PATIENT Li, Meng-Ju TUMOR TYPE
Uterus endometrial
adenocarcinoma endometrioid
COUNTRY CODE

REPORT DATE 28 Sep 2021

ORD-1194331-01

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT

DISEASE Uterus endometrial adenocarcinoma endometrioid

NAME Li, Meng-Ju

DATE OF BIRTH 12 December 1974

SEX Female

MEDICAL RECORD # 47551224

PHYSICIAN

ORDERING PHYSICIAN Chen, Yi-Jen

MEDICAL FACILITY Taipei Veterans General Hospital

ADDITIONAL RECIPIENT None MEDICAL FACILITY ID 205872 PATHOLOGIST Not Provided

SPECIMEN

SPECIMEN SITE Uterus

SPECIMEN ID S110-67168 R (PF21018)

SPECIMEN TYPE Slide Deck

DATE OF COLLECTION 18 August 2021 **SPECIMEN RECEIVED** 21 September 2021

Biomarker Findings

Tumor Mutational Burden - 372 Muts/Mb Microsatellite status - MS-Stable

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

FBXW7 R441W **CTNNA1** R451* PIK3CA R88Q, T1025A **CUL3** L602* **ATM R250* ESR1** R256Q **BRCA2** E1879D FAM123B E387* NF1 R1362*, R2450* **KEL** E494* CTNNB1 K335T KLHL6 D104N FANCL splice site 904-2A>C MAP3K1 G914* FGFR2 N549D MLL2 R5214C **PIK3CB** R3210 **MSH6** E641* PIK3R1 E476* POLE V411L

PTEN R130Q, R173H SETD2 splice site 5278-1G>T

RAD51D R253Q SPEN A2727V

TSC1 Q739* STAG2 splice site 463-1G>T

APC S2307L, R2204* **TNFAIP3** R87*

ATRX R907* **TP53** F113V - subclonal, T211A[†]

 CARD11 R271W
 XPO1 R749Q

 CREBBP E50*
 ZNF217 D326N

CTCF R129*

16 Therapies with Clinical Benefit

O Therapies with Resistance

56 Clinical Trials

PATHOLOGIST COMMENTS

Douglas Lin, M.D. 28-Sep-2021

The genomic profile is compatible with a POLE-ultramutated endometrial carcinoma.

BIO	MAR	KER	FIN	IDIN	IGS

Tumor Mutational Burden - 372 Muts/Mb

10 Trials see p. 37

Microsatellite status - MS-Stable

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)		THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)	
Dostarlimab	2A	Avelumab	2A
Pembrolizumab	2A	Nivolumab	2A
		Atezolizumab	
		Cemiplimab	
		Durvalumab	
		Nivolumab + Ipilimumab	

No therapies or clinical trials. see Biomarker Findings section

 $[\]dagger$ See About the Test in appendix for details.

TUMOR TYPE Uterus endometrial adenocarcinoma endometrioid COUNTRY CODE

REPORT DATE 28 Sep 2021

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GENOMIC FINDINGS	THERAPIES WITH CLINICAL RELEVA (IN PATIENT'S TUMOR TYPE)	ANCE THERAPIES WITH CLINICATION (IN OTHER TUMOR	AL RELEVANCI R TYPE)
FBXW7 - R441W	none	Everolimus	2A
10 Trials see p. 47		Temsirolimus	2A
PIK3CA - R88Q, T1025A	none	Everolimus	2A
10 Trials see <i>p. 53</i>		Temsirolimus	2A
ATM - R250*	none	Niraparib	
		Olaparib	
		Rucaparib	
10 Trials see p. 39		Talazoparib	
BRCA2 - E1879D	none	Niraparib	
		Olaparib	
		Rucaparib	
10 Trials see p. 41		Talazoparib	
NF1 - R1362*, R2450*	none	Selumetinib	
10 Trials see p. 51		Trametinib	
CTNNB1 - K335T	none	none	
9 Trials see p. 43			
FANCL - splice site 904-2A>C	none	none	
10 Trials see p. 45			
FGFR2 - N549D	none	none	
9 Trials see p. 49			
PIK3CB - R321Q	none	none	
10 Trials see <i>p. 55</i>			
PIK3R1 - E476*	none	none	
4 Trials see p. 57			
PTEN - R130Q, R173H	none	none	
10 Trials see p. 58			
RAD51D - R253Q	none	none	
10 Trials see p. 60			

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FOUNDATIONONE®CDx



PATIENT Li, Meng-Ju TUMOR TYPE
Uterus endometrial
adenocarcinoma endometrioid
COUNTRY CODE

REPORT DATE 28 Sep 2021

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GENOMIC FINDINGS	THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)
TSC1 - Q739*	none	none
10 Trials see <i>p.</i> 62		
		NCCN category

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING IN SELECT CANCER SUSCEPTIBILITY GENES

Findings below have been previously reported as pathogenic germline in the ClinVar genomic database and were detected at an allele frequency of >10%. See appendix for details.

ATM - R250* p. 5 **MSH6 -** E641* p. 20

This report does not indicate whether variants listed above are germline or somatic in this patient. In the appropriate clinical context, follow-up germline testing would be needed to determine whether a finding is germline or somatic.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS (CH)

Genomic findings below may include nontumor somatic alterations, such as CH. The efficacy of targeting such nontumor somatic alterations is unknown. This content should be interpreted based on clinical context. Refer to appendix for additional information on CH.

MLL2 - R5214C p. 19

GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

APC - S2307L, R2204*	p. 13	MAP3K1 - G914*	p. 19
ATRX - R907*			
CARD11 - R271W	p. 15	MSH6 - E641*	p. 20
CREBBP - E50*	p. 15	POLE - V411L	p. 21
CTCF - R129*	p. 16	SETD2 - splice site 5278-1G>T	p. 21
CTNNA1 - R451*	p. 16	SPEN - A2727V	p. 22
CUL3 - L602*	p. 17	STAG2 - splice site 463-1G>T	p. 22
<i>ESR1</i> - R256Q	p. 17	TNFAIP3 - R87*	p. 23
FAM123B - E387*	p. 18	<i>TP53</i> - F113V - subclonal, T211A	p. 24
KEL - E494*	p. 18	XPO1 - R749Q	p. 25
KLHL6 - D104N	p. 18	ZNF217 - D326N	p. 25

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patients tumor type. This report should be regarded and used as upplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.



BIOMARKER FINDINGS

BIOMARKER

Tumor Mutational Burden

RESULT 372 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L11-3, anti-PD-1 therapies1-4, and combination nivolumab and ipilimumab⁵⁻¹⁰. In multiple pan-tumor studies, higher TMB has been reported to be associated with increased ORR and OS from treatment with immune checkpoint inhibitors1-4,11. Higher TMB was found to be significantly associated with improved OS upon immune checkpoint inhibitor treatment for patients with 9 types of advanced tumors1. Analyses across several solid tumor types reported that patients with higher TMB (defined as ≥16-20 Muts/Mb) achieved greater clinical benefit from PD-1 or PD-L1-targeting monotherapy, compared

with patients with higher TMB treated with chemotherapy¹² or those with lower TMB treated with PD-1 or PD-L1-targeting agents². However, the KEYNOTE 158 trial of pembrolizumab monotherapy for patients with solid tumors found significant improvement in ORR for patients with TMB ≥10 Muts/Mb (based on this assay or others) compared to those with TMB <10 Muts/Mb, in a large cohort that included multiple tumor types; similar findings were observed in the KEYNOTE 028 and 012 trials⁴.¹¹¹. Together, these studies suggest that patients with TMB ≥10 Muts/Mb may derive clinical benefit from PD-1 or PD-L1 inhibitors.

FREQUENCY & PROGNOSIS

A large-scale genomic analysis found that endometrial adenocarcinomas harbored a median TMB of 4.5 Muts/Mb, and 15% of cases had an elevated TMB of greater than 20 Muts/Mb¹³. Another study evaluating TMB in endometrial adenocarcinoma reported that 24% of tumors had a mutational burden of greater than 10.4 Muts/Mb¹⁴. Increased tumor mutational burden (TMB) in endometrial carcinoma has been correlated with POLE mutation and advanced high-grade endometrioid subtypes¹¹5-¹8. Ultramutated endometrial tumors (elevated TMB with POLE

mutations) have also been associated with improved PFS¹6. The same study associated lower mutational burden, independent of PD-L1 status, in endometrial carcinomas with poorer prognosis¹6. For patients with advanced microsatellite-stable endometrial carcinoma not treated with immunotherapy, OS did not significantly differ between patients with TMBhigh (≥10 Muts/Mb) and TMB-low (11.4 vs. 13.5 months, adjusted HR=1.15) in 1 study¹9.

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma²⁰⁻²¹ and cigarette smoke in lung cancer²²⁻²³, treatment with temozolomide-based chemotherapy in glioma²⁴⁻²⁵, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes^{16,26-29}, and microsatellite instability (MSI)^{16,28-29}. This sample harbors a TMB level that may be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors in multiple solid tumor types^{2-4,11}.

BIOMARKER

Microsatellite status

RESULT MS-Stable

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors³⁰⁻³², including approved therapies nivolumab and pembrolizumab³³. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and experienced a significantly higher ORR

compared with non-MSI-H cases (70% vs. 12%, p=0.001) 34 .

FREQUENCY & PROGNOSIS

MSS has been reported in 73-89% of endometrial cancers ^{16,18,35-40}. Data regarding the role of MSI status on prognosis and survival in endometrial cancer are conflicting, with most studies finding no relationship between MSI-H endometrial cancers and survival ^{36-37,39,41-43}, and one study predicting improved disease-free and disease-specific survival ³⁵. However, these studies often evaluated endometrial cancers of all FIGO stages together. Studies specifically analyzing early stage endometrial cancer have reported a correlation between MSI-H and decreased survival ^{36,40,44-45}, thereby suggesting that MSI-H predicts for poor prognosis in this subset of endometrial tumors.

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor⁴⁶. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH₂, MSH₆, or PMS₂⁴⁶⁻⁴⁸. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor; one with mutations in none of the tested microsatellite markers⁴⁹⁻⁵¹. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins $^{46,48,50-51}$.



GENOMIC FINDINGS

GENE

ATM

ALTERATION R250*

TRANSCRIPT ID NM 000051

CODING SEQUENCE EFFECT

748C>T

VARIANT ALLELE FREQUENCY (% VAF) 34.1%

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies —

Loss of functional ATM results in a defective DNA damage response and homologous recombinationmediated DNA repair and may predict sensitivity to PARP inhibitors⁵². Clinical data in prostate cancer⁵³⁻⁵⁵, gastric cancer⁵⁶, colorectal cancer⁵⁷, breast cancer⁵⁷, papillary renal cell carcinoma⁵⁸, and cholangiocarcinoma⁵⁹ indicate that loss or inactivation of ATM may confer sensitivity to PARP inhibitors⁶⁰⁻⁶⁷. In Phase 1 trials of ATR inhibitors, a heavily pretreated patient with colorectal cancer who achieved a CR to berzosertib68 and 4 out of 4 patients with diverse solid tumors who achieved PRs to BAY189534469 harbored ATM inactivation or protein loss; studies showing reduced cell viability and increased DNA damage in preclinical models of solid tumors⁷⁰⁻⁷² and hematologic malignancies^{70,73} also support the increased sensitivity of ATM-deficient cells to ATR inhibitors. Preclinical experiments also indicate that loss of ATM causes dependency on DNA-PKcs

in cancer cells; DNA-PKcs inhibitors promoted apoptosis in ATM-deficient cells and were active in a lymphoma mouse model lacking ATM activity74.

FREQUENCY & PROGNOSIS

ATM mutation and homozygous deletion have been observed in 11-12% and up to 1.8% (5/761) samples of endometrial carcinoma, respectively (cBioPortal, COSMIC, Mar 2021)75-77. Altered expression of ATM has been reported in 4.5% (1/ 22) of endometrial hyperplasia samples and in 3.3% (2/61) of endometrioid-type endometrial cancer samples analyzed in one study78. Hypermethylation of ATM has been reported in 11% of endometrial carcinomas⁷⁹. Conflicting findings have been reported regarding the prognostic implications of ATM expression in endometrial carcinomas, with one study reporting a negative correlation between ATM expression and pathological grade80 and another a positive correlation between increased ATM expression and unfavorable clinicopathologic features, including increased tumor grade and reduced 5-year recurrence free survival81.

FINDING SUMMARY

ATM encodes the protein ataxia telangiectasia mutated, which is a serine/threonine protein kinase that plays a key role in the DNA damage response⁸². Loss of functional ATM promotes tumorigenesis83. Alterations such as seen here may disrupt ATM function or expression84-86.

POTENTIAL GERMLINE IMPLICATIONS

One or more of the ATM variants observed here has been described in the ClinVar database as a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with ataxia-telangiectasia syndrome (ClinVar, Mar 2021)87. Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. ATM mutation carriers have increased cancer risk, with female carriers displaying a 38% lifetime risk of breast cancer88. Biallelic mutations in ATM underlie the rare autosomal-recessive inherited disorder ataxia-telangiectasia (A-T), also referred to as genome instability or DNA damage response syndrome⁸⁹. This disease is characterized by genomic instability, sensitivity to DNA-damaging agents, and increased risk of developing cancer^{82,89}. The prevalence of A-T is estimated at 1:40,000 to 1:100,000 worldwide89. In the appropriate clinical context, germline testing of ATM is recommended.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion90-95. Comprehensive genomic profiling of solid tumors may detect nontumor alterations that are due to CH94,96-97. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to



GENOMIC FINDINGS

GENE

BRCA2

ALTERATION E1879D

TRANSCRIPT ID

CODING SEQUENCE EFFECT

5637G>T

VARIANT ALLELE FREQUENCY (% VAF) 10.1%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

Alterations that inactivate BRCA1 or BRCA2 may confer sensitivity to PARP inhibitors53,98-114 or ATR inhibitors^{69,115-116}. Clinical responses to PARP inhibitors have been reported for patients with either germline or somatic BRCA1/2 mutations^{53,103,106,113-114} and for patients with platinum-resistant or -refractory disease^{98,102,109,112}. In a case study, a patient with therapy-induced neuroendocrine prostate cancer and an inactivating BRCA2 rearrangement experienced a CR ongoing for 20 months to the ATR inhibitor berzosertib¹¹⁶. Preclinical studies of BRCA_{1/2} inactivation in T-cell acute lymphoblastic leukemia (T-ALL)117, ovarian carcinoma118, and triple-negative breast cancer (TNBC)119 showing reduced cell viability and increased DNA damage during ATR treatment further support the sensitivity of BRCA2-deficient cells to ATR inhibitors. The WEE1 inhibitor adavosertib has been evaluated as a monotherapy and in

combination with PARP-inhibitor, olaparib. In a Phase 2 study for patients with PARP-resistant ovarian cancer, the combination of olaparib and adavosertib elicited improved clinical benefit (ORR: 29%; DCR: 89%) compared to adavosertib alone (ORR: 23%; DCR: 63%); however, in the BRCA-mutated cohort, no significant difference in clinical benefit was observed between the combination (ORR: 19%) and monotherapy (ORR: 20%) treatments¹²⁰. In a Phase 1 monotherapy trial of adavosertib that included 9 patients with BRCA₁/₂-mutated solid tumors, 2 patients with BRCA₁-mutated cancers (1 with ovarian serous carcinoma and 1 with oral squamous cell carcinoma) achieved PRs, and a third patient with ovarian serous carcinoma harboring mutations in BRCA1 and TP53 experienced 14% tumor shrinkage prior to disease progression¹²¹. It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

Nontargeted Approaches —

Inactivation of BRCA2 may also predict sensitivity to DNA-damaging drugs such as trabectedin, lurbinectedin, and the platinum chemotherapies cisplatin and carboplatin¹²²⁻¹³².

FREQUENCY & PROGNOSIS

BRCA2 mutation has been reported in 10% of uterine corpus endometrial and endometrioid carcinoma cases analyzed in the TCGA dataset¹⁶. LOH of the wild type BRCA1/2 allele in patients with endometrial carcinoma harboring germline BRCA1/2 mutations has been associated with higher FIGO grade, nonendometrioid histology,

and frequent co-occurrence of TP53 mutation¹³³. Studies have reported that BRCA2 germline mutations do not increase the lifetime risk of endometrial cancer¹³⁴⁻¹³⁵.

FINDING SUMMARY

The BRCA2 tumor suppressor gene encodes a protein that regulates the response to DNA damage¹³⁶. Inactivating mutations in BRCA2 can lead to the inability to repair DNA damage and loss of cell cycle checkpoints, which can lead to tumorigenesis¹³⁷. Although alterations such as seen here have not been fully characterized and are of unknown functional significance, similar alterations have been previously reported in the context of cancer, which may indicate biological relevance.

POTENTIAL GERMLINE IMPLICATIONS

Inactivating germline mutations in BRCA1 or BRCA2 are associated with autosomal dominant hereditary breast and ovarian cancer 138-139, and the lifetime risk of breast and ovarian cancer in BRCA2 mutation carriers has been estimated to be as high as >80% and 23%, respectively¹⁴⁰. Elevated risk for other cancer types, including gastric, pancreatic, prostate, and colorectal, has also been identified, with an increase in risk ranging from 20 to 60%141. The estimated prevalence of deleterious germline BRCA1/2 mutations in the general population is between 1:400 and 1:800, with an approximately 10-fold higher prevalence in the Ashkenazi Jewish population^{140,142-147}. In the appropriate clinical context, germline testing of BRCA2 is recommended.



GENOMIC FINDINGS

GENE

FBXW7

ALTERATION R441W

TRANSCRIPT ID

CODING SEQUENCE EFFECT

1321C>T

VARIANT ALLELE FREQUENCY (% VAF)

34.9%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

FBXW7 inactivating alterations may indicate sensitivity to mTOR inhibitors¹⁴⁸⁻¹⁴⁹. Several case studies reported clinical benefit for patients with FBXW7-mutated cancers, including lung

adenocarcinoma¹⁵⁰, renal cell carcinoma⁵⁸, and cervical squamous cell carcinoma¹⁵¹. It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

Nontargeted Approaches —

FBXW7 inactivation may also result in resistance to anti-tubulin chemotherapies based on results from preclinical studies¹⁵².

FREQUENCY & PROGNOSIS

FBXW7 mutations have been reported in 14.2% of endometrial carcinomas (COSMIC, Feb 2021)⁷⁷. Studies have variously reported FBXW7 mutation in 18-29% of uterine serous carcinoma, 16% of endometrial cancer, and 2% of endometrioid endometrial cancer cases¹⁵³⁻¹⁵⁷. In primary endometrial carcinomas, FBXW7 mutations correlated with lymph node involvement¹⁵⁸.

Reduced FBXW7 expression has been associated with poor prognosis in some cancers such as colorectal cancer, gastric cancer, esophageal SCC, cervical SCC, melanoma, and non-small cell lung carcinoma¹⁵⁹⁻¹⁶⁶.

FINDING SUMMARY

FBXW7 encodes the F-box protein subunit of the SCF ubiquitin ligase complex, which targets proteins for degradation¹⁶⁷. FBXW7 inactivation is associated with chromosomal instability and with stabilization of proto-oncogenes, such as mTOR, MYC, cyclin E, NOTCH, and JUN; FBXW7 is therefore considered a tumor suppressor¹⁶⁷⁻¹⁶⁸. Although alterations such as seen here have not been fully characterized and are of unknown functional significance, similar alterations have been previously reported in the context of cancer, which may indicate biological relevance.

GENE

NF1

ALTERATION R1362*, R2450*

TRANSCRIPT IDNM_001042492, NM_001042492

CODING SEQUENCE EFFECT 4084C>T, 7348C>T

VARIANT ALLELE FREQUENCY (% VAF)

34.5%, 71.5%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

On the basis of clinical evidence in neurofibromatosis Type 1-associated neurofibroma¹⁶⁹⁻¹⁷², glioma or glioblastoma¹⁷²⁻¹⁷⁶, non-small cell lung cancer¹⁷⁷, NF1 inactivation may predict sensitivity to MEK inhibitors such as cobimetinib, trametinib, binimetinib, and selumetinib. Loss or inactivation of NF1 may also predict sensitivity to mTOR inhibitors, including the approved agents everolimus and temsirolimus, based on limited clinical data¹⁷⁸⁻¹⁸⁰ and strong

preclinical data in models of malignant peripheral nerve sheath tumor (MPNST)¹⁸¹⁻¹⁸². A preclinical study suggests that combined mTOR and MEK inhibition is effective in a model of NF1-deficient MPNST¹⁸³. Whereas frequent adverse events precluded a recommended Phase 2 dose and schedule for the combination of trametinib and everolimus in a Phase 1b trial for solid tumors¹⁸⁴, a retrospective study for heavily pretreated patients with solid tumors reported tolerable regimens of the combination for 23/31 patients, with 16 patients treated >3 months and evaluable patients achieving a median PFS of 6.5 months¹⁸⁵.

FREQUENCY & PROGNOSIS

NF1 alterations have been reported in 5-9% of endometrial carcinomas^{16,186-187}. Loss of NF1 has been reported in 13% (4/31) of endometrial carcinomas¹⁸⁸. Published data investigating the prognostic implications of NF1 mutation in endometrial carcinomas are limited (PubMed, Feb 2021).

FINDING SUMMARY

NF1 encodes neurofibromin, a GTPase-activating protein (GAP) that is a key negative regulator of

the RAS signaling pathway¹⁸⁹. Neurofibromin acts as a tumor suppressor by repressing RAS signaling¹⁹⁰. Alterations such as seen here may disrupt NF1 function or expression¹⁹⁰⁻¹⁹⁹.

POTENTIAL GERMLINE IMPLICATIONS

One or more of the NF1 variants observed here has been described in the ClinVar database as a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with neurofibromatosis type 1 (ClinVar, Mar 2021)87. Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. Germline mutations in NF1 cause the autosomal dominant disorder neurofibromatosis type 1, which is characterized in part by increased risk of developing various tumors, including sarcoma, glioma, breast carcinoma, and neuroendocrine and hematological neoplasms²⁰⁰⁻²⁰². Estimates for the prevalence of the disorder in the general population range from 1:2,500 to 1:3,000²⁰³⁻²⁰⁴, and in the appropriate clinical context, germline testing of NF1 is recommended.



GENOMIC FINDINGS

GENE

PIK3CA

ALTERATION R88Q, T1025A

R88Q, T1025A

TRANSCRIPT ID

NM_006218, NM_006218

CODING SEQUENCE EFFECT

263G>A, 3073A>G

VARIANT ALLELE FREQUENCY (% VAF)

36.3%, 37.0%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

Clinical and preclinical data in various tumor types indicate that PIK₃CA activating alterations may predict sensitivity to therapies targeting PI₃K²⁰⁵⁻²⁰⁷, AKT²⁰⁸⁻²⁰⁹, or mTOR^{178,210-216}. Results from the Phase 2 MATCH trial for patients with PIK₃CA-altered solid tumors found that 27% (18/65) of patients experienced PFS lasting at least 6 months after treatment with taselisib; however, no ORs were observed in this study²¹⁷. In a Phase 1

trial of the dual PI₃K/mTOR kinase inhibitor apitolisib, 79% (11/14) of patients with PIK₃CA-mutated advanced solid tumors experienced disease control (3 PRs, 8 SDs)²¹⁸. The PI₃K inhibitor alpelisib demonstrated an ORR of 6.0% (8/134) and a DCR of 58% (78/134) in a study of PIK₃CA-mutated solid tumors²¹⁹. However, the PI₃K inhibitor copanlisib exhibited limited efficacy in PIK₃CA-altered tumors²²⁰⁻²²¹.

Potential Resistance —

Activating mutations in PIK₃CA may confer resistance to HER₂-targeted therapies; combined inhibition of HER₂ and the PI₃K pathway may be required in HER₂-positive tumors with PIK₃CA mutation²²²⁻²²⁶.

FREQUENCY & PROGNOSIS

In the scientific literature, PIK₃CA mutations have been reported in 16-54% of endometrial carcinomas^{16,158,227}. In endometrial cancers, PIK₃CA mutations often co-occur with other mutations that activate the PI₃K-AKT-mTOR signaling axis, such as PTEN and KRAS alterations²²⁸⁻²²⁹. Overexpression of p110-alpha

has been reported in 72% of endometrial carcinomas²³⁰. One study reported PIK₃CA exon 9 or 20 mutations in 20% (20/99) of high-grade endometrial carcinomas; these mutations were associated with shorter patient survival within Grade 3 endometrioid carcinoma, but not within endometrial serous carcinoma²³¹.

FINDING SUMMARY

PIK₃CA encodes p₁₁₀-alpha, which is the catalytic subunit of phosphatidylinositol ₃-kinase (PI₃K). The PI₃K pathway is involved in cell signaling that regulates a number of critical cellular functions, including cell growth, proliferation, differentiation, motility, and survival²³²⁻²³³. PIK₃CA alterations that have been characterized as activating, such as R88Q observed here, are predicted to be oncogenic^{227,229,234-252}. Although alterations such as T₁₀₂5A seen here have not been fully characterized, they have been associated with sensitivity to targeted therapies or have shown cancer association, which may indicate biological relevance^{75-77,246,253-255}.



GENOMIC FINDINGS

GENE

CTNNB1

ALTERATION K335T

TRANSCRIPT ID

CODING SEQUENCE EFFECT

1004A>C

VARIANT ALLELE FREQUENCY (% VAF)

38.4%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

Mutation or activation of CTNNB1 signaling has been shown to increase mTOR signaling, promote tumorigenesis, and respond to mTOR inhibition in preclinical studies²⁵⁶⁻²⁵⁸. Small studies have reported clinical benefit following treatment of everolimus combined with other targeted agents for patients with CTNNB1-mutated hepatocellular carcinoma^{180,259} or endometrial carcinoma²⁵⁴. In preclinical studies, CTNNB1 activating mutations have been shown to increase expression of WNT pathway member DKK1, which may promote tumor cell proliferation and immune evasion²⁶⁰⁻²⁶². A Phase 1 trial of DKK1-targeting

antibody DKN-01 in combination with paclitaxel in esophageal cancer reported a PR rate in 2 out of 4 patients and SD rate of in 1 out of 4 patients with CTNNB1 activating mutations, compared with 24% (10/41) PR and 37% (15/41) SD in unselected patients²⁶³. Multiple preclinical studies in cancer models harboring CTNNB1 mutation or beta-catenin pathway activation have reported activation of the NOTCH pathway and sensitivity to pharmacologic inhibition of NOTCH signaling by gamma-secretase inhibitors²⁶⁴⁻²⁶⁷. Phase 1 and 2 clinical trials of gamma-secretase inhibitor PF-03084014 have shown high response rates in patients with desmoid tumors, which are driven by activating CTNNB1 mutations in the majority of cases²⁶⁸⁻²⁶⁹, suggesting CTNNB1-mutated tumors may be sensitive to gamma-secretase inhibitors. Although WNT pathway inhibitors have been explored preclinically in CTNNB1-mutated cells, clinical data supporting this therapeutic approach are lacking^{257,270-272}. It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

FREQUENCY & PROGNOSIS

CTNNB1 mutations have been reported in 7-45% of endometrial carcinomas (ECs)^{16,273-276}. CTNNB1 mutations are more common in Type 1 EC than

Type 2²⁷⁷⁻²⁷⁸. In addition, one study found that CTNNB1 mutations were identified more frequently in sporadic ECs (31%, 18/58), than in Lynch syndrome (LS)-associated ECs (6.9%, 2/ $29)^{276}$. Nuclear beta-catenin protein expression has been observed in 14.7-27.6% (33/224-55/199) of ECs, with a significantly higher incidence in Type 1 tumors^{274,279}. Multiple studies have reported that CTNNB1 mutation characterizes an aggressive subset of $EC^{44,280-281}$. One study found that that TP53 or CTNNB1 mutation was an independent marker of poor recurrence-free survival (HR=4.69) for patients with low grade, early stage EC²⁸⁰. Low membrane expression of beta-catenin has been linked with poor prognosis in EC and ovarian endometrioid carcinomas²⁸²⁻²⁸³.

FINDING SUMMARY

CTNNB1 encodes beta-catenin, a key downstream component of the WNT signaling pathway. Beta-catenin interacts with cadherin to regulate cell-cell adhesion; as a component of the WNT pathway, it also plays a role in development, cell proliferation, and cell differentiation²⁸⁴. Although alterations such as seen here have not been fully characterized and are of unknown functional significance, similar alterations have been previously reported in the context of cancer, which may indicate biological relevance.

GENE

FANCL

ALTERATION

splice site 904-2A>C

TRANSCRIPT ID

NM_018062

CODING SEQUENCE EFFECT

904-2A>C

VARIANT ALLELE FREQUENCY (% VAF)

32.6%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no targeted therapies that directly address genomic alterations in FANCL. Clinical

evidence in ovarian cancer indicates that FANCL inactivation may confer sensitivity to PARP inhibitors $^{285-286}$.

FREQUENCY & PROGNOSIS

FANCL mutations are most frequently observed in tumors of the prostate (5.9%), liver (4.1%), biliary tract (3.4%), endometrium (3.2%), pancreas (2.7%), large intestine (2.4%), and stomach (2.4%) (COSMIC, May 2021)⁷⁷. Published data investigating the prognostic implications of FANCL alterations in solid tumors and hematologic malignancies are limited (PubMed, May 2021). In a prospective study of 255 patients with follicular lymphoma, 2p gain, which includes VRK2, FANCL, and LINC01122, was associated with worse PFS and OS in multivariate analysis²⁸⁷.

FINDING SUMMARY

FANCL encodes a member of the Fanconi anemia nuclear complex, a multiprotein structure also including the products of FANCA, FANCC, FANCF and FANCG. The activity of this complex is essential to prevention of chromosome breakage caused by DNA damage²⁸⁸. Germline mutations in FANCL cause Fanconi anemia, a clinically heterogeneous disorder involving various developmental abnormalities as well as predisposition to cancer; underlying these phenotypes are defects in DNA repair²⁸⁹. Alterations such as seen here may disrupt FANCL function or expression²⁹⁰⁻²⁹⁷. Germline mutations in FANCL, such as T367fs*13, have been associated with Fanconi anemia, breast cancer, and ovarian cancer and with an increased risk of esophageal cancer and prostate cancer²⁹⁸⁻³⁰².



GENOMIC FINDINGS

GENE

FGFR2

ALTERATION N549D

TRANSCRIPT ID

NM_000141

CODING SEQUENCE EFFECT

1645A>G

VARIANT ALLELE FREQUENCY (% VAF)

34.8%

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies —

FGFR2 activating mutations, amplifications, or fusions may confer sensitivity to selective FGFR inhibitors such as erdafitinib³⁰³, pemigatinib^{304,305}, infigratinib³⁰⁶, E7090³⁰⁷, AZD4547³⁰⁸⁻³¹⁰, Debio 1347³¹¹⁻³¹², rogaratinib³¹³, futibatinib³¹⁴, and derazantinib³¹⁵ as well as to the multikinase inhibitors pazopanib³¹⁶⁻³¹⁷ and ponatinib³¹⁸. Cases of clinical response to selective

FGFR inhibitors have been reported for patients with various FGFR2-mutated tumors, including PRs for patients with FGFR2-mutated cholangiocarcinoma treated with AZD4547³¹⁹ or erdafitinib³²⁰ and a PR for a patient with FGFR2-mutated lung squamous cell carcinoma treated with infigratinib³²¹. A patient with endometrioid adenocarcinoma and cervical cancer and both an FGFR2 amplification and mutation experienced a PR following treatment with Debio-1347³¹¹. Additional clinical responses associated with FGFR2 mutation have been reported for a patient with HNSCC³¹⁷, and in a patient with solitary fibrous tumor treated with pazopanib³²².

FREQUENCY & PROGNOSIS

FGFR2 mutations have been reported in 12.5% of cases in the Uterine Corpus Endometrioid Carcinoma TCGA dataset¹⁶. In the scientific literature, mutation of FGFR2 has been identified in 10.3% (48/466) of endometrial carcinoma (EC) tumors in one study, and at similar frequencies in others³²³⁻³²⁵. FGFR2 expression has been identified

in 71.9% (23/32) of EC specimens in one study; however, levels of FGFR2 protein or FGFR2 mRNA have not been consistently demonstrated to be higher in tumor specimens compared to normal endometrial tissues³²⁶⁻³²⁸. FGFR2 mutations in endometrioid endometrial tumors have been correlated with decreased disease-free survival in one study³²⁵.

FINDING SUMMARY

FGFR2 encodes a tyrosine kinase cell surface receptor, which plays an important role in cell differentiation, growth, and angiogenesis³²⁹⁻³³⁰. FGFR2 mutations that have been characterized as activating, such as observed here, are predicted to be oncogenic^{324-325,331-341}. FGFR2 activating alterations that affect amino acid E565 and mutations such as S252W, P253R, W290C, S372C, Y375C, N549K, and K659E have been reported in patients with Apert, Pfeiffer, Beare-Stevenson, and Crouzon syndromes and syndromic craniosynostosis^{324,341-349}.

GENE

PIK3CB

ALTERATION

R321Q

TRANSCRIPT ID

CODING SEQUENCE FEFECT

962G>A

VARIANT ALLELE FREQUENCY (% VAF)

36.1%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

Preclinical evidence indicates that PTEN-deficient tumors depend on PI₃K-beta³⁵⁰⁻³⁵², and inhibitors selective for PI₃K-beta, including GSK2636771 and AZD8186, are in clinical trials for PTEN-deficient tumors³⁵³⁻³⁵⁴. PIK₃CB activating alterations may

further refine the use of PI₃K-beta-selective inhibitors³⁵⁵. Two patients with PTEN-deficient prostate tumors and concurrent PIK₃CB L₁₀₄₉R mutation or PIK₃CB amplification derived significant clinical benefit from treatment with GSK₂6₃6₇₇₁356</sup>. It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

FREQUENCY & PROGNOSIS

In the Uterine Corpus Endometrial Carcinoma TCGA dataset, PIK₃CB mutations and amplification have been observed in 5.4% and 2.1% of cases, respectively 16. Published data investigating the prognostic implications of PIK₃CB alterations in endometrial carcinoma are limited (PubMed, Oct 2020). However, a preclinical study reported that elevated PIK₃CB mRNA levels contributed to early endometrial tumorigenesis and increased cellular proliferation

of primary tumors357.

