

**ABOUT THE TEST** FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

**PATIENT**

**DISEASE** Uterus endometrial adenocarcinoma endometrioid  
**NAME** Lin, Hsiu-Chu  
**DATE OF BIRTH** 18 May 1962  
**SEX** Female  
**MEDICAL RECORD #** 46527757

**PHYSICIAN**

**ORDERING PHYSICIAN** Chen, Yi-Jen  
**MEDICAL FACILITY** Taipei Veterans General Hospital  
**ADDITIONAL RECIPIENT** None  
**MEDICAL FACILITY ID** 205872  
**PATHOLOGIST** Not Provided

**SPECIMEN**

**SPECIMEN SITE** Abdominal wall  
**SPECIMEN ID** S110-23268 B (PF21016)  
**SPECIMEN TYPE** Slide Deck  
**DATE OF COLLECTION** 13 August 2021  
**SPECIMEN RECEIVED** 21 September 2021

**Biomarker Findings**

**Microsatellite status** - MS-Stable  
**Tumor Mutational Burden** - 1 Muts/Mb

**Genomic Findings**

For a complete list of the genes assayed, please refer to the Appendix.

**AKT1** L52R  
**PIK3CA** H419\_C420del  
**ARID1A** rearrangement intron 2, Y1369\*  
**CTNNB1** S33Y  
**ASXL1** L542fs\*160

2 Therapies with Clinical Benefit  
 0 Therapies with Resistance

24 Clinical Trials

**BIOMARKER FINDINGS**

**Microsatellite status** - MS-Stable

**Tumor Mutational Burden** - 1 Muts/Mb

**GENOMIC FINDINGS**

**AKT1** - L52R

10 Trials *see p. 9*

**PIK3CA** - H419\_C420del

10 Trials *see p. 14*

**ARID1A** - rearrangement intron 2, Y1369\*

5 Trials *see p. 11*

**CTNNB1** - S33Y

9 Trials *see p. 12*

**THERAPY AND CLINICAL TRIAL IMPLICATIONS**

No therapies or clinical trials. *see Biomarker Findings section*

No therapies or clinical trials. *see Biomarker Findings section*

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)
none	Everolimus <span>2A</span>
	Temsirolimus <span>2A</span>
none	Everolimus <span>2A</span>
	Temsirolimus <span>2A</span>
none	none
none	none

  NCCN category

**VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS (CH)**

Genomic findings below may include nontumor somatic alterations, such as CH. The efficacy of targeting such nontumor somatic alterations is unknown. This content should be interpreted based on clinical context. Refer to appendix for additional information on CH.

**ASXL1** - L542fs\*160 ..... p. 6

**GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS**

*For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.*

**ASXL1 - L542fs\*160** ..... **p. 6**

**NOTE** Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

ORDERED TEST # ORD-1190754-01

## BIOMARKER FINDINGS

## BIOMARKER

## Microsatellite status

## RESULT

MS-Stable

### POTENTIAL TREATMENT STRATEGIES

#### — Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors<sup>1-3</sup>, including approved therapies nivolumab and pembrolizumab<sup>4</sup>. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and experienced a significantly higher ORR compared

with non-MSI-H cases (70% vs. 12%,  $p=0.001$ )<sup>5</sup>.

### FREQUENCY & PROGNOSIS

MSS has been reported in 73-89% of endometrial cancers<sup>6-13</sup>. Data regarding the role of MSI status on prognosis and survival in endometrial cancer are conflicting, with most studies finding no relationship between MSI-H endometrial cancers and survival<sup>8-9,11,14-16</sup>, and one study predicting improved disease-free and disease-specific survival<sup>7</sup>. However, these studies often evaluated endometrial cancers of all FIGO stages together. Studies specifically analyzing early stage endometrial cancer have reported a correlation between MSI-H and decreased survival<sup>8,12,17-18</sup>, thereby suggesting that MSI-H predicts for poor prognosis in this subset of endometrial tumors.

### FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor<sup>19</sup>. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2<sup>19-21</sup>. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers<sup>22-24</sup>. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins<sup>19,21,23-24</sup>.

