

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT	DISEASE Brain anaplastic oligodendroglioma	PHYSICIAN	ORDERING PHYSICIAN Yeh, Yi-Chen	SPECIMEN	SPECIMEN SITE Brain
	NAME Wu, Cheng-Yen		MEDICAL FACILITY Taipei Veterans General Hospital		SPECIMEN ID S111-21010 A (PF22067)
	DATE OF BIRTH 04 February 1994		ADDITIONAL RECIPIENT None		SPECIMEN TYPE Slide Deck
	SEX Male		MEDICAL FACILITY ID 205872		DATE OF COLLECTION 27 May 2022
	MEDICAL RECORD # 41441638		PATHOLOGIST Not Provided		SPECIMEN RECEIVED 17 June 2022

Biomarker Findings

Microsatellite status - MS-Stable
Tumor Mutational Burden - 0 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

IDH2 R172K

NOTCH1 C750fs*1, F357del, duplication exons 8-25

TERT promoter -124C>T

Report Highlights

- Variants with **diagnostic implications** that may indicate a specific cancer type: **IDH2** R172K (p. 3), **TERT** promoter -124C>T (p. 5)
- Variants that may inform **nontargeted treatment approaches** (e.g., chemotherapy) in this tumor type: **IDH2** R172K (p. 3)
- Evidence-matched **clinical trial options** based on this patient's genomic findings: (p. 6)
- Variants with **prognostic implications** for this tumor type that may impact treatment decisions: **IDH2** R172K (p. 3), **TERT** promoter -124C>T (p. 5)

BIOMARKER FINDINGS

Microsatellite status - MS-Stable

Tumor Mutational Burden - 0 Muts/Mb

GENOMIC FINDINGS

IDH2 - R172K

1 Trial see p. 6

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. see Biomarker Findings section

No therapies or clinical trials. see Biomarker Findings section

**THERAPIES WITH CLINICAL
RELEVANCE
(IN PATIENT'S TUMOR TYPE)**

none

**THERAPIES WITH CLINICAL
RELEVANCE
(IN OTHER TUMOR TYPE)**

none

GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

NOTCH1 - C750fs*1, F357del, duplication exons

8-25 p. 4

TERT - promoter -124C>T

..... p. 5

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 27 June 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1391890-01

BIOMARKER FINDINGS

BIOMARKER

Microsatellite status

RESULT

MS-Stable

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁻³, including approved therapies nivolumab and pembrolizumab⁴. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, $p=0.001$)⁵.

FREQUENCY & PROGNOSIS

Low-level MSI has been reported in 5-9% of glioblastoma (GBM) samples⁶⁻⁸. A large-scale study did not find high-level microsatellite instability (MSI-H) in any of 129 GBM samples⁶, although a small-scale study reported MSI-H in 4 of 15 pediatric GBMs and 1 of 12 adult GBMs⁹. A pediatric patient with Lynch syndrome but MSS was reported to develop anaplastic oligodendroglioma¹⁰. The frequency of MSI has been reported to be increased in relapsed compared to primary GBM⁶, in GBMs with a previous lower grade astrocytoma⁷, and in giant cell GBM compared to classic GBM⁸. A case study of a pediatric patient with glioblastoma with an oligodendroglioma component was reported to harbor MSI-high, which was proposed to be a possible mechanism of resistance to temozolomide treatment¹¹.

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor¹². Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2¹²⁻¹⁴. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers¹⁵⁻¹⁷. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{12,14,16-17}.

BIOMARKER

Tumor Mutational Burden

RESULT

0 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1¹⁸⁻²⁰, anti-PD-1 therapies¹⁸⁻²¹, and combination nivolumab and ipilimumab²²⁻²⁷. In glioma, a lack of association between TMB and clinical benefit from immune checkpoint inhibitors has been reported^{18,28-29}. However, multiple case studies have reported that patients with ultramutated gliomas driven by POLE

mutations have benefited from treatment with anti-PD-1³⁰⁻³¹ or anti-PD-L1³² therapies. Therefore, although increased TMB alone may not be a strong biomarker for PD-1 or PD-L1 inhibitors in this cancer type, these agents may have efficacy for patients with glioma harboring both high TMB and POLE mutation.

