Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

# ACTOnco® + Report

PATIENT						
Identifier: 林張敏珠	Patient ID: 44731933					
Date of Birth: Feb 27, 1937	Gender: Female					
Diagnosis: Lung adenocarcinoma						
ORDERING PHYSICIAN						
Name: 趙恒勝醫師	Tel: 886-228712121					
Facility: 臺北榮總						
Address: 臺北市北投區石牌路二段 201 號						
SPECIMEN						
Specimen ID: S11210979 Collection site	: Lung Type: FFPE tissue					
Date received: Mar 24, 2023 Lab ID: AA-23	-01763 D/ID: NA					

#### ABOUT ACTOnco®+

The test is a next-generation sequencing (NGS)-based assay developed for efficient and comprehensive genomic profiling of cancers. This test interrogates coding regions of 440 genes associated with cancer treatment, prognosis and diagnosis. Genetic mutations detected by this test include small-scale mutations like single nucleotide variants (SNVs), small insertions and deletions (InDels) (≤ 15 nucleotides) and large-scale genomic alterations like copy number alterations (CNAs). The test also includes an RNA test, detecting fusion transcripts of 13 genes.

# SUMMARY FOR ACTIONABLE VARIANTS VARIANTS/BIOMARKERS WITH EVIDENCE OF CLINICAL SIGNIFICANCE

Genomic	Probable Effects in F	Probable Sensitive in Other			
Alterations/Biomarkers	Sensitive Resistant		Cancer Types		
Not detected					

#### VARIANTS/BIOMARKERS WITH POTENTIAL CLINICAL SIGNIFICANCE

Genomic Alterations/Biomarkers	Possibly Sensitive	Possibly Resistant
MSH6 S200*	Olaparib	-

#### Note:

- The above summary tables present genomic variants and biomarkers based on the three-tiered approach proposed by US FDA for reporting tumor profiling NGS testing. "Variants/biomarkers with evidence of clinical significance" refers to mutations that are widely recognized as standard-of-care biomarkers (FDA level 2/AMP tier 1). "Variants/biomarkers with potential clinical significance" refers to mutations that are not included in the standard of care but are informational for clinicians, which are commonly biomarkers used as inclusion criterial for clinical trials (FDA level 3/AMP tier 2).
- The therapeutic agents and possible effects to a given drug are based on mapping the variants/biomarkers with ACT Genomics clinical knowledge database. The mapping results only provide information for reference, but not medical recommendation.
- Please refer to corresponding sections for more detailed information about genomic alteration and clinical relevance listed above.





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AG4-QP4001-02(07) page 1 of 18

Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

# ACTOnco® + Report

### **TESTING RESULTS**

### **VARIANT(S) WITH CLINICAL RELEVANCE**

### - Single Nucleotide and Small InDel Variants

Gene	Amino Acid Change	Allele Frequency	
MSH6	S200*	25.1%	
TP53	S314fs	35.8%	

### - Copy Number Alterations

Chromosome	Gene	Variation	Copy Number	
	Not	detected		

#### - Fusions

Fusion Gene & Exon	Transcript ID
	No fusion gene detected in this sample

### - Immune Checkpoint Inhibitor (ICI) Related Biomarkers

Biomarker	Results
Tumor Mutational Burden (TMB)	2.6 muts/Mb
Microsatellite Instability (MSI)	Microsatellite stable (MSS)

### Note:

- Variant(s) enlisted in the SNV table may currently exhibit no relevance to treatment response prediction. Please refer to INTERPRETATION for more biological information and/or potential clinical impacts of the variants.
- Loss of heterozygosity (LOH) information was used to infer tumor cellularity. Copy number alteration in the tumor was determined based on 34% tumor purity.
- TMB was calculated by using the sequenced regions of ACTOnco®+ to estimate the number of somatic nonsynonymous mutations per megabase of all protein-coding genes (whole exome). The threshold for high mutation load is set at ≥ 7.5 mutations per megabase. TMB, microsatellite status and gene copy number deletion cannot be determined if calculated tumor purity is < 30%.





