

**ABOUT THE TEST** FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

<b>PATIENT</b>	<b>DISEASE</b> Liver hepatocellular carcinoma (HCC)	<b>PHYSICIAN</b>	<b>ORDERING PHYSICIAN</b> Yeh, Yi-Chen	<b>SPECIMEN</b>	<b>SPECIMEN SITE</b> Soft Tissue
	<b>NAME</b> Su, Tung-Fa		<b>MEDICAL FACILITY</b> Taipei Veterans General Hospital		<b>SPECIMEN ID</b> S110-39510 A (PF22041)
	<b>DATE OF BIRTH</b> 29 May 1962		<b>ADDITIONAL RECIPIENT</b> None		<b>SPECIMEN TYPE</b> Slide Deck
	<b>SEX</b> Male		<b>MEDICAL FACILITY ID</b> 205872		<b>DATE OF COLLECTION</b> 14 December 2021
	<b>MEDICAL RECORD #</b> 47952034		<b>PATHOLOGIST</b> Not Provided		<b>SPECIMEN RECEIVED</b> 25 March 2022

## Biomarker Findings

**Microsatellite status** - MS-Stable  
**Tumor Mutational Burden** - 3 Muts/Mb

## Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

**ARID1A** G931\*

**CTNNB1** D32Y

**KEAP1** G477S - subclonal†

**TERT** promoter -124C>T

† See About the Test in appendix for details.

## Report Highlights

- Evidence-matched **clinical trial options** based on this patient's genomic findings: (p. 7)

### BIOMARKER FINDINGS

**Microsatellite status** - MS-Stable

**Tumor Mutational Burden** - 3 Muts/Mb

### GENOMIC FINDINGS

**ARID1A** - G931\*

8 Trials see p. 7

**CTNNB1** - D32Y

7 Trials see p. 9

### THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. see Biomarker Findings section

No therapies or clinical trials. see Biomarker Findings section

#### THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)

none

none

#### THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)

none

none

### GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

**KEAP1** - G477S - subclonal ..... p. 5    **TERT** - promoter -124C>T ..... p. 6

**NOTE** Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

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Electronically signed by Erik Williams, M.D. | 08 April 2022  
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1328300-01

## BIOMARKER FINDINGS

## BIOMARKER

## Microsatellite status

## RESULT

MS-Stable

### POTENTIAL TREATMENT STRATEGIES

#### — Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors<sup>1-3</sup>, including approved therapies nivolumab and pembrolizumab<sup>4</sup>. In a retrospective analysis of 361 patients with solid tumors treated

with pembrolizumab, 3% were MSI-H and experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%,  $p=0.001$ )<sup>5</sup>.

### FREQUENCY & PROGNOSIS

MSI-H was not detected in a study of 122 hepatocellular (HCC) samples<sup>6</sup>, although smaller studies have reported MSI in 0-18% of tumors<sup>7-11</sup>, and MSI at some level has been detected in a subset of HCC tumors<sup>6</sup>. The prognostic significance of MSI in HCC has not been determined (PubMed, Jun 2021).

### FINDING SUMMARY

Microsatellite instability (MSI) is a condition of

genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor<sup>12</sup>. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2<sup>12-14</sup>. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers<sup>15-17</sup>. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins<sup>12,14,16-17</sup>.

## BIOMARKER

## Tumor Mutational Burden

## RESULT

3 Muts/Mb

### POTENTIAL TREATMENT STRATEGIES

#### — Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1<sup>18-20</sup>, anti-PD-1 therapies<sup>18-21</sup>, and combination nivolumab and ipilimumab<sup>22-27</sup>. In multiple pan-tumor studies, higher TMB has been reported to be associated with increased ORR and OS from treatment with immune checkpoint inhibitors<sup>18-21,28</sup>. Higher TMB was found to be significantly associated with improved OS upon immune checkpoint inhibitor treatment for patients with 9 types of advanced tumors<sup>18</sup>. Analyses across several solid tumor types reported that patients with higher TMB (defined as  $\geq 16-20$

Muts/Mb) achieved greater clinical benefit from PD-1 or PD-L1-targeting monotherapy, compared with patients with higher TMB treated with chemotherapy<sup>29</sup> or those with lower TMB treated with PD-1 or PD-L1-targeting agents<sup>19</sup>. However, the KEYNOTE 158 trial of pembrolizumab monotherapy for patients with solid tumors found significant improvement in ORR for patients with TMB  $\geq 10$  Muts/Mb (based on this assay or others) compared to those with TMB  $< 10$  Muts/Mb, in a large cohort that included multiple tumor types; similar findings were observed in the KEYNOTE 028 and 012 trials<sup>21,28</sup>. Together, these studies suggest that patients with TMB  $\geq 10$  Muts/Mb may derive clinical benefit from PD-1 or PD-L1 inhibitors.

### FREQUENCY & PROGNOSIS

Hepatocellular carcinoma (HCC) harbors a median TMB of 3.6 mutations per megabase (mut/Mb), and 1% of cases have high TMB ( $> 20$  muts/Mb)<sup>30</sup>. In an analysis of the TCGA Liver HCC dataset, high TMB was associated with reduced PFS and OS<sup>31</sup>. A retrospective study of 128 patients with HCC who underwent curative resection reported decreased recurrence-free survival for patients

with high TMB ( $> 4.8$  Muts/Mb) compared to those with low TMB ( $\leq 4.8$  Muts/Mb) measured in tissue samples<sup>32</sup>.

### FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma<sup>33-34</sup> and cigarette smoke in lung cancer<sup>35-36</sup>, treatment with temozolomide-based chemotherapy in glioma<sup>37-38</sup>, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes<sup>39-43</sup>, and microsatellite instability (MSI)<sup>39,42-43</sup>. This sample harbors a TMB level associated with lower rates of clinical benefit from treatment with PD-1- or PD-L1-targeting immune checkpoint inhibitors compared with patients with tumors harboring higher TMB levels, based on several studies in multiple solid tumor types<sup>19-20,28</sup>.

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ORDERED TEST # ORD-1328300-01

**GENOMIC FINDINGS**
**GENE**
**ARID1A**
**ALTERATION**

G931\*

**TRANSCRIPT ID**

NM\_006015

**CODING SEQUENCE EFFECT**

2791G&gt;T

**VARIANT ALLELE FREQUENCY (% VAF)**

30.3%

**POTENTIAL TREATMENT STRATEGIES**
**— Targeted Therapies —**

There are no therapies approved to address the mutation or loss of ARID1A in cancer. However, on the basis of limited clinical and preclinical evidence, ARID1A inactivating mutations may lead to sensitivity to ATR inhibitors such as M6620 and ceralasertib<sup>44</sup>. In a Phase 2 study of ceralasertib in solid tumors, 2 patients with endometrial carcinoma in the cohort with loss of ARID1A expression achieved CRs on ceralasertib monotherapy; at least 1 of these 2 patients carried an inactivating ARID1A mutation. In contrast, no responses were observed for patients with normal ARID1A expression treated with ceralasertib combined with olaparib<sup>45</sup>. One patient with small cell lung cancer harboring an ARID1A mutation

experienced a PR when treated with M6620 combined with topotecan<sup>46</sup>. In a Phase 1 trial, a patient with metastatic colorectal cancer harboring both an ARID1A mutation and ATM loss treated with single-agent M6620 achieved a CR that was ongoing at 29 months<sup>47</sup>. On the basis of limited preclinical evidence from studies in ovarian cancer, ARID1A inactivation may predict sensitivity to EZH2 inhibitors<sup>48-49</sup>, which are under investigation in clinical trials. Other studies have reported that the loss of ARID1A may activate the PI3K-AKT pathway and be linked with sensitivity to inhibitors of this pathway<sup>50-52</sup>. Patients with ARID1A alterations in advanced or metastatic solid tumors may derive benefit from treatment with anti-PD-1 or anti-PD-L1 immunotherapy<sup>53</sup>. Loss of ARID1A expression has been associated with chemoresistance to platinum-based therapy for patients with ovarian clear cell carcinoma<sup>54-55</sup> and to 5-fluorouracil in colorectal cancer cell lines<sup>56</sup>.

**FREQUENCY & PROGNOSIS**

ARID1A alterations are particularly prevalent in ovarian clear cell carcinoma (46-50%), ovarian and uterine endometrioid carcinomas (24-44%), and cholangiocarcinoma (27%); they are also reported in up to 27% of gastric carcinoma, esophageal adenocarcinoma, Waldenstrom macroglobulinemia, pediatric Burkitt lymphoma, hepatocellular carcinoma, colorectal carcinoma,

and urothelial carcinoma samples analyzed (COSMIC, cBioPortal, Jan 2022)<sup>57-65</sup>. ARID1A loss is associated with microsatellite instability in ovarian and endometrial endometrioid adenocarcinomas<sup>53,66-69</sup>, CRC<sup>53,70-72</sup>, and gastric cancer<sup>53,73-77</sup>. ARID1A protein loss is associated with tumors of poor histological grade for many tumor types, including colorectal cancer (CRC)<sup>70-72</sup>, cervical cancer<sup>78-79</sup>, gastric cancer<sup>73-77</sup>, urothelial carcinoma<sup>80-82</sup>, ovarian and endometrial cancers<sup>55,66-69,83-87</sup>, breast carcinoma<sup>88-90</sup>, and clear cell renal cell carcinoma<sup>91</sup>; ARID1A mutation has been associated with poor outcomes for patients with cholangiocarcinoma<sup>92-95</sup>. However, prognostic data regarding patient survival are often mixed and conflicting.

**FINDING SUMMARY**

ARID1A encodes the AT-rich interactive domain-containing protein 1A, also known as Baf250a, a member of the SWI/SNF chromatin remodeling complex. Mutation, loss, or inactivation of ARID1A has been reported in many cancers, and the gene is considered a tumor suppressor<sup>61,76,89,96-101</sup>. ARID1A mutations, which are mostly truncating, have been identified along the entire gene and often correlate with ARID1A protein loss<sup>61,74,97-98,102</sup>, whereas ARID1A missense mutations are mostly uncharacterized.

ORDERED TEST # ORD-1328300-01

## GENOMIC FINDINGS

## GENE

## CTNNB1

## ALTERATION

D32Y

## TRANSCRIPT ID

NM\_001904

## CODING SEQUENCE EFFECT

94G&gt;T

## VARIANT ALLELE FREQUENCY (% VAF)

30.6%

## POTENTIAL TREATMENT STRATEGIES

## — Targeted Therapies —

Mutation or activation of CTNNB1 signaling has been shown to increase mTOR signaling, promote tumorigenesis, and respond to mTOR inhibition in preclinical studies<sup>103-105</sup>. Small studies have reported clinical benefit following treatment of everolimus combined with other targeted agents for patients with CTNNB1-mutated hepatocellular carcinoma<sup>106-107</sup> or endometrial carcinoma<sup>108</sup>. In preclinical studies, CTNNB1 activating mutations have been shown to increase expression of WNT pathway member DKK1, which may promote tumor cell proliferation and immune evasion<sup>109-111</sup>. A Phase 1 trial of DKK1-targeting antibody DKN-01 in combination with paclitaxel in esophageal cancer reported a PR rate in 2 out of 4 patients and SD rate of in 1 out of 4 patients with CTNNB1 activating mutations, compared with 24% (10/41) PR and 37% (15/41) SD in unselected patients<sup>112</sup>. Multiple preclinical studies in cancer