## BIOMARKER

## Tumor Mutational Burden

## RESULT

1 Muts/Mb

### POTENTIAL TREATMENT STRATEGIES

#### — Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1<sup>25-27</sup>, anti-PD-1 therapies<sup>25-28</sup>, and combination nivolumab and ipilimumab<sup>29-34</sup>. In multiple pan-tumor studies, higher TMB has been reported to be associated with increased ORR and OS from treatment with immune checkpoint inhibitors<sup>25-28,35</sup>. Higher TMB was found to be significantly associated with improved OS upon immune checkpoint inhibitor treatment for patients with 9 types of advanced tumors<sup>25</sup>. Analyses across several solid tumor types reported that patients with higher TMB (defined as  $\geq 16$ -20 Muts/Mb) achieved greater clinical benefit from PD-1 or PD-L1-targeting monotherapy, compared with patients with higher TMB treated with

chemotherapy<sup>36</sup> or those with lower TMB treated with PD-1 or PD-L1-targeting agents<sup>26</sup>. However, the KEYNOTE 158 trial of pembrolizumab monotherapy for patients with solid tumors found significant improvement in ORR for patients with TMB  $\geq 10$  Muts/Mb (based on this assay or others) compared to those with TMB  $< 10$  Muts/Mb, in a large cohort that included multiple tumor types; similar findings were observed in the KEYNOTE 028 and 012 trials<sup>28,35</sup>. Together, these studies suggest that patients with TMB  $\geq 10$  Muts/Mb may derive clinical benefit from PD-1 or PD-L1 inhibitors.

### FREQUENCY & PROGNOSIS

A large-scale genomic analysis found that endometrial adenocarcinomas harbored a median TMB of 4.5 Muts/Mb, and 15% of cases had an elevated TMB of greater than 20 Muts/Mb<sup>37</sup>. Another study evaluating TMB in endometrial adenocarcinoma reported that 24% of tumors had a mutational burden of greater than 10.4 Muts/Mb<sup>38</sup>. Increased tumor mutational burden (TMB) in endometrial carcinoma has been correlated with POLE mutation and advanced high-grade endometrioid subtypes<sup>6,13,39-40</sup>. Ultramutated endometrial tumors (elevated TMB with POLE mutations) have also been associated with improved PFS<sup>6</sup>. The same study associated lower

mutational burden, independent of PD-L1 status, in endometrial carcinomas with poorer prognosis<sup>6</sup>. For patients with advanced microsatellite-stable endometrial carcinoma not treated with immunotherapy, OS did not significantly differ between patients with TMB-high ( $\geq 10$  Muts/Mb) and TMB-low (11.4 vs. 13.5 months, adjusted HR=1.15) in 1 study<sup>41</sup>.

### FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma<sup>42-43</sup> and cigarette smoke in lung cancer<sup>44-45</sup>, treatment with temozolomide-based chemotherapy in glioma<sup>46-47</sup>, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes<sup>6,48-51</sup>, and microsatellite instability (MSI)<sup>6,50-51</sup>. This sample harbors a TMB level associated with lower rates of clinical benefit from treatment with PD-1- or PD-L1-targeting immune checkpoint inhibitors compared with patients with tumors harboring higher TMB levels, based on several studies in multiple solid tumor types<sup>26-27,35</sup>.

ORDERED TEST # ORD-1190754-01

## GENOMIC FINDINGS

## GENE

# AKT1

## ALTERATION

L52R

## TRANSCRIPT ID

NM\_001014431

## CODING SEQUENCE EFFECT

155T&gt;G

## VARIANT ALLELE FREQUENCY (% VAF)

87.2%

## POTENTIAL TREATMENT STRATEGIES

### — Targeted Therapies —

Mutations that activate AKT1 may predict activity of AKT1 inhibitors in various tumor types. Phase 3 trials of AKT inhibitor combination treatments for patients with advanced breast cancer (capivasertib) and prostate cancer (ipatasertib) are furthest in development<sup>52</sup>, and basket trials have also shown responses in a variety of other solid tumors<sup>53-54</sup>. An NCI-MATCH subprotocol of capivasertib for patients with breast cancer (18 patients) and other tumor types (17 patients) reported an ORR of 29% (10/35) with 1 CR experienced by a patient with endometrioid endometrial adenocarcinoma and PRs experienced