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AG4-QP4001-02(07) page **2** of **18** 

Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

# ACTOnco® + Report

# THERAPEUTIC IMPLICATIONS

### **TARGETED THERAPIES**

Genomic Alterations	Therapies	Effect	
Level 3B			
<b>MSH6</b> S200*	Olaparib	sensitive	

Therapies associated with benefit or lack of benefit are based on biomarkers detected in this tumor and published evidence in professional guidelines or peer-reviewed journals.

Level	Description			
1	FDA-recognized biomarkers predictive of response or resistance to FDA approved drugs in this indication			
2	Standard care biomarkers (recommended by the NCCN guideline) predictive of response or resistance to FDA approved drugs in this indication			
ЗА	Biomarkers predictive of response or resistance to therapies approved by the FDA or NCCN guideline in a different cancer type			
3B	Biomarkers that serve as inclusion criteria for clinical trials (minimal supportive data required)			
4	Biomarkers that show plausible therapeutic significance based on small studies, few case reports, or preclinical studies			





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AG4-QP4001-02(07) page 3 of 18

Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

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### **IMMUNE CHECKPOINT INHIBITORS (ICIs)**

No genomic alterations detected to confer sensitivity or lack of benefit to immune checkpoint therapies.

#### - Other Biomarkers with Potential Clinical Effects for ICIs

Genomic Alterations	Potential Clinical Effects
	Not detected

Note: Tumor non-genomic factors, such as patient germline genetics, PDL1 expression, tumor microenvironment, epigenetic alterations or other factors not provided by this test may affect ICI response.

#### **CHEMOTHERAPIES**

No genomic alterations detected in this tumor predicted to confer sensitivity or lack of benefit to chemotherapies.

#### **HORMONAL THERAPIES**

No genomic alterations detected in this tumor predicted to confer sensitivity or lack of benefit to hormonal therapies.

#### **OTHERS**

No genomic alterations detected in this tumor predicted to confer sensitivity or lack of benefit to other therapies.

#### Note:

Therapeutic implications provided in the test are based solely on the panel of 440 genes sequenced. Therefore, alterations in genes not covered in this panel, epigenetic and post-transcriptional and post-translational factors may also determine a patient's response to therapies. In addition, several other patient-associated clinical factors, including but not limited to, prior lines of therapies received, dosage and combinations with other therapeutic agents, patient's cancer types, sub-types, and/or stages, may also determine the patient's clinical response to therapies.





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AG4-QP4001-02(07) page 4 of 18

Project ID: C23-M001-00902 Report No.: AA-23-01763 ONC

Date Reported: Apr 07, 2023



### VARIANT INTERPRETATION

#### MSH6 S200\*

#### **Biological Impact**

The mutS homolog 6 (MSH6) gene encodes the DNA repair mismatch protein MSH6, a tumor suppressor involved in the DNA mismatch repair (MMR)[1]. Germline mutations in MMR pathway, including MSH6, are associated with susceptibility to hereditary nonpolyposis colon cancer (HNPCC), endometrial cancer[2] or sporadic cancers with microsatellite instability (MSI)[3].

S200\* mutation results in a premature truncation of the MSH6 protein at amino acid 200 (UniProtKB). This mutation is predicted to lead to a loss of MSH6 function, despite not having characterized in the literature.

#### Therapeutic and prognostic relevance

No drugs targeting MSH6 mutations have been currently approved. A screening test for microsatellite instability (MSI) is commonly used to identify a MMR-deficient tumor in clinic[4][5]. Tumors with mismatch-repair defects were associated with a large number of somatic mutations and have been predictive of clinical benefit to certain immune checkpoint blockade therapies [6][7]. The PD-1 antibodies pembrolizumab and nivolumab have been approved by the U.S. FDA for instability-high (MSI-H) or mismatch repair deficient (dMMR) solid tumors and MSI-H or dMMR colorectal cancer respectively. Down-regulation of genes involved in the MMR pathway such as MLH1, MSH2, and MSH6 in high-grade serous epithelial ovarian cancer cell lines rendered cells sensitive to PARP inhibitors[8].