models harboring CTNNB1 mutation or beta-catenin pathway activation have reported activation of the NOTCH pathway and sensitivity to pharmacologic inhibition of NOTCH signaling by gamma-secretase inhibitors<sup>113-116</sup>. Phase 1 and 2 clinical trials of gamma-secretase inhibitor PF-03084014 have shown high response rates in patients with desmoid tumors, which are driven by activating CTNNB1 mutations in the majority of cases<sup>117-118</sup>, suggesting CTNNB1-mutated tumors may be sensitive to gamma-secretase inhibitors. Although WNT pathway inhibitors have been explored preclinically in CTNNB1-mutated cells, clinical data supporting this therapeutic approach are lacking<sup>104,119-121</sup>.

## — Potential Resistance —

A preclinical study reported that expression of activated mutated CTNNB1 led to resistance to anti-PD-1 therapy in a mouse model of hepatocellular carcinoma (HCC)<sup>122</sup>. However, clinical studies have reported conflicting results. In one study of immune checkpoint inhibitors (ICPI) as treatment for HCC, 7/7 patients with CTNNB1 alterations experienced PD; additionally, patients with tumors with WNT pathway activation (including alterations in CTNNB1, AXIN1, AXIN2, or APC) experienced significantly shorter median PFS (2.0 vs. 7.4 months, HR=9.2) and numerically shorter median OS (9.1 vs. 15.2 months, HR=2.6, p=0.11) compared to those with non-WNT-pathway-altered tumors<sup>123</sup>. In contrast, another clinical study treating HCC with ICPI reported similar PD rates in patients with or without CTNNB1 alterations (2/4 vs. 13/24), as

well as no significant differences in median PFS or OS between patients with or without mutations in WNT pathway genes (PFS 2.5 vs 3.1 months, p=0.32; OS 11.5 vs 16.5 months, p=0.57)<sup>124</sup>. Whether CTNNB1 alterations promote innate resistance to ICPI therefore remains unclear.

## FREQUENCY &amp; PROGNOSIS

CTNNB1 mutations have been reported in 10-18% of hepatocellular carcinomas (HCC) in general, with one study identifying CTNNB1 as the most frequently mutated oncogene in HCC<sup>125-127</sup>. Some studies have reported that CTNNB1 mutation correlates with favorable prognosis in patients with HCC<sup>128-129</sup>, but another study suggested that beta-catenin nuclear accumulation, which was correlated with CTNNB1 exon 3 mutations, is associated with poor outcome in patients with this tumor type<sup>130</sup>. Solid tumors with WNT/beta-catenin pathway alterations, as seen here, were observed to have significantly less T-cell inflammation in one study<sup>131</sup>.

## FINDING SUMMARY

CTNNB1 encodes beta-catenin, a key downstream component of the WNT signaling pathway. Beta-catenin interacts with cadherin to regulate cell-cell adhesion; as a component of the WNT pathway, it also plays a role in development, cell proliferation, and cell differentiation<sup>132</sup>. CTNNB1 exon 3 mutations, such as observed here, lead to increased beta-catenin protein stability and activation of the WNT pathway, and are considered to be activating<sup>133-151</sup>.

ORDERED TEST # ORD-1328300-01

**GENOMIC FINDINGS**
**GENE**

# KEAP1

**ALTERATION**

G477S - subclonal

**TRANSCRIPT ID**

NM\_012289

**CODING SEQUENCE EFFECT**

1429G&gt;A

**VARIANT ALLELE FREQUENCY (% VAF)**

0.84%

**POTENTIAL TREATMENT STRATEGIES**
**— Targeted Therapies —**

A study of patients with localized non-small cell lung cancer (NSCLC) identified pathogenic KEAP1 and NFE2L2 mutations as predictors of local recurrence following radiotherapy but not surgery; limited preclinical data also showed that treatment with a glutaminase inhibitor sensitized KEAP1-mutated NSCLC cells to radiation<sup>152</sup>. In other preclinical studies, treatment with AKT inhibitors sensitized lung cancer cells harboring KEAP1 or NFE2L2 mutations to both chemotherapy and radiation therapy<sup>153-154</sup>. Mixed clinical data have been reported for the association between KEAP1 mutations and the response to immunotherapy. A pan-cancer study of immunotherapy showed that patients with KEAP1 mutations had shorter OS (10 vs. 20 months) than those without<sup>155</sup>. However, another study across solid tumors showed that KEAP1 mutations were associated with higher tumor mutational burden

(TMB) and PD-L1 expression, as well as improved survival outcomes with immunotherapy compared with other treatments (20.0 vs. 11.5 months)<sup>156</sup>. For patients with non-small cell lung cancer (NSCLC), a study of PD-L1 inhibitors showed that patients with concurrent mutations of STK11 and KEAP1 (n=39) experienced significantly shorter PFS (1.6 vs. 2.5 months, HR=1.5) and OS (4 vs. 11 months, HR=1.9) compared with patients with STK11- and KEAP1-wildtype tumors (n=210) despite significantly higher TMB in the group harboring STK11 and KEAP1 mutations (median 9.4 vs. 6.1 Muts/Mb)<sup>157</sup>. Retrospective analyses of patients with NSCLC who received immunotherapy reported reduced OS (p=0.040) for patients harboring KEAP1- or NFE2L2-mutated tumors<sup>158</sup> or STK11- or KEAP1-mutated tumors (p < 0.001)<sup>159</sup> compared with those without. Studies of immune checkpoint inhibitors for patients with lung adenocarcinoma showed that coexisting mutations between KEAP1, PBRM1, SMARCA4, STK11, and KRAS were associated with worse OS<sup>160</sup>. An exploratory analysis of a subset of patients with PD-L1-positive NSCLC treated in the first-line setting with pembrolizumab showed similar ORR, PFS, and OS when comparing patients with STK11 or KEAP1 mutations and those without<sup>161</sup>. In addition, preclinical data suggest that KEAP1 inactivation increases tumor demand for glutamine and increases tumor sensitivity to glutaminase inhibitors like telaglenastat<sup>162-164</sup>. Limited clinical data suggest that KEAP1 mutations may predict improved clinical benefit from combinations of glutaminase inhibitors and anti-PD-1 inhibitors<sup>165</sup>; a Phase 1/2 study of the

