

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT	DISEASE Testis germ cell tumor (seminoma)	PHYSICIAN	ORDERING PHYSICIAN Yeh, Yi-Chen	SPECIMEN	SPECIMEN SITE Testis
	NAME Lin, Yen-Wei		MEDICAL FACILITY Taipei Veterans General Hospital		SPECIMEN ID S107-36281 E (PF21077)
	DATE OF BIRTH 30 March 1993		ADDITIONAL RECIPIENT None		SPECIMEN TYPE Slide Deck
	SEX Male		MEDICAL FACILITY ID 205872		DATE OF COLLECTION 20 September 2018
	MEDICAL RECORD # 44954598		PATHOLOGIST Not Provided		SPECIMEN RECEIVED 28 December 2021

Biomarker Findings

Microsatellite status - MS-Stable
Tumor Mutational Burden - 0 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

KIT amplification
PDGFRA amplification
CCND2 amplification
KRAS amplification
FGF23 amplification
FGF6 amplification
KDR amplification

† See About the Test in appendix for details.

Report Highlights

- Targeted therapies with potential clinical benefit approved in another tumor type: Imatinib (p. 7), Nilotinib (p. 7), Sorafenib (p. 8), Sunitinib (p. 8)
- Evidence-matched clinical trial options based on this patient's genomic findings: (p. 9)

BIOMARKER FINDINGS

Microsatellite status - MS-Stable

Tumor Mutational Burden - 0 Muts/Mb

GENOMIC FINDINGS

KIT - amplification

10 Trials see p. 11

PDGFRA - amplification

1 Trial see p. 15

CCND2 - amplification

9 Trials see p. 9

KRAS - amplification

9 Trials see p. 13

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. see Biomarker Findings section

No therapies or clinical trials. see Biomarker Findings section

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)
none	Imatinib
	Nilotinib
	Sorafenib
	Sunitinib
none	Imatinib
none	none
none	none

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GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

FGF23 - amplification p. 5 **KDR - amplification** p. 6
FGF6 - amplification p. 6

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

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BIOMARKER FINDINGS

BIOMARKER

Microsatellite status

RESULT

MS-Stable

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁻³, including approved therapies nivolumab and pembrolizumab⁴. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, $p=0.001$)⁵.

FREQUENCY & PROGNOSIS

MSI positivity at any level has been reported in 0-31% ($n=17-188$) of testicular germ cell tumors (GCTs)⁶⁻¹⁴ and in 33% (12/36) of ovarian GCTs⁹. Within testicular GCTs, 12-31% of cases ($n=17-51$) have been classified as having low-level MSI (MSI-L) and 69-100% ($n=17-133$) have been classified as MSS^{10,12,14}, with MSI-high (MSI-H) rarely reported; one study identified MSI-L in 6% (6/100) and MSI-H in 0% (0/100) of unselected testicular GCTs, and MSI-L in 6% (2/35) and MSI-H in 26% (9/35) of chemotherapy-resistant cases¹⁵⁻¹⁶. The significance of MSI in the context of GCTs is not well established (PubMed, Nov 2021). One study reported MSI-H to be significantly more frequent in chemotherapy-resistant than unselected testicular GCT cases¹⁵⁻¹⁶, and some studies have associated MSI positivity with reduced PFS and/or increased risk of recurrence^{11,15-17}; however, another study found no correlation between MSI status and clinicopathologic features, including

chemotherapy sensitivity and outcome¹².

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor¹⁸. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2¹⁸⁻²⁰. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers²¹⁻²³. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{18,20,22-23}.

BIOMARKER

Tumor Mutational Burden

RESULT

0 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1²⁴⁻²⁶, anti-PD-1 therapies²⁴⁻²⁷, and combination nivolumab and ipilimumab²⁸⁻³³. In multiple pan-tumor studies, higher TMB has been reported to be associated with increased ORR and OS from treatment with immune checkpoint inhibitors^{24-27,34}. Higher TMB was found to be significantly associated with improved OS upon immune checkpoint inhibitor treatment for patients with 9 types of advanced tumors²⁴. Analyses across several solid tumor types reported

that patients with higher TMB (defined as $\geq 16-20$ Muts/Mb) achieved greater clinical benefit from PD-1 or PD-L1-targeting monotherapy, compared with patients with higher TMB treated with chemotherapy³⁵ or those with lower TMB treated with PD-1 or PD-L1-targeting agents²⁵. However, the KEYNOTE 158 trial of pembrolizumab monotherapy for patients with solid tumors found significant improvement in ORR for patients with TMB ≥ 10 Muts/Mb (based on this assay or others) compared to those with TMB < 10 Muts/Mb, in a large cohort that included multiple tumor types; similar findings were observed in the KEYNOTE 028 and 012 trials^{27,34}. Together, these studies suggest that patients with TMB ≥ 10 Muts/Mb may derive clinical benefit from PD-1 or PD-L1 inhibitors.

FREQUENCY & PROGNOSIS

Compared to other tumor types, germ cell tumors (GCTs) have been reported to have a low TMB (mean 0.51 mutations per megabase [mut/Mb] as measured in tissue), with a similar mutation rate reported for seminoma (0.50 muts/Mb) and non-seminoma (0.49 muts/Mb) tumors³⁶. Published

data investigating the prognostic implications of TMB in germ cell tumors are limited (PubMed, Sep 2021).

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma³⁷⁻³⁸ and cigarette smoke in lung cancer³⁹⁻⁴⁰, treatment with temozolomide-based chemotherapy in glioma⁴¹⁻⁴², mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes⁴³⁻⁴⁷, and microsatellite instability (MSI)^{43,46-47}. This sample harbors a TMB level associated with lower rates of clinical benefit from treatment with PD-1- or PD-L1-targeting immune checkpoint inhibitors compared with patients with tumors harboring higher TMB levels, based on several studies in multiple solid tumor types^{25-26,34}.

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GENOMIC FINDINGS

GENE

KIT

ALTERATION
amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence, primarily in GIST, AML, and systemic mastocytosis, KIT activating alterations are associated with sensitivity to KIT tyrosine kinase inhibitors including imatinib, sunitinib, sorafenib, dasatinib, nilotinib, pazopanib, regorafenib, ponatinib, midostaurin, avapritinib, and ripretinib⁴⁸⁻⁵⁵. The use of mTOR inhibitors as an alternative therapeutic strategy has demonstrated limited success in KIT-mutated, imatinib-resistant melanoma, with 1 PR and 3 SD observed for 4 patients treated with everolimus⁵⁶. However, no

responses were observed for 10 patients with mastocytosis following everolimus monotherapy, with 8/10 patients harboring the KIT D816V mutation⁵⁷. The role of KIT amplification as a biomarker for response to mTOR inhibitors has not been investigated (PubMed, Mar 2021). Clinical benefit has been observed for patients with KIT amplified or overexpressing tumors following treatment with imatinib⁵⁸⁻⁶⁸, nilotinib⁶⁹, sorafenib⁷⁰⁻⁷³, and sunitinib⁷⁴⁻⁷⁵, suggesting that KIT amplification may be sensitive to these inhibitors. However, evidence demonstrating clinical benefit for regorafenib, dasatinib, pazopanib, or ponatinib in the context of KIT amplified or overexpressing tumors is limited.