### **TP53 S314fs**

#### **Biological Impact**

TP53 encodes the p53 protein, a crucial tumor suppressor that orchestrates essential cellular processes including cell cycle arrest, senescence and apoptosis[9]. TP53 is a proto-typical haploinsufficient gene, such that loss of a single copy of TP53 can result in tumor formation<sup>[10]</sup>.

S314fs mutation results in a change in the amino acid sequence beginning at 314, likely to cause premature truncation of the functional p53 protein (UniProtKB). This mutation is predicted to lead to a loss of p53 protein function, despite not being characterized in the literature.

#### Therapeutic and prognostic relevance

Despite having a high mutation rate in cancers, there are currently no approved targeted therapies for TP53 mutations. A phase II trial demonstrated that Wee1 inhibitor (AZD1775) in combination with carboplatin was well tolerated and showed promising anti-tumor activity in TP53-mutated ovarian cancer refractory or resistant (< 3 months) to standard first-line therapy (NCT01164995)[11].

In a retrospective study (n=19), advanced sarcoma patients with TP53 loss-of-function mutations displayed improved progression-free survival (208 days versus 136 days) relative to patients with wild-type TP53 when treated with pazopanib<sup>[12]</sup>. Results from another Phase I trial of advanced solid tumors (n=78) demonstrated that TP53 hotspot mutations are associated with better clinical response to the combination of pazopanib and vorinostat[13].

Advanced solid tumor and colorectal cancer patients harboring a TP53 mutation have been shown to be more sensitive to bevacizumab when compared with patients harboring wild-type TP53<sup>[14][15][16]</sup>. In a pilot trial (n=21), TP53-negative breast cancer patients demonstrated increased survival following treatment with bevacizumab in combination with chemotherapy agents, Adriamycin (doxorubicin) and Taxotere (docetaxel)[17]. TP53 mutations were correlated with poor survival of advanced breast cancer patients receiving tamoxifen or primary chemotherapy[18][19].





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AG4-QP4001-02(07) page 5 of 18

Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

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In a retrospective study of non-small cell lung cancer (NSCLC), TP53 mutations were associated with high expression of VEGF-A, the primary target of bevacizumab, offering a mechanistic explanation for why patients exhibit improved outcomes after bevacizumab treatment when their tumors harbor mutant TP53 versus wild-type TP53<sup>[20]</sup>.





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AG4-QP4001-02(07) page 6 of 18

Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

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# US FDA-APPROVED DRUG(S)

Olaparib (LYNPARZA)

Olaparib is an oral, small molecule inhibitor of poly (ADP-ribose) polymerase-1, -2, and -3 (PARP-1, -2, -3). Olaparib is developed by KuDOS Pharmaceuticals and marketed by AstraZeneca under the trade name LYNPARZA.