glutaminase inhibitor telaglenastat (CB-839) plus nivolumab to treat advanced NSCLC reported better clinical benefit rates and median PFS for patients with KEAP1 mutations (75% [3/4] vs. 15% [2/13], 6.4 vs. 3.7 months), KRAS mutations (38% [3/8] vs. 20% [2/10], 4.5 vs. 3.7 months), or KEAP1 and KRAS concurrent mutations (100% [2/2] vs. 13% [1/8], 7.2 vs. 3.7 months) compared with patients without these mutations<sup>165</sup>. The KEAP1 mutation has also been identified as a potential biomarker for sensitivity to combined AKT and TXNRD1 inhibition in lung cancer<sup>166</sup>.

**FREQUENCY & PROGNOSIS**

Somatic mutation of KEAP1 occurs in a range of solid tumors, including gastric, hepatocellular, colorectal, and lung cancers<sup>167</sup>. KEAP1 mutations are rare in hematological malignancies, occurring in fewer than 1% of samples analyzed (COSMIC, 2022)<sup>57</sup>. In a retrospective analysis of the pan-solid MSKCC dataset, KEAP1 mutation correlated with reduced OS (13.28 vs. 26.53 months)<sup>156</sup>.

**FINDING SUMMARY**

KEAP1 encodes a substrate adaptor protein that regulates the cellular response to oxidative stress by providing substrate specificity for a CUL3-dependent ubiquitin ligase<sup>168</sup>. KEAP1 exerts anti-tumor effects through negative regulation of NRF2, a transcription factor encoded by NFE2L2<sup>169-171</sup>; KEAP1 inactivation promotes cancer progression through NRF2-mediated chemoresistance and cell growth<sup>170-171</sup>.

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**GENOMIC FINDINGS**
**GENE**
**TERT**
**ALTERATION**

promoter -124C&gt;T

**TRANSCRIPT ID**

NM\_198253

**CODING SEQUENCE EFFECT**

-124C&gt;T

**VARIANT ALLELE FREQUENCY (% VAF)**

34.2%

**POTENTIAL TREATMENT STRATEGIES**
**— Targeted Therapies —**

Therapeutic options for targeting tumors with TERT mutations are limited, although a variety of approaches have been investigated, including immunotherapies using TERT as a tumor-associated antigen and antisense oligonucleotide- or peptide-based therapies. TERT peptide vaccines

showed limited anticancer efficacy in clinical trials<sup>172</sup>; however, in one preclinical study, the combination of a TERT peptide vaccine and anti-CTLA-4 therapy suppressed tumor growth<sup>173</sup>. A Phase 2 study of the TERT inhibitor imetelstat for patients with advanced non-small cell lung cancer reported no improvement in PFS or OS<sup>174</sup>.

**FREQUENCY & PROGNOSIS**

TERT is one of the most commonly mutated genes in hepatocellular carcinoma (HCC), with mutations, predominately in the promoter region, reported in 25-59% of cases<sup>175-177</sup>. TERT promoter mutation has been reported to be the earliest genetic event in HCC development, identified in pre-neoplastic cirrhotic lesions<sup>175,177</sup>. The frequency of these mutations increases from low-grade pre-malignant tumors such as adenomas and dysplastic nodules to high-grade pre-malignant tumors to carcinoma, suggesting a role for TERT promoter mutation in HCC progression<sup>175,177-178</sup>. A large-scale study reported

that TERT promoter alterations are associated with the presence of hepatitis C infection but the absence of hepatitis B infection, and are not associated with prognostic factors such as tumor size, grade, stage, or recurrence in patients with HCC<sup>176</sup>.

**FINDING SUMMARY**

Telomerase reverse transcriptase (TERT, or hTERT) is a catalytic subunit of the telomerase complex, which is required to maintain appropriate chromosomal length<sup>179</sup>. Activation of TERT is a hallmark of cancer, being detected in up to 80-90% of malignancies and absent in quiescent cells<sup>180-182</sup>. Mutations within the promoter region of TERT that confer enhanced TERT promoter activity have been reported in two hotspots, located at -124 bp and -146 bp upstream of the transcriptional start site (also termed C228T and C250T, respectively)<sup>183-185</sup>, as well as tandem mutations at positions -124/-125 bp and -138/-139 bp<sup>183</sup>.



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**CLINICAL TRIALS**

**NOTE** Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://clinicaltrials.gov). Or visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

**GENE**  
**ARID1A**
**RATIONALE**  
 ARID1A loss or inactivation may predict

sensitivity to ATR inhibitors.