FREQUENCY & PROGNOSIS

KIT amplification was reported in 21% of seminoma and 9% of nonseminoma germ cell tumors⁷⁶. In seminoma, KIT amplification was reported to be associated with increased gene expression⁷⁶. KIT gene expression was reported to

be increased in seminoma compared with non-seminoma⁷⁶. In the Testicular Germ Cell TCGA and the Germ Cell Tumors MSKCC datasets, KIT amplification has been observed in 2.0% (3/149) and 2.2% (4/180) of cases, respectively (cBioPortal, Apr 2021)⁷⁷⁻⁷⁸. The prognostic significance of KIT alterations in germ cell neoplasms has not been established (PubMed, Apr 2021).

FINDING SUMMARY

KIT (also called c-KIT) encodes a cell surface tyrosine kinase receptor that, upon ligand binding and dimerization, activates the PI3K-AKT and RAS-MAPK signaling pathways⁷⁹. KIT aberrations, including point mutations, translocations, amplification, and overexpression, have been associated with various malignancies, and KIT is considered an oncoprotein⁸⁰. KIT has been reported to be amplified in cancer⁷⁸ and may be biologically relevant in this context⁸¹⁻⁸².

GENE

PDGFRA

ALTERATION
amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of extensive clinical evidence in solid tumors and hematologic cancers, PDGFRA activating alterations are associated with sensitivity to imatinib⁸³⁻¹²⁰. Sorafenib has shown clinical and preclinical activity against the FIP1L1-PDGFRA fusion in chronic eosinophilic leukemia (CEL) and mutations associated with clinical resistance to imatinib and sunitinib in both CEL and gastrointestinal stromal tumor (GIST)¹²¹⁻¹²⁶. Complete responses to nilotinib have been reported in patients with CEL or hypereosinophilic syndrome with FIP1L1-PDGFRA or activating mutations^{99,127-128}; preclinical evidence has reported efficacy of

nilotinib in the context of PDGFRA mutations associated with GIST¹²⁹⁻¹³⁰. Patients with GIST harboring PDGFRA activating mutations have been reported to derive clinical benefit from treatment with sunitinib¹³¹ or regorafenib¹³²⁻¹³³. Preclinical studies have reported sensitivity of activating PDGFRA mutations and FIP1L1-PDGFRA fusion to dasatinib^{123,129}. PDGFRA D842 mutations were reported to be sensitive to avapritinib in clinical¹⁴⁸ and preclinical¹⁴⁸ studies of GIST, and demonstrated sensitivity to ripretinib for 1 patient¹³⁴.

FREQUENCY & PROGNOSIS

PDGFRA amplification has been reported in 1.7% of testicular germ cell tumors^{77-78,135}. In a study of 49 central nervous system germinomas, amplification of 4q12, the region where PDGFRA is located, was reported in 1 pineal gland germinoma¹³⁶. One study reported that PDGFRA gene expression was not associated with PDGFRA amplification in germ cell tumors⁷⁶. Amplification of PDGFRA or 4q12 was not reported in a study of 87 ovarian germ cell tumors¹³⁷. The frequency of

PDGFRA amplification in mediastinal germinomas and retroperitoneal germ cell tumors has not been evaluated (cBioPortal, PubMed, Nov 2021)⁷⁷⁻⁷⁸. Published data investigating the prognostic implications of PDGFRA alterations in germ cell tumors are limited (PubMed, Nov 2018). Published data investigating the prognostic implications of PDGFRA alterations in germ cell tumors are limited (PubMed, Jul 2021).

FINDING SUMMARY

PDGFRA encodes platelet-derived growth factor receptor alpha (PDGFR-alpha), a tyrosine kinase receptor that, upon binding of cognate ligands (PDGFA or PDGFB), activates several signaling pathways, including PI3K and MAPK¹³⁸. PDGFR aberrations, including point mutations, translocations, amplification, and/or overexpression, have been associated with various malignancies⁸⁰. Amplification of PDGFRA, frequently occurring with amplification of the genes KDR and KIT, has been associated with increased PDGFRA expression¹³⁹⁻¹⁴² and poor prognosis^{139,143-145} in some subtypes of glioma.

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GENOMIC FINDINGS
GENE

CCND2

ALTERATION
 amplification

abemaciclib, palbociclib, and ribociclib. Clinical studies of CDK4/6 inhibitors have shown the most promise for estrogen receptor-positive breast cancer¹⁴⁸⁻¹⁴⁹.

and CCND2 was reported to be one of the six most highly overexpressed genes in this tumor type¹⁵²⁻¹⁵⁴. Published data investigating the prognostic implications of CCND2 in germ cell tumors are limited (PubMed, Mar 2021).

FREQUENCY & PROGNOSIS

CCND2 in germ cell neoplasms has been most extensively studied in the context of testicular germ cell tumors. The amplification of the chromosomal region that includes CCND2 has been reported in at least 10% of testicular germ cell tumors¹⁵⁰⁻¹⁵². Overexpression of CCND2, at an average of 8-fold, has been reported in 69% (31/45) to 83% (25/30) of testicular germ cell tumors,

FINDING SUMMARY

CCND2 encodes the protein cyclin D2, which binds and regulates the cyclin-dependent kinases that control cell cycle progression, and is a downstream target of cancer signaling pathways including hedgehog and PI3K¹⁵⁵⁻¹⁵⁶. CCND2 has been reported to be amplified in cancer⁷⁸, and may be biologically relevant in this context⁸¹⁻⁸².

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

Although preclinical studies suggest that cyclin D2 activates CDK4/6¹⁴⁶⁻¹⁴⁷, it is unknown whether CCND2 amplification or activating mutation predicts response to CDK4/6 inhibitors such as

GENE

KRAS

ALTERATION
 amplification

1/2 patients with KRAS amplification experienced a PR¹⁶³. A patient with cervical cancer harboring both KRAS and PIK3CA amplification treated with the combination of trametinib and the AKT inhibitor GSK2141795 achieved a SD¹⁶⁴.

germ cell tumors but were found not to provide predictive information regarding response to therapy and patient survival^{16,171-172}.

FREQUENCY & PROGNOSIS

KRAS in germ cell neoplasms has been most extensively studied in the context of testicular germ cell tumors. KRAS amplification and mutation have been observed in 8.1-20.0% and 0.0-40.0% of testicular germ cell tumors, respectively (cBioPortal, Feb 2021)^{77-78,135,165-167}. KRAS mutations have been infrequently reported in ovarian germ cell tumors, but have been identified in malignant tumors arising from teratomas¹⁶⁸⁻¹⁷⁰. KRAS amplification and mutation have been linked to invasive growth of testicular

FINDING SUMMARY

KRAS encodes a member of the RAS family of small GTPases. Activating mutations in RAS genes can cause uncontrolled cell proliferation and tumor formation^{158,173}. In numerous cancer type-specific studies as well as a large-scale pan-cancer analysis, KRAS amplification was shown to correlate with increased expression^{166,174-176}. Additionally, KRAS amplification correlated with sensitivity of cancer cell lines to KRAS knockdown, suggesting that amplified KRAS is an oncogenic driver¹⁷⁶.

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

Preclinical evidence suggests that KRAS activation may predict sensitivity to MEK inhibitors, such as trametinib, binimetinib, cobimetinib, and selumetinib¹⁵⁷⁻¹⁶². Clinical evidence that KRAS amplification in the absence of a concurrent KRAS activating mutation is sensitive to MEK inhibitors is limited. A Phase 2 study of selumetinib plus docetaxel in patients with gastric cancer reported

GENE

FGF23

ALTERATION
 amplification

activity in FGF amplified cancers; following treatment with a selective pan-FGFR inhibitor, a patient with head and neck squamous cell carcinoma (HNSCC) and amplification of 11q13 (FGF3, FGF4, FGF19) and 12p13 (FGF6 and FGF23) experienced a radiologic CR¹⁷⁷.

