

PATIENT Chien, Li-Shu TUMOR TYPE
Nasopharynx and paranasal
sinuses undifferentiated
carcinoma
COUNTRY CODE

REPORT DATE 05 Oct 2021

ORDERED TEST # ORD-1194451-01

**ABOUT THE TEST** FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

**PATIENT** 

**DISEASE** Nasopharynx and paranasal sinuses undifferentiated

carcinoma

NAME Chien, Li-Shu

DATE OF BIRTH 01 December 1966

**SEX** Female

MEDICAL RECORD # 19906060

**PHYSICIAN** 

ORDERING PHYSICIAN Mu-Hsin Chang, Peter

**MEDICAL FACILITY** Taipei Veterans General Hospital

ADDITIONAL RECIPIENT None MEDICAL FACILITY ID 205872

PATHOLOGIST Not Provided

**SPECIMEN** 

SPECIMEN SITE Lung

SPECIMEN ID \$109-16673C (PF21020)

SPECIMEN TYPE Slide Deck

DATE OF COLLECTION 05 June 2020

SPECIMEN RECEIVED 28 September 2021

**Biomarker Findings** 

TW

Microsatellite status - MS-Stable

Tumor Mutational Burden - 0 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

**SMARCB1** loss

CHEK2 loss exons 2-10

1 Therapies with Clinical Benefit0 Therapies with Resistance

18 Clinical Trials

BIOMARKER FINDINGS

Microsatellite status - MS-Stable

Tumor Mutational Burden - 0 Muts/Mb

**GENOMIC FINDINGS** 

**SMARCB1 - loss** 

9 Trials see p. 8

CHEK2 - loss exons 2-10

10 Trials see p. 6

No therapies or clinical trials. see Biomarker Findings section		
No therapies or clinical trials. see Biomarker Findings section		
THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)	
none	Tazemetostat	
none	none	

THERAPY AND CLINICAL TRIAL IMPLICATIONS

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patients tumor type. This report should be regarded and used as upplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.



ORDERED TEST # ORD-1194451-01

**BIOMARKER FINDINGS** 

#### BIOMARKER

# Microsatellite status

RESULT MS-Stable

#### **POTENTIAL TREATMENT STRATEGIES**

#### - Targeted Therapies -

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors<sup>1-3</sup>, including approved therapies nivolumab and pembrolizumab<sup>4</sup>. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and

experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, p=0.001)<sup>5</sup>.

#### **FREQUENCY & PROGNOSIS**

MSI-high (MSI-H) has been observed at high frequency in endometrial cancers  $(14-33\%)^{6-13}$ , colorectal cancers (CRCs;  $10-15\%)^{3,14-17}$ , and gastric cancers ( $12-35\%)^{18-21}$  and at lower frequencies in many other tumor types, including esophageal<sup>22</sup>, small bowel<sup>23-27</sup>, hepatobiliary<sup>28-34</sup>, prostate<sup>35-37</sup>, and urinary tract carcinomas<sup>38-40</sup>. One retrospective study of patients with Stage 2-4a nasopharyngeal carcinoma treated with intensity-modulated radiotherapy in China reported that deficiency of mismatch repair protein expression independently correlated with better distant metastasis-free survival (HR=0.25)<sup>41</sup>.

#### **FINDING SUMMARY**

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor<sup>16</sup>. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH<sub>2</sub>, MSH<sub>6</sub>, or PMS<sub>2</sub><sup>16,42-43</sup>. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers<sup>15,44-45</sup>. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins15-16,43,45.

#### BIOMARKER

# Tumor Mutational Burden

RESULT 0 Muts/Mb

# POTENTIAL TREATMENT STRATEGIES

# Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1<sup>46-48</sup>, anti-PD-1 therapies<sup>46-49</sup>, and combination nivolumab and ipilimumab<sup>50-55</sup>. In multiple pan-tumor studies, higher TMB has been reported to be associated with increased ORR and OS from treatment with immune checkpoint inhibitors<sup>46-49,56</sup>. Higher TMB was found to be significantly associated with improved OS upon immune checkpoint inhibitor treatment for patients with 9 types of advanced tumors<sup>46</sup>.

Analyses across several solid tumor types reported that patients with higher TMB (defined as ≥16-20 Muts/Mb) achieved greater clinical benefit from PD-1 or PD-L1-targeting monotherapy, compared with patients with higher TMB treated with chemotherapy<sup>57</sup> or those with lower TMB treated with PD-1 or PD-L1-targeting agents<sup>47</sup>. However, the KEYNOTE 158 trial of pembrolizumab monotherapy for patients with solid tumors found significant improvement in ORR for patients with TMB ≥10 Muts/Mb (based on this assay or others) compared to those with TMB <10 Muts/Mb, in a large cohort that included multiple tumor types; similar findings were observed in the KEYNOTE 028 and 012 trials<sup>49,56</sup>. Together, these studies suggest that patients with TMB ≥10 Muts/Mb may derive clinical benefit from PD-1 or PD-L1 inhibitors.

