

PATIENT Chang, Chin-Wei TUMOR TYPE
Colon adenocarcinoma (CRC)
COUNTRY CODE
TW

REPORT DATE
10 Mar 2022
ORDERED TEST #
ORD-1314037-01

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT

DISEASE Colon adenocarcinoma (CRC)
NAME Chang, Chin-Wei
DATE OF BIRTH 18 May 1959
SEX Male
MEDICAL RECORD # 44137044

PHYSICIAN

ORDERING PHYSICIAN Yeh, Yi-Chen
MEDICAL FACILITY Taipei Veterans General Hospital
ADDITIONAL RECIPIENT None
MEDICAL FACILITY ID 205872
PATHOLOGIST Not Provided

SPECIMEN

SPECIMEN SITE Chest Wall
SPECIMEN ID S111-01929C
SPECIMEN TYPE Slide Deck
DATE OF COLLECTION 13 January 2022
SPECIMEN RECEIVED 04 March 2022

Biomarker Findings

Microsatellite status - MS-Stable
Tumor Mutational Burden - 5 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

KRAS G12D NRAS wildtype APC T1556fs*3 PIK3CA E545K CDKN2A/R CDK

CDKN2A/B CDKN2B loss, CDKN2A loss SMAD4 E538*

SOX9 Q411*

2 Disease relevant genes with no reportable alterations: *BRAF*, *NRAS*

† See About the Test in appendix for details.

Report Highlights

- Evidence-matched clinical trial options based on this patient's genomic findings: (p. 10)

BIOMARKER FINDINGS

Microsatellite status - MS-Stable

Tumor Mutational Burden - 5 Muts/Mb	
GENOMIC FINDINGS	
KRAS - G12D	
10 Trials see p. 11	
NRAS - wildtype	
0 Trials	
APC - T1556fs*3	
4 Trials see p. 10	
PIK3CA - E545K	
10 Trials see p. 13	

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. see Biomarker Findings section

No therapies or clinical trials. see Biomarker Findings section

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)
Cetuximab 🔀	none
Panitumumab 🗴	
Cetuximab 🗴	none
Panitumumab 🗴	
none	none
none	none

Extensive evidence showing variant(s) in this sample may confer resistance to this therapy





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GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

CDKN2A/B - CDKN2B loss, CDKN2A loss p. 7 *SOX9* - Q411* p. 8 *SMAD4* - E538* p. 8

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

The rapies contained in this report may have been approved by the $\ensuremath{\mathsf{US}}\xspace \ensuremath{\mathsf{FDA}}\xspace.$



BIOMARKER FINDINGS

BIOMARKER

Microsatellite status

RESULT MS-Stable

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁻³, including approved therapies nivolumab and pembrolizumab⁴. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, p=0.001)⁵. For patients with chemotherapy-refractory microsatellite-stable (MSS) metastatic colorectal cancer (CRC), a Phase 3 trial reported no OS advantage from the combination of the PD-L1 inhibitor atezolizumab plus cobimetinib relative to regorafenib (8.9 vs. 8.5 months, HR=1.00);

atezolizumab monotherapy similarly did not prolong OS (7.1 vs. 8.5 months, HR=1.19)⁶. For patients with MSS CRC, a Phase 2 study combining ipilimumab and nivolumab reported an overall DCR of 25% $(10/40)^7$. Two Phase 1 studies for patients with MSS CRC treated with regorafenib and nivolumab reported PFSs of 7.9 months⁸ and 5.7 months⁹, and a patient with MSS CRC refractory to chemotherapy treated with the PD-1 inhibitor sintilimab and regorafenib reported a CR^{10} .

Nontargeted Approaches —

MSI has not been found to be a predictive biomarker for combination chemotherapy regimens, including FOLFOX¹¹⁻¹² and FOLFIRI¹³⁻¹⁴. Patients with MSS CRC are more likely to benefit from postsurgical fluorouracil (FU)-based adjuvant therapy¹⁵⁻¹⁶ but less likely to benefit from irinotecan chemotherapy¹⁷.

FREQUENCY & PROGNOSIS

MSS colorectal cancers (CRCs) make up 70-85% of CRC cases^{3,18-22}. MSS colorectal cancers are

molecularly heterogeneous, driven by diverse mechanisms such as extensive DNA methylation, oncogenic mutations in KRAS or BRAF, or chromosomal instability²². Multiple studies have shown that MSS CRCs have a worse prognosis than MSI-high tumors^{18,23-29}.

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor²⁰. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH₂, MSH₆, or PMS₂^{20,30-31}. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers^{19,32-33}. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{19-20,31,33}.

BIOMARKER FINDINGS

BIOMARKER

Tumor Mutational Burden

RESULT 5 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L134-36, anti-PD-1 therapies34-37, and combination nivolumab and ipilimumab $^{38-43}$. In multiple pan-tumor studies, higher TMB has been reported to be associated with increased ORR and OS from treatment with immune checkpoint inhibitors34-37,44. Higher TMB was found to be significantly associated with improved OS upon immune checkpoint inhibitor treatment for patients with 9 types of advanced tumors³⁴. Analyses across several solid tumor types reported that patients with higher TMB (defined as ≥16-20 Muts/Mb) achieved greater clinical benefit from PD-1 or PD-L1-targeting monotherapy, compared with patients with higher TMB treated with chemotherapy⁴⁵ or those with lower TMB treated with PD-1 or PD-L1-targeting agents³⁵. However, the KEYNOTE 158 trial of pembrolizumab monotherapy for patients with solid tumors found significant improvement in ORR for patients with

TMB ≥10 Muts/Mb (based on this assay or others) compared to those with TMB <10 Muts/Mb, in a large cohort that included multiple tumor types; similar findings were observed in the KEYNOTE 028 and 012 trials37,44. Together, these studies suggest that patients with TMB ≥10 Muts/Mb may derive clinical benefit from PD-1 or PD-L1 inhibitors. In CRC specifically, a retrospective analysis of immune checkpoint inhibitor efficacy reported significantly improved OS for patients with tumors harboring TMB ≥9.8 Muts/MB compared with those with tumors with TMB < 9.8 Muts/Mb (~ equivalency <12 Muts/Mb as measured by this assay)34. Another retrospective study reported that a TMB ≥12 Muts/Mb cutoff identifies >99% of MSI-High CRC cases but only 3% of MSS cases, indicating the utility of this cutoff for identification of patients with CRC likely to benefit from treatment with immune checkpoint inhibitors⁴⁶.

FREQUENCY & PROGNOSIS

Elevated tumor mutational burden (TMB) has been reported in 8-25% of colorectal cancer (CRC) samples^{21,47-48}. Multiple studies have reported that up to 90% of hypermutated CRC cases exhibit high levels of microsatellite instability (MSI-H) and mismatch repair deficiency (MMR-D)^{21,47}. Increased TMB is significantly associated with MSI-H and MMR-D, with studies reporting that 100% of MSI-H CRCs harbor elevated TMB and conversely that 100% of tumors with low TMB harbor intact MMR⁴⁷. A subset of CRCs that harbor increased TMB but not MSI-H are driven

by mutations in POLE, which leads to an "ultramutated" phenotype with especially high TMB^{21,47}. Tumors with increased TMB harbor BRAF V6ooE mutations more frequently than those with low TMB^{21,47}, whereas TMB-low tumors more frequently harbor mutations in TP53 and APC²¹. In a study for 61 patients with metastatic, microsatellite stable (MSS) CRC treated with best standard of care, plasma TMB scores ≥28 muts/Mb (approximately 14 muts/Mb as measured by this assay) were associated with reduced OS as compared with plasma TMB scores <28 muts/Mb (3.0 vs. 5.3 months, HR 0.76, p=0.007), whereas tissue TMB was not found to be prognostic in this population⁴⁹.

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma⁵⁰⁻⁵¹ and cigarette smoke in lung cancer⁵²⁻⁵³, treatment with temozolomide-based chemotherapy in glioma⁵⁴⁻⁵⁵, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes^{21,56-59}, and microsatellite instability (MSI)^{21,56,59}. This sample harbors a TMB below levels that would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents^{34,44,46}.