(3.7%), and head and neck squamous cell carcinoma (3.4%) (cBioPortal, 2022)⁷⁷⁻⁷⁸.

FREQUENCY & PROGNOSIS

FGF23 alterations have been reported with highest incidence in uterine carcinosarcoma (7.0%), ovarian carcinoma (6.5%), testicular germ cell cancer (5.4%), cutaneous melanoma (5.0%), low-grade glioma (4.9%), lung squamous cell carcinoma (4.5%), sarcoma (4.3%), colorectal adenocarcinoma (4.2%), lung adenocarcinoma

FINDING SUMMARY

FGF23 encodes a member of the fibroblast growth factor protein family that plays a central role in phosphate homeostasis¹⁷⁸. Overexpression of FGF23 by tumor cells can cause hypophosphatemia through excessive renal phosphate clearance¹⁷⁹, while germline gain-of-function (protein stabilizing) mutations in FGF23 cause autosomal dominant hypophosphatemic rickets¹⁸⁰.

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

There are no targeted therapies that directly address genomic alterations in FGF23. Inhibitors of FGF receptors, however, are undergoing clinical trials in a number of different cancers. Limited data suggest that pan-FGFR inhibitors show

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GENOMIC FINDINGS
GENE
FGF6
ALTERATION
 amplification

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

There are no targeted therapies that directly address genomic alterations in FGF6. Inhibitors of FGF receptors, however, are undergoing clinical trials in a number of different cancers. Limited data suggest that pan-FGFR inhibitors show activity in FGF amplified cancers; following treatment with a selective pan-FGFR inhibitor, a patient with head and neck squamous cell carcinoma (HNSCC) and amplification of 11q13 (FGF3, FGF4, FGF19) and 12p13 (FGF6 and FGF23) experienced a radiologic CR¹⁷⁷.

FREQUENCY & PROGNOSIS

Somatic alterations affecting FGF6 are infrequently documented, with the highest rates reported in penile cancer (4%), cutaneous melanoma (1-3%), stomach carcinoma (1-3%) and colorectal cancer (1%) (cBioPortal, COSMIC, Jan 2022)^{77-78,181}. Amplification of FGF6 has been frequently observed in testicular germ cell cancer (5%) and ovarian serous cystadenocarcinoma (5%), and in 2-6% of lower-grade gliomas, glioblastomas, sarcomas, breast invasive carcinomas, uterine carcinosarcomas, lung squamous cell carcinomas (SCC), head and neck SCC, pancreatic adenocarcinomas, and esophageal carcinomas (cBioPortal, Jan 2022)⁷⁷⁻⁷⁸. FGF6 is co-localized with FGF23 and CCND2 at chromosomal locus 12p13 and has been reported to be co-amplified with these genes in 1.3% of

patients with breast cancer¹⁸². FGF6 expression has been reported in 54% (14/26) of prostate cancer samples, which also frequently express FGFR4¹⁸³. FGF6 expression has also been observed in 71% (12/17) of patients with childhood acute lymphoblastic leukemia¹⁸⁴.

FINDING SUMMARY

FGF6 (also known as HST-2) encodes a member of the fibroblast growth factor protein family and is hypothesized to play a role in muscle tissue regeneration¹⁸⁵ by signaling through FGFR4, and to a lesser extent FGFR1 and FGFR2¹⁸⁶. FGF6 expression has been observed in several cancers^{183-184,187} and was shown to be oncogenic in preclinical models¹⁸⁷⁻¹⁸⁸. FGF6 has been reported as amplified in cancer⁷⁸ and may be biologically relevant in this context⁸¹⁻⁸².

GENE
KDR
ALTERATION
 amplification

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

On the basis of clinical benefit for patients with ccRCC¹⁸⁹⁻¹⁹³ and a patient with breast angiosarcoma¹⁹⁴, high VEGFR-2 expression has been associated with sensitivity to sunitinib. However, because data supporting concordance between VEGFR-2 expression and KDR genomic biomarkers are limited, it is unclear whether these

therapeutic strategies would be beneficial in this case. On the basis of extensive clinical evidence across multiple tumor types, expression of plasma or tumor VEGFR-1 or VEGFR-2 has not been established as a reliable biomarker to predict response to the VEGFA-targeted agent bevacizumab¹⁹⁵⁻²¹⁴.

FREQUENCY & PROGNOSIS

KDR amplification has been reported in 2.2% of testicular germ cell tumors^{77-78,135}. In a study of 49 central nervous system germinomas, amplification of 4q12, the region where KDR is located, was reported in 1 pineal gland germinoma¹³⁶. Amplification of KDR or 4q12 was not reported in a study of 87 ovarian germ cell tumors¹³⁷. The frequency of KDR amplification in mediastinal

germinomas and retroperitoneal germ cell tumors has not been evaluated (cBioPortal, PubMed, Nov 2021)⁷⁷⁻⁷⁸. Published data investigating the prognostic implications of KDR alterations in germ cell tumors are limited (PubMed, Nov 2021).

FINDING SUMMARY

KDR encodes vascular endothelial growth factor receptor 2 (VEGFR2), a member of the vascular endothelial growth factor receptor (VEGFR) family. It is a receptor tyrosine kinase that transmits signals from VEGFA and is involved in both tumor angiogenesis and vasculogenesis during development²¹⁵. KDR amplification has been reported in many tumor types and may be oncogenic²¹⁵.

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THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Imatinib

Assay findings association
KIT
amplification

PDGFRA
amplification

AREAS OF THERAPEUTIC USE

Imatinib targets the BCR-ABL fusion protein, PDGFR, and KIT. It is FDA approved for the treatment of KIT-positive gastrointestinal stromal tumors (GIST), Ph+ chronic myeloid leukemia (CML) and acute lymphoblastic leukemia (ALL), myelodysplastic syndrome/myeloproliferative syndrome (MDS/MPS), aggressive systemic mastocytosis without a D816V KIT mutation, hypereosinophilic syndrome and/or chronic eosinophilic leukemia, and dermatofibrosarcoma protuberans. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical and preclinical data in KIT-

mutated^{59-60,95,216}, KIT-amplified⁵⁸⁻⁶¹, or KIT-expressing tumors^{63-68,217-218}, KIT activating alterations may confer sensitivity to imatinib. PDGFRA amplification may predict sensitivity to tyrosine kinase inhibitors such as imatinib; a patient with Merkel cell carcinoma expressing PDGFRA achieved a complete response to imatinib⁹³.

SUPPORTING DATA

Although a Phase 2 trial of imatinib in chemotherapy-refractory germ cell tumors expressing KIT reported no significant clinical benefit²¹⁹, two case studies have reported responses of KIT-expressing testicular seminomas to imatinib, including one complete response for 28 months²²⁰⁻²²¹.