FREQUENCY & PROGNOSIS

The median TMB of oligodendrogliomas is 2.7 mutations per megabase (mut/Mb), and 8.4% of cases have high TMB (>20 muts/Mb)³³. For pediatric patients, high TMB has been reported in a subset of high-grade gliomas, frequently in association with mutations in mismatch repair or proofreading genes and in TP53, whereas BRAF alterations or other oncogene fusions were observed more frequently in brain tumors harboring low TMB³⁴⁻³⁵. Increased TMB has been reported to correlate with higher tumor grade in glioma³⁶ and glioblastoma (GBM) tissue samples with biallelic mismatch repair deficiency

(bMMRD)³⁰, as well as with shorter OS of patients with diffuse glioma³⁷.

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma³⁸⁻³⁹ and cigarette smoke in lung cancer⁴⁰⁻⁴¹, treatment with temozolomide-based chemotherapy in glioma⁴²⁻⁴³, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes⁴⁴⁻⁴⁸, and microsatellite instability (MSI)^{44,47-48}. This sample harbors a TMB below levels that would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents^{18,28-32}.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 27 June 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1391890-01

GENOMIC FINDINGS

GENE

IDH2

ALTERATION

R172K

TRANSCRIPT ID

NM_002168

CODING SEQUENCE EFFECT

515G>A

VARIANT ALLELE FREQUENCY (% VAF)

38.6%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of preclinical data and clinical responses for patients with acute myeloid leukemia (AML), IDH2 mutations may predict response to selective IDH2 inhibitors such as enasidenib⁴⁹⁻⁵¹. In Phase 1/2 studies of enasidenib for patients with IDH2-mutated advanced hematological malignancies, overall response rates of 40% and 53% were achieved for patients with relapsed/refractory AML and myelodysplastic syndrome (MDS), respectively⁴⁹. On the basis of extensive clinical data in AML and limited data in MDS, IDH2 mutations may also predict response to the combination of the BCL-2 inhibitor venetoclax and a DNA methyltransferase inhibitor such as azacitidine or decitabine⁵²⁻⁵⁸, or to single-agent venetoclax⁵⁹⁻⁶⁰, azacitidine, or decitabine⁶¹⁻⁶⁶. Limited clinical data also indicate that IDH2 mutations may predict sensitivity to the

combination of enasidenib and azacitidine⁶⁷ or to glutaminase inhibitors such as telaglenastat⁶⁸. A patient with previously treated IDH2-mutated AML experienced a CR longer than 14 months with incomplete hematologic recovery on telaglenastat monotherapy⁶⁸.

— Nontargeted Approaches —

IDH1/2 mutations are associated with improved survival outcomes for patients with glioma treated with radiation or alkylating chemotherapy (NCCN CNS Cancers Guidelines, v2.2021).

FREQUENCY & PROGNOSIS

IDH2 mutations have been reported in 5-7% of oligodendrogliomas analyzed in the COSMIC database, nearly all of which were substitutions at R172 (COSMIC, Jun 2022)⁶⁹. Another large study in glioma reported IDH2 mutation in 2.5% (5/203) of cases⁷⁰. Studies have reported IDH mutation (IDH1 or IDH2) in 54% (23/43) and 100% (16/16) of oligodendrogliomas analyzed, although IDH1 mutation was more prevalent, with IDH2 in just 1/43 and 2/16 samples⁷¹⁻⁷². Studies have reported that IDH mutation (IDH1 or IDH2) was associated with improved overall survival in glioma, including oligodendroglioma^{70-71,73}. This improvement in overall survival may be due to increased radiation sensitivity in gliomas with IDH mutation⁷⁴. In the context of IDH-mutated gliomas, TERT mutations are associated with improved OS (NCCN CNS Cancers Guidelines, v2.2021)⁷⁵. IDH1/2 mutations are a strong favorable prognostic marker for OS in Grade 2-3

glioma, particularly in combination with 1p/19q codeletion (NCCN CNS Cancers Guidelines, v2.2021).