**ALTERATION**  
 G931\*

**NCT04768296**
**PHASE 2**

Berzosertib + Topotecan in Relapsed Platinum-Resistant Small-Cell Lung Cancer (DDriver SCLC 250)

**TARGETS**  
 TOP1, ATR

**LOCATIONS:** Hangzhou (China), Nanjing (China), Wuhan (China), Xi'an (China), Osaka (Japan), Beijing (China), Hirakata-shi (Japan), Takatsuki-shi (Japan), Chengdu (China), Chuo-ku (Japan)

**NCT02264678**
**PHASE 1/2**

Ascending Doses of AZD6738 in Combination With Chemotherapy and/or Novel Anti Cancer Agents

**TARGETS**  
 ATR, PARP, PD-L1

**LOCATIONS:** Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Goyang-si (Korea, Republic of), Cambridge (United Kingdom), Withington (United Kingdom), Manchester (United Kingdom), Coventry (United Kingdom), Sutton (United Kingdom), Oxford (United Kingdom), Villejuif (France)

**NCT04657068**
**PHASE 1/2**

A Study of ART0380 for the Treatment of Advanced or Metastatic Solid Tumors

**TARGETS**  
 ATR

**LOCATIONS:** London (United Kingdom), Colorado, Oklahoma, Tennessee, Florida

**NCT04802174**
**PHASE 1/2**

Lurbinectedin With Berzosertib, an ATR Kinase Inhibitor in Small Cell Cancers and High-Grade Neuroendocrine Cancers

**TARGETS**  
 ATR

**LOCATIONS:** Maryland

**NCT02595931**
**PHASE 1**

ATR Kinase Inhibitor VX-970 and Irinotecan Hydrochloride in Treating Patients With Solid Tumors That Are Metastatic or Cannot Be Removed by Surgery

**TARGETS**  
 ATR

**LOCATIONS:** California, Missouri, Pennsylvania, Massachusetts, Connecticut, Tennessee

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**CLINICAL TRIALS**
**NCT04514497**
**PHASE 1**

Testing the Addition of an Anti-cancer Drug, BAY 1895344, to Usual Chemotherapy for Advanced Stage Solid Tumors, With a Specific Focus on Patients With Small Cell Lung Cancer, Poorly Differentiated Neuroendocrine Cancer, and Pancreatic Cancer

**TARGETS**  
 ATR, TOP1

**LOCATIONS:** Arizona, Minnesota, Oklahoma, Connecticut, Tennessee, Florida

**NCT04266912**
**PHASE 1/2**

Avelumab and M6620 for the Treatment of DDR Deficient Metastatic or Unresectable Solid Tumors

**TARGETS**  
 ATR, PD-L1

**LOCATIONS:** Texas

**NCT03669601**
**PHASE 1**

AZD6738 &amp; Gemcitabine as Combination Therapy

**TARGETS**  
 ATR

**LOCATIONS:** Cambridge (United Kingdom)

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**CLINICAL TRIALS**
**GENE**
**CTNNB1**
**RATIONALE**

 Based on clinical and preclinical evidence, tumors sensitive to mTOR inhibitors.  
 with activating CTNNB1 alterations may be

**ALTERATION**
**D32Y**
**NCT03591965**
**PHASE 2**

Dual TORC1/TORC2 Inhibitor ATG-008 (CC-223) in HBV Positive Advanced Hepatocellular Carcinoma (HCC) Subjects

**TARGETS**  
 mTORC1, mTORC2

**LOCATIONS:** Taipei (Taiwan), Taoyuan (Taiwan), Taichung (Taiwan), Fuzhou (China), Hangzhou (China), Shanghai (China), Nanjing (China), Hefei (China), Guangzhou (China), Changsha (China)

**NCT04337463**
**PHASE NULL**

ATG-008 Combined With Toripalimab in Advanced Solid Tumors

**TARGETS**  
 mTORC1, mTORC2, PD-1

**LOCATIONS:** Chongqing (China), Chengdu (China)

**NCT04803318**
**PHASE 2**

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

**TARGETS**  
 mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK

**LOCATIONS:** Guangzhou (China)

**NCT03065062**
**PHASE 1**

Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head &amp; Neck and Other Solid Tumors

**TARGETS**  
 PI3K-alpha, PI3K-gamma, mTORC1, mTORC2, CDK4, CDK6

**LOCATIONS:** Massachusetts

**NCT01582191**
**PHASE 1**

A Phase 1 Trial of Vandetanib (a Multi-kinase Inhibitor of EGFR, VEGFR and RET Inhibitor) in Combination With Everolimus (an mTOR Inhibitor) in Advanced Cancer

**TARGETS**  
 mTOR, EGFR, SRC, RET, VEGFRs

**LOCATIONS:** Texas

**NCT02321501**
**PHASE 1**

Phase I/Ib Dose Escalation &amp; Biomarker Study of Ceritinib (LDK378) + Everolimus for Locally Advanced or Metastatic Solid Tumors With an Expansion in Non-Small Cell Lung Cancer (NSCLC) Characterized by Abnormalities in Anaplastic Lymphoma Kinase (ALK) Expression

**TARGETS**  
 ROS1, ALK, mTOR

**LOCATIONS:** Texas

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 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

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**CLINICAL TRIALS**
**NCT03203525**
**PHASE 1**

 Combination Chemotherapy and Bevacizumab With the NovoTTF-100L(P) System in Treating  
 Participants With Advanced, Recurrent, or Refractory Hepatic Metastatic Cancer

**TARGETS**  
 VEGFA, mTOR

**LOCATIONS:** Texas

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**APPENDIX**
**Variants of Unknown Significance**

**NOTE** One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

**ATR**  
A344T

**MAP2K1 (MEK1)**  
T55fs\*12

**SNCAIP**  
rearrangement

**STK11**  
F354L

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**APPENDIX**
**Genes Assayed in FoundationOne®CDx**