Nilotinib

Assay findings association
KIT
amplification

AREAS OF THERAPEUTIC USE

Nilotinib targets tyrosine kinases such as ABL (including BCR-ABL), PDGFRs, KIT, CSF1R, DDR1, and DDR2. It is FDA approved to treat newly diagnosed pediatric or adult patients with Philadelphia chromosome-positive (Ph+) chronic myeloid leukemia (CML) in chronic phase, adults with Ph+ CML in chronic or accelerated phase with resistance or intolerance to prior therapy including imatinib, and pediatric patients with Ph+ CML in chronic phase with resistance or intolerance to prior tyrosine-kinase inhibitor therapy. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical and preclinical data in KIT-mutated^{69,222-225}, KIT-amplified⁶⁹, or KIT-expressing tumors²²⁶, KIT activating alterations may confer sensitivity to nilotinib.

SUPPORTING DATA

Clinical data on the efficacy of nilotinib for the treatment of germ cell tumors are limited (PubMed, Nov 2021). Nilotinib has been primarily investigated as a therapeutic

option for the treatment of CML or gastrointestinal stromal tumors (GIST). In the context of CML, a Phase 3 clinical trial of Ph+ patients treated with imatinib or nilotinib (300 mg or 400 mg) reported progression-free survival (PFS) rates of 93% and 97-98% and overall survival (OS) rates of 93% and 94-97%, respectively, at 4 years²²⁷. For imatinib-resistant Japanese patients with CML, a Phase 2 trial reported a 47.8% major medical response rate to treatment with nilotinib at 12 months²²⁸. A Phase 3 clinical trial of single-agent nilotinib in 240 patients with advanced GIST who failed prior treatment with imatinib or sunitinib reported no significant difference in progression-free survival between nilotinib and the best supportive care, but did report increased overall survival for nilotinib-treated patients²²⁹. A Phase 2 trial has shown that nilotinib was well tolerated and suggested it may be particularly useful for treating patients with GIST harboring mutations in KIT exon 17²³⁰. Preclinical, cell-based assays have reported efficacy for nilotinib alone and in combination with additional therapies in the context of leiomyosarcoma and synovial sarcoma²³¹.

ORDERED TEST # ORD-1269426-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Sorafenib

Assay findings association
KIT
amplification

AREAS OF THERAPEUTIC USE

Sorafenib is a kinase inhibitor that targets the RAF kinases, KIT, FLT3, RET, VEGFRs, and PDGFRs. It is FDA approved for the treatment of unresectable hepatocellular carcinoma, advanced renal cell carcinoma, and recurrent or metastatic differentiated thyroid carcinoma. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical and preclinical data in KIT-mutated²³²⁻²³⁹ or KIT-expressing tumors⁷⁰⁻⁷³, KIT activating alterations may predict sensitivity to sorafenib.

SUPPORTING DATA

In a Phase 2 study for patients with relapsed or refractory testicular germ cell tumors, none of the 18 patients experienced a PR or CR, but 3 experienced SD lasting >11 months²⁴⁰. One patient with a KRAS-amplified testicular germ cell tumor who was refractory to chemotherapy and immunotherapy experienced CRs and PRs in metastatic lesions in the lung and brain upon treatment with the combination of carboplatin, paclitaxel, and sorafenib²⁴¹. Phase 2 studies in non-small cell lung cancer (NSCLC) report that single-agent sorafenib improved disease

control rates and that the addition of sorafenib to erlotinib increased survival in EGFR wild-type patients²⁴²⁻²⁴³. A Phase 1 trial of everolimus combined with sorafenib reported 11% partial responses and 77% stable disease²⁴⁴. In the context of small cell lung carcinoma, sorafenib combined with cisplatin plus etoposide was highly toxic and ineffective²⁴⁵. In HER2-negative breast cancer, Phase 2b trials found improved progression-free survival for sorafenib added to capecitabine, but not when added to paclitaxel²⁴⁶⁻²⁴⁷. Phase 2 studies of sorafenib in biliary tract cancer reported disease control rates of 33-39%²⁴⁸. Three patients with cholangiocarcinoma derived clinical benefit from sorafenib²⁴⁹⁻²⁵⁰. However, the addition of sorafenib to gemcitabine did not improve outcome in patients with biliary tract tumors compared with gemcitabine alone²⁵¹. A Phase 2 study of sorafenib and bicalutamide in castration-resistant prostate cancer (CRPC) observed a PSA response or stable disease (>6 months) in 47% (18/39) of patients²⁵². Single-agent sorafenib was moderately active as second-line treatment for CRPC (3.7 months PFS and 18.0 months OS)²⁵³. For the treatment of glioblastoma or high-grade gliomas, sorafenib alone or combined with temozolomide/radiotherapy or erlotinib did not show efficacy²⁵⁴⁻²⁵⁷.

Sunitinib

Assay findings association
KIT
amplification

AREAS OF THERAPEUTIC USE

Sunitinib is a small-molecule tyrosine kinase inhibitor that targets PDGFRs, VEGFRs, KIT, FLT3, CSF-1R, and RET. It is FDA approved for the treatment of advanced or metastatic pancreatic neuroendocrine tumors, gastrointestinal stromal tumors (GISTs) in patients who have progressed on or are intolerant to imatinib, and advanced renal cell carcinoma (RCC) as well as for the adjuvant treatment of patients at high risk of recurrent RCC after nephrectomy. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical and preclinical data in KIT-

mutated^{74,258-262} or KIT-expressing tumors⁷⁴⁻⁷⁵, KIT activating alterations may predict sensitivity to sunitinib.

SUPPORTING DATA

One study of sunitinib in cisplatin-refractory or multiply relapsed germ cell tumors reported a total response rate of 13% in 33 patients, including three confirmed partial responses (PR) and one unconfirmed PR²⁶³. However, a Phase 2 trial of sunitinib in patients with relapsed or refractory germ cell tumors found no objective responses in ten patients, with five cases of stable disease and five exhibiting progressive disease²⁶⁴. A case study has reported an exceptional response to sunitinib in a patient with RET-amplified germ cell tumor²⁶⁵.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

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Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1269426-01

CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov. Or visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

GENE
CCND2
RATIONALE
 CCND2 amplification or activation may predict

sensitivity to CDK4/6 inhibitors.

ALTERATION
 amplification

NCT04594005
PHASE 1/2

CDK4/6 Tumor, Abemaciclib, Paclitaxel

TARGETS
 CDK4, CDK6

LOCATIONS: Seoul (Korea, Republic of)

NCT03099174
PHASE 1

This Study in Patients With Different Types of Cancer (Solid Tumours) Aims to Find a Safe Dose of Xentuzumab in Combination With Abemaciclib With or Without Hormonal Therapies. The Study Also Tests How Effective These Medicines Are in Patients With Lung and Breast Cancer.