FINDING SUMMARY

PIK3CB (PI3K-beta, p110-beta) encodes a member of the class IA phosphoinositide 3-kinases (PIK3CA, PIK3CB, PIK3CD, and PIK3CG), which are essential regulators of cellular proliferation, survival, metabolism, and motility. Dysregulation of the PI₃K signaling pathway is observed in many human cancers and occurs most frequently through loss of the tumor suppressor PTEN or activating mutations in PIK₃CA (p₁₁₀-alpha). Contrary to PIK3CA, which is frequently mutated within hot spots causing increased kinase activity, mutation of PIK₃CB is quite $rare^{355,358-359}$. Although alterations such as seen here have not been fully characterized and are of unknown functional significance, similar alterations have been previously reported in the context of cancer, which may indicate biological relevance.



GENOMIC FINDINGS

GENE

PIK3R1

ALTERATION E476*

TRANSCRIPT ID NM_181523

CODING SEQUENCE EFFECT

1426G>T

VARIANT ALLELE FREQUENCY (% VAF)

34.8%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

On the basis of clinical³⁶⁰⁻³⁶¹ and preclinical³⁶²⁻³⁶³ data, PIK₃R₁ alteration may predict sensitivity to pan-PI₃K or PI₃K-alpha-selective inhibitors. In patients with PIK₃R₁ mutation and no other alterations in the PI₃K-AKT-mTOR pathway, 2

CRs have been achieved by patients with endometrial cancer treated with the pan-PI₃K inhibitor pilaralisib³⁶⁰, and 1 PR has been achieved by a patient with breast cancer treated with the PI₃K-alpha inhibitor alpelisib in combination with ribociclib and letrozole³⁶⁴. Limited clinical and preclinical data suggest that PIK₃R₁ alterations may also be sensitive to inhibitors of mTOR^{186,215,363,365-366} or AKT³⁶⁷⁻³⁶⁸. One preclinical study reported that PIK₃R₁ truncation mutations in the 299–370 range confer sensitivity to MEK inhibitors³⁶⁹.

FREQUENCY & PROGNOSIS

In the TCGA datasets, PIK₃R₁ mutation is most frequently observed in endometrial carcinoma (33%)¹⁶, glioblastoma (GBM; 11%)³⁷⁰, uterine carcinosarcoma (11%)(cBioPortal, 2021)⁷⁵⁻⁷⁶, and lower grade glioma (5%)³⁷¹. PIK₃R₁ is often inactivated by in-frame insertions or deletions (indels), and the majority of this class of mutation

(80%) was observed in endometrial carcinoma³⁷²⁻³⁷⁴, although PIK₃R₁ indels have been reported in other cancer types such as GBM, cervical squamous cell carcinoma, and urothelial bladder carcinoma³⁷². On the basis of limited clinical data, reduced PIK₃R₁ expression has been associated with reduced disease-free survival in prostate cancer³⁷⁵ and metastasis-free survival in breast cancer³⁷⁶. PIK₃R₁ expression is not associated with overall survival in neuroendocrine tumors³⁷⁷.

FINDING SUMMARY

PIK₃R₁ encodes the p8₅-alpha regulatory subunit of phosphatidylinositol ₃-kinase (PI₃K)³⁷⁸. Loss of PIK₃R₁ has been shown to result in increased PI₃K signaling³⁷⁹⁻³⁸², promote tumorigenesis^{362,367,379}, and promote hyperplasia in the context of PTEN-deficiency³⁸³. Alterations such as seen here may disrupt PIK₃R₁ function or expression^{240,252,363,368-369,373-374,384-390}.

GENE

PTEN

ALTERATION R130Q, R173H

TRANSCRIPT ID NM_000314, NM_000314

CODING SEQUENCE EFFECT

389G>A, 518G>A

VARIANT ALLELE FREQUENCY (% VAF)

35.0%, 32.4%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

PTEN loss or mutation leads to activation of the PI₃K-AKT-mTOR pathway and may predict sensitivity to inhibitors of this pathway^{221,391-393}. Clinical studies in endometrial cancer have not observed an association between PTEN deficiency and response to the mTOR inhibitors everolimus or temsirolimus^{186,394-400}. Preclinical data indicate that PTEN loss or inactivation may predict sensitivity to PARP inhibitors⁴⁰¹⁻⁴⁰⁵, and clinical benefit has been observed for patients with PTEN-altered breast cancer including triple negative breast cancer⁴⁰⁶, ovarian cancer²⁸⁶, uterine leiomyosarcoma⁴⁰⁷, and endometrial cancer⁴⁰⁵

treated with PARP inhibitors. However, some studies have reported a lack of association between PTEN mutation and PARP inhibitor sensitivity^{102,408}. In a Phase 1 study of patients treated with PARP and AKT inhibitors olaparib and capivasertib, two patients with PTEN-mutated ovarian cancer and a patient with PTEN-mutated endometrial cancer achieved clinical benefit (CR, PR, or SD >4 months)⁴⁰⁹.

FREQUENCY & PROGNOSIS

PTEN mutation has been observed in 55-65% of endometrial cancers and in 70% of primary endometrioid endometrial cancers, whereas PTEN loss has been reported in 5% of endometrial carcinoma cases 16,410-411. PTEN mutation or loss has been suggested to play a role in endometrial cancer progression; high PTEN expression has been observed in Grade 1 and 2 tumors but not in Grade 3 tumors 230,412-414. Loss of PTEN protein expression has been correlated with increased survival of patients with Stage 3 or 4 endometrial carcinoma, but not Stage 1 or 2²³⁰.

FINDING SUMMARY

PTEN encodes an inositol phosphatase that functions as a tumor suppressor by negatively regulating the PI₃K-AKT-mTOR pathway; loss of PTEN can lead to uncontrolled cell growth and

suppression of apoptosis³⁹². Alterations such as seen here may disrupt PTEN function or expression⁴¹⁵⁻⁴⁵⁵.

POTENTIAL GERMLINE IMPLICATIONS

One or more of the PTEN variants observed here has been described in the ClinVar database as a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with hamartoma tumor syndrome (ClinVar, Mar 2021)87. Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. PTEN mutations underlie several inherited disorders, collectively termed PTEN hamartoma tumor syndrome (PHTS), which include Cowden syndrome (CS) and its variant Lhermitte-Duclos disease (LD), Bannayan-Riley-Ruvalcaba syndrome (BRRS), PTEN-related Proteus syndrome (PS), and Proteus-like syndrome⁴⁵⁶⁻⁴⁵⁷. The mutation rate for PTEN in these disorders ranges from 20 to 85% of patients 456,458 . The estimated incidence of Cowden syndrome is 1/ 200,000, which may be an underestimate due to the high variability of this disorder⁴⁵⁶. Given the association between PTEN and these inherited syndromes, in the appropriate clinical context, germline testing for mutations affecting PTEN is recommended.



GENOMIC FINDINGS

GENE

RAD51D

ALTERATION R253O

TRANSCRIPT ID

NM_002878

CODING SEQUENCE EFFECT

758G>A

VARIANT ALLELE FREQUENCY (% VAF)

1.0%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

Limited preclinical data⁴⁵⁹⁻⁴⁶⁰ and clinical evidence in ovarian cancer^{103,461} indicate that loss or inactivation of RAD₅₁D may confer sensitivity to PARP inhibitors. Loss of functional RAD₅₁D may also predict sensitivity to DNA-damaging drugs such as mitomycin C and cisplatin^{459,462-464}.

It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

FREQUENCY & PROGNOSIS

RAD51D mutation has been observed in 3.1% of endometrioid carcinomas, but not in any samples of other endometrial histologies analyzed in the COSMIC database (Sep 2021)⁷⁷. Published data investigating the prognostic implications of RAD51D alteration in endometrial carcinomas are limited (PubMed, Sep 2021).

FINDING SUMMARY

RAD51D, also known as RAD51L3, is involved in homologous recombination-mediated DNA repair and telomere maintenance⁴⁶⁵⁻⁴⁶⁸. Germline mutations in RAD51D have been associated with hereditary breast and ovarian cancer^{460,469-472}, and RAD51D mutation carriers have an increased lifetime risk of ovarian cancer, estimated to be

10-12% 460.473. Although alterations such as seen here have not been fully characterized and are of unknown functional significance, similar alterations have been previously reported in the context of cancer, which may indicate biological relevance.

POTENTIAL GERMLINE IMPLICATIONS

Inactivating germline mutations in RAD51D are associated with hereditary breast and ovarian cancer (HBOC) syndrome, an autosomal dominant disorder that predisposes patients to breast and ovarian malignancies⁴⁷⁴⁻⁴⁷⁵. The risk of ovarian cancer in RAD51D mutation carriers has been estimated to be 10 to 12%^{460,473}. Germline RAD51D mutation has been reported at frequencies of up to 1% in breast and ovarian familial cancer populations without BRCA1/2 mutation^{472,476-477}. In the appropriate clinical context, germline testing of RAD51D is recommended.

GENE

TSC1

ALTERATION

Q739*

TRANSCRIPT ID NM 000368

CODING SEQUENCE EFFECT

2215C>T

VARIANT ALLELE FREQUENCY (% VAF)

35.6%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

Loss or inactivation of TSC1 can activate mTOR signaling⁴⁷⁸⁻⁴⁷⁹; however, response rates for patients with TSC1-mutated solid tumors treated with MTOR inhibitors such as everolimus and temsirolimus have been low⁴⁸⁰⁻⁴⁸². In the prospective NCI-MATCH study, the ORR for

patients with various TSC1-mutated solid tumors treated with everolimus was 7.7% (1/13); the single response was reported for a patient with urothelial cancer⁴⁸⁰. In TSC1-mutated renal cell carcinoma (RCC), responses to MTOR inhibitors have been described in multiple case series and reports^{178,483-486}, but retrospective analysis of a broader cohort showed no responses in TSC1-mutated RCC (o/7)⁴⁸¹. Retrospective analyses of the RECORD-3, GRANITE-1, and EVOLVE-1 studies of everolimus to treat patients with RCC, gastric cancer, or hepatocellular carcinoma, respectively, showed no significant association between alterations in MTOR, TSC1, or TSC2 and median PFS⁴⁸².

FREQUENCY & PROGNOSIS

TSC1 mutations have been reported in 5.2% of endometrial carcinomas analyzed in COSMIC (Sep 2021)⁷⁷. Published data investigating the prognostic implications of TSC1 alterations in endometrial cancer are limited (PubMed, Oct

2020).

FINDING SUMMARY

TSC1 encodes the protein Hamartin, which interacts with Tuberin, the gene product of TSC2, to inhibit and regulate mTOR activity^{478,487}. Alterations such as seen here may disrupt TSC1 function or expression⁴⁸⁸⁻⁴⁹⁰.

POTENTIAL GERMLINE IMPLICATIONS

Inactivating germline mutations in TSC1 are associated with the autosomal dominant disorder tuberous sclerosis complex, which results in the development of hamartomas in multiple organ systems and an increased risk of developing renal cell carcinoma⁴⁹¹⁻⁴⁹². TSC1 mutations account for approximately 10 to 30% of reported sporadic cases⁴⁹³. Prevalence for this disorder in the general population is estimated to be 1/6,000 from birth and 1/12,000 to 1/14,000 in children under 10 years of age⁴⁹⁴. In the appropriate clinical context, germline testing of TSC1 is recommended.



GENOMIC FINDINGS

GENE

APC

ALTERATION S2307L, R2204*

TRANSCRIPT ID

NM_000038, NM_000038

CODING SEQUENCE EFFECT 6920C>T. 6610C>T

VARIANT ALLELE FREQUENCY (% VAF) 35.6%, 36.5%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no approved drugs targeted to APC defects or WNT upregulation in solid tumors. Preclinical studies have reported that APC inactivation or beta-catenin activation confer synthetic lethality when TRAIL receptors are upregulated and the TRAIL death receptor program is activated⁴⁹⁵. In addition, the COX-2 inhibitor celecoxib was shown to reduce WNT signaling in cancer cell lines⁴⁹⁶⁻⁴⁹⁷. A preclinical study has found that a small-molecule tankyrase inhibitor shows some activity in APC-mutant CRC models⁴⁹⁸. It is not known whether these

therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

FREQUENCY & PROGNOSIS

In the Uterine Corpus Endometrial Carcinoma TCGA dataset, APC mutations are reported in 12% of cases16. Studies have reported APC mutations at varying frequencies in endometrial carcinoma, from no mutations being detected in 128 samples to 43% (12/28) of analyzed endometrial carcinomas having APC mutations⁴⁹⁹⁻⁵⁰⁰. APC methylation has been detected at a higher frequency in endometrial adenocarcinomas with microsatellite instability (MSI) (43%, 17/40), than in cases without MSI (16%, 12/74)⁵⁰⁰⁻⁵⁰¹. Two studies of endometrial carcinomas have reported the absence of APC protein expression in 8% (2/ 24) and 71% (30/42) of cases^{499,502}. A preclinical study using a mouse model demonstrated that loss of APC could induce endometrial cancer⁵⁰³. The prognostic significance of APC mutations in endometrial cancer remains unclear (PubMed, Jul 2021). Solid tumors with WNT/beta-catenin pathway alterations, as seen here, were observed to have significantly less T-cell inflammation in one study 504 .

FINDING SUMMARY

APC (adenomatous polyposis coli) encodes a tumor suppressor with critical roles in regulating cell division and adhesion. APC interacts with beta-catenin and controls signaling in the WNT pathway, which regulates embryonic development and cell differentiation⁵⁰⁵. Although alterations such as seen here have not been fully characterized and are of unknown functional significance, similar alterations have been previously reported in the context of cancer, which may indicate biological relevance.

POTENTIAL GERMLINE IMPLICATIONS

One or more of the APC variants observed here has been described in the ClinVar database as a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with familial adenomatous polyposis (ClinVar, Mar 2021)⁸⁷. Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. Germline mutations in APC are found in more than 90% of patients with familial adenomatous polyposis (FAP)⁵⁰⁶⁻⁵⁰⁸. The prevalence for FAP in the general population is estimated to be 1:8,300 from birth⁵⁰⁹, and in the appropriate clinical context germline testing of APC is recommended.



GENOMIC FINDINGS

GENE

ATRX

ALTERATION R907*

TRANSCRIPT ID

CODING SEQUENCE EFFECT 2719C>T

VARIANT ALLELE FREQUENCY (% VAF) 30.7%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

No targeted therapies are available to directly address ATRX inactivation. Based on preclinical⁵¹⁰⁻⁵¹¹ and limited clinical data⁵¹², ATRX alterations may confer sensitivity to combination strategies involving WEE1 inhibition. In a Phase 2 study evaluating the WEE1 inhibitor adavosertib plus irinotecan for the treatment of pediatric patients with neuroblastoma, prolonged SD was reported for 44% (4/9) of patients with ATRX-deficient tumors and responses were seen in two tumors that had evidence of ALT⁵¹². Preclinical evidence also suggests that ATRX deficiency may impart sensitivity to synthetic lethal approaches

involving PARP inhibition and irinotecan⁵¹³, combined PARP and ATR inhibition⁵¹¹, or double-strand break-induction with agents such as doxorubicin, irinotecan, and topotecan⁵¹⁴; however, these approaches have not been demonstrated clinically.

FREQUENCY & PROGNOSIS

Somatic mutation of ATRX has been reported in a number of solid tumor types, often associated with ALT515. ATRX mutation correlating with ALT has been reported in 10-20% of pancreatic neuroendocrine tumors (PNETs) $^{515-517}$, 12.6% of pheochromocytomas and paragangliomas⁵¹⁸, and 48% of adolescent and young adult (AYA) patients with glioblastoma (GBM) or neuroblastoma⁵¹⁹⁻⁵²³. ATRX loss in PNET516,524 and melanoma525 and mutation in other neuroendocrine tumors 518 is associated with poor prognosis. Pediatric patients with high-grade glioma and ATRX mutation were shown to have more aggressive disease but are more responsive to treatment with double-strand break therapy⁵¹⁴. ATRX mutation or loss of expression is more frequent in Grade 2/3 astrocytoma and secondary GBM than primary GBM, oligodendroglioma, and oligoastrocytoma526-529 and has been proposed as a distinguishing biomarker⁵²⁷⁻⁵²⁹. ATRX mutation has not been detected in concurrence with MYCN

amplification in glioma and neuroblastoma⁵²⁰⁻⁵²³. Low-grade gliomas with both IDH1/2 mutation and ATRX mutation are associated with worse prognosis than those with IDH1/2 mutation but no ATRX mutation⁵²⁷. Loss of ATRX protein expression has been reported in 33-39% of incidences of leiomyosarcoma (LMS) associating with ALT, a poor prognostic factor across all LMS subtypes, and with poor prognosis in extrauterine LMS but not in uterine LMS⁵³⁰⁻⁵³¹.

FINDING SUMMARY

ATRX encodes a SWI/SNF chromatin remodeling protein implicated in histone variant H₃·3 deposition, transcriptional regulation, and telomere maintenance⁵³²⁻⁵³³. ATRX inactivation or loss of expression is associated with alternative lengthening of telomeres (ALT)^{515,531,534-535}. Alterations that disrupt the ADD domain (aa 167-270) or helicase domain (aa 2010-2280) of ATRX are predicted to result in loss of ATRX function is not sufficient to induce ALT, which requires other undetermined factors^{532,539}. Germline mutations in ATRX give rise to alpha-thalassemia X-linked intellectual disability syndrome (ATR-X syndrome)⁵⁴⁰.



GENOMIC FINDINGS

GENE

CARD11

ALTERATION R271W

TRANSCRIPT ID NM_032415

CODING SEQUENCE EFFECT

811C>T

VARIANT ALLELE FREQUENCY (% VAF)

2.3%

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies —

Although there are no therapies targeting CARD11 alterations, a number of drugs that block NF-kB activation are under investigation⁵⁴¹⁻⁵⁴³. Preclinical evidence from models of mantle cell lymphoma suggests that tumors with CARD11 mutations may exhibit reduced sensitivity to inhibitors of BCR/NF-kB signaling including

ibrutinib and lenalidomide⁵⁴⁴⁻⁵⁴⁵. CARD11 mutations have also been correlated with reduced sensitivity to sotrastaurin (protein kinase C inhibitor) in preclinical models of DLBCL⁵⁴⁶.

FREQUENCY & PROGNOSIS

CARD11 alterations have been most frequently reported in B-cell lymphomas, including diffuse large B-cell lymphoma (DLBCL; 11-23%)⁵⁴⁷⁻⁵⁴⁹, follicular lymphoma (21-25%)550, primary central nervous system lymphoma (16-30%)551-552, splenic marginal zone lymphoma (7-9%)553-554, and mantle cell lymphoma (6%)544. CARD11 mutations have also been detected in T-cell lymphomas, specifically in angioimmunoblastic T-cell lymphomas (AITLs), peripheral T-cell lymphomas (PTCLs), and cases of Sezary syndrome⁵⁵⁵⁻⁵⁵⁷. Copy number gains and consequent CARD11 overexpression have been reported in 71% (12/17) of patients with aggressive AITL and in 41% (30/ 73) of patients with PTCL558-559. Increased CARD11 expression has also been observed in T-ALL560. CARD11 amplification in DLBCL and

increased CARD11 protein expression in AITL or PTCL is associated with reduced overall survival^{547,559}. Although CARD11 mutations have been detected in a variety of solid tumors⁵⁶¹⁻⁵⁶⁶, the prognostic implications of CARD11 alterations in non-hematological malignancies are unclear.

FINDING SUMMARY

CARD11 (also known as CARMA1) is a scaffold protein critical for B- and T-cell receptor-mediated NF-kappaB (NF-kB) activation⁵⁶⁷⁻⁵⁶⁹. Activating CARD11 mutations, often occurring within the CARD (aa 18-110) or coiled-coil (aa 130-499) domains, can facilitate constitutive NF-kB signaling, proliferation, and cell survival in lymphomas^{548,555,557,569-576}. Preclinical evidence suggests that activating CARD11 mutations may reduce sensitivity to ibrutinib, lenalidomide, and sotrastaurin in lymphoma⁵⁴⁴⁻⁵⁴⁶. Some germline CARD11 mutations have been found to underlie B-cell expansion with NF-kB and T-cell anergy (BENTA), a disorder characterized by congenital B-cell lymphocytosis^{569,572,577}.

GENE

CREBBP

ALTERATION

E50*

TRANSCRIPT ID NM_004380

CODING SEQUENCE EFFECT

1/19/5\7

VARIANT ALLELE FREQUENCY (% VAF)

38.4%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no targeted therapies available to address genomic alterations in CREBBP. The use of histone deacetylase (HDAC) inhibitors are being investigated in clinical trials that are recruiting patients with either lymphoma or urothelial carcinoma harboring CREBBP alterations. However, it has been reported that there is no correlation between CREBBP mutation status and

response to HDAC inhibitors in DLBCL578.

FREQUENCY & PROGNOSIS

CREBBP mutations have been observed at high frequency in follicular lymphoma (FL, 39%) and diffuse large B-cell lymphoma (DLBCL, 17%), and at lower frequency in acute lymphoblastic leukemia (ALL, 6%), and tumors of the urinary tract (15%), skin (11%), endometrium (9%), liver (9%), and stomach (9%) (COSMIC, 2021)⁷⁷. These mutations include missense substitutions clustered in the CREBBP histone acetyltransferase domain and truncating mutations throughout the gene sequence, suggesting a role for CREBBP inactivation in these diseases. CREBBP mutations have been reported to occur in the transition from prostate acinar carcinoma to squamous cell carcinoma (SCC)579, which may indicate significance for CREBBP in SCC. In two cases of relapsed pediatric B-cell ALL, CREBBP mutation conferred resistance to glucocorticoid therapy⁵⁸⁰. Reports have found CREBBP mutation in 62-68% of patients with FL581-582, which was associated with immune evasion⁵⁸¹. AML with MYST₃/

CREBBP fusion was reported to occur in 60-80% of cases 9-72 months after adjuvant chemotherapy for breast cancer and was associated with a poor prognosis⁵⁸³⁻⁵⁸⁴.

FINDING SUMMARY

CREBBP encodes a ubiquitously expressed transcriptional coregulatory protein that interacts with multiple transcription factors and can couple control of gene expression to chromatin remodeling via its histone acetyltransferase activity. Inherited microdeletions and truncating point mutations in CREBBP are reported to be causal in approximately 20% of cases of Rubinstein-Taybi syndrome⁵⁸⁵. The chromosomal rearrangement t(8;16)(p11;p13) is characteristic of the M4/M5 subtype of acute myeloid leukemia (AML) and results in a chimeric gene fusing MYST₃/MOZ (a gene essential for the development of the hematopoietic system and maintenance of hematopoietic stem cells) to CREBBP586. CREBBP-BCORL1 fusion has been reported in patients with ossifying fibromyxoid tumors⁵⁸⁷⁻⁵⁸⁸.



GENOMIC FINDINGS

GENE

CTCF

ALTERATION R129*

TRANSCRIPT ID NM_001191022

CODING SEQUENCE EFFECT

385C>T

VARIANT ALLELE FREQUENCY (% VAF)

36.3%

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies —

There are no targeted therapies available to address genomic alterations in CTCF.

FREQUENCY & PROGNOSIS

Somatic mutations in CTCF are infrequently reported in most cancers, but have been observed more commonly (24%) in uterine corpus endometrial carcinoma (cBioPortal, 2021)⁷⁵⁻⁷⁶; nearly half of the observed mutations were truncating, suggesting a tumor suppressor role for CTCF in this disease. In addition, CTCF has been found to act as a tumor suppressor in breast cancer cell line studies⁵⁸⁹⁻⁵⁹⁰.

FINDING SUMMARY

CTCF encodes an 11-zinc-finger protein that is implicated in a number of regulatory roles, including gene activation and repression, imprinting, insulation, methylation, and X chromosome inactivation⁵⁹¹. CTCF plays a role in transcriptional regulation of a number of key cancer-associated genes, including the oncogene MYC⁵⁹² and tumor suppressor TP53⁵⁹³, via maintenance of local DNA methylation status. The decreased expression levels of CTCF and/or BORIS, another 11-zinc-finger transcriptional regulator, were reported to be closely associated with global DNA methylation variability and decreased overall survival in epithelial ovarian cancer⁵⁹⁴⁻⁵⁹⁵.

GENE

CTNNA1

ALTERATION

R451*

TRANSCRIPT ID

NM_001903

CODING SEQUENCE EFFECT

1351C>T

VARIANT ALLELE FREQUENCY (% VAF)

32.4%

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies —

There are no available targeted therapies to address genomic alterations in CTNNA1.

FREQUENCY & PROGNOSIS

CTNNA1 mutations have been observed with

highest incidence in endometrial carcinoma (6.8%)⁵⁹⁶, cutaneous melanoma (6.4%)⁵⁹⁷, colorectal adenocarcinoma (4.4%)596, and stomach adenocarcinoma (4.0%) TCGA datasets (cBioPortal, 2021)⁷⁵⁻⁷⁶. CTNNA1 mutations have been observed in patients with hereditary diffuse gastric carcinoma without CDH1 mutations⁵⁹⁸⁻⁵⁹⁹. Reduced CTNNA1 expression in patients with breast cancer has been correlated with a poor clinical outcome and breast cancer brain metastasis⁶⁰⁰⁻⁶⁰¹. Deletion and hypermethylation of CTNNA1 has been observed in up to 22% (18/ 83) of myelodysplastic syndrome (MDS) cases and associated with poor clinicopathological features⁶⁰²⁻⁶⁰⁴ and a trend for inferior survival⁶⁰². Loss of CTNNA1 expression via 5q deletion or hypermethylation has been reported as a frequent event in acute myeloid leukemia and associated with shorter relapse-free survival in one studv604-606.

FINDING SUMMARY

CTNNA1 encodes alpha-catenin, a member of the cadherin family that functions in cell adhesion. Alpha-catenin acts as a tumor suppressor, through mechanisms that can vary by tissue⁶⁰⁷⁻⁶⁰⁸. Alphacatenin is one of three catenin proteins that are in complex with E-cadherin to help mediate cell-cell adhesion in epithelial tumor suppression⁶⁰⁷⁻⁶⁰⁸; loss of cell adhesion may contribute to cancer cell invasiveness and formation of metastases. In epidermal cells, alpha-catenin acts as a tumor suppressor by inducing YAP1 phosphorylation and cytoplasmic localization^{601,609}. Alpha-catenin also acts as a tumor suppressor by interacting with IKBalpha to influence the NF-KB pathway in Ecadherin-negative basal-like breast cancer cells⁶⁰¹. Loss of alpha-catenin expression is also hypothesized to alter the balance between the cytoplasmic (cell adhesion) and nuclear (cell proliferation) functions of beta-catenin, further contributing to oncogenesis⁶¹⁰.



GENOMIC FINDINGS

GENE

CUL3

ALTERATION

TRANSCRIPT ID

CODING SEQUENCE EFFECT

1805T>G

VARIANT ALLELE FREQUENCY (% VAF)

34.8%

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies —

There are no therapies available to directly address genomic alterations in CUL₃.

FREQUENCY & PROGNOSIS

Mutations affecting CUL₃-KEAP₁ complex formation and regulation of the stress-regulated transcription factor NRF₂ have been found in 4/5 cases of sporadic papillary renal cell carcinoma⁶¹¹. Mutations affecting the CUL₃-KEAP₁ complex have also been reported in lung cancer, and linked

to increased NF-kB signaling through upregulation of IkkB protein expression⁶¹². Decreased CUL₃ expression has been linked to an aggressive phenotype in bladder cancer models⁶¹³ and to formation of hepatocellular carcinomas through regulation of cyclin E expression⁶¹⁴.

FINDING SUMMARY

CUL3 encodes the RING ubiquitin ligase cullin 3, which has been shown to form complexes regulating diverse cellular processes, including development and stress responses.

GENE

ESR1

ALTERATION R256Q

TRANSCRIPT ID

NM_000125

CODING SEQUENCE EFFECT

767G>A

VARIANT ALLELE FREQUENCY (% VAF)

36.6%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

Therapies that directly target ER-alpha, such as selective ER modulators (SERMs) and the selective ER degrader (SERD) fulvestrant, as well as aromatase inhibitors (AIs) that inhibit estrogen production, are approved to treat ER-positive (ER+) and/or hormone receptor-positive (HR+) breast cancer (NCCN Guidelines v6.2020). AI treatment has also been reported to provide clinical benefit in a subset of HR+ gynecologic malignancies 615-619. Combinations of fulvestrant

and CDK₄/6 inhibitors such as abemaciclib, palbociclib, and ribociclib, have also demonstrated efficacy for patients with ESR1-mutated breast cancer⁶²⁰. Clinical data suggest that ESR1 mutations may confer sensitivity to the firstgeneration SERD fulvestrant in breast cancer⁶²¹⁻⁶²². A retrospective analysis of ESR1 mutations in gynecologic malignancies reported clinical benefit for patients with ESR1 mutations and fulvestrant treatment as a monotherapy or in combination, including 1 patient with peritoneal serous carcinoma and an ESR1 Y537N mutation who experienced prolonged clinical benefit (48+ months) from fulvestrant monotherapy⁶²³. It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

- Potential Resistance -

Extensive clinical evidence indicates that ESR1 mutations arise primarily during treatment with AIs, especially relative to other ER-targeting therapies, and ESR1 ligand-independent activating alterations may predict lack of benefit from subsequent AI treatment in breast carcinoma^{621,624-625}. In gynecologic malignancies,

activating ESR1 Y537S and D538G mutations have been reported to confer resistance to AI treatment, particularly for patients who have already received AI treatment^{623,626}.

FREQUENCY & PROGNOSIS

ESR1 mutation has been reported in 2.5-7.1% of endometrial carcinomas analyzed (COSMIC, cBioPortal, Nov 2020)⁷⁵⁻⁷⁷. Lower levels of ERalpha protein and/or ESR1 mRNA expression have been associated with a reduced survival in endometrial carcinoma⁶²⁷⁻⁶²⁸.

FINDING SUMMARY

ESR1 encodes estrogen receptor alpha (ER-alpha), one of the major estrogen receptor isoforms in humans. Along with co-activator proteins, the ER complex promotes transcription of genes involved in cell cycle progression and survival⁶²⁹. Although alterations such as seen here have not been fully characterized and are of unknown functional significance, similar alterations have been previously reported in the context of cancer, which may indicate biological relevance.



GENOMIC FINDINGS

GENE

FAM123B

ALTERATION F387*

TRANSCRIPT ID

CODING SEQUENCE EFFECT

1159G>T

VARIANT ALLELE FREQUENCY (% VAF)

35.5%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no targeted therapies available to address genomic alterations in FAM123B.

FREQUENCY & PROGNOSIS

Somatic mutation of FAM123B is rare in most cancers (COSMIC, 2021)⁷⁷, but is observed at rates ranging from 5-30% in Wilms tumor⁶³⁰⁻⁶³². No association between FAM123B alteration and clinical features or outcomes of Wilms tumor has

been documented.

FINDING SUMMARY

FAM123B, also known as AMER1, encodes the protein WTX, which binds to beta-catenin, enhancing its proteasomal degradation and thereby exerting a repressive effect on WNT pathway signaling⁶³³. Germline mutation or deletion of FAM123B causes osteopathia striata with cranial sclerosis⁶³⁴⁻⁶³⁵.

GENE

KEL

ALTERATION

E494*
TRANSCRIPT ID

NM 000420

CODING SEQUENCE EFFECT

1480G>T

VARIANT ALLELE FREQUENCY (% VAF)

32.6%

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies —

There are no therapies available to target genomic alterations in KEL.

FREQUENCY & PROGNOSIS

KEL mutations have been reported in tumors of the skin, lung, endometrium, stomach, large intestine, soft tissue, and liver at rates of 1.9-8.4%; up to 1.2% of acute lymphoblastic leukemia (ALL), acute myeloid leukemia (AML), and chronic

lymphocytic leukemia-small lymphocytic lymphoma (CLL/SLL) samples (COSMIC, 2021)⁷⁷. However, the mechanism by which KEL mutations contribute to tumor formation is not known.

FINDING SUMMARY

KEL encodes a transmembrane glycoprotein with similarities to zinc-dependent metalloproteases; this glycoprotein is highly polymorphic and forms the Kell blood group antigen⁶³⁶.

GENE

KLHL6

ALTERATION

D104N

TRANSCRIPT ID

NM_130446

CODING SEQUENCE EFFECT

310G>A

VARIANT ALLELE FREQUENCY (% VAF)

1.2%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

No targeted therapies are available to address genomic alterations in KLHL6.

FREQUENCY & PROGNOSIS

KLHL6 lies in a segment of chromosome 3q that has been observed to exhibit significant amplification in both squamous cell lung⁶³⁷ and ovarian⁶³⁸ carcinoma.

FINDING SUMMARY

KLHL6 encodes a protein that exhibits homology to the Drosophila Kelch gene and is hypothesized to play a role in germinal center B-cell differentiation⁶³⁹. Whole genome sequencing identified somatic alteration of KLHL6 in a small number of chronic lymphocytic leukemias⁶⁴⁰, whereas an exome-sequencing effort identified similar alterations in a follicular lymphoma case⁶⁴¹.



GENOMIC FINDINGS

GENE

MAP3K1

ALTERATION G914*

TRANSCRIPT ID

NM_005921

CODING SEQUENCE EFFECT

2740G>T

VARIANT ALLELE FREQUENCY (% VAF)

33.3%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no targeted therapies available to address genomic alterations in MAP₃K₁.

FREQUENCY & PROGNOSIS

Somatic alterations of MAP₃K₁, including missense mutations, truncating alterations, and loss of heterozygosity, have been observed most frequently in breast and uterine carcinomas (cBioPortal, COSMIC, 2021)⁷⁵⁻⁷⁷.

