

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT

DISEASE Brain anaplastic astrocytoma
NAME Ou, Yueh-Hsing
DATE OF BIRTH 01 April 1962
SEX Female
MEDICAL RECORD # 10184506

PHYSICIAN

ORDERING PHYSICIAN Hsu, Pin-Chuan
MEDICAL FACILITY Taipei Veterans General Hospital
ADDITIONAL RECIPIENT None
MEDICAL FACILITY ID 205872
PATHOLOGIST Not Provided

SPECIMEN

SPECIMEN SITE Brain
SPECIMEN ID S110-16112 A (PF21005)
SPECIMEN TYPE Slide Deck
DATE OF COLLECTION 08 May 2021
SPECIMEN RECEIVED 27 August 2021

Biomarker Findings

Microsatellite status - MS-Stable
Tumor Mutational Burden - 0 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

EGFR amplification, EGFR-FAM19A2 rearrangement, rearrangement intron 24
CDK4 amplification
MDM2 amplification
PTEN D116fs*18

2 Therapies with Clinical Benefit

27 Clinical Trials

0 Therapies with Resistance

BIOMARKER FINDINGS

Microsatellite status - MS-Stable

Tumor Mutational Burden - 0 Muts/Mb

GENOMIC FINDINGS

EGFR - amplification, EGFR-FAM19A2 rearrangement, rearrangement intron 24

5 Trials *see p. 9*

CDK4 - amplification

10 Trials *see p. 7*

MDM2 - amplification

4 Trials *see p. 10*

PTEN - D116fs*18

10 Trials *see p. 11*

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. *see Biomarker Findings section*

No therapies or clinical trials. *see Biomarker Findings section*

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)
none	Cetuximab
	Panitumumab
none	none
none	none
none	none
none	none

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

ORDERED TEST # ORD-1173008-01

BIOMARKER FINDINGS

BIOMARKER

Microsatellite status

RESULT

MS-Stable

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁻³, including approved therapies nivolumab and pembrolizumab⁴. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and

experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, $p=0.001$)⁵.

FREQUENCY & PROGNOSIS

MSI-High has been reported in 3-8% of adult or pediatric astrocytomas and was generally not associated with Lynch syndrome⁶⁻⁸. Low-level MSI has been reported in 5-9% of glioblastoma (GBM) samples⁹⁻¹¹. A large-scale study did not find high-level microsatellite instability (MSI-H) in any of 129 GBM samples⁹, although a small-scale study reported MSI-H in 4 of 15 pediatric GBMs and 1 of 12 adult GBMs¹². The frequency of MSI has been reported to be increased in relapsed compared to primary GBM⁹, in GBMs with a previous lower grade astrocytoma¹⁰, and in giant cell GBM compared to classic GBM¹¹.

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor¹³. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2¹³⁻¹⁵. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers¹⁶⁻¹⁸. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{13,15,17-18}.

BIOMARKER

Tumor Mutational Burden

RESULT

0 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1¹⁹⁻²¹, anti-PD-1 therapies¹⁹⁻²², and combination nivolumab and ipilimumab²³⁻²⁸. In glioma, a lack of association between TMB and clinical benefit from immune checkpoint inhibitors has been reported^{19,29-30}. However, multiple case studies have reported that patients with ultramutated gliomas driven by POLE

mutations have benefited from treatment with anti-PD-1³¹⁻³² or anti-PD-L1³³ therapies. Therefore, although increased TMB alone may not be a strong biomarker for PD-1 or PD-L1 inhibitors in this cancer type, these agents may have efficacy for patients with glioma harboring both high TMB and POLE mutation.

FREQUENCY & PROGNOSIS

Anaplastic astrocytoma harbors a median TMB of 1.8 mutations per megabase (mut/Mb), and 2% of cases have high TMB (>20 mut/Mb)³⁴. For pediatric patients, high TMB has been reported in a subset of high-grade gliomas, frequently in association with mutations in mismatch repair or proofreading genes and in TP53, whereas BRAF alterations or other oncogene fusions were observed more frequently in brain tumors harboring low TMB³⁵⁻³⁶. Increased TMB has been reported to correlate with higher tumor grade in glioma³⁷ and glioblastoma (GBM) tissue samples with biallelic mismatch repair deficiency

(bMMRD)³¹, as well as with shorter OS of patients with diffuse glioma³⁸.

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma³⁹⁻⁴⁰ and cigarette smoke in lung cancer⁴¹⁻⁴², treatment with temozolomide-based chemotherapy in glioma⁴³⁻⁴⁴, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes⁴⁵⁻⁴⁹, and microsatellite instability (MSI)^{45,48-49}. This sample harbors a TMB below levels that would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents^{19,29-33}.

ORDERED TEST # ORD-1173008-01

GENOMIC FINDINGS

GENE

EGFR

ALTERATION

amplification, EGFR-FAM19A2 rearrangement, rearrangement intron 24

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

A patient with multiple glioblastoma (GBM) tumors, one of which harbored EGFRvIII, experienced progression of the EGFRvIII-positive tumor during treatment with osimertinib⁵⁰. Clinical studies of the second-generation EGFR TKIs afatinib and dacomitinib for patients with EGFR-amplified gliomas have shown limited efficacy⁵¹⁻⁵⁵; however, a small subset of patients has experienced clinical benefit⁵¹⁻⁵³. The second-generation EGFR TKIs afatinib and dacomitinib have shown minimal efficacy for patients with EGFRvIII glioblastoma (GBM)^{51,53-54,56}. A Phase 1/2 study of afatinib, temozolomide, or the combination for patients with GBM reported clinical benefit, including for patients with EGFRvIII; however, temozolomide alone and in combination exhibited better responses than afatinib monotherapy^{54,56}. A Phase 2 trial of dacomitinib for patients with EGFR-amplified GBM reported a DCR of 26% (5/19) among patients with EGFR amplification and EGFRvIII; however, the trial failed to meet its primary endpoint of 6-month PFS⁵¹. A retrospective biomarker analysis of another Phase 2 study of dacomitinib for patients with GBM found no association between EGFRvIII and clinical benefit⁵³. A Phase 1 trial of ABT-414, an EGFR-targeted antibody-drug conjugate with a toxic payload, in patients with GBM reported 2 complete responses (CR) and 5 partial responses (PR) in 18 patients with EGFR amplification (39% response rate); no CR or PR were observed in 28 patients without EGFR amplification⁵⁷. A clinical study of patients with GBM treated with gefitinib or erlotinib found no correlation between EGFR amplification or mutation and response to the therapy, but sensitivity to EGFR kinase inhibitors was associated with the co-expression of the EGFRvIII alteration and PTEN⁵⁸. Activation of multiple ERBB family receptors or activation of the PI3K pathway may be responsible for resistance to EGFR-targeted therapy in GBM;