GENOMIC FINDINGS

GENE

KRAS

ALTERATION

TRANSCRIPT ID

CODING SEQUENCE EFFECT

35G>A

VARIANT ALLELE FREQUENCY (% VAF)

83.4%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

Preclinical evidence suggests that KRAS activation may predict sensitivity to MEK inhibitors, such as trametinib, binimetinib, cobimetinib, and selumetinib⁶⁰⁻⁶⁵. However, multiple clinical trials have reported lack of efficacy of trametinib and other MEK inhibitors when used as monotherapy for treatment of patients with KRAS-mutant CRC66-70. Both clinical71-72 and preclinical73-74 studies suggest that combinatorial approaches including MEK inhibitors are likely to be more effective for the treatment of CRC, including strategies such as combination of MEK inhibitors with PI₃K inhibitors⁷², RAF inhibitors⁷³, pan-ERBB inhibitors⁷⁴, or chemotherapeutic agents⁷¹. In a Phase 1 study evaluating the MEK-pan-RAF dual inhibitor CH5126766, 6 patients harboring KRAS mutations experienced PRs, including 3

with non-small cell lung cancer (NCSLC), 1 with low-grade serous ovarian carcinoma (LGSOC), 1 with endometrial adenocarcinoma, and 1 with multiple myeloma⁷⁵. Combination of CH₅₁₂6766 with the FAK inhibitor defactinib elicited PR rates of 50% (4/8) for patients with KRAS-mutated low-grade serous ovarian cancer and 12% (2/17) for patients with KRAS-mutated non-small cell lung cancer (NSCLC) in a Phase 1 study76-77. Preclinical and clinical data suggest that KRAS mutations may predict clinical benefit from SHP2 inhibitors⁷⁸⁻⁷⁹. A Phase 1 study of RMC-4630 for relapsed/refractory solid tumors reported a DCR of 58% (23/40) for patients with NSCLC and KRAS mutations and a DCR of 75% (12/16) for patients with NSCLC and KRAS G12C mutations⁸⁰. Interim results from a Phase 1/2 study of RMC-4630 plus cobimetinib reported tumor reduction in 3 of 8 patients with KRASmutated colorectal cancer⁸¹. Preclinical data suggest that KRAS mutation may confer sensitivity to SOS1 inhibitors⁸²⁻⁸³. Phase 1 studies of the SOS1 inhibitor BI 1701963 alone or in combination with MEK inhibitors, KRAS G12C inhibitors, or irinotecan are recruiting for patients with solid tumors harboring KRAS mutations⁸⁴⁻⁸⁵ . Preclinical and limited clinical evidence suggest that KRAS mutation may predict sensitivity to PLK1 inhibitors⁸⁶. A Phase 1b/2 study of PLK1 inhibitor onvansertib in combination with FOLFIRI and bevacizumab for patients with KRAS-mutated metastatic CRC previously treated with chemotherapy reported an 87.5% (7/8; 3 PR, 4 SD) clinical benefit rate, with 1 patient going on

to successful curative surgery87.

- Potential Resistance -

Activating mutations in KRAS or NRAS are associated with lack of clinical benefit from cetuximab⁸⁸⁻⁹¹ or panitumumab⁹²⁻⁹⁴ for patients with CRC. Therefore, activating mutations in either gene indicate against the use of cetuximab and panitumumab (NCCN Colon Cancer Guidelines, v_{3.2021}).

FREQUENCY & PROGNOSIS

Mutations in KRAS have been reported in approximately 35-50% of colorectal cancers (CRCs)⁹⁵⁻¹⁰³. Numerous studies have reported that KRAS mutations are associated with increased metastasis, adverse clinicopathological features, and shorter survival of patients with CRC^{97-100,104-105}.

FINDING SUMMARY

KRAS encodes a member of the RAS family of small GTPases. Activating mutations in RAS genes can cause uncontrolled cell proliferation and tumor formation 61,106. KRAS alterations affecting amino acids G12, G13, Q22, P34, A59, Q61, and A146, as well as mutations G10_A11insG, G10_A11insAG (also reported as G10_A11dup and G12_G13insAG), A18D, L19F, D33E, G60_A66dup/E62_A66dup, E62K, E63K, R68S, and K117N have been characterized as activating and oncogenic 61,107-129.

GENE

NRAS

ALTERATION wildtype

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies

Lack of mutations in KRAS or NRAS is associated with clinical benefit of treatment with EGFR-

targeting antibodies cetuximab⁸⁸⁻⁹¹ or panitumumab⁹²⁻⁹⁴ for patients with CRC. Therefore, these agents are indicated to treat patients with CRC lacking such mutations (NCCN Guidelines v₃.2021).

FREQUENCY & PROGNOSIS

The majority of colorectal cancers (CRCs) (91-98%) have been reported to lack NRAS mutations $^{21,103,130-135}.$ NRAS wild-type status has been reported to be associated with decreased frequency of metastasis 103 and longer survival $^{135-136}$

of patients with CRC.

FINDING SUMMARY

NRAS encodes a member of the RAS family of small GTPases that mediate transduction of growth signals. Activation of RAS signaling causes cell growth, differentiation, and survival by activating the RAF-MAPK-ERK, PI₃K, and other pathways⁶¹. No alterations in NRAS were identified in this case.



GENOMIC FINDINGS

GENE

APC

ALTERATION T1556fs*3

TRANSCRIPT ID

NM_000038

CODING SEQUENCE EFFECT

4666_4667insA

VARIANT ALLELE FREQUENCY (% VAF)

72.4%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no approved drugs targeting APC inactivation in cancer. Loss of APC function leads to accumulation of beta-catenin and upregulation of WNT pathway transcription programs¹³⁷, and potential therapeutic approaches to target this pathway include CBP/beta-catenin antagonists, which interfere with the ability of beta-catenin to interact with transcriptional co-activator

CBP¹³⁸⁻¹³⁹. In a Phase 1 trial of the CBP/beta-catenin antagonist $E_{73}86$, 1 patient with APC-mutated small bowel adenocarcinoma achieved a PR with tumor shrinkage of -69% and response duration of 165 days¹⁴⁰; preclinical data support sensitivity of APC-deficient gastric or colorectal cancer models to $E_{73}86^{141-142}$.

FREQUENCY & PROGNOSIS

APC alterations have been found in 77% of tumors in the Colorectal Adenocarcinoma TCGA dataset²¹. Inactivation of APC leads to activation of the Wnt/beta-catenin pathway, which is thought to play a role in the adenoma-carcinoma transition in some cancers, including colorectal cancer (CRC)¹⁴³. The prognostic significance of APC mutations in sporadic CRC remains unclear¹⁴⁴. Solid tumors with WNT/beta-catenin pathway alterations, as seen here, were observed to have significantly less T-cell inflammation in one study¹⁴⁵.

FINDING SUMMARY

APC (adenomatous polyposis coli) encodes a tumor suppressor with critical roles in regulating

cell division and adhesion. APC interacts with beta-catenin and controls signaling in the WNT pathway, which regulates embryonic development and cell differentiation¹⁴⁶. Alterations such as seen here may disrupt APC function or expression¹⁴⁷⁻¹⁵¹.

POTENTIAL GERMLINE IMPLICATIONS

One or more of the APC variants observed here has been described in the ClinVar database as a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with familial adenomatous polyposis (ClinVar, Sep 2021)¹⁵². Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. Germline mutations in APC are found in more than 90% of patients with familial adenomatous polyposis (FAP)¹⁵³⁻¹⁵⁵. The prevalence for FAP in the general population is estimated to be 1:8,300 from birth¹⁵⁶, and in the appropriate clinical context germline testing of APC is recommended.

GENE

PIK3CA

ALTERATION

F545K

TRANSCRIPT ID

NM_006218

CODING SEQUENCE EFFECT

1633G>A

VARIANT ALLELE FREQUENCY (% VAF)

27.4%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

Clinical and preclinical data in various tumor types indicate that PIK₃CA activating alterations may predict sensitivity to therapies targeting PI₃K¹⁵⁷⁻¹⁵⁹, AKT¹⁶⁰⁻¹⁶¹, or mTOR¹⁶²⁻¹⁶⁹. Emerging

evidence suggests that the glutaminase inhibitor telaglenastat has clinical activity in PIK₃CA-mutated colorectal cancer (CRC). A Phase 1 trial of telaglenastat and capecitabine for patients with CRC who progressed on fluoropyrimidine chemotherapy observed numerically increased median PFS for patients with PIK₃CA mutation compared with patients with wildtype PIK₃CA status (24.8 vs. 16 weeks, n=7 vs. n=4), including SD >30 weeks for 3 patients with PIK₃CA mutation¹⁷⁰.

Potential Resistance

Multiple clinical studies report that inhibitors of the PI₃K-AKT-mTOR pathway have not produced significant clinical benefit as monotherapies to treat CRC, even for tumors that harbor alterations in PIK₃CA or PTEN; data are more limited for alterations in other genes in this pathway^{166,171-172}.

