
**PHASE 2**

A Study Evaluating the Activity of Anti-cancer Treatments Targeting Tumor Molecular Alterations/ Characteristics in Advanced / Metastatic Tumors.

**TARGETS**  
CDK6, CDK4, MDM2, MET, ROS1, RET, VEGFRs

**LOCATIONS:** Nice (France), Lyon (France), Marseille (France), Toulouse (France), Bordeaux (France)

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**ORDERED TEST #** ORD-1285685-01

**CLINICAL TRIALS**
**NCT04400474**
**PHASE 2**

Trial of Cabozantinib Plus Atezolizumab in Advanced and Progressive Neoplasms of the Endocrine System. The CABATEN Study

**TARGETS**  
PD-L1, MET, ROS1, RET, VEGFRs

**LOCATIONS:** Badalona (Spain), Barcelona (Spain), L'Hospitalet de Llobregat (Spain), Zaragoza (Spain), Santander (Spain), Oviedo (Spain), Alicante (Spain), Madrid (Spain), Murcia (Spain), Málaga (Spain)

**NCT04079712**
**PHASE 2**

Testing the Combination of XL184 (Cabozantinib), Nivolumab, and Ipilimumab for Poorly Differentiated Neuroendocrine Tumors

**TARGETS**  
PD-1, CTLA-4, MET, ROS1, RET, VEGFRs

**LOCATIONS:** Utah, California, Kansas, Missouri

**NCT04412629**
**PHASE 2**

Cabozantinib in High Grade Neuroendocrine Neoplasms

**TARGETS**  
MET, ROS1, RET, VEGFRs

**LOCATIONS:** Missouri

**NCT04197310**
**PHASE 2**

Cabozantinib and Nivolumab for Carcinoid Tumors

**TARGETS**  
PD-1, MET, ROS1, RET, VEGFRs

**LOCATIONS:** Massachusetts

**NCT04693468**
**PHASE 1**

Talazoparib and Palbociclib, Axitinib, or Crizotinib for the Treatment of Advanced or Metastatic Solid Tumors, TalaCom Trial

**TARGETS**  
PARP, CDK4, CDK6, VEGFRs, ALK, ROS1, AXL, TRKA, MET, TRKC

**LOCATIONS:** Texas

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**ORDERED TEST #** ORD-1285685-01

**CLINICAL TRIALS**
**GENE**
**RAF1**
**RATIONALE**

Activating RAF1 rearrangements may predict sensitivity to MEK inhibitors. RAF1 amplification

may predict sensitivity to pan-RAF inhibitors.

**ALTERATION**

amplification, RAF1-PHACTR1 non-canonical fusion

**NCT04803318**
**PHASE 2**

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

**TARGETS**

mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK

**LOCATIONS:** Guangzhou (China)

**NCT03989115**
**PHASE 1/2**

Dose-Escalation and Dose-Expansion of RMC-4630 and Cobimetinib in Relapsed/Refractory Solid Tumors

**TARGETS**

SHP2, MEK

**LOCATIONS:** Seoul (Korea, Republic of), Oregon, California, Colorado, Arizona, Wisconsin, Illinois

**NCT03284502**
**PHASE 1**

Cobimetinib and HM95573 in Patients With Locally Advanced or Metastatic Solid Tumors

**TARGETS**

MEK, RAFs

**LOCATIONS:** Hwasun (Korea, Republic of), Pusan (Korea, Republic of), Seongnam (Korea, Republic of), Seoul (Korea, Republic of), Goyang-si (Korea, Republic of)

**NCT04801966**
**PHASE NULL**

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

**TARGETS**

CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF

**LOCATIONS:** Melbourne (Australia)

**NCT03905148**
**PHASE 1/2**

Study of the Safety and Pharmacokinetics of BGB-283 and PD-0325901 in Patients With Advanced or Refractory Solid Tumors

**TARGETS**

RAFs, EGFR, MEK

**LOCATIONS:** Nedlands (Australia), Blacktown (Australia), Randwick (Australia), Melbourne (Australia), Texas

**NCT02407509**
**PHASE 1**

Phase I Trial of RO5126766

**TARGETS**

RAFs, MEK, mTOR

**LOCATIONS:** London (United Kingdom), Sutton (United Kingdom)

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**CLINICAL TRIALS**
**NCT02070549**
**PHASE 1**

Trametinib in Treating Patients With Advanced Cancer With or Without Hepatic Dysfunction

**TARGETS**  
**MEK**
**LOCATIONS:** Toronto (Canada)

**NCT03162627**
**PHASE 1**

Selumetinib and Olaparib in Solid Tumors

**TARGETS**  
**MEK, PARP**
**LOCATIONS:** Texas

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**APPENDIX**
**Variants of Unknown Significance**

**NOTE** One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

**ACVR1B**  
amplification

**AKT2**  
S474F

**APC**  
D1841H

**AXIN1**  
R739C

**BCOR**  
S1266C

**BRCA1**  
R1835Q

**BRIP1**  
I857L

**CALR**  
K385N and rearrangement

**DDR2**  
I29M, S131F and  
amplification

**FOXL2**  
Q219\*

**GATA6**  
S184N

**JAK3**  
rearrangement

**KDM5C**  
E1505K

**MLL2**  
E878K, Q3967E and S3696L

**PPARG**  
amplification

**PTPRO**  
D975N

**SDHC**  
amplification

**SMO**  
amplification

**SRC**  
E22\_N23insPAE

**TET2**  
G1861E

**VHL**  
amplification

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**APPENDIX**

Genes assayed in FoundationOne®Liquid CDx

FoundationOne Liquid CDx interrogates 324 genes, including 309 genes with complete exonic (coding) coverage and 15 genes with only select non-coding coverage (indicated with an \*); 75 genes (indicated in bold) are captured with increased sensitivity and have complete exonic (coding) coverage unless otherwise noted.