FINDING SUMMARY

The isocitrate dehydrogenases IDH1 and IDH2 encode highly homologous enzymes that are involved in the citric acid (TCA) cycle and other metabolic processes, playing roles in normal cellular metabolism and in protection against oxidative stress and apoptosis⁷⁶. Amino acids 140 and 172 are hotspots for cancer-related mutations in IDH2⁷⁷. Functional studies have reported that mutation of R140 or R172, such as observed here, alters IDH2 enzymatic activity, resulting in gain-of-function activity and the production of the potential oncometabolite, D-2-hydroxyglutarate (2-HG)⁷⁶⁻⁸¹. This leads to downstream effects that are associated with tumorigenesis^{79,82}, and research suggests that hotspot IDH gene mutations could be early stage events in specific cancers⁸²⁻⁸³.

POTENTIAL DIAGNOSTIC IMPLICATIONS

Co-occurring TERT mutation, IDH mutation, and 1p/19q co-deletion is indicative of oligodendroglioma (NCCN CNS Cancers Guidelines, v2.2021)⁸⁴. IDH1/2 mutation is associated with Grade 2 and 3 astrocytomas and oligodendrogliomas, and distinguishes secondary glioblastoma (GBM) from primary GBM (NCCN CNS Cancers Guidelines, v2.2021).

ORDERED TEST # ORD-1391890-01

GENOMIC FINDINGS

GENE

NOTCH1

ALTERATION

C750fs*1, F357del, duplication exons 8-25

TRANSCRIPT ID

NM_017617, NM_017617

CODING SEQUENCE EFFECT

2250_2251delTG, 1070_1072delTCT

VARIANT ALLELE FREQUENCY (% VAF)

14.8%, 28.3%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

NOTCH1 inhibitors and gamma-secretase inhibitors (GSIs) may be potential therapeutic approaches in the case of NOTCH1 activating mutations⁸⁵⁻⁹³. In a Phase 2 study, the GSI AL101 (BMS-906024) elicited PRs in 15% (6/39) and SDs in 54% (21/39) of patients with metastatic adenoid cystic carcinoma (ACC) harboring NOTCH1 activating alterations⁹⁴. Additional responses to AL101 have been reported in a patient with gastroesophageal junction adenocarcinoma harboring multiple NOTCH1 mutations, a patient with T-cell acute lymphoblastic leukemia (T-ALL) harboring a NOTCH1 HD domain mutation, and a patient with ACC harboring a single NOTCH1 mutation⁹⁵. A Phase 1 study of the pan-NOTCH inhibitor CB-103 for patients with advanced or recurrent solid tumors reported a preliminary mPFS of 21.7 weeks for patients with ACC, with 2 patients harboring NOTCH1-mutated ACC demonstrating SD > 6 months as best response⁹⁶. On the basis of clinical data in non-Hodgkin lymphoma, NOTCH1 activating alterations may be

associated with sensitivity to the approved PI3K inhibitor copanlisib⁹⁷; this is further supported by limited preclinical data that suggest that NOTCH1 may be a negative regulator of PTEN⁹⁸⁻⁹⁹. It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here. While activating mutations may be targeted via gamma-secretase inhibitors or PI3K inhibitors, there are no therapies available to address NOTCH1 inactivation, as seen here.