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

**DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS**

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTK	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL	CCND1
CCND2	CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2	DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FAM46C	FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3
FGFR4	FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3
GATA4	GATA6	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3F3A
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDMSA	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKKN1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKB1A	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD3 (WHSC1L1)	NTSC2	NTRK1	NTRK2	NTRK3	P2RY8	PALB2
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA
PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET
RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2
SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOC3
SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU
SYK	TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53
TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1	WT1	XPO1
XRCC2	ZNF217	ZNF703						

**DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS**

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV1
ETV4	ETVS	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT
KMT2A (MLL)	MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA
RAF1	RARA	RET	ROS1	RSP02	SDC4	SLC34A2	TERC*	TERT**
TPRSS2								

\*TERC is an NCRNA

\*\*Promoter region of TERT is interrogated

**ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS**

Loss of Heterozygosity (LOH) score

Microsatellite (MS) status

Tumor Mutational Burden (TMB)

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## APPENDIX

## About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.



## ABOUT FOUNDATIONONE CDx

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:  
[www.rochefoundationmedicine.com/f1cdxtech](http://www.rochefoundationmedicine.com/f1cdxtech).

## INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

## TEST PRINCIPLES

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

## THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

### Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

### Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

## Ranking of Therapies and Clinical Trials

### Ranking of Therapies in Summary Table

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

### Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

## NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) ([www.nccn.org](http://www.nccn.org)). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to [NCCN.org](http://NCCN.org). NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

## Limitations

1. In the fractional-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-

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About FoundationOne®CDx

- Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.
- TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation [https://www.accessdata.fda.gov/cdrh\\_docs/pdf17/P170019B.pdf](https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf). The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
  - The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
  - Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
  - Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine

whether the patient is a candidate for biopsy.

- Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of HER2 overexpression as 20% in breast cancer. Based on the F1CDx HER2 CDx concordance study, approximately 10% of HER2 amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

## REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

## VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

## Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

\*Interquartile Range = 1<sup>st</sup> Quartile to 3<sup>rd</sup> Quartile

## VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

## VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear

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About FoundationOne®CDx

cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

#### LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

#### NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

#### NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

#### TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

#### SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

MR Suite Version 6.1.0

**The median exon coverage for this sample is 1,044x**

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 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531  
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531  
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