<b>ABL1</b> Exons 4-9	ACVR1B	<b>AKT1</b> Exon 3	AKT2	AKT3	<b>ALK</b> Exons 20-29, Introns 18, 19	ALOX12B	AMER1 (FAM123B)	<b>APC</b>
<b>AR</b>	<b>ARAF</b> Exons 4, 5, 7, 11, 13, 15, 16	ARFRP1	ARID1A	ASXL1	<b>ATM</b>	<b>ATR</b>	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BCR* Introns 8, 13, 14	<b>BRAF</b> Exons 11-18, Introns 7-10	<b>BRCA1</b> Introns 2, 7, 8, 12, 16, 19, 20	<b>BRCA2</b> Intron 2	BRD4	BRIP1	BTG1
BTG2	<b>BTK</b> Exons 2, 15	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL
<b>CCND1</b>	CCND2	CCND3	CCNE1	CD22	CD70	CD74* Introns 6-8	CD79A	CD79B
<b>CD274</b> (PD-L1)	CDC73	<b>CDH1</b>	<b>CDK12</b>	<b>CDK4</b>	<b>CDK6</b>	CDK8	CDKN1A	CDKN1B
<b>CDKN2A</b>	CDKN2B	CDKN2C	CEBPA	CHEK1	<b>CHEK2</b>	CIC	CREBBP	<b>CRKL</b>
CSF1R	CSF3R	CTCF	CTNNA1	<b>CTNNB1</b> Exon 3	CUL3	CUL4A	CXCR4	CYP17A1
DAXX	DDR1	<b>DDR2</b> Exons 5, 17, 18	DIS3	DNMT3A	DOT1L	EED	<b>EGFR</b> Introns 7, 15, 24-27	EP300
EPHA3	EPHB1	EPHB4	<b>ERBB2</b>	<b>ERBB3</b> Exons 3, 6, 7, 8, 10, 12, 20, 21, 23, 24, 25	ERBB4	ERCC4	ERG	<b>ERRFI1</b>
<b>ESR1</b> Exons 4-8	ETV4* Intron 8	ETV5* Introns 6, 7	<b>ETV6*</b> Introns 5, 6	EWSR1* Introns 7-13	<b>EZH2</b> Exons 4, 16, 17, 18	EZR* Introns 9-11	FAM46C	FANCA
FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12	FGF14	FGF19
FGF23	FGF3	FGF4	FGF6	<b>FGFR1</b> Introns 1, 5, Intron 17	<b>FGFR2</b> Intron 1, Intron 17	<b>FGFR3</b> Exons 7, 9 (alternative designation exon 10), 14, 18, Intron 17	FGFR4	FH
FLCN	FLT1	<b>FLT3</b> Exons 14, 15, 20	<b>FOXL2</b>	FUBP1	GABRA6	GATA3	GATA4	GATA6
<b>GNA11</b> Exons 4, 5	GNA13	<b>GNAQ</b> Exons 4, 5	<b>GNAS</b> Exons 1, 8	GRM3	GSK3B	H3F3A	HDAC1	HGF
HNFI1A	<b>HRAS</b> Exons 2, 3	HSD3B1	ID3	<b>IDH1</b> Exon 4	<b>IDH2</b> Exon 4	IGF1R	IKBKE	IKZF1
INPP4B	IRF2	IRF4	IRS2	JAK1	<b>JAK2</b> Exon 14	<b>JAK3</b> Exons 5, 11, 12, 13, 15, 16	JUN	KDM5A
KDM5C	KDM6A	KDR	KEAP1	KEL	<b>KIT</b> Exons 8, 9, 11, 12, 13, 17, Intron 16	KLHL6	KMT2A (MLL) Introns 6, 8-11, Intron 7	KMT2D (MLL2)

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**APPENDIX**

Genes assayed in FoundationOne®Liquid CDx

FoundationOne Liquid CDx interrogates 324 genes, including 309 genes with complete exonic (coding) coverage and 15 genes with only select non-coding coverage (indicated with an \*); 75 genes (indicated in bold) are captured with increased sensitivity and have complete exonic (coding) coverage unless otherwise noted.

<b>KRAS</b>	LTK	LYN	MAF	<b>MAP2K1</b> (MEK1) Exons 2, 3	<b>MAP2K2</b> (MEK2) Exons 2-4, 6, 7	MAP2K4	MAP3K1	MAP3K13
MAPK1	MCL1	<b>MDM2</b>	MDM4	MED12	MEF2B	MEN1	MERTK	<b>MET</b>
MITF	MKNK1	MLH1	<b>MPL</b> Exon 10	MRE11A	MSH2 Intron 5	MSH3	MSH6	MST1R
MTAP	<b>MTOR</b> Exons 19, 30, 39, 40, 43-45, 47, 48, 53, 56	MUTYH	MYB* Intron 14	<b>MYC</b> Intron 1	MYCL (MYCL1)	<b>MYCN</b>	<b>MYD88</b> Exon 4	NBN
<b>NF1</b>	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2 Intron 26	NOTCH3	<b>NPM1</b> Exons 4-6, 8, 10
<b>NRAS</b> Exons 2, 3	NSD3 (WHSC1L1)	NTSC2	<b>NTRK1</b> Exons 14, 15, Introns 8-11	NTRK2 Intron 12	<b>NTRK3</b> Exons 16, 17	NUTM1* Intron 1	P2RY8	<b>PALB2</b>
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	<b>PDCD1LG2</b> (PD-L2)	<b>PDGFRA</b> Exons 12, 18, Introns 7, 9, 11
<b>PDGFRB</b> Exons 12-21, 23	PDK1	PIK3C2B	PIK3C2G	<b>PIK3CA</b> Exons 2, 3, 5-8, 10, 14, 19, 21 (Coding Exons 1, 2, 4-7, 9, 13, 18, 20) PPP2R2A	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
<b>PTEN</b>	<b>PTPN11</b>	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	<b>RAF1</b> Exons 3, 4, 6, 7, 10, 14, 15, 17, Introns 4-8	RARA Intron 2	<b>RB1</b>	RBM10	REL	<b>RET</b> Introns 7, 8, Exons 11, 13-16, Introns 9-11
RICTOR	RNF43	<b>ROS1</b> Exons 31, 36-38, 40, Introns 31-35	RPTOR	RSP02* Intron 1	SDC4* Intron 2	SDHA	SDHB	SDHC
SDHD	SETD2	SF3B1	SGK1	SLC34A2* Intron 4	SMAD2	SMAD4	SMARCA4	SMARCB1
<b>SMO</b>	SNCAIP	SOC1	SOX2	SOX9	SPEN	SPOP	SRC	STAG2
STAT3	<b>STK11</b>	SUFU	SYK	TBX3	TEK	TERC* ncRNA	<b>TERT*</b> Promoter	TET2
TGFBR2	TIPARP	<b>TMPRSS2*</b> Introns 1-3	TNFAIP3	TNFRSF14	<b>TP53</b>	TSC1	TSC2	TYRO3
U2AF1	<b>VEGFA</b>	VHL	WHSC1	WT1	XPO1	XRCC2	ZNF217	ZNF703

**ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS**

Microsatellite (MS) status

Blood Tumor Mutational Burden (bTMB)

Tumor Fraction

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FoundationOne Liquid CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium. The CE-IVD regulatory status of FoundationOne Liquid CDx is applicable in countries that accept and/or recognize the CE mark.