FREQUENCY & PROGNOSIS

NOTCH1 mutation has been observed in 11% of lower grade glioma cases analyzed^{42,100}. NOTCH1 mutation is rare in glioblastoma and has been observed in 0-1.7% of cases, but has been reported in 19-27% of oligodendrogliomas (COSMIC, cBioPortal, Mar 2022)^{69,72,101-104}. NOTCH1 expression has been observed in 60%, 67-68%, 85-93%, and 92-100% of grades 1, 2, 3, and 4 astrocytoma samples, respectively¹⁰⁵⁻¹⁰⁶. In one study of 18 pilocytic astrocytomas (PA) and 4 low grade astrocytomas, NOTCH1 mRNA over-expression was observed in 95% (21/22) of cases¹⁰⁷. In addition, the NOTCH1 target, HES1, exhibited moderate to strong protein expression via immunohistochemistry in 66% (40/61) PA samples compared to non-neoplastic brain samples¹⁰⁷. NOTCH1 expression was significantly increased from Grade 1 to Grade 4 gliomas and was associated with glioma progression¹⁰⁸, as well as with tumor grade and tumor proliferation activity in astrocytoma¹⁰⁵⁻¹⁰⁶.

FINDING SUMMARY

NOTCH1 encodes a member of the NOTCH family of receptors, which are involved in cell fate

determination and various developmental processes. Depending on cellular context, NOTCH1 can act as either a tumor suppressor or an oncogene¹⁰⁹⁻¹¹⁰. Upon binding of membrane-bound ligands, the NOTCH1 intracellular domain (NICD) is cleaved and forms part of a transcription factor complex that regulates downstream target genes involved in cell fate determination, proliferation, and apoptosis¹¹¹⁻¹¹². NOTCH1 rearrangements and fusions, including SEC16A-NOTCH1, that eliminate the NOTCH1 extracellular domain but retain the transmembrane and intracellular domains have been observed in patients with breast cancer¹¹³⁻¹¹⁴ and T-cell acute lymphoblastic leukemia (T-ALL)¹¹⁵, and have been reported to cause ligand-independent activation of NOTCH1 transcriptional activity and are gamma-secretase inhibitor (GSI)-sensitive¹¹⁴⁻¹¹⁸. One or more alterations observed here have not been fully characterized or form fusions with non-canonical breakpoints, and the effect on NOTCH1 function is unclear; however, similar NOTCH1 rearrangements have been reported in the context of cancer, which may indicate biological relevance. NOTCH1 alterations that disrupt ligand binding¹¹⁹⁻¹²¹ or remove the transmembrane domain (amino acids 1736-1756), RAM domain (amino acids 1757-1926), ankyrin repeats (amino acids 1927-2122) and/or transactivation domain (amino acids 2123-2374) that are necessary for NOTCH1 function, such as observed here, are predicted to be inactivating^{112,122-124}. Several point mutations, including D469G, A465T, C478F, R1594Q, and P1770S, have also been reported to inactivate NOTCH1^{109,125-126}. One or more of the NOTCH1 alterations observed here have not been characterized, and the effect on NOTCH1 function is not known.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 27 June 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1391890-01

GENOMIC FINDINGS

GENE

TERT

ALTERATION

promoter -124C>T

TRANSCRIPT ID

NM_198253

CODING SEQUENCE EFFECT

-124C>T

VARIANT ALLELE FREQUENCY (% VAF)

45.3%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Therapeutic options for targeting tumors with TERT mutations are limited, although a variety of approaches have been investigated, including immunotherapies using TERT as a tumor-associated antigen and antisense oligonucleotide- or peptide-based therapies. TERT peptide vaccines showed limited anticancer efficacy in clinical trials¹²⁷; however, in one preclinical study, the combination of a TERT peptide vaccine and anti-CTLA-4 therapy suppressed tumor growth¹²⁸. A

Phase 2 study of the TERT inhibitor imetelstat for patients with advanced non-small cell lung cancer reported no improvement in PFS or OS¹²⁹.