TARGETS
 CDK4, CDK6, IGF-1, IGF-2, Aromatase, ER

LOCATIONS: Seoul (Korea, Republic of), Goyang (Korea, Republic of), Aichi, Nagoya (Japan), Kanagawa, Isehara (Japan), Tokyo, Chuo-ku (Japan), Tokyo, Koto-ku (Japan), Chiba, Kashiwa (Japan), Helsinki (Finland), Tampere (Finland), Turku (Finland)

NCT04801966
PHASE NULL

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

TARGETS
 CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF

LOCATIONS: Melbourne (Australia)

NCT03994796
PHASE 2

Genetic Testing in Guiding Treatment for Patients With Brain Metastases

TARGETS
 TRKB, ALK, TRKC, ROS1, TRKA, CDK4, CDK6, PI3K, mTOR

LOCATIONS: Alaska, Washington

NCT02896335
PHASE 2

Palbociclib In Progressive Brain Metastases

TARGETS
 CDK4, CDK6

LOCATIONS: Massachusetts

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ORDERED TEST # ORD-1269426-01

CLINICAL TRIALS
NCT03310879
PHASE 2

Study of the CDK4/6 Inhibitor Abemaciclib in Solid Tumors Harboring Genetic Alterations in Genes Encoding D-type Cyclins or Amplification of CDK4 or CDK6

TARGETS
 CDK4, CDK6

LOCATIONS: Massachusetts

NCT03065062
PHASE 1

Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head & Neck and Other Solid Tumors

TARGETS
 PI3K-alpha, PI3K-gamma, mTORC1, mTORC2, CDK4, CDK6

LOCATIONS: Massachusetts

NCT03454035
PHASE 1

Ulixertinib/Palbociclib in Patients With Advanced Pancreatic and Other Solid Tumors

TARGETS
 MAPK3, MAPK1, CDK4, CDK6

LOCATIONS: North Carolina

NCT02897375
PHASE 1

Palbociclib With Cisplatin or Carboplatin in Advanced Solid Tumors

TARGETS
 CDK4, CDK6

LOCATIONS: Georgia

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ORDERED TEST # ORD-1269426-01

CLINICAL TRIALS
GENE
KIT
ALTERATION
 amplification

RATIONALE

KIT amplification or activating mutations may predict sensitivity to small molecule tyrosine kinase inhibitors. Also, because KIT activation

leads to activation of the PI3K-AKT-mTOR pathway, PI3K and mTOR pathway inhibitors may be relevant in a tumor with KIT activation.

NCT04337463
PHASE NULL

ATG-008 Combined With Toripalimab in Advanced Solid Tumors

TARGETS
 mTORC1, mTORC2, PD-1

LOCATIONS: Chongqing (China), Chengdu (China)

NCT04803318
PHASE 2

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

TARGETS
 mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK

LOCATIONS: Guangzhou (China)

NCT02461849
PHASE 2

Patients With Refractory, Metastatic Cancer Harboring KIT Mutation or Amplification to Investigate the Clinical Efficacy and Safety of Imatinib Therapy

TARGETS
 KIT, ABL

LOCATIONS: Seoul (Korea, Republic of)

NCT03564691
PHASE 1

Study of MK-4830 as Monotherapy and in Combination With Pembrolizumab (MK-3475) in Participants With Advanced Solid Tumors (MK-4830-001)

TARGETS
 ITL4, FGFRs, RET, PDGFRA, VEGFRs, KIT, PD-1

LOCATIONS: Seoul (Korea, Republic of), Tokyo (Japan), Haifa (Israel), Petah Tikva (Israel), Ramat Gan (Israel), Tel Aviv (Israel), Warszawa (Poland), Gdansk (Poland), Heraklion (Greece), Washington

NCT04008797
PHASE 1

A Study of E7386 in Combination With Other Anticancer Drug in Participants With Solid Tumor

TARGETS
 CBP, Beta-catenin, FGFRs, RET, PDGFRA, VEGFRs, KIT

LOCATIONS: Osakasayama (Japan), Chuo-Ku (Japan), Kashiwa (Japan)

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ORDERED TEST # ORD-1269426-01

CLINICAL TRIALS
NCT03297606
PHASE 2

Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)

TARGETS

VEGFRs, ABL, SRC, ALK, ROS1, AXL, TRKA, MET, TRKC, DDR2, KIT, EGFR, PD-1, CTLA-4, PARP, CDK4, CDK6, FLT3, CSF1R, RET, mTOR, ERBB2, ERBB3, MEK, BRAF, SMO

LOCATIONS: Vancouver (Canada), Edmonton (Canada), Saskatoon (Canada), Regina (Canada), Ottawa (Canada), Montreal (Canada), Toronto (Canada), Kingston (Canada), London (Canada)

NCT04729348
PHASE 2

Pembrolizumab And Lenvatinib In Leptomeningeal Metastases

TARGETS

PD-1, KIT, VEGFRs, FGFRs, PDGFRA, RET

LOCATIONS: Massachusetts

NCT03711058
PHASE 1/2

Study of PI3Kinase Inhibition (Copanlisib) and Anti-PD-1 Antibody Nivolumab in Relapsed/Refractory Solid Tumors With Expansions in Mismatch-repair Proficient (MSS) Colorectal Cancer

TARGETS

PD-1, PI3K

LOCATIONS: Maryland

NCT04449549
PHASE 2

Rapid Analysis and Response Evaluation of Combination Anti-Neoplastic Agents in Rare Tumors (RARE CANCER) Trial: RARE 1 Nilotinib and Paclitaxel

TARGETS

ABL, KIT

LOCATIONS: Maryland

NCT03065062
PHASE 1

Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head & Neck and Other Solid Tumors

TARGETS

PI3K-alpha, PI3K-gamma, mTORC1, mTORC2, CDK4, CDK6

LOCATIONS: Massachusetts

ORDERED TEST # ORD-1269426-01

CLINICAL TRIALS
GENE
KRAS
RATIONALE

KRAS activating mutations or amplification may predict sensitivity to inhibitors of MAPK pathway

components, including MEK inhibitors.

ALTERATION

amplification

NCT04803318
PHASE 2

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

TARGETS

mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK

LOCATIONS: Guangzhou (China)

NCT03989115
PHASE 1/2

Dose-Escalation and Dose-Expansion of RMC-4630 and Cobimetinib in Relapsed/Refractory Solid Tumors

TARGETS

SHP2, MEK

LOCATIONS: Seoul (Korea, Republic of), Oregon, California, Colorado, Arizona, Wisconsin, Illinois

NCT03284502
PHASE 1

Cobimetinib and HM95573 in Patients With Locally Advanced or Metastatic Solid Tumors

TARGETS

MEK, RAFs

LOCATIONS: Hwasun (Korea, Republic of), Pusan (Korea, Republic of), Seongnam (Korea, Republic of), Seoul (Korea, Republic of), Goyang-si (Korea, Republic of)

NCT04801966
PHASE NULL

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

TARGETS

CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF

LOCATIONS: Melbourne (Australia)

NCT03905148
PHASE 1/2

Study of the Safety and Pharmacokinetics of BGB-283 and PD-0325901 in Patients With Advanced or Refractory Solid Tumors

TARGETS

RAFs, EGFR, MEK

LOCATIONS: Nedlands (Australia), Blacktown (Australia), Randwick (Australia), Melbourne (Australia), Texas

NCT02407509
PHASE 1

Phase I Trial of RO5126766

TARGETS

RAFs, MEK, mTOR

LOCATIONS: London (United Kingdom), Sutton (United Kingdom)

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CLINICAL TRIALS
NCT04800822
PHASE 1

PF-07284892 in Participants With Advanced Solid Tumors

TARGETS

SHP2, ROS1, ALK, MEK, BRAF, EGFR

LOCATIONS: California, Michigan, New York, Tennessee, Texas

NCT02070549
PHASE 1

Trametinib in Treating Patients With Advanced Cancer With or Without Hepatic Dysfunction

TARGETS

MEK

LOCATIONS: Toronto (Canada)

NCT03162627
PHASE 1

Selumetinib and Olaparib in Solid Tumors

TARGETS

MEK, PARP

LOCATIONS: Texas

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CLINICAL TRIALS
GENE
PDGFRA
RATIONALE

PDGFRA amplification may predict sensitivity to imatinib and to anti-PDGFRA antibodies.