#### ABOUT FOUNDATIONONE LIQUID CDx

FoundationOne Liquid CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne Liquid CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details.

#### INTENDED USE

FoundationOne Liquid CDx is a next generation sequencing based *in vitro* diagnostic device that analyzes 324 genes. Substitutions and insertion and deletion alterations (indels) are reported in 311 genes, copy number alterations (CNAs) are reported in 310 genes, and gene rearrangements are reported in 324 genes. The test also detects the genomic signatures blood tumor mutational burden (bTMB), microsatellite instability (MSI), and tumor fraction. FoundationOne Liquid CDx utilizes circulating cell-free DNA (cfDNA) isolated from plasma derived from the anti-coagulated peripheral whole blood of cancer patients. The test is intended to be used as a companion diagnostic to identify patients who may benefit from treatment with targeted therapies in accordance with the approved therapeutic product labeling. Additionally, FoundationOne Liquid CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with malignant neoplasms.

#### TEST PRINCIPLES

The FoundationOne Liquid CDx assay is performed exclusively as a laboratory service using circulating cell-free DNA (cfDNA) isolated from plasma derived from anti-coagulated peripheral whole blood from patients with solid malignant neoplasms. The assay employs a single DNA extraction method to obtain cfDNA from plasma from whole blood. Extracted

cfDNA undergoes whole-genome shotgun library construction and hybridization-based capture of 324 cancer-related genes including coding exons and select introns of 309 genes, as well as only select intronic regions or non-coding regions of 15 genes. Hybrid-capture selected libraries are sequenced with deep coverage using the NovaSeq® 6000 platform. Sequence data are processed using a customized analysis pipeline designed to accurately detect genomic alterations, including base substitutions, indels, select copy number variants, and select genomic rearrangements. Substitutions and insertion and deletion alterations (indels) are reported in 311 genes, copy number alterations (CNAs) are reported in 310 genes, and gene rearrangements are reported in 324 genes. The assay also reports tumor fraction, and genomic signatures including MSI and bTMB. A subset of targeted regions in 75 genes is baited for increased sensitivity.

#### THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. *Note:* A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

#### QUALIFIED ALTERATION CALLS (EQUIVOCAL)

All equivocal calls, regardless of alteration type, imply that there is adequate evidence to call the alteration with confidence. However, the repeatability of equivocal calls may be lower than non-equivocal calls.

#### RANKING OF THERAPIES AND CLINICAL TRIALS

##### Ranking of Therapies in Summary Table

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

##### Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

#### LIMITATIONS

1. For *in vitro* diagnostic use.
2. For prescription use only. This test must be ordered by a qualified medical professional in accordance with clinical laboratory regulations.
3. A negative result does not rule out the presence of a mutation below the limits of detection of the assay. Patients for whom no companion diagnostic alterations are detected should be considered for confirmation with an appropriately validated tumor tissue test, if available.
4. The FoundationOne Liquid CDx assay does not detect heterozygous deletions.
5. The test is not intended to provide information on cancer predisposition.
6. Performance has not been validated for cfDNA input below the specified minimum input.
7. Tissue TMB and blood TMB (bTMB) are estimated from the number of synonymous and nonsynonymous single-nucleotide variants (SNVs) and insertions and deletions (indels) per area of coding genome sampled, after the removal of known and likely oncogenic driver events and germline SNPs. Tissue TMB is calculated based on variants with an allele frequency of  $\geq 5\%$ , and bTMB is calculated based on variants with an allele frequency of  $\geq 0.5\%$ .
8. Tumor fraction is the percentage of circulating tumor DNA (ctDNA) present in a cell-free DNA (cfDNA) sample. The tumor fraction estimate is computationally derived from the observed level of aneuploidy in the sample. Tumor fraction is considered elevated when ctDNA levels are high enough that aneuploidy can be detected and is significantly distinct from that typically found in non-tumor samples.
9. Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the tumor genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor. The MSI algorithm is based on genome wide analysis of 1765 microsatellite loci and not based on the 5 or 7 MSI loci described in current clinical practice guidelines for solid tissue testing.
10. Genomic findings from circulating cell-free DNA (cfDNA) may originate from circulating tumor DNA fragments, germline alterations, or non-tumor somatic alterations, such as clonal hematopoiesis of indeterminate potential (CHIP). Genes with alterations that may be derived from CHIP include, but are not limited

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**APPENDIX**
**About FoundationOne® Liquid CDx**

to: *ASXL1*, *ATM*, *CBL*, *CHEK2*, *DNMT3A*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, *TP53*, and *U2AF1*.

11. Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. If a reported alteration is suspected to be germline, confirmatory testing should be considered in the appropriate clinical context.
12. The test is not intended to replace germline testing or to provide information about cancer predisposition.

## REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

## VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >30%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to

distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

## VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *ATM*, *CBL*, *CHEK2*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

## NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) ([www.nccn.org](http://www.nccn.org)). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2021. All rights reserved. To view the most recent and complete version of the guidelines, go online to [NCCN.org](http://NCCN.org). NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

## LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

## NO GUARANTEE OF CLINICAL BENEFIT

This report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

## NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne Liquid CDx.

## TREATMENT DECISIONS ARE THE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this test or the information contained in this report.

Certain sample of variant characteristics may result in reduced sensitivity. These include: low sample quality, deletions and insertions >40bp, or repetitive/high homology sequences. FoundationOne Liquid CDx is performed using cell-free DNA, and as such germline events may not be reported.