by 7 patients with breast cancer, 1 patient with uterine leiomyosarcoma, and 1 patient with oncocytic carcinoma of the parotid gland<sup>54</sup>. On the basis of clinical data in endometrial cancers, AKT1 may predict activity to AKT1 inhibitors such as capivasertib. One patient with endometrial endometrioid adenocarcinoma experienced a CR in a basket trial of capivasertib<sup>54</sup>, and another basket trial of capivasertib for heavily pre-treated patients included 2 PRs in 8 AKT1-mutated endometrial carcinomas (25%)<sup>53</sup>. A Phase 1 study combining the mTORC1/2 inhibitor sapanisertib with metformin reported 1 PR in a patient with endometrial cancer harboring concurrent AKT and mTOR alterations<sup>55</sup>. On the basis of clinical data in solid tumors, AKT1 activating mutations may be sensitive to mTOR inhibitors such as everolimus and temsirolimus<sup>56-60</sup>. In an exploratory analysis, a study for patients with AKT1-mutated hormone-receptor-positive (HR+), HER2-negative breast cancer treated with the investigational ATP-competitive MTOR-inhibitor sapanisertib and exemestane or fulvestrant reported a positive association between best overall response (CR+PR) and AKT1-mutated status (n=11) compared with patients with AKT1-wildtype status (n=42) (p<0.03)<sup>61</sup>. In a retrospective analysis of clinical outcomes for patients with HR+ breast cancer, AKT1 E17K was associated with significantly increased median

duration of treatment with everolimus-containing regimens<sup>62</sup>.

## FREQUENCY & PROGNOSIS

In the scientific literature, AKT1 mutation has been identified in 2-4% of endometrial carcinoma cases, with studies predominately reporting on the incidence of the most common AKT1 mutation in this disease, E17K<sup>56,63-65</sup>. Elevated AKT1 activity has been reported in endometrial cancer tissues, with one study citing AKT1 activation in 53% (19/35) cases<sup>56,66-67</sup>. Published data investigating the prognostic implications of AKT1 alterations in endometrial cancer are limited (PubMed, Dec 2020).

## FINDING SUMMARY

AKT1 encodes an intracellular serine/threonine kinase and is one of three members of the AKT gene family. AKT activation promotes cell survival via inhibition of apoptosis and also contributes to cell proliferation through several interactions with the cell cycle machinery; inappropriate activation of AKT can therefore lead to tumor formation<sup>68</sup>. Missense mutations and in-frame duplications that occur in the pleckstrin homology (PH) domain of AKT1, as seen here, have been shown to transform cells and activate AKT signaling and are therefore considered to be oncogenic<sup>69-75</sup>.

ORDERED TEST # ORD-1190754-01

**GENOMIC FINDINGS**
**GENE**
**PIK3CA**
**ALTERATION**

H419\_C420del

**TRANSCRIPT ID**

NM\_006218

**CODING SEQUENCE EFFECT**

1256\_1261delACTGTC

**VARIANT ALLELE FREQUENCY (% VAF)**

35.3%

the dual PI3K/mTOR kinase inhibitor apitolisib, 79% (11/14) of patients with PIK3CA-mutated advanced solid tumors experienced disease control (3 PRs, 8 SDs)<sup>90</sup>. The PI3K inhibitor alpelisib demonstrated an ORR of 6.0% (8/134) and a DCR of 58% (78/134) in a study of PIK3CA-mutated solid tumors<sup>91</sup>. However, the PI3K inhibitor copanlisib exhibited limited efficacy in PIK3CA-altered tumors<sup>92-93</sup>. It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

that activate the PI3K-AKT-mTOR signaling axis, such as PTEN and KRAS alterations<sup>101-102</sup>. Overexpression of p110- $\alpha$  has been reported in 72% of endometrial carcinomas<sup>103</sup>. One study reported PIK3CA exon 9 or 20 mutations in 20% (20/99) of high-grade endometrial carcinomas; these mutations were associated with shorter patient survival within Grade 3 endometrioid carcinoma, but not within endometrial serous carcinoma<sup>104</sup>.