#### **FREQUENCY & PROGNOSIS**

Nasopharyngeal and sinonasal undifferentiated carcinoma harbors a median TMB of 2.7 mutations per megabase (muts/Mb), and 1.9% of cases have high TMB (>20 muts/Mb)<sup>58</sup>. Published

data investigating the prognostic implications of TMB in nasopharyngeal and sinonasal carcinoma are limited (PubMed, Oct 2021).

#### **FINDING SUMMARY**

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma<sup>59-60</sup> and cigarette smoke in lung cancer<sup>61-62</sup>, treatment with temozolomide-based chemotherapy in glioma<sup>63-64</sup>, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes<sup>17,65-68</sup>, and microsatellite instability (MSI)<sup>17,65,68</sup>. This sample harbors a TMB level associated with lower rates of clinical benefit from treatment with PD-1- or PD-L1-targeting immune checkpoint inhibitors compared with patients with tumors harboring higher TMB levels, based on several studies in multiple solid tumor types<sup>47-48,56</sup>.



ORDERED TEST # ORD-1194451-01

**GENOMIC FINDINGS** 

**GENE** 

# **SMARCB1**

ALTERATION loss

# **POTENTIAL TREATMENT STRATEGIES**

#### Targeted Therapies —

Clinical and preclinical evidence suggests that tumors with loss of SMARCB1 may be sensitive to a variety of targeted therapies, including EZH2 inhibitors<sup>69-72</sup>, CDK<sub>4</sub>/6 inhibitors<sup>73-76</sup>, Aurora kinase inhibitors<sup>77</sup>, and anti-PD-1 immune checkpoint inhibitors 78-81. The EZH2 inhibitor tazemetostat has shown efficacy in adult and pediatric patients with INI1-negative solid tumors across Phase 1 and 2 trials, predominantly in sarcoma subtypes<sup>69</sup>. A clinical trial of the Aurora kinase A inhibitor alisertib for pediatric patients with atypical teratoid rhabdoid tumors, which are characterized by SMARCB1 loss or inactivation, reported disease stabilization or regression in 100% (4/4) of patients, and 2 patients achieved disease regression lasting more than 1 year82. A study of INI1-negative pediatric solid tumors observed frequent signs of T-cell inflammation, including PD-L1 expression in 47% (14/30) of

patients, and reported 3 patients with metastatic epithelioid sarcoma, anaplastic chordoma, or malignant rhabdoid tumor who experienced disease control on pembrolizumab or nivolumab for at least 12, 9, and 3 months, respectively<sup>78</sup>. Case studies have reported a patient with treatment-refractory metastatic epithelioid sarcoma had a PR on nivolumab plus ipilimumab, with the addition of pazopanib leading to a CR of >12 months<sup>83</sup>; a patient with SMARCB<sub>1</sub>-mutated metastatic renal medullary carcinoma (RMC) had SD for 7 months with nivolumab84; and a patient with metastatic RMC and concurrent SMARCB1 loss and ARID2 mutation had a CR with 9 months of nivolumab treatment<sup>85</sup>. Preclinical evidence also supports PD-1 immune checkpoint blockade as treatment approach for rhabdoid tumors that are characterized by SMARCB1 inactivation and T-cell inflammation<sup>79-80</sup>.

#### **FREQUENCY & PROGNOSIS**

The loss of SMARCB1 protein expression was reported in 6% of 142 sinonasal carcinomas in another study, with genomic loss of SMARCB1 identified in 75% (6/8) of patients in that subset<sup>86</sup>. In a genomic analysis of 56 nasopharyngeal carcinoma cases, alterations of SMARCB1 was not detected in any of the samples<sup>87</sup>. Loss of SMARCB1 has been reported in all three sinonasal

basaloid carcinomas analyzed in one study<sup>88</sup>. In head and neck carcinoma, SMARCB1 inactivation may indicate an INI1-deficient subtype of sinonasal carcinoma characterized by poor differentiation and aggressive clinicopathologic features<sup>86,89</sup>.

#### **FINDING SUMMARY**

The SMARCB1 gene encodes the SNF5 protein, also known as INI1, one of three core subunits of the SWI/SNF family of chromatin remodeling complexes<sup>90</sup>. SNF5, as a member of the remodeling complex, plays a key role in cell cycle control and can act as a tumor suppressor<sup>91</sup>. Alterations such as seen here may disrupt SMARCB1 function or expression<sup>92-97</sup>.

#### POTENTIAL DIAGNOSTIC IMPLICATIONS

SMARCB1 mutation and loss of protein expression are characteristic of malignant rhabdoid tumors, INI1-deficient sinonasal carcinomas, epithelioid sarcomas, kidney medullary carcinoma and atypical teratoid/rhabdoid tumors, particularly in cases with low TMB and a paucity of co-occurring alterations (NCCN Soft Tissue Sarcoma Guidelines, v2.2021)<sup>78,86,89,98-103</sup>.