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APPENDIX

About FoundationOne®Liquid CDx

**SELECT ABBREVIATIONS**

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
Muts/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

MR Suite Version 5.2.0

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**APPENDIX** **References**

1. Gandara DR, et al. Nat. Med. (2018) PMID: 30082870
2. Wang Z, et al. JAMA Oncol (2019) PMID: 30816954
3. Aggarwal C, et al. Clin. Cancer Res. (2020) PMID: 32102950
4. Li et al., 2020; ASCO Abstract 6511
5. Cerami E, et al. Cancer Discov (2012) PMID: 22588877
6. Gao J, et al. Sci Signal (2013) PMID: 23550210
7. Tate JG, et al. Nucleic Acids Res. (2019) PMID: 30371878
8. Shao C, et al. JAMA Netw Open (2020) PMID: 33119110
9. Reikhtman N, et al. Clin. Cancer Res. (2016) PMID: 26960398
10. Harms PW, et al. Cancer Res. (2015) PMID: 26238782
11. Goh G, et al. Oncotarget (2016) PMID: 26655088
12. Wong SQ, et al. Cancer Res. (2015) PMID: 26627015
13. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
14. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
15. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
16. Rizvi NA, et al. Science (2015) PMID: 25765070
17. Johnson BE, et al. Science (2014) PMID: 24336570
18. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
19. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
20. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
21. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
22. Nature (2012) PMID: 22810696
23. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
24. Bronkhorst AJ, et al. Biomol Detect Quantif (2019) PMID: 30923679
25. Raja R, et al. Clin. Cancer Res. (2018) PMID: 30093454
26. Hrebien S, et al. Ann. Oncol. (2019) PMID: 30860573
27. Choudhury AD, et al. JCI Insight (2018) PMID: 30385733
28. Goodall J, et al. Cancer Discov (2017) PMID: 28450425
29. Goldberg SB, et al. Clin. Cancer Res. (2018) PMID: 29330207
30. Bettegowda C, et al. Sci Transl Med (2014) PMID: 24553385
31. Lapin M, et al. J Transl Med (2018) PMID: 30400802
32. Shulman DS, et al. Br. J. Cancer (2018) PMID: 30131550
33. Stover DG, et al. J. Clin. Oncol. (2018) PMID: 29298117
34. Hemming ML, et al. JCO Precis Oncol (2019) PMID: 30793095
35. Egyud M, et al. Ann. Thorac. Surg. (2019) PMID: 31059681
36. Fan G, et al. PLoS ONE (2017) PMID: 28187169
37. Vu et al., 2020; DOI: 10.1200/PO.19.00204
38. Li G, et al. J Gastrointest Oncol (2019) PMID: 31602320
39. Zhang EW, et al. Cancer (2020) PMID: 32757294
40. Butler TM, et al. Cold Spring Harb Mol Case Stud (2019) PMID: 30833418
41. Mao JH, et al. Science (2008) PMID: 18787170
42. Yang H, et al. Oncotarget (2015) PMID: 25749036
43. Villarruz LC, et al. Lung Cancer (2014) PMID: 24360397
44. Olson D, et al. Clin Genitourin Cancer (2016) PMID: 27079472
45. Kulkarni et al., 2020; <https://doi.org/10.1016/j.jgyno.2020.05.244>
46. Wertz IE, et al. Nature (2011) PMID: 21368834
47. Park HY, et al. Hum. Pathol. (2019) PMID: 30851333
48. Simbolo M, et al. Virchows Arch. (2018) PMID: 30219970
49. Tu K, et al. Hepatol. Res. (2012) PMID: 22548670
50. Iwatsuki M, et al. Int. J. Cancer (2010) PMID: 19739118
51. Yokobori T, et al. Int. J. Oncol. (2012) PMID: 22576686
52. Yokobori T, et al. Cancer Res. (2009) PMID: 19366810
53. Yokobori T, et al. Mol. Cancer Res. (2014) PMID: 24165483
54. Rajagopalan H, et al. Nature (2004) PMID: 14999283
55. Cheng Y, et al. J. Invest. Dermatol. (2013) PMID: 23381582
56. Xu Y, et al. Biomarkers (2016) PMID: 26954701
57. Welcker M, et al. Nat. Rev. Cancer (2008) PMID: 18094723
58. Akhondji S, et al. Cancer Res. (2007) PMID: 17909001
59. Welcker M, et al. Genes Dev. (2013) PMID: 24298052
60. Welcker M, et al. Cell Div (2007) PMID: 17298674
61. Strohmaier H, et al. Nature (2001) PMID: 11565034
62. Pashkova N, et al. Mol. Cell (2010) PMID: 21070969
63. O'Neil J, et al. J. Exp. Med. (2007) PMID: 17646409
64. Malyskova A, et al. Leukemia (2013) PMID: 23228967
65. Thompson BJ, et al. J. Exp. Med. (2007) PMID: 17646408
66. Ou SH, et al. J Thorac Oncol (2011) PMID: 21623265
67. Schwab R, et al. Lung Cancer (2014) PMID: 24192513
68. Le X, et al. Clin Lung Cancer (2015) PMID: 25922291
69. Schrock AB, et al. J Thorac Oncol (2017) PMID: 28315738
70. Lennerz JK, et al. J. Clin. Oncol. (2011) PMID: 22042947
71. Chi AS, et al. J. Clin. Oncol. (2012) PMID: 22162573
72. Palma NA, et al. Case Rep Oncol (2014) PMID: 25232318
73. Schuler et al., 2016; ASCO Abstract 9067
74. Wu et al., 2018; WCLC Abstract P1.01-97
75. Wu YL, et al. J. Clin. Oncol. (2018) PMID: 30156984
76. Gainor JF, et al. J Thorac Oncol (2020) PMID: 31864558
77. Gautschi O, et al. J Thorac Oncol (2020) PMID: 31864554