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**APPENDIX**
**References**

1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) pmid: 25392179
2. Kroemer G, et al. Oncoimmunology (2015) pmid: 26140250
3. Lal N, et al. Oncoimmunology (2015) pmid: 25949894
4. Le DT, et al. N. Engl. J. Med. (2015) pmid: 26028255
5. Ayers et al., 2016; ASCO-SITC Abstract P60
6. Goumard C, et al. Cancer Genomics Proteomics ( ) pmid: 28871000
7. Gross-Goupil M, et al. Int. J. Cancer (2003) pmid: 12640682
8. Zhang SH, et al. World J. Gastroenterol. (2005) pmid: 15918185
9. Chiappini F, et al. Carcinogenesis (2004) pmid: 14656944
10. Salvucci M, et al. Oncogene (1999) pmid: 9926933
11. Wang L, et al. Int. J. Oncol. (2001) pmid: 11494037
12. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) pmid: 26337942
13. You JF, et al. Br. J. Cancer (2010) pmid: 21081928
14. Bairwa NK, et al. Methods Mol. Biol. (2014) pmid: 24623249
15. Boland CR, et al. Cancer Res. (1998) pmid: 9823339
16. Pawlik TM, et al. Dis. Markers (2004) pmid: 15528785
17. Boland CR, et al. Gastroenterology (2010) pmid: 20420947
18. Samstein RM, et al. Nat. Genet. (2019) pmid: 30643254
19. Goodman AM, et al. Mol. Cancer Ther. (2017) pmid: 28835386
20. Goodman AM, et al. Cancer Immunol Res (2019) pmid: 31405947
21. Cristescu R, et al. Science (2018) pmid: 30309915
22. Ready N, et al. J. Clin. Oncol. (2019) pmid: 30785829
23. Hellmann MD, et al. N. Engl. J. Med. (2018) pmid: 29658845
24. Hellmann MD, et al. Cancer Cell (2018) pmid: 29657128
25. Hellmann MD, et al. Cancer Cell (2018) pmid: 29731394
26. Rozeman EA, et al. Nat Med (2021) pmid: 33558721
27. Sharma P, et al. Cancer Cell (2020) pmid: 32916128
28. Marabelle A, et al. Lancet Oncol. (2020) pmid: 32919526
29. Legrand et al., 2018; ASCO Abstract 12000
30. Chalmers ZR, et al. Genome Med (2017) pmid: 28420421
31. Shrestha R, et al. Front Oncol (2018) pmid: 30057891
32. Cai H, et al. J Surg Oncol (2020) pmid: 31995247
33. Pfeifer GP, et al. Mutat. Res. (2005) pmid: 15748635
34. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) pmid: 23875803
35. Pfeifer GP, et al. Oncogene (2002) pmid: 12379884
36. Rizvi NA, et al. Science (2015) pmid: 25765070
37. Johnson BE, et al. Science (2014) pmid: 24336570
38. Choi S, et al. Neuro-oncology (2018) pmid: 29452419
39. Cancer Genome Atlas Research Network, et al. Nature (2013) pmid: 23636398
40. Briggs S, et al. J. Pathol. (2013) pmid: 23447401
41. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) pmid: 24583393
42. Nature (2012) pmid: 22810696
43. Roberts SA, et al. Nat. Rev. Cancer (2014) pmid: 25568919
44. Williamson CT, et al. Nat Commun (2016) pmid: 27958275
45. Aggarwal et al., 2021; ESMO Abstract 5120
46. Thomas A, et al. J. Clin. Oncol. (2018) pmid: 29252124
47. Yap TA, et al. J Clin Oncol (2020) pmid: 32568634
48. Bitler BG, et al. Nat. Med. (2015) pmid: 25686104
49. Kim KH, et al. Nat. Med. (2015) pmid: 26552009
50. Wiegand KC, et al. BMC Cancer (2014) pmid: 24559118
51. Huang HN, et al. Mod. Pathol. (2014) pmid: 24336158
52. Samartzis EP, et al. Oncotarget (2014) pmid: 24979463
53. Okamura R, et al. J Immunother Cancer (2020) pmid: 32111729
54. Yokoyama Y, et al. J Gynecol Oncol (2014) pmid: 24459582
55. Katagiri A, et al. Mod. Pathol. (2012) pmid: 22101352
56. Xie C, et al. Tumour Biol. (2014) pmid: 24833095
57. Tate JG, et al. Nucleic Acids Res. (2019) pmid: 30371878
58. Cerami E, et al. Cancer Discov (2012) pmid: 22588877
59. Gao J, et al. Sci Signal (2013) pmid: 23550210
60. Wu RC, et al. Cancer Biol. Ther. (2014) pmid: 24618703
61. Jones S, et al. Hum. Mutat. (2012) pmid: 22009941
62. Dulak AM, et al. Nat. Genet. (2013) pmid: 23525077
63. Streppel MM, et al. Oncogene (2014) pmid: 23318448
64. Jiao Y, et al. J. Pathol. (2014) pmid: 24293293
65. Ross JS, et al. Oncologist (2014) pmid: 24563076
66. Huang HN, et al. Histopathology (2015) pmid: 25195947
67. Hussein YR, et al. Mod. Pathol. (2015) pmid: 25394778
68. Bosse T, et al. Mod. Pathol. (2013) pmid: 23702729
69. Allo G, et al. Mod. Pathol. (2014) pmid: 23887303
70. Chou A, et al. Hum. Pathol. (2014) pmid: 24925223
71. Ye J, et al. Hum. Pathol. (2014) pmid: 25311944
72. Wei XL, et al. World J. Gastroenterol. (2014) pmid: 25561809
73. Chen K, et al. Proc. Natl. Acad. Sci. U.S.A. (2015) pmid: 25583476
74. Wang K, et al. Nat. Genet. (2011) pmid: 22037554
75. Abe H, et al. Virchows Arch. (2012) pmid: 22915242
76. Wang DD, et al. PLoS ONE (2012) pmid: 22808142
77. Wiegand KC, et al. Hum. Pathol. (2014) pmid: 24767857
78. Katagiri A, et al. Int. J. Gynecol. Cancer (2012) pmid: 22274316
79. Cho H, et al. Hum. Pathol. (2013) pmid: 23427874
80. Gui Y, et al. Nat. Genet. (2011) pmid: 21822268
81. Balbás-Martínez C, et al. PLoS ONE (2013) pmid: 23650517
82. Faraj SF, et al. Hum. Pathol. (2014) pmid: 25175170
83. Rahman M, et al. Hum. Pathol. (2013) pmid: 22939958
84. Maeda D, et al. Int J Mol Sci (2010) pmid: 21614196
85. Lowery WJ, et al. Int. J. Gynecol. Cancer (2012) pmid: 22193641
86. Fadare O, et al. Mod. Pathol. (2013) pmid: 23524907
87. Mao TL, et al. Am. J. Surg. Pathol. (2013) pmid: 24076775
88. Zhang X, et al. Cancer Epidemiol (2012) pmid: 21889920
89. Mamo A, et al. Oncogene (2012) pmid: 21892209
90. Zhao J, et al. Tumour Biol. (2014) pmid: 24430365
91. Lichner Z, et al. Am. J. Pathol. (2013) pmid: 23416164
92. Feng F, et al. Int J Clin Oncol (2021) pmid: 33387086
93. Conci S, et al. Updates Surg (2020) pmid: 32020551
94. Simbolo M, et al. Sci Rep (2018) pmid: 29740198
95. Ruzzenente A, et al. Ann. Surg. Oncol. (2016) pmid: 26717940
96. Guan B, et al. Cancer Res. (2011) pmid: 21900401
97. Wiegand KC, et al. N. Engl. J. Med. (2010) pmid: 20942669
98. Jones S, et al. Science (2010) pmid: 20826764
99. Yan HB, et al. Carcinogenesis (2014) pmid: 24293408
100. Huang J, et al. Nat. Genet. (2012) pmid: 22922871
101. Chan-On W, et al. Nat. Genet. (2013) pmid: 24185513
102. Zang ZJ, et al. Nat. Genet. (2012) pmid: 22484628
103. Tanwar PS, et al. Biol. Reprod. (2009) pmid: 19403928
104. Tanwar PS, et al. PLoS ONE (2011) pmid: 21695255
105. Fujishita T, et al. Proc. Natl. Acad. Sci. U.S.A. (2008) pmid: 18768809
106. Bhoori S, et al. J. Hepatol. (2010) pmid: 20347502
107. Janku F, et al. Oncotarget (2014) pmid: 24931142
108. Slomovitz BM, et al. J. Clin. Oncol. (2015) pmid: 25624430
109. Niida A, et al. Oncogene (2004) pmid: 15378020
110. Chamorro MN, et al. EMBO J. (2005) pmid: 15592430
111. Kagey MH, et al. Br. J. Pharmacol. (2017) pmid: 28574171
112. Kagey et al., 2017; AACR Abstract 369
113. Kwon C, et al. Nat. Cell Biol. (2011) pmid: 21841793
114. Arcaroli JJ, et al. Br. J. Cancer (2013) pmid: 23868008
115. Shang H, et al. Cancer (2015) pmid: 26349011
116. Kode A, et al. Nature (2014) pmid: 24429522
117. Kummur et al., 2015; ASCO Abstract 10563
118. Messersmith WA, et al. Clin. Cancer Res. (2015) pmid: 25231399
119. Zhu J, et al. Carcinogenesis (2012) pmid: 22964660
120. Kogan Y, et al. Biochem. J. (2012) pmid: 22356261
121. Lachenmayer A, et al. Clin. Cancer Res. (2012) pmid: 22811581
122. Ruiz de Galarreta M, et al. Cancer Discov (2019) pmid: 31186238
123. Harding JJ, et al. Clin. Cancer Res. (2018) pmid: 30373752
124. von Felden J, et al. Oncogene (2021) pmid: 33097857
125. Chu HH, et al. Clin Mol Hepatol (2013) pmid: 23837144
126. Kan Z, et al. Genome Res. (2013) pmid: 23788652
127. Nault JC, et al. Int J Hepatol (2013) pmid: 23401783
128. Yuan RH, et al. PLoS ONE (2013) pmid: 23785431
129. Hsu HC, et al. Am. J. Pathol. (2000) pmid: 10980116
130. Nhieu JT, et al. Am. J. Pathol. (1999) pmid: 10487827
131. Luke JJ, et al. Clin Cancer Res (2019) pmid: 30635339
132. Biochem. Biophys. Res. Commun. (2000) pmid: 10679188
133. Anastas JN, et al. Nat. Rev. Cancer (2013) pmid: 23258168
134. Fukuchi T, et al. Cancer Res. (1998) pmid: 9721853
135. Cancer Sci. (2003) pmid: 12824913
136. Takahashi Y, et al. Virchows Arch. (2006) pmid: 16523258
137. Tanaka Y, et al. Cancer Res. (2001) pmid: 11731417
138. Abraham SC, et al. Am. J. Pathol. (2002) pmid: 11943721
139. Austinat M, et al. Mol. Cancer (2008) pmid: 18282277
140. Wu G, et al. Mol. Cell (2003) pmid: 12820959
141. Provost E, et al. Oncogene (2005) pmid: 15829978
142. Curr. Opin. Genet. Dev. (1999) pmid: 10072352
143. Segditsas S, et al. Oncogene (2006) pmid: 17143297
144. Barth AI, et al. J. Cell Biol. (1997) pmid: 9024698
145. Harada N, et al. EMBO J. (1999) pmid: 10545105
146. Hsu SC, et al. Mol. Cell. Biol. (1998) pmid: 9671490
147. Breuhahn K, et al. J. Pathol. (2008) pmid: 18491352
148. Soon PS, et al. Oncologist (2008) pmid: 18515740
149. Tacon LJ, et al. Oncologist (2011) pmid: 21212436
150. Simon DP, et al. Mol. Cell. Endocrinol. (2012) pmid: 22266195
151. Hirotsu Y, et al. Hepatol. Res. (2016) pmid: 26850916
152. Binkley MS, et al. Cancer Discov (2020) pmid: 33071215
153. Chowdhry S, et al. Oncogene (2013) pmid: 22964642
154. Abazeed ME, et al. Cancer Res. (2013) pmid: 23980093
155. Chen X, et al. Ann Transl Med (2020) pmid: 32175433
156. Xu X, et al. Oncologist (2020) pmid: 32272498
157. Arbour et al., 2018; IASLC WCLC Abstract MA19.09