therefore, inhibition of ERBB family members or treatment with PI3K/AKT inhibitors or mTOR inhibitors such as everolimus or temsirolimus in combination with an EGFR-targeted treatment, may be a therapeutic option⁵⁹⁻⁶⁰. In multiple glioblastoma (GBM) studies, the presence of EGFRvIII has not predicted clinical benefit from first-generation EGFR TKIs such as erlotinib⁶¹⁻⁶⁶ or gefitinib^{64,67}. However, case reports have described patients with EGFRvIII-positive GBM responding to erlotinib⁶⁸⁻⁷¹. In a retrospective study of patients with GBM treated with erlotinib or gefitinib, co-expression of EGFRvIII with PTEN protein was the strongest predictor of response ($P < 0.001$)⁵⁸, suggesting that activity in this setting is dependent on PTEN status⁷²⁻⁷³. However, a prospective Phase 2 trial testing erlotinib monotherapy for patients with EGFRvIII and PTEN-positive recurrent glioblastoma reported minimal efficacy and was terminated⁶⁶. Multiple studies have failed to find a positive association between increased EGFR expression and clinical benefit from erlotinib or gefitinib for patients with glioblastoma^{58,74-76}. Case studies of patients with cancers harboring EGFR rearrangements treated with osimertinib have reported mixed results. Of 3 patients with non-small cell lung cancer (NSCLC) with EGFR kinase domain duplication (KDD), 2 attained PRs with osimertinib, whereas the third experienced PD⁷⁷. A patient with multiple glioblastoma (GBM) tumors, one of which harbored EGFRvIII, experienced progression of the EGFRvIII-positive tumor during treatment with osimertinib⁵⁰. Third-generation EGFR inhibitors, such as osimertinib, selectively target mutated EGFR, including EGFR T790M⁷⁸⁻⁷⁹. EGFR amplification or expression may be associated with benefit from anti-EGFR antibodies, such as cetuximab⁸⁰⁻⁸³, panitumumab⁸¹, or necitumumab⁸⁴. Necitumumab is an anti-EGFR antibody that is approved to treat metastatic squamous NSCLC in combination with gemcitabine and cisplatin⁸⁵⁻⁸⁶ that has also shown benefit in patients with CRC and melanoma⁸⁷⁻⁸⁸. Irreversible EGFR inhibitors, as well as HSP90 inhibitors, may be appropriate for patients with de novo or acquired resistance to EGFR-targeted therapy⁸⁹⁻⁹². Preclinical studies have reported that EGFR-mutant cells⁸⁹⁻⁹¹, including cells with exon 20 insertions⁹³, are sensitive to HSP90 inhibitors. Consistent with preclinical data demonstrating that the EGFR inhibitor AZD3759 is capable of penetrating the blood-brain barrier and reducing the volume of brain and leptomeningeal

metastases, preliminary results from a Phase 1 trial evaluating single-agent AZD3759 reported a reduction in the volume of brain metastases in 40.0% (8/20) of patients with previously treated NSCLC harboring either EGFR L858R or EGFR exon 19 deletion, including 3 confirmed PRs and 3 unconfirmed PRs⁹⁴⁻⁹⁵. In a Phase 1/2 trial for advanced NSCLC, the brain-penetrant third-generation EGFR TKI lazertinib enabled ORRs of 54.3% (69/127) for all evaluable patients and 44.4% (8/18, intracranial) for patients with brain metastases⁹⁶. The reovirus Reolysin targets cells with activated RAS signaling⁹⁷⁻⁹⁹ and is in clinical trials for patients with some tumor types. Reolysin has demonstrated mixed clinical efficacy, with the highest rate of response reported for patients with head and neck cancer¹⁰⁰⁻¹⁰⁸.

FREQUENCY & PROGNOSIS

Across several genomic studies of CNS tumors, EGFR alterations have been reported in 13.2% of anaplastic astrocytomas, 5.3-15.9% of glioblastoma multiformes (GBMs), and 0% of pilocytic astrocytomas¹⁰⁹⁻¹¹². Across several genomic studies of CNS tumors, EGFR amplification has been reported in 16.9% of anaplastic astrocytomas, and 39.7% of glioblastoma multiformes (GBMs)¹⁰⁹⁻¹¹². EGFR amplification and/or EGFR expression in glioma has been correlated with poor overall survival in patients under 60 years of age, prolonged survival in patients over the age of 60, and tumor grade¹¹³⁻¹¹⁶.

FINDING SUMMARY

EGFR encodes the epidermal growth factor receptor, which belongs to a class of proteins called receptor tyrosine kinases. In response to signals from the environment, EGFR passes biochemical messages to the cell that stimulate it to grow and divide¹¹⁷. Amplification of EGFR has been associated with increased expression of EGFR mRNA and protein in several cancer types¹¹⁸⁻¹²⁰. Disruption of the EGFR C-terminal region through deletion¹²¹⁻¹²³, truncation^{121-122,124}, splicing errors^{121,124}, or gene fusion¹²⁵⁻¹²⁶, has been demonstrated to be activating and is likely to be oncogenic. These mutations have been shown to cause cellular transformation and tumor formation and to be sensitive to EGFR-targeting therapies, including erlotinib, lapatinib, and cetuximab^{121-123,125-126}. One or more of the alterations observed here are predicted to be activating.

© 2021 Foundation Medicine, Inc. All rights reserved.

ORDERED TEST # ORD-1173008-01

GENOMIC FINDINGS

GENE

CDK4

ALTERATION

amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

CDK4 amplification or activation may predict sensitivity to CDK4/6 inhibitors such as abemaciclib, palbociclib, and ribociclib¹²⁷⁻¹³⁰. Clinical benefit has been reported for limited tumor types including patients with CDK4-amplified liposarcoma and sarcoma in response to treatment with abemaciclib¹³¹, palbociclib^{127,132}, and ribociclib¹³³.

— Potential Resistance —

On the basis of a Phase 1b study, PTEN loss of expression may be associated with resistance to combination therapy with CDK4/6 inhibitors such as ribociclib and aromatase inhibitors such as letrozole¹³⁴.

FREQUENCY & PROGNOSIS

Across TCGA and MKSCC studies, CDK4 amplification has been reported in 4.0-9.4% of glioma cases and 14% of glioblastoma multiforme cases (cBioPortal, Sep 2021)^{109-111,135-136}. A study has reported amplification of the 12q14-15 region, where CDK4 and MDM2 reside, in 5% (2/42) of glioblastomas¹³⁷. Amplification of CDK4 and corresponding increased CDK4 protein expression has been reported to be associated with a poorer patient outcome in anaplastic astrocytoma and

glioblastoma¹³⁸⁻¹⁴¹.

FINDING SUMMARY

CDK4 encodes the cyclin-dependent kinase 4, which regulates the cell cycle, senescence, and apoptosis¹⁴². CDK4 and its functional homolog CDK6 are activated by D-type cyclins and promote cell cycle progression by inactivating the tumor suppressor Rb¹⁴³⁻¹⁴⁴. Amplification of the chromosomal region that includes CDK4 has been reported in multiple cancer types, including lung cancer, glioblastoma, and liposarcoma, and has been associated with overexpression of CDK4 protein^{127,145-151}.