**GENOMIC FINDINGS** 

ORDERED TEST # ORD-1194451-01

FOUNDATION**ONE®CD**x

**GENE** CHEK2

ALTERATION loss exons 2-10

#### POTENTIAL TREATMENT STRATEGIES

# - Targeted Therapies -

Limited clinical data indicate that CHEK2 inactivation may predict sensitivity to PARP inhibitors. Patients with CHEK2-altered prostate cancer have experienced clinical responses to PARP inhibitors<sup>104-106</sup>. Clinical benefit has been observed for patients with ovarian107 and testicular108 cancers treated with PARP inhibitors. One study of patients with breast cancer reported that carriers of the CHEK2 H371Y mutation have a higher likelihood of response to neoadjuvant chemotherapy<sup>109</sup>, whereas another study found that CHEK2 mutation carriers have a lower frequency of objective clinical responses to neoadjuvant therapy110. A third study reported

that the CHEK2 1100delC mutation is not associated with differential efficacy of chemotherapy and endocrine therapy in patients with metastatic breast cancer<sup>111</sup>.

#### **FREQUENCY & PROGNOSIS**

CHEK2 mutations have been reported in 8.3% of glioblastoma (GBM) samples and in carcinomas of the endometrium (3.8%), urinary tract (3.5%), ovary (3.4%), and large intestine (3.1%), and at low frequency in a variety of solid and hematologic cancer types (COSMIC, 2021)<sup>112</sup>. In breast cancer, certain CHEK2 mutations are associated with higher grade and larger tumors as well as bilateral disease<sup>113</sup>. A study reported that a polymorphism in CHEK2 was associated with worse survival of patients with GBM, but this association lost significance after adjusting for other prognostic factors<sup>114-115</sup>. Another study in prostate cancer reported that CHEK2 expression is decreased in higher grade tumors and that CHEK2 is a tumor suppressor that decreases the growth of prostate cancer cells and regulates androgen receptor signaling<sup>116</sup>.

#### **FINDING SUMMARY**

CHEK2 encodes the protein checkpoint kinase 2, a serine/threonine kinase that plays an important role in the DNA-damage response; it is a putative tumor suppressor<sup>117-120</sup>. Alterations such as seen here may disrupt CHEK2 function or expression<sup>121-131</sup>.

#### **POTENTIAL GERMLINE IMPLICATIONS**

Germline CHEK2 mutation has been associated with cancer susceptibility of low to moderate penetrance, especially in hereditary breast cancer132. CHEK2 germline mutation has been identified in approximately 2.5% of familial or high-risk breast cancer cases<sup>133-134</sup>. Although heterozygous germline CHEK2 mutation increases breast cancer risk two- to three-fold, it is not associated with younger age at diagnosis134-135. In the appropriate clinical context, germline testing of CHEK2 is recommended.

REPORT DATE 05 Oct 2021

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

ORDERED TEST # ORD-1194451-01

FOUNDATION**ONE®CD**x

# **Tazemetostat**

Assay findings association

**SMARCB1** loss

#### **AREAS OF THERAPEUTIC USE**

Tazemetostat is an inhibitor of the histone methyltransferase EZH2. It is FDA approved as a monotherapy for the treatment of metastatic or locally advanced epithelioid sarcoma and relapsed or refractory follicular lymphoma. Please see the drug label for full prescribing information.

#### **GENE ASSOCIATION**

On the basis of Phase 2 clinical data across sarcoma subtypes and sinonasal carcinoma  $^{70,136-138}$ , and preclinical evidence<sup>72,139</sup>, loss or inactivation of SMARCB1 may predict sensitivity to tazemetostat.

#### **SUPPORTING DATA**

Clinical data on the efficacy of tazemetostat for the treatment of head and neck cancer are limited (PubMed, Aug 2021). In solid tumors, tazemetostat has been primarily investigated in the context of INI1-deficient sarcomas. A Phase 2 study for patients with INI1-negative tumors treated with tazemetostat reported ORRs (all PRs) of 15% (9/62) for the epithelioid sarcoma cohort  $^{136}$ , 0% (0/ 11) for the malignant rhabdoid tumor (MRT) cohort<sup>140</sup>, and 9.4% (3/32) for those with other solid tumors, including PRs in 2 of 13 patients with sarcoma (both spindle cell sarcoma) and 1 of 16 patients with non-mesenchymal tumors (a sinonasal carcinoma)137. For patients with synovial sarcoma, a Phase 2 study reported that treatment with tazemetostat monotherapy achieved SD as best response in 33% (11/33) of patients, with 5 SDs lasting ≥16 weeks<sup>141</sup>. Additional Phase 1/2 trials for tazemetostat have reported clinical benefit for adult patients with small cell carcinoma of the ovary, hypercalcemic type (2 PRs)70,140,142 malignant mesothelioma (2 PRs)<sup>143</sup>, and MRT (1 CR)<sup>70</sup>. A Phase 1 trial for 46 pediatric patients with synovial sarcoma or INI1-negative tumors reported clinical benefit in the context of epithelioid sarcoma (1 CR), chordoma (1 CR, 1PR), or atypical teratoid rhabdoid tumor (ATRT, 1 CR) across tazemetostat dosage groups<sup>138</sup>.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

TUMOR TYPE

carcinoma

Nasopharynx and paranasal

sinuses undifferentiated

REPORT DATE 05 Oct 2021

**CLINICAL TRIALS** 

# ORDERED TEST # ORD-1194451-01

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

FOUNDATION**ONE®CD**x

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity > Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov. Or visit https://www.foundationmedicine.com/genomictesting#support-services.