FREQUENCY & PROGNOSIS

TERT promoter mutations have been reported in 51-59% of gliomas¹³⁰⁻¹³¹, most frequently in glioblastoma (GBM, 54-84%), gliosarcoma (81%), oligodendroglioma (78%), and historically in oligoastrocytomas (25-31%) but less frequently in lower grade astrocytomas (10-18%) and in only 1% of ependymomas¹³⁰⁻¹³⁴. In the context of IDH-mutated gliomas, TERT mutations are associated with improved OS (NCCN CNS Cancers Guidelines, v2.2021)⁷⁵. TERT promoter mutation has been shown to be significantly associated with increased TERT gene expression in astrocytoma, oligodendroglioma, and GBM¹³⁵. TERT promoter mutations significantly associate with poor prognosis in patients with GBM, although this correlation may be due to the association with primary GBM as opposed to IDH-positive secondary GBM^{130,132,135-136}. In the context of IDH-wildtype glioma, TERT mutations are associated with reduced OS (NCCN CNS Cancers Guidelines, v2.2021).

FINDING SUMMARY

Telomerase reverse transcriptase (TERT, or hTERT) is a catalytic subunit of the telomerase complex, which is required to maintain appropriate chromosomal length¹³⁷. Activation of TERT is a hallmark of cancer, being detected in up to 80-90% of malignancies and absent in quiescent cells¹³⁸⁻¹⁴⁰. Mutations within the promoter region of TERT that confer enhanced TERT promoter activity have been reported in two hotspots, located at -124 bp and -146 bp upstream of the transcriptional start site (also termed C228T and C250T, respectively)¹⁴¹⁻¹⁴³, as well as tandem mutations at positions -124/-125 bp and -138/-139 bp¹⁴¹.

POTENTIAL DIAGNOSTIC IMPLICATIONS

Co-occurring TERT mutation, IDH mutation, and 1p/19q co-deletion is indicative of oligodendroglioma (NCCN CNS Cancers Guidelines, v2.2021)⁸⁴. TERT mutations are associated with 1p/19q co-deletion in oligodendrogliomas, and are highly recurrent in IDH/ATRX-wildtype glioblastoma (GBM) (NCCN CNS Cancers Guidelines, v2.2021)⁸⁴.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 27 June 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1391890-01

CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov. Or visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

GENE
IDH2
RATIONALE
IDH2 mutations may predict sensitivity to IDH2 inhibitors.

ALTERATION
R172K

NCT04762602
PHASE 1

A Study of HMPL-306 in Advanced Solid Tumors With IDH Mutations

TARGETS
IDH2, IDH1

LOCATIONS: Barcelona (Spain), Madrid (Spain), California, Iowa, Pennsylvania, Kentucky, Texas, Georgia

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 27 June 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1391890-01

APPENDIX
Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

BRCA2
A2786T

KMT2A (MLL)
Y2473C

MET
L211W

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 27 June 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1391890-01

APPENDIX
Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B or WTX)	
APC	AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX
AURKA	AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2
BCL6	BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1
BTG2	BTK	CALR	CARD11	CASP8	CBFB	CBL	CCND1	CCND2
CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73	CDH1
CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B	CDKN2C
CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R	CTCF
CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1	DDR2
DIS3	DNMT3A	DOT1L	EED	EGFR	EMSY (C11orf30)	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12	FGF14
FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3	FGFR4
FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3	GATA4
GATA6	GID4 (C17orf39)	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3-3A (H3F3A)
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDM5A	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKKN1	MLH1	MPL	MRE11 (MRE11A)	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD2 (WHSC1 or MMSET)	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK1	NTRK2	NTRK3
P2RY8	PALB2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)
PDGFRA	PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1
PMS2	POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI
PRKN (PARK2)	PTCH1	PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51
RAD51B	RAD51C	RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10
REL	RET	RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC
SDHD	SETD2	SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO
SNCAIP	SOC1	SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3
STK11	SUFU	SYK	TBX3	TEK	TENTSC (FAM46C)	TET2	TGFB2	TIPARP
TNFAIP3	TNFRSF14	TP53	TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL
WT1	XPO1	XRCC2	ZNF217	ZNF703				

DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV1
ETV4	ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT
KMT2A (MLL)	MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA
RAF1	RARA	RET	ROS1	RSPO2	SDC4	SLC34A2	TERC*	TERT**
TPRSS2								

*TERC is an NCRNA

**Promoter region of TERT is interrogated

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Homologous Recombination status
Loss of Heterozygosity (LOH) score
Microsatellite (MS) status
Tumor Mutational Burden (TMB)

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 27 June 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1391890-01

APPENDIX

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.