GENE

MDM2

ALTERATION

amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

MDM2 antagonists disrupt the MDM2-p53 interaction, thereby stabilizing p53¹⁵². Preclinical studies have suggested that the amplification of MDM2, in the absence of concurrent TP53 mutations, may increase sensitivity to these agents¹⁵³⁻¹⁵⁴. Preliminary Phase 1 studies of the MDM2-p53 antagonist alrizomadlin (APG-115) reported a PR in a patient with liposarcoma harboring an MDM2 amplification and wildtype for TP53 and SD in 21%-38% (6/28 and 5/13, respectively) of patients in genomically unselected solid tumors¹⁵⁵⁻¹⁵⁶. A Phase 2 trial of alrizomadlin in combination with pembrolizumab reported a PR in 1 of 3 patients with malignant peripheral nerve sheath tumor that had failed standard therapy, as well as PRs in patients with multiple

types of solid tumors that had failed immunotherapy, including 1 out of 14 patients with non-small cell lung cancer; 1 out of 5 patients with urothelial carcinoma; and 2 out of 5, 1 out of 5, and 1 out of 11 patients with mucosal, uveal, and cutaneous melanoma, respectively¹⁵⁷. Phase 1b studies of the MDM2 inhibitor idasanutlin for refractory AML in combination with cytarabine or venetoclax reported anti-leukemic response rates of 33% (25/75) and 37% (11/30), respectively¹⁵⁸⁻¹⁵⁹; clinical benefit (58% ORR, 7/12) with idasanutlin monotherapy has been reported for patients with polycythemia vera¹⁶⁰. The dual MDM2/MDM4 inhibitor ALRN-6924 led to an ORR of 27% (4/15) for patients with TP53 wildtype peripheral T-cell lymphoma in a Phase 2 study¹⁶¹; responses have also been observed in TP53 wildtype AML, MDS, Merkel cell carcinoma, colorectal cancer, and liposarcoma¹⁶²⁻¹⁶³.

FREQUENCY & PROGNOSIS

In the Glioblastoma Multiforme (GBM) TCGA dataset, amplification of MDM2 has been found in 8% of cases¹¹⁰. A study has reported amplification of the 12q14-15 region, where MDM2 and CDK4 reside, in 5% (2/42) of GBMs¹³⁷. Amplification of

MDM2 has been associated with poor survival in patients with glioblastoma^{137,164}.

FINDING SUMMARY

MDM2 encodes an E3 ubiquitin protein ligase, which mediates the ubiquitination and subsequent degradation of p53, Rb1, and other proteins¹⁶⁵⁻¹⁶⁷. MDM2 acts to prevent the activity of the tumor suppressor p53; therefore, overexpression or amplification of MDM2 may be oncogenic¹⁶⁸⁻¹⁶⁹. Overexpression or amplification of MDM2 is frequent in cancer¹⁷⁰. Although two retrospective clinical studies suggest that MDM2 amplification may predict a short time-to-treatment failure on anti-PD-1/PD-L1 immune checkpoint inhibitors, with 4/5 patients with MDM2 amplification¹⁷¹ and 2/3 patients with MDM2 or MDM4 amplification¹⁷² experiencing tumor hyperprogression, amplification of MDM2 or MDM4 was not associated with shorter progression-free survival (PFS) in a retrospective analysis of non-small cell lung cancer (NSCLC) outcomes with immune checkpoint inhibitors (hazard ratio of 1.4, p=0.44)¹⁷³. The latter study reported PFS of >2 months for 5/8 patients with MDM2/MDM4 amplification¹⁷³.

ORDERED TEST # ORD-1173008-01

GENOMIC FINDINGS

GENE

PTEN

ALTERATION

D116fs*18

TRANSCRIPT ID

NM_000314

CODING SEQUENCE EFFECT

346delG

VARIANT ALLELE FREQUENCY (% VAF)

45.1%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

PTEN loss or mutation leads to activation of the PI3K-AKT-mTOR pathway and may predict sensitivity to inhibitors of this pathway¹⁷⁴⁻¹⁷⁷. Across various tissues, most clinical studies have not observed an association between PTEN deficiency and response to inhibitors of the PI3K-AKT-mTOR pathway. However, limited studies in prostate cancer¹⁷⁸⁻¹⁸¹, renal cell carcinoma¹⁸², breast cancer¹⁸³⁻¹⁸⁴, and colorectal cancer¹⁸⁵ have reported an association between PTEN deficiency and response to inhibitors targeting the PI3K-AKT-mTOR pathway. Preclinical data indicate that PTEN loss or inactivation may predict sensitivity to PARP inhibitors¹⁸⁶⁻¹⁹⁰, and clinical benefit has been observed for patients with PTEN-altered breast cancer including triple negative breast cancer¹⁹¹, ovarian cancer¹⁹², uterine leiomyosarcoma¹⁹³, and endometrial cancer¹⁹⁰ treated with PARP inhibitors. However, some studies have reported a lack of association

between PTEN mutation and PARP inhibitor sensitivity¹⁹⁴⁻¹⁹⁵.

— Potential Resistance —

On the basis of a Phase 1b study, PTEN loss of expression may be associated with resistance to combination therapy with CDK4/6 inhibitors such as ribociclib and aromatase inhibitors such as letrozole¹³⁴. Limited clinical evidence in glioblastoma²⁹, leiomyosarcoma¹⁹⁶, and melanoma¹⁹⁷ suggests that PTEN alterations may predict a lack of response to anti-PD-1 therapy. In an analysis of 39 patients with metastatic melanoma treated with pembrolizumab or nivolumab, patients with PTEN-expressing tumors achieved significantly greater reduction of tumor size than those with reduction or loss of PTEN expression¹⁹⁷. In a retrospective analysis of 66 patients with glioblastoma, tumors from nivolumab or pembrolizumab non-responders were significantly enriched for PTEN mutations²⁹. In a patient with uterine leiomyosarcoma treated with pembrolizumab monotherapy, a treatment-resistant tumor arose that harbored PTEN loss¹⁹⁶.

FREQUENCY & PROGNOSIS

Studies in the literature have indicated that PTEN alterations (mutation or homozygous deletion) occur most frequently in glioblastoma (GBM), less frequently in anaplastic astrocytoma, and rarely in lower grade glioma subtypes including low grade astrocytoma, oligodendroglioma, oligoastrocytoma, and ependymoma^{113,198-204}. One study detected PTEN mutation in 42% (97/232) and loss in 10% (24/232) of IDH-wildtype GBM samples analyzed²⁰⁵. In the TCGA dataset, PTEN

mutation was observed in 23% of GBM cases and PTEN deletion was reported in 7% of cases¹¹⁰, while in the Lower Grade Glioma TCGA dataset, PTEN mutation was observed in 4% of cases and homozygous deletion observed in 1.2% of cases²⁰⁶. Loss of PTEN correlated with significantly worse prognosis in all grades of gliomas^{201,207}.

FINDING SUMMARY

PTEN encodes an inositol phosphatase that functions as a tumor suppressor by negatively regulating the PI3K-AKT-mTOR pathway; loss of PTEN can lead to uncontrolled cell growth and suppression of apoptosis¹⁷⁵. Alterations such as seen here may disrupt PTEN function or expression^{60,203,208-246}.

POTENTIAL GERMLINE IMPLICATIONS

PTEN mutations underlie several inherited disorders, collectively termed PTEN hamartoma tumor syndrome (PHTS), which include Cowden syndrome (CS) and its variant Lhermitte-Duclos syndrome (LD), Bannayan-Riley-Ruvalcaba syndrome (BRRS), PTEN-related Proteus syndrome (PS), and Proteus-like syndrome²⁴⁷⁻²⁴⁸. The mutation rate for PTEN in these disorders ranges from 20 to 85% of patients^{247,249}. The estimated incidence of Cowden syndrome is 1/200,000, which may be an underestimate due to the high variability of this disorder²⁴⁷. Given the association between PTEN and these inherited syndromes, in the appropriate clinical context, germline testing for mutations affecting PTEN is recommended.