ALTERATION

amplification

NCT01738139
PHASE 1

Ipilimumab and Imatinib Mesylate in Advanced Cancer

TARGETS

KIT, ABL, CTLA-4

LOCATIONS: Texas

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APPENDIX
Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

BARD1
G82V

C11ORF30 (EMSY)
S1076G

CDKN1B
amplification

CEBPA
A358G

ERBB4
E133K

FLT1
P470H

MST1R
V670G

PDGFRA
rearrangement

PIK3C2B
H1145L

PIK3C2G
amplification

PTPRO
amplification

RAD52
amplification

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APPENDIX
Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTK	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL	CCND1
CCND2	CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2	DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FAM46C	FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3
FGFR4	FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3
GATA4	GATA6	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3F3A
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDMSA	KDMS5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKKN1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKB1A	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD3 (WHSC1L1)	NTSC2	NTRK1	NTRK2	NTRK3	P2RY8	PALB2
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA
PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET
RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2
SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOC3
SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU
SYK	TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53
TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1	WT1	XPO1
XRCC2	ZNF217	ZNF703						

DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSP02	SDC4	SLC34A2	TERC*	TERT**	TPR52

*TERC is an NCRNA

**Promoter region of TERT is interrogated

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Loss of Heterozygosity (LOH) score

Microsatellite (MS) status

Tumor Mutational Burden (TMB)

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APPENDIX

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.



ABOUT FOUNDATIONONE CDx

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:
www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLES

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials

Ranking of Therapies in Summary Table

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2021. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

1. In the fractional-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-

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- Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.
- TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
 - The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
 - Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
 - Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine

whether the patient is a candidate for biopsy.

- Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of *HER2* overexpression as 20% in breast cancer. Based on the F1CDx *HER2* CDx concordance study, approximately 10% of *HER2* amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

*Interquartile Range = 1st Quartile to 3rd Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear

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cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