FREQUENCY & PROGNOSIS

PIK₃CA mutations have been reported in 10-20% of colorectal cancers^{21,96,173-174}. A meta-analysis of 864 patients with colorectal cancer treated with cetuximab- or panitumumab-based therapy showed that PIK₃CA mutations, particularly in exon 20 (H₁₀4₇R), are significantly associated with worse response¹⁷⁵ and shorter progression-free and overall survival¹³³.

FINDING SUMMARY

PIK₃CA encodes p₁₁₀-alpha, which is the catalytic subunit of phosphatidylinositol ₃-kinase (PI₃K). The PI₃K pathway is involved in cell signaling that regulates a number of critical cellular functions, including cell growth, proliferation, differentiation, motility, and survival¹⁷⁶⁻¹⁷⁷. PIK₃CA alterations that have been characterized as activating, such as observed here, are predicted to be oncogenic¹⁷⁸⁻¹⁹⁹.



GENOMIC FINDINGS

GENE

CDKN2A/B

ALTERATION

CDKN2B loss, CDKN2A loss

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

Preclinical data suggest that tumors with loss of p16INK4a function may be sensitive to CDK4/6 inhibitors, such as abemaciclib, ribociclib, and palbociclib²⁰⁰⁻²⁰³. Although case studies have reported that patients with breast cancer or uterine leiomyosarcoma harboring CDKN2A loss responded to palbociclib treatment²⁰⁴⁻²⁰⁵, multiple other clinical studies have shown no significant correlation between p16INK4a loss or inactivation and the rapeutic benefit of these agents $^{206\text{--}212};$ it is not known whether CDK4/6 inhibitors would be beneficial in this case. Although preclinical studies have suggested that loss of p14ARF function may be associated with reduced sensitivity to MDM2 inhibitors²¹³⁻²¹⁴, the clinical relevance of p14ARF as a predictive biomarker is not clear. There are no drugs that directly target the mutation or loss of CDKN2B in cancer. Because the p15INK4b protein encoded by CDKN2B is known to inhibit CDK4,

tumors with CDKN2B mutation or loss may predict sensitivity to CDK4/6 inhibitors, such as ribociclib, abemaciclib, and palbociclib^{207,209-210,215-217}.

FREQUENCY & PROGNOSIS

Homozygous deletion of both CDKN2A and CDKN2B has been found in 1% of colorectal adenocarcinomas^{21,218}. Inactivation of CDKN2A and CDKN2B by promoter methylation, also leading to loss of p15INK4b and p16INK4a expression, has been reported in colorectal adenocarcinoma²¹⁹⁻²²⁰. p16INK4a expression status has not been associated with patient prognosis in colorectal adenocarcinoma²¹⁹⁻²²¹.

FINDING SUMMARY

CDKN2A encodes two different, unrelated tumor suppressor proteins, p16INK4a and p14ARF, whereas CDKN2B encodes the tumor suppressor p15INK4b²²²⁻²²³. Both p15INK4b and p16INK4a bind to and inhibit CDK4 and CDK6, thereby maintaining the growth-suppressive activity of the Rb tumor suppressor; loss or inactivation of either p15INK4b or p16INK4a contributes to dysregulation of the CDK4/6-cyclin-Rb pathway and loss of cell cycle control²²⁴⁻²²⁵. The tumor suppressive functions of p14ARF involve

stabilization and activation of p53, via a mechanism of MDM2 inhibition²²⁶⁻²²⁷. One or more alterations observed here are predicted to result in p16INK4a loss of function²²⁸⁻²⁴⁹. One or more alterations seen here are predicted to result in p14ARF loss of function^{232,249-252}. CDKN2B alterations such as seen here are predicted to inactivate p15INK4b²⁵³.

POTENTIAL GERMLINE IMPLICATIONS

Germline CDKN2A mutation is associated with melanoma-pancreatic cancer syndrome, a condition marked by increased risk of developing malignant melanoma and/or pancreatic cancer²⁵⁴. Mutation carriers within families may develop either or both types of cancer, and melanoma cases may be referred to as familial or hereditary melanoma²⁵⁵⁻²⁵⁶. CDKN₂A is the most implicated gene in familial melanoma, with germline mutations present in 16% to 20% of familial melanoma cases²⁵⁷⁻²⁵⁹. CDKN₂A alteration has also been implicated in familial melanomaastrocytoma syndrome, an extremely rare tumor association characterized by dual predisposition to melanoma and nervous system tumors²⁶⁰⁻²⁶². In the appropriate clinical context, germline testing of CDKN2A is recommended.



GENOMIC FINDINGS

GENE

SMAD4

ALTERATION

E538*

TRANSCRIPT ID

NM_005359

CODING SEQUENCE EFFECT

1612G>T

VARIANT ALLELE FREQUENCY (% VAF)

81.9%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no therapies to address SMAD4 alterations in cancer. Preclinical studies²⁶³⁻²⁶⁴ and a clinical study of pancreatic cancer suggest that low SMAD4 expression exhibit increased responsiveness to chemotherapeutic agents such as cisplatin and irinotecan²⁶⁵.

FREQUENCY & PROGNOSIS

SMAD4 mutation or homozygous deletion is most frequently observed in pancreatic adenocarcinoma (43%)²⁶⁶, pancreatic acinar cell carcinoma²⁶⁷, cholangiocarcinoma (25%)²⁶⁸, appendiceal adenocarcinoma (14-20% mutation; 57% deletion)269-270, colorectal adenocarcinoma (CRC; 14%)²¹, esophageal adenocarcinoma (14%)²⁷¹, and stomach adenocarcinoma (13%)²⁷². In preclinical studies, SMAD4 loss of function has been implicated in the development of mucinous neoplasms of the pancreas, including mucinous cystic neoplasms (MCN)²⁷³ and intraductal papillary mucinous neoplasms (IPMN)²⁷⁴; in clinical samples, SMAD4 homozygous deletion has been observed in 10% of IPMNs and 8% of MCNs, and mutation was also observed in 5% of IPMNs²⁷⁵. SMAD4 gene alterations have been associated with reduced overall survival for patients with pancreatic adenocarcinoma²⁷⁶. Reduced SMAD4 expression has been associated with worse prognosis in various cancer types, including CRC²⁷⁷⁻²⁷⁹, appendiceal mucinous neoplasm²⁸⁰, gastric adenocarcinoma²⁸¹⁻²⁸², esophageal adenocarcinoma²⁸³, esophageal

squamous cell carcinoma²⁸⁴, breast cancer²⁸⁵, and prostate cancer²⁸⁶.

FINDING SUMMARY

SMAD4, also known as DPC4, encodes a tumor suppressor that regulates transcriptional activity downstream of TGF-beta receptor signaling²⁸⁷⁻²⁸⁸. SMAD4 alterations that result in loss or disruption of the MH1 domain (aa 18-142), MH2 domain (aa 323-552), or SAD domain (aa 275-320) are predicted to be inactivating²⁸⁹⁻³⁰².

POTENTIAL GERMLINE IMPLICATIONS

Germline SMAD4 mutations, including those at the R₃61 hotspot, have been observed in patients with juvenile polyposis syndrome³⁰³⁻³⁰⁵, which is associated with an increased risk of gastrointestinal cancers³⁰⁶. The penetrance of deleterious SMAD4 mutations in patients with colon cancer is estimated at 20% by age 35 and 70% by age 65³⁰⁷. In the appropriate clinical context, germline testing of SMAD4 is recommended.

GENE

SOX9

ALTERATION

Q411*

TRANSCRIPT ID

NM_000346

CODING SEQUENCE EFFECT

1231C>T

VARIANT ALLELE FREQUENCY (% VAF)

53.7%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no therapies available to directly address genomic alterations in SOX9.

FREQUENCY & PROGNOSIS

Mutation of SOX9 in cancer is typically rare, but it has been reported in 3-11% of colorectal carcinomas, most of which were truncating or frameshift alterations, and fewer than 4% of other tumor types (cBioPortal, COSMIC, Jan 2022)³⁰⁸⁻³¹⁰.

Increased expression of SOX9 has been associated with tumor development and/or increased aggressiveness of prostate cancer, pancreatic ductal adenocarcinoma, ovarian cancer, glioma, and esophageal adenocarcinoma³¹¹⁻³¹⁴.

FINDING SUMMARY

SOX9 encodes a transcription factor important for the development and differentiation of multiple tissues, including cartilage, testis, and prostate³¹⁵.