ORDERED TEST # ORD-1173008-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Cetuximab

Assay findings association

EGFR

amplification, EGFR-FAM19A2 rearrangement, rearrangement intron 24

AREAS OF THERAPEUTIC USE

Cetuximab is a monoclonal antibody that targets EGFR. It is FDA approved for the treatment of head and neck squamous cell carcinoma (HNSCC) and KRAS-wild-type, EGFR-expressing metastatic colorectal cancer (CRC). Please see the drug label for full prescribing information.

GENE ASSOCIATION

For patients with metastatic CRC receiving cetuximab or panitumumab as mono- or combination therapy, increased EGFR copy number associated with improved OS (HR=0.62) in a meta-analysis, although increased survival was not seen in populations that received first-line treatment with EGFR antibodies⁸¹.

SUPPORTING DATA

A clinical trial of cetuximab with bevacizumab (an anti-VEGF monoclonal antibody) in patients with glioblastoma (GBM) did not show improved efficacy compared with bevacizumab alone²⁵⁰. In preclinical trials, cetuximab, matuzumab, and panitumumab were reported to be ineffective at blocking EGFR dimerization and activation in GBM cells expressing EGFR extracellular domain mutations²⁵¹. However, another study demonstrated that in patients with GBM harboring EGFR amplification but lacking expression of the EGFRvIII variant, treatment with cetuximab resulted in significantly better progression-free survival (PFS) and numerical (although not statistically significant) improvement in overall survival (OS)²⁵².

Panitumumab

Assay findings association

EGFR

amplification, EGFR-FAM19A2 rearrangement, rearrangement intron 24

AREAS OF THERAPEUTIC USE

Panitumumab is a monoclonal antibody that targets EGFR. It is FDA approved to treat KRAS wild-type and NRAS wild-type metastatic colorectal cancer (CRC) combined with chemotherapy or as monotherapy for patients who have progressed on prior chemotherapy. Please see the drug label for full prescribing information.

GENE ASSOCIATION

For patients with metastatic CRC receiving cetuximab or panitumumab as mono- or combination therapy, increased EGFR copy number associated with improved OS (HR=0.62) in a meta-analysis, although increased survival was not seen in populations that received first-line treatment with EGFR antibodies⁸¹.

SUPPORTING DATA

A Phase 1 trial of EnGeneIC delivery vehicle (EDV) targeting EGFR with panitumumab in combination with doxorubicin for 14 patients with glioblastoma (GBM) reported no responses and 28% (4/14) SDs²⁵³. Two Phase 2 studies of panitumumab and chemotherapy in biliary tract cancer, including cholangiocarcinoma, reported encouraging efficacy and manageable toxicity²⁵⁴⁻²⁵⁵. In a Phase 2 trial of advanced NSCLC, the addition of panitumumab to paclitaxel/carboplatin did not result in improved clinical benefit²⁵⁶. A Phase 1 study of panitumumab for patients with metastatic renal cell carcinoma resulted in a response rate of 6% and stable disease in 50% of patients²⁵⁷.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

ORDERED TEST # ORD-1173008-01

CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://www.foundationmedicine.com/genomic-testing#support-services). Or visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

GENE
CDK4
RATIONALE
 CDK4 amplification may predict sensitivity to

CDK4/6 inhibitors.

ALTERATION
 amplification

NCT03239015
PHASE 2

Efficacy and Safety of Targeted Precision Therapy in Refractory Tumor With Druggable Molecular Event

TARGETS
 EGFR, ERBB2, ERBB4, PARP, mTOR, MET, RET, ROS1, VEGFRs, BRAF, CDK4, CDK6

LOCATIONS: Shanghai (China)

NCT03099174
PHASE 1

This Study in Patients With Different Types of Cancer (Solid Tumours) Aims to Find a Safe Dose of Xentuzumab in Combination With Abemaciclib With or Without Hormonal Therapies. The Study Also Tests How Effective These Medicines Are in Patients With Lung and Breast Cancer.

TARGETS
 CDK4, CDK6, IGF-1, IGF-2, Aromatase, ER

LOCATIONS: Seoul (Korea, Republic of), Goyang (Korea, Republic of), Aichi, Nagoya (Japan), Kanagawa, Isehara (Japan), Tokyo, Chuo-ku (Japan), Tokyo, Koto-ku (Japan), Chiba, Kashiwa (Japan), Helsinki (Finland), Tampere (Finland), Turku (Finland)

NCT04594005
PHASE 1/2

CDK4/6 Tumor, Abemaciclib, Paclitaxel

TARGETS
 CDK4, CDK6

LOCATIONS: Seoul (Korea, Republic of)

NCT03834740
PHASE NULL

Ph0/2 Ribociclib & Everolimus

TARGETS
 CDK6, CDK4, mTOR

LOCATIONS: Arizona

NCT02933736
PHASE NULL

Ribociclib (LEE011) in Preoperative Glioma and Meningioma Patients

TARGETS
 CDK6, CDK4

LOCATIONS: Arizona

ORDERED TEST # ORD-1173008-01

CLINICAL TRIALS
NCT04801966
PHASE NULL

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

TARGETS
 CDK4, CDK6, PI3K-alpha, PD-L1, MEK,
 PARP, PD-1, BRAF

LOCATIONS: Melbourne (Australia)

NCT03994796
PHASE 2

Genetic Testing in Guiding Treatment for Patients With Brain Metastases

TARGETS
 ALK, ROS1, TRKA, TRKB, TRKC, CDK4,
 CDK6, PI3K, mTOR

LOCATIONS: Alaska, Washington

NCT04116541
PHASE 2

 A Study Evaluating the Activity of Anti-cancer Treatments Targeting Tumor Molecular Alterations/
 Characteristics in Advanced / Metastatic Tumors.

TARGETS
 CDK6, CDK4, MDM2, MET, RET, ROS1,
 VEGFRs

LOCATIONS: Nice (France), Lyon (France), Marseille (France), Toulouse (France), Bordeaux (France)

NCT02981940
PHASE 2

A Study of Abemaciclib in Recurrent Glioblastoma

TARGETS
 CDK4, CDK6

LOCATIONS: Utah, California, Massachusetts

NCT02896335
PHASE 2

Palbociclib In Progressive Brain Metastases

TARGETS
 CDK4, CDK6

LOCATIONS: Massachusetts

ORDERED TEST # ORD-1173008-01

CLINICAL TRIALS

GENE EGFR	RATIONALE EGFR activating mutations, rearrangements, or amplification may predict sensitivity to EGFR-targeted therapies. Strategies to overcome	resistance to current agents include next-generation EGFR inhibitors and combination therapies.
ALTERATION amplification, EGFR-FAM19A2 rearrangement, rearrangement intron 24		

NCT03829436
PHASE 1

TPST-1120 as Monotherapy and in Combination With (Nivolumab, Docetaxel or Cetuximab) in Subjects With Advanced Cancers