MR Suite Version 5.2.0

The median exon coverage for this sample is 755x

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References

1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) PMID: 25392179
2. Kroemer G, et al. Oncoimmunology (2015) PMID: 26140250
3. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
4. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
5. Ayers et al., 2016; ASCO-SITC Abstract P60
6. Lothe RA, et al. Cancer Res. (1993) PMID: 8261392
7. Murty VV, et al. Cancer Res. (1994) PMID: 8033127
8. Huddart RA, et al. Br. J. Cancer (1995) PMID: 7669575
9. Faulkner SW, et al. Gynecol. Oncol. (2000) PMID: 11006028
10. Devouassoux-Shisheboran M, et al. Mol. Hum. Reprod. (2001) PMID: 11719586
11. Velasco A, et al. Cancer Biol. Ther. (2004) PMID: 15492498
12. Olasz J, et al. Anticancer Res. () PMID: 16309235
13. Vladušić T, et al. Anticancer Res. (2014) PMID: 25075023
14. Cárcano FM, et al. Andrology (2016) PMID: 27153176
15. Mayer F, et al. Cancer Res. (2002) PMID: 12019150
16. Honecker F, et al. J. Clin. Oncol. (2009) PMID: 19289622
17. Velasco A, et al. Int. J. Cancer (2008) PMID: 18076065
18. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
19. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
20. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
21. Boland CR, et al. Cancer Res. (1998) PMID: 9823339
22. Pawlik TM, et al. Dis. Markers (2004) PMID: 15528785
23. Boland CR, et al. Gastroenterology (2010) PMID: 20420947
24. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
25. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
26. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
27. Cristescu R, et al. Science (2018) PMID: 30309915
28. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
29. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
30. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
31. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
32. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
33. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
34. Marabelle A, et al. Lancet Oncol. (2020) PMID: 32919526
35. Legrand et al., 2018; ASCO Abstract 12000
36. Litchfield K, et al. Nat Commun (2015) PMID: 25609015
37. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
38. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
39. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
40. Rizvi NA, et al. Science (2015) PMID: 25765070
41. Johnson BE, et al. Science (2014) PMID: 24336570
42. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
43. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
44. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
45. Heitzner E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
46. Nature (2012) PMID: 22810696
47. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
48. Evans EK, et al. Sci Transl Med (2017) PMID: 29093181
49. Abbaspour Babaei M, et al. Drug Des Devel Ther (2016) PMID: 27536065
50. Ramaswamy A, et al. J Gastrointest Oncol (2016) PMID: 27563456
51. Demetri GD, et al. Lancet (2013) PMID: 23177515
52. Gotlib J, et al. N. Engl. J. Med. (2016) PMID: 27355533
53. Jawhar M, et al. Blood (2017) PMID: 28424161
54. Xu X, et al. Int J Clin Exp Pathol (2014) PMID: 25031773
55. Gotlib J, et al. Blood (2005) PMID: 15972446
56. Si L, et al. J. Clin. Oncol. (2012) PMID: 22162580
57. Parikh SA, et al. Leuk Lymphoma (2010) PMID: 20038218
58. Wei X, et al. Oncol. Res. (2019) PMID: 30075827
59. Hodi FS, et al. J. Clin. Oncol. (2013) PMID: 23775962
60. Carvajal RD, et al. JAMA (2011) PMID: 21642685
61. Guo J, et al. J. Clin. Oncol. (2011) PMID: 21690468
62. Debiec-Rychter M, et al. Gastroenterology (2005) PMID: 15685537
63. Dematteo RP, et al. Lancet (2009) PMID: 19303137
64. Faivre S, et al. J. Clin. Oncol. (2005) PMID: 16135502
65. Hotte SJ, et al. J. Clin. Oncol. (2005) PMID: 15659505
66. Alcedo JC, et al. Head Neck (2004) PMID: 15350030
67. Brandwein JM, et al. Leukemia (2011) PMID: 21403650
68. Reardon DA, et al. Br. J. Cancer (2009) PMID: 19904263
69. Lee SJ, et al. Oncologist (2015) PMID: 26424760
70. Llovet JM, et al. Clin. Cancer Res. (2012) PMID: 22374331
71. Zhang HL, et al. Clin Genitourin Cancer (2013) PMID: 23058498
72. Seino S, et al. Gastroenterology (2014) PMID: 25450081
73. Li XF, et al. Med. Oncol. (2009) PMID: 18846437
74. Minor DR, et al. Clin. Cancer Res. (2012) PMID: 22261812
75. Mahipal A, et al. Melanoma Res. (2012) PMID: 23114504
76. McIntyre A, et al. Cancer Res. (2005) PMID: 16166280
77. Cerami E, et al. Cancer Discov (2012) PMID: 22588877
78. Gao J, et al. Sci Signal (2013) PMID: 23550210
79. Int. J. Biochem. Cell Biol. (1999) PMID: 10582339
80. Semin. Oncol. (2004) PMID: 15175998
81. Zack TI, et al. Nat. Genet. (2013) PMID: 24071852
82. Beroukhir M, et al. Nature (2010) PMID: 20164920
83. Arefi M, et al. Int. J. Hematol. (2012) PMID: 22806436
84. Baccarani M, et al. Haematologica (2007) PMID: 17666373
85. Cassier PA, et al. Clin. Cancer Res. (2012) PMID: 22718859
86. Chalmers ZR, et al. Blood Cancer J (2015) PMID: 25658984
87. Cools J, et al. N. Engl. J. Med. (2003) PMID: 12660384
88. Curtis CE, et al. Br. J. Haematol. (2007) PMID: 17555450
89. Debiec-Rychter M, et al. Eur. J. Cancer (2004) PMID: 15010069
90. Dileo P, et al. Int. J. Cancer (2011) PMID: 20473908
91. Fanta PT, et al. J. Clin. Oncol. (2015) PMID: 24638008
92. Florian S, et al. Leuk. Res. (2006) PMID: 16406018
93. Frenard C, et al. JAAD Case Rep (2016) PMID: 27051816
94. Griffin JH, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) PMID: 12808148
95. Heinrich MC, et al. J. Clin. Oncol. (2003) PMID: 14645423
96. Helbig G, et al. Br. J. Haematol. (2009) PMID: 19120352
97. Helbig G, et al. Am. J. Hematol. (2014) PMID: 24009127
98. Hus M, et al. Leuk. Res. (2011) PMID: 21093052
99. Ikezoe T, et al. Leuk. Res. (2010) PMID: 20303172
100. Intermetoli T, et al. Br. J. Haematol. (2009) PMID: 19735261
101. Jain N, et al. Leuk. Res. (2009) PMID: 19013640
102. Jovanovic JV, et al. Blood (2007) PMID: 17299092
103. Kang HJ, et al. Acta Oncol (2012) PMID: 22150077
104. Klion AD, et al. Blood (2004) PMID: 14504092
105. Kobayashi M, et al. Respiratory (2009) PMID: 19192229
106. Kocáková I, et al. Klin Onkol (2014) PMID: 24635438
107. Metzgeroth G, et al. Br. J. Haematol. (2008) PMID: 18950453
108. Murayama Y, et al. World J Gastrointest Oncol (2012) PMID: 22645636
109. Ogbogu PU, et al. J. Allergy Clin. Immunol. (2009) PMID: 19910029
110. Ohnishi H, et al. Br. J. Haematol. (2006) PMID: 16856885
111. Pardanani A, et al. Blood (2003) PMID: 12842979
112. Pardanani A, et al. Blood (2004) PMID: 15284118
113. Qu SQ, et al. Oncotarget (2016) PMID: 27120808
114. Score J, et al. Leukemia (2006) PMID: 16498388
115. Shah S, et al. J Hematol Oncol (2014) PMID: 24669761
116. Sugimoto Y, et al. Cancer Genet (2015) PMID: 26319757
117. Volz HC, et al. Int. J. Cardiol. (2011) PMID: 20609486
118. von Bubnoff N, et al. Leukemia (2005) PMID: 15618966
119. Walz C, et al. Genes Chromosomes Cancer (2006) PMID: 16845659
120. Yoo C, et al. Cancer Res Treat (2016) PMID: 26130666
121. Al-Riyami AZ, et al. Leuk. Lymphoma (2013) PMID: 23157309
122. Lierman E, et al. Blood (2006) PMID: 16645167
123. Lierman E, et al. Leukemia (2009) PMID: 19212337
124. Metzgeroth G, et al. Leukemia (2012) PMID: 21818111
125. Roubaud G, et al. Ann. Oncol. (2012) PMID: 22294526
126. von Bubnoff N, et al. Oncogene (2011) PMID: 20972453
127. Hochhaus A, et al. J. Cancer Res. Clin. Oncol. (2013) PMID: 24057647
128. Tabouret E, et al. Leuk. Res. (2011) PMID: 20832858
129. Dewaele B, et al. Clin. Cancer Res. (2008) PMID: 18794084
130. Weisberg E, et al. Gastroenterology (2006) PMID: 17087936
131. Brohl AS, et al. Clin Sarcoma Res (2015) PMID: 26396737
132. Grellety T, et al. Future Sci OA (2015) PMID: 28031906
133. Kollár A, et al. Clin Sarcoma Res (2014) PMID: 25905001
134. Jaku et al., 2017; ASCO Abstract 2515
135. Bagrodia A, et al. J. Clin. Oncol. (2016) PMID: 27646943
136. Schulte SL, et al. Oncotarget (2016) PMID: 27391150
137. Van Nieuwenhuysen E, et al. Gynecol. Oncol. (2018) PMID: 30170975
138. Andrae J, et al. Genes Dev. (2008) PMID: 18483217
139. Burford A, et al. PLoS ONE (2013) PMID: 23990986
140. Flavahan WA, et al. Nature (2016) PMID: 26700815
141. Roszik J, et al. Sci Rep (2016) PMID: 26787600
142. Verhaak RG, et al. Cancer Cell (2010) PMID: 20129251
143. Koschmann C, et al. Oncotarget (2016) PMID: 27582545
144. Phillips JJ, et al. Brain Pathol. (2013) PMID: 23438035
145. Puget S, et al. PLoS ONE (2012) PMID: 22389665
146. Busk PK, et al. Exp. Cell Res. (2005) PMID: 15707582
147. Busk PK, et al. Cell Cycle () PMID: 12695654
148. Finn RS, et al. Lancet Oncol. (2015) PMID: 25524798
149. DeMichele A, et al. Clin. Cancer Res. (2015) PMID: 25501126
150. Rodriguez S, et al. Oncogene (2003) PMID: 12660824
151. Korkola JE, et al. Cancer Res. (2006) PMID: 16424014
152. Skotheim RI, et al. Cell. Oncol. (2006) PMID: 17167184
153. Schmidt BA, et al. Cancer Res. (2001) PMID: 11358847
154. Kukoski R, et al. Appl. Immunohistochem. Mol. Morphol. (2003) PMID: 12777997
155. Katoh Y, et al. Curr. Mol. Med. (2009) PMID: 19860666
156. White PC, et al. Oncogene (2006) PMID: 16301994