**GENE** CHEK2

On the basis of clinical evidence in prostate and other solid cancers, CHEK2 loss or inactivation

may confer sensitivity to PARP inhibitors.

ALTERATION loss exons 2-10

NCT04123366 PHASE 2

**RATIONALE** 

Study of Olaparib (MK-7339) in Combination With Pembrolizumab (MK-3475) in the Treatment of Homologous Recombination Repair Mutation (HRRm) and/or Homologous Recombination Deficiency (HRD)-Positive Advanced Cancer (MK-7339-007/KEYLYNK-007)

**TARGETS** PARP, PD-1

LOCATIONS: Fukuoka (Japan), Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Okayama (Japan), Nagoya (Japan), Tokyo (Japan), Kashiwa (Japan), Sapporo (Japan), Nedlands (Australia), Southport (Australia)

NCT03742895 PHASE 2

Efficacy and Safety of Olaparib (MK-7339) in Participants With Previously Treated, Homologous Recombination Repair Mutation (HRRm) or Homologous Recombination Deficiency (HRD) Positive Advanced Cancer (MK-7339-002 / LYNK-002)

**TARGETS PARP** 

LOCATIONS: Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Chelyabinsk (Russian Federation), Nedlands (Australia), Kazan (Russian Federation), Arkhangelsk (Russian Federation), Port Macquarie (Australia), Ryazan (Russian Federation), Darlinghurst (Australia), Moscow (Russian Federation)

NCT02264678 PHASE 1/2

Ascending Doses of AZD6738 in Combination With Chemotherapy and/or Novel Anti Cancer Agents **TARGETS** 

ATR, PARP, PD-L1

LOCATIONS: Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Cambridge (United Kingdom), Withington (United Kingdom), London (United Kingdom), Sutton (United Kingdom), Villejuif (France), Saint Herblain (France), California

NCT04635631 **PHASE 1** 

STUDY OF TALAZOPARIB MONOTHERAPY IN CHINESE PARTICIPANTS WITH ADVANCED SOLID **TUMORS** 

**TARGETS PARP** 

LOCATIONS: Beijing (China), Changchun (China)

NCT03772561 **PHASE 1** 

Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid **Tumor Malignancies** 

**TARGETS** 

PARP, AKTs, PD-L1

**LOCATIONS:** Singapore (Singapore)



ORDERED TEST # ORD-1194451-01

CLINICAL TRIALS

NCT04801966	PHASE NULL			
Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study	TARGETS CDK4, CDK6, PI3K-alpha, PD-L1, MEK PARP, PD-1, BRAF			
LOCATIONS: Melbourne (Australia)				
NCT02769962	PHASE 1/2			
Trial of CRLX101, a Nanoparticle Camptothecin With Olaparib in People With Relapsed/Refractory Small Cell Lung Cancer	TARGETS PARP, TOP1			
LOCATIONS: Maryland				
NCT04497116	PHASE 1/2			
Study of RP-3500 in Advanced Solid Tumors	TARGETS ATR, PARP			
LOCATIONS: Copenhagen (Denmark), Newcastle Upon Tyne (United Kingdom), Manchester (United (Canada), Massachusetts, Rhode Island, New York, Tennessee, Texas	Kingdom), London (United Kingdom), Toronto			
NCT03127215	PHASE 2			
Study of Olaparib/Trabectedin vs. Doctor's Choice in Solid Tumors	TARGETS FUS-DDIT3, PARP			
LOCATIONS: Dresden (Germany), München (Germany), Frankfurt (Germany), Essen (Germany), Ma	FUS-DDIT3, PARP			
<b>LOCATIONS:</b> Dresden (Germany), München (Germany), Frankfurt (Germany), Essen (Germany), Ma (Germany), Tuebingen (Germany), Freiburg (Germany)	FUS-DDIT3, PARP			
Study of Olaparib/Trabectedin vs. Doctor's Choice in Solid Tumors  LOCATIONS: Dresden (Germany), München (Germany), Frankfurt (Germany), Essen (Germany), Ma (Germany), Tuebingen (Germany), Freiburg (Germany)  NCT03297606  Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)	FUS-DDIT3, PARP inz (Germany), Heidelberg (Germany), Stuttgart			



ORDERED TEST # ORD-1194451-01

CLINICAL TRIALS

GENE	
<b>SMA</b>	RCB1

ALTERATION loss

#### **RATIONALE**

On the basis of clinical and preclinical evidence, loss or inactivation of SMARCB1 may be associated with sensitivity to EZH2 inhibitors, CDK4/6 inhibitors, and Aurora kinase inhibitors.