TARGETS
 PD-1, PPARalpha, EGFR

LOCATIONS: California, Michigan, Oklahoma, Pennsylvania, New York, Tennessee, Maryland, North Carolina, Florida

NCT02800486
PHASE 2

Super Selective Intra-arterial Repeated Infusion of Cetuximab (Erbix) With Reirradiation for Treatment of Relapsed/Refractory GBM, AA, and AOA

TARGETS
 EGFR

LOCATIONS: New York

NCT03783403
PHASE 1

 A Study of CC-95251, a Monoclonal Antibody Directed Against SIRP α , in Subjects With Advanced Solid and Hematologic Cancers

TARGETS
 CD20, EGFR, SIRP-alpha

LOCATIONS: Heidelberg (Australia), Melbourne (Australia), Edmonton (Canada), California, Colorado, Arizona, Toronto (Canada), Oklahoma, Texas, Pennsylvania

NCT02451553
PHASE 1

Afatinib Dimaleate and Capecitabine in Treating Patients With Advanced Refractory Solid Tumors, Pancreatic Cancer or Biliary Cancer

TARGETS
 EGFR, ERBB2, ERBB4

LOCATIONS: Washington

NCT01552434
PHASE 1

Bevacizumab, Temsirolimus Alone and in Combination With Valproic Acid or Cetuximab in Patients With Advanced Malignancy and Other Indications

TARGETS
 VEGFA, HDAC, mTOR, EGFR

LOCATIONS: Texas

ORDERED TEST # ORD-1173008-01

CLINICAL TRIALS

GENE

MDM2

ALTERATION

amplification

RATIONALE

Inhibitors of the MDM2-p53 interaction are being tested in clinical trials. Overexpression or

amplification of MDM2 may increase sensitivity to these agents, but more data are required.

NCT04589845

PHASE 2

Tumor-Agnostic Precision Immuno-Oncology and Somatic Targeting Rational for You (TAPISTRY) Platform Study

TARGETS

ALK, ROS1, TRKA, TRKB, TRKC, RET, PD-L1, AKTs, ERBB2, MDM2, PI3K-alpha

LOCATIONS: Zhongzheng Dist. (Taiwan), Taipei City (Taiwan), Tainan (Taiwan), Seoul (Korea, Republic of), Beijing (China), Woolloongabba (Australia), Darlinghurst (Australia), Randwick (Australia), Melbourne (Australia), Haifa (Israel)

NCT03449381

PHASE 1

This Study Aims to Find the Best Dose of BI 907828 in Patients With Different Types of Advanced Cancer (Solid Tumors)

TARGETS

MDM2

LOCATIONS: Tokyo, Chuo-ku (Japan), Ottawa (Canada), Connecticut, New York, Tennessee, Florida

NCT03611868

PHASE 1/2

A Study of APG-115 in Combination With Pembrolizumab in Patients With Metastatic Melanomas or Advanced Solid Tumors

TARGETS

MDM2, PD-1

LOCATIONS: Brisbane (Australia), California, Arizona, Missouri, Arkansas, Pennsylvania, New York, Tennessee, Texas

NCT03725436

PHASE 1

ALRN-6924 and Paclitaxel in Treating Patients With Advanced, Metastatic, or Unresectable Solid Tumors

TARGETS

MDM2, MDM4

LOCATIONS: Texas

ORDERED TEST # ORD-1173008-01

CLINICAL TRIALS
GENE
PTEN
ALTERATION
D116fs*18

RATIONALE
PTEN loss or inactivating mutations may lead to increased activation of the PI3K-AKT-mTOR pathway and may indicate sensitivity to inhibitors

of this pathway. PTEN loss or inactivation may also predict sensitivity to PARP inhibitors.

NCT04337463
PHASE NULL

ATG-008 Combined With Toripalimab in Advanced Solid Tumors

TARGETS
mTORC1, mTORC2, PD-1

LOCATIONS: Chongqing (China), Chengdu (China)

NCT02264678
PHASE 1/2

Ascending Doses of AZD6738 in Combination With Chemotherapy and/or Novel Anti Cancer Agents

TARGETS
ATR, PARP, PD-L1

LOCATIONS: Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Cambridge (United Kingdom), Withington (United Kingdom), London (United Kingdom), Sutton (United Kingdom), Villejuif (France), Saint Herblain (France), California

NCT04740190
PHASE 2

Talazoparib - Carboplatin for Recurrent High-grade Glioma With DDRd

TARGETS
PARP

LOCATIONS: Hong Kong (Hong Kong)

NCT04001569
PHASE 1/2

AZD8186 and Paclitaxel in Advanced Gastric Cancer

TARGETS
PI3K-beta

LOCATIONS: Seongnam-si (Korea, Republic of)

NCT04635631
PHASE 1

STUDY OF TALAZOPARIB MONOTHERAPY IN CHINESE PARTICIPANTS WITH ADVANCED SOLID TUMORS

TARGETS
PARP

LOCATIONS: Beijing (China), Changchun (China)

NCT03772561
PHASE 1

Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies

TARGETS
PARP, AKTs, PD-L1

LOCATIONS: Singapore (Singapore)

ORDERED TEST # ORD-1173008-01

CLINICAL TRIALS
NCT04801966
PHASE NULL

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

TARGETS
 CDK4, CDK6, PI3K-alpha, PD-L1, MEK,
 PARP, PD-1, BRAF

LOCATIONS: Melbourne (Australia)

NCT03994796
PHASE 2

Genetic Testing in Guiding Treatment for Patients With Brain Metastases

TARGETS
 ALK, ROS1, TRKA, TRKB, TRKC, CDK4,
 CDK6, PI3K, mTOR

LOCATIONS: Alaska, Washington

NCT04632992
PHASE 2

A Study Evaluating Targeted Therapies in Participants Who Have Advanced Solid Tumors With Genomic Alterations or Protein Expression Patterns Predictive of Response

TARGETS
 ALK, ROS1, TRKA, TRKB, TRKC, PD-L1,
 ERBB2, ERBB3, PI3K-alpha, RET, AKTs

LOCATIONS: Alaska, Washington, Oregon, California, Montana

NCT04497116
PHASE 1/2

Study of RP-3500 in Advanced Solid Tumors

TARGETS
 PARP

LOCATIONS: Copenhagen (Denmark), Newcastle Upon Tyne (United Kingdom), Manchester (United Kingdom), London (United Kingdom), Toronto (Canada), Massachusetts, Rhode Island, New York, Tennessee, Texas

ORDERED TEST # ORD-1173008-01

APPENDIX
Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

ATM
Q95K

EGFR
rearrangement

MDM2
rearrangement and
rearrangement

NOTCH3
R1175W

PDCD1LG2 (PD-L2)
F236L

STK11
F354L

ORDERED TEST # ORD-1173008-01

APPENDIX
Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTB	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL	CCND1
CCND2	CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2	DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FAM46C	FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3
FGFR4	FH	FLCN	FLT1	FLT3	FOXO2	FUBP1	GABRA6	GATA3
GATA4	GATA6	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3F3A
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDMSA	KDMS5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKB1A	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3	P2RY8	PALB2
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA
PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET
RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2
SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOC3
SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU
SYK	TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53
TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1	WT1	XPO1
XRCC2	ZNF217	ZNF703						

DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSP02	SDC4	SLC34A2	TERC*	TERT**	TPRSS2

*TERC is an NCRNA

**Promoter region of TERT is interrogated

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Loss of Heterozygosity (LOH) score

Microsatellite (MS) status

Tumor Mutational Burden (TMB)

ORDERED TEST # ORD-1173008-01

APPENDIX

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.