**POTENTIAL TREATMENT STRATEGIES**
**— Targeted Therapies —**

Clinical and preclinical data in various tumor types indicate that PIK3CA activating alterations may predict sensitivity to therapies targeting PI3K<sup>76-78</sup>, AKT<sup>79-80</sup>, or mTOR<sup>81-88</sup>. Results from the Phase 2 MATCH trial for patients with PIK3CA-altered solid tumors found that 27% (18/65) of patients experienced PFS lasting at least 6 months after treatment with taselisib; however, no ORs were observed in this study<sup>89</sup>. In a Phase 1 trial of

**— Potential Resistance —**

Activating mutations in PIK3CA may confer resistance to HER2-targeted therapies; combined inhibition of HER2 and the PI3K pathway may be required in HER2-positive tumors with PIK3CA mutation<sup>94-98</sup>.

**FREQUENCY & PROGNOSIS**

In the scientific literature, PIK3CA mutations have been reported in 16-54% of endometrial carcinomas<sup>6,99-100</sup>. In endometrial cancers, PIK3CA mutations often co-occur with other mutations

**FINDING SUMMARY**

PIK3CA encodes p110- $\alpha$ , which is the catalytic subunit of phosphatidylinositol 3-kinase (PI3K). The PI3K pathway is involved in cell signaling that regulates a number of critical cellular functions, including cell growth, proliferation, differentiation, motility, and survival<sup>105-106</sup>. Although alterations such as seen here have not been fully characterized, they have been associated with sensitivity to targeted therapies or have shown cancer association, which may indicate biological relevance<sup>107-113</sup>.

**GENE**
**ARID1A**
**ALTERATION**

rearrangement intron 2, Y1369\*

**TRANSCRIPT ID**

NM\_006015

**CODING SEQUENCE EFFECT**

4107C&gt;A

**VARIANT ALLELE FREQUENCY (% VAF)**

43.6%

topotecan<sup>114-115</sup>. On the basis of limited preclinical evidence from studies in ovarian cancer, ARID1A inactivation may predict sensitivity to inhibitors of EZH2<sup>116-117</sup>, which are under investigation in clinical trials. Other studies have reported that loss of ARID1A may activate the PI3K-AKT pathway and be linked with sensitivity to inhibitors of this pathway<sup>118-120</sup>. Loss of ARID1A expression has been associated with chemoresistance to platinum-based therapy in patients with ovarian clear cell carcinoma<sup>121-122</sup> and to 5-fluorouracil (5-FU) in CRC cell lines<sup>123</sup>.

loss is associated with microsatellite instability in ovarian and endometrial endometrioid adenocarcinomas<sup>40,130-133</sup>, CRC<sup>133-136</sup>, and gastric cancer<sup>133,137-141</sup>. Several studies have reported no correlation between ARID1A loss and clinicopathological parameters in ovarian clear cell or endometrioid carcinomas or other endometrial cancers<sup>142-145</sup>, whereas others suggest that ARID1A loss is a negative prognostic factor<sup>122,146</sup>.

**POTENTIAL TREATMENT STRATEGIES**
**— Targeted Therapies —**

There are no therapies approved to address the mutation or loss of ARID1A in cancer. However, on the basis of limited clinical and preclinical evidence, ARID1A inactivating mutations may lead to sensitivity to ATR inhibitors such as M6620; 1 patient with small cell lung cancer harboring an ARID1A mutation experienced a PR when treated with M6620 combined with

**FREQUENCY & PROGNOSIS**

ARID1A alterations are particularly prevalent in ovarian clear cell carcinoma (46-50%), ovarian and uterine endometrioid carcinomas (24-44%), and cholangiocarcinoma (27%); they are also reported in up to 27% of gastric carcinoma, esophageal adenocarcinoma, Waldenstrom macroglobulinemia, pediatric Burkitt lymphoma, hepatocellular carcinoma, colorectal carcinoma, and urothelial carcinoma samples analyzed (COSMIC, cBioPortal, 2021)<sup>107,112-113,124-129</sup>. ARID1A

**FINDING SUMMARY**

ARID1A encodes the AT-rich interactive domain-containing protein 1A, also known as Baf250a, a member of the SWI/SNF chromatin remodeling complex. Mutation, loss, or inactivation of ARID1A has been reported in many cancers, and the gene is considered a tumor suppressor<sup>125,140,147-153</sup>. ARID1A mutations, which are mostly truncating, have been identified along the entire gene and often correlate with ARID1A protein loss<sup>125,138,148-149,154</sup>, whereas ARID1A missense mutations are mostly uncharacterized.