Patients with certain SMARCB1-deficient malignant tumors have benefited from anti-PD-1 immune checkpoint inhibitors.

NCT03099174	PHASE 1		
This Study in Patients With Different Types of Cancer (Solid Tumours) Aims to Find a Safe Dose of Xentuzumab in Combination With Abemaciclib With or Without Hormonal Therapies. The Study Also Tests How Effective These Medicines Are in Patients With Lung and Breast Cancer.	TARGETS CDK4, CDK6, IGF-1, IGF-2, Aromatase, ER		
<b>LOCATIONS:</b> Seoul (Korea, Republic of), Goyang (Korea, Republic of), Aichi, Nagoya (Japan), Kanaga Koto-ku (Japan), Chiba, Kashiwa (Japan), Helsinki (Finland), Tampere (Finland), Turku (Finland)	wa, Isehara (Japan), Tokyo, Chuo-ku (Japan), Tokyo,		
NCT04801966	PHASE NULL		
Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study	TARGETS CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF		
LOCATIONS: Melbourne (Australia)			
NCT04104776	PHASE 1/2		
A Study of CPI-0209 in Patients With Advanced Solid Tumors	TARGETS EZH2, TOP1		
LOCATIONS: Washington, Michigan, Ohio, Massachusetts, New Jersey, Texas, Virginia, Georgia			
NCT02896335	PHASE 2		
Palbociclib In Progressive Brain Metastases	TARGETS CDK4, CDK6		
LOCATIONS: Massachusetts			
NCT04555837	PHASE 1/2		
Alisertib and Pembrolizumab for the Treatment of Patients With Rb-deficient Head and Neck quamous Cell Cancer  TARGETS Aurora kinase A, PD-1			
LOCATIONS: Texas			
NCT04241835	PHASE 1		
A Study of Oral Tazemetostat in Subjects With Moderate and Severe Hepatic Impairment With	TARGETS		

Cedex (France), Nevada, Illinois



ORDERED TEST # ORD-1194451-01

CLINICAL TRIALS

NCT03065062	PHASE 1			
Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head & Neck and Other Solid Tumors	TARGETS PI3K-alpha, PI3K-gamma, mTORC1, mTORC2, CDK4, CDK6			
LOCATIONS: Massachusetts				
NCT03454035	PHASE 1			
Ulixertinib/Palbociclib in Patients With Advanced Pancreatic and Other Solid Tumors	TARGETS MAPK3, MAPK1, CDK4, CDK6			
LOCATIONS: North Carolina				
NCT02897375	PHASE 1			
Palbociclib With Cisplatin or Carboplatin in Advanced Solid Tumors	TARGETS CDK4, CDK6			
LOCATIONS: Georgia				



PATIENT Chien, Li-Shu TUMOR TYPE
Nasopharynx and paranasal
sinuses undifferentiated
carcinoma

REPORT DATE 05 Oct 2021

ORDERED TEST # ORD-1194451-01

APPENDIX

Variants of Unknown Significance

**NOTE** One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

**ABL1** T984M **AKT1** R174H **MSH6** K1358fs\*2 SETD2 P190L

SPEN G1286W

TUMOR TYPE Nasopharynx and paranasal sinuses undifferentiated carcinoma

**APPENDIX** 

Genes Assayed in FoundationOne®CDx

ORDERED TEST # ORD-1194451-01

FOUNDATION ONE ® CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

#### DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTK	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL	CCND1
CCND2	CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2	DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRFI1	ESR1	EZH2
FAM46C	FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3
FGFR4	FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3
GATA4	GATA6	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3F3A
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDM5A	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3	P2RY8	PALB2
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA
PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET
RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2
SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOCS1
SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU
SYK	TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53
TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1	WT1	XPO1
XRCC2	ZNF217	ZNF703						
	T: FOR THE DETE			ENTS				
ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSPO2	SDC4	SLC34A2	TERC*	TERT**	TMPRSS2

<sup>\*</sup>TERC is an NCRNA

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Loss of Heterozygosity (LOH) score Microsatellite (MS) status Tumor Mutational Burden (TMB)

<sup>\*\*</sup>Promoter region of TERT is interrogated

APPENDIX

About FoundationOne®CDx

ORDERED TEST # ORD-1194451-01

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.

FOUNDATIONONE®CDx

#### **ABOUT FOUNDATIONONE CDX**

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details: www.rochefoundationmedicine.com/f1cdxtech.

#### **INTENDED USE**

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffinembedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

# **TEST PRINCIPLES**

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture–selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g.,gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

# THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

#### **Diagnostic Significance**

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials Ranking of Therapies in Summary Table
Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials
Pediatric trial qualification → Geographical
proximity → Later trial phase.