ABOUT FOUNDATIONONE CDx

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:
www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLES

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials
Ranking of Therapies in Summary Table

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2021. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

1. The MSI-H/MSS designation by FMI F1CDx test is based on genome wide analysis of 95 microsatellite loci and not based on the 5 or 7 MSI loci described in current clinical practice guidelines. The threshold for MSI-H/MSS was determined by analytical concordance to comparator assays (IHC and PCR) using uterine, cecum and colorectal cancer FFPE tissue. The clinical validity of the qualitative MSI designation has not been established. For Microsatellite Instability (MSI) results, confirmatory testing using a validated orthogonal method should be considered.
2. TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics

© 2021 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 07 September 2021
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Shakti Ramkissoon, M.D., Ph.D., M.M. Sc, Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1173008-01

APPENDIX

About FoundationOne®CDx

of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.

- The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
- Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
- Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine whether the patient is a candidate for biopsy.
- Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive,

and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of *HER2* overexpression as 20% in breast cancer. Based on the F1CDx *HER2* CDx concordance study, approximately 10% of *HER2* amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

*Interquartile Range = 1st Quartile to 3rd Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*,

RAD51C, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating

ORDERED TEST # ORD-1173008-01

APPENDIX

About FoundationOne®CDx

physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/m	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

MR Suite Version 5.0.0

The median exon coverage for this sample is 468x

© 2021 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 07 September 2021
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Shakti Ramkissoon, M.D., Ph.D., M.M. Sc, Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1173008-01

APPENDIX
References

1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) PMID: 25392179
2. Kroemer G, et al. Oncoimmunology (2015) PMID: 26140250
3. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
4. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
5. Ayers et al., 2016; ASCO-SITC Abstract P60
6. Alonso M, et al. Cancer Res. (2001) PMID: 11280776
7. Rodríguez-Hernández I, et al. PLoS ONE (2013) PMID: 24073290
8. Vladimirova V, et al. Neuropathol. Appl. Neurobiol. (2008) PMID: 18053027
9. Martínez R, et al. Oncology (2004) PMID: 15331927
10. Martínez R, et al. J. Cancer Res. Clin. Oncol. (2005) PMID: 15672285
11. Martínez R, et al. Cancer Genet. Cytogenet. (2007) PMID: 17498554
12. Szybka M, et al. Clin. Neuropathol. () PMID: 12908754
13. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
14. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
15. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
16. Boland CR, et al. Cancer Res. (1998) PMID: 9823339
17. Pawlik TM, et al. Dis. Markers (2004) PMID: 15528785
18. Boland CR, et al. Gastroenterology (2010) PMID: 20420947
19. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
20. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
21. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
22. Cristescu R, et al. Science (2018) PMID: 30309915
23. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
24. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
25. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
26. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
27. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
28. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
29. Zhao J, et al. Nat. Med. (2019) PMID: 30742119
30. Touat M, et al. Nature (2020) PMID: 32322066
31. Bouffet E, et al. J. Clin. Oncol. (2016) PMID: 27001570
32. Johanns TM, et al. Cancer Discov (2016) PMID: 27683556
33. Lukas RV, et al. J. Neurooncol. (2018) PMID: 30073642
34. Chalmers ZR, et al. Genome Med (2017) PMID: 28420421
35. Patel RR, et al. Pediatr Blood Cancer (2020) PMID: 32386112
36. Johnson A, et al. Oncologist (2017) PMID: 28912153
37. Draaisma K, et al. Acta Neuropathol Commun (2015) PMID: 26699864
38. Wang L, et al. BMC Cancer (2020) PMID: 32164609
39. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
40. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
41. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
42. Rizvi NA, et al. Science (2015) PMID: 25765070
43. Johnson BE, et al. Science (2014) PMID: 24336570
44. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
45. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
46. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
47. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
48. Nature (2012) PMID: 22810696
49. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
50. Makhlin I, et al. CNS Oncol (2019) PMID: 31769726
51. Sepúlveda-Sánchez JM, et al. Neuro-oncology (2017) PMID: 28575464
52. Tanaka S, et al. Sci Rep (2019) PMID: 30644426
53. Chi AS, et al. JCO Precis Oncol (2020) PMID: 32923886