THERAPIES ASSOCIATED WITH RESISTANCE

IN PATIENT'S TUMOR TYPE

Cetuximab



Resistance of variant(s) to associated therapy is likely

Assay findings association

KRAS G12D

NRAS wildtype

AREAS OF THERAPEUTIC USE

Cetuximab is a monoclonal antibody that targets EGFR. It is FDA approved for the treatment of head and neck squamous cell carcinoma (HNSCC) and KRAS-wild-type, EGFR-expressing metastatic colorectal cancer (CRC). Please see the drug label for full prescribing information.

GENE ASSOCIATION

Therapies targeting EGFR, including cetuximab, have been shown to have significant clinical activity for patients with CRC^{88-91,316-317}; wild-type KRAS and NRAS are predictive biomarkers for the efficacy of cetuximab in metastatic CRC (NCCN Colon Cancer Guidelines v3.2021). Activating mutations in either KRAS⁸⁸⁻⁹¹ or NRAS^{133,318}, which function downstream of EGFR, are associated with lack of benefit of cetuximab for patients with CRC and indicate against the use of cetuximab (NCCN Guidelines v3.2021).

SUPPORTING DATA

Cetuximab has been shown to improve OS, PFS, and response rate for patients with KRAS-wild-type CRC, both as first-line combination therapy with FOLFIRI or

 ${
m FOLFOX4^{88-89,317}}$ and as monotherapy or combination therapy with irinotecan for chemotherapy-refractory patients^{90-91,316}. A prospective study of first-line cetuximab for patients with KRAS/NRAS/BRAF mutation-negative metastatic CRC resulted in limited efficacy, with 10.5% (2/19) of participants experiencing PRs and 57.9% (11/19) experiencing SDs³¹⁹. The Phase 2 AVETUX trial of cetuximab combined with avelumab and mFOLFOX6 for patients with RAS- and BRAF-wild-type metastatic CRC resulted in an ORR of 79.5% (6 CR and 25 PRs, n=39) and a DCR of $92.3\%^{320}$. In the Phase 3 ASPECCT study, panitumumab was found to be noninferior to cetuximab with respect to median OS (10.4 vs. 10.0 months, HR=0.97) for patients with previously treated KRAS exon 2 wildtype metastatic colorectal cancer; median PFS was also similar between the two treatment groups (4.4 vs. 4.1 months, HR=1.00)321. In a similar patient population, a Phase 2 study of combination panitumumab and irinotecan versus combination cetuximab and irinotecan also demonstrated non-inferiority with respect to median PFS (5.4 vs. 4.3 months, HR = 0.64) and median OS (14.9 vs. 11.5 months, HR=0.66)322.

Panitumumab



Resistance of variant(s) to associated therapy is likely

Assay findings association

KRAS G12D

NRAS wildtype

AREAS OF THERAPEUTIC USE

Panitumumab is a monoclonal antibody that targets EGFR. It is FDA approved to treat KRAS wild-type and NRAS wild-type metastatic colorectal cancer (CRC) combined with chemotherapy or as monotherapy for patients who have progressed on prior chemotherapy. Please see the drug label for full prescribing information.

GENE ASSOCIATION

Therapies targeting EGFR, including panitumumab, have been shown to have significant clinical activity for patients with CRC^{92,321,323}; wild-type KRAS and NRAS are predictive biomarkers for the efficacy of panitumumab in metastatic CRC (NCCN Colon Cancer Guidelines v3.2021). Activating mutations in either KRAS⁹²⁻⁹⁴ or NRAS^{93,324}, which function downstream of EGFR, are associated with lack of benefit of panitumumab for patients with CRC and indicate against the use of panitumumab (NCCN Guidelines v3.2021).

SUPPORTING DATA

Panitumumab has been shown to improve OS, PFS, and

ORR for patients with KRAS wild-type CRC, both as first-line combination therapy with FOLFOX492 and as monotherapy for chemotherapy-refractory patients^{321,323}. An open-label, randomized Phase 2 trial reported that for patients with unresectable RAS-wild-type colorectal adenocarcinoma treated with first-line panitumumab plus FOLFOX4, maintenance with a combination of panitumumab plus fluorouracil and leucovorin was superior to panitumumab monotherapy (10-month PFS 59% vs. 49%)325. In the Phase 3 ASPECCT study, panitumumab was found to be non-inferior to cetuximab with respect to median OS (10.4 vs. 10.0 months, HR=0.97) for patients with previously treated KRAS exon 2 wildtype metastatic colorectal cancer; median PFS was also similar between the two treatment groups (4.4 vs. 4.1 months, HR=1.00)321. In a similar patient population, a Phase 2 study of combination panitumumab and irinotecan versus combination cetuximab and irinotecan also demonstrated non-inferiority with respect to median PFS (5.4 vs. 4.3 months, HR = 0.64) and median OS (14.9 vs. 11.5 months, HR=0.66)322.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.



TUMOR TYPE Colon adenocarcinoma (CRC) REPORT DATE 10 Mar 2022



ORDERED TEST # ORD-1314037-01

CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial \Rightarrow Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov. Or visit https://www.foundationmedicine.com/genomictesting#support-services.

GENE APC

ALTERATION T1556fs*3

RATIONALE

LOCATIONS: Fukuoka (Japan), Nagaizumi-cho (Japan), Chuo Ku (Japan), Kashiwa (Japan)

LOCATIONS: Osakasayama (Japan), Chuo-Ku (Japan), Kashiwa (Japan)

Based on preclinical and limited clinical data, APC CBP/beta-catenin interaction inhibitors. inactivation may be associated with sensitivity to

NCT03833700	PHASE 1
A Study of E7386 in Participants With Advanced Solid Tumor Including Colorectal Cancer (CRC)	TARGETS CBP, Beta-catenin

NCT05091346	PHASE 1/2
A Study of E7386 in Combination With Pembrolizumab in Previously Treated Participants With Selected Solid Tumors	TARGETS CBP, Beta-catenin, PD-1
LOCATIONS: Tokyo (Japan)	

NCT04008797	PHASE 1
,	TARGETS CBP, Beta-catenin, FGFRs, RET, PDGFRA, VEGFRs, KIT

NCT03264664	PHASE 1
Study of E7386 in Participants With Selected Advanced Neoplasms	TARGETS CBP, Beta-catenin
LOCATIONS: Glasgow (United Kingdom), Manchester (United Kingdom), Sutton (United Kingdom)	

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CLINICAL TRIALS

GENE	
KRA	S

ALTERATION G12D

RATIONALE

KRAS activating mutations or amplification may predict sensitivity to inhibitors of MAPK pathway components, including MEK inhibitors. KRAS mutation may predict sensitivity to PLK1 inhibitors. Limited clinical and preclinical studies indicate KRAS mutations may predict sensitivity

to MEK-pan-RAF dual inhibitors. Multiple clinical studies have reported lack of efficacy of MEK inhibitors as monotherapy for treatment of KRAS-mutant colorectal cancer; combination therapies may be more effective.

NCT04803318	PHASE 2
Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors	TARGETS mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK
LOCATIONS: Guangzhou (China)	

NCT03284502	PHASE 1
Cobimetinib and HM95573 in Patients With Locally Advanced or Metastatic Solid Tumors	TARGETS MEK, RAFs

LOCATIONS: Hwasun (Korea, Republic of), Pusan (Korea, Republic of), Seongnam (Korea, Republic of), Seoul (Korea, Republic of), Goyang-si (Korea, Republic of)

NCT04303403	PHASE 1
Study of Trametinib and Ruxolitinib in Colorectal Cancer and Pancreatic Adenocarcinoma	TARGETS JAK2, JAK1, MEK

NCT04801966	PHASE NULL
Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study	TARGETS CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF

NCT03905148	PHASE 1/2
Study of the Safety and Pharmacokinetics of BGB-283 and PD-0325901 in Patients With Advanced or Refractory Solid Tumors	TARGETS RAFs, EGFR, MEK

LOCATIONS: Nedlands (Australia), Blacktown (Australia), Randwick (Australia), Melbourne (Australia), California, Texas

NCT02079740	PHASE 1/2
Trametinib and Navitoclax in Treating Patients With Advanced or Metastatic Solid Tumors	TARGETS BCL2, BCL-XL, BCL-W, MEK
LOCATIONS: Massachusetts	

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LOCATIONS: Singapore (Singapore)

LOCATIONS: Melbourne (Australia)



TUMOR TYPE
Colon adenocarcinoma (CRC)