# NATIONAL COMPREHENSIVE CANCER NETWORK\* (NCCN\*) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2021. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

#### Limitations

- 1. The MSI-H/MSS designation by FMI F1CDx test is based on genome wide analysis of 95 microsatellite loci and not based on the 5 or 7 MSI loci described in current clinical practice guidelines. The threshold for MSI-H/MSS was determined by analytical concordance to comparator assays (IHC and PCR) using uterine, cecum and colorectal cancer FFPE tissue. The clinical validity of the qualitative MSI designation has not been established. For Microsatellite Instability (MSI) results, confirmatory testing using a validated orthogonal method should be considered.
- 2. TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics

ORDERED TEST # ORD-1194451-01

FOUNDATIONONE®CDx

**APPENDIX** 

About FoundationOne®CDx

of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh\_docs/ pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.

- 3. The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding armand chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
- 4. Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
- 5. Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine whether the patient is a candidate for biopsy.
- 6. Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an ERBB2 amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive,

and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of ERBB2 copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in

https://www.mycancergenome.org/content/ disease/breast-cancer/ERBB2/238/ report the frequency of HER2 overexpression as 20% in breast cancer. Based on the F1CDx HER2 CDx concordance study, approximately 10% of HER2 amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

#### **VARIANT ALLELE FREQUENCY**

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*		
Repeatability	5.11 - 10.40		
Reproducibility	5.95 - 12.31		
INDELS	%CV*		
INDELS Repeatability	%CV*		

<sup>\*</sup>Interquartile Range =  $1^{st}$  Quartile to  $3^{rd}$  Quartile

# **VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING**

The variants indicated for consideration of followup germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are ATM, BAP1, BRCA1, BRCA2, BRIP1, CHEK2, FH, FLCN, MLH1, MSH2, MSH6, MUTYH, PALB2, PMS2, POLE,

RAD51C, RAD51D, RET, SDHA, SDHB, SDHC, SDHD, TSC2, and VHL, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

#### **VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS**

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are ASXL1, CBL, DNMT3A, IDH2, JAK2, KMT2D (MLL2), MPL, MYD88, SF3B1, TET2, and U2AF1 and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

#### LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

#### **NO GUARANTEE OF CLINICAL BENEFIT**

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

#### NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

#### TREATMENT DECISIONS ARE **RESPONSIBILITY OF PHYSICIAN**

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating

TUMOR TYPE

**APPENDIX** 

About FoundationOne®CDx

ORDERED TEST # ORD-1194451-01

physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

FOUNDATION ONE ® CDx

#### **SELECT ABBREVIATIONS**

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
muts/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
os	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
ткі	Tyrosine kinase inhibitor

MR Suite Version 5.0.0

The median exon coverage for this sample is 955x



#### ORDERED TEST # ORD-1194451-01

# Nasopharynx and paranasal sinuses undifferentiated carcinoma

**APPENDIX** 

References

- 1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) pmid: 25392179
- 2. Kroemer G, et al. Oncoimmunology (2015) pmid: 26140250
- 3. Lal N, et al. Oncoimmunology (2015) pmid: 25949894
- 4. Le DT, et al. N. Engl. J. Med. (2015) pmid: 26028255
- 5. Ayers et al., 2016; ASCO-SITC Abstract P60
- 6. Zighelboim I, et al. J. Clin. Oncol. (2007) pmid: 17513808
- 7. Hampel H, et al. Cancer Res. (2006) pmid: 16885385
- 8. Stelloo E, et al. Clin. Cancer Res. (2016) pmid:
- 9. Kanopienė D, et al. Medicina (Kaunas) (2014) pmid: 25458958
- 10. Black D, et al. J. Clin. Oncol. (2006) pmid: 16549821
- 11. Nout RA, et al. Gynecol. Oncol. (2012) pmid: 22609107
- 12. Steinbakk A, et al. Cell Oncol (Dordr) (2011) pmid: 21547578
- 13. Bilbao C, et al. Int. J. Radiat. Oncol. Biol. Phys. (2010) pmid: 20005452
- 14. Guastadisegni C, et al. Eur. J. Cancer (2010) pmid: 20627535
- 15. Pawlik TM, et al. Dis. Markers (2004) pmid: 15528785
- 16. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) pmid: 26337942
- 17. Nature (2012) pmid: 22810696
- 18. Hiyama T, et al. J. Gastroenterol. Hepatol. (2004) pmid: 15209621
- 19. Wu MS, et al. Cancer Res. (1998) pmid: 9537253
- 20. dos Santos NR, et al. Gastroenterology (1996) pmid: 8536886
- 21. Fang WL, et al. Biomed Res Int (2013) pmid: 23555086
- 22. Farris AB, et al. Am. J. Surg. Pathol. (2011) pmid: 21422910
- 23. Agaram NP, et al. Am. J. Clin. Pathol. (2010) pmid: 20395525
- Ruemmele P, et al. Am. J. Surg. Pathol. (2009) pmid: 19252434
- 25. Planck M, et al. Cancer (2003) pmid: 12627520
- 26. Hibi K, et al. Jpn. J. Cancer Res. (1995) pmid: 7775257
- 27. Muneyuki T, et al. Dig. Dis. Sci. (2000) pmid: 11117578 28. Zhang SH, et al. World J. Gastroenterol. (2005) pmid:
- 29. Chiappini F, et al. Carcinogenesis (2004) pmid:
- 14656944
- **30.** Suto T, et al. J Surg Oncol (2001) pmid: 11223838
- 31. Momoi H, et al. J. Hepatol. (2001) pmid: 11580146
- 32. Liengswangwong U, et al. Int. J. Cancer (2003) pmid: 14506736
- 33. Moy AP, et al. Virchows Arch. (2015) pmid: 25680569
- 34. Yoshida T, et al. J. Gastroenterol. (2000) pmid:
- 35. Pritchard CC, et al. Nat Commun (2014) pmid: 25255306
- **36.** Azzouzi AR, et al. BJU Int. (2007) pmid: 17233803
- 37. Burger M, et al. J. Mol. Med. (2006) pmid: 16924473
- 38. Bai S, et al. Am. J. Clin. Pathol. (2013) pmid: 23690119 39. Giedl J, et al. Am. J. Clin. Pathol. (2014) pmid: 25319978
- 40. Yamamoto Y, et al. Clin. Cancer Res. (2006) pmid: 16675567
- 41. Chen FM, et al. Sci Rep (2020) pmid: 32546739
- 42. You JF, et al. Br. J. Cancer (2010) pmid: 21081928
- 43. Bairwa NK, et al. Methods Mol. Biol. (2014) pmid: 24623249
- 44. Boland CR, et al. Cancer Res. (1998) pmid: 9823339
- 45. Boland CR, et al. Gastroenterology (2010) pmid: 20420947