FINDING SUMMARY

MAP₃K₁ encodes a multifunctional protein kinase and E₃ ubiquitin ligase involved in several signal transduction pathways central to cancer cell biology⁶⁴². Different MAP₃K₁ protein isoforms have been suggested to exert both pro- and antiapoptotic influences⁶⁴³⁻⁶⁴⁴. Germline polymorphism in MAP₃K₁ has been hypothesized to associate with risk for at least some subtypes of breast carcinoma⁶⁴⁵, but the extent of effect is small and experimental results have been inconsistently replicated⁶⁴⁶.

GENE

MLL2

ALTERATION R5214C

TRANSCRIPT ID

NM_003482

CODING SEQUENCE EFFECT

15640C>T

VARIANT ALLELE FREQUENCY (% VAF)

1.2%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no targeted therapies available to address genomic alterations in MLL2.

FREQUENCY & PROGNOSIS

MLL2 alterations are observed in a number of solid tumor contexts (COSMIC, 2021)⁷⁷, and are especially prevalent in lung squamous cell carcinoma (SCC)⁶³⁷ and small cell lung carcinoma (SCLC)⁶⁴⁷. MLL2 mutation was found to be an independent prognostic factor of poor PFS and OS in non-small cell lung cancer, but not in SCLC⁶⁴⁸. One study reported that MLL2 truncating mutations were more common in recurrent ovary granulosa cell tumors (GCT) compared with primary GCTs (24% [10/42] vs. 3.0% [1/32])⁶⁴⁹. In a study of esophageal SCC, high MLL2 expression positively correlated with tumor stage, differentiation, and size, and negatively correlated with OS⁶⁵⁰.

FINDING SUMMARY

MLL2 encodes an H₃K₄-specific histone methyltransferase that is involved in the transcriptional response to progesterone signaling⁶⁵¹. Germline de novo mutations of MLL2 are responsible for the majority of cases of Kabuki syndrome, a complex and phenotypically distinctive developmental disorder⁶⁵². A significant number of inactivating MLL2 alterations have been observed in multiple tumor types, suggesting a tumor suppressor role⁶⁵³.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion⁹⁰⁻⁹⁵. Comprehensive genomic profiling of solid tumors may detect nontumor alterations that are due to CH^{94,96-97}. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.



GENOMIC FINDINGS

GENE

MSH₆

ALTERATION F641*

TRANSCRIPT ID

CODING SEQUENCE EFFECT

1921G>T

VARIANT ALLELE FREQUENCY (% VAF)

34.2%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

Numerous studies in various cancer types have shown that MSH6 loss or inactivation is associated with MSI and increased mutation burden^{28,47,654-657}. Clinical studies have shown that MSI is associated with patient responses to anti-programmed death 1 (PD-1) immune checkpoint inhibitors pembrolizumab^{33,658} and nivolumab⁶⁵⁹. Higher mutation burden was also reported to be associated with response to pembrolizumab²³. Furthermore, MSI status correlates with higher PD-1 and PD-L1 expression³⁰, potential biomarkers of response to PD-1 targeted

immunotherapies. Therefore, inactivation of MSH6 may confer sensitivity to anti-PD-1 immune checkpoint inhibitors.

FREQUENCY & PROGNOSIS

MSH6 mutations have been reported in 17% of endometrioid carcinomas¹⁶. Multiple studies have cited an increased risk (16-44%) of endometrial cancer for female carriers of germline MSH6 mutations⁶⁶⁰⁻⁶⁶². Endometrial tumors harboring MMR protein alterations have been associated with worse overall and progression-free survival⁶⁶³.

FINDING SUMMARY

MSH6 encodes MutS homolog 6 protein, a member of the mismatch repair (MMR) gene family. Defective MMR occurring as a result of mutation(s) in the MMR family (MLH1, MSH2, MSH6, or PMS2) can result in microsatellite instability (MSI), common in colon, endometrium, and stomach cancers⁴⁷. Alterations such as seen here may disrupt MSH6 function or expression⁶⁶⁴⁻⁶⁶⁹.

POTENTIAL GERMLINE IMPLICATIONS

One or more of the MSH6 variants observed here

has been described in the ClinVar database as a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with hereditary cancer-predisposing syndrome (ClinVar, Mar 2021)87. Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. Germline mutations in MSH6 are associated with both "typical" and "atypical" forms of autosomal dominant Lynch syndrome (also known as hereditary nonpolyposis colorectal cancer or HNPCC), which accounts for 1-7% of all colorectal cancers⁶⁷⁰. Approximately 10% of all Lynch syndrome-associated mutations have been attributed to alterations in MSH6671. Carriers of mutations in MSH6 have a 60-80% risk of colorectal cancer⁶⁷². Lynch syndrome has an estimated prevalence in the general population ranging from 1:600 to 1:2000^{670,673-674}. Biallelic germline mutation of MSH6 has been shown to account for 20% of cases of the very rare syndrome Constitutional Mismatch Repair Deficiency (CMMRD), which is characterized by a 95% incidence rate of childhood onset lymphoma, leukemia and brain tumors, followed by earlyonset colorectal cancer⁶⁷⁵⁻⁶⁷⁹. Given the association between MSH6 and these inherited syndromes, in the appropriate clinical context, germline testing of MSH6 is recommended.



GENOMIC FINDINGS

GENE

POLE

ALTERATION V411I

TRANSCRIPT ID NM_006231

CODING SEQUENCE EFFECT

1231G>C

VARIANT ALLELE FREQUENCY (% VAF) 34.7%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no targeted therapies that directly address POLE mutations. However, increased mutation load, such as may occur in "ultramutated" cancers harboring deleterious mutations in POLE, has been reported to be associated with response to the anti-programmed death 1 (PD-1) immune checkpoint inhibitors pembrolizumab15,23 and nivolumab680-681. In particular, a patient with non-small cell lung cancer harboring a deleterious POLE mutation achieved durable clinical benefit on

pembrolizumab²³; two patients with POLEmutated endometrial cancer responded to pembrolizumab¹⁵ or nivolumab⁶⁸²; two patients with POLE-mutated, TMB-high, MSS colorectal cancer responded to pembrolizumab⁶⁸³; and two patients with biallelic mismatch repair deficiency (bMMRD)-associated glioblastoma harboring POLE mutations experienced clinically and radiologically significant responses to nivolumab680. Furthermore, POLE-mutated endometrial cancers have been shown to have higher predicted neoantigen load, increased numbers of tumor-infiltrated lymphocytes (TILs), and higher expression of PD-1 and PD-L1 in the TILs⁶⁸⁴, which are potential biomarkers of response to anti-PD-1 immunotherapies.

FREQUENCY & PROGNOSIS

POLE alterations have been reported in 6-15% of endometrial carcinomas (COSMIC, Feb 2021)18,77,685-686. In the context of endometrial carcinoma, POLE mutations are associated with high tumor grade¹⁸ and correlate with better prognosis, with the most favorable prognosis seen for high-grade tumors (NCCN Uterine Neoplasms Guidelines, v2.2021)16,685-689.

FINDING SUMMARY

POLE encodes the catalytic subunit A of DNA polymerase epsilon, which plays roles in DNA replication and repair⁶⁹⁰. Deleterious mutations in POLE, mainly located within the exonuclease domain (amino acids 268-471) and reported at hotspot residues F104, D275, P286, S297, N363, D368, V411, L424, P436, R446, A456, Y458, S459, and S461, are predicted to disrupt the proofreading function of the enzyme, resulting in a high mutation rate and contributing to the development of "ultramutated," microsatellitestable cancers16,18,26-28,680-681,691-698.

POTENTIAL GERMLINE IMPLICATIONS

Germline mutations in POLE underlie polymerase proofreading-associated polyposis (PPAP), a highly penetrant, autosomal-dominant disorder characterized by the development of adenomatous polyps and an increased risk of intestinal, colorectal, and endometrial cancers^{26-27,691-692,694,699}. PPAP underlies 0.1% to 0.4% of familial cancers⁷⁰⁰. The lifetime risk of colorectal cancer for a POLE mutation carrier is estimated to be 21% to 28%⁷⁰¹. In the appropriate clinical context, germline testing of POLE is recommended.

GENE

SETD2

ALTERATION

splice site 5278-1G>T

TRANSCRIPT ID

NM 014159

CODING SEQUENCE EFFECT

5278-1G>T

VARIANT ALLELE FREQUENCY (% VAF)

35.8%

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies —

There are no targeted therapies available to address genomic alterations in SETD2.

FREQUENCY & PROGNOSIS

Somatic inactivating alterations of SETD2 are documented to occur at low frequency in a number of solid tumors, most commonly in renal carcinoma⁷⁰². SETD2 has been associated with favorable prognosis in gastric cancer⁷⁰³. SETD2 has also been associated with poor prognosis in RCC and MDS⁷⁰⁴, while data in other tumor types is limited (PubMed, Jun 2021).

FINDING SUMMARY

SETD2 encodes a histone lysine-36 methyltransferase⁷⁰⁵ that preferentially interacts with the expanded N-terminal polyglutamine tracts present in mutant huntingtin, implicating it in the pathogenesis of Huntington disease⁷⁰⁶. SETD2 mRNA expression has been observed to be consistently reduced in breast tumors relative to adjacent non-tumor tissue, suggesting a potential tumor suppressor role⁷⁰⁷. Although alterations such as seen here have not been fully characterized and are of unknown functional significance, similar alterations have been previously reported in the context of cancer, which may indicate biological relevance.



GENOMIC FINDINGS

GENE

SPEN

ALTERATION A2727V

TRANSCRIPT ID

NM_015001

CODING SEQUENCE EFFECT

8180C>T

VARIANT ALLELE FREQUENCY (% VAF) 35.9%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no targeted therapies available to address

SPEN inactivating mutations. Although gammasecretase inhibitors are in clinical development to target NOTCH activation, it is not known if these therapies would be beneficial in the context of SPEN mutation.

FREQUENCY & PROGNOSIS

SPEN truncating mutations have been reported in adenoid cystic carcinoma (ACC) (21%)⁷⁰⁸ and splenic marginal zone lymphoma (SMZL) (5%)⁵⁵³; NOTCH pathway gene mutations were frequent in both ACC and SMZL and observed in approximately 30% of cases^{553,708}.

FINDING SUMMARY

SPEN (also known as MINT or SHARP) encodes a transcriptional regulator that interacts with

HDAC1 and the SMRT/NcoR corepressors⁷⁰⁹⁻⁷¹⁰. SPEN represses the transcriptional activity of the NOTCH signaling pathway⁷¹¹⁻⁷¹². Activation of NOTCH signaling results in binding of the transcription factor RBPJ to the NOTCH intracellular domain and consequent activation of the NOTCH transcriptional program⁷¹³. SPEN binding to RBPJ has been shown to repress NOTCH-mediated transcription⁷¹¹⁻⁷¹². SPEN alterations that result in loss of the RBPJ-interaction domain (aa 2804-2816)⁷¹¹⁻⁷¹² or the SPOC domain (aa 3498-3664)⁷¹⁰ are predicted to disrupt binding of SPEN to RBPJ or corepressors and are likely to be inactivating.

GENE

STAG2

ALTERATION splice site 463-1G>T

TRANSCRIPT ID NM_006603

CODING SEQUENCE EFFECT 463-1G>T

VARIANT ALLELE FREQUENCY (% VAF) 32.5%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no therapies that directly target STAG2. However, in preclinical studies, STAG2 inactivation by mutation or knockdown resulted in increased sensitivity to PARP inhibitors⁷¹⁴ or oxaliplatin⁷¹⁵.

FREQUENCY & PROGNOSIS

STAG2 mutations have been observed most

frequently in urothelial bladder carcinoma (16-35%)^{566,716-719}, Ewing sarcoma (13-22%)⁷²⁰⁻⁷²¹, upper urinary tract urothelial carcinoma (11%)⁷²², myeloid malignancies (6%)723-724, and glioblastoma (6%)⁷²⁵. STAG2 truncation mutations are associated with loss of protein expression^{716-717,719,721}. In patients with Ewing sarcoma, STAG2 and TP53 mutations often cooccur and are associated with decreased overall survival, although mutation of either STAG2 or TP53 alone was not demonstrated to affect survival⁷²⁰⁻⁷²¹. STAG2 mutation in patients with myelodysplastic syndrome is associated with decreased overall survival and has also been associated with increased response to treatment with azacitidine or decitabine in patients with myeloid malignancies⁷²³. The data on the prognostic significance of STAG2 mutation or loss of STAG2 protein expression in the context of urothelial bladder carcinoma are conflicting⁷¹⁶⁻⁷¹⁹. In patients with pancreatic ductal adenocarcinoma, loss of STAG2 staining was significantly associated with decreased overall survival, but was also associated with survival benefit from adjuvant chemotherapy⁷¹⁵. An

inactivating STAG2 mutation was identified in a patient with melanoma that acquired resistance to vemurafenib and preclinical evidence suggests that loss of STAG2 expression decreases the sensitivity of BRAF V600E-positive melanoma cells to vemurafenib, dabrafenib, and trametinib⁷²⁶.

FINDING SUMMARY

STAG2 encodes a subunit of the cohesin complex, which maintains sister chromatid cohesion. The cohesin complex includes four subunits: SMC1A, SMC3, RAD21, and either STAG1 or STAG2⁷²⁷. Cohesin is also involved in transcriptional regulation, DNA replication and DNA repair⁷²⁷. STAG2 mutations, which are mostly truncating, or loss of STAG2 protein expression have been reported in multiple cancer types⁷²⁷⁻⁷²⁸. STAG2 deletion has been shown to promote tumorigenesis in preclinical studies⁷¹⁵, and STAG2 inactivation has been proposed to promote tumorigenesis via a mechanism that involves increased aneuploidy^{716,718,725} or altered transcriptional regulation^{717,719,723-724}.



GENOMIC FINDINGS

GENE

TNFAIP3

ALTERATION R87*

TRANSCRIPT ID

CODING SEQUENCE EFFECT

259C>T

VARIANT ALLELE FREQUENCY (% VAF)

35.8%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no therapies that address the loss of TNFAIP3. A20 has multiple functions and is subject to a wide range of genomic lesions, thereby making it challenging to develop a unified therapeutic approach. Potential avenues targeting dysregulation of ubiquitination pathways include anti-CD20 therapies, such as rituximab, and proteasome inhibitors, such as bortezomib⁷²⁹.

RNAi-mediated downregulation of TNFAIP3 has been reported to sensitize multiple myeloma cells to bortezomib⁷³⁰.

FREQUENCY & PROGNOSIS

In the COSMIC dataset, TNFAIP3 mutations have been reported in 3.5% of prostate, 3.1% of endometrial, 2.8% of skin, 2.7% of gastric, and 2.4% of large intestine cancers (Jan 2021)77. Overexpression of TNFAIP3 has been associated with aggressive high-grade ER-/PR-negative breast tumors731, resistance to TNF-alpha and TRAIL-induced apoptosis in glioblastoma⁷³²⁻⁷³³ and to chemotherapy in acute lymphoblastic leukemia⁷³⁴, and poor prognosis in adrenocortical carcinoma735. Loss of heterozygosity in the genomic region including TNFAIP3 has been found in 16.8% (25/149) of colorectal adenocarcinomas, and significantly decreased TNFAIP3 mRNA expression has been observed in colorectal cancer (CRC) tumors compared with adjacent non-neoplastic mucosa⁷³⁶. Reduced A20 expression has been suggested as a marker of poor prognosis in CRC737.

FINDING SUMMARY

TNFAIP3 encodes tumor necrosis factor alphainduced protein 3, also known as A20, a regulator of NF-kB signaling and apoptosis738 that has both ubiquitin ligase and deubiquitinase activities⁷³⁹⁻⁷⁴⁰ and whose loss or inactivation may be tumorigenic⁷⁴¹. TNFAIP3 is frequently deleted or mutated in lymphoma, where it functions as a tumor suppressor741, but its expression and function are context dependent in solid tumors^{738,742-745}, leukemia^{734,746-747}, and multiple myeloma⁷⁴⁸⁻⁷⁴⁹. TNFAIP3 mutations that disrupt the A20p37 chain (amino acids 371-790), which mediates ubiquitin ligase activity and interaction with the cIAP1/TRAF2 complex 739,750 , are predicted to be inactivating. In T-cells, cleavage of A20 codon R439 by MALT1 has been shown to upregulate NFkB signaling; R439A has been shown to block MALT1-mediated NF-kB activation⁷⁵¹; however, the function of R439 mutations outside of the context of T-cell lymphoma has not been reported.



GENOMIC FINDINGS

GENE

TP53

ALTERATION F113V - subclonal, T211A

TRANSCRIPT ID

NM_000546, NM_000546

CODING SEQUENCE EFFECT 337T>G, 631A>G

VARIANT ALLELE FREQUENCY (% VAF)

0.77%, 36.0%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor adavosertib⁷⁵²⁻⁷⁵⁵, or p53 gene therapy and immunotherapeutics such as SGT-53⁷⁵⁶⁻⁷⁶⁰ and ALT-801⁷⁶¹. In a Phase 1 study, adavosertib in combination with gemcitabine, cisplatin, or carboplatin elicited PRs in 9.7% (17/ 176) and SDs in 53.4% (94/176) of patients with solid tumors; the response rate was 21.1% (4/19) in patients with TP53 mutations versus 12.1% (4/ 33) in patients who were TP53 wild-type762. A Phase 2 trial of adavosertib in combination with chemotherapy (gemcitabine, carboplatin, paclitaxel, or doxorubicin) reported a 31.9% (30/ 94, 3 CR) ORR and a 73.4% (69/94) DCR in patients with platinum-refractory TP53-mutated ovarian, Fallopian tube, or peritoneal cancer⁷⁶³. A smaller Phase 2 trial of adavosertib in combination with carboplatin achieved a 42.9% (9/21, 1 CR) ORR and a 76.2% (16/21) DCR in patients with platinum-refractory TP53-mutated ovarian cancer⁷⁶⁴. The combination of adavosertib with paclitaxel and carboplatin in patients with TP53-mutated ovarian cancer also significantly increased PFS compared with paclitaxel and carboplatin alone⁷⁶⁵. In the Phase 2 VIKTORY trial, patients with TP53-mutated metastatic and/ or recurrent gastric cancer experienced a 24.0% (6/25) ORR with adavosertib combined with paclitaxel766. A Phase 1 trial of neoadjuvant

adavosertib in combination with cisplatin and docetaxel for head and neck squamous cell carcinoma (HNSCC) elicited a 71.4% (5/7) response rate for patients with TP53 alterations⁷⁶⁷. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel in patients with solid tumors, 75.0% (9/12) of evaluable patients experienced clinical benefit, including 2 confirmed and 1 unconfirmed PRs and 2 instances of SD with significant tumor shrinkage⁷⁶⁰. Additionally, the combination of a CHK1 inhibitor and irinotecan reportedly reduced tumor growth and prolonged survival in a TP53-mutated, but not TP53-wildtype, breast cancer xenotransplant mouse model⁷⁶⁸. Missense mutations leading to TP₅₃ inactivation may also be sensitive to therapies that reactivate mutated p53 such as APR-246⁷⁶⁹⁻⁷⁷¹. In a Phase 1b trial for patients with p53-positive highgrade serous ovarian cancer, APR-246 combined with carboplatin and pegylated liposomal doxorubicin achieved a 52% (11/21) response rate and 100% DCR772. ATR inhibitor treatment of chronic lymphocytic leukemia (CLL) cells with biallelic inactivation of TP53 suppressed cell viability, promoted DNA damage, and attenuated xenograft growth in preclinical studies73,773; however, ATR inhibitors as monotherapy had little effect on these parameters in solid tumor models in other preclinical studies⁷⁷⁴⁻⁷⁷⁵. Therefore, it is unclear whether TP53 inactivation predicts sensitivity to ATR inhibition.

FREQUENCY & PROGNOSIS

TP53 mutations have been reported in 29% of endometrial carcinomas analyzed in COSMIC, including 60% of serous carcinomas, 36% of clear cell carcinomas, and 23% of endometrioid carcinomas (Feb 2021)⁷⁷. In one large study, high (pathologic) expression of p53 was found in 24% of endometrial carcinoma samples and was associated with non-endometrioid histology, high grade (Grade 3 vs. Grade 1-2), and advanced FIGO stage, as well as with lymph node metastasis and poor disease-specific survival, but was not an independent factor for poor prognosis in multivariate analysis⁷⁷⁶. In other studies, p53 alterations (defined as TP53 mutation or p53

nuclear expression) have been found to be associated with poor prognosis in patients with endometrial cancer⁷⁷⁷⁻⁷⁷⁸.

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers⁷⁷⁹. Alterations such as seen here may disrupt TP53 function or expression⁷⁸⁰⁻⁷⁸⁴.

POTENTIAL GERMLINE IMPLICATIONS

Germline mutations in TP53 are associated with the very rare autosomal dominant disorder Li-Fraumeni syndrome and the early onset of many cancers⁷⁸⁵⁻⁷⁸⁷, including sarcomas⁷⁸⁸⁻⁷⁸⁹. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000⁷⁹⁰ to 1:20,000⁷⁸⁹. For pathogenic TP53 mutations identified during tumor sequencing, the rate of germline mutations was 1% in the overall population and 6% in tumors arising before age 30⁷⁹¹. In the appropriate clinical context, germline testing of TP53 is recommended.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion90-95. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy⁹⁰⁻⁹¹. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease⁷⁹². Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH94,96-97. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.



GENOMIC FINDINGS

GENE

XPO1

ALTERATION R749O

TRANSCRIPT ID NM_003400

CODING SEQUENCE EFFECT

2246G>A

VARIANT ALLELE FREQUENCY (% VAF)
0.90%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

XPO1 activation may predict sensitivity to XPO1 inhibitors⁷⁹³, such as the selective inhibitors of nuclear export (SINEs)⁷⁹⁴ KPT-330⁷⁹⁵ and KPT-8602⁷⁹⁶⁻⁷⁹⁷, which are in clinical trials in solid tumors and hematological cancers. In a Phase 1 trial of KPT-330 for unselected advanced solid tumors, one of 157 evaluable patients achieved a complete response, six achieved partial responses

(4%), and 67 (43%) achieved stable disease (SD), which was durable for ≥4 months for 27 patients (17%)⁷⁹⁸. A patient with accelerated-phase chronic myelogenous leukemia (CML) refractory to available tyrosine kinase inhibitors demonstrated short-term anti-leukemic activity from KPT-330⁷⁹⁹. Inhibitors of XPO1 have shown efficacy in mouse models and cultured cells of CML, T-cell acute lymphoblastic leukemia, acute myeloid leukemia, mantle cell lymphoma, multiple myeloma, and CLL^{795,797,799-804}, as well as in GIST, a subset of sarcomas⁸⁰⁵, and ESCCs⁸⁰⁶.

FREQUENCY & PROGNOSIS

XPO1 mutations have been reported at highest frequency in chronic lymphocytic leukemia (2–15%) and endometrial tumors (3–5%) and at lower incidences in other tumor types (COSMIC, 2021)^{16,77,640,807-809}. One study reported XPO1 mutation at high frequency in cases of Hodgkin lymphoma (26%, 5/19) and primary mediastinal B-cell lymphoma (24%, 28/117)⁸¹⁰. In CLL, XPO1 correlates with unmutated IGHV status, presence of NOTCH1 mutations, increased white blood cell

count, and decreased progression-free survival after certain chemoimmunotherapy regimens^{640,807,809,811}. Increased levels of nuclear exportin-1 have been reported to correlate with clinical stage in human cholangiocarcinoma samples, and reduced exportin-1 expression suppressed tumor growth in a xenograft model of the disease⁸¹².

FINDING SUMMARY

XPO1 encodes exportin-1, also known as CRM1, a nuclear pore protein that controls nuclear export and localization of multiple cell cycle- and proliferation-associated proteins⁸¹³. Exportin-1 may also be required for efficient centrosome maintenance by BRCA1⁸¹⁴. XPO1 residue E571 is a mutational hotspot in chronic lymphocytic leukemia (CLL) and other tumor types^{640,807,809-811,815}, although mutations at this site have not been functionally characterized. XPO1 D624G has been reported in the context of esophageal squamous cell carcinoma (ESCC) and has been suggested to lead to gain of function⁸⁰⁶.

GENE

ZNF217

ALTERATION

D326N

TRANSCRIPT ID

CODING SEQUENCE EFFECT

976G>A

VARIANT ALLELE FREQUENCY (% VAF)

34.3%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no available targeted therapies to address genomic alterations in ZNF217. Expression of ZNF217 may predict relapse of estrogen receptor (ER)-positive breast cancer under hormone

therapy through its direct interaction with ERalpha⁸¹⁶⁻⁸¹⁷. ZNF217 overexpression has also been associated with resistance to paclitaxel⁸¹⁸ and doxorubicin⁸¹⁹ in breast cancer cell lines. ZNF217 has been suggested as a potential biomarker for treatment with the DNA synthesis inhibitor and AKT inhibitor triciribine in breast cancer based on preclinical findings in cultured cells and xenografts expressing high levels of ZNF217; triciribine treatment also restored sensitivity to doxorubicin in these cells⁸²⁰.

FREQUENCY & PROGNOSIS

Amplification and/or overexpression of ZNF217 has been reported in breast⁸²¹, ovarian⁸²²⁻⁸²³, gastric⁸²⁴⁻⁸²⁵, colon⁸²⁶, prostate⁸²⁷, esophageal⁸²⁸, and urothelial carcinomas⁸²⁹, glioblastoma⁸³⁰, and ovarian carcinosarcomas⁸³¹. Overexpression in these tumors has generally been linked with aggressive tumor behavior and poor clinical

prognosis. High levels of ZNF217 expression result in dysregulation of a broad range of genes that may contribute to tumorigenesis⁸³²⁻⁸³⁴, and increased expression or activation of ERBB3^{821,835}, FAK⁸²¹, Aurora kinase A⁸¹⁸, AKT⁸¹⁹, and TGF-beta/SMAD signaling⁸²¹ has been demonstrated in ZNF217-expressing tumors or cells.

FINDING SUMMARY

ZNF217 encodes a candidate oncogene that has likely roles in histone modification and transcriptional repression^{819,836}. ZNF217 amplification has been correlated with protein overexpression in breast carcinoma tumors and cell lines⁸³⁷. The role of ZNF217 in promoting tumorigenesis was established in preclinical studies demonstrating that expression of ZNF217 results in the immortalization of both human mammary epithelial cells and ovarian surface epithelial cells in culture⁸³⁸⁻⁸³⁹.



THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

Dostarlimab

Assay findings association

Tumor Mutational Burden 372 Muts/Mb

AREAS OF THERAPEUTIC USE

Dostarlimab is a monoclonal antibody that binds to the PD-1 receptor and blocks its interaction with PD-L1 and PD-L2, reducing inhibition of the antitumor response. It is FDA approved to treat patients with mismatch repair deficient recurrent or advanced endometrial cancer or solid tumors. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical data across solid tumors^{2-4,12,840}, TMB of ≥10 Muts/Mb (based on this assay or others) may predict sensitivity to immune checkpoint inhibitors targeting PD-1 or PD-L1. An association between higher TMB and improved OS, median PFS, and ORR has been observed in large pan-solid tumor studies for patients

treated with immune checkpoint inhibitors²⁻³.

SUPPORTING DATA

Single-agent dostarlimab has been approved to treat patients with mismatch repair deficient (dMMR) endometrial cancer based on results observed in the Phase 1 GARNET study. In the study, dostarlimab elicited an ORR of 42% (30/71; 9 CRs, 21 PRs) for patients with endometrial cancer; median PFS, median OS, and median duration of response (DOR) have not been reached⁸⁴¹. In a Phase 2 study of combination niraparib and dostarlimab for patients with endometrial cancer, patients experienced an ORR of 14% (3/22) and a DCR of 32%, while patients treated with niraparib experienced an ORR of 4.0% (1/23) and a DCR of 20%⁸⁴².

Pembrolizumab

Assay findings association

Tumor Mutational Burden 372 Muts/Mb

AREAS OF THERAPEUTIC USE

Pembrolizumab is a monoclonal antibody that binds to the PD-1 receptor and blocks its interaction with the ligands PD-L1 and PD-L2 to enhance antitumor immune responses. It is FDA approved for patients with tumor mutational burden-high (TMB-High; ≥10 Muts/Mb), microsatellite instability-high (MSI-High), or mismatch repair deficient (dMMR) solid tumors, or PD-L1-positive non-small cell lung cancer (NSCLC), head and neck squamous cell cancer (HNSCC), classical Hodgkin lymphoma, cervical cancer, gastric cancer, esophageal cancer, or gastroesophageal junction (GEJ) carcinoma. It is also approved in various treatment settings for patients with melanoma, NSCLC, small cell lung cancer, HNSCC, urothelial carcinoma, hepatocellular carcinoma, Merkel cell carcinoma, or cutaneous squamous cell carcinoma (CSCC). Combination treatments with pembrolizumab are approved for patients with NSCLC, renal cell carcinoma, endometrial carcinoma that is not MSI-High or dMMR, or triple-negative breast cancer (TNBC). Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical data across solid tumors^{2-4,12,840}, TMB of ≥10 Muts/Mb (based on this assay or others) may predict sensitivity to immune checkpoint inhibitors targeting PD-1 or PD-L1. An association between higher TMB and improved OS, median PFS, and ORR has been observed in large pan-solid tumor studies for patients treated with immune checkpoint inhibitors²⁻³.

SUPPORTING DATA

In the Phase 3 KEYNOTE-775 study, combination

pembrolizumab and lenvatinib treatment improved median PFS (mPFS, 7.2 vs. 3.8 months, HR=0.56) and median OS (mOS, 18.3 vs. 11.4 months, HR=0.62) for patients with advanced endometrial cancer and prior platinum chemotherapy, compared with physician's choice of chemotherapy; benefit was observed in both mismatch repair deficient (dMMR, mPFS HR=0.36, mOS HR=0.37) and proficient (pMMR, mPFS HR=0.60, mOS HR=0.68) $subgroups ^{843}. \ Similar \ results \ for \ pembrolizum ab \ with$ lenvatinib were reported in the Phase 1b/2 KEYNOTE-146 study, with a trend toward an improved ORR for patients who were dMMR compared with pMMR (64% vs. 37%) but a similar ORR for patients who were PD-L1 positive (CPS ≥1.0%) compared with PD-L1 negative (36% vs. 40%)844. Single-agent pembrolizumab elicited clinical benefit for 26% (3/23 PRs and 3/23 SDs) of patients with advanced PD-L1-positive (CPS ≥1.0%) endometrial cancer included in the Phase 1b KEYNOTE-028 study, with a mPFS of 1.8 months⁸⁴⁵. In the Phase 2 KEYNOTE 158 multi-solid tumor trial, treatment with the PD-1 inhibitor pembrolizumab led to improved ORR for patients with TMB of 10 Muts/Mb or higher compared those with TMB <10 Muts/Mb (28.3% [34/120] vs. 6.5% [41/635])¹¹. In the KEYNOTE 028/012 pan-solid tumor trials, a similar improvement in ORR was reported for patients with >103 non-synonymous mutations/exome (~ equivalency >8 Muts/Mb as measured by this assay) compared to those with <103 non-synonymous mutations/exome (30.6% [11/36] vs. 6.5% [5/77])4. A patient with PD-L1-positive, ultramutated endometrial adenocarcinoma harboring POLE mutation experienced a PR to pembrolizumab for more than 14 months¹⁵.



THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Atezolizumab

Assay findings association

Tumor Mutational Burden 372 Muts/Mb

AREAS OF THERAPEUTIC USE

Atezolizumab is a monoclonal antibody that binds to PDL1 and blocks its interaction with PD-1 to enhance antitumor immune responses. It is FDA approved to treat patients with non-small cell lung cancer (NSCLC) and urothelial carcinoma, depending on treatment setting. Atezolizumab is also approved in combination with other therapies to treat patients with non-squamous NSCLC lacking EGFR or ALK alterations, small cell lung cancer, triple-negative breast cancer, hepatocellular carcinoma, and BRAF V600-positive melanoma. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical data across solid tumors^{2-4,12,840}, TMB of ≥10 Muts/Mb (based on this assay or others) may predict sensitivity to immune checkpoint inhibitors targeting PD-1 or PD-L1. An association between higher TMB and improved OS, median PFS, and ORR has been observed in large pan-solid tumor studies for patients treated with immune checkpoint inhibitors²⁻³.

SUPPORTING DATA

In a Phase 1 study of single-agent atezolizumab, patients with advanced or recurrent uterine cancer achieved an ORR of 13.3% (2/15), a median PFS of 1.4 months, and a median OS of 9.6 months; both objective responses occurred in PD-L1-positive patients (\geq 5% of immune cells), and one of these patients was also MSI-High⁸⁴⁶. In a retrospective analysis of patients with 17 solid tumor types (comprised of 47% NSCLC, 40% urothelial carcinoma, and 13% encompassing 15 other solid tumors), TMB of 16 Muts/Mb or greater was reported to be

associated with an improved ORR to atezolizumab compared to chemotherapy (30% vs. 14%) 12 . Atezolizumab has been studied primarily for the treatment of non-small cell lung cancer (NSCLC)847-852 and urothelial carcinoma⁸⁵³⁻⁸⁵⁶ . A study of atezolizumab as monotherapy for patients with advanced solid tumors reported a median PFS of 18 weeks and an ORR of 21%, including confirmed responses in 25.6% (11/43) of melanomas, 12.5% (7/56) of renal cell carcinomas (RCC) and 16.7% (1/6) of colorectal cancers (CRCs)852. As singleagent therapy in genomically unselected young patients (<30 years old) with relapsed or refractory cancers, atezolizumab elicited an ORR of 1.5% (1/67) for patients with solid tumors, with similar safety and pharmacokinetics as seen in adults⁸⁵⁷. A Phase 1a study of atezolizumab reported an ORR of 14.5% (9/62), a median PFS of 5.6 months, and a median OS of 28.9 months for patients with clear cell RCC858. A Phase 1b study evaluated atezolizumab combined with nab-paclitaxel for patients with previously treated metastatic triple-negative breast cancer (mTNBC) and reported confirmed objective responses for 41.7% (10/24) of patients; no dose-limiting toxicities were observed⁸⁵⁹. A Phase 1b study that evaluated atezolizumab in combination with the MEK inhibitor cobimetinib for advanced solid tumors reported an ORR of 8.3% (7/84) in patients with CRC, 40.9% (9/22) in patients with melanoma, 17.9% (5/28) in patients with NSCLC, and 18.8% (3/16) in patients with other tumors (ovarian cancer, clear-cell sarcoma, and RCC); there was no association between BRAF or KRAS mutation status and response rate in any disease setting, and no doselimiting toxicities were encountered860-861.



THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Avelumab

Assay findings association

Tumor Mutational Burden 372 Muts/Mb

AREAS OF THERAPEUTIC USE

Avelumab is a monoclonal antibody that binds to PD-L1 and blocks its interaction with PD-1 in order to enhance antitumor immune responses. It is FDA approved to treat patients 12 years and older with Merkel cell carcinoma, or for urothelial carcinoma in various treatment settings. The combination of avelumab and axitinib is FDA approved for patients with renal cell carcinoma (RCC). Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical data across solid tumors^{2-4,12,840}, TMB of ≥10 Muts/Mb (based on this assay or others) may predict sensitivity to immune checkpoint inhibitors targeting PD-1 or PD-L1. An association between higher TMB and improved OS, median PFS, and ORR has been observed in large pan-solid tumor studies for patients treated with immune checkpoint inhibitors²⁻³.

SUPPORTING DATA

In a Phase 2 trial of avelumab in recurrent endometrial

cancer, patients with MMR deficiency or POLE mutation achieved an ORR of 26.7% (4/15) and a median PFS of 4.4 months; for patients with MMR-proficient tumors, the ORR was 6.2% (1/16) and the median PFS was 1.9 months 862 . The JAVELIN Phase 1b study has demonstrated clinical benefit from single-agent avelumab in a variety of solid tumor types, including non-small cell lung carcinoma (NSCLC)863, gastric carcinoma and gastroesophageal junction (GEJ) adenocarcinoma⁸⁶⁴, urothelial carcinoma⁸⁶⁵, mesothelioma⁸⁶⁶, ovarian carcinoma⁸⁶⁷, and breast cancer⁸⁶⁸, and from avelumab combined with axitinib in renal cell carcinoma869. Emerging clinical data show a positive trend toward the association of tumor cell PD-L1 expression and improved objective response rate, progression-free survival, or overall survival in NSCLC in the first-line setting and in ovarian and breast cancer^{863,867-868}. Limited clinical data indicate activity of avelumab in adrenocortical carcinoma, metastatic castration-resistant prostate cancer, and thymic cancer870-872.

Cemiplimab

Assay findings association

Tumor Mutational Burden 372 Muts/Mb

AREAS OF THERAPEUTIC USE

Cemiplimab is a monoclonal antibody that binds to the PD-1 receptor and blocks its interaction with the ligands PD-L1 and PD-L2 to enhance antitumor immune responses. It is FDA approved to treat patients with NSCLC with high PD-L1 expression (TPS \geq 50%), cutaneous squamous cell carcinoma (CSCC), or basal cell carcinoma (BCC). Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical data across solid tumors^{2-4,12,840}, TMB of ≥10 Muts/Mb (based on this assay or others) may predict sensitivity to immune checkpoint inhibitors targeting PD-1 or PD-L1. An association between higher TMB and improved OS, median PFS, and ORR has been observed in large pan-solid tumor studies for patients treated with immune checkpoint inhibitors²⁻³.

SUPPORTING DATA

Clinical data on the efficacy of cemiplimab for the treatment of endometrial cancer are limited (PubMed, May 2021). Cemiplimab has been studied primarily in advanced cutaneous squamous cell carcinoma (CSCC), where it elicited a combined ORR of 48% (41/85) in Phase 1 and 2 studies⁸⁷³. A Phase 2 trial of cemiplimab in patients with basal cell carcinoma (BCC) reported ORRs of 31% (5 CRs and 21 PRs) in patients with locally advanced BCC and 21% (6 PRs) in patients with metastatic BCC⁸⁷⁴⁻⁸⁷⁵. The Phase 3 EMPOWER-Lung 1 trial for advanced non-small cell lung cancer (NSCLC) with PD-L1 expression ≥50% reported that cemiplimab is associated with improved PFS (8.2 vs. 5.7 months), OS (not reached vs. 14.2 months), and ORR (37% vs. 21%) compared with chemotherapy⁸⁷⁶.



THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Durvalumab

Assay findings association

Tumor Mutational Burden 372 Muts/Mb

AREAS OF THERAPEUTIC USE

Durvalumab is a monoclonal antibody that binds to PD-L1 and blocks its interaction with PD-1 to enhance antitumor immune responses. It is FDA approved to treat patients with non-small cell lung cancer (NSCLC) and small cell lung cancer (SCLC). Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical data across solid tumors^{2-4,12,840}, TMB of ≥10 Muts/Mb (based on this assay or others) may predict sensitivity to immune checkpoint inhibitors targeting PD-1 or PD-L1. An association between higher TMB and improved OS, median PFS, and ORR has been observed in large pan-solid tumor studies for patients

treated with immune checkpoint inhibitors²⁻³.

SUPPORTING DATA

The PHAEDRA Phase 2 trial of durvalumab in advanced endometrial cancer reported an ORR of 43% and a DCR of 66% (5 CRs, 10 PRs, 8 SDs in 35 patients) for the MMR-deficient cohort, and an ORR of 3% and a DCR of 29% (1 PR, 9 SDs in 35 patients) for the MMR-proficient cohort using iRECIST criteria⁸⁷⁷. A Phase 2 study for patients with endometrial carcinoma and carcinosarcoma evaluated durvalumab with or without tremelimumab and reported ORR of 14.8% (4/27 with 1 CR and 3 PRs) versus 11.1% (3/27 with 2 CRs and 1 PR), median PFS of 7.6 weeks versus 8.1 weeks, and median duration of response of 16 weeks versus 8 weeks⁸⁷⁸.

Everolimus

Assay findings association

FBXW7 R441W

PIK3CA R88Q, T1025A

AREAS OF THERAPEUTIC USE

Everolimus is an orally available mTOR inhibitor that is FDA approved to treat renal cell carcinoma (RCC) following antiangiogenic therapy; pancreatic neuroendocrine tumors; and well-differentiated nonfunctional neuroendocrine tumors of the lung or gastrointestinal tract. Everolimus is also approved to treat either renal angiomyolipoma or subependymal giant cell astrocytoma in association with tuberous sclerosis complex (TSC). Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of several case reports describing clinical benefit for patients with FBXW7-mutated cancers treated with mTOR inhibitors, including lung adenocarcinoma¹⁵⁰, renal cell carcinoma⁵⁸, and cervical squamous cell carcinoma⁸⁷⁹, FBXW7 loss or inactivation may predict sensitivity to everolimus or temsirolimus. On the basis of clinical evidence^{178,210-216}, PIK₃CA activating mutations may predict sensitivity to mTOR inhibitors such as everolimus and temsirolimus. Studies have reported modest activity of these therapies as single agents (ORRs of o-4%), but improved activity has been observed when they are combined with other agents such as bevacizumab and doxorubicin (ORRs of 25-44%), for patients with PIK₃CA-mutated solid tumors^{213-216,880-884}.

SUPPORTING DATA

Clinical benefit has been reported for several patients with PIK₃CA-mutated endometrial cancer treated with everolimus as a single agent^{186,212,399} or combined with hormone therapy^{186,254}. In a Phase 2 clinical trial of

recurrent endometrial cancer, 43% (12/28) of patients reported SD at 8 weeks and 21% (6/28) of patients achieved clinical benefit at 20 weeks upon administration of everolimus monotherapy⁴⁰⁰. Combination with the aromatase inhibitor letrozole for the same disease population achieved an ORR of 31% (11/35), with 9 CRs²⁵⁴. Further addition of metformin to this regimen led to a clinical benefit rate (CR+PR+SD) of 67% (32/48), including PR in 29% (14/48) of cases; no significant difference was observed between cases with and without KRAS mutation⁷⁶⁵. Everolimus achieved PR or SD in 35% of patients with recurrent endometrial carcinoma; KRAS mutation was associated with reduced median PFS (3.1 vs. 1.0 months) and median OS (9.3 vs. 2.3 months)³⁹⁹. Another study investigating estrogen and/or progesterone receptor-positive gynecologic or breast malignancies featuring mutation or loss of genes in the PI₃K-AKT-mTOR pathway, including PIK₃CA, AKT₁, or PTEN, observed SD in 17% (1/6) of patients with endometrial cancer following combined treatment with everolimus and anastrozole⁸⁸⁵. No response was seen in a patient with endometrial stromal sarcoma and Peutz-Jeghers Syndrome associated with a germline STK11 mutation treated with a combination of everolimus and anastrozole 886 . Whereas frequent adverse events precluded a recommended Phase 2 dose and schedule for the combination of trametinib and everolimus in a Phase 1b trial for solid tumors¹⁸⁴, a retrospective study for heavily pretreated patients with solid tumors reported tolerable regimens of the combination for 23/31 patients, with 16 patients treated >3 months and evaluable patients achieving a median PFS of 6.5 months¹⁸⁵.



THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Niraparib

Assay findings association

ATM R250*

BRCA2 E1879D

AREAS OF THERAPEUTIC USE

The PARP inhibitor niraparib is FDA approved to treat patients with epithelial ovarian, fallopian tube, or primary peritoneal cancer, with or without homologous recombination deficiency (HRD)-positive status. Please see the drug label for full prescribing information.

GENE ASSOCIATION

ATM inactivation may predict sensitivity to PARP inhibitors. ATM alterations have been associated with clinical benefit from PARP inhibitors in solid tumors, including prostate cancer^{53-55,887}, colorectal cancer⁵⁷, breast cancer⁵⁷, gastric cancer⁵⁶, cholangiocarcinoma⁵⁹, and papillary renal cell carcinoma⁵⁸. On the basis of clinical evidence in ovarian and breast cancers^{101-102,888}, loss or inactivation of either BRCA1 or BRCA2 may confer sensitivity to PARP inhibitors such as niraparib.

SUPPORTING DATA

In a Phase 2 study of niraparib for patients with endometrial cancer, patients experienced an ORR of 4.0% (1/23) and a DCR of 20%, while patients treated with

niraparib and dostarlimab experienced an ORR of 14% (3/ 22) and a DCR of 32%842. Niraparib has been primarily evaluated in the context of ovarian cancer. In a Phase 3 study of patients with platinum-sensitive, recurrent ovarian cancer, niraparib significantly increased median PFS, as compared to placebo, in patients with germline BRCA mutations (21 vs. 5.5 months) and in patients without germline BRCA mutations (9.3 vs. 3.9 months) as well as in a subgroup of the patients without germline BRCA mutations with homologous recombinationdeficient tumors (12.9 vs. 3.8 months)101. In a Phase 1 study of niraparib treatment for patients with solid tumors, 40% (8/20) of patients with ovarian cancer and BRCA mutations and 50% (2/4) of patients with breast cancer and BRCA mutations experienced a PR, and 43% (9/21) of patients with castration-resistant prostate cancer and 100% (2/2) of patients with non-small cell lung cancer achieved SD102. A Phase 1 study of the combination of niraparib and bevacizumab in patients with platinum-sensitive, high-grade ovarian cancer reported a DCR of 91% (10/11), with a response rate of 45% (5/11)889.



THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Nivolumab

Assay findings association

Tumor Mutational Burden 372 Muts/Mb

AREAS OF THERAPEUTIC USE

Nivolumab is a monoclonal antibody that binds to the PD-1 receptor and blocks its interaction with PD-L1 and PD-L2, reducing inhibition of the antitumor immune response. It is FDA approved in various treatment settings for patients with melanoma, renal cell carcinoma (RCC), non-small cell lung cancer (NSCLC), head and neck squamous cell carcinoma (HNSCC), urothelial carcinoma, classical Hodgkin lymphoma (cHL), gastric cancer, gastroesophageal junction cancer, and esophageal adenocarcinoma or squamous cell carcinoma (ESCC). Furthermore, nivolumab is approved to treat patients with mismatch repair-deficient (dMMR) or microsatellite instability-high (MSI-H) metastatic colorectal cancer (CRC) that has progressed on fluoropyrimidine, oxaliplatin, and irinotecan. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical data across solid tumors^{2-4,12,840}, TMB of ≥10 Muts/Mb (based on this assay or others) may predict sensitivity to immune checkpoint inhibitors targeting PD-1 or PD-L1. An association between higher TMB and improved OS, median PFS, and ORR has been observed in large pan-solid tumor studies for patients treated with immune checkpoint inhibitors²⁻³.

SUPPORTING DATA

A Phase 2 study of cabozantinib combined with nivolumab reported improved median PFS (5.3 vs. 1.9 months) and ORR (25% [9/36] vs. 16.7% [3/18]) compared with nivolumab monotherapy for patients with endometrial cancer; OS was also improved with combination therapy (13.0 vs. 7.9 months), although the data have not matured yet⁸⁹⁰. A retrospective analysis of patients with recurrent and refractory endometrial cancer treated with nivolumab alone or in combination with other agents reported a CR of >20 months for a patient

treated with nivolumab and everolimus and an overall clinical benefit rate of 75% (9/12), including 8 SDs of >3 months⁸⁹¹. A case study reported 2 PRs to nivolumab for patients with endometrial carcinoma harboring high tumor mutation burden; response was ongoing at 7-9 months⁶⁸². Nivolumab monotherapy has been reported to elicit clinical benefit for patients with multiple types of solid tumors, including melanoma $(27-31\% \text{ ORR})^{892-896}$, non-small cell lung carcinoma (NSCLC; 17-20% ORR and 9-10 months mOS in unselected patients)897-903, urothelial carcinoma (20-26% ORR in unselected patients)904-905, renal cell carcinoma (RCC; 26% ORR)906-911, MSI-High colorectal cancer (CRC; 58% $ORR)^{912-914}$, head and neck squamous cell carcinoma (11-17% ORR)915-917, ovarian cancer (6-15% ORR)918-920, small cell lung cancer (SCLC; 10-12% ORR)866,921-922, gastroesophageal carcinoma (12-18% ORR)923-925, and cancer of unknown primary (CUP; 10% ORR in unselected, previously treated patients)888, as well as with Hodgkin lymphoma (66-87% ORR)926-928. Combination treatment with nivolumab plus the CTLA-4 inhibitor ipilimumab has achieved further efficacy in melanoma (up to 61% ORR; mOS > 60 months for the combination vs. 37 months for nivolumab monotherapy) $\!^{892,929\text{-}931}$, NSCLC (17 months mOS)932, MSI-High CRC (64% ORR)933, RCC (42% ORR)934-935, SCLC (19-25% ORR)866,922, urothelial carcinoma (38% ORR in unselected patients; 58% ORR in patients with ≥1% tumor PD-L1 expression)904, and other solid tumors. Combinations of nivolumab with various targeted therapies, such as pazopanib (1a PR in a patient with epithelioid sarcoma)936, sunitinib (9% ORR in unselected patients with sarcoma)937, cabozantinib (29% ORR in patients with immunotherapy-refractory urothelial carcinoma)938, or vemurafenib (1a durable PR in a patient with BRAF V600E-mutated and PD-L1 positive anaplastic thyroid cancer)939, have also shown evidence of efficacy and continue to undergo clinical investigation.



THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Nivolumab + Ipilimumab

Assay findings association

Tumor Mutational Burden 372 Muts/Mb

AREAS OF THERAPEUTIC USE

Nivolumab is a monoclonal antibody that binds to the PD-1 receptor and blocks its interaction with PD-L1 and PD-L2, reducing inhibition of the antitumor immune response, and ipilimumab is a cytotoxic T-lymphocyte antigen 4 (CTLA-4)-blocking antibody. The combination is FDA approved in various treatment settings for patients with melanoma, renal cell carcinoma (RCC), non-small cell lung cancer (NSCLC), hepatocellular carcinoma (HCC), and pleural mesothelioma. Furthermore, nivolumab is approved in combination with ipilimumab to treat patients with mismatch repair-deficient (dMMR) or microsatellite instability-high (MSI-H) metastatic colorectal cancer (CRC) that has progressed on fluoropyrimidine, oxaliplatin, and irinotecan. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical data across solid tumors $^{5-6,940}$, a TMB score of \geq 10 Muts/Mb (as measured by this assay) may predict sensitivity to combination nivolumab and ipilimumab treatment.

SUPPORTING DATA

A Phase 2 trial treating patients with rare gynecological malignancies with combination nivolumab and ipilimumab reported objective responses in 27% (11/41) of patients, including those with vaginal SCC, ovarian and uterine carcinosarcoma, uterine serous cancer, uterine clear cell carcinoma, and uterine leiomyosarcoma⁹⁴¹.



THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Olaparib

Assay findings association

ATM R250*

BRCA2 E1879D

AREAS OF THERAPEUTIC USE

The PARP inhibitor olaparib is FDA approved to treat patients with epithelial ovarian, Fallopian tube, or primary peritoneal cancer, patients with deleterious or suspected deleterious gBRCA-mutated pancreatic adenocarcinoma or HER2-negative breast cancer, and patients with prostate cancer and mutations in homologous recombination repair genes. Olaparib is also approved in combination with bevacizumab to treat patients with ovarian, Fallopian tube, or primary peritoneal cancer with deleterious or suspected deleterious somatic or gBRCA mutation and/or genomic instability. Please see the drug label for full prescribing information.

GENE ASSOCIATION

ATM inactivation may predict sensitivity to PARP inhibitors. ATM alterations have been associated with clinical benefit from PARP inhibitors in solid tumors, including prostate cancer⁵³-55,887, colorectal cancer⁵⁷, breast cancer⁵⁷, gastric cancer⁵⁶, cholangiocarcinoma⁵⁹, and papillary renal cell carcinoma⁵⁸. On the basis of extensive clinical evidence in ovarian cancer¹⁰⁷⁻¹¹¹ as well as strong clinical evidence in multiple other cancer types^{53,98-99,107,110,114,942}, loss or inactivation of either BRCA1 or BRCA2 may confer sensitivity to olaparib.

SUPPORTING DATA

A heavily pretreated patient with low-grade endometrioid endometrial adenocarcinoma and germline and somatic BRCA2 mutations experienced ongoing SD for 15 months from single-agent olaparib⁹⁴³. A patient with high-grade serous endometrial cancer harboring a somatic BRIP1 frameshift alteration experienced a CR following olaparib monotherapy⁹⁴⁴. A patient with endometrial adenocarcinoma and PTEN loss experienced clinical benefit for 8 months following olaparib therapy. Although BRCA alterations were not detected, only a subset of common BRCA mutations were assessed, and the authors

could not exclude the possibility of other BRCA alterations contributing to the response⁴⁰⁵. Olaparib has been studied primarily for the treatment of ovarian cancer, with response rates often significantly higher for patients with BRCA mutations than for those without 107,110; higher response rates have also been observed for patients with platinum-sensitive versus platinum-resistant cancer^{109-110,112,945}. As maintenance therapy for patients with newly diagnosed or platinumsensitive relapsed ovarian cancer, olaparib has demonstrated significantly improved median PFS and median OS compared with placebo in the Phase 3 SOLO-1 study 113 and in multiple later-phase studies $^{105\text{-}106,946\text{-}947}$. Phase 3 studies of olaparib for patients with BRCAmutated metastatic breast¹⁰⁰ or pancreatic cancer¹¹⁴ or for patients with metastatic castration-resistant prostate cancer and BRCA or ATM alterations948 have also reported significantly longer median PFS compared with chemotherapy, placebo, or hormone therapy. Additionally, olaparib has demonstrated clinical activity for patients with other solid tumors harboring BRCA mutations. including leiomyosarcoma $^{949}\!,$ cholangiocarcinoma $^{950}\!,$ and bladder cancer951 in smaller studies. Olaparib in combination with the AKT inhibitor capivasertib has demonstrated clinical benefit for patients with solid tumors; a Phase 1 trial reported a 45% (25/56) DCR, including 14 PRs and 11 SDs, and 14 of those experiencing clinical benefit had germline BRCA1/2 mutated-solid tumors⁹⁵². In a Phase 2 study of olaparib plus pembrolizumab for patients with advanced solid tumors, those with BRCA1 or BRCA2 mutations and those with homologous recombination deficient tumors reported the highest ORRs of 29% (6/21) and 21% (16/76), respectively. Patients with homologous recombination repair deficient tumors, including and excluding patients with BRCA1/2 mutations, reported lower ORRs of 15% (8/53) and 6.3% (2/32), respectively⁹⁵³.



THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Rucaparib

Assay findings association

ATM R250*

BRCA2 E1879D

AREAS OF THERAPEUTIC USE

The PARP inhibitor rucaparib is FDA approved to treat patients with metastatic castration-resistant prostate cancer (mCRPC) or epithelial ovarian, Fallopian tube, or primary peritoneal cancer and deleterious somatic or germline BRCA mutations. Rucaparib is also approved as a maintenance treatment of patients with recurrent epithelial ovarian, Fallopian tube, or primary peritoneal cancer. Please see the drug label for full prescribing information.

GENE ASSOCIATION

ATM inactivation may predict sensitivity to PARP inhibitors. ATM alterations have been associated with clinical benefit from PARP inhibitors in solid tumors, including prostate cancer^{53-55,887}, colorectal cancer⁵⁷, breast cancer⁵⁶, cholangiocarcinoma⁵⁹, and papillary renal cell carcinoma⁵⁸. On the basis of strong clinical evidence in ovarian cancer^{103-104,763}, as well as clinical data in other cancer types^{104,954-955}, loss or inactivation of either BRCA1 or BRCA2 may confer sensitivity to rucaparib.

SUPPORTING DATA

Clinical data on the efficacy of rucaparib for the treatment of endometrial cancers are limited (PubMed, May 2021). Rucaparib has primarily been evaluated in the context of ovarian carcinoma, breast carcinoma, pancreatic carcinoma, and melanoma. In a Phase 2 study of rucaparib for recurrent, platinum-sensitive ovarian, peritoneal, or fallopian tube carcinoma, median PFS was significantly longer in patients with BRCA1/2 mutations (12.8 months) or high loss of heterozygosity (LOH; 5.7 months) compared to patients with low LOH (5.2 months).

Objective responses were observed for 80% (32/40) of patients with BRCA1/2 mutations, for 29% (24/82) with high LOH, and for 10% (7/10) with low LOH103. In heavily pretreated patients with a germline BRCA1/2 mutation who had received 2-4 prior chemotherapy treatments and had a progression free interval of greater than 6 months, 65% (17/26) of patients achieved an objective response with rucaparib treatment⁷⁶³. In a Phase 2 study evaluating rucaparib for patients with advanced breast or ovarian cancer and germline BRCA1/2 mutations, disease control was observed in 92% (12/13) of patients with ovarian cancer treated with oral rucaparib dosed continuously, but no objective responses were reported in breast cancer patients (n=23). However, 39% (9/23) of evaluable patients with breast cancer achieved SD lasting 12 weeks or more¹⁰⁴. In a Phase 1 study of rucaparib treatment in patients with solid tumors, 3/4 patients with ovarian cancer and 1/1 patient with breast cancer given the recommended Phase 2 dose reported objective responses; all responders harbored BRCA1/2 mutations954. A Phase 2 study of rucaparib treatment for patients with relapsed pancreatic cancer reported 1/19 CR, 2/19 PR (one unconfirmed) and 4/19 SD. Of the 19 patients treated in the study, 15 (79%) had a BRCA2 mutation 955 . In a Phase 2 study of intravenous rucaparib in combination with temozolomide for patients with metastatic melanoma, 8/ 46 patients achieved a PR and 8/46 had SD956; a Phase 1 study reported 1 CR, 1 PR, and 4 SD lasting six months or longer in patients with metastatic melanoma $^{957}\!.$ A Phase 1 study of intravenous and oral rucaparib in combination with chemotherapy for the treatment of advanced solid tumors reported a disease control rate of 68.8% (53/77), including 1 CR and 9 PRs958.



THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Selumetinib

Assay findings association

NF1 R1362*, R2450*

NE1

AREAS OF THERAPEUTIC USE

Selumetinib is a MEK inhibitor that is FDA approved to treat pediatric patients 2 years of age and older with neurofibromatosis type 1 (NF1) who have symptomatic, inoperable plexiform neurofibromas (PNs). Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical evidence in neurofibromatosis type 1 (NF1)-associated neurofibroma^{169-172,959-963}, glioma^{172-176,964}, and non-small cell lung cancer¹⁷⁷, NF1 inactivation may predict sensitivity to MEK inhibitors.

SUPPORTING DATA

A Phase 2 study evaluating selumetinib in patients with recurrent endometrial cancer who had received prior treatment with 1-2 therapies did not reach its primary endpoints of efficacy and 6-month event-free survival, reporting mPFS of 2.3 months and mOS of 8.5 months with 1 CR and 2 PR observed in 52 patients⁹⁶⁵. Selumetinib has demonstrated efficacy in NF1-associated

neurofibroma in Phase 2 studies^{170,959-960} and a Phase 1 study¹⁶⁹. Phase 2 studies reported clinical responses in low-grade glioma^{173,966}, melanoma⁹⁶⁷⁻⁹⁷¹, and in lung^{177,972-973} and endometrial cancer⁹⁶⁵. A Phase 2 study of selumetinib for patients with activating alterations in the MAPK pathway reported a DCR of 15% (3/20), with no objective responses observed974. Phase 1 studies of selumetinib to treat patients with solid tumors reported 1/15 PR for a patient with colorectal cancer (CRC) and 5/ 15 SDs for patients with tonsil squamous cell carcinoma (SCC), non-small cell lung cancer (NSCLC), and CRC975; 2/39 PRs (for patients with CRC) and 18/39 SDs were achieved when selumetinib was administered in combination with cyclosporin A⁹⁷⁶. Multiple Phase 1 studies combining selumetinib with erlotinib or temsirolimus⁹⁷⁷, docetaxel or dacarbazine⁹⁷⁸, AKT inhibitors979, or cixutumumab (an anti-IGF-1R antibody)980 reported clinical responses for patients with advanced solid tumors including NSCLC, thyroid carcinoma, tongue SCC, and ovarian cancer.

Talazoparib

Assay findings association

ATM R250*

BRCA2 E1879D

AREAS OF THERAPEUTIC USE

The PARP inhibitor talazoparib is FDA approved to treat HER2-negative locally advanced or metastatic breast cancer with deleterious or suspected deleterious germline BRCA mutations. Please see the drug label for full prescribing information.

GENE ASSOCIATION

ATM inactivation may predict sensitivity to PARP inhibitors. ATM alterations have been associated with clinical benefit from PARP inhibitors in solid tumors, including prostate cancer^{53-55,887}, colorectal cancer⁵⁷, breast cancer⁵⁷, gastric cancer⁵⁶, cholangiocarcinoma⁵⁹, and papillary renal cell carcinoma⁵⁸. On the basis of strong clinical data in breast cancer⁹⁸¹⁻⁹⁸³ and additional clinical evidence in ovarian, pancreatic, and prostate cancer⁹⁸⁴⁻⁹⁸⁷, loss or inactivation of either BRCA1 or BRCA2 may confer sensitivity to talazoparib.

SUPPORTING DATA

A Phase 1 study reported a PR for a patient with uterine

carcinoma and a BRCA2 germline mutation treated with talazoparib plus carboplatin and paclitaxel followed by talazoparib monotherapy988. Talazoparib has been studied primarily in the context of BRCA-mutated, HER2-negative breast cancer, where patients achieved significantly longer median PFS (8.6 vs. 5.6 months, HR=0.54), a higher ORR (62.6% vs. 27.2%), and improved quality of life on talazoparib compared with standard chemotherapy in a Phase 3 study⁹⁸²⁻⁹⁸³. In a Phase 2 study of talazoparib for BRCA1/2-wildtype patients with homologous recombination pathway alterations, the best outcome in non-breast tumors was SD \geq 6 months for 2/7patients who had colon cancer with germline ATM alteration or testicular cancer with germline CHEK2 and somatic ATM alteration⁵⁷. Clinical activity of single-agent talazoparib has been observed in numerous other solid tumors, including responses in BRCA-mutated ovarian, pancreatic, prostate, and ampulla of Vater cancers; PALB2-mutated pancreatic and bladder cancers; ATMmutated cholangiocarcinoma; and small cell lung cancer^{984-986,989}

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Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531



THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Temsirolimus

Assay findings association

FBXW7 R441W

PIK3CA R88Q, T1025A

AREAS OF THERAPEUTIC USE

Temsirolimus is an intravenous mTOR inhibitor that is FDA approved for the treatment of advanced renal cell carcinoma. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of several case reports describing clinical benefit for patients with FBXW7-mutated cancers treated with mTOR inhibitors, including lung adenocarcinoma¹⁵⁰, renal cell carcinoma⁵⁸, and cervical squamous cell carcinoma⁸⁷⁹, FBXW7 loss or inactivation may predict sensitivity to everolimus or temsirolimus. On the basis of clinical evidence^{178,210-216}, PIK₃CA activating mutations may predict sensitivity to mTOR inhibitors such as everolimus and temsirolimus. Studies have reported modest activity of these therapies as single agents (ORRs of o-4%), but improved activity has been observed when they are combined with other agents such as bevacizumab and doxorubicin (ORRs of 25-44%), for patients with PIK₃CA-mutated solid tumors^{213-216,880-884}.

SUPPORTING DATA

In a pooled analysis, 14% (3/21) of patients with PIK3CA-

mutated endometrial cancer treated with temsirolimus or ridaforolimus achieved objective response and 29% (6/21) experienced disease progression³⁹⁶. A case report described a patient with heavily pretreated PIK3CAmutated endometrial cancer who had SD for 17 months with temsirolimus alone followed by combination with letrozole990. A Phase 2 clinical trial of temsirolimus in recurrent or metastatic endometrial cancer reported PR in 4/29 (14%) chemotherapy-naïve patients and 4% (1/25) of chemotherapy-treated patients, with SD reported in 69% (20/29) of chemotherapy-naïve patients and 48% (12/25) of chemotherapy-treated patients; however, response in this study was found to be independent of molecular markers of PI₃K-AKT-mTOR pathway activation³⁹⁵. Another Phase 2 study of temsirolimus in patients with endometrial cancer reported PFS of >15 months in 6 patients and associated clinical benefit and longer PFS with mutation of AKT1 or CTNNB1, respectively⁹⁹¹. Temsirolimus combined with carboplatin and paclitaxel achieved objective partial responses in 82% (9/11) of patients with endometrial cancer⁹⁹². A Phase 2 trial of temsirolimus in combination with bevacizumab in patients with endometrial carcinoma reported clinical response in 25% of patients⁹⁹³.

Trametinib

Assay findings association

NF1 R1362*, R2450*

AREAS OF THERAPEUTIC USE

Trametinib is a MEK inhibitor that is FDA approved as a monotherapy to treat patients with melanoma with BRAF V600E or V600K mutations. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical evidence in neurofibromatosis type 1 (NF1)-associated neurofibroma $^{169-172,959-963}$, glioma $^{172-176,964}$, and non-small cell lung cancer 177 , NF1 inactivation may predict sensitivity to MEK inhibitors.

SUPPORTING DATA

Although very little clinical data has been reported on the use of MEK inhibitors in endometrial or uterine cancer, a Phase 1 trial of trametinib in combination with an AKT inhibitor (GSK2141795), reported 3/3 objective responses in uterine/endometrial tumors, with 2 patients achieving stable disease (SD) and 1 partial response (PR) occurring in a patient with mutant KRAS and PTEN loss⁹⁹⁴. In other solid tumor types, a Phase 1 trial of trametinib in 206 patients with solid tumors reported 21 (10%) objective responses⁹⁹⁵. Phase 1 monotherapy trials of RO4987655, another MEK inhibitor, have shown significant response

rates in patients with melanoma, including those with BRAF and NRAS mutations, but very low response rates in patients with other solid tumors, including those with KRAS mutations996-997. A Phase 1b trial of trametinib in combination with gemcitabine in patients with solid tumors showed a complete response in a patient with breast cancer, as well as partial responses in patients with pancreatic or salivary gland cancer⁹⁹⁸. A Phase 1b trial of combination treatment with the MEK inhibitor MEK162 and the PI₃K-alpha inhibitor BYL₇₁₉ reported disease control (partial responses or stable disease) in 47% (21/45) of patients, including partial responses in 2 of 3 patients with KRAS-mutant ovarian cancer and 1 of 3 patients with NRAS-mutant melanoma; a 43% rate of stable disease was observed in patients with KRAS-mutant colorectal cancer, with responses independent of PIK3CA mutation status⁹⁹⁹. Whereas frequent adverse events precluded a recommended Phase 2 dose and schedule for the combination of trametinib and everolimus in a Phase 1b trial for solid tumors¹⁸⁴, a retrospective study for heavily pretreated patients with solid tumors reported tolerable regimens of the combination for 23/31 patients, with 16 patients treated >3 months and evaluable patients achieving a median PFS of 6.5 months 185.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.



CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity > Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov. Or visit https://www.foundationmedicine.com/genomictesting#support-services.

BIOMARKER

Tumor Mutational

Burden

RESULT 372 Muts/Mb **RATIONALE**

Increased tumor mutational burden may predict response to anti-PD-1 (alone or in combination

with anti-CTLA-4) or anti-PD-L1 immune checkpoint inhibitors.