54. Reardon DA, et al. Neuro-oncology (2015) PMID: 25140039
55. Blumenthal DT, et al. J. Neurooncol. (2016) PMID: 27531351
56. Alshami J, et al. Oncotarget (2015) PMID: 26423602
57. Gan et al., 2015; ASCO Abstract 2016
58. Mellinghoff IK, et al. N. Engl. J. Med. (2005) PMID: 16282176
59. Clark PA, et al. Neoplasia (2012) PMID: 22745588
60. Fenton TR, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) PMID: 22891331
61. van den Bent MJ, et al. J Clin Oncol (2009) PMID: 19204207
62. Haas-Kogan DA, et al. J Natl Cancer Inst (2005) PMID: 15956649
63. Brown PD, et al. J Clin Oncol (2008) PMID: 18955445
64. Preusser M, et al. J Neurooncol (2008) PMID: 18458820
65. Wen PY, et al. Neuro-oncology (2014) PMID: 24470557
66. Gallego O, et al. J Neurooncol (2014) PMID: 24352766
67. Uhm JH, et al. Int J Radiat Oncol Biol Phys (2011) PMID: 20510539
68. Doyle SP, et al. Oxf Med Case Reports (2018) PMID: 30410775
69. D'Alessandris QG, et al. Acta Neurochir (Wien) (2013) PMID: 23132371
70. Custodio A, et al. Clin Transl Oncol (2010) PMID: 20462843
71. D'Alessandris QG, et al. Acta Neurochir (Wien) (2018) PMID: 30306271
72. Mellinghoff IK, et al. Clin Cancer Res (2007) PMID: 17255257
73. Arif SH, et al. Asian J Neurosurg () PMID: 29492119
74. Franceschi E, et al. Br. J. Cancer (2007) PMID: 17353924
75. Chakravarti A, et al. Int. J. Radiat. Oncol. Biol. Phys. (2013) PMID: 23182702
76. Hegi ME, et al. Mol. Cancer Ther. (2011) PMID: 21471286
77. Wang J, et al. Int. J. Cancer (2019) PMID: 30255937
78. Jänne PA, et al. N. Engl. J. Med. (2015) PMID: 25923549
79. Soria JC, et al. N. Engl. J. Med. (2018) PMID: 29151359
80. Pirker R, et al. Lancet Oncol. (2012) PMID: 22056021
81. Jiang Z, et al. PLoS ONE (2013) PMID: 23441167
82. Licitra L, et al. Ann. Oncol. (2011) PMID: 21048039
83. Herbst RS, et al. Lancet Oncol. (2018) PMID: 29169877
84. Paz-Ares L, et al. Ann. Oncol. (2016) PMID: 27207107
85. Thatcher N, et al. Lancet Oncol. (2015) PMID: 26045340
86. Paz-Ares L, et al. Lancet Oncol. (2015) PMID: 25701171
87. Elez E, et al. Br. J. Cancer (2016) PMID: 26766738
88. Kuenen B, et al. Clin. Cancer Res. (2010) PMID: 20197484
89. Shimamura T, et al. Cancer Res. (2005) PMID: 16024644
90. Shimamura T, et al. Cancer Res. (2008) PMID: 18632637
91. Sawai A, et al. Cancer Res. (2008) PMID: 18199556
92. Bernardes CE, et al. J Phys Condens Matter (2015) PMID: 25923649
93. Xu W, et al. Br. J. Cancer (2007) PMID: 17712310
94. Zeng Q, et al. J. Med. Chem. (2015) PMID: 26313252
95. Yang Z, et al. Sci Transl Med (2016) PMID: 27928026
96. Ahn et al., 2019; ASCO 31587882
97. Strong JE, et al. EMBO J. (1998) PMID: 9628872
98. Coffey MC, et al. Science (1998) PMID: 9812900
99. Gong J, et al. Front Oncol (2014) PMID: 25019061
100. Forsyth P, et al. Mol. Ther. (2008) PMID: 18253152
101. Vidal L, et al. Clin. Cancer Res. (2008) PMID: 18981012
102. Gollamudi R, et al. Invest New Drugs (2010) PMID: 19572105
103. Harrington KJ, et al. Clin. Cancer Res. (2010) PMID: 20484020
104. Comins C, et al. Clin. Cancer Res. (2010) PMID: 20926400
105. Lolkema MP, et al. Clin. Cancer Res. (2011) PMID: 21106728
106. Galanis E, et al. Mol. Ther. (2012) PMID: 22871663
107. Karapanagiotou EM, et al. Clin. Cancer Res. (2012) PMID: 22316603
108. Morris DG, et al. Invest New Drugs (2013) PMID: 22886613
109. Jonsson P, et al. Clin. Cancer Res. (2019) PMID: 31263031
110. Brennan CW, et al. Cell (2013) PMID: 24120142
111. Ceccarelli M, et al. Cell (2016) PMID: 26824661
112. Thomas AA, et al. Neuro-oncology (2017) PMID: 28472509
113. Smith JS, et al. J. Natl. Cancer Inst. (2001) PMID: 11504770
114. Shinojima N, et al. Cancer Res. (2003) PMID: 14583498
115. Ambrose MM, et al. Asian Pac. J. Cancer Prev. (2010) PMID: 21133628
116. Hobbs J, et al. Am. J. Surg. Pathol. (2012) PMID: 22472960
117. Ciardiello F, et al. N. Engl. J. Med. (2008) PMID: 18337605
118. Liang Z, et al. BMC Cancer (2010) PMID: 20637128
119. Bhargava R, et al. Mod. Pathol. (2005) PMID: 15920544
120. Yang YL, et al. Chin. Med. J. (2012) PMID: 22490401
121. Cho J, et al. Cancer Res. (2011) PMID: 22001862
122. Imielinski M, et al. Cell (2012) PMID: 22980975
123. Pines G, et al. Oncogene (2010) PMID: 20676128
124. Wang Q, et al. Exp. Cell Res. (2007) PMID: 17643422
125. Frattini V, et al. Nat. Genet. (2013) PMID: 23917401
126. Konduri K, et al. Cancer Discov (2016) PMID: 27102076
127. Dickson MA, et al. J. Clin. Oncol. (2013) PMID: 23569312
128. Flaherty KT, et al. Clin. Cancer Res. (2012) PMID: 22090362
129. Patnaik A, et al. Cancer Discov (2016) PMID: 27217383
130. Infante JR, et al. Clin. Cancer Res. (2016) PMID: 27542767
131. Dickson et al., 2019; ASCO Abstract 11004
132. Dickson MA, et al. JAMA Oncol (2016) PMID: 27124835
133. Peguero et al., 2016; ASCO Abstract 2528
134. Costa C, et al. Cancer Discov (2019) PMID: 31594766
135. Cerami E, et al. Cancer Discov (2012) PMID: 22588877
136. Gao J, et al. Sci Signal (2013) PMID: 23550210
137. Zheng S, et al. Genes Dev. (2013) PMID: 23796897
138. Kim H, et al. Proc. Natl. Acad. Sci. U.S.A. (2010) PMID: 20080666
139. Ruano Y, et al. Am. J. Clin. Pathol. (2009) PMID: 19141386
140. Fischer U, et al. Mol. Cancer Res. (2008) PMID: 18403636
141. Bäcklund LM, et al. Br. J. Cancer (2005) PMID: 15970925
142. Choi YJ, et al. Oncogene (2014) PMID: 23644662
143. Cell (1995) PMID: 7736585
144. Musgrave EA, et al. Nat. Rev. Cancer (2011) PMID: 21734724
145. Wikman H, et al. Genes Chromosomes Cancer (2005) PMID: 15543620
146. Rao SK, et al. J. Neurooncol. (2010) PMID: 19609742