# - FDA Approval Summary of Olaparib (LYNPARZA)

Ob	Her2-negative high-risk early breast cancer (Approved on 2022/03/11)
OlympiA	HER2-/gBRCA mutation
NCT02032823	Olaparib vs. Placebo [ invasive disease-free survival (IDFS)(M): ]
PPO6 #[21]	Prostate cancer (Approved on 2020/05/19)
PROfound <sup>[21]</sup>	HRR genes mutation
NCT02987543	Olaparib vs. Enzalutamide or abiraterone acetate [PFS(M): 5.8 vs. 3.5]
DAOL A 4[22]	Ovarian cancer (Approved on 2020/05/08)
PAOLA-1 <sup>[22]</sup>	HRD+
NCT02477644	Olaparib + bevacizumab vs. Placebo + bevacizumab [PFS(M): 37.2 vs. 17.7]
POLO <sup>[23]</sup>	Pancreatic adenocarcinoma (Approved on 2019/12/27)
NCT02184195	gBRCA mutation
NC102184195	Olaparib vs. Placebo [ORR(%): 23.0 vs. 12.0, PFS(M): 7.4 vs. 3.8]
201 0 4[24]	Ovarian cancer, Fallopian tube cancer, Peritoneal carcinoma (Approved on 2018/12/19)
<b>SOLO-1</b> <sup>[24]</sup> NCT01844986	gBRCA mutation or sBRCA mutation
NC101844986	Olaparib vs. Placebo [PFS(M): NR vs. 13.8]
OI :AD[25]	Breast cancer (Approved on 2018/02/06)
OlympiAD <sup>[25]</sup>	HER2-/gBRCA mutation
NCT02000622	Olaparib vs. Chemotherapy [PFS(M): 7 vs. 4.2]
001 0 0/FN00T 0: 04 <sup>[26]</sup>	Ovarian cancer, Fallopian tube cancer, Peritoneal carcinoma (Approved on 2017/08/17)
<b>SOLO-2/ENGOT-Ov21</b> <sup>[26]</sup> NCT01874353	gBRCA mutation
NC101874353	Olaparib vs. Placebo [PFS(M): 19.1 vs. 5.5]
Of a select O[27]	Ovarian cancer, Fallopian tube cancer, Peritoneal carcinoma (Approved on 2017/08/17)
Study19 <sup>[27]</sup>	
NCT00753545	Olaparib vs. Placebo [PFS(M): 8.4 vs. 4.8]

D=day; W=week; M=month





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AG4-QP4001-02(07) page **7** of **18** 

Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

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# **ONGOING CLINICAL TRIALS**

Trials were searched by applying filters: study status, patient's diagnosis, intervention, location and/or biomarker(s). Please visit <a href="https://clinicaltrials.gov">https://clinicaltrials.gov</a> to search and view for a complete list of open available and updated matched trials.

No trial has been found.





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AG4-QP4001-02(07) page 8 of 18

Project ID: C23-M001-00902 Report No.: AA-23-01763 ONC Date Reported: Apr 07, 2023

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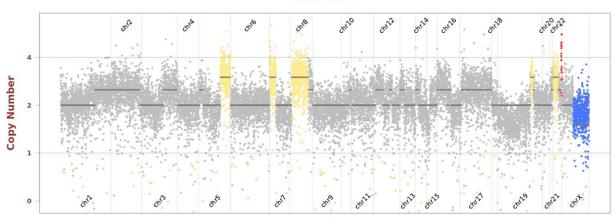
# SUPPLEMENTARY INFORMATION OF TESTING RESULTS DETAILED INFORMATION OF VARIANTS WITH CLINICAL RELEVANCE

### - Single Nucleotide and Small InDel Variants

Gene	Amino Acid Change	Exon	cDNA Change	Accession Number	COSMIC ID	Allele Frequency	Coverage
MSH6	S200*	3	c.599C>G	NM_000179	-	25.1%	1574
TP53	S314fs	9	c.940_952del	NM_000546	-	35.8%	1540

### - Copy Number Alterations

Observed copy number (CN) for each evaluated position is shown on the y-axis. Regions referred to as amplification or deletion are shown in color. Regions without significant changes are represented in gray.









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AG4-QP4001-02(07) page 9 of 18

Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

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### **OTHER DETECTED VARIANTS**