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 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309  
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Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531  
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**References**

158. Zhang C, et al. J Thorac Oncol (2020) PMID: 32471565
159. Shang et al., 2020; WCLC Abstract P75.02
160. Marinelli D, et al. Ann Oncol (2020) PMID: 32866624
161. Cho et al., 2020; AACR Abstract CT084
162. Gwinn DM, et al. Cancer Cell (2018) PMID: 29316436
163. Sayin VI, et al. Elife (2017) PMID: 28967864
164. Romero R, et al. Nat. Med. (2017) PMID: 28967920
165. Skoulidis et al., 2021; ASCO Abstract TPS9627
166. Dai B, et al. Cancer Res. (2013) PMID: 23824739
167. Yoo NJ, et al. Histopathology (2012) PMID: 22348534
168. Lo SC, et al. J. Biol. Chem. (2006) PMID: 17046835
169. Wakabayashi N, et al. Nat. Genet. (2003) PMID: 14517554
170. Kansanen E, et al. Redox Biol (2013) PMID: 24024136
171. Hast BE, et al. Cancer Res. (2013) PMID: 23382044
172. Nat Rev Clin Oncol (2017) PMID: 27245281
173. Duperret EK, et al. Mol Ther (2018) PMID: 29249395
174. Chiappori AA, et al. Ann Oncol (2015) PMID: 25467017
175. Nault JC, et al. Nat Commun (2013) PMID: 23887712
176. Chen YL, et al. Int J Surg (2014) PMID: 24866078
177. Nault JC, et al. Hepatology (2014) PMID: 25123086
178. Pilati C, et al. Cancer Cell (2014) PMID: 24735922
179. Shay JW, et al. Semin. Cancer Biol. (2011) PMID: 22015685
180. Shay JW, et al. Eur. J. Cancer (1997) PMID: 9282118
181. Kim NW, et al. Science (1994) PMID: 7605428
182. Hanahan D, et al. Cell (2000) PMID: 10647931
183. Horn S, et al. Science (2013) PMID: 23348503
184. Huang FW, et al. Science (2013) PMID: 23348506
185. Vinagre J, et al. Nat Commun (2013) PMID: 23887589

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