ABOUT FOUNDATIONONE CDx

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:
www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLES

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials

Ranking of Therapies in Summary Table

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

1. In the fractional-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 27 June 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1391890-01

APPENDIX

About FoundationOne®CDx

- Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.
- TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
 - Homologous Recombination status may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas (Coleman et al., 2017; 28916367). Samples with deleterious *BRCA1/2* alteration and/or Loss of Heterozygosity (LOH) score $\geq 16\%$ will be reported as "HRD Positive" and samples with absence of these findings will be reported as "HRD Not Detected," agnostic of potential secondary *BRCA1/2* reversion alterations. Certain potentially deleterious missense or small in-frame deletions in *BRCA1/2* may not be classified as deleterious and, in the absence of an elevated LOH profile, samples with such mutations may be classified as "HRD Not Detected." A result of "HRD Not Detected" does not rule out the presence of a *BRCA1/2* alteration or an elevated LOH profile outside the assay performance characteristic limitations.
 - The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian,

peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.

- Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
- Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine whether the patient is a candidate for biopsy.
- Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of *HER2* overexpression as 20% in breast cancer. Based on the F1CDx *HER2* CDx concordance study, approximately 10% of *HER2* amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant

patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

*Interquartile Range = 1st Quartile to 3rd Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 27 June 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 | CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 | CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 | CLIA: 22D2027531

ORDERED TEST # ORD-1391890-01

APPENDIX
About FoundationOne®CDx

CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D (MLL2)*, *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking

into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

REFERENCE SEQUENCE INFORMATION

Sequence data is mapped to the human genome, Genome Reference Consortium Human Build 37 (GRCh37), also known as hg19.

MR Suite Version 6.3.0

The median exon coverage for this sample is 757x

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 27 June 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1391890-01