78. Faivre et al., 2021; ASCO GI Abstract 329
79. Le et al., 2021; ASCO Abstract 9021
80. Yang et al., 2019; AACR Abstract CT193
81. Park et al., 2019; ESMO Abstract 4770
82. Wu et al., 2019; IASLC Abstract MA09.09
83. Gan HK, et al. Clin. Cancer Res. (2019) PMID: 30952639
84. Lee J, et al. Cancer Discov (2019) PMID: 31315834
85. Kim ST, et al. Transl Oncol (2019) PMID: 30695737
86. Kwak et al., 2015; ASCO GI Abstract 01
87. Spigel DR, et al. J. Clin. Oncol. (2013) PMID: 24101053
88. Catenacci DV, et al. Cancer Discov (2011) PMID: 21389872
89. Harding JJ, et al. Clin. Cancer Res. (2019) PMID: 31142504
90. Strickler JH, et al. J. Clin. Oncol. (2018) PMID: 30285518
91. Camidge et al., 2021; AACR Abstract CT179
92. Song J, et al. Arch. Pathol. Lab. Med. (2010) PMID: 21043826
93. Rossi G, et al. J. Clin. Oncol. (2005) PMID: 16314638
94. Miao L, et al. Oncotarget (2017) PMID: 28903317
95. J. Clin. Oncol. (2011) PMID: 22042966
96. Jung KH, et al. Arch. Pharm. Res. (2012) PMID: 22553051
97. Ang CS, et al. Anticancer Res. (2013) PMID: 23898085
98. Abou-Bakr AA, et al. Gulf J Oncolog (2013) PMID: 23996864
99. Ho JC, et al. Semin Respir Crit Care Med (2013) PMID: 24258573
100. Dziadziszko R, et al. J Thorac Oncol (2012) PMID: 22237262
101. Madoz-Gúrpide J, et al. J Transl Med (2015) PMID: 26319934
102. Ali SM, et al. Oncologist (2015) PMID: 25882375
103. Kwak EL, et al. Cancer Discov (2015) PMID: 26432108
104. Basho RK, et al. JAMA Oncol (2017) PMID: 27893038
105. Hirai H, et al. Mol. Cancer Ther. (2010) PMID: 20571069
106. Banck MS, et al. J. Clin. Invest. (2013) PMID: 23676460
107. Liu AX, et al. Cancer Res. (1998) PMID: 9679957
108. Vivanco I, et al. Nat. Rev. Cancer (2002) PMID: 12094235
109. Chin YR, et al. Cell Adh Migr ( ) PMID: 21519185
110. Cheng JQ, et al. Proc. Natl. Acad. Sci. U.S.A. (1992) PMID: 14096633
111. Thompson FH, et al. Cancer Genet. Cytogenet. (1996) PMID: 8646743
112. Altomare DA, et al. Oncogene (2005) PMID: 16288292
113. Int. J. Biol. Markers ( ) PMID: 18409144
114. Scrima M, et al. PLoS ONE (2012) PMID: 22363436
115. Lin AB, et al. Clin. Cancer Res. (2017) PMID: 28331049
116. Chen X, et al. Clin Cancer Res (2018) PMID: 30181387
117. Mörry T, et al. Int. J. Biochem. Cell Biol. (2004) PMID: 15147722
118. Lee JM, et al. Lancet Oncol. (2018) PMID: 29361470
119. Lheureux S, et al. Lancet (2021) PMID: 33485453
120. Oza AM, et al. Clin Cancer Res (2020) PMID: 32611648
121. Fu et al., 2021; AACR abstract 974
122. Toledo LI, et al. Nat. Struct. Mol. Biol. (2011) PMID: 21552262
123. Buisson R, et al. Mol. Cell (2015) PMID: 26365377
124. Yang L, et al. Oncotarget (2015) PMID: 26204491
125. Kok YP, et al. Oncogenesis (2020) PMID: 33028815
126. Taylor-Harding B, et al. Oncotarget (2015) PMID: 25557169
127. Etemadmoghadam D, et al. Clin. Cancer Res. (2013) PMID: 24004674
128. Scaltriti M, et al. Proc. Natl. Acad. Sci. U.S.A. (2011) PMID: 21321214
129. Nanos-Webb A, et al. Breast Cancer Res. Treat. (2012) PMID: 21695458
130. Ma T, et al. Mol. Cancer Ther. (2013) PMID: 23686769
131. Zakka K, et al. Oncotarget (2020) PMID: 32477464
132. Grabowski P, et al. Clin. Cancer Res. (2008) PMID: 19010853
133. Salton C, et al. Oncogene (2007) PMID: 17471231
134. Liu SZ, et al. Asian Pac. J. Cancer Prev. (2013) PMID: 23534765
135. Leung SY, et al. Mod. Pathol. (2006) PMID: 16575401
136. Lin L, et al. Cancer Res. (2000) PMID: 11156406
137. Mayr D, et al. Am. J. Clin. Pathol. (2006) PMID: 16753589
138. Nakayama N, et al. Cancer (2010) PMID: 20336784
139. Stamatakis M, et al. World J Surg Oncol (2010) PMID: 21176227
140. Wilson MA, et al. Clin. Cancer Res. (2016) PMID: 26307133
141. Lei J, et al. Oncotarget (2016) PMID: 26981887
142. Mehnert et al., 2016; EORTC-NCI-AACR Abstract 435
143. Pacaud A, et al. Eur J Cancer (2021) PMID: 33684875
144. Nakama K, et al. J Dermatol (2021) PMID: 33768587
145. McEvoy CR, et al. J. Clin. Invest. (2019) PMID: 30835257
146. Touat et al., 2019; DOI: 10.1200/PO.18.00298
147. Yde CW, et al. Cancer Genet (2016) PMID: 27810072
148. George J, et al. Nature (2015) PMID: 26168399
149. Peifer M, et al. Nat. Genet. (2012) PMID: 22941188
150. Rudin CM, et al. Nat. Genet. (2012) PMID: 22941189
151. Gollob JA, et al. Semin. Oncol. (2006) PMID: 16890795
152. Maurer G, et al. Oncogene (2011) PMID: 21577205
153. Zack TI, et al. Nat. Genet. (2013) PMID: 24071852
154. Beroukhir R, et al. Nature (2010) PMID: 20164920
155. Tran NH, et al. J. Biol. Chem. (2003) PMID: 12551923
156. Stanton VP, et al. Mol. Cell. Biol. (1989) PMID: 2710120
157. Palanisamy N, et al. Nat. Med. (2010) PMID: 20526349
158. Jones DT, et al. Oncogene (2009) PMID: 19363522
159. Konecny GE, et al. Clin. Cancer Res. (2011) PMID: 21278246