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CLINICAL TRIALS

NCT02613650	PHASE 1
A Trial of mFOLFIRI With MEK162 in Patients With Advanced KRAS Positive Metastatic Colorectal Cancers	TARGETS MEK
LOCATIONS: Utah	
NCT04965818	PHASE 1/2
Phase 1b/2 Study of Futibatinib in Combination With Binimetinib in Patients With Advanced KRAS Mutant Cancer	TARGETS MEK, FGFRs
LOCATIONS: California, Indiana, Texas	
NCT03829410	PHASE 1/2
Onvansertib in Combination With FOLFIRI and Bevacizumab for Second Line Treatment of Metastatic Colorectal Cancer Patients With a Kras Mutation	TARGETS PLK1, VEGFA
LOCATIONS: California, Arizona, Minnesota, Kansas, Arkansas, Virginia, Florida	
NCT04800822	PHASE 1
PF-07284892 in Participants With Advanced Solid Tumors	TARGETS SHP2, ROS1, ALK, MEK, BRAF, EGFR
LOCATIONS: California, Michigan, New York, Tennessee, Texas	



CLINICAL TRIALS

PIK3CA

ALTERATION E545K

RATIONALE

PIK3CA activating mutations may lead to activation of the PI₃K-AKT-mTOR pathway and may therefore indicate sensitivity to inhibitors of this pathway. Strong clinical data support sensitivity of PIK3CA-mutated solid tumors to the PI₃K-alpha inhibitor alpelisib. Several clinical studies have shown that inhibitors of the PI₃K-AKT-mTOR pathway have not produced

significant clinical benefit when used as a monotherapy in patients with colorectal cancer; combination therapies may be required to overcome this lack of response. On the basis of preclinical and limited clinical data, PIK₃CA activating mutations may predict sensitivity to glutaminase inhibitors.

NCT04589845	PHASE 2
Tumor-Agnostic Precision Immuno-Oncology and Somatic Targeting Rational for You (TAPISTRY) Platform Study	TARGETS TRKB, ALK, TRKC, ROS1, TRKA, RET, PD-L1, AKTs, ERBB2, MDM2, PI3K- alpha

LOCATIONS: Zhongzheng Dist. (Taiwan), Taipei City (Taiwan), Taoyuan County (Taiwan), Tainan (Taiwan), Hong Kong (Hong Kong), Seoul (Korea, Republic of), Xi'an (China), Tianjin (China), Beijing (China), Chengdu City (China)

NCT04337463	PHASE NULL
ATG-008 Combined With Toripalimab in Advanced Solid Tumors	TARGETS mTORC1, mTORC2, PD-1
LOCATIONS: Chongqing (China), Chengdu (China)	

NCT04803318	PHASE 2
	TARGETS mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK

NCT03772561	PHASE 1
Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies	TARGETS PARP, AKTs, PD-L1
LOCATIONS: Singapore (Singapore)	

NCT04801966	PHASE NULL
Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study	TARGETS CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF
LOCATIONS: Melbourne (Australia)	

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LOCATIONS: Guangzhou (China)

PHASE 1/2



ORDERED TEST # ORD-1314037-01

NCT03711058

CLINICAL TRIALS

110103711030	PRASE 1/2
Study of PI3Kinase Inhibition (Copanlisib) and Anti-PD-1 Antibody Nivolumab in Relapsed/Refractory Solid Tumors With Expansions in Mismatch-repair Proficient (MSS) Colorectal Cancer	TARGETS PD-1, PI3K
LOCATIONS: Maryland	
NCT03006172	PHASE 1
To Evaluate the Safety, Tolerability, and Pharmacokinetics of GDC-0077 Single Agent in Participants With Solid Tumors and in Combination With Endocrine and Targeted Therapies in Participants With Breast Cancer	TARGETS PI3K-alpha, Aromatase, ER, CDK6, CDK4
LOCATIONS: London (United Kingdom), Surrey (United Kingdom), Bordeaux (France), Barcelona (Spai Massachusetts, New York, Tennessee	in), Valencia (Spain), Toronto (Canada),
NCT03673787	PHASE 1/2
A Trial of Ipatasertib in Combination With Atezolizumab	TARGETS AKTs, PD-L1
LOCATIONS: Sutton (United Kingdom)	
NCT03065062	PHASE 1
Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head & Neck and Other Solid Tumors	TARGETS PI3K-alpha, PI3K-gamma, mTORC1, mTORC2, CDK4, CDK6
LOCATIONS: Massachusetts	
NCT03217669	PHASE 1
Epacadostat (INCB24360) in Combination With Sirolimus in Advanced Malignancy	TARGETS IDO1, mTOR
LOCATIONS: Kansas	



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APPENDIX

Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

AR	BCOR	BCORL1	BRIP1
S176R	V878A	V872G	N775S
FLT3	INPP4B	JAK1	PARP3
N484S	Y919N	S537G	R408H
ROS1 D2213E and Q1708L	SPEN N2999K and T796S	TIPARP G307*	



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APPENDIX

Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

AND COLL MON	DER ALIERATIO	10						
ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTK	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL	CCND1
CCND2	CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2	DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRFI1	ESR1	EZH2
FAM46C	FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3
FGFR4	FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3
GATA4	GATA6	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3F3A
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDM5A	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3	P2RY8	PALB2
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA
PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET
RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2
SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOCS1
SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU
SYK	TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53
TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1	WT1	XPO1
XRCC2	ZNF217	ZNF703						
DNA GENE LIST:	FOR THE DETEC	TION OF SELECT	REARRANGEME	NTS				
ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
DADA	DET	POS1	PCPO2	SDCA	SICSANS	TEDC*	TEDT**	TMDDCC2

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSPO2	SDC4	SLC34A2	TERC*	TERT**	TMPRSS2

^{*}TERC is an NCRNA

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Loss of Heterozygosity (LOH) score Microsatellite (MS) status Tumor Mutational Burden (TMB)

^{**}Promoter region of TERT is interrogated

APPENDIX

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.

ABOUT FOUNDATIONONE CDX

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details: www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffinembedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLES

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture–selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g.,gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials Ranking of Therapies in Summary Table
Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials
Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

1. In the fractional-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-



APPENDIX

About FoundationOne®CDx

- Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.
- 2. TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh_docs/ pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
- 3. The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding armand chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
- 4. Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
- 5. Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine

whether the patient is a candidate for biopsy. Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an ERBB2 amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of ERBB2 copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in https://www.mycancergenome.org/content/ disease/breast-cancer/ERBB2/238/ report the frequency of HER2 overexpression as 20% in breast cancer. Based on the F1CDx HER2 CDx concordance study, approximately 10% of HER2 amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be

approximately 2%. **REPORT HIGHLIGHTS**

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
INDELS Repeatability	%CV*

^{*}Interquartile Range = 1^{st} Quartile to 3^{rd} Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of followup germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are ATM, BAP1, BRCA1, BRCA2, BRIP1, CHEK2, FH, FLCN, MLH1, MSH2, MSH6, MUTYH, PALB2, PMS2, POLE, RAD51C, RAD51D, RET, SDHA, SDHB, SDHC, SDHD, TSC2, and VHL, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are ASXL1, CBL, DNMT3A, IDH2, JAK2, KMT2D (MLL2), MPL, MYD88, SF3B1, TET2, and U2AF1 and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear



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About FoundationOne®CDx

cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
muts/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
os	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
ткі	Tyrosine kinase inhibitor