NCT04237649	PHASE NULL
KAZ954 Alone and With PDR001, NZV930 and NIR178 in Advanced Solid Tumors	TARGETS ADORA2A, CD73, PD-1

LOCATIONS: Taipei (Taiwan), Sunto Gun (Japan), Singapore (Singapore), Milano (Italy), Barcelona (Spain), California, Illinois, Missouri, Connecticut, Texas

NCT04589845	PHASE 2
Tumor-Agnostic Precision Immuno-Oncology and Somatic Targeting Rational for You (TAPISTRY) Platform Study	TARGETS ALK, ROS1, TRKA, TRKB, TRKC, RET, PD-L1, AKTs, ERBB2, MDM2, PI3K- alpha

LOCATIONS: Zhongzheng Dist. (Taiwan), Taipei City (Taiwan), Tainan (Taiwan), Seoul (Korea, Republic of), Beijing (China), Woolloongabba (Australia), Darlinghurst (Australia), Randwick (Australia), Melbourne (Australia), Haifa (Israel)

NCT04181788	PHASE 1/2
Sasanlimab (PF-06801591, PD-1 Inhibitor) in Participants With Advanced Malignancies	TARGETS PD-1

LOCATIONS: Taipei (Taiwan), Kaohsiung (Taiwan), Shanghai (China), Nanjing (China), Incheon (Korea, Republic of), Seoul (Korea, Republic of), Chongqing (China), Beijing (China), Chuo-ku (Japan), Kopeysk (Russian Federation)

NCT02829723	PHASE 1/2
Phase I/II Study of BLZ945 Single Agent or BLZ945 in Combination With PDR001 in Advanced Solid Tumors	TARGETS PD-1, CSF1R

LOCATIONS: Taipei (Taiwan), Tainan (Taiwan), Nagoya (Japan), Koto ku (Japan), Singapore (Singapore), Tel Aviv (Israel), Zurich (Switzerland), Rozzano (Italy), Barcelona (Spain), Hospitalet de LLobregat (Spain)

NCT03530397	PHASE 1
A Study to Evaluate MEDI5752 in Subjects With Advanced Solid Tumors	TARGETS PD-L1, PD-1, CTLA-4

LOCATIONS: Taipei (Taiwan), Cheongju-si (Korea, Republic of), Incheon (Korea, Republic of), Seoul (Korea, Republic of), Gyeonggi-do (Korea, Republic of), Amsterdam (Netherlands), Napoli (Italy), Roma (Italy), Villejuif Cedex (France), Barcelona (Spain)



CLINICAL TRIALS

NCT02628067	PHASE 2
Study of Pembrolizumab (MK-3475) in Participants With Advanced Solid Tumors (MK-3475-158/KEYNOTE-158)	TARGETS PD-1

LOCATIONS: Taipei (Taiwan), Makati (Philippines), Seoul (Korea, Republic of), Beijing (China), Chiyoda-Ku, Tokyo (Japan), North Ryde (Australia), Moscow (Russian Federation), Hod Hasharon (Israel), Drammen (Norway), Glostrup (Denmark)

NCT03192345	PHASE 1
A First-in-human Study of the Safety, Pharmacokinetics, Pharmacodynamics and Anti-tumor Activity of SAR439459 Monotherapy and Combination of SAR439459 and Cemiplimab in Patients With Advanced Solid Tumors	TARGETS PD-1, TGF-beta
LOCATIONS: Tainei 100 (Taiwan) Tainan (Taiwan) Kashsiung (Taiwan) Social (Korea Bonublic of) Ho	sidolborg Most (Australia) Molbourno (Australia)

LOCATIONS: Taipei 100 (Taiwan), Tainan (Taiwan), Kaohsiung (Taiwan), Seoul (Korea, Republic of), Heidelberg West (Australia), Melbourne (Australia), Tallinn (Estonia), Hannover (Germany), Essen (Germany), Utrecht (Netherlands)

NCT04261439	PHASE 1
A Phase I/Ib Study of NIZ985 Alone and in Combination With Spartalizumab	TARGETS PD-1

LOCATIONS: Taipei (Taiwan), Chuo ku (Japan), Essen (Germany), Napoli (Italy), Leuven (Belgium), Barcelona (Spain), California, Texas

NCT03565445	PHASE 1
A Study of ASP1948, Targeting an Immune Modulatory Receptor, in Subjects With Advanced Solid Tumors	TARGETS PD-1, NRP1

LOCATIONS: Taipei (Taiwan), Seoul (Korea, Republic of), Tokyo (Japan), Chiba (Japan), Meldola (Italy), Modena (Italy), Newcastle upon Tyne (United Kingdom), Monza (Italy), Milano (Italy), Glasgow (United Kingdom)

NCT03799003	PHASE 1
A Study of ASP1951 in Subjects With Advanced Solid Tumors	TARGETS PD-1, TNFRSF18

LOCATIONS: Taipei (Taiwan), Taichung (Taiwan), Daegu (Korea, Republic of), Chungcheongbukdo (Korea, Republic of), Seoul (Korea, Republic of), Gyeonggi-do (Korea, Republic of), Washington, California, Nevada



CLINICAL TRIALS

GENE ATM **RATIONALE**

Loss or inactivation of ATM may increase sensitivity to PARP inhibitors, ATR inhibitors, or

DNA-PKcs inhibitors.

ALTERATION R250*

NCT04269200

Durvalumab With or Without Olaparib as Maintenance Therapy After First-Line Treatment of Advanced and Recurrent Endometrial Cancer

TARGETS PD-L1, PARP

LOCATIONS: Nakagami-gun (Japan), Shanghai (China), Guangdong (China), Hong Kong (Hong Kong), HKG (Hong Kong), Changchun (China), Wuhan (China), Kurume-shi (Japan), Gyeongsangnam-do (Korea, Republic of), Suwon (Korea, Republic of)

NCTO4123366

Study of Olaparib (MK-7339) in Combination With Pembrolizumab (MK-3475) in the Treatment of Homologous Recombination Repair Mutation (HRRm) and/or Homologous Recombination Deficiency (HRD)-Positive Advanced Cancer (MK-7339-007/KEYLYNK-007)

TARGETS
PARP, PD-1

LOCATIONS: Fukuoka (Japan), Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Okayama (Japan), Nagoya (Japan), Tokyo (Japan), Kashiwa (Japan), Sapporo (Japan), Nedlands (Australia), Southport (Australia)

NCT03742895

Efficacy and Safety of Olaparib (MK-7339) in Participants With Previously Treated, Homologous Recombination Repair Mutation (HRRm) or Homologous Recombination Deficiency (HRD) Positive Advanced Cancer (MK-7339-002 / LYNK-002)

TARGETS PARP

LOCATIONS: Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Chelyabinsk (Russian Federation), Nedlands (Australia), Kazan (Russian Federation), Arkhangelsk (Russian Federation), Port Macquarie (Australia), Ryazan (Russian Federation), Darlinghurst (Australia), Moscow (Russian Federation)

NCTO4716686

Niraparib Monotherapy as Maintain and Recurrent Treatment of Endometrial Serous Carcinoma
TARGETS
PARP

LOCATIONS: Jinan (China)

NCT02264678 PHASE 1/2

Ascending Doses of AZD6738 in Combination With Chemotherapy and/or Novel Anti Cancer Agents

TARGETS

ATR, PARP, PD-L1

LOCATIONS: Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Cambridge (United Kingdom), Withington (United Kingdom), London (United Kingdom), Sutton (United Kingdom), Villejuif (France), Saint Herblain (France), California

NCTO2630199

Study of AZD6738, DNA Damage Repair/Novel Anti-cancer Agent, in Combination With Paclitaxel, in Refractory Cancer

LOCATIONS: Seoul (Korea, Republic of)



NCT04635631	PHASE 1
STUDY OF TALAZOPARIB MONOTHERAPY IN CHINESE PARTICIPANTS WITH ADVANCED SOLID TUMORS	TARGETS PARP
LOCATIONS: Beijing (China), Changchun (China)	
NCT03188965	PHASE 1
First-in-human Study of ATR Inhibitor BAY1895344 in Patients With Advanced Solid Tumors and Lymphomas	TARGETS ATR
LOCATIONS: Sunto (Japan), Chuo-ku (Japan), Kashiwa (Japan), Singapore (Singapore), St. Gallen (Sv. Tyne (United Kingdom), Genève (Switzerland), Sutton (United Kingdom), Edmonton (Canada)	witzerland), Bellinzona (Switzerland), Newcastle Upoi
NCT03772561	PHASE 1
Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies	TARGETS PARP, AKTs, PD-L1
LOCATIONS: Singapore (Singapore)	
NCT04801966	PHASE NULL
Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study	TARGETS CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF
LOCATIONS: Melbourne (Australia)	



CLINICAL TRIALS

GENE		
BR	CA	2

ALTERATION E1879D

RATIONALE

BRCA2 loss or inactivating alterations may predict sensitivity to PARP inhibitors or to ATR inhibitors. It is not known whether these

therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

NCT04269200	PHASE 3
Durvalumab With or Without Olaparib as Maintenance Therapy After First-Line Treatment of Advanced and Recurrent Endometrial Cancer	TARGETS PD-L1, PARP

LOCATIONS: Nakagami-gun (Japan), Shanghai (China), Guangdong (China), Hong Kong (Hong Kong), HKG (Hong Kong), Changchun (China), Wuhan (China), Kurume-shi (Japan), Gyeongsangnam-do (Korea, Republic of), Suwon (Korea, Republic of)

NCT03239015	PHASE 2
Efficacy and Safety of Targeted Precision Therapy in Refractory Tumor With Druggable Molecular Event	TARGETS EGFR, ERBB2, ERBB4, PARP, mTOR, MET, RET, ROS1, VEGFRS, BRAF, CDK4, CDK6

LOCATIONS: Shanghai (China)

NCT04123366	PHASE 2
Study of Olaparib (MK-7339) in Combination With Pembrolizumab (MK-3475) in the Treatment of Homologous Recombination Repair Mutation (HRRm) and/or Homologous Recombination Deficiency (HRD)-Positive Advanced Cancer (MK-7339-007/KEYLYNK-007)	TARGETS PARP, PD-1

LOCATIONS: Fukuoka (Japan), Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Okayama (Japan), Nagoya (Japan), Tokyo (Japan), Kashiwa (Japan), Sapporo (Japan), Nedlands (Australia), Southport (Australia)

NCT03742895	PHASE 2
Efficacy and Safety of Olaparib (MK-7339) in Participants With Previously Treated, Homologous Recombination Repair Mutation (HRRm) or Homologous Recombination Deficiency (HRD) Positive Advanced Cancer (MK-7339-002 / LYNK-002)	TARGETS PARP

LOCATIONS: Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Chelyabinsk (Russian Federation), Nedlands (Australia), Kazan (Russian Federation), Arkhangelsk (Russian Federation), Port Macquarie (Australia), Ryazan (Russian Federation), Darlinghurst (Australia), Moscow (Russian Federation)

NCT04716686	PHASE 2
Niraparib Monotherapy as Maintain and Recurrent Treatment of Endometrial Serous Carcinoma	TARGETS PARP
LOCATIONS: Jinan (China)	



NCT02264678	PHASE 1/2
Ascending Doses of AZD6738 in Combination With Chemotherapy and/or Novel Anti Cancer Agents	TARGETS ATR, PARP, PD-L1
LOCATIONS: Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Cambridge (United Kingdom), Sutton (United Kingdom), Villejuif (France), Saint Herblain (France), California	om), Withington (United Kingdom), London (United
NCT02630199	PHASE 1
Study of AZD6738, DNA Damage Repair/Novel Anti-cancer Agent, in Combination With Paclitaxel, in Refractory Cancer	TARGETS ATR
LOCATIONS: Seoul (Korea, Republic of)	
NCT04635631	PHASE 1
STUDY OF TALAZOPARIB MONOTHERAPY IN CHINESE PARTICIPANTS WITH ADVANCED SOLID TUMORS	TARGETS PARP
LOCATIONS: Beijing (China), Changchun (China)	
NCT03188965	PHASE 1
First-in-human Study of ATR Inhibitor BAY1895344 in Patients With Advanced Solid Tumors and Lymphomas	TARGETS ATR
LOCATIONS: Sunto (Japan), Chuo-ku (Japan), Kashiwa (Japan), Singapore (Singapore), St. Gallen (Sw Tyne (United Kingdom), Genève (Switzerland), Sutton (United Kingdom), Edmonton (Canada)	itzerland), Bellinzona (Switzerland), Newcastle Upon
NCT03772561	PHASE 1
Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies	TARGETS PARP, AKTs, PD-L1
LOCATIONS: Singapore (Singapore)	



CLINICAL TRIALS

GE	ΝE				
C	T	N	N	В	1

ALTERATION K335T

RATIONALE

Based on clinical and preclinical evidence, tumors with activating CTNNB1 alterations may be sensitive to mTOR inhibitors. It is not known

whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

NCT04337463	PHASE NULL	
ATG-008 Combined With Toripalimab in Advanced Solid Tumors	TARGETS mTORC1, mTORC2, PD-1	
LOCATIONS: Chongqing (China), Chengdu (China)		
NCT04803318	PHASE 2	
Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors	TARGETS mTOR, FGFRs, KIT, PDGFRA, RET, VEGFRs, MEK	
LOCATIONS: Guangzhou (China)		
NCT03008408 PHASE 2		
Phase II Ribociclib, Everolimus and Letrozole in Endometrial Cancer TARGETS Aromatase, mTOR, CDK		
LOCATIONS: Texas		
NCT03065062	PHASE 1	
Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head & Neck and Other Solid Tumors TARGETS PI3K-alpha, PI3K-gamm mTORC2, CDK4, CDK6		
LOCATIONS: Massachusetts		
NCT01582191	PHASE 1	
A Phase 1 Trial of Vandetanib (a Multi-kinase Inhibitor of EGFR, VEGFR and RET Inhibitor) in Combination With Everolimus (an mTOR Inhibitor) in Advanced Cancer	TARGETS mTOR, EGFR, RET, SRC, VEGFRS	
LOCATIONS: Texas		
NCT02159989	PHASE 1	
Sapanisertib and Ziv-Aflibercept in Treating Patients With Recurrent Solid Tumors That Are Metastatic or Cannot Be Removed by Surgery	TARGETS PIGF, VEGFA, VEGFB, mTORC1, mTORC2	



NCT02321501	PHASE 1
Phase I/Ib Dose Escalation & Biomarker Study of Ceritinib (LDK378) + Everolimus for Locally Advanced or Metastatic Solid Tumors With an Expansion in Non-Small Cell Lung Cancer (NSCLC) Characterized by Abnormalities in Anaplastic Lymphoma Kinase (ALK) Expression	TARGETS ROS1, ALK, mTOR
LOCATIONS: Texas	
NCT03017833	PHASE 1
Sapanisertib and Metformin in Treating Patients With Advanced or Metastatic Relapsed or Refractory Cancers	TARGETS mTORC1, mTORC2
LOCATIONS: Texas	
NCT03430882	PHASE 1
Sapanisertib, Carboplatin, and Paclitaxel in Treating Patients With Recurrent or Refractory Malignant Solid Tumors	TARGETS mTORC1, mTORC2
LOCATIONS: Texas	



CLINICAL TRIALS

FANCL

RATIONALE

On the basis of clinical evidence in ovarian cancer, to PARP inhibitors.

FANCL loss or inactivation may confer sensitivity

ALTERATION splice site 904-2A>C

NCT04269200 PHASE 3

Durvalumab With or Without Olaparib as Maintenance Therapy After First-Line Treatment of Advanced and Recurrent Endometrial Cancer

TARGETS
PD-L1, PARP

LOCATIONS: Nakagami-gun (Japan), Shanghai (China), Guangdong (China), Hong Kong (Hong Kong), HKG (Hong Kong), Changchun (China), Wuhan (China), Kurume-shi (Japan), Gyeongsangnam-do (Korea, Republic of), Suwon (Korea, Republic of)

NCT04123366 PHASE 2

Study of Olaparib (MK-7339) in Combination With Pembrolizumab (MK-3475) in the Treatment of Homologous Recombination Repair Mutation (HRRm) and/or Homologous Recombination Deficiency (HRD)-Positive Advanced Cancer (MK-7339-007/KEYLYNK-007)

LOCATIONS: Fukuoka (Japan), Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Okayama (Japan), Nagoya (Japan), Tokyo (Japan), Kashiwa (Japan), Sapporo (Japan), Nedlands (Australia), Southport (Australia)

NCT03742895 PHASE 2

Efficacy and Safety of Olaparib (MK-7339) in Participants With Previously Treated, Homologous
Recombination Repair Mutation (HRRm) or Homologous Recombination Deficiency (HRD) Positive
Advanced Cancer (MK-7339-002 / LYNK-002)

LOCATIONS: Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Chelyabinsk (Russian Federation), Nedlands (Australia), Kazan (Russian Federation), Arkhangelsk (Russian Federation), Port Macquarie (Australia), Ryazan (Russian Federation), Darlinghurst (Australia), Moscow (Russian Federation)

NCT04716686 PHASE 2

Niraparib Monotherapy as Maintain and Recurrent Treatment of Endometrial Serous Carcinoma

TARGETS
PARP

LOCATIONS: Jinan (China)

NCT02264678 PHASE 1/2

Ascending Doses of AZD6738 in Combination With Chemotherapy and/or Novel Anti Cancer Agents TARGETS

ATR, PARP, PD-L1

LOCATIONS: Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Cambridge (United Kingdom), Withington (United Kingdom), London (United Kingdom), Sutton (United Kingdom), Villejuif (France), Saint Herblain (France), California

NCT04635631 PHASE 1

STUDY OF TALAZOPARIB MONOTHERAPY IN CHINESE PARTICIPANTS WITH ADVANCED SOLID TUMORS

TARGETS
PARP

LOCATIONS: Beijing (China), Changchun (China)



NCT03772561	PHASE 1
Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies	t TARGETS PARP, AKTs, PD-L1
LOCATIONS: Singapore (Singapore)	
NCT04801966	PHASE NULL
Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study	TARGETS CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF
LOCATIONS: Melbourne (Australia)	
NCT04497116	PHASE 1/2
Study of RP-3500 in Advanced Solid Tumors	TARGETS ATR, PARP
LOCATIONS: Copenhagen (Denmark), Newcastle Upon Tyne (United Kingdom), Manchester (United Canada), Massachusetts, Rhode Island, New York, Tennessee, Texas	ited Kingdom), London (United Kingdom), Toronto
NCT04159155	PHASE 2/3
A Study of Various Treatments in Serous or p53 Abnormal Endometrial Cancer	TARGETS PARP
LOCATIONS: Toronto (Canada)	



CLINICAL TRIALS

GEN	E		
FB	X	W	7

ALTERATION R441W

RATIONALE

Loss or inactivation of FBXW7 may lead to increased mTOR activation and may predict sensitivity to mTOR inhibitors. It is not known

whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

PHASE 2	
TARGETS EGFR, ERBB2, ERBB4, PARP, mTOR, MET, RET, ROS1, VEGFRS, BRAF, CDK4 CDK6	
PHASE NULL	
TARGETS mTORC1, mTORC2, PD-1	
PHASE 2	
TARGETS mTOR, FGFRs, KIT, PDGFRA, RET, VEGFRs, MEK	
PHASE 2	
TARGETS VEGFRs, ABL, SRC, ALK, AXL, MET, ROS1, TRKA, TRKC, DDR2, KIT, EGFR, PD-1, CTLA-4, PARP, CDK4, CDK6, CSF1R, FLT3, RET, mTOR, ERBB2, ERBB3, MEK, BRAF, SMO	
wa (Canada), Montreal (Canada), Toronto (Canada	
PHASE 2	
TARGETS mTOR	



NCT03008408	PHASE 2
Phase II Ribociclib, Everolimus and Letrozole in Endometrial Cancer	TARGETS Aromatase, mTOR, CDK4, CDK6
LOCATIONS: Texas	
NCT03217669	PHASE 1
Epacadostat (INCB24360) in Combination With Sirolimus in Advanced Malignancy	TARGETS IDO1, mTOR
LOCATIONS: Kansas	
NCT03065062	PHASE 1
Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head & Neck and Other Solid Tumors	TARGETS PI3K-alpha, PI3K-gamma, mTORC1, mTORC2, CDK4, CDK6
LOCATIONS: Massachusetts	
NCT01582191	PHASE 1
A Phase 1 Trial of Vandetanib (a Multi-kinase Inhibitor of EGFR, VEGFR and RET Inhibitor) in Combination With Everolimus (an mTOR Inhibitor) in Advanced Cancer	TARGETS mTOR, EGFR, RET, SRC, VEGFRs
LOCATIONS: Texas	
NCT02159989	PHASE 1
Sapanisertib and Ziv-Aflibercept in Treating Patients With Recurrent Solid Tumors That Are Metastatic or Cannot Be Removed by Surgery	TARGETS PIGF, VEGFA, VEGFB, mTORC1, mTORC2
LOCATIONS: Texas	



CLINICAL TRIALS

GENE	
FGI	FR2

RATIONALE

FGFR inhibitors may be relevant in tumors with

alterations that activate FGFR2.

ALTERATION N549D

NCT04865289	PHASE 3
Pembrolizumab (MK-3475) Plus Lenvatinib (E7080/MK-7902) Versus Chemotherapy for Endometrial Carcinoma (ENGOT-en9 / MK-7902-001) -China Extension Study	TARGETS FGFRS, KIT, PD-1, PDGFRA, RET, VEGFRS

LOCATIONS: Hangzhou (China), Shanghai (China), Nanchang (China), Nanjing (China), Hefei (China), Guangzhou (China), Changsha (China), Wuhan (China), Nanning (China), Chongqing (China)

NCT02450136	PHASE NULL
Single-arm Study to Evaluate the Safety and Efficacy of Pazopanib, in Subjects With FGFR2 Amplification, FGFR2 Mutation Refractory Solid Tumors	TARGETS FGFR1, FGFR2, FGFR3, KIT, VEGFRS
LOCATIONS: Seoul (Korea, Republic of)	

NCT04803318	PHASE 2
Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors	TARGETS mTOR, FGFRs, KIT, PDGFRA, RET, VEGFRs, MEK
LOCATIONS: Guangzhou (China)	

NCT03564691	PHASE 1
Study of MK-4830 as Monotherapy and in Combination With Pembrolizumab (MK-3475) in Participants With Advanced Solid Tumors (MK-4830-001)	TARGETS ITL4, FGFRs, KIT, PDGFRA, RET, VEGFRs, PD-1

LOCATIONS: Seoul (Korea, Republic of), Tokyo (Japan), Haifa (Israel), Petah Tikva (Israel), Ramat Gan (Israel), Tel Aviv (Israel), Warszawa (Poland), Gdansk (Poland), Heraklion (Greece), Washington

NCT04042116	PHASE 1/2
A Study to Evaluate Lucitanib in Combination With Nivolumab in Patients With a Solid Tumor	TARGETS FGFRs, VEGFRs, PD-1

LOCATIONS: Innsbruck (Austria), Essen (Germany), Bologna (Italy), Naples (Italy), Leuven (Belgium), Brussels (Belgium), Ghent (Belgium), Washington, Barcelona (Spain), Madrid (Spain)

NCT04565275	PHASE 1/2
A Study of ICP-192 in Patients With Advanced Solid Tumors	TARGETS FGFR1, FGFR2, FGFR3, FGFR4
LOCATIONS: Colorado, Minnesota, Arizona, Florida	



NCT02272998	PHASE 2
Ponatinib for Patients Whose Advanced Solid Tumor Cancer Has Activating Mutations Involving the Following Genes: FGFR1, FGFR2, FGFR3, FGFR4, RET, KIT.	TARGETS ABL, FGFRs, FLT3, KIT, RET, VEGFRs
LOCATIONS: Ohio	
NCT02856425	PHASE 1
Trial Of Pembrolizumab And Nintedanib	TARGETS FGFR1, FGFR2, FGFR3, FLT3, LCK, LYN, SRC, VEGFRs, PD-1
LOCATIONS: Villejuif (France)	
NCT01543763	PHASE 1
Phase I Tolerability, Efficacy, and Safety Study of Pazopanib in Combination With PCI-24781 in Patients With Metastatic Solid Tumors	TARGETS HDAC, FGFR1, FGFR2, FGFR3, KIT, VEGFRs
LOCATIONS: California	



CLINICAL TRIALS

GENE	:
NF	1

INF I

ALTERATION R1362*, R2450*

RATIONALE

On the basis of clinical evidence and strong preclinical evidence, NF1 inactivation may predict sensitivity to MEK inhibitors. Limited clinical

data and strong preclinical data indicate that loss or inactivation of NF1 may also predict sensitivity to mTOR inhibitors.

PHASE 2	
TARGETS EGFR, ERBB2, ERBB4, PARP, mTOR, MET, RET, ROS1, VEGFRS, BRAF, CDK4, CDK6	
PHASE NULL	
TARGETS mTORC1, mTORC2, PD-1	
PHASE 2	
TARGETS mTOR, FGFRs, KIT, PDGFRA, RET, VEGFRs, MEK	
PHASE 1/2	
TARGETS SHP2, MEK	
PHASE NULL	
TARGETS CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF	
PHASE 1/2	
TARGETS RAFs, EGFR, MEK	
LOCATIONS: Nedlands (Australia), Blacktown (Australia), Randwick (Australia), Melbourne (Australia), Texas	



LOCATIONS: Toronto (Canada)

NCT03297606	PHASE 2
Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)	TARGETS VEGFRS, ABL, SRC, ALK, AXL, MET, ROS1, TRKA, TRKC, DDR2, KIT, EGFR, PD-1, CTLA-4, PARP, CDK4, CDK6, CSF1R, FLT3, RET, mTOR, ERBB2, ERBB3, MEK, BRAF, SMO
LOCATIONS: Vancouver (Canada), Edmonton (Canada), Saskatoon (Canada), Regina (Canada), Ottawa (Canada), Montreal (Canada), Toronto (Canada), Kingston (Canada), London (Canada)	
NCT04800822	PHASE 1
PF-07284892 in Participants With Advanced Solid Tumors	TARGETS SHP2, ROS1, ALK, BRAF, EGFR, MEK
LOCATIONS: California, Michigan, New York, Tennessee, Texas	
NCT02397083	PHASE 2
Phase II Study of Intrauterine Device (IUD) Alone or in Combination With Everolimus in Endometrial Cancer	TARGETS mTOR
LOCATIONS: Colorado, Michigan, Virginia, Texas	
NCT02070549	PHASE 1
Trametinib in Treating Patients With Advanced Cancer With or Without Hepatic Dysfunction	TARGETS MEK



CLINICAL TRIALS

GENE	
PIK3CA	١

ALTERATION R88Q, T1025A

RATIONALE

PIK₃CA activating mutations may lead to activation of the PI₃K-AKT-mTOR pathway and may therefore indicate sensitivity to inhibitors of

this pathway. Strong clinical data support sensitivity of PIK₃CA-mutated solid tumors to the PI₃K-alpha inhibitor alpelisib.

•	
NCT04589845	PHASE 2
Tumor-Agnostic Precision Immuno-Oncology and Somatic Targeting Rational for You (TAPISTRY) Platform Study	TARGETS ALK, ROS1, TRKA, TRKB, TRKC, RET, PD-L1, AKTs, ERBB2, MDM2, PI3K- alpha
LOCATIONS: Zhongzheng Dist. (Taiwan), Taipei City (Taiwan), Tainan (Taiwan), Seoul (Korea, Republi Darlinghurst (Australia), Randwick (Australia), Melbourne (Australia), Haifa (Israel)	ic of), Beijing (China), Woolloongabba (Australia),
NCT03239015	PHASE 2
Efficacy and Safety of Targeted Precision Therapy in Refractory Tumor With Druggable Molecular Event	TARGETS EGFR, ERBB2, ERBB4, PARP, mTOR, MET, RET, ROS1, VEGFRS, BRAF, CDK4, CDK6
LOCATIONS: Shanghai (China)	
NCT04337463	PHASE NULL
ATG-008 Combined With Toripalimab in Advanced Solid Tumors	TARGETS mTORC1, mTORC2, PD-1
LOCATIONS: Chongqing (China), Chengdu (China)	
NCT02688881	PHASE 4
Study to Evaluate the Safety and Efficacy of Sirolimus, in Subject With Refractory Solid Tumors	TARGETS mTOR
LOCATIONS: Seoul (Korea, Republic of)	
NCT04803318	PHASE 2
Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors	TARGETS mTOR, FGFRs, KIT, PDGFRA, RET, VEGFRs, MEK
LOCATIONS: Guangzhou (China)	
NCT03772561	PHASE 1
Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies	TARGETS PARP, AKTs, PD-L1
LOCATIONS: Singapore (Singapore)	



NCT04801966	PHASE NULL
Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study	TARGETS CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF
LOCATIONS: Melbourne (Australia)	
NCT04632992	PHASE 2
A Study Evaluating Targeted Therapies in Participants Who Have Advanced Solid Tumors With Genomic Alterations or Protein Expression Patterns Predictive of Response	TARGETS ALK, ROS1, TRKA, TRKB, TRKC, PD-L1, ERBB2, ERBB3, PI3K-alpha, RET, AKTs
LOCATIONS: Alaska, Washington, Oregon, California, Montana	
NCT03994796	PHASE 2
Genetic Testing in Guiding Treatment for Patients With Brain Metastases	TARGETS ALK, ROS1, TRKA, TRKB, TRKC, CDK4, CDK6, PI3K, mTOR
LOCATIONS: Alaska, Washington	
NCT03297606	PHASE 2
Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)	TARGETS VEGFRS, ABL, SRC, ALK, AXL, MET, ROS1, TRKA, TRKC, DDR2, KIT, EGFR, PD-1, CTLA-4, PARP, CDK4, CDK6, CSF1R, FLT3, RET, mTOR, ERBB2, ERBB3, MEK, BRAF, SMO
LOCATIONS: Vancouver (Canada), Edmonton (Canada), Saskatoon (Canada), Regina (Canada), C Kingston (Canada), London (Canada)	Ottawa (Canada), Montreal (Canada), Toronto (Canada)



CLINICAL TRIALS

GENE	
PIK3CE	

ALTERATION R321Q

RATIONALE

Preclinical evidence suggests that overexpression of PIK₃CB promotes cellular transformation, and that PTEN-deficient cancers may be dependent on PIK₃CB for growth. Therefore, inhibitors of PIK₃CB or downstream pathway components

including AKT may be beneficial in tumors harboring PIK₃CB alterations. It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

R321Q	PIK ₃ CB or downstream pathway components	been fully characterized, as seen here.
NCT03239015		PHASE 2
Efficacy and Safety of Targeted Precision Thera Event	py in Refractory Tumor With Druggable Molecular	TARGETS EGFR, ERBB2, ERBB4, PARP, mTOR, MET, RET, ROS1, VEGFRS, BRAF, CDK4, CDK6
LOCATIONS: Shanghai (China)		
NCT04337463		PHASE NULL
ATG-008 Combined With Toripalimab in Advan	ced Solid Tumors	TARGETS mTORC1, mTORC2, PD-1
LOCATIONS: Chongqing (China), Chengdu (Chi	ina)	
NCT04803318		PHASE 2
Trametinib Combined With Everolimus and Len Tumors	vatinib for Recurrent/Refractory Advanced Solid	TARGETS mTOR, FGFRs, KIT, PDGFRA, RET, VEGFRs, MEK
LOCATIONS: Guangzhou (China)		
NCT04001569		PHASE 1/2
AZD8186 and Paclitaxel in Advanced Gastric Ca	incer	TARGETS PI3K-beta
LOCATIONS: Seongnam-si (Korea, Republic of)		
NCT03772561		PHASE 1
Phase I Study of AZD5363 + Olaparib + Durvalu Tumor Malignancies	mab in Patients With Advanced or Metastatic Solid	TARGETS PARP, AKTs, PD-L1
LOCATIONS: Singapore (Singapore)		
NCT03994796		PHASE 2
Genetic Testing in Guiding Treatment for Patien	ts With Brain Metastases	TARGETS ALK, ROS1, TRKA, TRKB, TRKC, CDK4, CDK6, PI3K, mTOR
LOCATIONS: Alaska, Washington		



LOCATIONS: Maryland

NCT03297606	PHASE 2
Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)	TARGETS VEGFRS, ABL, SRC, ALK, AXL, MET, ROS1, TRKA, TRKC, DDR2, KIT, EGFR, PD-1, CTLA-4, PARP, CDK4, CDK6, CSF1R, FLT3, RET, mTOR, ERBB2, ERBB3, MEK, BRAF, SMO
LOCATIONS: Vancouver (Canada), Edmonton (Canada), Saskatoon (Canada), Regina (Canada), Ottaw. Kingston (Canada), London (Canada)	a (Canada), Montreal (Canada), Toronto (Canada),
NCT03673787	PHASE 1/2
A Trial of Ipatasertib in Combination With Atezolizumab	TARGETS AKTs, PD-L1
LOCATIONS: Sutton (United Kingdom)	
NCT02397083	PHASE 2
Phase II Study of Intrauterine Device (IUD) Alone or in Combination With Everolimus in Endometrial Cancer	TARGETS mTOR
LOCATIONS: Colorado, Michigan, Virginia, Texas	
NCT03711058	PHASE 1/2
Study of PI3Kinase Inhibition (Copanlisib) and Anti-PD-1 Antibody Nivolumab in Relapsed/Refractory Solid Tumors With Expansions in Mismatch-repair Proficient (MSS) Colorectal Cancer	TARGETS PD-1, PI3K



CLINICAL TRIALS

GEI	NE	
PI	K3R	1

RATIONALE

On the basis of clinical and strong preclinical data, sensitivity to pan-PI₃K or PI₃K-alpha-selective PIK₃R₁ loss or inactivation may indicate

inhibitors.

ALTERATION E476*

L4/0	
NCT04801966	PHASE NULL
Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study	TARGETS CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF
LOCATIONS: Melbourne (Australia)	
NCT03711058	PHASE 1/2
Study of PI3Kinase Inhibition (Copanlisib) and Anti-PD-1 Antibody Nivolumab in Relapsed/Refractory Solid Tumors With Expansions in Mismatch-repair Proficient (MSS) Colorectal Cancer	TARGETS PD-1, PI3K
LOCATIONS: Maryland	

NCT03502733	PHASE 1
Copanlisib and Nivolumab in Treating Patients With Metastatic Solid Tumors or Lymphoma	TARGETS PI3K, PD-1

LOCATIONS: Maryland, Texas

NCT03586661	PHASE 1
Niraparib and Copanlisib in Treating Participants With Recurrent Endometrial, Ovarian, Primary Peritoneal, or Fallopian Tube Cancer	TARGETS PI3K, PARP
LOCATIONS: Texas	



CLINICAL TRIALS

GENE PTEN

ALTERATION R130Q, R173H

RATIONALE

PTEN loss or inactivating mutations may lead to increased activation of the PI₃K-AKT-mTOR pathway and may indicate sensitivity to inhibitors

of this pathway. PTEN loss or inactivation may also predict sensitivity to PARP inhibitors.