- 46. Samstein RM, et al. Nat. Genet. (2019) pmid: 30643254
- 47. Goodman AM, et al. Mol. Cancer Ther. (2017) pmid: 28835386
- 48. Goodman AM, et al. Cancer Immunol Res (2019) pmid: 31405947
- 49. Cristescu R, et al. Science (2018) pmid: 30309915
- 50. Ready N, et al. J. Clin. Oncol. (2019) pmid: 30785829
- 51. Hellmann MD, et al. N. Engl. J. Med. (2018) pmid: 29658845
- 52. Hellmann MD, et al. Cancer Cell (2018) pmid: 29657128
- 53. Hellmann MD, et al. Cancer Cell (2018) pmid: 29731394
- 54. Rozeman EA, et al. Nat Med (2021) pmid: 33558721
- 55. Sharma P, et al. Cancer Cell (2020) pmid: 32916128
- 56. Marabelle A, et al. Lancet Oncol. (2020) pmid: 32919526
- 57. Legrand et al., 2018; ASCO Abstract 12000
- 58. Chalmers ZR, et al. Genome Med (2017) pmid: 28420421
- 59. Pfeifer GP, et al. Mutat. Res. (2005) pmid: 15748635
- 60. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) pmid: 23875803
- 61. Pfeifer GP, et al. Oncogene (2002) pmid: 12379884
- 62. Rizvi NA, et al. Science (2015) pmid: 25765070
- 63. Johnson BE, et al. Science (2014) pmid: 24336570
- 64. Choi S, et al. Neuro-oncology (2018) pmid: 29452419
- 65. Cancer Genome Atlas Research Network, et al. Nature (2013) pmid: 23636398
- 66. Briggs S. et al. J. Pathol. (2013) pmid: 23447401
- 67. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) pmid: 24583393
- 68. Roberts SA, et al. Nat. Rev. Cancer (2014) pmid: 25568919
- 69. Gounder M, et al. Lancet Oncol (2020) pmid: 33035459
- 70. Italiano A, et al. Lancet Oncol. (2018) pmid: 29650362
- 71. Alimova I, et al. Neuro-oncology (2013) pmid: 23190500
- 72. Knutson SK, et al. Proc. Natl. Acad. Sci. U.S.A. (2013) pmid: 23620515
- 73. Alarcon-Vargas D, et al. Oncogene (2006) pmid: 16302003
- 74. Smith ME, et al. Proc. Natl. Acad. Sci. U.S.A. (2011) pmid: 21173237
- Geoerger B, et al. Clin. Cancer Res. (2017) pmid: 28432176
- 76. Hashizume R, et al. Neuro-oncology (2016) pmid: 27370397
- 77. Lee S, et al. Cancer Res. (2011) pmid: 21521802
- 78. Forrest SJ, et al. Clin Cancer Res (2020) pmid: 32122923
- 79. Leruste A, et al. Cancer Cell (2019) pmid: 31708437
- 80. Chun HE, et al. Cell Rep (2019) pmid: 31708418
- 81. Panwalkar P, et al. Neuro Oncol (2020) pmid: 31912158
- 82. Wetmore C, et al. Neuro-oncology (2015) pmid: 25688119
- 83. Pecora A, et al. J Immunother () pmid: 32815894
- 84. Sodji Q, et al. J Immunother Cancer (2017) pmid: 28807004
- 85. Beckermann KE, et al. J Immunother Cancer (2017) pmid: 28105368
- 86. Bishop JA, et al. Am. J. Surg. Pathol. (2014) pmid: 25007146
- 87. Lin DC, et al. Nat. Genet. (2014) pmid: 24952746
- Agaimy A, et al. Am. J. Surg. Pathol. (2014) pmid: 24832165
- 89. Kakkar A, et al. Hum. Pathol. (2019) pmid: 30120966
- 90. Wilson BG, et al. Nat. Rev. Cancer (2011) pmid: 21654818
- 91. Versteege I, et al. Oncogene (2002) pmid: 12226744