Gene Amino Acid Exc		Exon	cDNA Change	Accession Number	COSMIC ID	Allele Frequency	Coverage	
ARAF	L161M	6	c.481T>A	NM_001654	COSM6913205	24.7%	2201	
ATM	N1650S	33	c.4949A>G	NM_000051	-	37.7%	1669	
CALR	D335N	8	c.1003G>A	NM_004343	COSM3528934	36.6%	2168	
CBL	V431A	9	c.1292T>C	NM_005188	-	5.8%	3153	
CREBBP	S78T	2	c.233G>C	NM_004380	-	55.0%	1230	
CSF1R	Splice region	-	c.2132+5C>A	NM_005211	-	39.9%	1155	
EP300	R568Q	8	c.1703G>A	NM_001429	-	49.1%	3995	
FLT4	H1224R	27	c.3671A>G	NM_182925	-	40.4%	1829	
IDH1	199M	4	c.297A>G	NM_005896	COSM53043	68.0%	2045	
IL7R	A254T	6	c.760G>A	NM_002185	-	59.9%	1835	
KMT2C	C385F	8	c.1154G>T	NM_170606	COSM9180259	6.1%	3999	
MLH1	R217C	8	c.649C>T	NM_000249	COSM186809	54.2%	3910	
MTOR	L1184V	23	c.3550C>G	NM_004958	-	55.5%	1008	
MUC16	R12563C	24	c.37687C>T	NM_024690	COSM4933144	8.8%	1962	
MUC16	K10060N	3	c.30180G>C	NM_024690	-	29.1%	1536	
NOTCH2	Splice region	-	c.2480-4C>G	NM_024408	-	41.3%	1471	
PDGFRA	N328D	7	c.982A>G	NM_006206	-	38.5%	3834	
PTPRD	F74V	11	c.220T>G	NM_002839	-	5.1%	2448	
SYNE1	N4590S	78	c.13769A>G	NM_182961	COSM2152615	63.3%	1221	
USH2A	E4264K	63	c.12790G>A	NM 206933	COSM213160	49.0%	2332	

#### Note:

- This table enlists variants detected by the panel other than those with clinical relevance (reported in Testing Result section).

The clinical impact of a genetic variant is determined according to ACT Genomics in-house clinical knowledge database. A negative result does not necessarily indicate absence of biological effect on the tumor. Some variants listed here may possibly have preclinical data or may show potential clinical relevance in the future.





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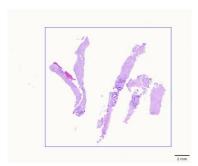
AG4-QP4001-02(07) page 10 of 18

Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

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# TEST DETAILS SPECIMEN RECEIVED AND PATHOLOGY REVIEW





- Collection date: Mar 14, 2023

Facility retrieved: 臺北榮總

H&E-stained section No.: S11210979

Collection site: Lung

- Examined by: Dr. Yeh-Han Wang

- 1. The percentage of viable tumor cells in total cells in the whole slide (%): 30%
- 2. The percentage of viable tumor cells in total cells in the encircled areas in the whole slide (%): 30%
- 3. The percentage of necrotic cells (including necrotic tumor cells) in total cells in the whole slide (%): 0%
- The percentage of necrotic cells (including necrotic tumor cells) in total cells in the encircled areas in the whole slide (%): 0%
- 5. Additional comment: NA
- Manual macrodissection: Not performed
- The outline highlights the area of malignant neoplasm annotated by a pathologist.

### **RUN QC**

Panel: ACTOnco®+

#### **DNA** test

Mean Depth: 1628x

Target Base Coverage at 100x: 96%

#### **RNA** test

Average unique RNA Start Sites per control GSP2: 139

### **LIMITATIONS**

- 1. This test does not provide information of variant causality and does not detect variants in non-coding regions that could affect gene expression. This report does not report polymorphisms and we do not classify whether a mutation is germline or somatic. Variants identified by this assay were not subject to validation by Sanger or other technologies.
- 2. The possibility cannot be excluded that certain pathogenic variants detected by other sequencing tools may not be reported in





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AG4-QP4001-02(07) page 11 of 18

Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

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the test because of technical limitation of bioinformatics algorithm or the NGS sequencing platform, e.g. low coverage.

3. This test has been designed to detect fusions in 13 genes sequenced. Therefore, fusion in genes not covered by this test would not be reported. For novel fusions detected in this test, Sanger sequencing confirmation is recommended if residue specimen is available.

### **NEXT-GENERATION SEQUENCING (NGS) METHODS**

#### DNA test

Extracted genomic DNA was amplified using primers targeting coding exons of analyzed genes and subjected to library construction. Barcoded libraries were subsequently conjugated with sequencing beads by emulsion PCR and enriched using Ion Chef system. Sequencing was performed according to Ion Proton or Ion S5 sequencer protocol (Thermo Fisher Scientific).