MR Suite Version 6.0.0

The median exon coverage for this sample is 1,030x

Electronically signed by Erik Williams, M.D. | 10 March 2022

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APPENDIX

References

- Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) pmid: 25392179
- 2. Kroemer G, et al. Oncoimmunology (2015) pmid: 26140250
- 3. Lal N, et al. Oncoimmunology (2015) pmid: 25949894
- 4. Le DT, et al. N. Engl. J. Med. (2015) pmid: 26028255
- 5. Ayers et al., 2016; ASCO-SITC Abstract P60
- 6. Ciardiello et al., 2018; ESMO Abstract LBA-004
- 7. Parikh et al., 2021; DOI: 10.1038/s43018-021-00269-7
- 8. Fukuoka S, et al. J. Clin. Oncol. (2020) pmid: 32343640
- 9. Kim et al., 2020; DOI: 10.1016/j.annonc.2020.04.073
- 10. Zhang Y, et al. BMC Gastroenterol (2021) pmid: 34688262
- 11. Sinicrope FA, et al. J. Clin. Oncol. (2013) pmid: 24019539
- Gavin PG, et al. Clin. Cancer Res. (2012) pmid: 23045248
- Bertagnolli MM, et al. J. Clin. Oncol. (2009) pmid: 13.
- Van Cutsem E, et al. J. Clin. Oncol. (2009) pmid:
- 15. Ribic CM, et al. N. Engl. J. Med. (2003) pmid: 12867608
- 16. Sargent DJ, et al. J. Clin. Oncol. (2010) pmid: 20498393
- 17. Fallik D, et al. Cancer Res. (2003) pmid: 14522894
- 18. Guastadisegni C, et al. Eur. J. Cancer (2010) pmid: 20627535
- 19. Pawlik TM, et al. Dis. Markers (2004) pmid: 15528785
- 20. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) pmid: 26337942
- 21. Nature (2012) pmid: 22810696
- 22. Histopathology (2007) pmid: 17204026
- Samowitz WS, et al. Cancer Epidemiol. Biomarkers Prev. (2001) pmid: 11535541
- Elsaleh H, et al. Clin Colorectal Cancer (2001) pmid: 24. 12445368
- 25. Brueckl WM, et al. Anticancer Res. () pmid: 12820457
- 26. Guidoboni M, et al. Am. J. Pathol. (2001) pmid:
- 27. Gryfe R, et al. N. Engl. J. Med. (2000) pmid: 10631274
- Sinicrope FA, et al. Gastroenterology (2006) pmid:
- 29. Laghi L, et al. Dig Dis (2012) pmid: 22722556
- 30. You JF, et al. Br. J. Cancer (2010) pmid: 21081928
- 31. Bairwa NK, et al. Methods Mol. Biol. (2014) pmid: 24623249
- 32. Boland CR, et al. Cancer Res. (1998) pmid: 9823339
- Boland CR, et al. Gastroenterology (2010) pmid: 20420947
- Samstein RM, et al. Nat. Genet. (2019) pmid: 30643254
- Goodman AM, et al. Mol. Cancer Ther. (2017) pmid: 28835386
- Goodman AM, et al. Cancer Immunol Res (2019) pmid: 31405947
- 37. Cristescu R, et al. Science (2018) pmid: 30309915
- Ready N, et al. J. Clin. Oncol. (2019) pmid: 30785829
- 39. Hellmann MD, et al. N. Engl. J. Med. (2018) pmid: 29658845
- 40. Hellmann MD, et al. Cancer Cell (2018) pmid: 29657128 41. Hellmann MD, et al. Cancer Cell (2018) pmid: 29731394
- 42. Rozeman EA, et al. Nat Med (2021) pmid: 33558721
- 43. Sharma P, et al. Cancer Cell (2020) pmid: 32916128
- 44. Marabelle A, et al. Lancet Oncol. (2020) pmid: 32919526
- 45. Legrand et al., 2018; ASCO Abstract 12000
- 46. Fabrizio DA, et al. J Gastrointest Oncol (2018) pmid:
- 47. Stadler ZK, et al. J. Clin. Oncol. (2016) pmid: 27022117
- 48. Shao C, et al. JAMA Netw Open (2020) pmid: 33119110

- 49. Chen EX, et al. JAMA Oncol (2020) pmid: 32379280
- 50. Pfeifer GP, et al. Mutat. Res. (2005) pmid: 15748635
- 51. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) pmid: 23875803
- 52. Pfeifer GP, et al. Oncogene (2002) pmid: 12379884
- 53. Rizvi NA, et al. Science (2015) pmid: 25765070
- 54. Johnson BE, et al. Science (2014) pmid: 24336570 55. Choi S, et al. Neuro-oncology (2018) pmid: 29452419
- Cancer Genome Atlas Research Network, et al. Nature (2013) pmid: 23636398
- 57. Briggs S, et al. J. Pathol. (2013) pmid: 23447401
- 58. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) pmid:
- 59. Roberts SA, et al. Nat. Rev. Cancer (2014) pmid:
- 60. Nakano H, et al. Proc. Natl. Acad. Sci. U.S.A. (1984) pmid: 6320174
- 61. Pylayeva-Gupta Y, et al. Nat. Rev. Cancer (2011) pmid: 21993244
- 62. Yamaguchi T, et al. Int. J. Oncol. (2011) pmid: 21523318
- 63. Watanabe M, et al. Cancer Sci. (2013) pmid: 23438367
- 64. Gilmartin AG, et al. Clin. Cancer Res. (2011) pmid:
- 65. Yeh JJ, et al. Mol. Cancer Ther. (2009) pmid: 19372556
- 66. Tsimberidou et al., 2013: ASCO Abstract e22086
- 67. Infante JR, et al. Lancet Oncol. (2012) pmid: 22805291
- Zimmer L, et al. Clin. Cancer Res. (2014) pmid: 24947927
- Bennouna J, et al. Invest New Drugs (2011) pmid: 20127139
- Weekes CD, et al. Clin. Cancer Res. (2013) pmid: 23434733
- 71. Hochster et al., 2013; ASCO GI Abstract 380
- 72. Juric et al., 2014; ASCO Abstract 9051
- 73. Lamba S, et al. Cell Rep (2014) pmid: 25199829
- 74. Sun C. et al. Cell Rep (2014) pmid: 24685132
- 75. Guo C. et al. Lancet Oncol (2020) pmid: 33128873
- 76. Krebs et al., 2021; AACR Abstract CT019
- 77. Shinde et al., 2020; AACR Abstract CT143
- 78. Lu H, et al. Mol Cancer Ther (2019) pmid: 31068384
- 79. Mainardi S, et al. Nat Med (2018) pmid: 29808006
- 80. Koczywas et al., 2021; AACR Abstract LB001
- 81. Bendell et al., 2020: FORTC-NCI-AACR Abstract 5 Hillig RC, et al. Proc Natl Acad Sci U S A (2019) pmid: 30683722
- 83. Hofmann MH, et al. Cancer Discov (2021) pmid:
- 84. Hofmann et al., 2021; AACR Abstract CT210
- 85. Gort et al., 2020; ASCO Abstract TPS3651
- 86. Luo J, et al. Cell (2009) pmid: 19490893
- 87. Barzi et al., 2020; AACR Abstract CT235
- 88. Van Cutsem E, et al. J. Clin. Oncol. (2011) pmid:
- 89. Bokemeyer C, et al. Ann. Oncol. (2011) pmid: 21228335
- 90. Karapetis CS, et al. N. Engl. J. Med. (2008) pmid: 18946061
- 91. De Roock W. et al. Ann. Oncol. (2008) pmid: 17998284
- 92. Douillard JY, et al. Ann. Oncol. (2014) pmid: 24718886
- Douillard JY, et al. N. Engl. J. Med. (2013) pmid: 24024839
- 94. Amado RG, et al. J. Clin. Oncol. (2008) pmid: 18316791 95. Lièvre A, et al. Cancer Res. (2006) pmid: 16618717
- 96. De Roock W, et al. Lancet Oncol. (2011) pmid: 21163703
- 97. Chen J, et al. BMC Cancer (2014) pmid: 25367198
- 98. Li W, et al. BMC Cancer (2015) pmid: 25929517
- 99. Hu J, et al. Medicine (Baltimore) (2016) pmid: 27977612