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APPENDIX
References

157. Nakano H, et al. Proc. Natl. Acad. Sci. U.S.A. (1984) pmid: 6320174
158. Pylayeva-Gupta Y, et al. Nat. Rev. Cancer (2011) pmid: 21993244
159. Yamaguchi T, et al. Int. J. Oncol. (2011) pmid: 21523318
160. Watanabe M, et al. Cancer Sci. (2013) pmid: 23438367
161. Gilmartin AG, et al. Clin. Cancer Res. (2011) pmid: 21245089
162. Yeh JJ, et al. Mol. Cancer Ther. (2009) pmid: 19372556
163. Lee et al., 2018; ASCO Abstract 4061
164. Liu JF, et al. Gynecol. Oncol. (2019) pmid: 31118140
165. Goddard NC, et al. Int. J. Androl. (2007) pmid: 17573850
166. McIntyre A, et al. Neoplasia (2005) pmid: 16354586
167. Moul JW, et al. Genes Chromosomes Cancer (1992) pmid: 1381946
168. Li Y, et al. J Ovarian Res (2014) pmid: 25297496
169. Hershkovitz D, et al. Pathol. Int. (2013) pmid: 24422958
170. Stanojevic B, et al. BMC Cancer (2012) pmid: 22682753
171. Mayer F, et al. BJU Int. (2011) pmid: 20955261
172. Roelofs H, et al. Am. J. Pathol. (2000) pmid: 11021820
173. Kahn S, et al. Anticancer Res. () pmid: 3310850
174. Mita H, et al. BMC Cancer (2009) pmid: 19545448
175. Birkeland E, et al. Br. J. Cancer (2012) pmid: 23099803
176. Chen Y, et al. PLoS ONE (2014) pmid: 24874471
177. Dumbrava et al., 2018; doi/full/10.1200/PO.18.00100
178. Jonsson KB, et al. N. Engl. J. Med. (2003) pmid: 12711740
179. Shimada T, et al. Proc. Natl. Acad. Sci. U.S.A. (2001) pmid: 11344269
180. Yu X, et al. Cytokine Growth Factor Rev. (2005) pmid: 15863037
181. Tate JG, et al. Nucleic Acids Res. (2019) pmid: 30371878
182. Parish A, et al. Cell Cycle (2015) pmid: 25950492
183. Ropiquet F, et al. Cancer Res. (2000) pmid: 10945637
184. Niini T, et al. Leukemia (2002) pmid: 12399964
185. Neuhaus P, et al. Mol. Cell. Biol. (2003) pmid: 12917328
186. Ornitz DM, et al. J. Biol. Chem. (1996) pmid: 8663044
187. Iida S, et al. Oncogene (1992) pmid: 1549352
188. Marics I, et al. Oncogene (1989) pmid: 2649847
189. Beuselinck B, et al. Acta Oncol (2018) pmid: 29095068
190. Song Y, et al. Chin. Med. J. (2015) pmid: 26228213
191. Dornbusch J, et al. PLoS ONE (2013) pmid: 24086736
192. Terakawa T, et al. Urol. Oncol. (2013) pmid: 21478036
193. You D, et al. World J Urol (2015) pmid: 24710685
194. Silva E, et al. Breast J () pmid: 25639617
195. Baumgarten P, et al. Neuro-oncology (2016) pmid: 26627848
196. Sathornsumetee S, et al. J. Clin. Oncol. (2008) pmid: 18182667
197. Olafson LR, et al. J Clin Neurosci (2019) pmid: 31582283
198. Duda DG, et al. Oncologist (2010) pmid: 20484123
199. Stremtitz S, et al. Mol. Cancer Ther. (2016) pmid: 27535973
200. Weickhardt AJ, et al. Br. J. Cancer (2015) pmid: 26125443
201. Kopetz S, et al. J. Clin. Oncol. (2010) pmid: 20008624
202. Miles DW, et al. Br. J. Cancer (2013) pmid: 23422754
203. Fountzilias G, et al. Anticancer Res. (2011) pmid: 21868552
204. Gianni L, et al. J. Clin. Oncol. (2013) pmid: 23569311
205. Sánchez-Rovira P, et al. Clin Transl Oncol (2013) pmid: 23397155
206. Cameron D, et al. Lancet Oncol. (2013) pmid: 23932548
207. Mok T, et al. J Thorac Oncol (2014) pmid: 24807156
208. An SJ, et al. Cancer Gene Ther. (2014) pmid: 24577128
209. Bais C, et al. J. Natl. Cancer Inst. (2017) pmid: 29059426
210. Cohen EE, et al. Lancet Oncol. (2009) pmid: 19201650
211. Van Cutsem E, et al. J. Clin. Oncol. (2012) pmid: 22565005
212. Lee EQ, et al. Clin. Cancer Res. (2018) pmid: 29941486
213. Xu L, et al. Cancer Res. (2009) pmid: 19826039
214. Heist RS, et al. Proc. Natl. Acad. Sci. U.S.A. (2015) pmid: 25605928
215. Biol. Pharm. Bull. (2011) pmid: 22130231
216. Debiec-Rychter M, et al. Eur. J. Cancer (2006) pmid: 16624552
217. Kamenz T, et al. World J. Gastroenterol. (2006) pmid: 16570351
218. Wang YY, et al. Proc. Natl. Acad. Sci. U.S.A. (2005) pmid: 15650049
219. Einhorn LH, et al. Am. J. Clin. Oncol. (2006) pmid: 16462496
220. Pedersini R, et al. Lancet Oncol. (2007) pmid: 17976614
221. Okamura A, et al. Ann. Oncol. (2010) pmid: 20080832
222. Carvajal RD, et al. Clin. Cancer Res. (2015) pmid: 25695690
223. Hochhaus A, et al. J. Cancer Res. Clin. Oncol. (2015) pmid: 26002753
224. Blay JY, et al. Lancet Oncol. (2015) pmid: 25882987
225. Kajimoto N, et al. Int J Clin Exp Pathol (2015) pmid: 26722383
226. Sako H, et al. PLoS ONE (2014) pmid: 25221952
227. Hughes TP, et al. Blood (2014) pmid: 24335106
228. Takahashi N, et al. Biomark Res (2014) pmid: 24650752
229. Reichardt P, et al. Ann. Oncol. (2012) pmid: 22357255
230. Cauchi C, et al. Cancer Chemother. Pharmacol. (2012) pmid: 22119758
231. Villar VH, et al. PLoS ONE (2012) pmid: 22662203
232. Quintás-Cardama A, et al. Nat Clin Pract Oncol (2008) pmid: 18936790
233. Bisagni G, et al. J Thorac Oncol (2009) pmid: 19461405
234. Handolias D, et al. Br. J. Cancer (2010) pmid: 20372153
235. Dişel U, et al. Lung Cancer (2011) pmid: 20970876
236. Park SH, et al. Invest New Drugs (2012) pmid: 22270258
237. Catania C, et al. Onco Targets Ther (2014) pmid: 24855380
238. Guo T, et al. Clin. Cancer Res. (2007) pmid: 17699867
239. Hu S, et al. Mol. Cancer Ther. (2008) pmid: 18483300
240. Skoneczna et al., 2014; ASCO Abstract 367
241. Lian B, et al. Oncologist (2019) pmid: 31492770
242. Wakelee HA, et al. J Thorac Oncol (2012) pmid: 22982658
243. Spigel DR, et al. J. Clin. Oncol. (2011) pmid: 21576636
244. You et al., 2016; ASCO Abstract 2532
245. Sharma N, et al. Invest New Drugs (2014) pmid: 24420556
246. Baselga J, et al. J. Clin. Oncol. (2012) pmid: 22412143
247. Gradishar WJ, et al. Eur. J. Cancer (2013) pmid: 22954665
248. Bengala C, et al. Br. J. Cancer (2010) pmid: 19935794
249. Qun W, et al. Hepatogastroenterology () pmid: 20698202
250. LaRocca RV, et al. J Gastrointest Cancer (2007) pmid: 19089671
251. Moehler M, et al. Eur. J. Cancer (2014) pmid: 25446376
252. Beardsley EK, et al. Invest New Drugs (2012) pmid: 21785998
253. Aragon-Ching JB, et al. BJU Int. (2009) pmid: 19154507
254. Reardon DA, et al. J. Neurooncol. (2011) pmid: 20443129
255. Lee EQ, et al. Neuro-oncology (2012) pmid: 23099651
256. Peereboom DM, et al. Neuro-oncology (2013) pmid: 23328813
257. Hottinger AF, et al. Br. J. Cancer (2014) pmid: 24786603
258. Heinrich MC, et al. J. Clin. Oncol. (2008) pmid: 18955458
259. Buchbinder EI, et al. Cancer (2015) pmid: 26264378
260. Reichardt P, et al. BMC Cancer (2016) pmid: 26772734
261. Hirai F, et al. Mol Clin Oncol (2016) pmid: 27073655
262. Goemans BF, et al. Leuk. Res. (2010) pmid: 20435347
263. Oechsle K, et al. Ann. Oncol. (2011) pmid: 21415240
264. Feldman DR, et al. Invest New Drugs (2010) pmid: 19547919
265. Subbiah V, et al. J Hematol Oncol (2014) pmid: 25085632

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