NCT04269200	PHASE 3
Durvalumab With or Without Olaparib as Maintenance Therapy After First-Line Treatment of Advanced and Recurrent Endometrial Cancer	TARGETS PD-L1, PARP

LOCATIONS: Nakagami-gun (Japan), Shanghai (China), Guangdong (China), Hong Kong (Hong Kong), HKG (Hong Kong), Changchun (China), Wuhan (China), Kurume-shi (Japan), Gyeongsangnam-do (Korea, Republic of), Suwon (Korea, Republic of)

NCT04337463	PHASE NULL
ATG-008 Combined With Toripalimab in Advanced Solid Tumors	TARGETS mTORC1, mTORC2, PD-1
LOCATIONS: Chongqing (China), Chengdu (China)	

NCT04716686	PHASE 2
Niraparib Monotherapy as Maintain and Recurrent Treatment of Endometrial Serous Carcinoma	TARGETS PARP

LOCATIONS: Jinan (China)

NCT04001569	PHASE 1/2
AZD8186 and Paclitaxel in Advanced Gastric Cancer	TARGETS PI3K-beta

LOCATIONS: Seongnam-si (Korea, Republic of)

NCT02264678	PHASE 1/2
Ascending Doses of AZD6738 in Combination With Chemotherapy and/or Novel Anti Cancer Agents	TARGETS ATR, PARP, PD-L1

LOCATIONS: Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Cambridge (United Kingdom), Withington (United Kingdom), London (United Kingdom), Sutton (United Kingdom), Villejuif (France), Saint Herblain (France), California

NCT04635631	PHASE 1
STUDY OF TALAZOPARIB MONOTHERAPY IN CHINESE PARTICIPANTS WITH ADVANCED SOLID TUMORS	TARGETS PARP
LOCATIONS: Beijing (China), Changchun (China)	



NCT03772561	PHASE 1	
Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies	d TARGETS PARP, AKTs, PD-L1	
LOCATIONS: Singapore (Singapore)		
NCT04801966	PHASE NULL	
Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study	TARGETS CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF	
LOCATIONS: Melbourne (Australia)		
NCT04632992	PHASE 2	
A Study Evaluating Targeted Therapies in Participants Who Have Advanced Solid Tumors With Genomic Alterations or Protein Expression Patterns Predictive of Response	TARGETS ALK, ROS1, TRKA, TRKB, TRKC, PD-L1, ERBB2, ERBB3, PI3K-alpha, RET, AKTs	
LOCATIONS: Alaska, Washington, Oregon, California, Montana		
NCT04497116	PHASE 1/2	
Study of RP-3500 in Advanced Solid Tumors	TARGETS ATR, PARP	
LOCATIONS: Copenhagen (Denmark), Newcastle Upon Tyne (United Kingdom), Manchester (Unite (Canada), Massachusetts, Rhode Island, New York, Tennessee, Texas	d Kingdom), London (United Kingdom), Toronto	



CLINICAL TRIALS

GENE RAD51D

ALTERATION R253Q

RATIONALE

Inactivation of RAD51D may predict sensitivity to PARP inhibitors. It is not known whether these therapeutic approaches would be relevant in the

context of alterations that have not been fully characterized, as seen here.

NCT04269200 PHASE 3

Durvalumab With or Without Olaparib as Maintenance Therapy After First-Line Treatment of **TARGETS** Advanced and Recurrent Endometrial Cancer PD-L1, PARP

LOCATIONS: Nakagami-gun (Japan), Shanghai (China), Guangdong (China), Hong Kong (Hong Kong), HKG (Hong Kong), Changchun (China), Wuhan (China), Kurume-shi (Japan), Gyeongsangnam-do (Korea, Republic of), Suwon (Korea, Republic of)

NCT04123366 PHASE 2

Study of Olaparib (MK-7339) in Combination With Pembrolizumab (MK-3475) in the Treatment of **TARGETS** Homologous Recombination Repair Mutation (HRRm) and/or Homologous Recombination Deficiency PARP, PD-1 (HRD)-Positive Advanced Cancer (MK-7339-007/KEYLYNK-007)

LOCATIONS: Fukuoka (Japan), Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Okayama (Japan), Nagoya (Japan), Tokyo (Japan), Kashiwa (Japan), Sapporo (Japan), Nedlands (Australia), Southport (Australia)

NCT03742895 PHASE 2

Efficacy and Safety of Olaparib (MK-7339) in Participants With Previously Treated, Homologous **TARGETS** Recombination Repair Mutation (HRRm) or Homologous Recombination Deficiency (HRD) Positive **PARP** Advanced Cancer (MK-7339-002 / LYNK-002)

LOCATIONS: Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Chelyabinsk (Russian Federation), Nedlands (Australia), Kazan (Russian Federation), Arkhangelsk (Russian Federation), Port Macquarie (Australia), Ryazan (Russian Federation), Darlinghurst (Australia), Moscow (Russian

NCT04716686 PHASE 2

TARGETS Niraparib Monotherapy as Maintain and Recurrent Treatment of Endometrial Serous Carcinoma **PARP**

LOCATIONS: Jinan (China)

Federation)

NCT02264678 **PHASE 1/2**

Ascending Doses of AZD6738 in Combination With Chemotherapy and/or Novel Anti Cancer Agents **TARGETS**

ATR, PARP, PD-L1

LOCATIONS: Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Cambridge (United Kingdom), Withington (United Kingdom), London (United Kingdom), Sutton (United Kingdom), Villejuif (France), Saint Herblain (France), California

NCT04635631 **PHASE 1** STUDY OF TALAZOPARIB MONOTHERAPY IN CHINESE PARTICIPANTS WITH ADVANCED SOLID **TARGETS TUMORS PARP**

LOCATIONS: Beijing (China), Changchun (China)



NCT03772561	PHASE 1
Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies	t TARGETS PARP, AKTs, PD-L1
LOCATIONS: Singapore (Singapore)	
NCT04801966	PHASE NULL
Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study	TARGETS CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF
LOCATIONS: Melbourne (Australia)	
NCT04497116	PHASE 1/2
Study of RP-3500 in Advanced Solid Tumors	TARGETS ATR, PARP
LOCATIONS: Copenhagen (Denmark), Newcastle Upon Tyne (United Kingdom), Manchester (United Canada), Massachusetts, Rhode Island, New York, Tennessee, Texas	ited Kingdom), London (United Kingdom), Toronto
NCT04159155	PHASE 2/3
A Study of Various Treatments in Serous or p53 Abnormal Endometrial Cancer	TARGETS PARP
LOCATIONS: Toronto (Canada)	



CLINICAL TRIALS

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RATIONALE

Inactivating TSC1 alterations may lead to increased mTOR activation and predict sensitivity

to mTOR inhibitors.

ALTERATION 0739*

NCT03239015	PHASE 2
Efficacy and Safety of Targeted Precision Therapy in Refractory Tumor With Druggable Molecular Event	TARGETS EGFR, ERBB2, ERBB4, PARP, mTOR, MET, RET, ROS1, VEGFRS, BRAF, CDK4 CDK6
LOCATIONS: Shanghai (China)	
NCT04337463	PHASE NULL
ATG-008 Combined With Toripalimab in Advanced Solid Tumors	TARGETS mTORC1, mTORC2, PD-1
LOCATIONS: Chongqing (China), Chengdu (China)	
NCT04803318	PHASE 2
Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors	TARGETS mTOR, FGFRs, KIT, PDGFRA, RET, VEGFRs, MEK
LOCATIONS: Guangzhou (China)	
NCT02693535	PHASE 2
TAPUR: Testing the Use of Food and Drug Administration (FDA) Approved Drugs That Target a Specific Abnormality in a Tumor Gene in People With Advanced Stage Cancer	TARGETS VEGFRs, ABL, SRC, ALK, AXL, MET, ROS1, TRKA, TRKC, CDK4, CDK6, CSF1R, FLT3, KIT, RET, mTOR, EGFR, ERBB2, ERBB3, MEK, BRAF, SMO, DDR2, PARP, PD-1, CTLA-4, ERBB4
LOCATIONS: Hawaii, Washington, Oregon, California	
NCT03297606	PHASE 2
Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)	TARGETS VEGFRS, ABL, SRC, ALK, AXL, MET, ROS1, TRKA, TRKC, DDR2, KIT, EGFR, PD-1, CTLA-4, PARP, CDK4, CDK6, CSF1R, FLT3, RET, mTOR, ERBB2, ERBB3, MEK, BRAF, SMO

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Kingston (Canada), London (Canada)



NCT02397083	PHASE 2
Phase II Study of Intrauterine Device (IUD) Alone or in Combination With Everolimus in Endometrial Cancer	TARGETS mTOR
LOCATIONS: Colorado, Michigan, Virginia, Texas	
NCT03008408	PHASE 2
Phase II Ribociclib, Everolimus and Letrozole in Endometrial Cancer	TARGETS Aromatase, mTOR, CDK4, CDK6
LOCATIONS: Texas	
NCT03217669	PHASE 1
Epacadostat (INCB24360) in Combination With Sirolimus in Advanced Malignancy	TARGETS IDO1, mTOR
LOCATIONS: Kansas	
NCT03065062	PHASE 1
Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head & Neck and Other Solid Tumors	TARGETS PI3K-alpha, PI3K-gamma, mTORC1, mTORC2, CDK4, CDK6
LOCATIONS: Massachusetts	
NCT01582191	PHASE 1
A Phase 1 Trial of Vandetanib (a Multi-kinase Inhibitor of EGFR, VEGFR and RET Inhibitor) in Combination With Everolimus (an mTOR Inhibitor) in Advanced Cancer	TARGETS mTOR, EGFR, RET, SRC, VEGFRs
LOCATIONS: Texas	



APPENDIX

Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

ABL1	AKT3	ALK	APC L2136I, L23F and L2574I
V43fs*4	L277R, R241W, V419A and splice site 697-2A>C	N285Y	
AR A871V, D551E and Y364D	ARID1A G851D	ASXL1 E824D, G1401E and R394H	ATM E2052K and R493C
ATR E254D, L108P, R1300C, R635Q and S1701Y	ATRX D1875Y, K467Q, R701H and T480R	BAP1 D608N	BCOR E27K, L1334V and V293I
BCORL1 R1226H, S244L and S663Y	BRAF R389C	BRCA1 D749Y	BRCA2 E1455_E1456>DK, E2020K, L3317R, N1316H and T3414A
BRD4	BRIP1	BTK	C11ORF30 (EMSY)
S499L	L892I and R356Q	L23I, R13*, R288W and T33I	E924K and S335N
CARD11 E420D and P567L	CASP8	CBFB	CBL
	E311D	A99T	E394G and R462Q
CCND2	CDK12	CDK6	CDK8 N372S, V78I, splice site 1110+1G>T and splice site 205-1G>A
K71T	K726T	R214C	
CHEK1	CREBBP	CRKL	CTCF
F70L	G211E and T872M	A208T and F134V	K208N
CTNNA1 K867T	CUL3 F224C and F377C	DAXX K662N and Q469H	DIS3 E117D and R108H
DNMT3A D702N	DOT1L S631L	EED L425F	EP300 E1531G, L68I and M205R
EPHA3	EPHB1 E116D, R663W and V234A	ERBB2	ERBB3
E647* and V626A		R1161Q	H578N
ERBB4 A1219T	ERG F430C	ESR1 D218N	FAM123B K170Q and S286N
FANCG	FAS	FGF12	FGF23
V64A and Y268H	R279C	R224M and Y155D	A213V
FGF3	FGF4	FGFR4	FH P461T
R151I and S108L	G37D	L113F	



APPENDIX

Variants of Unknown Significance

FLT1 F333Y and M945fs*15	FLT3 F710V	GABRA6 L424I and Y233C	GATA6 E460K and N510D
GNAS A475V, F315C and L132M	GSK3B A22T and R92I	HGF L565I and P299H	HSD3B1 L167M
IDH1 R222C	IDH2 D200Y	IGF1R E83K	IKBKE E705K
IKZF1 K275N, S105P and S364L	INPP4B E807D, L277V and L344R	IRS2 D276N, E1199K, P813S and S1174L	JAK1 R93C
JAK2 R803Q	KDM5A E1098*, R893Q and R902W	KDM5C K101N	KDM6A R519G
KDR E685K, G335D, R652I and V1251fs*5	KEAP1 R614Q	KEL A311T, D497N, E352D and Y714H	KIT R956W
KLHL6 N200H	KMT2A (MLL) I3660V, L950I, S941Y and T3574M	LTK K827T and S830N	MAF N365K
MAP2K4 R304Q	MAP3K1 R532Q	MED12 L1300M, R422W, R438C and R470H	MERTK D579Y and P81S
MET R1166*	MITF splice site 760-2A>C	MLH1 D63Y	MLL2 A781T, E1588K and L3776R
MSH6 A780T, E995K, K431T and R1242H	MST1R F574L and G356D	MTOR Q2223R	NBN E474D
NF1 K63N, L2639I and W837L	NF2 R187I and R418C	NKX2-1 D356G	NOTCH1 A105T, D464N and G2333D
NRAS Y4C	NSD3 (WHSC1L1) G748V	NT5C2 K244E and K45N	NTRK1 M566T
NTRK2 E70K and R124*	NTRK3 E559G, P725H and R721C	P2RY8 S58L	PALB2 K279N
PARP1 A194V	PARP2 E222Q	PBRM1 R876C	PDCD1 (PD-1) A50T
PDGFRA E556*, M1073V, R346S and splice site 1122-1G>T	PDK1 K73N and R341I	PIK3CA L285V, R230* and splice site 352+1G>A	PIK3CB A655V, E225*, N377H and R955*



APPENDIX

Variants of Unknown Significance

POLD1 R177C	POLE L12351	PPP2R2A E338A	PTCH1 L223I
PTEN E157D and I5T	PTPN11	PTPRO	RAD51
	K590T	L832R	L2741
RAD51B	RAD51C	RAD52	RAF1
L321R	S163R	R55H	E104K
RBM10 E705D	REL R45Q	RET R474W	RICTOR H1453R, K1155T, L612R, L804I, R1110C and R222Q
ROS1 E402D, E908*, I883L, K1533N, N399H, R1212C, R922I and V785M	SETD2 P190L	SF3B1 E809* and R451Q	SGK1 L363I
SMAD2	SPEN	SPOP	STAG2 D153N and F1214L
S276L and S464A	R535Q	R284C	
STAT3 T494fs*55	TBX3 A418T, E720K, R304W and S363L	TEK splice site 3063-1G>A	TET2 A241V and Y1935H
TNFAIP3	TSC1	TSC2	U2AF1
H289Q	K475Q, L930P and R228Q	D8Y and K69N	R234C

VEGFA

P363S and S186Y



APPENDIX

Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTK	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL	CCND1
CCND2	CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2	DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	ЕРНАЗ	ЕРНВ1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRFI1	ESR1	EZH2
FAM46C	FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3
FGFR4	FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3
GATA4	GATA6	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3F3A
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDM5A	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3	P2RY8	PALB2
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA
PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET
RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2
SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOCS1
SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU
SYK	TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53
TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1	WT1	XPO1
XRCC2	ZNF217	ZNF703						
DNA GENE LIS	T: FOR THE DETE	CTION OF SELECT	T REARRANGE	MENTS				
ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSPO2	SDC4	SLC34A2	TERC*	TERT**	TMPRSS2

^{*}TERC is an NCRNA

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Loss of Heterozygosity (LOH) score Microsatellite (MS) status Tumor Mutational Burden (TMB)

^{**}Promoter region of TERT is interrogated



APPENDIX

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.

ABOUT FOUNDATIONONE CDX

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details: www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffinembedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLES

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture–selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g.,gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials Ranking of Therapies in Summary Table
Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials
Pediatric trial qualification → Geographical
proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK* (NCCN*) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2021. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

- 1. The MSI-H/MSS designation by FMI F1CDx test is based on genome wide analysis of 95 microsatellite loci and not based on the 5 or 7 MSI loci described in current clinical practice guidelines. The threshold for MSI-H/MSS was determined by analytical concordance to comparator assays (IHC and PCR) using uterine, cecum and colorectal cancer FFPE tissue. The clinical validity of the qualitative MSI designation has not been established. For Microsatellite Instability (MSI) results, confirmatory testing using a validated orthogonal method should be considered.
- 2. TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics



APPENDIX

About FoundationOne®CDx

of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh_docs/ pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.

- 3. The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding armand chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
- 4. Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
- 5. Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine whether the patient is a candidate for biopsy.
- 6. Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive,

and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in

https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/ report the frequency of HER2 overexpression as 20% in breast cancer. Based on the F1CDx HER2 CDx concordance study, approximately 10% of HER2 amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*	
Repeatability	5.11 - 10.40	
Reproducibility	5.95 - 12.31	
INDELS	%CV*	
INDELS Repeatability	%CV*	

^{*}Interquartile Range = 1st Quartile to 3rd Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of followup germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are ATM, BAP1, BRCA1, BRCA2, BRIP1, CHEK2, FH, FLCN, MLH1, MSH2, MSH6, MUTYH, PALB2, PMS2, POLE,

RAD51C, RAD51D, RET, SDHA, SDHB, SDHC, SDHD, TSC2, and VHL, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are ASXL1, CBL, DNMT3A, IDH2, JAK2, KMT2D (MLL2), MPL, MYD88, SF3B1, TET2, and U2AF1 and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating



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physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
muts/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
os	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
ткі	Tyrosine kinase inhibitor

MR Suite Version 5.0.0

The median exon coverage for this sample is 994x

APPENDIX

References

- 1. Samstein RM, et al. Nat. Genet. (2019) pmid: 30643254
- 2. Goodman AM, et al. Mol. Cancer Ther. (2017) pmid: 28835386
- Goodman AM, et al. Cancer Immunol Res (2019) pmid: 31405947
- 4. Cristescu R, et al. Science (2018) pmid: 30309915
- 5. Ready N, et al. J. Clin. Oncol. (2019) pmid: 30785829
- Hellmann MD, et al. N. Engl. J. Med. (2018) pmid: 29658845
- 7. Hellmann MD, et al. Cancer Cell (2018) pmid: 29657128
- 8. Hellmann MD, et al. Cancer Cell (2018) pmid: 29731394
- 9. Rozeman EA, et al. Nat Med (2021) pmid: 33558721
- 10. Sharma P, et al. Cancer Cell (2020) pmid: 32916128
- Marabelle A, et al. Lancet Oncol. (2020) pmid: 32919526
- 12. Legrand et al., 2018; ASCO Abstract 12000
- Chalmers ZR, et al. Genome Med (2017) pmid: 28420421
- 14. Santin et al., 2016; ASCO Abstract 5591
- 15. Mehnert JM, et al. J. Clin. Invest. (2016) pmid: 27159395
- Cancer Genome Atlas Research Network, et al. Nature (2013) pmid: 23636398
- 17. Hussein YR, et al. Mod. Pathol. (2015) pmid: 25394778
- Church DN, et al. Hum. Mol. Genet. (2013) pmid: 23528559
- 19. Shao C, et al. JAMA Netw Open (2020) pmid: 33119110
- **20.** Pfeifer GP, et al. Mutat. Res. (2005) pmid: 15748635
- 21. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) pmid: 23875803
- 22. Pfeifer GP, et al. Oncogene (2002) pmid: 12379884
- 23. Rizvi NA, et al. Science (2015) pmid: 25765070
- 24. Johnson BE, et al. Science (2014) pmid: 24336570
- 25. Choi S, et al. Neuro-oncology (2018) pmid: 29452419
- **26.** Briggs S, et al. J. Pathol. (2013) pmid: 23447401
- Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) pmid: 24583393
- 28. Nature (2012) pmid: 22810696
- 29. Roberts SA, et al. Nat. Rev. Cancer (2014) pmid: 25568919
- 30. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) pmid: 25392179
- **31.** Kroemer G, et al. Oncoimmunology (2015) pmid: 26140250
- **32.** Lal N, et al. Oncoimmunology (2015) pmid: 25949894
- 33. Le DT, et al. N. Engl. J. Med. (2015) pmid: 26028255
- **34.** Ayers et al., 2016; ASCO-SITC Abstract P60
- **35.** Black D, et al. J. Clin. Oncol. (2006) pmid: 16549821
- **36.** Mackay HJ, et al. Eur. J. Cancer (2010) pmid: 20304627
- Kanopienė D, et al. Medicina (Kaunas) (2014) pmid: 25458958
- **38.** Hampel H, et al. Cancer Res. (2006) pmid: 16885385
- **39.** Steinbakk A, et al. Cell Oncol (Dordr) (2011) pmid: 21547578
- Bilbao C, et al. Int. J. Radiat. Oncol. Biol. Phys. (2010) pmid: 20005452
- 41. Zighelboim I, et al. J. Clin. Oncol. (2007) pmid: 17513808
- **42.** Bilbao-Sieyro C, et al. Oncotarget (2014) pmid: 25026289
- **43.** Arabi H, et al. Gynecol. Oncol. (2009) pmid: 19275958
- **44.** Stelloo E, et al. Clin. Cancer Res. (2016) pmid: 27006490
- 45. Nout RA, et al. Gynecol. Oncol. (2012) pmid: 22609107
- 46. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) pmid: 26337942
- **47**. You JF, et al. Br. J. Cancer (2010) pmid: 21081928
- 48. Bairwa NK, et al. Methods Mol. Biol. (2014) pmid:

- 24623249
- 49. Boland CR, et al. Cancer Res. (1998) pmid: 9823339
- 50. Pawlik TM, et al. Dis. Markers (2004) pmid: 15528785
- 51. Boland CR, et al. Gastroenterology (2010) pmid: 20420947
- 52. Michels J, et al. Oncogene (2014) pmid: 24037533
- 53. Mateo J, et al. N. Engl. J. Med. (2015) pmid: 26510020
- **54.** Mateo J, et al. Lancet Oncol. (2019) pmid: 31806540
- Abida W, et al. Clin. Cancer Res. (2020) pmid: 32086346
- 56. Bang YJ, et al. J. Clin. Oncol. (2015) pmid: 26282658
- 57. Gruber et al., 2019; ASCO Abstract 3006
- 58. Olson D, et al. Clin Genitourin Cancer (2016) pmid: 27079472
- 59. Piha-Paul et al., 2018; AACR-NCI-EORTC Abstract A096
- 60. Weston VJ, et al. Blood (2010) pmid: 20739657
- **61.** Williamson CT, et al. Mol. Cancer Ther. (2010) pmid: 20124459
- **62.** Gilardini Montani MS, et al. J. Exp. Clin. Cancer Res. (2013) pmid: 24252502
- **63.** Bryant HE, et al. Nucleic Acids Res. (2006) pmid: 16556909
- **64.** Ihnen M, et al. Mol. Cancer Ther. (2013) pmid: 23729402
- Williamson CT, et al. EMBO Mol Med (2012) pmid: 22416035
- 66. Kubota E, et al. Cell Cycle (2014) pmid: 24841718
- **67.** Huehls AM, et al. Mol. Pharmacol. (2012) pmid: 22833573
- 68. O'Carrigan et al., 2016; ASCO Abstract 2504
- **69.** Yap TA, et al. Cancer Discov (2021) pmid: 32988960
- **70.** Menezes DL, et al. Mol. Cancer Res. (2015) pmid: 25232030
- 71. Vendetti FP, et al. Oncotarget (2015) pmid: 26517239
- 72. Min A, et al. Mol. Cancer Ther. (2017) pmid: 28138034
- 73. Kwok M, et al. Blood (2016) pmid: 26563132
- 74. Riabinska A, et al. Sci Transl Med (2013) pmid: 23761041
- 75. Cerami E, et al. Cancer Discov (2012) pmid: 22588877
- **76.** Gao J, et al. Sci Signal (2013) pmid: 23550210
- 77. Tate JG, et al. Nucleic Acids Res. (2019) pmid: 30371878
- 78. Tsuda H, et al. Oncology (2003) pmid: 12931023
- Suehiro Y, et al. Clin. Cancer Res. (2008) pmid: 18519763
- 80. Shan W, et al. Tumour Biol. (2015) pmid: 25608836
- 81. Mhawech-Fauceglia P, et al. Gynecol. Oncol. (2014) pmid: 24508840
- 82. Shiloh Y, et al. Nat. Rev. Mol. Cell Biol. (2013) pmid: 23847781
- 83. Cremona CA, et al. Oncogene (2014) pmid: 23851492
- 84. Jiang X. et al. J. Biol. Chem. (2006) pmid: 16603769
- 85. Fernandes N, et al. J. Biol. Chem. (2005) pmid: 15713674
- 86. Scott SP, et al. Proc. Natl. Acad. Sci. U.S.A. (2002) pmid: 11805335
- 87. Landrum MJ, et al. Nucleic Acids Res. (2018) pmid: 29165669
- 88. van Os NJ, et al. Clin Genet (2016) pmid: 26662178
- 89. Rothblum-Oviatt C, et al. Orphanet J Rare Dis (2016) pmid: 27884168
- **90.** Jaiswal S, et al. N. Engl. J. Med. (2014) pmid: 25426837
- 91. Genovese G, et al. N. Engl. J. Med. (2014) pmid: 25426838
- 92. Xie M, et al. Nat. Med. (2014) pmid: 25326804
- 93. Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) pmid: 28669404
- 94. Severson EA, et al. Blood (2018) pmid: 29678827
- 95. Fuster JJ, et al. Circ. Res. (2018) pmid: 29420212
- 96. Chabon JJ, et al. Nature (2020) pmid: 32269342

- 97. Razavi P, et al. Nat. Med. (2019) pmid: 31768066
- 98. Kaufman B, et al. J. Clin. Oncol. (2015) pmid: 25366685
- 99. Tutt A, et al. Lancet (2010) pmid: 20609467
- 100. Robson M, et al. N. Engl. J. Med. (2017) pmid: 28578601
- **101.** Mirza MR, et al. N. Engl. J. Med. (2016) pmid: 27717299
- 102. Sandhu SK, et al. Lancet Oncol. (2013) pmid: 23810788
- 103. Swisher EM, et al. Lancet Oncol. (2017) pmid: 27908594104. Drew Y, et al. Br. J. Cancer (2016) pmid: 27002934
- **105.** Pujade-Lauraine E, et al. Lancet Oncol. (2017) pmid: 28754483
- **106.** Ledermann JA, et al. Lancet Oncol. (2016) pmid: 27617661
- **107.** Fong PC, et al. N. Engl. J. Med. (2009) pmid: 19553641
- 108. Audeh MW, et al. Lancet (2010) pmid: 20609468
- 109. Fong PC, et al. J. Clin. Oncol. (2010) pmid: 20406929
- 110. Gelmon KA, et al. Lancet Oncol. (2011) pmid: 21862407
- 111. Kaye SB, et al. J. Clin. Oncol. (2012) pmid: 22203755
- 112. Domchek SM, et al. Gynecol. Oncol. (2016) pmid:
- 113. Moore K, et al. N. Engl. J. Med. (2018) pmid: 30345884
- 114. Golan T, et al. N. Engl. J. Med. (2019) pmid: 31157963
- 115. Thomas A, et al. J. Clin. Oncol. (2018) pmid: 29252124
- 116. Saito YD, et al. Cancer Treat Res Commun (2018) pmid: 31299005
- 117. Pouliot GP, et al. PLoS ONE (2019) pmid: 31721781
- 118. Kim H, et al. Clin. Cancer Res. (2017) pmid: 27993965
- 119. Jin J, et al. Neoplasia (2018) pmid: 29605721
- 120. Westin et al., 2021; ASCO Abstract 5505
- 121. Do K. et al. J. Clin. Oncol. (2015) pmid: 25964244
- 122. Isakoff SJ, et al. J. Clin. Oncol. (2015) pmid: 25847936
- 123. Dann RB, et al. Gynecol. Oncol. (2012) pmid: 23406760
- **124.** Evers B, et al. Clin. Cancer Res. (2008) pmid: 18559613
- 125. Soares DG, et al. Proc. Natl. Acad. Sci. U.S.A. (2007) pmid: 17656556
- 126. Miolo G, et al. Cancer Biol. Ther. (2016) pmid: 27561088
- 127. Ghouadni A, et al. Breast (2017) pmid: 28467918
- 128. Case Rep Oncol () pmid: 28626402
- **129.** Monk BJ, et al. Gynecol. Oncol. (2020) pmid: 31924332 **130.** Harnicek D, et al. Int. J. Cancer (2016) pmid: 26933761
- 131. Cruz C, et al. J. Clin. Oncol. (2018) pmid: 30240327
- 132. Poveda A, et al. Ann. Oncol. (2017) pmid: 28368437
- 133. de Jonge MM, et al. Clin. Cancer Res. (2019) pmid: 31492746
- 134. Levine DA, et al. Gynecol. Oncol. (2001) pmid: 11263938
- 135. Segev Y, et al. Gynecol. Oncol. (2013) pmid: 23562522
- 136. Yang H, et al. Science (2002) pmid: 12228710137. Nat. Struct. Mol. Biol. (2011) pmid: 21731065
- 138. Miki Y. et al. Science (1994) pmid: 7545954
- 139. Wooster R, et al. Nature () pmid: 8524414
- 140. King MC, et al. Science (2003) pmid: 14576434141. MedGenMed (2005) pmid: 16369438
- 142. Ford D, et al. Lancet (1994) pmid: 7907678
- 142. Ford D, et al. Lancet (1994) pmid: 7907678

 143. Whittemore AS, et al. Am. J. Hum. Genet. (1997) pmid:
- 9042908 144. Claus EB, et al. Cancer (1996) pmid: 8635102
- 145. Struewing JP, et al. N. Engl. J. Med. (1997) pmid: 9145676
- 146. Oddoux C, et al. Nat. Genet. (1996) pmid: 8841192147. Hall MJ, et al. Cancer (2009) pmid: 19241424
- **148.** Mao JH, et al. Science (2008) pmid: 18787170
- 149. Yang H, et al. Oncotarget (2015) pmid: 25749036150. Villaruz LC, et al. Lung Cancer (2014) pmid: 24360397

 Kulkarni et al., 2020; https://doi.org/10.1016/ j.ygyno.2020.05.244

APPENDIX

References

- 152. Wertz IE, et al. Nature (2011) pmid: 21368834
- 153. Zhao S, et al. Proc. Natl. Acad. Sci. U.S.A. (2013) pmid: 23359684
- 154. Le Gallo M, et al. Nat. Genet. (2012) pmid: 23104009
- 155. Kuhn E, et al. J. Natl. Cancer Inst. (2012) pmid: 22923510
- 156. Spruck CH, et al. Cancer Res. (2002) pmid: 12183400
- 157. Cassia R, et al. J. Pathol. (2003) pmid: 14648662
- 158. Garcia-Dios DA, et al. Gynecol. Oncol. (2013) pmid: 23219661
- 159. Tu K, et al. Hepatol. Res. (2012) pmid: 22548670
- 160. Iwatsuki M, et al. Int. J. Cancer (2010) pmid: 19739118
- 161. Yokobori T, et al. Int. J. Oncol. (2012) pmid: 22576686
- 162. Yokobori T, et al. Cancer Res. (2009) pmid: 19366810
- 163. Yokobori T, et al. Mol. Cancer Res. (2014) pmid: 24165483
- 164. Rajagopalan H, et al. Nature (2004) pmid: 14999283
- 165. Cheng Y, et al. J. Invest. Dermatol. (2013) pmid:
- 166. Xu Y. et al. Biomarkers (2016) pmid: 26954701
- 167. Welcker M, et al. Nat. Rev. Cancer (2008) pmid: 18094723
- 168. Akhoondi S, et al. Cancer Res. (2007) pmid: 17909001
- 169. Dombi E, et al. N. Engl. J. Med. (2016) pmid: 28029918
- 170. Schalkwijk S, et al. Cancer Chemother Pharmacol (2021) pmid: 33903938
- 171. Toledano H, et al. Childs Nerv Syst (2021) pmid:
- 172. Ronsley R. et al. Cancer Med (2021) pmid: 33939292
- 173. Fangusaro J, et al. Lancet Oncol. (2019) pmid: 31151904
- 174. Manoharan N, et al. J Neurooncol (2020) pmid:
- 175. Kondyli M, et al. J Neurooncol (2018) pmid: 30097824
- 176. Awada G, et al. Case Rep Oncol () pmid: 33082744
- 177. Middleton G. et al. Nature (2020) pmid: 32669708
- 178. Lim SM, et al. Oncotarget (2016) pmid: 26859683
- 179. Weiss B, et al. Neuro-oncology (2015) pmid: 25314964
- 180. Janku F, et al. Oncotarget (2014) pmid: 24931142
- 181. Johannessen CM, et al. Curr. Biol. (2008) pmid: 18164202
- 182. Johannessen CM, et al. Proc. Natl. Acad. Sci. U.S.A. (2005) pmid: 15937108
- 183. Malone CF, et al. Cancer Discov (2014) pmid: 24913553
- 184. Tolcher AW, et al. Ann. Oncol. (2015) pmid: 25344362
- 185. Patterson et al., 2018; AACR Abstract 3891
- 186. Myers AP, et al. Gynecol, Oncol. (2016) pmid: 27016228 187. Soumerai TE, et al. Clin. Cancer Res. (2018) pmid:
- 30068706 188. Murayama-Hosokawa S, et al. Oncogene (2010) pmid: 20062086
- 189. Hattori S, et al. Biochem. Biophys. Res. Commun. (1991) pmid: 1904223
- 190. Morcos P, et al. Mol. Cell. Biol. (1996) pmid: 8628317
- 191. Ballester R, et al. Cell (1990) pmid: 2121371
- 192. Xu GF, et al. Cell (1990) pmid: 2116237
- 193. Martin GA, et al. Cell (1990) pmid: 2121370
- 194. Thomas L. et al. Hum. Mutat. (2012) pmid: 22807134
- Skuse GR, et al. Hum. Mol. Genet. (1997) pmid: 195. 9300663
- 196. Messiaen LM, et al. Genet. Med. () pmid: 11258625
- 197. Ars E, et al. Hum. Mol. Genet. (2000) pmid: 10607834
- 198. Messiaen LM, et al. J. Med. Genet. (2005) pmid: 15863657
- 199. Poullet P, et al. Mol. Cell. Biol. (1994) pmid: 8264648
- 200. Jett K, et al. Genet. Med. (2010) pmid: 20027112
- 201. Patil S, et al. Oncologist (2012) pmid: 22240541
- 202. Evans DG, et al. Clin Sarcoma Res (2012) pmid:

- 23036231
- 203. Upadhyaya M, et al. J. Med. Genet. (1995) pmid: 8544190
- 204. Williams VC, et al. Pediatrics (2009) pmid: 19117870
- 205. Fritsch C, et al. Mol. Cancer Ther. (2014) pmid: 24608574
- 206. Juric D, et al. J. Clin. Oncol. (2018) pmid: 29401002 207. Gallant JN, et al. NPJ Precis Oncol (2019) pmid:
- 30793038 208. André F, et al. N. Engl. J. Med. (2019) pmid: 31091374
- 209. Smyth LM, et al. NPJ Breast Cancer (2021) pmid: 33863913
- 210. Park HS, et al. PLoS ONE (2016) pmid: 27105424
- 211. Hou MM, et al. Oncotarget (2014) pmid: 25426553
- 212. Varnier R. et al. Eur J Cancer (2019) pmid: 31351267
- 213. Janku F, et al. Cell Rep (2014) pmid: 24440717
- 214. Moroney J, et al. Clin. Cancer Res. (2012) pmid: 22927482
- 215. Basho RK, et al. JAMA Oncol (2017) pmid: 27893038
- 216. Moroney JW, et al. Clin. Cancer Res. (2011) pmid: 21890452
- 217. Krop et al., 2018; ASCO Abstract 101
- 218. Dolly SO, et al. Clin. Cancer Res. (2016) pmid: 26787751
- 219. Aust Fam Physician (1986) pmid: 2941002
- 220. Santin AD, et al. Gynecol Oncol Rep (2020) pmid: 31934607
- 221. Patnaik A, et al. Ann. Oncol. (2016) pmid: 27672108
- 222. Esteva FJ, et al. Am. J. Pathol. (2010) pmid: 20813970
- 223. Baselga J, et al. J. Clin. Oncol. (2014) pmid: 25332247
- 224. Chakrabarty A. et al. Oncogene (2010) pmid: 20581867
- 225. Kataoka Y, et al. Ann. Oncol. (2010) pmid: 19633047 226. Wang L. et al. BMC Cancer (2011) pmid: 21676217
- 227. Rudd ML, et al. Clin. Cancer Res. (2011) pmid: 21266528
- 228. Oda K, et al. Cancer Res. (2005) pmid: 16322209
- 229. Oda K, et al. Cancer Res. (2008) pmid: 18829572
- 230. Akiyama-Abe A, et al. Br. J. Cancer (2013) pmid:
- 231. McIntyre JB, et al. Gynecol. Oncol. (2014) pmid: 24262879
- 232. Samuels Y, et al. Cancer Cell (2005) pmid: 15950905
- 233. Nat. Rev. Cancer (2009) pmid: 19629070
- **234.** Kang S, et al. Proc. Natl. Acad. Sci. U.S.A. (2005) pmid: 15647370
- 235. Ikenoue T, et al. Cancer Res. (2005) pmid: 15930273
- 236. Gymnopoulos M. et al. Proc. Natl. Acad. Sci. U.S.A. (2007) pmid: 17376864
- 237. Horn S, et al. Oncogene (2008) pmid: 18317450
- 238. Hon WC, et al. Oncogene (2012) pmid: 22120714
- 239. Burke JE, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) pmid: 22949682
- 240. Wu H, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) pmid: 19915146
- 241. Laurenti R, et al. Rev Saude Publica (1990) pmid: 2103068
- 242. Dan S, et al. Cancer Res. (2010) pmid: 20530683
- **243.** Zhao L, et al. Oncogene (2008) pmid: 18794883
- 244. Lui VW, et al. Cancer Discov (2013) pmid: 23619167
- 245. Ross RL, et al. Oncogene (2013) pmid: 22430209
- 246. Rivière JB, et al. Nat. Genet. (2012) pmid: 22729224
- 247. Shibata T, et al. Cancer Lett. (2009) pmid: 19394761
- 248. Dogruluk T, et al. Cancer Res. (2015) pmid: 26627007
- 249. Croessmann S, et al. Clin. Cancer Res. (2018) pmid: 29284706
- 250. Ng PK, et al. Cancer Cell (2018) pmid: 29533785
- 251. Spangle JM, et al. (2020) pmid: 32929011
- 252. Chen L, et al. Nat Commun (2018) pmid: 29636477

- 253. Akgumus G, et al. J Mol Diagn (2017) pmid: 28502730
- 254. Slomovitz BM, et al. J. Clin. Oncol. (2015) pmid:
- 255. Al-Rohil RN, et al. Cancer (2016) pmid: 26479420
- 256. Tanwar PS, et al. Biol. Reprod. (2009) pmid: 19403928
- 257. Tanwar PS, et al. PLoS ONE (2011) pmid: 21695255
- 258. Fujishita T, et al. Proc. Natl. Acad. Sci. U.S.A. (2008) pmid: 18768809
- 259. Bhoori S, et al. J. Hepatol. (2010) pmid: 20347502
- 260. Niida A, et al. Oncogene (2004) pmid: 15378020
- 261. Chamorro MN, et al. EMBO J. (2005) pmid: 15592430
- 262. Kagey MH, et al. Br. J. Pharmacol. (2017) pmid: 28574171
- 263. Kagey et al., 2017; AACR Abstract 369
- 264. Kwon C, et al. Nat. Cell Biol. (2011) pmid: 21841793
- 265. Arcaroli JJ, et al. Br. J. Cancer (2013) pmid: 23868008
- 266. Shang H, et al. Cancer (2015) pmid: 26349011
- 267. Kode A, et al. Nature (2014) pmid: 24429522
- 268. Kummar et al., 2015: ASCO Abstract 10563
- Messersmith WA, et al. Clin. Cancer Res. (2015) pmid: 25231399
- 270. Zhu J, et al. Carcinogenesis (2012) pmid: 22964660
- 271. Kogan Y, et al. Biochem. J. (2012) pmid: 22356261
- 272. Lachenmayer A, et al. Clin. Cancer Res. (2012) pmid:
- 273. Dellinger TH, et al. Expert Rev Anticancer Ther (2012) pmid: 22149432
- 274. Saegusa M, et al. Br. J. Cancer (2001) pmid: 11161379
- 275. McConechy MK, et al. Mod. Pathol. (2014) pmid:
- 276. Huang M, et al. Cancer (2013) pmid: 23760948
- 277. Yeramian A, et al. Oncogene (2013) pmid: 22430211
- 278. Machin P, et al. Hum. Pathol. (2002) pmid: 11957146
- 279. Peiró G, et al. Hum. Pathol. (2013) pmid: 22955108 280. Kurnit KC, et al. Mod. Pathol. (2017) pmid: 28281553
- 281. Liu Y, et al. J. Natl. Cancer Inst. (2014) pmid: 25214561
- 282. Rosen DG, et al. Mod. Pathol. (2010) pmid: 19820688
- 283. Athanassiadou P, et al. Int. J. Gynecol. Cancer () pmid: 17504383
- 284. Biochem. Biophys. Res. Commun. (2000) pmid: 10679188
- 285. O'Malley et al., 2017; AACR-NCI-EORTC Abstract LB-A12
- 286. Dougherty et al., 2014; ASCO Abstract 5536
- 287. Qu X, et al. Blood (2019) pmid: 30446494
- 288. Pace P, et al. EMBO J. (2002) pmid: 12093742 289. Deakyne JS, et al. Biochemistry Mosc. (2011) pmid:
- 21568838
- 290. Alpi A, et al. Mol. Cell. Biol. (2007) pmid: 17938197
- 291. Alpi AF, et al. Mol. Cell (2008) pmid: 19111657
- 292. Meetei AR, et al. Nat. Genet. (2003) pmid: 12973351 293. Machida YJ, et al. Mol. Cell (2006) pmid: 16916645
- 294. Hodson C. et al. J. Biol. Chem. (2011) pmid: 21775430 295. Hodson C, et al. Structure (2014) pmid: 24389026
- 296. Seki S, et al. Genes Cells (2007) pmid: 17352736
- 297. Ali AM, et al. Hum. Mutat. (2009) pmid: 19405097 298. Fostira F, et al. Breast Cancer Res. Treat. (2018) pmid:
- 29335925
- 299. Hart SN, et al. BMJ Open (2016) pmid: 27084275 300. Del Valle J, et al. Cancers (Basel) (2020) pmid: 32235514
- 301. Chandrasekharappa SC, et al. Blood (2013) pmid: 23613520
- 302. Dicks E, et al. Oncotarget (2017) pmid: 28881617
- 303. Tabernero J, et al. J. Clin. Oncol. (2015) pmid: 26324363 304. Krook MA, et al. Cold Spring Harb Mol Case Stud (2019) pmid: 31371345



APPENDIX

References

- **305.** Abou-Alfa GK, et al. Lancet Oncol. (2020) pmid: 32203698
- **306.** Nogova L, et al. J. Clin. Oncol. (2017) pmid: 27870574
- 307. Morizane et al., 2020; ASCO GI Abstract 538
- 308. Pearson A, et al. Cancer Discov (2016) pmid: 27179038
- **309.** Van Cutsem E, et al. Ann. Oncol. (2017) pmid: 29177434
- **310.** Aggarwal C, et al. J Thorac Oncol (2019) pmid: 31195180
- 311. Voss MH, et al. Clin. Cancer Res. (2019) pmid: 30745300 312. Cleary JM, et al. Cancer Discov (2021) pmid: 33926920
- 313. Schuler M, et al. Lancet Oncol. (2019) pmid: 31405822
- **314.** Goyal L, et al. Cancer Discov (2019) pmid: 31109923
- 315. Mazzaferro V. et al. Br. J. Cancer (2019) pmid: 30420614
- **316.** Borad MJ, et al. PLoS Genet. (2014) pmid: 24550739
- 317. Liao RG, et al. Cancer Res. (2013) pmid: 23786770
- **318.** Gozgit JM, et al. Mol. Cancer Ther. (2012) pmid: 22238366
- 319. Chae et al., 2018; ASCO Abstract 2503
- 320. Soria et al., 2017; ASCO Abstract 4074
- 321. Slosberg ED, et al. Oncotarget (2018) pmid: 29765547
- 322. Yue et al., 2016; AMP Abstract S06
- 323. Krakstad C, et al. PLoS ONE (2012) pmid: 23300780
- **324.** Dutt A, et al. Proc. Natl. Acad. Sci. U.S.A. (2008) pmid: 18552176
- 325. Byron SA, et al. PLoS ONE (2012) pmid: 22383975
- **326.** Gatius S, et al. Mod. Pathol. (2011) pmid: 21725289
- **327.** Ishikawa A, et al. Int. J. Oncol. (2008) pmid: 18292933
- **328.** Siegfried S, et al. Cancer (1997) pmid: 9070494
- **329.** Powers CJ, et al. Endocr. Relat. Cancer (2000) pmid: 11021964
- **330.** Turner N, et al. Nat. Rev. Cancer (2010) pmid: 20094046
- 331. Reintjes N, et al. PLoS ONE (2013) pmid: 23527311
- 332. Ratisoontorn C, et al. Connect. Tissue Res. (2003) pmid: 12952211
- Mohammadi M, et al. Cytokine Growth Factor Rev. (2005) pmid: 15863029
- 334. Ibrahimi OA, et al. Hum. Mol. Genet. (2004) pmid: 15282208
- **335.** Pollock PM, et al. Oncogene (2007) pmid: 17525745
- **336.** Kapeli K, et al. J. Biol. Chem. (2011) pmid: 21908617
- **337.** Eathiraj S, et al. J. Biol. Chem. (2011) pmid: 21454610
- Tanizaki J, et al. Cancer Res. (2015) pmid: 26048680
 Lorenzi MV, et al. Oncogene (1997) pmid: 9266968
- **340.** Itoh H, et al. Cancer Res. (1994) pmid: 8205545
- 341. Chen H, et al. Mol. Cell (2007) pmid: 17803937
- **342.** Park WJ, et al. Am. J. Hum. Genet. (1995) pmid: 7668257
- 343. Wilkie AO, et al. Nat. Genet. (1995) pmid: 7719344
- **344.** Lajeunie E, et al. Eur. J. Hum. Genet. (2006) pmid: 16418739
- 345. Chen CP, et al. Taiwan J Obstet Gynecol (2013) pmid: 24411056
- **346.** Slavotinek A, et al. Am. J. Med. Genet. A (2009) pmid: 19610084
- **347.** Gozgit JM, et al. Cancer Chemother. Pharmacol. (2013) pmid: 23468082
- **348.** Przylepa KA, et al. Nat. Genet. (1996) pmid: 8696350
- **349.** Kan SH, et al. Am. J. Hum. Genet. (2002) pmid: 11781872
- **350.** Wee S, et al. Proc. Natl. Acad. Sci. U.S.A. (2008) pmid: 18755892
- 351. Jia S, et al. Nature (2008) pmid: 18594509
- **352.** Schmit F, et al. Proc. Natl. Acad. Sci. U.S.A. (2014) pmid: 24737887
- 353. Arkenau et al., 2014; ASCO Abstract 2514
- 354. Siu et al., 2015; AACR Abstract CT329
- **355.** Pazarentzos E, et al. Oncogene (2016) pmid: 25982275
- 356. de Bono et al., 2015; AACR Abstract CT328

- 357. Karlsson T, et al. Oncotarget (2017) pmid: 28002804
- **358.** Phillips WA, et al. Int. J. Cancer (2006) pmid: 16380997
- **359.** Dbouk HA, et al. PLoS ONE (2013) pmid: 23734178 **360.** Matulonis U, et al. Gynecol. Oncol. (2015) pmid:
- **360.** Matulonis U, et al. Gynecol. Oncol. (2015) pmid 25528496
- **361.** Pitz MW, et al. Neuro-oncology (2015) pmid: 25605819
- **362.** Thorpe LM, et al. Proc. Natl. Acad. Sci. U.S.A. (2017) pmid: 28630349
- **363.** Sun M, et al. Proc. Natl. Acad. Sci. U.S.A. (2010) pmid: 20713702
- 364. Juric et al., 2016; SABCS Abstract P3-14-01
- 365. Day TA, et al. Clin. Cancer Res. (2019) pmid: 30420444
- 366. Ou O, et al. Cancer Lett. (2014) pmid: 25193464
- 367. Li X, et al. Nat Commun (2019) pmid: 30755611
- **368.** Quayle SN, et al. PLoS ONE (2012) pmid: 23166678 **369.** Cheung LW, et al. Cancer Cell (2014) pmid: 25284480
- **370.** Brennan CW, et al. Cell (2013) pmid: 24120142
- 371. Cancer Genome Atlas Research Network, et al. N. Engl. J. Med. (2015) pmid: 26061751
- 372. Ye K, et al. Nat. Med. (2016) pmid: 26657142
- 373. Cheung LW, et al. Cancer Discov (2011) pmid: 21984976
- **374.** Urick ME, et al. Cancer Res. (2011) pmid: 21478295
- 375. Munkley J, et al. Oncoscience (2015) pmid: 26501081
- 376. Cizkova M, et al. BMC Cancer (2013) pmid: 24229379
- 377. Qian ZR, et al. J. Clin. Oncol. (2013) pmid: 23980085
- 378. Huang CH, et al. Cell Cycle (2008) pmid: 18418043
- 379. Taniguchi CM, et al. Cancer Res. (2010) pmid:
- 20530665 **380.** Luo J, et al. Cell Metab. (2006) pmid: 16679293
- **381.** Ueki K, et al. J. Biol. Chem. (2003) pmid: 14504291
- **382.** Mauvais-Jarvis F, et al. J. Clin. Invest. (2002) pmid: 11781359
- **383.** Luo J, et al. Proc. Natl. Acad. Sci. U.S.A. (2005) pmid: 16006513
- 384. Jaiswal BS, et al. Cancer Cell (2009) pmid: 19962665
- 385. Ko HR, et al. Cell Death Dis (2014) pmid: 24651434
- **386.** Huang CH, et al. Science (2007) pmid: 18079394
- **387.** Bousquet C, et al. EMBO J. (2006) pmid: 16917505 **388.** Oliver MD, et al. Biosci. Rep. (2017) pmid: 28143957
- **389.** Philp AJ, et al. Cancer Res. (2001) pmid: 11606375
- **390.** Lucas CL, et al. J. Exp. Med. (2014) pmid: 25488983
- 391. Courtney KD, et al. J. Clin. Oncol. (2010) pmid:
- 392. Simpson L, et al. Exp. Cell Res. (2001) pmid: 11237521
- 393. Milella M, et al. Sci Rep (2017) pmid: 28220839
- **394.** Meyer LA, et al. Int J Gynecol Cancer (2014) pmid: 24651628
- 395. Oza AM, et al. J. Clin. Oncol. (2011) pmid: 21788564
- **396.** Mackay HJ, et al. Cancer (2014) pmid: 24166148
- **397.** Fleming GF, et al. Gynecol. Oncol. (2014) pmid: 24456823
- 398. Tsoref D, et al. Gynecol. Oncol. (2014) pmid: 25173583
- **399.** Trédan O, et al. Target Oncol (2013) pmid: 23238879
- 400. Slomovitz BM, et al. Cancer (2010) pmid: 20681032
- **401.** Mendes-Pereira AM, et al. EMBO Mol Med (2009) pmid: 20049735
- 402. Shen Y, et al. Clin. Cancer Res. (2013) pmid: 23881923
- 403. Chatterjee P, et al. PLoS ONE (2013) pmid: 23565244
- **404.** McCormick A, et al. Int. J. Gynecol. Cancer (2016) pmid: 26905328
- **405.** Forster MD, et al. Nat Rev Clin Oncol (2011) pmid: 21468130
- 406. Eikesdal HP, et al. Ann Oncol (2021) pmid: 33242536
- **407.** Pan M, et al. Perm J (2021) pmid: 33970096
- 408. Romero I, et al. Gynecol Oncol (2020) pmid: 32988624

- **409.** Yap TA, et al. Cancer Discov (2020) pmid: 32532747
- 410. Byron SA, et al. Cancer Res. (2008) pmid: 18757403
- Peterson LM, et al. Int. J. Gynecol. Pathol. (2012) pmid: 22498935
- 412. Daniilidou K, et al. J BUON () pmid: 23613406
- 413. van der Zee M, et al. J. Pathol. (2013) pmid: 23288720
- **414.** Joshi A, et al. Am. J. Pathol. (2012) pmid: 22503752
- **415.** Campbell RB, et al. J. Biol. Chem. (2003) pmid: 12857747
- 416. Rodríguez-Escudero I, et al. Hum. Mol. Genet. (2011) pmid: 21828076
- 417. He X, et al. Cancer Res. (2013) pmid: 23475934
- 418. Han SY, et al. Cancer Res. (2000) pmid: 10866302
- 419. Myers MP, et al. Proc. Natl. Acad. Sci. U.S.A. (1998) pmid: 9811831
- 420. Pradella LM, et al. BMC Cancer (2014) pmid: 24498881
- 421. Kim JS, et al. Mol. Cell. Biol. (2011) pmid: 21536651
- 422. Denning G, et al. Oncogene (2007) pmid: 17213812
- 423. Hlobilkova A, et al. Anticancer Res. () pmid: 16619501
- 424. Redfern RE, et al. Protein Sci. (2010) pmid: 20718038
- **425.** Shenov S, et al. PLoS ONE (2012) pmid: 22505997
- **426.** Wang Y, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) pmid: 19329485
- 427. Okumura K, et al. J. Biol. Chem. (2006) pmid: 16829519
- **428.** Lee JO, et al. Cell (1999) pmid: 10555148
- **429.** Maxwell GL, et al. Cancer Res. (1998) pmid: 9635567
- 430. Risinger JI, et al. Clin. Cancer Res. (1998) pmid: 9865913
- 431. Kato H, et al. Clin. Cancer Res. (2000) pmid: 11051241
- **432.** Fenton TR, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) pmid: 22891331
- **433.** Ngeow J, et al. J. Clin. Endocrinol. Metab. (2012) pmid: 23066114
- **434.** Lobo GP, et al. Hum. Mol. Genet. (2009) pmid: 19457929
- **435.** Liu J, et al. Oncogene (2014) pmid: 23995781
- **436.** Maehama T, et al. Annu. Rev. Biochem. (2001) pmid:
- **437.** De Vivo I, et al. J. Med. Genet. (2000) pmid: 10807691
- **438.** Ramaswamy S, et al. Proc. Natl. Acad. Sci. U.S.A. (1999) pmid: 10051603
- **439.** Liu JL, et al. Mol. Cell. Biol. (2005) pmid: 15988030
- **440.** Karoui M, et al. Br. J. Cancer (2004) pmid: 15026806
- **440.** Karoui M, et al. Br. J. Cancer (2004) pmid: 150 **441.** Gil A, et al. PLoS ONE (2015) pmid: 25875300
- 442. Furnari FB, et al. Cancer Res. (1998) pmid: 9823298
- **443.** Spinelli L, et al. J. Med. Genet. (2015) pmid: 25527629 **444.** Mingo J, et al. Eur. J. Hum. Genet. (2018) pmid:
- 29706633 445. Wang Q, et al. J. Mol. Graph. Model. (2010) pmid:
- 20538496 **446.** Andrés-Pons A, et al. Cancer Res. (2007) pmid:
- 17942903 447. Butler MG. et al. J. Med. Genet. (2005) pmid: 15805158
- 448. Georgescu MM, et al. Proc. Natl. Acad. Sci. U.S.A. (1999)
- pmid: 10468583 449. Staal FJ, et al. Br. J. Cancer (2002) pmid: 12085208
- **450.** Nguyen HN, et al. Oncogene (2014) pmid: 24292679
- Nguyen HN, et al. Oncogene (2014) pmid: 2429267
 Rahdar M, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) pmid: 19114656
- **452.** Das S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) pmid: 12808147
- **453.** Wang X, et al. Biochem, J. (2008) pmid: 18498243
- **454.** Valiente M, et al. J. Biol. Chem. (2005) pmid: 15951562
- **455.** Nguyen HN, et al. Oncogene (2015) pmid: 25263454 **456.** Blumenthal GM, et al. Eur. J. Hum. Genet. (2008) pmid: 18781191
- 457. Orloff MS, et al. Oncogene (2008) pmid: 18794875



APPENDIX

References

- 458. Zbuk KM, et al. Nat. Rev. Cancer (2007) pmid: 17167516
- 459. Rivera B, et al. Cancer Res. (2017) pmid: 28646019
- 460. Loveday C, et al. Nat. Genet. (2011) pmid: 21822267
- 461. Kondrashova O, et al. Cancer Discov (2017) pmid:
- 462. Hinz JM, et al. Nucleic Acids Res. (2006) pmid: 16522646
- 463. Wiese C, et al. Nucleic Acids Res. (2006) pmid: 16717288
- 464. Wickramanayake A, et al. Gynecol. Oncol. (2012) pmid:
- 465. Tarsounas M, et al. Cell (2004) pmid: 15109494
- 466. Suwaki N, et al. Semin. Cell Dev. Biol. (2011) pmid: 21821141
- Sasaki MS, et al. Cytogenet. Genome Res. (2004) pmid: 467. 15162012
- 468. Smiraldo PG, et al. Cancer Res. (2005) pmid: 15781618
- 469. Graffeo R, et al. Breast Cancer Res. Treat. (2016) pmid: 27734215
- 470. Osher DJ. et al. Br. J. Cancer (2012) pmid: 22415235
- 471. Pelttari LM, et al. J. Med. Genet. (2012) pmid: 22652533
- 472. Gutiérrez-Enríquez S, et al. Int. J. Cancer (2014) pmid:
- 473. Song H, et al. J. Clin. Oncol. (2015) pmid: 26261251
- 474. Janatova M, et al. PLoS ONE (2015) pmid: 26057125
- 475. Stafford JL, et al. PLoS ONE (2017) pmid: 28591191
- 476. Konstanta I, et al. J. Hum. Genet. (2018) pmid: 30111881
- 477. Chen X. et al. Ann. Oncol. (2018) pmid: 30165555
- 478. Tee AR, et al. Curr. Biol. (2003) pmid: 12906785
- 479. Mallela K, et al. Mol Cell Biochem (2021) pmid: 33575875
- 480. Adib E, et al. Clin Cancer Res (2021) pmid: 33727259
- 481. Nassar AH, et al. Mol Cancer Ther (2020) pmid: 31653662
- 482. Voss MH, et al. Clin. Cancer Res. (2018) pmid: 30327302
- 483. Ali SM, et al. Eur. Urol. (2015) pmid: 25796537
- Kwiatkowski DJ, et al. Clin. Cancer Res. (2016) pmid:
- 485. Hamieh L, et al. PLoS Genet (2018) pmid: 30256787
- Roldan-Romero JM, et al. Int J Cancer (2020) pmid: 31335987
- 487. Inoki K, et al. Genes Dev. (2003) pmid: 12869586
- 488. Miloloza A, et al. Hum. Mol. Genet. (2000) pmid: 10915759
- Hoogeveen-Westerveld M, et al. Biochim. Biophys. Acta (2010) pmid: 20547222
- 490. Hodges AK, et al. Hum. Mol. Genet. (2001) pmid: 11741833
- 491. Ann. N. Y. Acad. Sci. (1991) pmid: 2039135
- 492. van Slegtenhorst M, et al. Science (1997) pmid:
- 493. Crino PB, et al. N. Engl. J. Med. (2006) pmid: 17005952
- 494. Curatolo P, et al. Lancet (2008) pmid: 18722871
- 495. Zhang L, et al. Nature (2010) pmid: 20348907
- 496. Lu W, et al. Eur. J. Pharmacol. (2009) pmid: 19026633
- 497. Tuynman JB, et al. Cancer Res. (2008) pmid: 18281498
- 498. Lau T. et al. Cancer Res. (2013) pmid: 23539443
- Pijnenborg JM, et al. Int. J. Gynecol. Cancer () pmid: 15361208
- 500. Moreno-Bueno G, et al. Oncogene (2002) pmid: 12439748
- 501. Zysman M, et al. Cancer Res. (2002) pmid: 12097272
- 502. Singh M, et al. Gynecol. Oncol. (2011) pmid: 21813170
- 503. Tanwar PS, et al. Cancer Res. (2011) pmid: 21363919
- 504. Luke JJ, et al. Clin Cancer Res (2019) pmid: 30635339
- 505. Logan CY, et al. Annu. Rev. Cell Dev. Biol. (2004) pmid: 15473860

- 506. Kerr SE, et al. J Mol Diagn (2013) pmid: 23159591
- 507. Annu Rev Pathol (2011) pmid: 21090969
- 508. Kastritis E, et al. Int. J. Cancer (2009) pmid: 18844223
- 509. Half E, et al. Orphanet J Rare Dis (2009) pmid:
- 510. Liang J, et al. Cancer Res (2020) pmid: 31551363
- 511. Garbarino J, et al. Transl Oncol (2021) pmid: 34118569
- 512. Cole et al., 2021; AACR Abstract CT059
- 513. George SL, et al. EBioMedicine (2020) pmid: 32846370
- 514. Koschmann C, et al. Sci Transl Med (2016) pmid: 26936505
- 515. Heaphy CM, et al. Science (2011) pmid: 21719641
- 516. Singhi et al., 2015; USCAP Abstract 1797
- 517. Jiao Y, et al. Science (2011) pmid: 21252315
- 518. Fishbein L, et al. Nat Commun (2015) pmid: 25608029
- 519. Morosini et al., 2014: ASCO Abstract 11008
- 520. Cheung NK, et al. JAMA (2012) pmid: 22416102
- 521. Molenaar JJ, et al. Nature (2012) pmid: 22367537
- 522. Pugh TJ, et al. Nat. Genet. (2013) pmid: 23334666 523. Cheung NK, et al. Nat. Rev. Cancer (2013) pmid:
- 23702928
- 524. Marinoni I, et al. Gastroenterology (2014) pmid: 24148618 525. Qadeer ZA, et al. J. Invest. Dermatol. (2014) pmid:
- 24468746
- 526. Kannan K, et al. Oncotarget (2012) pmid: 23104868
- 527. Haberler C, et al. Clin. Neuropathol. () pmid: 24559763 528. Reuss DE, et al. Acta Neuropathol. (2015) pmid: 25427834
- 529. Sahm F, et al. Acta Neuropathol. (2014) pmid: 25143301
- 530. Singhi et al., 2015; USCAP Abstract 93
- 531. Liau JY, et al. Am. J. Surg. Pathol. (2015) pmid: 25229770
- 532. Clynes D, et al. Trends Biochem. Sci. (2013) pmid: 23916100
- 533. Ratnakumar K, et al. Epigenetics (2013) pmid:
- 534. Lovejoy CA, et al. PLoS Genet. (2012) pmid: 22829774
- 535. Bower K, et al. PLoS ONE (2012) pmid: 23185534
- 536. Nan X, et al. Proc. Natl. Acad. Sci. U.S.A. (2007) pmid: 17296936
- 537. Garrick D, et al. Gene (2004) pmid: 14729260
- 538. Eustermann S, et al. Nat. Struct. Mol. Biol. (2011) pmid:
- 539. Flynn RL, et al. Science (2015) pmid: 25593184
- 540. Gibbons RJ, et al. Cell (1995) pmid: 7697714
- 541. Thome M, et al. Cold Spring Harb Perspect Biol (2010) pmid: 20685844
- 542. Lim KH, et al. Immunol. Rev. (2012) pmid: 22435566
- 543. Milhollen MA, et al. Blood (2010) pmid: 20525923
- 544. Wu C, et al. Oncotarget (2016) pmid: 27224912
- **545.** Nagel D, et al. Oncotarget (2015) pmid: 26540570
- 546. Naylor TL, et al. Cancer Res. (2011) pmid: 21324920
- 547. Bu R, et al. Leuk. Lymphoma (2012) pmid: 22397314
- **548.** Lenz G, et al. Science (2008) pmid: 18323416
- 549. Morin RD, et al. Blood (2013) pmid: 23699601
- 550. Morin RD, et al. Nature (2011) pmid: 21796119
- 551. Montesinos-Rongen M, et al. Acta Neuropathol. (2010) pmid: 20544211
- **552.** Braggio E, et al. Clin. Cancer Res. (2015) pmid: 25991819
- 553. Rossi D, et al. J. Exp. Med. (2012) pmid: 22891273
- 554. Yan O. et al. Haematologica (2012) pmid: 22102703
- 555. Vallois D, et al. Blood (2016) pmid: 27369867
- 556. Wang L. et al. Nat. Genet. (2015) pmid: 26551670
- 557. da Silva Almeida AC, et al. Nat. Genet. (2015) pmid: 26551667

- 558. Oshiro A, et al. Blood (2006) pmid: 16484591
- 559. Fujiwara SI, et al. Leukemia (2008) pmid: 18633432
- 560. Ma Y, et al. Eur. J. Med. Res. (2014) pmid: 25384343
- 561. Krauthammer M, et al. Nat. Genet. (2012) pmid:
- 562. Hodis E. et al. Cell (2012) pmid: 22817889
- 563. Witkiewicz AK, et al. Nat Commun (2015) pmid: 25855536
- 564. Nature (2014) pmid: 25079317
- 565. Nature (2014) pmid: 25079552
- 566. Nature (2014) pmid: 24476821
- 567. Weil R, et al. Curr. Opin. Immunol. (2004) pmid: 15134788
- 568. Paul S, et al. Trends Immunol. (2013) pmid: 23474202
- **569.** Juilland M, et al. Curr. Opin. Hematol. (2016) pmid: 27135977
- 570. Bognar MK, et al. Oncogene (2016) pmid: 26776161
- 571. Chan W, et al. Mol. Cell. Biol. (2013) pmid: 23149938
- 572. Brohl AS, et al. J. Clin. Immunol. (2015) pmid: 25352053
- 573. Jeelall YS, et al. J. Exp. Med. (2012) pmid: 23027925
- 574. Lamason RL, et al. Biochemistry (2010) pmid: 20799731
- 575. Dong G, et al. Clin. Cancer Res. (2011) pmid: 21266526
- 576. Compagno M, et al. Nature (2009) pmid: 19412164
- 577. Snow AL, et al. J. Exp. Med. (2012) pmid: 23129749 578. Mensah AA, et al. Oncotarget (2015) pmid: 25671298
- 579. Grasso CS, et al. Ann. Oncol. (2015) pmid: 25735316 580. Ma X. et al. Nat Commun (2015) pmid: 25790293
- 581. Green MR, et al. Proc. Natl. Acad. Sci. U.S.A. (2015)
- pmid: 25713363
- 582. Loeffler M, et al. Leukemia (2015) pmid: 25027518 583. Gervais C, et al. Leukemia (2008) pmid: 18528428
- 584. Haferlach T, et al. Leukemia (2009) pmid: 19194466
- 585. Petrij F, et al. J. Med. Genet. (2000) pmid: 10699051 586. Borrow J, et al. Nat. Genet. (1996) pmid: 8782817
- 587. Kao YC, et al. Genes Chromosomes Cancer (2017) pmid: 27537276
- 588. Hofvander J, et al. Mod. Pathol. (2020) pmid: 31932680
- 589. Méndez-Catalá CF, et al. Neoplasia (2013) pmid:
- 590. Tiffen JC, et al. Int. J. Cancer (2013) pmid: 23553099
- 591. Phillips JE, et al. Cell (2009) pmid: 19563753
- 592. Gombert WM, et al. PLoS ONE (2009) pmid: 19568426
- 593. Soto-Reyes E, et al. Oncogene (2010) pmid: 20101205
- 594. Woloszynska-Read A, et al. Clin. Cancer Res. (2011) pmid: 21296871
- 595. Kemp CJ, et al. Cell Rep (2014) pmid: 24794443
- 596. Hoadley KA, et al. Cell (2018) pmid: 29625048
- 597. Van Allen EM, et al. Cancer Discov (2014) pmid: 24265153
- **598.** Hansford S, et al. JAMA Oncol (2015) pmid: 26182300
- 599. Majewski IJ, et al. J. Pathol. (2013) pmid: 23208944
- 600. Ding L, et al. Nature (2010) pmid: 20393555
- 601. Piao HL, et al. Nat. Cell Biol. (2014) pmid: 24509793 602. Qian J, et al. Clin. Chem. Lab. Med. (2014) pmid:
- 603. Ye Y, et al. Cancer Res. (2009) pmid: 19826047
- 604. Liu TX, et al. Nat. Med. (2007) pmid: 17159988 605. Chen XX, et al. Leuk. Res. (2014) pmid: 24685333
- 606. Li M, et al. Oncotarget (2016) pmid: 27129146 607. Kobielak A, et al. Nat. Rev. Mol. Cell Biol. (2004) pmid:
- 15366705 608. Bajpai S, et al. J. Biol. Chem. (2009) pmid: 19458087
- **609.** Silvis MR, et al. Sci Signal (2011) pmid: 21610251 610. Inge LJ, et al. Mol. Cancer Ther. (2008) pmid: 18566211