- 100. Zekri J, et al. Genet. Mol. Res. (2017) pmid: 28218784
- 101. Staudacher JJ, et al. Clin Transl Gastroenterol (2017)
- 102. Wang Y, et al. Virchows Arch. (2018) pmid: 29705968
- 103. Guo F, et al. Sci Rep (2018) pmid: 29666387
- 104. Mármol I, et al. Int J Mol Sci (2017) pmid: 28106826
- 105. Kwak MS, et al. Medicine (Baltimore) (2017) pmid: 28858102
- Kahn S, et al. Anticancer Res. () pmid: 3310850
- Akagi K, et al. Biochem. Biophys. Res. Commun. (2007) pmid: 17150185
- Bollag G, et al. J. Biol. Chem. (1996) pmid: 8955068
- 109. Buhrman G, et al. Proc. Natl. Acad. Sci. U.S.A. (2010) pmid: 20194776
- 110. Sci. STKE (2004) pmid: 15367757
- Edkins S. et al. Cancer Biol. Ther. (2006) pmid: 16969076
- 112. Feig LA, et al. Mol. Cell. Biol. (1988) pmid: 3043178
- 113. Gremer L. et al. Hum. Mutat. (2011) pmid: 20949621
- 114. Janakiraman M, et al. Cancer Res. (2010) pmid: 20570890
- 115. Kim E. et al. Cancer Discov (2016) pmid: 27147599
- Lukman S, et al. PLoS Comput. Biol. (2010) pmid: 20838576
- 117. Naguib A, et al. J Mol Signal (2011) pmid: 21371307
- 118. Prior IA, et al. Cancer Res. (2012) pmid: 22589270
- 119. Privé GG, et al. Proc. Natl. Acad. Sci. U.S.A. (1992) pmid: 1565661
- 120. Scheffzek K, et al. Science (1997) pmid: 9219684
- 121. Scholl C, et al. Cell (2009) pmid: 19490892 122. Smith G. et al. Br. J. Cancer (2010) pmid: 20147967
- 123. Tyner JW, et al. Blood (2009) pmid: 19075190
- 124. Valencia A, et al. Biochemistry (1991) pmid: 2029511
- 125. White Y. et al. Nat Commun (2016) pmid: 26854029
- 126. Wiest JS, et al. Oncogene (1994) pmid: 8058307
- 127. Angeles AKJ, et al. Oncol Lett (2019) pmid: 31289513 Tong JH, et al. Cancer Biol. Ther. (2014) pmid: 24642870
- 129. Loree JM, et al. Clin Cancer Res (2021) pmid: 34117033 Pentheroudakis G, et al. BMC Cancer (2013) pmid:
- 23374602 Vaughn CP, et al. Genes Chromosomes Cancer (2011) pmid: 21305640
- 132. Janku F, et al. Target Oncol (2013) pmid: 23400451
- 133. De Roock W, et al. Lancet Oncol. (2010) pmid: 20619739
- Irahara N, et al. Diagn. Mol. Pathol. (2010) pmid: 20736745
- Schirripa M, et al. Int. J. Cancer (2015) pmid: 24806288
- Cercek A, et al. Clin. Cancer Res. (2017) pmid: 136. 28446505
- Zhan T, et al. Oncogene (2017) pmid: 27617575
- 138. Jung YS, et al. Exp Mol Med (2020) pmid: 32037398
- Krishnamurthy N, et al. Cancer Treat Rev (2018) pmid: 29169144
- 140. Kawazoe et al., 2021; ESMO Abstract 473P
- 141. Yamada K, et al. Cancer Res (2021) pmid: 33408116 Kanda Y, et al. Biochem Biophys Res Commun (2022) pmid: 34837838
- 143. Fu Y, et al. Int. J. Cancer (2011) pmid: 21455986
- 144. Quyn AJ, et al. Surgeon (2008) pmid: 19110823
- 145. Luke JJ, et al. Clin Cancer Res (2019) pmid: 30635339
- Logan CY, et al. Annu. Rev. Cell Dev. Biol. (2004) pmid:
- 147. Eklof Spink K, et al. EMBO J. (2001) pmid: 11707392 148. Liu J, et al. J. Mol. Biol. (2006) pmid: 16753179
- 149. Dikovskaya D, et al. J. Cell. Sci. (2010) pmid: 20144988

150. Murphy SJ, et al. Dig. Dis. Sci. (2007) pmid: 17410430

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APPENDIX

References

- 151. Aretz S, et al. Hum. Mutat. (2004) pmid: 15459959
- 152. Landrum MJ, et al. Nucleic Acids Res. (2018) pmid:
- 153. Kerr SE, et al. J Mol Diagn (2013) pmid: 23159591
- 154. Annu Rev Pathol (2011) pmid: 21090969
- 155. Kastritis E, et al. Int. J. Cancer (2009) pmid: 18844223
- 156. Half E, et al. Orphanet J Rare Dis (2009) pmid: 19822006
- Fritsch C, et al. Mol. Cancer Ther. (2014) pmid: 24608574
- 158. Juric D. et al. J. Clin. Oncol. (2018) pmid: 29401002
- Gallant JN, et al. NPJ Precis Oncol (2019) pmid: 30793038
- 160. André F, et al. N. Engl. J. Med. (2019) pmid: 31091374
- Smyth LM, et al. NPJ Breast Cancer (2021) pmid: 33863913
- 162. Park HS, et al. PLoS ONE (2016) pmid: 27105424
- 163. Lim SM, et al. Oncotarget (2016) pmid: 26859683
- 164. Hou MM, et al. Oncotarget (2014) pmid: 25426553
- 165. Varnier R, et al. Eur J Cancer (2019) pmid: 31351267
- 166. Janku F, et al. Cell Rep (2014) pmid: 24440717
- 167. Moroney J. et al. Clin. Cancer Res. (2012) pmid: 22927482
- 168. Basho RK, et al. JAMA Oncol (2017) pmid: 27893038
- Moroney JW, et al. Clin. Cancer Res. (2011) pmid: 169.
- 170. Zhao Y, et al. Cancer Res (2020) pmid: 32907836
- 171. Ng K, et al. Clin. Cancer Res. (2013) pmid: 23743569
- Ganesan P, et al. Mol. Cancer Ther. (2013) pmid: 24092809
- 173. Li HT, et al. Oncol. Rep. (2011) pmid: 21424126
- 174. Brannon AR, et al. Genome Biol. (2014) pmid: 25164765
- 175. Huang L, et al. Arch Med Sci (2014) pmid: 24701207
- 176. Samuels Y, et al. Cancer Cell (2005) pmid: 15950905
- 177. Nat. Rev. Cancer (2009) pmid: 19629070
- Kang S, et al. Proc. Natl. Acad. Sci. U.S.A. (2005) pmid: 15647370
- 179 Ikenoue T, et al. Cancer Res. (2005) pmid: 15930273
- Gymnopoulos M, et al. Proc. Natl. Acad. Sci. U.S.A. (2007) pmid: 17376864
- Horn S, et al. Oncogene (2008) pmid: 18317450
- 182. Rudd ML, et al. Clin. Cancer Res. (2011) pmid: 21266528
- 183. Hon WC, et al. Oncogene (2012) pmid: 22120714
- Burke JE, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) pmid:
- Wu H, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) pmid: 185. 19915146
- 186. Laurenti R, et al. Rev Saude Publica (1990) pmid:
- 187. Dan S, et al. Cancer Res. (2010) pmid: 20530683
- 188. Oda K. et al. Cancer Res. (2008) pmid: 18829572
- 189. Zhao L, et al. Oncogene (2008) pmid: 18794883 190. Lui VW, et al. Cancer Discov (2013) pmid: 23619167
- 191. Ross RL, et al. Oncogene (2013) pmid: 22430209
- 192. Rivière JB, et al. Nat. Genet. (2012) pmid: 22729224
- 193. Shibata T, et al. Cancer Lett. (2009) pmid: 19394761
- 194. Dogruluk T, et al. Cancer Res. (2015) pmid: 26627007
- Croessmann S, et al. Clin. Cancer Res. (2018) pmid: 195.
- 196. Ng PK, et al. Cancer Cell (2018) pmid: 29533785
- 197. Spangle JM, et al. (2020) pmid: 32929011
- Chen L, et al. Nat Commun (2018) pmid: 29636477
- 199. Jin N, et al. J Clin Invest (2021) pmid: 34779417
- Konecny GE, et al. Clin. Cancer Res. (2011) pmid: 200.
- 201. Katsumi Y, et al. Biochem. Biophys. Res. Commun.