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**APPENDIX**
**References**

160. Katsumi Y, et al. Biochem. Biophys. Res. Commun. (2011) PMID: 21871868
161. Cen L, et al. Neuro-oncology (2012) PMID: 22711607
162. Logan JE, et al. Anticancer Res. (2013) PMID: 23898052
163. Elvin JA, et al. Oncologist (2017) PMID: 28283584
164. Gao J, et al. Curr Oncol (2015) PMID: 26715889
165. Gopalan et al., 2014; ASCO Abstract 8077
166. Peguero et al., 2016; ASCO Abstract 2528
167. Konecny et al., 2016; ASCO Abstract 5557
168. DeMichele A, et al. Clin. Cancer Res. (2015) PMID: 25501126
169. Finn RS, et al. Lancet Oncol. (2015) PMID: 25524798
170. Infante JR, et al. Clin. Cancer Res. (2016) PMID: 27542767
171. Johnson DB, et al. Oncologist (2014) PMID: 24797823
172. Lubomierski N, et al. Cancer Res. (2001) PMID: 11479232
173. Chaussade L, et al. Oncogene (2001) PMID: 11641784
174. Moore PS, et al. Br. J. Cancer (2001) PMID: 11161385
175. Dacic S, et al. Hum. Pathol. (2002) PMID: 12378519
176. Virmani AK, et al. Genes Chromosomes Cancer (1998) PMID: 9559342
177. Gazzeri S, et al. Cancer Res. (1998) PMID: 9731504
178. Arnold CN, et al. Int. J. Cancer (2008) PMID: 18646189
179. Toyooka S, et al. Mol. Cancer Ther. (2001) PMID: 12467239
180. Muscarella P, et al. Cancer Res. (1998) PMID: 9443399
181. Houben R, et al. Exp. Dermatol. (2009) PMID: 19400830
182. Lassacher A, et al. J. Invest. Dermatol. (2008) PMID: 18219279
183. Simon B, et al. Ann. N. Y. Acad. Sci. (2004) PMID: 15153447
184. House MG, et al. Ann. Surg. (2003) PMID: 14501508
185. Quelle DE, et al. Cell (1995) PMID: 8521522
186. Mutat. Res. (2005) PMID: 15878778
187. Gazzeri S, et al. Oncogene (1998) PMID: 9484839
188. Oncogene (1999) PMID: 10498883
189. Sherr CJ, et al. Cold Spring Harb. Symp. Quant. Biol. (2005) PMID: 16869746
190. Ozanne P, et al. Int. J. Cancer (2010) PMID: 20549699
191. Ruas M, et al. Oncogene (1999) PMID: 10498896
192. Jones R, et al. Cancer Res. (2007) PMID: 17909018
193. Haferkamp S, et al. Aging Cell (2008) PMID: 18843795
194. Huot TJ, et al. Mol. Cell. Biol. (2002) PMID: 12417717
195. Rizos H, et al. J. Biol. Chem. (2001) PMID: 11518711
196. Gombart AF, et al. Leukemia (1997) PMID: 9324288
197. Yang R, et al. Cancer Res. (1995) PMID: 7780957
198. Parry D, et al. Mol. Cell. Biol. (1996) PMID: 8668202
199. Greenblatt MS, et al. Oncogene (2003) PMID: 12606942
200. Yarbrough WG, et al. J. Natl. Cancer Inst. (1999) PMID: 10491434
201. Poi MJ, et al. Mol. Carcinog. (2001) PMID: 11255261
202. Byeon IJ, et al. Mol. Cell (1998) PMID: 9660926
203. Kannengiesser C, et al. Hum. Mutat. (2009) PMID: 19260062
204. Lal G, et al. Genes Chromosomes Cancer (2000) PMID: 10719365
205. Koh J, et al. Nature (1995) PMID: 7777061
206. McKenzie HA, et al. Hum. Mutat. (2010) PMID: 20340136
207. Miller PJ, et al. Hum. Mutat. (2011) PMID: 21462282
208. Kutschers CL, et al. Physiol. Behav. (1977) PMID: 905385
209. Scaini MC, et al. Hum. Mutat. (2014) PMID: 24659262
210. Jenkins NC, et al. J. Invest. Dermatol. (2013) PMID: 23190892
211. Walker GJ, et al. Int. J. Cancer (1999) PMID: 10389768
212. Rutter JL, et al. Oncogene (2003) PMID: 12853981
213. Whelan AJ, et al. N Engl J Med (1995) PMID: 7666917
214. Adv Exp Med Biol (2010) PMID: 20687502
215. Hogg D, et al. J Cutan Med Surg (1998) PMID: 9479083
216. De Unamuno B, et al. Melanoma Res (2018) PMID: 29543703
217. Soura E, et al. J Am Acad Dermatol (2016) PMID: 26892650
218. Huerta C, et al. Acta Derm Venereol (2018) PMID: 29405243
219. Kaufman DK, et al. Neurology (1993) PMID: 8414022
220. Bahuau M, et al. Cancer Res (1998) PMID: 9622062
221. Chan AK, et al. Clin Neuropathol ( ) PMID: 28699883
222. Black LE, et al. Am. J. Pathol. (2019) PMID: 31351986
223. Baselga J, et al. Nat. Rev. Cancer (2009) PMID: 19536107
224. Jaiswal BS, et al. Cancer Cell (2013) PMID: 23680147
225. Jura N, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) PMID: 20007378
226. Choudhury NJ, et al. J. Clin. Oncol. (2016) PMID: 27044931
227. Verlingue L, et al. Eur. J. Cancer (2018) PMID: 29413684
228. Bidard FC, et al. Ann. Oncol. (2015) PMID: 25953157
229. Umelo I, et al. Oncotarget (2016) PMID: 26689995
230. Mishra R, et al. Oncotarget (2018) PMID: 29963236
231. Perez EA, et al. BMC Cancer (2019) PMID: 31146717
232. Baselga J, et al. J. Clin. Oncol. (2014) PMID: 25332247
233. Kim SB, et al. Int. J. Cancer (2016) PMID: 27428671
234. Baselga J, et al. Clin. Cancer Res. (2016) PMID: 26920887
235. Shah MA, et al. Gastric Cancer (2019) PMID: 30706247
236. Kurzeder C, et al. J. Clin. Oncol. (2016) PMID: 27269942