ORDERED TEST # ORD-1190754-01

GENOMIC FINDINGS

GENE

CTNNB1

ALTERATION

S33Y

TRANSCRIPT ID

NM\_001904

CODING SEQUENCE EFFECT

98C>A

VARIANT ALLELE FREQUENCY (% VAF)

42.0%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Mutation or activation of CTNNB1 signaling has been shown to increase mTOR signaling, promote tumorigenesis, and respond to mTOR inhibition in preclinical studies<sup>155-157</sup>. Small studies have reported clinical benefit following treatment of everolimus combined with other targeted agents for patients with CTNNB1-mutated hepatocellular carcinoma<sup>158-159</sup> or endometrial carcinoma<sup>110</sup>. In preclinical studies, CTNNB1 activating mutations have been shown to increase expression of WNT pathway member DKK1, which may promote tumor cell proliferation and immune evasion<sup>160-162</sup>. A Phase 1 trial of DKK1-targeting antibody

DKN-01 in combination with paclitaxel in esophageal cancer reported a PR rate in 2 out of 4 patients and SD rate of 1 out of 4 patients with CTNNB1 activating mutations, compared with 24% (10/41) PR and 37% (15/41) SD in unselected patients<sup>163</sup>. Multiple preclinical studies in cancer models harboring CTNNB1 mutation or beta-catenin pathway activation have reported activation of the NOTCH pathway and sensitivity to pharmacologic inhibition of NOTCH signaling by gamma-secretase inhibitors<sup>164-167</sup>. Phase 1 and 2 clinical trials of gamma-secretase inhibitor PF-03084014 have shown high response rates in patients with desmoid tumors, which are driven by activating CTNNB1 mutations in the majority of cases<sup>168-169</sup>, suggesting CTNNB1-mutated tumors may be sensitive to gamma-secretase inhibitors. Although WNT pathway inhibitors have been explored preclinically in CTNNB1-mutated cells, clinical data supporting this therapeutic approach are lacking<sup>156,170-172</sup>.

FREQUENCY & PROGNOSIS

CTNNB1 mutations have been reported in 7-45% of endometrial carcinomas (ECs)<sup>6,173-176</sup>. CTNNB1 mutations are more common in Type 1 EC than Type 2<sup>177-178</sup>. In addition, one study found that CTNNB1 mutations were identified more frequently in sporadic ECs (31%, 18/58), than in

Lynch syndrome (LS)-associated ECs (6.9%, 2/29)<sup>176</sup>. Nuclear beta-catenin protein expression has been observed in 14.7-27.6% (33/224-55/199) of ECs, with a significantly higher incidence in Type 1 tumors<sup>174,179</sup>. Multiple studies have reported that CTNNB1 mutation characterizes an aggressive subset of EC<sup>17,180-181</sup>. One study found that that TP53 or CTNNB1 mutation was an independent marker of poor recurrence-free survival (HR=4.69) for patients with low grade, early stage EC<sup>180</sup>. Low membrane expression of beta-catenin has been linked with poor prognosis in EC and ovarian endometrioid carcinomas<sup>182-183</sup>. Solid tumors with WNT/beta-catenin pathway alterations, as seen here, were observed to have significantly less T-cell inflammation in one study<sup>184</sup>.

FINDING SUMMARY

CTNNB1 encodes beta-catenin, a key downstream component of the WNT signaling pathway. Beta-catenin interacts with cadherin to regulate cell-cell adhesion; as a component of the WNT pathway, it also plays a role in development, cell proliferation, and cell differentiation<sup>185</sup>. CTNNB1 exon 3 mutations, such as observed here, lead to increased beta-catenin protein stability and activation of the WNT pathway, and are considered to be activating<sup>186-204</sup>.

GENE

ASXL1

ALTERATION

L542fs\*160

TRANSCRIPT ID

NM\_015338

CODING SEQUENCE EFFECT

1624\_1627delCTTG

VARIANT ALLELE FREQUENCY (% VAF)

38.0%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no targeted therapies available to address genomic alterations in ASXL1.