Raw reads generated by the sequencer were mapped to the hg19 reference genome using the Ion Torrent Suite. Coverage depth was calculated using Torrent Coverage Analysis plug-in. Single nucleotide variants (SNVs) and short insertions/deletions (InDels) were identified using the Torrent Variant Caller plug-in. VEP (Variant Effect Predictor) was used to annotate every variant using databases from Clinvar, COSMIC and Genome Aggregation database. Variants with coverage  $\geq$  20, allele frequency  $\geq$  5% and actionable variants with allele frequency  $\geq$  2% were retained. This test provides uniform coverage of the targeted regions, enabling target base coverage at  $100x \geq 85\%$  with a mean coverage  $\geq$  500x.

Variants reported in Genome Aggregation database with > 1% minor allele frequency (MAF) were considered as polymorphisms. ACT Genomics in-house database was used to determine technical errors. Clinically actionable and biologically significant variants were determined based on the published medical literature.

The copy number alterations (CNAs) were predicted as described below:

Amplicons with read counts in the lowest 5th percentile of all detectable amplicons and amplicons with a coefficient of variation ≥ 0.3 were removed. The remaining amplicons were normalized to correct the pool design bias. ONCOCNV (an established method for calculating copy number aberrations in amplicon sequencing data by Boeva et al., 2014) was applied for the normalization of total amplicon number, amplicon GC content, amplicon length, and technology-related biases, followed by segmenting the sample with a gene-aware model. The method was used as well for establishing the baseline of copy number variations.

Tumor mutational burden (TMB) was calculated by using the sequenced regions of ACTOnco®+ to estimate the number of somatic nonsynonymous mutations per megabase of all protein-coding genes (whole exome). The TMB calculation predicted somatic variants and applied a machine learning model with a cancer hotspot correction. TMB may be reported as "TMB-High", "TMB-Low" or "Cannot Be Determined". TMB-High corresponds to ≥ 7.5 mutations per megabase (Muts/Mb); TMB-Low corresponds to < 7.5 Muts/Mb. TMB is reported as "Cannot Be Determined" if the tumor purity of the sample is < 30%.

Classification of microsatellite instability (MSI) status is determined by a machine learning prediction algorithm. The change of a number of repeats of different lengths from a pooled microsatellite stable (MSS) baseline in > 400 genomic loci are used as the features for the algorithm. The final output of the results is either microsatellite Stable (MSS) or microsatellite instability high (MSI-H).

#### RNA test

Extracted RNA was reverse-transcribed and subjected to library construction. Sequencing was performed according to lon Proton or lon S5 sequencer protocol (Thermo Fisher Scientific). To ensure sequencing quality for fusion variant analysis, the average unique RNA Start Sites (SS) per control Gene Specific Primer 2 (GSP 2) should be ≥ 10.

The fusion analysis pipeline aligned sequenced reads to the human reference genome, identified regions that map to noncontiguous regions of the genome, applied filters to exclude probable false-positive events and, annotated previously characterized fusion events according to Quiver Gene Fusion Database, a curated database owned and maintained by ArcherDX. In general, samples with detectable fusions need to meet the following criteria: (1) Number of unique start sites (SS) for the GSP2  $\geq$  3; (2) Number of supporting reads spanning the fusion junction  $\geq$  5; (3) Percentage of supporting reads spanning the fusion junction  $\geq$  10%; (4) Fusions annotated





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AG4-QP4001-02(07) page 12 of 18

Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

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in Quiver Gene Fusion Database.

### **DATABASE USED**

- Reference genome: Human genome sequence hg19
- COSMIC v.92
- Genome Aggregation database r2.1.1
- ClinVar (version 20210404)
- ACT Genomics in-house database
- Quiver Gene Fusion Database version 5.1.18

**Variant Analysis:** 

醫檢師黃靖婷 博士 Ching-Ting Huang Ph.D. 檢字第 016511 號 CTHUANG

Sign Off

解剖病理專科醫師王業翰 Yeh-Han Wang M.D. 病解字第 000545 號







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AG4-QP4001-02(07) page **13** of **18** 

Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

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# GENE LIST SNV & CNV

ABCB1*	ABCC2*	ABCG2*	ABL1	ABL2	ADAMTS1	ADAMTS13	ADAMTS15	ADAMTS16	ADAMTS18	ADAMTS6	ADAMTSS
ADAMTSL1	ADGRA2	ADH1C*	AKT1	AKT2	AKT3	ALDH1A1*	ALK	AMER1	APC	AR	ARAF
ARID1A	ARID1B	ARID2	ASXL1	ATM	ATR	ATRX	AURKA	AURKB	AXIN1	AXIN2	AXL
B2M	BAP1	BARD1	BCL10	BCL2*	BCL2L1	BCL2L2*	BCL6	BCL9	BCOR	BIRC2	BIRC3
BLM	BMPR1A	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2*	ВТК	BUB1B	CALR
CANX	CARD11	CASP8	CBFB	CBL	CCNA1	CCNA	CCNB1	CCNB2	CCNB3	CCND1	CCND2
CCND3	CCNE1	CCNE2	CCNH	CD19	CD274	CD58	CD70*	CD79A	CD79B	CDC73	CDH1
CDK1	CDK12	CDK2	CDK4	CDK5	CDK6	CDK7	CDK8	CDK9	CDKN1A	CDKN1B	CDKN2A
CDKN2B	CDKN2C	CEBPA*	CHEK1	CHEK2	CIC	CREBBP	CRKL	CRLF2	CSF1R	CTCF	CTLA4
CTNNA1	CTNNB1	CUL3	CYLD	CYP1A1*	CYP2B6*	CYP2C19*	CYP2C8*	CYP2D6	CYP2E1*	CYP3A4*	CYP3A5*
DAXX	DCUN1D1	DDR2	DICER1	DNMT3A	DOT1L	DPYD	DTX1	E2F3	EGFR	EP300	EPCAM
EPHA2	ЕРНА3	EPHA5	ЕРНА7	EPHB1	ERBB2	ERBB3	ERBB4	ERCC1	ERCC2	ERCC3	ERCC4
ERCC5	ERG	ESR1	ESR2	ETV1	ETV4	EZH2	FAM46C	FANCA	FANCC	FANCD2	FANCE
FANCF	FANCG	FANCL	FAS	FAT1	FBXW7	FCGR2B	FGF1*	FGF10	FGF14	FGF19*	FGF23
FGF3	FGF4*	FGF6	FGFR1	FGFR2	FGFR3	FGFR4	FH	FLCN	FLT1	FLT3	FLT4
FOXL2*	FOXP1	FRG1	FUBP1	GATA1	GATA2	GATA3	GNA11	GNA13	GNAQ	GNAS	GREM1
GRIN2A	GSK3B	GSTP1*	GSTT1*	HGF	HIF1A	HIST1H1C*	HIST1H1E*	HNF1A	HR	HRAS*	HSP90AA
HSP90AB1	HSPA4	HSPA5	IDH1	IDH2	IFNL3*	IGF1	IGF1R	IGF2	IKBKB	IKBKE	IKZF1
IL6	IL7R	INPP4B	INSR	IRF4	IRS1	IRS2*	JAK1	JAK2	JAK3	JUN*	KAT6A
KDM5A	KDM5C	KDM6A	KDR	KEAP1	KIT	KMT2A	КМТ2С	KMT2D	KRAS	LCK	LIG1
LIG3	LMO1	LRP1B	LYN	MALT1	MAP2K1	MAP2K2	MAP2K4	MAP3K1	MAP3K7	MAPK1	МАРК3
MAX	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1	MET	MITF	MLH1	MPL	MRE11
MSH2	MSH6	MTHFR*	MTOR	MUC16	MUC4	мис6	МИТҮН	MYC	MYCL	MYCN	MYD88
NAT2*	NBN	NEFH	NF1	NF2	NFE2L2	NFKB1	NFKBIA	NKX2-1*	NOTCH1	NOTCH2	<i>NOTCH3</i>
NOTCH4	NPM1	NQ01*	NRAS	NSD1	NTRK1	NTRK2	NTRK3	PAK3	PALB2	PARP1	PAX5
PAX8	PBRM1	PDCD1	PDCD1LG2	PDGFRA	PDGFRB	PDIA3	PGF	PHOX2B*	PIK3C2B	PIK3C2G	РІКЗСЗ
PIK3CA	РІКЗСВ	PIK3CD	PIK3CG	PIK3R1	PIK3R2	PIK3R3	PIM1	PMS1	PMS2	POLB	POLD1
POLE	PPARG	PPP2R1A	PRDM1	PRKAR1A	PRKCA	PRKCB	PRKCG	PRKCI	PRKCQ	PRKDC	PRKN
PSMB8	PSMB9	PSME1	PSME2	PSME3	PTCH1	PTEN	PTGS2	PTPN11	PTPRD	PTPRT	RAC1
RAD50	RAD51	RAD51B	RAD51C	RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10	RECQL4
REL	RET	RHOA	RICTOR	RNF43	ROS1	RPPH1	RPTOR	RUNX1	RUNX1T1	RXRA	SDHA
SDHB	SDHC	SDHD	SERPINB3	SERPINB4	SETD2	SF3B1	SGK1	SH2D1A*	SLC19A1*	SLC22A2*	SLCO1B1
SLCO1B3*	SMAD2	SMAD3	SMAD4	SMARCA4	SMARCB1	SMO	SOCS1*	SOX2*	SOX9	SPEN	SPOP
SRC	STAG2	STAT3	STK11	SUFU	SYK	SYNE1	TAF1	TAP1	TAP2	TAPBP	TBX3
TEK	TERT	TET1	TET2	TGFBR2	TMSB4X*	TNF	TNFAIP3	TNFRSF14	TNFSF11	TOP1	TP53
TPMT*	TSC1	TSC2	TSHR	TYMS	U2AF1	UBE2A*	UBE2K	UBR5	UGT1A1*	USH2A	VDR*
VEGFA	VEGFB	VHL	WT1	XIAP	XPO1	XRCC2	ZNF217				