237. Cohen EE, et al. Ann. Oncol. (2017) PMID: 28961833
238. LaBonte MJ, et al. Mol. Cancer Ther. (2016) PMID: 27325685
239. Nishimura R, et al. Oncology (2017) PMID: 28478451
240. Duchnowska R, et al. Oncotarget (2017) PMID: 29262628
241. Fabi A, et al. Expert Opin Pharmacother (2013) PMID: 23472669
242. Beltran H, et al. Nat. Med. (2016) PMID: 26855148
243. Jiao Y, et al. Science (2011) PMID: 21252315
244. Cao Y, et al. Nat Commun (2013) PMID: 24326773
245. Sheng Q, et al. Br. J. Cancer (2011) PMID: 21364581
246. Sassen A, et al. Breast Cancer Res. (2008) PMID: 18182100
247. Nature (2012) PMID: 22960745
248. Augert A, et al. J Thorac Oncol (2017) PMID: 28007623
249. Ardeshir-Larijani F, et al. Clin Lung Cancer (2018) PMID: 29627316
250. Hillman RT, et al. Nat Commun (2018) PMID: 29950560
251. Abudurehman A, et al. J. Cancer Res. Clin. Oncol. (2018) PMID: 29532228
252. Vicent GP, et al. Genes Dev. (2011) PMID: 21447625
253. Hannibal MC, et al. Am. J. Med. Genet. A (2011) PMID: 21671394
254. Fagan RJ, et al. Cancer Lett. (2019) PMID: 31128216
255. Jaiswal S, et al. N. Engl. J. Med. (2014) PMID: 25426837
256. Genovesi G, et al. N. Engl. J. Med. (2014) PMID: 25426838
257. Xie M, et al. Nat. Med. (2014) PMID: 25326804
258. Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) PMID: 28669404
259. Severson EA, et al. Blood (2018) PMID: 29678827
260. Fuster JJ, et al. Circ. Res. (2018) PMID: 29420212
261. Chabon JJ, et al. Nature (2020) PMID: 32269342
262. Razavi P, et al. Nat. Med. (2019) PMID: 31768066
263. Hirai H, et al. Cancer Biol. Ther. (2010) PMID: 20107315
264. Bridges KA, et al. Clin. Cancer Res. (2011) PMID: 21799033
265. Rajeshkumar NV, et al. Clin. Cancer Res. (2011) PMID: 21389100
266. Osman AA, et al. Mol. Cancer Ther. (2015) PMID: 25504633
267. Xu L, et al. Mol. Cancer Ther. (2002) PMID: 12489850
268. Xu L, et al. Mol. Med. (2001) PMID: 11713371
269. Camp ER, et al. Cancer Gene Ther. (2013) PMID: 23470564
270. Kim SS, et al. Nanomedicine (2015) PMID: 25240597
271. Pirolo KF, et al. Mol. Ther. (2016) PMID: 27357628
272. Hajdenberg et al., 2012; ASCO Abstract e15010
273. Leijen S, et al. J. Clin. Oncol. (2016) PMID: 27601554
274. Moore et al., 2019; ASCO Abstract 5513
275. Leijen S, et al. J. Clin. Oncol. (2016) PMID: 27998224
276. Oza et al., 2015; ASCO Abstract 5506
277. Méndez E, et al. Clin. Cancer Res. (2018) PMID: 29535125
278. Seligmann JF, et al. J Clin Oncol (2021) PMID: 34538072
279. Kwok M, et al. Blood (2016) PMID: 26563132
280. Boudny M, et al. Haematologica (2019) PMID: 30975914
281. Dillon MT, et al. Mol. Cancer Ther. (2017) PMID: 28062704
282. Middleton FK, et al. Cancers (Basel) (2018) PMID: 30127241
283. Fernandez-Cuesta L, et al. Nat Commun (2014) PMID: 24670920
284. Higaki-Mori H, et al. Hum. Pathol. (2012) PMID: 22795182
285. Rodig SJ, et al. J. Clin. Invest. (2012) PMID: 23114601
286. Takahashi T, et al. Oncogene (1991) PMID: 1656362
287. Chen H, et al. Endocr. Relat. Cancer (2012) PMID: 22389383
288. Wistuba II, et al. Gynecol. Oncol. (1999) PMID: 9889022
289. Tan HL, et al. Clin. Cancer Res. (2014) PMID: 24323898
290. Yachida S, et al. Am. J. Surg. Pathol. (2012) PMID: 22251937
291. Kobayashi Y, et al. Cancer Sci. (2004) PMID: 15072592
292. Przygodzki RM, et al. Am. J. Pathol. (1996) PMID: 8623922
293. Onuki N, et al. Cancer (1999) PMID: 10091733
294. O'Toole D, et al. Endocr. Relat. Cancer (2010) PMID: 20570957
295. Erler BS, et al. Tumour Biol. (2011) PMID: 21058037
296. Safatle-Ribeiro AV, et al. Eur J Gastroenterol Hepatol (2007) PMID: 17206073
297. Brown CJ, et al. Nat. Rev. Cancer (2009) PMID: 19935675
298. Joerger AC, et al. Annu. Rev. Biochem. (2008) PMID: 18410249
299. Kato S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) PMID: 12826609
300. Kamada R, et al. J. Biol. Chem. (2011) PMID: 20978130
301. Zerdoumi Y, et al. Hum. Mol. Genet. (2017) PMID: 28472496
302. Yamada H, et al. Carcinogenesis (2007) PMID: 17690113
303. Pavel M, et al. Ann Oncol (2020) PMID: 32272208
304. Baudin E, et al. Ann Oncol (2021) PMID: 33482246
305. Rindi G, et al. Mod Pathol (2018) PMID: 30140036
306. Nagtegaal ID, et al. Histopathology (2020) PMID: 31433515
307. Bougeard G, et al. J. Clin. Oncol. (2015) PMID: 26014290
308. Sorrell AD, et al. Mol Diagn Ther (2013) PMID: 23355100
309. Nichols KE, et al. Cancer Epidemiol. Biomarkers Prev. (2001) PMID: 11219776
310. Kleihues P, et al. Am. J. Pathol. (1997) PMID: 9006316
311. Gonzalez KD, et al. J. Clin. Oncol. (2009) PMID: 19204208
312. Lalloo F, et al. Lancet (2003) PMID: 12672316
313. Mandelker D, et al. Ann. Oncol. (2019) PMID: 31050713