FREQUENCY & PROGNOSIS

ASXL1 alterations occur infrequently across various solid tumor types<sup>205</sup> and are not known to act as drivers in any specific solid cancer type<sup>206</sup>. Published data investigating the prognostic implications of ASXL1 alterations in solid tumors are limited (PubMed, May 2021). In the context of clonal hematopoiesis, ASXL1 mutations are significantly enriched in current or former smokers<sup>207</sup>.

FINDING SUMMARY

ASXL1 regulates epigenetic marks and transcription through interaction with polycomb complex proteins and various transcription activators and repressors<sup>208-210</sup>. Alterations such as seen here may disrupt ASXL1 function or expression<sup>211-213</sup>.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion<sup>214-219</sup>. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy<sup>214-215</sup>. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease<sup>220</sup>. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH<sup>218,221-222</sup>. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.



ORDERED TEST # ORD-1190754-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

## Everolimus

*Assay findings association*
**AKT1**  
L52R

**PIK3CA**  
H419\_C420del

### AREAS OF THERAPEUTIC USE

Everolimus is an orally available mTOR inhibitor that is FDA approved to treat renal cell carcinoma (RCC) following antiangiogenic therapy; pancreatic neuroendocrine tumors; and well-differentiated non-functional neuroendocrine tumors of the lung or gastrointestinal tract. Everolimus is also approved to treat either renal angiomyolipoma or subependymal giant cell astrocytoma in association with tuberous sclerosis complex (TSC). Please see the drug label for full prescribing information.

### GENE ASSOCIATION

AKT1 activating mutations may predict sensitivity to mTOR inhibitors. A clinicogenomic registry study showed significantly increased median duration of treatment with everolimus-containing regimens for patients with AKT1 E17K-mutated breast cancer compared with AKT1-wildtype disease<sup>62</sup>. Individual patients with AKT1-mutated endometrial cancer<sup>56-57</sup>, papillary thyroid cancer<sup>58</sup>, ovarian cancer<sup>59</sup>, or thymoma<sup>60</sup> have achieved objective response or disease control with mTOR inhibitor treatment. On the basis of clinical evidence<sup>81-88</sup>, PIK3CA activating mutations may predict sensitivity to mTOR inhibitors such as everolimus and temsirolimus. Studies have reported modest activity of these therapies as single agents (ORRs of 0-4%), but improved activity has been observed when they are combined with other agents such as bevacizumab and doxorubicin (ORRs of 25-44%), for patients with PIK3CA-mutated solid tumors<sup>85-88,223-227</sup>.

### SUPPORTING DATA

Clinical benefit has been reported for several patients with PIK3CA-mutated endometrial cancer treated with

everolimus as a single agent<sup>56,84,228</sup> or combined with hormone therapy<sup>110,228</sup>. In a Phase 2 clinical trial of recurrent endometrial cancer, 43% (12/28) of patients reported SD at 8 weeks and 21% (6/28) of patients achieved clinical benefit at 20 weeks upon administration of everolimus monotherapy<sup>229</sup>. Combination with the aromatase inhibitor letrozole for the same disease population achieved an ORR of 31% (11/35), with 9 CRs<sup>110</sup>. Further addition of metformin to this regimen led to a clinical benefit rate (CR+PR+SD) of 67% (32/48), including PR in 29% (14/48) of cases; no significant difference was observed between cases with and without KRAS mutation<sup>230</sup>. Everolimus achieved PR or SD in 35% of patients with recurrent endometrial carcinoma; KRAS mutation was associated with reduced median PFS (3.1 vs. 1.0 months) and median OS (9.3 vs. 2.3 months)<sup>56</sup>. Another study investigating estrogen and/or progesterone receptor-positive gynecologic or breast malignancies featuring mutation or loss of genes in the PI3K-AKT-mTOR pathway, including PIK3CA, AKT1, or PTEN, observed SD in 17% (1/6) of patients with endometrial cancer following combined treatment with everolimus and anastrozole<sup>231</sup>. No response was seen in a patient with endometrial stromal sarcoma and Peutz-Jeghers Syndrome associated with a germline STK11 mutation treated with a combination of everolimus and anastrozole<sup>232</sup>. Whereas frequent adverse events precluded a recommended Phase 2 dose and schedule for the combination of trametinib and everolimus in a Phase 1b trial for solid tumors<sup>233</sup>, a retrospective study for heavily pretreated patients with solid tumors reported tolerable regimens of the combination for 23/31 patients, with 16 patients treated >3 months and evaluable patients achieving a median PFS of 6.5 months<sup>234</sup>.