© 2021 Foundation Medicine, Inc. All rights reserved.

 Electronically signed by Erik Williams, M.D. | 07 September 2021
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Shakti Ramkissoon, M.D., Ph.D., M.M. Sc, Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

 Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1173008-01

APPENDIX
References

147. Chung L, et al. Am. J. Surg. Pathol. (2009) pmid: 19574885
148. Ragazzini P, et al. Histol. Histopathol. (2004) pmid: 15024701
149. Dujardin F, et al. Mod. Pathol. (2011) pmid: 21336260
150. Zhang K, et al. Cancer Res. (2013) pmid: 23393200
151. Horvai AE, et al. Mod. Pathol. (2009) pmid: 19734852
152. Cheok CF, et al. Nat Rev Clin Oncol (2011) pmid: 20975744
153. Ohnstad HO, et al. Cancer (2013) pmid: 23165797
154. Gamble LD, et al. Oncogene (2012) pmid: 21725357
155. Zhang et al., 2019; ASCO Abstract 3124
156. Rasco et al., 2019; ASCO Abstract 3126
157. Tolcher et al., 2021; ASCO Abstract 2506
158. Martinelli et al., 2016; EHA21 Abstract S504
159. Daver et al., 2018; ASH Abstract 767
160. Mascarenhas et al., 2019; ASH Abstract 134
161. Shustov et al., 2018; ASH Abstract 1623
162. Sallman et al., 2018; ASH Abstract 4066
163. Meric-Bernstam et al., 2017; ASCO Abstract 2505
164. Fischer U, et al. Int. J. Cancer (2010) pmid: 19839052
165. Sdek P, et al. Mol. Cell (2005) pmid: 16337594
166. Brady M, et al. Mol. Cell. Biol. (2005) pmid: 15632057
167. Li M, et al. Mol. Cell (2004) pmid: 15053880
168. Brown CJ, et al. Nat. Rev. Cancer (2009) pmid: 19935675
169. Cordon-Cardo C, et al. Cancer Res. (1994) pmid: 8306343
170. Beroukhi R, et al. Nature (2010) pmid: 20164920
171. Kato S, et al. Clin. Cancer Res. (2017) pmid: 28351930
172. Singavi et al., 2017; ESMO Abstract 1140PD
173. Rizvi H, et al. J. Clin. Oncol. (2018) pmid: 29337640
174. Courtney KD, et al. J. Clin. Oncol. (2010) pmid: 20085938
175. Simpson L, et al. Exp. Cell Res. (2001) pmid: 11237521
176. Patnaik A, et al. Ann. Oncol. (2016) pmid: 27672108
177. Milella M, et al. Sci Rep (2017) pmid: 28220839
178. Templeton AJ, et al. Eur. Urol. (2013) pmid: 23582881
179. Sweeney C, et al. Lancet (2021) pmid: 34246347
180. de Bono JS, et al. Clin. Cancer Res. (2019) pmid: 30037818
181. Saura C, et al. Cancer Discov (2017) pmid: 27872130
182. Voss MH, et al. Clin. Cancer Res. (2018) pmid: 30327302
183. André F, et al. J. Clin. Oncol. (2016) pmid: 27091708
184. Schmid P, et al. J. Clin. Oncol. (2019) pmid: 31841354
185. Weldon Gilcrease G, et al. Invest New Drugs (2019) pmid: 30302599
186. Mendes-Pereira AM, et al. EMBO Mol Med (2009) pmid: 20049735
187. Shen Y, et al. Clin. Cancer Res. (2013) pmid: 23881923
188. Chatterjee P, et al. PLoS ONE (2013) pmid: 23565244
189. McCormick A, et al. Int. J. Gynecol. Cancer (2016) pmid: 26905328
190. Forster MD, et al. Nat Rev Clin Oncol (2011) pmid: 21468130
191. Eikesdal HP, et al. Ann Oncol (2021) pmid: 33242536
192. Dougherty et al., 2014; ASCO Abstract 5536
193. Pan M, et al. Perm J (2021) pmid: 33970096
194. Sandhu SK, et al. Lancet Oncol. (2013) pmid: 23810788
195. Romero I, et al. Gynecol Oncol (2020) pmid: 32988624
196. George S, et al. Immunity (2017) pmid: 28228279
197. Peng W, et al. Cancer Discov (2016) pmid: 26645196
198. Zhou XP, et al. Int. J. Cancer (1999) pmid: 10096247
199. Rasheed BK, et al. Cancer Res. (1997) pmid: 9331072
200. Davies MP, et al. Br. J. Cancer (1999) pmid: 10188904
201. Lin H, et al. Clin. Cancer Res. (1998) pmid: 9796977
202. Schmidt EE, et al. J. Neuropathol. Exp. Neurol. (1999) pmid: 10560660
203. Kato H, et al. Clin. Cancer Res. (2000) pmid: 11051241
204. Furnari FB, et al. Genes Dev. (2007) pmid: 17974913
205. Yan et al. 2020; DOI:10.1200/PO.19.00385
206. Cancer Genome Atlas Research Network, et al. N. Engl. J. Med. (2015) pmid: 26061751
207. Srividya MR, et al. Neuropathology (2011) pmid: 21134002
208. Campbell RB, et al. J. Biol. Chem. (2003) pmid: 12857747
209. Rodriguez-Escudero I, et al. Hum. Mol. Genet. (2011) pmid: 21828076
210. He X, et al. Cancer Res. (2013) pmid: 23475934
211. Han SY, et al. Cancer Res. (2000) pmid: 10866302
212. Myers MP, et al. Proc. Natl. Acad. Sci. U.S.A. (1998) pmid: 9811831
213. Pradella LM, et al. BMC Cancer (2014) pmid: 24498881
214. Kim JS, et al. Mol. Cell. Biol. (2011) pmid: 21536651
215. Denning G, et al. Oncogene (2007) pmid: 17213812
216. Hlobilkova A, et al. Anticancer Res. (2010) pmid: 16619501
217. Redfern RE, et al. Protein Sci. (2010) pmid: 20718038
218. Shenoy S, et al. PLoS ONE (2012) pmid: 22505997
219. Wang Y, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) pmid: 19329485
220. Okumura K, et al. J. Biol. Chem. (2006) pmid: 16829519
221. Lee JO, et al. Cell (1999) pmid: 10555148
222. Maxwell GL, et al. Cancer Res. (1998) pmid: 9635567
223. Risinger JI, et al. Clin. Cancer Res. (1998) pmid: 9865913
224. Ngeow J, et al. J. Clin. Endocrinol. Metab. (2012) pmid: 23066114
225. Lobo GP, et al. Hum. Mol. Genet. (2009) pmid: 19457929
226. Liu J, et al. Oncogene (2014) pmid: 23995781
227. Maehama T, et al. Annu. Rev. Biochem. (2001) pmid: 11395408
228. De Vivo I, et al. J. Med. Genet. (2000) pmid: 10807691
229. Ramaswamy S, et al. Proc. Natl. Acad. Sci. U.S.A. (1999) pmid: 10051603
230. Liu JL, et al. Mol. Cell. Biol. (2005) pmid: 15988030
231. Karoui M, et al. Br. J. Cancer (2004) pmid: 15026806
232. Gil A, et al. PLoS ONE (2015) pmid: 25875300
233. Furnari FB, et al. Cancer Res. (1998) pmid: 9823298
234. Spinelli L, et al. J. Med. Genet. (2015) pmid: 25527629
235. Mingo J, et al. Eur. J. Hum. Genet. (2018) pmid: 29706633
236. Wang Q, et al. J. Mol. Graph. Model. (2010) pmid: 20538496
237. Andrés-Pons A, et al. Cancer Res. (2007) pmid: 17942903
238. Butler MG, et al. J. Med. Genet. (2005) pmid: 15805158
239. Georgescu MM, et al. Proc. Natl. Acad. Sci. U.S.A. (1999) pmid: 10468583
240. Staal FJ, et al. Br. J. Cancer (2002) pmid: 12085208
241. Nguyen HN, et al. Oncogene (2014) pmid: 24292679
242. Rahdar M, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) pmid: 19114656
243. Das S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) pmid: 12808147
244. Wang X, et al. Biochem. J. (2008) pmid: 18498243
245. Valiente M, et al. J. Biol. Chem. (2005) pmid: 15951562
246. Nguyen HN, et al. Oncogene (2015) pmid: 25263454
247. Blumenthal GM, et al. Eur. J. Hum. Genet. (2008) pmid: 18781191
248. Orloff MS, et al. Oncogene (2008) pmid: 18794875
249. Zbuk KM, et al. Nat. Rev. Cancer (2007) pmid: 17167516
250. Hasselbalch B, et al. Neuro-oncology (2010) pmid: 20406901
251. Gajadhar AS, et al. Mol. Cancer Res. (2012) pmid: 22232519
252. Lv S, et al. Int. J. Oncol. (2012) pmid: 22752145
253. Whittle JR, et al. J Clin Neurosci (2015) pmid: 26279503
254. Jensen LH, et al. Ann. Oncol. (2012) pmid: 22367707
255. Sohal DP, et al. Ann. Oncol. (2013) pmid: 24146220
256. Crawford J, et al. J Thorac Oncol (2013) pmid: 24389433
257. Rowinsky EK, et al. J. Clin. Oncol. (2004) pmid: 15210739