<sup>\*</sup>Analysis of copy number alterations NOT available.

### **FUSION**

												1
ALK	BRAF	EGFR	FGFR1	FGFR2	FGFR3	MET	NRG1	NTRK1	NTRK2	NTRK3	RET	ROS1





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AG4-QP4001-02(07) page **14** of **18** 

Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

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### **APPENDIX**

POSSIBLE THERAPEUTIC IMPLICATIONS FOR HETEROZYGOUS DELETION

Not Applicable.

SIGNALING PATHWAYS AND MOLECULAR-TARGETED AGENTS

Not Applicable.





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AG4-QP4001-02(07) page 15 of 18

Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

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### **DISCLAIMER**

#### 法律聲明

本檢驗報告僅提供專業醫療參考,結果需經專業醫師解釋及判讀。基因突變資訊非必具備藥物或治療有效性指標,反之亦然。本檢驗報 告提供之用藥指引不聲明或保證其臨床有效性,反之亦然。本基因檢測方法係由本公司研究開發,已經過有效性測試。

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#### 醫療決策需由醫師決定

任何治療與用藥需經由醫師在考慮病患所有健康狀況相關資訊包含健檢、其他檢測報告和病患意願後,依照該地區醫療照護標準由醫師獨立判斷。醫師不應僅依據單一報告結果(例如本檢測或本報告書內容)做決策。

# 基因突變與用藥資訊並非依照有效性排序

本報告中列出之生物標記變異與藥物資訊並非依照潛在治療有效性排序。

#### 證據等級

藥物潛在臨床效益(或缺乏潛在臨床效益)的實證證據是依據至少一篇臨床療效個案報告或臨床前試驗做為評估。本公司盡力提供適時及 準確之資料,但由於醫學科技之發展日新月異,本公司不就本報告提供的資料是否為準確、適宜或最新作保證。

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AG4-QP4001-02(07) page 16 of 18

Project ID: C23-M001-00902 Report No.: AA-23-01763\_ONC Date Reported: Apr 07, 2023

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AG4-QP4001-02(07) page 17 of 18

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AG4-QP4001-02(07) page 18 of 18