APPENDIX
References

1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) PMID: 25392179
2. Kroemer G, et al. Oncoimmunology (2015) PMID: 26140250
3. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
4. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
5. Ayers et al., 2016; ASCO-SITC Abstract P60
6. Martinez R, et al. Oncology (2004) PMID: 15331927
7. Martinez R, et al. J. Cancer Res. Clin. Oncol. (2005) PMID: 15672285
8. Martinez R, et al. Cancer Genet. Cytogenet. (2007) PMID: 17498554
9. Szybka M, et al. Clin. Neuropathol. () PMID: 12908754
10. Heath JA, et al. Pediatr Blood Cancer (2013) PMID: 23255519
11. Mizoguchi M, et al. Neuropathology (2013) PMID: 23530875
12. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
13. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
14. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
15. Boland CR, et al. Cancer Res. (1998) PMID: 9823339
16. Pawlik TM, et al. Dis. Markers (2004) PMID: 15528785
17. Boland CR, et al. Gastroenterology (2010) PMID: 20420947
18. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
19. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
20. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
21. Cristescu R, et al. Science (2018) PMID: 30309915
22. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
23. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
24. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
25. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
26. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
27. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
28. Zhao J, et al. Nat. Med. (2019) PMID: 30742119
29. Touat M, et al. Nature (2020) PMID: 32322066
30. Bouffett E, et al. J. Clin. Oncol. (2016) PMID: 27001570
31. Johanns TM, et al. Cancer Discov (2016) PMID: 27683556
32. Lukas RV, et al. J. Neurooncol. (2018) PMID: 30073642
33. Chalmers ZR, et al. Genome Med (2017) PMID: 28420421
34. Patel RR, et al. Pediatr Blood Cancer (2020) PMID: 32386112
35. Johnson A, et al. Oncologist (2017) PMID: 28912153
36. Draaisma K, et al. Acta Neuropathol Commun (2015) PMID: 26699864
37. Wang L, et al. BMC Cancer (2020) PMID: 32164609
38. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
39. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
40. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
41. Rizvi NA, et al. Science (2015) PMID: 25765070
42. Johnson BE, et al. Science (2014) PMID: 24336570
43. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
44. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
45. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
46. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
47. Nature (2012) PMID: 22810696
48. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
49. Stein EM, et al. Blood (2017) PMID: 28588020
50. Yen K, et al. Cancer Discov (2017) PMID: 28193778
51. Amatangelo MD, et al. Blood (2017) PMID: 28588019
52. Aldoss I, et al. Haematologica (2018) PMID: 29545346
53. DiNardo CD, et al. Am. J. Hematol. (2018) PMID: 29218851
54. Andreani G, et al. Am J Hematol (2019) PMID: 30431666
55. Winters AC, et al. Blood Adv (2019) PMID: 31648312
56. DiNardo CD, et al. Blood (2018) PMID: 30361262
57. Wang YW, et al. Ann Hematol (2020) PMID: 31965269
58. Murrell GA, et al. J Prosthet Dent (1988) PMID: 3283329
59. Konopleva M, et al. Cancer Discov (2016) PMID: 27520294
60. Chan SM, et al. Nat. Med. (2015) PMID: 25599133
61. Figueroa ME, et al. Cancer Cell (2010) PMID: 21130701
62. Turcan S, et al. Nature (2012) PMID: 22343889
63. Emadi A, et al. Am. J. Hematol. (2015) PMID: 25651001
64. DiNardo CD, et al. Leuk. Lymphoma (2014) PMID: 24138309
65. Metzeler KH, et al. Leukemia (2012) PMID: 22124213
66. Traina F, et al. Leukemia (2014) PMID: 24045501
67. DiNardo et al., 2017; ASH Abstract 639
68. Wang et al., 2015; ASH Abstract 2566
69. Tate JG, et al. Nucleic Acids Res. (2019) PMID: 30371878
70. Qi ST, et al. Oncol. Rep. (2011) PMID: 21874255
71. Frenel JS, et al. J. Neurooncol. (2013) PMID: 23681562
72. Yip S, et al. J. Pathol. (2012) PMID: 22072542
73. Shibahara I, et al. Int. J. Clin. Oncol. (2012) PMID: 21971842
74. Li S, et al. Neuro-oncology (2013) PMID: 23115158