- (2011) pmid: 21871868
- 202. Cen L, et al. Neuro-oncology (2012) pmid: 22711607
- 203. Logan JE, et al. Anticancer Res. (2013) pmid: 23898052
- 204. Elvin JA, et al. Oncologist (2017) pmid: 28283584
- 205. Gao J, et al. Curr Oncol (2015) pmid: 26715889
- 206. Gopalan et al., 2014; ASCO Abstract 8077
- 207. Peguero et al., 2016; ASCO Abstract 2528 208. Konecny et al., 2016; ASCO Abstract 5557
- 209. DeMichele A, et al. Clin. Cancer Res. (2015) pmid:
- 25501126 210. Finn RS, et al. Lancet Oncol. (2015) pmid: 25524798
- 211. Infante JR, et al. Clin. Cancer Res. (2016) pmid:
- 212. Johnson DB, et al. Oncologist (2014) pmid: 24797823
- 213. Van Maerken T, et al. Mol. Cancer Ther. (2011) pmid:
- 214. Gamble LD, et al. Oncogene (2012) pmid: 21725357
- 215. Shapiro et al., 2013; ASCO Abstract 2500
- 216. Flaherty KT, et al. Clin. Cancer Res. (2012) pmid: 22090362
- 217. Dickson MA, et al. J. Clin. Oncol. (2013) pmid: 23569312
- 218. Giannakis M, et al. Cell Rep (2016) pmid: 27149842
- 219. Shima K. et al. Int. J. Cancer (2011) pmid: 20473920
- Nieminen TT, et al. Cancer Epidemiol. Biomarkers Prev. (2012) pmid: 22028395
- 221. Huh JW, et al. Hepatogastroenterology () pmid:
- 222. Quelle DE, et al. Cell (1995) pmid: 8521522
- 223. Mutat. Res. (2005) pmid: 15878778
- 224. Gazzeri S, et al. Oncogene (1998) pmid: 9484839
- 225. Oncogene (1999) pmid: 10498883
- Sherr CJ, et al. Cold Spring Harb. Symp. Quant. Biol. 226. (2005) pmid: 16869746
- 227. Ozenne P, et al. Int. J. Cancer (2010) pmid: 20549699
- 228. Ruas M, et al. Oncogene (1999) pmid: 10498896
- Jones R, et al. Cancer Res. (2007) pmid: 17909018
- 230. Haferkamp S, et al. Aging Cell (2008) pmid: 18843795 231. Huot TJ, et al. Mol. Cell. Biol. (2002) pmid: 12417717
- 232. Rizos H, et al. J. Biol. Chem. (2001) pmid: 11518711
- 233. Gombart AF, et al. Leukemia (1997) pmid: 9324288
- 234. Yang R, et al. Cancer Res. (1995) pmid: 7780957 235. Parry D, et al. Mol. Cell. Biol. (1996) pmid: 8668202
- 236. Greenblatt MS, et al. Oncogene (2003) pmid: 12606942
- 237. Yarbrough WG, et al. J. Natl. Cancer Inst. (1999) pmid: 10491434
- 238. Poi MJ, et al. Mol. Carcinog. (2001) pmid: 11255261
- 239. Byeon IJ, et al. Mol. Cell (1998) pmid: 9660926
- 240. Kannengiesser C, et al. Hum. Mutat. (2009) pmid: 19260062
- 241. Lal G. et al. Genes Chromosomes Cancer (2000) pmid: 10719365
- 242. Koh J, et al. Nature (1995) pmid: 7777061
- 243. McKenzie HA, et al. Hum. Mutat. (2010) pmid:
- 244. Miller PJ, et al. Hum. Mutat. (2011) pmid: 21462282
- 245. Kutscher CL, et al. Physiol. Behav. (1977) pmid: 905385
- 246. Scaini MC, et al. Hum. Mutat. (2014) pmid: 24659262
- 247. Jenkins NC, et al. J. Invest. Dermatol. (2013) pmid:
- 248. Walker GJ, et al. Int. J. Cancer (1999) pmid: 10389768
- 249. Rutter JL, et al. Oncogene (2003) pmid: 12853981 250. Itahana K, et al. Cancer Cell (2008) pmid: 18538737
- 251. Zhang Y, et al. Mol. Cell (1999) pmid: 10360174
- 252. Zhang Y, et al. Cell (1998) pmid: 9529249 253. Jafri M. et al. Cancer Discov (2015) pmid: 25873077

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- 254. Whelan AJ, et al. N Engl J Med (1995) pmid: 7666917
- 255. Adv Exp Med Biol (2010) pmid: 20687502
- Hogg D, et al. J Cutan Med Surg (1998) pmid: 9479083
- De Unamuno B, et al. Melanoma Res (2018) pmid: 29543703
- 258. Soura E, et al. J Am Acad Dermatol (2016) pmid: 26892650
- Huerta C, et al. Acta Derm Venereol (2018) pmid: 259.
- 260. Kaufman DK, et al. Neurology (1993) pmid: 8414022
- 261. Bahuau M, et al. Cancer Res (1998) pmid: 9622062
- 262. Chan AK, et al. Clin Neuropathol () pmid: 28699883 263. Cui Y, et al. Clin. Cancer Res. (2012) pmid: 22753594
- 264. Haeger SM, et al. Oncogene (2016) pmid: 25893305
- 265. Bachet JB, et al. Ann. Oncol. (2012) pmid: 22377565 Witkiewicz AK, et al. Nat Commun (2015) pmid: 25855536
- 267. Jiao Y, et al. J. Pathol. (2014) pmid: 24293293
- 268. Churi CR, et al. PLoS ONE (2014) pmid: 25536104
- 269. Liu X. et al. Clin. Chem. (2014) pmid: 24821835
- 270. Maru D, et al. Oncogene (2004) pmid: 14647445
- 271. Wang K, et al. Oncologist (2015) pmid: 26336083
- 272. Nature (2014) pmid: 25079317
- 273. Izeradjene K, et al. Cancer Cell (2007) pmid: 17349581
- 274. Bardeesy N, et al. Genes Dev. (2006) pmid: 17114584
- Springer S, et al. Gastroenterology (2015) pmid: 275. 26253305
- Blackford A, et al. Clin. Cancer Res. (2009) pmid:
- 19584151 Yan P, et al. Clin. Cancer Res. (2016) pmid: 26861460
- Kozak MM, et al. J. Clin. Pathol. (2015) pmid: 25681512
- Roth AD, et al. J. Natl. Cancer Inst. (2012) pmid:
- Davison JM, et al. Am. J. Surg. Pathol. (2014) pmid: 280. 24618609
- 281. Kim YH, et al. Ann. Oncol. (2004) pmid: 15033661
- Xiangming C, et al. Clin. Cancer Res. (2001) pmid: 282. 11234879
- Singhi AD, et al. Am. J. Surg. Pathol. (2015) pmid:
- 25634752 Natsugoe S, et al. Clin. Cancer Res. (2002) pmid:
- 12060625
- de Kruijf EM, et al. Ann. Oncol. (2013) pmid: 23022998 286. Shipitsin M, et al. Br. J. Cancer (2014) pmid: 25032733
- Nat. Rev. Mol. Cell Biol. (2012) pmid: 22992590
- 288. Cell (2008) pmid: 18662538 289. Massagué J, et al. Genes Dev. (2005) pmid: 16322555
- 290. Morén A, et al. Oncogene (2000) pmid: 10980615 291. Xu J. et al. Proc. Natl. Acad. Sci. U.S.A. (2000) pmid:
- 292. Luo K, et al. Genes Dev. (1999) pmid: 10485843
- Jones JB, et al. Nucleic Acids Res. (2000) pmid:
- 294. Fink SP, et al. Cancer Res. (2001) pmid: 11196171 295. De Bosscher K, et al. Biochem, J. (2004) pmid: 14715079
- 296. Shi Y, et al. Nature (1997) pmid: 9214508
- 297. Miyaki M, et al. Oncogene (1999) pmid: 10340381
- 298. Prokova V. et al. Biochemistry (2007) pmid: 17994767 299. Wu JW, et al. J. Biol. Chem. (2001) pmid: 11274206
- 300. Ding L, et al. J. Clin. Invest. (2009) pmid: 19139564
- **301.** Kuang C, et al. Oncogene (2004) pmid: 14647410 302. Watanabe M, et al. EMBO Rep. (2000) pmid: 11265759 Houlston R, et al. Hum. Mol. Genet. (1998) pmid:
- Woodford-Richens K, et al. Gut (2000) pmid: 10764709

305. Howe JR, et al. J. Med. Genet. (2004) pmid: 15235019

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TUMOR TYPE
Colon adenocarcinoma (CRC)

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APPENDIX

References

- **306.** Brosens LA, et al. World J. Gastroenterol. (2011) pmid: 22171123
- **307.** Kalia SS, et al. Genet. Med. (2017) pmid: 27854360
- 308. Cerami E, et al. Cancer Discov (2012) pmid: 22588877
- **309.** Gao J, et al. Sci Signal (2013) pmid: 23550210
- 310. Tate JG, et al. Nucleic Acids Res. (2019) pmid: 30371878
- 311. Kopp JL, et al. Cancer Cell (2012) pmid: 23201164 312. Zhong WD, et al. BMC Cancer (2012) pmid: 22703285
- 313. Wang L, et al. Med. Oncol. (2012) pmid: 22714060
- 314. Clemons NJ, et al. Am. J. Physiol. Gastrointest. Liver Physiol. (2012) pmid: 23064761
- **315.** Pritchett J, et al. Trends Mol Med (2011) pmid: 21237710
- **316.** Cunningham D, et al. N. Engl. J. Med. (2004) pmid: 15269313
- 317. Jonker DJ, et al. N. Engl. J. Med. (2007) pmid: 18003960
- **318.** Di Bartolomeo M, et al. Target Oncol (2014) pmid: 23821376
- 319. Moiseyenko VM, et al. Clin Drug Investig (2018) pmid: 29470838
- 320. Stein et al., 2020; ASCO GI Abstract 96
- **321.** Price TJ, et al. Lancet Oncol. (2014) pmid: 24739896
- 322. Sakai D, et al. Eur J Cancer (2020) pmid: 32526634
- **323.** Van Cutsem E, et al. J. Clin. Oncol. (2007) pmid: 17470858
- Peeters M, et al. Clin. Cancer Res. (2013) pmid: 23325582
- **325.** Pietrantonio F, et al. JAMA Oncol (2019) pmid: 31268481