- 92. Morozov A, et al. Proc. Natl. Acad. Sci. U.S.A. (1998) pmid: 9448295
- 93. Das S, et al. J. Biol. Chem. (2009) pmid: 19398554
- 94. Zhang ZK, et al. Mol. Cell. Biol. (2002) pmid: 12138206
- 95. Valencia AM, et al. Cell (2019) pmid: 31759698
- 96. Kondelin J. et al. EMBO Mol Med (2018) pmid: 30108113
- 97. Vries RG, et al. Genes Dev (2005) pmid: 15769941 98. Hollmann TJ, et al. Am. J. Surg. Pathol. (2011) pmid:
- 21934399
- 99. Oda Y. et al. Cancer Sci. (2009) pmid: 19076980
- 100. Hasselblatt M, et al. Genes Chromosomes Cancer (2013) pmid: 23074045
- 101. Rizzo D, et al. Am. J. Surg. Pathol. (2012) pmid: 22614000
- 102. Sullivan LM, et al. Mod. Pathol. (2013) pmid: 23060122
- 103. Arnold MA, et al. Hum. Pathol. (2013) pmid: 23245672
- 104. Abida W, et al. Clin. Cancer Res. (2020) pmid: 32086346
- 105. Mateo J. et al. Lancet Oncol. (2019) pmid: 31806540
- 106. Mateo J, et al. N. Engl. J. Med. (2015) pmid: 26510020
- 107. Swisher EM, et al. Lancet Oncol. (2017) pmid: 27908594
- 108. Gruber et al., 2019; ASCO Abstract 3006
- 109. Liu Y, et al. BMC Cancer (2015) pmid: 25884806
- 110. Pfeifer W, et al. Breast Cancer Res. Treat. (2014) pmid: 25414026
- 111. Kriege M, et al. J. Cancer Res. Clin. Oncol. (2015) pmid:
- 112. Tate JG, et al. Nucleic Acids Res. (2019) pmid: 30371878
- 113. Kilpivaara O, et al. Int. J. Cancer (2005) pmid: 15472904
- 114. Simon M, et al. Neurosurgery (2006) pmid: 17016233
- 115. Dong YS, et al. Tumour Biol. (2014) pmid: 24532427
- 116. Ta HO, et al. Cancer Res. (2015) pmid: 26573794 117. Tomlinson IP, et al. Mutagenesis (2012) pmid: 22294770
- 118. Xu HP, et al. Nucleic Acids Res. (1990) pmid: 2205842
- 119. van der Groep P, et al. Cell Oncol (Dordr) (2011) pmid: 21336636
- 120. Schutte M, et al. Am. J. Hum. Genet. (2003) pmid: 12610780
- 121. Xu X, et al. Mol. Cell. Biol. (2002) pmid: 12024051
- 122. Lee CH, et al. J. Biol. Chem. (2001) pmid: 11390408
- 123. Schwarz JK, et al. Mol. Cancer Res. (2003) pmid: 12805407
- 124. Ng CP, et al. J. Biol. Chem. (2004) pmid: 14681223
- 125. Matsuoka S, et al. Proc. Natl. Acad. Sci. U.S.A. (2000) pmid: 10973490
- 126. Zhou BB, et al. J. Biol. Chem. (2000) pmid: 10744722
- 127. Ahn JY, et al. Cancer Res. (2000) pmid: 11085506
- 128. Melchionna R, et al. Nat. Cell Biol. (2000) pmid: 11025670
- 129. Yoda A, et al. J. Biol. Chem. (2006) pmid: 16798742
- 130. Chehab NH, et al. Genes Dev. (2000) pmid: 10673500
- 131. Lee SB, et al. Cancer Res. (2001) pmid: 11719428 132. Mahdavi M. et al. J Cell Physiol (2019) pmid: 30552672
- 133. Kleiblová P, et al. Klin Onkol (2019) pmid: 31409080
- 134. Hauke J. et al. Cancer Med (2018) pmid: 29522266
- 135. Lu HM, et al. JAMA Oncol (2019) pmid: 30128536
- 136. Stacchiotti et al., 2019: ASCO Abstract 11003 137. Stacchiotti et al., 2018; ESMO Abstract 1611PD
- 138. Chi et al., 2017: AACR-NCI-EORTC Abstract A175
- 139. Kawano S, et al. PLoS ONE (2016) pmid: 27391784
- 140. Jones et al., 2018; ESMO Abstract 1612PD 141. Schoffski et al., 2017; ASCO Abstract 11057
- 142. Penebre et al., 2015; EORTC Abstract C87 143. Zauderer et al., 2018; ASCO Abstract 8515