75. Arita H, et al. Acta Neuropathol Commun (2020) PMID: 33228806
76. Reitman ZJ, et al. J. Natl. Cancer Inst. (2010) PMID: 20513808
77. Jin G, et al. PLoS ONE (2011) PMID: 21326614
78. Kranendijk M, et al. Biochim. Biophys. Acta (2011) PMID: 21889589
79. Gross S, et al. J. Exp. Med. (2010) PMID: 20142433
80. Ward PS, et al. Cancer Cell (2010) PMID: 20171147
81. Dang L, et al. Trends Mol Med (2010) PMID: 20692206
82. Amary MF, et al. J. Pathol. (2011) PMID: 21598255
83. Cardaci S, et al. Int J Cell Biol (2012) PMID: 22888353
84. Weller M, et al. Nat Rev Clin Oncol (2021) PMID: 33293629
85. Debeb BG, et al. Breast Cancer Res. Treat. (2012) PMID: 22547109
86. Fouladi M, et al. J. Clin. Oncol. (2011) PMID: 21825264
87. Groth C, et al. Semin. Cell Dev. Biol. (2012) PMID: 22309842
88. Kamstrup MR, et al. Blood (2010) PMID: 20538790
89. Kridel R, et al. Blood (2012) PMID: 22210878
90. Krop I, et al. J. Clin. Oncol. (2012) PMID: 22547604
91. Rosati E, et al. Int. J. Cancer (2013) PMID: 23001755
92. Samon JB, et al. Mol. Cancer Ther. (2012) PMID: 22504949
93. Lehal R, et al. Proc Natl Acad Sci U S A (2020) PMID: 32601208
94. Ferrarotto et al., 2020; ESMO Abstract 919MO
95. Knoechel B, et al. Cold Spring Harb Mol Case Stud (2015) PMID: 27148573
96. Lopez-Miranda et al., 2021; ASCO Abstract 3020
97. Dreyling M, et al. Ann. Oncol. (2017) PMID: 28633365
98. Palomero T, et al. Nat. Med. (2007) PMID: 17873882
99. Liu S, et al. Urol. Oncol. (2013) PMID: 21993533
100. Cancer Genome Atlas Research Network, et al. N. Engl. J. Med. (2015) PMID: 26061751
101. Brennan CW, et al. Cell (2013) PMID: 24120142
102. Nature (2008) PMID: 18772890
103. Cerami E, et al. Cancer Discov (2012) PMID: 22588877
104. Gao J, et al. Sci Signal (2013) PMID: 23550210
105. Xu P, et al. J. Neurooncol. (2010) PMID: 19771395
106. Xu P, et al. Pathol. Oncol. Res. (2009) PMID: 19424825
107. Brandt WD, et al. J. Neuropathol. Exp. Neurol. (2015) PMID: 25575134
108. Jiang L, et al. J Clin Neurosci (2011) PMID: 21251836
109. Wang NJ, et al. Proc. Natl. Acad. Sci. U.S.A. (2011) PMID: 22006338
110. Klinakis A, et al. Nature (2011) PMID: 21562564
111. Penton AL, et al. Semin. Cell Dev. Biol. (2012) PMID: 22306179
112. Kopan R, et al. Cell (2009) PMID: 19379690
113. Edwards PA, et al. Breast Cancer Res. (2012) PMID: 22424054
114. Robinson DR, et al. Nat. Med. (2011) PMID: 22101766
115. Haydu JE, et al. Blood (2012) PMID: 22510873
116. Palomero T, et al. Leukemia (2006) PMID: 16688224
117. Ashworth TD, et al. Blood (2010) PMID: 20852131
118. Sakamoto K, et al. Exp. Cell Res. (2005) PMID: 15561108
119. Andrawes MB, et al. J. Biol. Chem. (2013) PMID: 23839946
120. Rebay I, et al. Cell (1991) PMID: 1657403
121. Ge C, et al. BMC Dev. Biol. (2008) PMID: 18445292
122. Aster JC, et al. Mol. Cell. Biol. (2000) PMID: 11003647
123. Weng AP, et al. Science (2004) PMID: 15472075
124. Deregowski V, et al. J. Bone Miner. Res. (2006) PMID: 16869730
125. Uchibori M, et al. Oncol. Rep. (2017) PMID: 28791383
126. Liu J, et al. Proc Natl Acad Sci U S A (2013) PMID: 24277854
127. Nat Rev Clin Oncol (2017) PMID: 27245281
128. Duperret EK, et al. Mol Ther (2018) PMID: 29249395
129. Chiappori AA, et al. Ann Oncol (2015) PMID: 25467017
130. Killela PJ, et al. Proc. Natl. Acad. Sci. U.S.A. (2013) PMID: 23530248
131. Killela PJ, et al. Oncotarget (2014) PMID: 24722048
132. Nonoguchi N, et al. Acta Neuropathol. (2013) PMID: 23955565
133. Liu X, et al. Cell Cycle (2013) PMID: 23603989
134. Koelsche C, et al. Acta Neuropathol. (2013) PMID: 24154961
135. Arita H, et al. Acta Neuropathol. (2013) PMID: 23764841
136. Reitman ZJ, et al. Acta Neuropathol. (2013) PMID: 24217890
137. Shay JW, et al. Semin. Cancer Biol. (2011) PMID: 22015685
138. Shay JW, et al. Eur. J. Cancer (1997) PMID: 9282118
139. Kim NW, et al. Science (1994) PMID: 7605428
140. Hanahan D, et al. Cell (2000) PMID: 10647931
141. Horn S, et al. Science (2013) PMID: 23348503
142. Huang FW, et al. Science (2013) PMID: 23348506
143. Vinagre J, et al. Nat Commun (2013) PMID: 23887589

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 27 June 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531