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**APPENDIX**
**References**

314. Hematology Am Soc Hematol Educ Program (2018) pmid: 30504320
315. Paik PK, et al. Cancer Discov (2015) pmid: 25971939
316. Klempner SJ, et al. J Thorac Oncol (2017) pmid: 27693535
317. Yakes FM, et al. Mol. Cancer Ther. (2011) pmid: 21926191
318. Weber H, et al. J Biomol Screen (2014) pmid: 25260782
319. Navis AC, et al. PLoS ONE (2013) pmid: 23484006
320. Yeh I, et al. Nat Commun (2015) pmid: 26013381
321. Lee YH, et al. Cancers (Basel) (2014) pmid: 25534569
322. Torres KE, et al. Clin. Cancer Res. (2011) pmid: 21540237
323. Sameni M, et al. Clin. Cancer Res. (2016) pmid: 26432786
324. Schöffski P, et al. Eur. J. Cancer (2017) pmid: 29059635
325. Hellerstedt BA, et al. Clin Lung Cancer (2019) pmid: 30528315
326. Nokihara H, et al. Clin Lung Cancer (2019) pmid: 30718102
327. Elisei R, et al. J. Clin. Oncol. (2013) pmid: 24002501
328. Abou-Alfa GK, et al. N. Engl. J. Med. (2018) pmid: 29972759
329. Choueiri TK, et al. J. Clin. Oncol. (2017) pmid: 28199818
330. Italiano A, et al. Lancet Oncol. (2020) pmid: 32078813
331. Wu et al., 2018; WLCL Abstract P1.01-97
332. Wolf et al., 2020; ASCO Abstract 9509
333. Schuler M, et al. Ann. Oncol. (2020) pmid: 32240796
334. Wolf J, et al. N Engl J Med (2020) pmid: 32877583
335. van den Bent M, et al. J. Neurooncol. (2020) pmid: 31776899
336. Bang YJ, et al. Cancer Sci. (2020) pmid: 31778267
337. Esaki T, et al. Cancer Sci. (2019) pmid: 30724423
338. Vassal et al., 2015; ASCO Abstract 2595
339. Li et al., 2015; ASCO Abstract 8090
340. Frampton GM, et al. Cancer Discov (2015) pmid: 25971938
341. Benderra MA, et al. J Thorac Oncol (2016) pmid: 26845121
342. Waqar SN, et al. J Thorac Oncol (2015) pmid: 25898962
343. Mendenhall MA, et al. J Thorac Oncol (2015) pmid: 25898965
344. Jenkins RW, et al. Clin Lung Cancer (2015) pmid: 25769807
345. Stein MN, et al. Eur. Urol. (2015) pmid: 25457019
346. Awad et al., 2017; ASCO Abstract 8511
347. Wang S, et al. Clin Lung Cancer (2021) pmid: 32651063
348. Nakajima M, et al. Intern Med () pmid: 27803410
349. Shaw et al., 2016; ASCO Abstract 9066
350. Lu et al., 2016; ASCO Abstract 9058
351. Yoshida T, et al. J. Clin. Oncol. (2016) pmid: 27354483
352. Solomon BJ, et al. N. Engl. J. Med. (2014) pmid: 25470694
353. Shaw AT, et al. N. Engl. J. Med. (2013) pmid: 23724913
354. Moro-Sibilot et al., 2015; ASCO Abstract 8065
355. Goto et al., 2016; ASCO Abstract 9022
356. Shaw AT, et al. N. Engl. J. Med. (2014) pmid: 25264305
357. Mazières J, et al. J. Clin. Oncol. (2015) pmid: 25667280
358. Scheffler M, et al. Oncotarget (2015) pmid: 25868855
359. Vaishnavi A, et al. Nat. Med. (2013) pmid: 24162815
360. Drilon et al., 2016; ASCO Abstract 108
361. Camidge et al., 2014; ASCO Abstract 8001
362. Schrock AB, et al. J Thorac Oncol (2016) pmid: 27343443
363. Jorge SE, et al. Lung Cancer (2015) pmid: 26791794
364. Mahjoubi L, et al. Invest New Drugs (2016) pmid: 26892698
365. Awad MM, et al. J. Clin. Oncol. (2016) pmid: 26729443
366. Zhang Y, et al. J Thorac Oncol (2016) pmid: 26724472
367. Diamond JR, et al. J. Clin. Oncol. (2013) pmid: 23610116
368. Gambacorti-Passerini et al., 2017; ASH Abstract 4128
369. Mossé YP, et al. Lancet Oncol. (2013) pmid: 23598171
370. Kulkarni et al., 2020; DOI: 10.1016/j.jgyno.2020.05.244
371. Yao JC, et al. N. Engl. J. Med. (2011) pmid: 21306238
372. Yao JC, et al. Lancet (2016) pmid: 26703889
373. Bajetta et al., 2016; ASCO Abstract 4092
374. Pavel et al., 2017; Pavel et al.
375. Pavel ME, et al. Lancet (2011) pmid: 22119496
376. Fazio N, et al. Chest (2013) pmid: 23187897
377. Yao JC, et al. J. Clin. Oncol. (2010) pmid: 19933912
378. Tolcher AW, et al. Ann. Oncol. (2015) pmid: 25344362
379. Patterson et al., 2018; AACR Abstract 3891
380. Duran I, et al. Br. J. Cancer (2006) pmid: 17031397
381. Hobday et al., 2013; ASCO Abstract 4032
382. Ganesan P, et al. Invest New Drugs (2013) pmid: 23982248
383. Hobday TJ, et al. J. Clin. Oncol. (2015) pmid: 25488966
384. Pandya KJ, et al. J Thorac Oncol (2007) pmid: 17975496
385. Paik PK, et al. N. Engl. J. Med. (2020) pmid: 32469185
386. Mazieres et al., 2020; ESMO Abstract 1283P
387. Ryoo et al., 2018; ESMO Abstract 186P
388. Ryoo et al., 2018; ESMO Abstract 621PD
389. Decaens et al., 2019; ESMO Abstract 698P
390. Falchook GS, et al. Clin. Cancer Res. (2020) pmid: 31822497
391. Shitara K, et al. Jpn. J. Clin. Oncol. (2020) pmid: 32328660

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