611. Ooi A, et al. Cancer Res. (2013) pmid: 23365135

APPENDIX

References

- **612.** Genschik P, et al. EMBO J. (2013) pmid: 23912815
- **613.** Grau L, et al. PLoS ONE (2013) pmid: 23308193
- 614. Kossatz U, et al. J. Clin. Invest. (2010) pmid: 20978349
- **615.** Straubhar A, et al. Gynecol Oncol Rep (2017) pmid: 28560298
- **616.** Thangavelu A, et al. Gynecol. Oncol. (2013) pmid: 24076063
- Gershenson DM, et al. J. Clin. Oncol. (2017) pmid: 28221866
- 618. Esfahani K, et al. BMJ Case Rep (2014) pmid: 24925537
- **619.** Ramirez PT, et al. Gynecol. Oncol. (2008) pmid: 18457865
- **620.** Turner NC, et al. N. Engl. J. Med. (2018) pmid: 30345905
- **621.** Fribbens C, et al. J. Clin. Oncol. (2016) pmid: 27269946
- **622.** Spoerke JM, et al. Nat Commun (2016) pmid: 27174596
- **623.** Gaillard SL, et al. Gynecol. Oncol. (2019) pmid: 30987772
- **624.** Schiavon G, et al. Sci Transl Med (2015) pmid: 26560360
- **625.** Chandarlapaty S, et al. JAMA Oncol (2016) pmid: 27532364
- **626.** Stover EH, et al. JCO Precis Oncol (2018) pmid: 30828692
- 627. Wik E, et al. Clin. Cancer Res. (2013) pmid: 23319822
- **628.** Rahman MT, et al. Anticancer Res. (2013) pmid: 24023309
- **629.** Pearce ST, et al. Crit. Rev. Oncol. Hematol. (2004) pmid: 15094156
- 630. Perotti D, et al. Oncogene (2008) pmid: 18391980
- **631.** Ruteshouser EC, et al. Genes Chromosomes Cancer (2008) pmid: 18311776
- 632. Rivera MN, et al. Science (2007) pmid: 17204608
- 633. Major MB, et al. Science (2007) pmid: 17510365
- **634.** Jenkins ZA, et al. Nat. Genet. (2009) pmid: 19079258
- **635.** Perdu B, et al. J. Bone Miner. Res. (2010) pmid: 20209645
- **636.** Clapéron A, et al. J. Biol. Chem. (2005) pmid: 15769748
- **637.** Nature (2012) pmid: 22960745
- 638. Nature (2011) pmid: 21720365
- **639.** Gupta-Rossi N, et al. Mol. Immunol. (2003) pmid: 12617994
- 640. Puente XS, et al. Nature (2011) pmid: 21642962
- **641.** Weigert O, et al. Cancer Discov (2012) pmid: 22585168
- 642. Xia Y, et al. Genes Dev. (1998) pmid: 9808624
- 643. Lu Z, et al. Mol. Cell (2002) pmid: 12049732
- **644.** Schlesinger TK, et al. J. Biol. Chem. (2002) pmid: 11782455
- **645.** Garcia-Closas M, et al. PLoS Genet. (2008) pmid: 18437204
- 646. Slattery ML, et al. Breast Cancer Res. Treat. (2011) pmid: 21475998
- **647.** Augert A, et al. J Thorac Oncol (2017) pmid: 28007623
- 648. Ardeshir-Larijani F, et al. Clin Lung Cancer (2018) pmid: 29627316
- 649. Hillman RT, et al. Nat Commun (2018) pmid: 29950560
- 650. Abudureheman A, et al. J. Cancer Res. Clin. Oncol. (2018) pmid: 29532228
- **651.** Vicent GP, et al. Genes Dev. (2011) pmid: 21447625
- **652.** Hannibal MC, et al. Am. J. Med. Genet. A (2011) pmid: 21671394
- **653.** Fagan RJ, et al. Cancer Lett. (2019) pmid: 31128216
- 654. Joly MO, et al. Hum. Mutat. (2015) pmid: 25504677
- **655.** Pritchard CC, et al. Nat Commun (2014) pmid: 25255306
- 656. Rosty C, et al. Fam. Cancer (2014) pmid: 25117503
- **657.** McConechy MK, et al. Gynecol. Oncol. (2015) pmid: 25636458

- 658. Le et al., 2015: ASCO Abstract LBA100
- **659.** Lipson EJ, et al. Clin. Cancer Res. (2013) pmid: 23169436
- 660. Bonadona V, et al. JAMA (2011) pmid: 21642682
- **661.** Baglietto L, et al. J. Natl. Cancer Inst. (2010) pmid: 20028993
- **662.** Lu KH, et al. Fam. Cancer (2013) pmid: 23765559
- 663. Shih KK, et al. Gynecol. Oncol. (2011) pmid: 21742371
- 664. Li F, et al. Cell (2013) pmid: 23622243
- 665. Wu H, et al. PLoS ONE (2011) pmid: 21720545
- 666. Edelbrock MA, et al. Mutat. Res. () pmid: 23391514
- Berends MJ, et al. Am. J. Hum. Genet. (2002) pmid: 11709755
- 668. Warren JJ, et al. Mol. Cell (2007) pmid: 17531815
- 669. Geng H, et al. J. Biol. Chem. (2012) pmid: 22277660
- 670. Silva FC, et al. Sao Paulo Med J (2009) pmid: 19466295
- 671. Raevaara TE, et al. Gastroenterology (2005) pmid: 16083711
- 672. Kastrinos F, et al. Semin. Oncol. (2007) pmid: 17920897
- 673. Sehgal R, et al. Genes (Basel) (2014) pmid: 24978665
- 674. Fam. Cancer (2005) pmid: 16136383
- 675. Wimmer K, et al. Hum. Genet. (2008) pmid: 18709565
- 676. Wimmer K, et al. J. Med. Genet. (2014) pmid: 24737826
- **677.** Scott RH, et al. Nat Clin Pract Oncol (2007) pmid: 17759933
- 678. Ripperger T, et al. Haematologica (2010) pmid:
- **679.** Baris HN, et al. Pediatr Blood Cancer (2016) pmid: 26544533
- **680.** Bouffet E, et al. J. Clin. Oncol. (2016) pmid: 27001570
- **681.** Shlien A, et al. Nat. Genet. (2015) pmid: 25642631
- **682.** Santin AD, et al. Clin. Cancer Res. (2016) pmid: 27486176
- 683. Gong J, et al. J Natl Compr Canc Netw (2017) pmid: 28188185
- 684. Howitt BE, et al. JAMA Oncol (2015) pmid: 26181000
- **685.** Billingsley CC, et al. Int. J. Gynecol. Cancer (2016) pmid: 26937754
- **686.** McConechy MK, et al. Clin. Cancer Res. (2016) pmid: 26763250
- **687.** Church DN, et al. J. Natl. Cancer Inst. (2015) pmid: 25505230
- 688. Meng B, et al. Gynecol. Oncol. (2014) pmid: 24844595
- **689.** Murali R, et al. Lancet Oncol. (2014) pmid: 24872110
- 690. Popanda O, et al. Biochim. Biophys. Acta (1992) pmid: 1730053
- 691. Palles C, et al. Nat. Genet. (2013) pmid: 23263490
- 692. Valle L, et al. Hum. Mol. Genet. (2014) pmid: 24501277
- 693. Rayner E, et al. Nat. Rev. Cancer (2016) pmid: 26822575
- **694.** Spier I, et al. Int. J. Cancer (2015) pmid: 25529843
- 695. Shinbrot E, et al. Genome Res. (2014) pmid: 25228659
- 696. Rohlin A, et al. Int. J. Oncol. (2014) pmid: 24788313
- 697. Abdus A, et al. Int. J. Oncol. (2014) pinto. 24788313
- 698. Murphy K, et al. Genome (2006) pmid: 16699561
- 699. Dis. Colon Rectum (2014) pmid: 24509466
- 700. Mur P. et al. Genet Med (2020) pmid: 32792570
- 701. Buchanan DD, et al. Genet Med (2018) pmid: 29120461
- **702.** Varela I, et al. Nature (2011) pmid: 21248752
- 703. Chen Z, et al. Biochem Biophys Res Commun (2018) pmid: 29522714
- **704.** Chen BY, et al. Blood (2020) pmid: 32202636
- 705. Sun XJ, et al. J. Biol. Chem. (2005) pmid: 16118227
- **706.** Faber PW, et al. Hum. Mol. Genet. (1998) pmid: 9700202
- **707.** Al Sarakbi W, et al. BMC Cancer (2009) pmid: 19698110
- 708. Stephens PJ, et al. J. Clin. Invest. (2013) pmid: 23778141

- **709.** Shi Y, et al. Genes Dev. (2001) pmid: 11331609
- 710. Ariyoshi M, et al. Genes Dev. (2003) pmid: 12897056
- 711. Kuroda K, et al. Immunity (2003) pmid: 12594956
- 712. Oswald F, et al. EMBO J. (2002) pmid: 12374742713. Kopan R, et al. Cell (2009) pmid: 19379690
- **714.** Bailey ML, et al. Mol. Cancer Ther. (2014) pmid: 24356817
- 715. Evers L, et al. Genome Med (2014) pmid: 24484537
- 716. Solomon DA, et al. Nat. Genet. (2013) pmid: 24121789
- 717. Balbás-Martínez C, et al. Nat. Genet. (2013) pmid: 24121791
- 718. Guo G, et al. Nat. Genet. (2013) pmid: 24121792
- **719.** Taylor CF, et al. Hum. Mol. Genet. (2014) pmid: 24270882
- 720. Tirode F, et al. Cancer Discov (2014) pmid: 25223734
- **721.** Brohl AS, et al. PLoS Genet. (2014) pmid: 25010205
- 722. Hoang ML, et al. Sci Transl Med (2013) pmid: 23926200
- 723. Thota S, et al. Blood (2014) pmid: 25006131
- 724. Kon A, et al. Nat. Genet. (2013) pmid: 23955599
- **725.** Solomon DA, et al. Science (2011) pmid: 21852505
- **726.** Shen CH, et al. Nat. Med. (2016) pmid: 27500726
- **727.** Solomon DA, et al. BMB Rep (2014) pmid: 24856830 **728.** Kandoth C, et al. Nature (2013) pmid: 24132290
- 729. Hymowitz SG, et al. Nat. Rev. Cancer (2010) pmid: 20383180
- 730. Zhu YX, et al. Blood (2011) pmid: 21289309
- 731. Vendrell JA. et al. Oncogene (2007) pmid: 17297453
- 732. Hjelmeland AB, et al. PLoS Biol. (2010) pmid: 20186265
- 733. Bellail AC, et al. Cancer Discov (2012) pmid: 22585859
- 734. Chen S, et al. Leuk. Res. (2015) pmid: 26159495
 735. Hantel C, et al. Mol. Cell. Endocrinol. (2016) pmid: 26768118
- **736.** Ungerbäck J, et al. Carcinogenesis (2012) pmid:
- 22843550 **737.** Bavi P, et al. Clin Epigenetics (2011) pmid: 22704353
- 738. Harhai EW. et al. Cell Res. (2011) pmid: 21119682
- 739. Wertz IE, et al. Nature (2004) pmid: 15258597
- 740. Wertz IE, et al. Nature (2015) pmid: 26649818741. Honma K, et al. Blood (2009) pmid: 19608751
- **742.** Wang X, et al. Oncotarget (2016) pmid: 26909601
- 743. Couch G, et al. Cancer Prev Res (Phila) (2016) pmid: 27072986
- **744.** Wang Y, et al. PLoS ONE (2015) pmid: 26485275
- 745. Salim H, et al. Genes Chromosomes Cancer (2013) pmid: 23929716
- **746.** Saitoh Y. et al. Leukemia (2016) pmid: 26437781
- 747. Johansson P, et al. Int. J. Cancer (2016) pmid: 26199174
- **748.** Broyl A, et al. Blood (2010) pmid: 20574050
- **749.** Troppan K, et al. PLoS ONE (2015) pmid: 25856582
- **750.** Yamaguchi N, et al. Sci Rep (2013) pmid: 24008839 **751.** Coornaert B, et al. Nat. Immunol. (2008) pmid:
- 18223652
- 752. Hirai H, et al. Cancer Biol. Ther. (2010) pmid: 20107315753. Bridges KA, et al. Clin. Cancer Res. (2011) pmid: 20107315
- 21799033 **754.** Rajeshkumar NV, et al. Clin. Cancer Res. (2011) pmid:
- 21389100 **755.** Osman AA, et al. Mol. Cancer Ther. (2015) pmid:
- 25504633 **756.** Xu L, et al. Mol. Cancer Ther. (2002) pmid: 12489850
- **757.** Xu L, et al. Mol. Med. (2001) pmid: 11713371
- 758. Camp ER, et al. Cancer Gene Ther. (2013) pmid: 23470564
- **759.** Kim SS, et al. Nanomedicine (2015) pmid: 25240597 **760.** Pirollo KF, et al. Mol. Ther. (2016) pmid: 27357628



APPENDIX

References

- 761. Hajdenberg et al., 2012; ASCO Abstract e15010
- 762. Leijen S, et al. J. Clin. Oncol. (2016) pmid: 27601554
- 763. Moore et al., 2019; ASCO Abstract 5513
- 764. Leijen S, et al. J. Clin. Oncol. (2016) pmid: 27998224
- **765.** Oza et al., 2015; ASCO Abstract 5506
- 766. Lee J, et al. Cancer Discov (2019) pmid: 31315834
- **767.** Méndez E, et al. Clin. Cancer Res. (2018) pmid: 29535125
- 768. Ma CX, et al. J. Clin. Invest. (2012) pmid: 22446188
- 769. Lehmann S, et al. J. Clin. Oncol. (2012) pmid: 22965953
- **770.** Mohell N, et al. Cell Death Dis (2015) pmid: 26086967
- **771.** Fransson Å, et al. J Ovarian Res (2016) pmid: 27179933
- 772. Gourley et al., 2016; ASCO Abstract 5571
- 773. Boudny M, et al. Haematologica (2019) pmid: 30975914
- **774.** Dillon MT, et al. Mol. Cancer Ther. (2017) pmid: 28062704
- 775. Middleton FK, et al. Cancers (Basel) (2018) pmid: 30127241
- 776. Trovik J, et al. Eur. J. Cancer (2013) pmid: 23932335
- 777. Wild PJ, et al. EMBO Mol Med (2012) pmid: 22678923
- 778. Lee EJ, et al. Gynecol. Oncol. (2010) pmid: 20006376
- 779. Brown CJ, et al. Nat. Rev. Cancer (2009) pmid: 19935675
- 780. Joerger AC, et al. Annu. Rev. Biochem. (2008) pmid: 18410249
- **781.** Kato S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) pmid: 12826609
- **782.** Kamada R, et al. J. Biol. Chem. (2011) pmid: 20978130
- **783.** Zerdoumi Y, et al. Hum. Mol. Genet. (2017) pmid: 28472496
- 784. Yamada H, et al. Carcinogenesis (2007) pmid: 17690113
- 785. Bougeard G, et al. J. Clin. Oncol. (2015) pmid: 26014290
- **786.** Sorrell AD, et al. Mol Diagn Ther (2013) pmid: 23355100
- Nichols KE, et al. Cancer Epidemiol. Biomarkers Prev. (2001) pmid: 11219776
- **788.** Kleihues P, et al. Am. J. Pathol. (1997) pmid: 9006316
- **789.** Gonzalez KD, et al. J. Clin. Oncol. (2009) pmid: 19204208
- 790. Lalloo F, et al. Lancet (2003) pmid: 12672316
- 791. Mandelker D, et al. Ann. Oncol. (2019) pmid: 31050713
- **792.** Hematology Am Soc Hematol Educ Program (2018) pmid: 30504320
- 793. Neggers JE, et al. Chem. Biol. (2015) pmid: 25579209
- **794.** Ranganathan P, et al. Blood (2012) pmid: 22677130
- 795. Etchin J, et al. Br. J. Haematol. (2013) pmid: 23373539
- **796.** Hing ZA, et al. Leukemia (2016) pmid: 27323910
- **797.** Etchin J, et al. Leukemia (2017) pmid: 27211268
- **798.** Abdul Razak AR, et al. J. Clin. Oncol. (2016) pmid: 26926685
- **799.** Walker CJ, et al. Blood (2013) pmid: 23970380
- 800. Etchin J, et al. Leukemia (2016) pmid: 26202935
- 801. Ranganathan P, et al. Clin. Cancer Res. (2016) pmid: 27358488
- **802.** Zhang K, et al. Exp. Hematol. (2013) pmid: 22986101
- **803.** Schmidt J, et al. Leukemia (2013) pmid: 23752175
- **804.** Lapalombella R, et al. Blood (2012) pmid: 23034282
- 805. Nakayama R, et al. Oncotarget (2016) pmid: 26918731
- **806.** Lin DC, et al. Nat. Genet. (2014) pmid: 24686850
- 807. Vollbrecht C, et al. PLoS ONE (2015) pmid: 26053404
- 808. Guièze R, et al. Blood (2015) pmid: 26316624
- **809.** Herling CD, et al. Blood (2016) pmid: 27226433
- 810. Jardin F, et al. Am. J. Hematol. (2016) pmid: 27312795811. Jeromin S, et al. Leukemia (2014) pmid: 24113472
- 812. Luo J, et al. Int. J. Mol. Med. (2016) pmid: 27279267
- 813. Kırlı K, et al. Elife (2015) pmid: 26673895
- 814. Brodie KM, et al. J. Biol. Chem. (2012) pmid: 22262852

- 815. Luthra R, et al. Haematologica (2014) pmid: 24142997
- **816.** Nguyen NT, et al. Mol Oncol (2014) pmid: 24973012
- 817. Frietze S, et al. BMC Genomics (2014) pmid: 24962896 818. Thollet A, et al. Mol. Cancer (2010) pmid: 21059223
- 819. Huang G, et al. Hum. Mol. Genet. (2005) pmid: 16203743
- 820. Littlepage LE, et al. Cancer Discov (2012) pmid: 22728437
- 821. Vendrell JA, et al. Cancer Res. (2012) pmid: 22593193
- 822. Li J, et al. Int J Clin Exp Pathol (2014) pmid: 25031722
- 823. Rahman MT, et al. Anticancer Res. (2012) pmid: 22843878
- 824. Yang SH, et al. Clin. Cancer Res. (2005) pmid: 15701848
- 825. Shida A, et al. Anticancer Res. (2014) pmid: 25202062
- **826.** Rooney PH, et al. J. Pathol. (2004) pmid: 15476264
- 827. Szczyrba J, et al. Int. J. Cancer (2013) pmid: 22815235 828. Geppert CI, et al. Br. J. Cancer (2014) pmid: 24853183
- 626. Geppert CI, et al. Br. J. Calicer (2014) pilliu. 24655165
- **829.** Toncheva D, et al. Tumour Biol. () pmid: 15897688
- 830. Mao XG, et al. Lab. Invest. (2011) pmid: 21483406
- 831. Schipf A, et al. Virchows Arch. (2008) pmid: 18193277
- **832.** Quinlan KG, et al. Biochim. Biophys. Acta (2007) pmid: 17572303
- 833. Krig SR, et al. J. Biol. Chem. (2007) pmid: 17259635
- 834. Cowger JJ, et al. Oncogene (2007) pmid: 17130829
- 835. Krig SR, et al. Oncogene (2010) pmid: 20661224
- **836.** Banck MS, et al. Epigenetics (2009) pmid: 19242095
- 837. Collins C, et al. Proc. Natl. Acad. Sci. U.S.A. (1998) pmid: 9671742
- 838. Nonet GH, et al. Cancer Res. (2001) pmid: 11245413
- 839. Li P, et al. Int. J. Cancer (2007) pmid: 17266044
- 840. Marabelle et al., 2019; ESMO Abstract 11920
- 841. Oaknin A, et al. JAMA Oncol (2020) pmid: 33001143
- 842. Madariaga et al., 2021; ASCO Abstract 5574
- **843.** Makker et al., 2021; SGO Abstract 11512
- 844. Makker V, et al. J. Clin. Oncol. (2020) pmid: 32167863
- 845. Ott PA, et al. J Clin Oncol (2017) pmid: 28489510
- 846. Liu JF, et al. Gynecol. Oncol. (2019) pmid: 31204078
- **847.** Smith et al., 2016; ASCO Abstract 9028
- 848. Mazieres et al., 2016; ASCO Abstract 9032
- 849. Besse et al., 2015; ECC Abstract 16LBA
- **850.** Spigel et al., 2015; ASCO Abstract 8028
- 851. Fehrenbacher L, et al. Lancet (2016) pmid: 26970723
- 852. Herbst RS, et al. Nature (2014) pmid: 25428504
- 853. Balar et al., 2016; ASCO Abstract LBA4500
- 854. Dreicer et al., 2016: ASCO Abstract 4515
- **855.** Rosenberg JE, et al. Lancet (2016) pmid: 26952546
- **856.** Powles T, et al. Nature (2014) pmid: 25428503
- 857. Geoerger B, et al. Lancet Oncol. (2020) pmid: 31780255
- 858. McDermott DF, et al. J. Clin. Oncol. (2016) pmid: 26755520
- 859. Adams et al., 2016; ASCO Abstract 1009
- **860.** Hellmann MD, et al. Ann. Oncol. (2019) pmid: 30918950
- 861. Bendell et al., 2016; ASCO Abstract 3502
- **862.** Konstantinopoulos PA, et al. J. Clin. Oncol. (2019) pmid: 31461377
- 863. Verschraegen et al., 2016; ASCO Abstract 9036
- 864. Chung et al., 2016; ASCO Abstract 4009
- **865.** Patel et al., 2016; ESMO Abstract 777PD **866.** Hellmann et al., 2017; ASCO Abstract 8503
- 867. Disis et al. 2016: ASCO Abstract 5533
- 868. Dirix et al., 2016; SABCS Abstract S1-04
- **869.** Larkin et al., 2016; ESMO Abstract 775PD **870.** Le Tourneau et al., 2016; ASCO Abstract 4516
- 871. Fakhrejahani et al., 2017; ASCO GU Abstract 159

- 872. Rajan et al., 2016; ASCO Abstract e20106
- 873. Migden MR, et al. N. Engl. J. Med. (2018) pmid: 29863979
- 874. Stratigos et al., 2020; EMSO Abstract LBA47
- 875. Lewis et al. 2020; doi: 10.1136/jitc-2020-SITC2020.0428
- 876. Sezer et al., 2020: ESMO Abstract LBA52
- 877. Antill et al., 2019; ASCO Abstract 5501
- 878. Rubinstein et al., 2019: ASCO Abstract 5582
- 879. Kulkarni et al., 2020; DOI: 10.1016/j.ygyno.2020.05.244
- 880. Janku F, et al. Cancer Res. (2013) pmid: 23066039
- 881. Janku F, et al. J. Clin. Oncol. (2012) pmid: 22271473
- 883. Moulder S, et al. Ann. Oncol. (2015) pmid: 25878190
- 884. Byeon et al., 2020; doi: 10.21037/tcr.2020.04.07
- 885. Wheler JJ, et al. Oncotarget (2014) pmid: 24912489
- 886. Noriega-Iriondo MF, et al. Hered Cancer Clin Pract (2015) pmid: 25649062
- 887. de Bono et al., 2020; ASCO GU Abstract 119
- 888. Konstantinopolous et al., 2018; ASCO Abstract 106
- 889. Mirza et al., 2016; ASCO Abstract 5555
- **890.** Lheureux et al., 2020; ASCO Abstract 6010
- 891. Takano et al., 2019; ASCO Abstract e17124
- 892. Postow MA, et al. N. Engl. J. Med. (2015) pmid: 25891304
- 893. Larkin J, et al. N. Engl. J. Med. (2015) pmid: 26027431
- **894.** Robert C, et al. N. Engl. J. Med. (2015) pmid: 25399552
- 895. Topalian SL, et al. J. Clin. Oncol. (2014) pmid: 24590637
- 896. Weber JS, et al. Lancet Oncol. (2014) pmid: 2439003
- **897.** Brahmer et al., 2014; ASCO Abstract 8112
- 898. Gettinger et al., 2014; ASCO Abstract 8024
- 899. Antonia et al., 2014; ASCO Abstract 8023
- **900.** Rizvi et al., 2014; ASCO Abstract 8022
- **901.** Antonia et al., 2014; ASCO Abstract 8113
- Brahmer J, et al. N. Engl. J. Med. (2015) pmid: 26028407
 Borghaei H, et al. N. Engl. J. Med. (2015) pmid:
- 26412456
- **904.** Sharma P, et al. J. Clin. Oncol. (2019) pmid: 31100038
- **905.** Sharma P, et al. Lancet Oncol. (2017) pmid: 28131785
- **906.** Amin et al., 2014; ASCO Abstract 5010
- 907. McDermott et al., 2016, ASCO Abstract 4507
- Motzer RJ, et al. J. Clin. Oncol. (2015) pmid: 25452452
 Topalian SL, et al. N. Engl. J. Med. (2012) pmid:
- 22658127 910. Brahmer JR, et al. J. Clin. Oncol. (2010) pmid: 20516446
- 910. Branmer JR, et al. J. Clin. Oncol. (2010) pmid: 2051644
- 911. Motzer RJ, et al. N. Engl. J. Med. (2015) pmid: 26406148 912. Overman MJ, et al. Lancet Oncol. (2017) pmid:
- 28734759 913. Overman MJ, et al. J. Clin. Oncol. (2018) pmid:
- 29355075 914. Overman et al., 2019; ASCO GI Abstract 481
- 915. Gillison et al., 2016: AACR Abstract CT099
- 916. Ferris et al., 2016; ASCO Abstract 6009
- 917. Ferris et al. 2019; 31239321918. Brahmer JR, et al. N. Engl. J. Med. (2012) pmid:
- 22658128
- 919. Hamanishi J, et al. J. Clin. Oncol. (2015) pmid: 26351349 920. Normann MC, et al. J Gynecol Oncol (2019) pmid:
- 31074244
- 921. Calvo et al., 2015; ECC Abstract 3098922. Antonia SJ, et al. Lancet Oncol. (2016) pmid: 27269741
- 923. Le et al., 2016; ASCO GI Cancers Abstract 06 924. Kojima et al., 2016; ASCO GI Cancers Abstract TPS175
- 925. Janjigian YY, et al. J. Clin. Oncol. (2018) pmid: 30110194 926. Ansell et al., 2015; ASH Abstract 583



APPENDIX References

- **927.** Ansell SM, et al. N. Engl. J. Med. (2015) pmid: 25482239
- **928.** Younes A, et al. Lancet Oncol. (2016) pmid: 27451390
- **929.** Hodi FS, et al. Lancet Oncol. (2018) pmid: 30361170
- **930.** Hodi FS, et al. Lancet Oncol. (2016) pmid: 27622997
- 931. Larkin J, et al. N. Engl. J. Med. (2019) pmid: 31562797
- 932. Hellmann MD, et al. N. Engl. J. Med. (2019) pmid: 31562796
- 933. Lenz et al., 2020; ASCO GI Abstract 11
- 934. Motzer RJ, et al. Lancet Oncol. (2019) pmid: 31427204
- 935. Motzer RJ, et al. N. Engl. J. Med. (2018) pmid: 29562145
- 936. Paoluzzi et al., 2016; ASCO Abstract 11047
- **937.** Broto et al., 2019; ESMO Abstract 16690
- 938. Nadal et al., 2018; ASCO Abstract 4528
- 939. Kollipara R, et al. Oncologist (2017) pmid: 28778959
- 940. Hodi et al., 2019; AACR abstract CT037
- 941. Klein et al., 2020: ASCO Abstract 6091
- 942. Del Conte G, et al. Br. J. Cancer (2014) pmid: 25025963
- **943.** Gockley AA, et al. Gynecol. Oncol. (2018) pmid: 29937315
- 944. Nakamura et al., 2020; DOI: 10.1200/P0.19.00368
- 945. Matulonis UA, et al. Ann. Oncol. (2016) pmid: 26961146
- 946. Ledermann J, et al. N. Engl. J. Med. (2012) pmid: 22452356
- **947.** Ledermann J, et al. Lancet Oncol. (2014) pmid: 24882434
- **948.** de Bono J, et al. N. Engl. J. Med. (2020) pmid: 32343890
- 949. Seligson ND, et al. Oncologist (2019) pmid: 30541756
- **950.** Lin J, et al. Clin. Cancer Res. (2019) pmid: 31068370
- **951.** Necchi A, et al. Eur. J. Cancer (2018) pmid: 29680362 **952.** Vinitski S, et al. Heart Vessels (1988) pmid: 3253274
- 953. Maio et al., 2021; AACR abstract CT178

- 954. Kristeleit et al., 2014: ASCO Abstract 2573
- 955. Domcheck et al., 2016; ASCO Abstract 4110
- 956. Plummer R, et al. Cancer Chemother. Pharmacol. (2013) pmid: 23423489
- 957. Plummer R, et al. Clin. Cancer Res. (2008) pmid: 19047122
- 958. Wilson RH, et al. Br. J. Cancer (2017) pmid: 28222073
- 959. Glassberg et al., 2020; ASPHO Abstract 2015
- **960.** Coyne et al., 2020; ASCO Abstract 3612
- 961. McCowage et al., 2018; ASCO Abstract 10504
- 962. Mueller et al., 2020; SNO Abstract NFB-17
- 963. Waldner et al., 2020; DOI: 10.1055/s-0040-1715638
- 964. Romo et al., 2019; SNO Abstract RARE-54
- **965.** Coleman RL, et al. Gynecol. Oncol. (2015) pmid: 25887099
- 966. Banerjee A, et al. Neuro-oncology (2017) pmid: 28339824
- 967. Gupta A, et al. Ann. Oncol. (2014) pmid: 24567366
- 968. Robert C, et al. Lancet Oncol. (2013) pmid: 23735514
- **969.** Kirkwood JM, et al. Clin. Cancer Res. (2012) pmid: 22048237
- 970. Banerji U, et al. Clin. Cancer Res. (2010) pmid: 20179232
- 971. Boers-Sonderen MJ, et al. Anticancer Drugs (2012) pmid: 22293660
- 972. Lopez-Chavez A, et al. J. Clin. Oncol. (2015) pmid: 25667274
- 973. Hainsworth JD, et al. J Thorac Oncol (2010) pmid: 20802351
- 974. Allen et al., 2021; ASCO Abstract 10008
- **975.** Deming DA, et al. Invest New Drugs (2016) pmid:
- Krishnamurthy A, et al. Cancer Res. (2018) pmid: 30042150

- Infante JR, et al. Invest New Drugs (2017) pmid: 28424891
- 978. LoRusso PM, et al. BMC Cancer (2017) pmid: 28264648
- 979. Tolcher AW, et al. Clin. Cancer Res. (2015) pmid:
- 980. Wilky BA, et al. Br. J. Cancer (2015) pmid: 25268371
- 981. Turner et al., 2017; ASCO Abstract 1007
- 982. Litton JK, et al. N. Engl. J. Med. (2018) pmid: 30110579
- 983. Ettl J, et al. Ann. Oncol. (2018) pmid: 30124753
- 984. Meehan et al., 2017; AACR Abstract 4687
- 985. de Bono J, et al. Cancer Discov (2017) pmid: 28242752
- 986. Lu E, et al. J Natl Compr Canc Netw (2018) pmid: 30099369
- 987. De Bono et al., 2020; ASCO Abstract 5566
- 988. Turk et al., 2018; ASCO Abstract 2548
- 989. Piha-Paul et al., 2017; EORTC-NCI-AACR Abstract A096
- 990. Dhami J, et al. Cold Spring Harb Mol Case Stud (2018) pmid: 29588307
- 991. Myers et al., 2015; ASCO Annual Meeting Abstract 5592
- 992. Kollmannsberger C, et al. Ann. Oncol. (2012) pmid: 21447615
- 993. Alvarez EA, et al. Gynecol. Oncol. (2013) pmid: 23262204
- 994. Kurzrock et al., 2011; ASCO abstract 3085
- 995. Infante JR, et al. Lancet Oncol. (2012) pmid: 22805291
- 996. Leijen S, et al. Clin. Cancer Res. (2012) pmid: 22767668
- 997. Zimmer L, et al. Clin. Cancer Res. (2014) pmid: 24947927
- 998. Infante JR, et al. Eur. J. Cancer (2013) pmid: 23583440
- 999. Juric et al., 2014: ASCO Abstract 9051