ORDERED TEST # ORD-1190754-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

## Temsirolimus

*Assay findings association*
**AKT1**  
L52R

**PIK3CA**  
H419\_C420del

### AREAS OF THERAPEUTIC USE

Temsirolimus is an intravenous mTOR inhibitor that is FDA approved for the treatment of advanced renal cell carcinoma. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

AKT1 activating mutations may predict sensitivity to mTOR inhibitors. A clinicogenomic registry study showed significantly increased median duration of treatment with everolimus-containing regimens for patients with AKT1 E17K-mutated breast cancer compared with AKT1-wildtype disease<sup>62</sup>. Individual patients with AKT1-mutated endometrial cancer<sup>56-57</sup>, papillary thyroid cancer<sup>58</sup>, ovarian cancer<sup>59</sup>, or thymoma<sup>60</sup> have achieved objective response or disease control with mTOR inhibitor treatment. On the basis of clinical evidence<sup>81-88</sup>, PIK3CA activating mutations may predict sensitivity to mTOR inhibitors such as everolimus and temsirolimus. Studies have reported modest activity of these therapies as single agents (ORRs of 0-4%), but improved activity has been observed when they are combined with other agents such as bevacizumab and doxorubicin (ORRs of 25-44%), for patients with PIK3CA-mutated solid tumors<sup>85-88,223-227</sup>.

### SUPPORTING DATA

In a pooled analysis, 14% (3/21) of patients with PIK3CA-mutated endometrial cancer treated with temsirolimus or ridaforolimus achieved objective response and 29% (6/21) experienced disease progression<sup>235</sup>. A case report described a patient with heavily pretreated PIK3CA-mutated endometrial cancer who had SD for 17 months with temsirolimus alone followed by combination with letrozole<sup>236</sup>. A Phase 2 clinical trial of temsirolimus in recurrent or metastatic endometrial cancer reported PR in 4/29 (14%) chemotherapy-naïve patients and 4% (1/25) of chemotherapy-treated patients, with SD reported in 69% (20/29) of chemotherapy-naïve patients and 48% (12/25) of chemotherapy-treated patients; however, response in this study was found to be independent of molecular markers of PI3K-AKT-mTOR pathway activation<sup>237</sup>. Another Phase 2 study of temsirolimus in patients with endometrial cancer reported PFS of >15 months in 6 patients and associated clinical benefit and longer PFS with mutation of AKT1 or CTNNB1, respectively<sup>238</sup>. Temsirolimus combined with carboplatin and paclitaxel achieved objective partial responses in 82% (9/11) of patients with endometrial cancer<sup>239</sup>. A Phase 2 trial of temsirolimus in combination with bevacizumab in patients with endometrial carcinoma reported clinical response in 25% of patients<sup>240</sup>.

**NOTE** Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.



ORDERED TEST # ORD-1190754-01

**CLINICAL TRIALS**

**NOTE** Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://www.foundationmedicine.com/genomic-testing#support-services). Or visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

**GENE**  
**AKT1**
**ALTERATION**  
**L52R**
**RATIONALE**  
 AKT1 amplification or mutation may lead to activation of AKT signaling and therefore may result in sensitivity to AKT pathway inhibitors.

Inhibitors of AKT and the downstream protein mTOR are under investigation in clinical trials.

**NCT04589845**
**PHASE 2**

Tumor-Agnostic Precision Immuno-Oncology and Somatic Targeting Rational for You (TAPISTRY) Platform Study

**TARGETS**  
 ALK, ROS1, TRKA, TRKB, TRKC, RET, PD-L1, AKTs, ERBB2, MDM2, PI3K-alpha

**LOCATIONS:** Zhongzheng Dist. (Taiwan), Taipei City (Taiwan), Tainan (Taiwan), Seoul (Korea, Republic of), Beijing (China), Woolloongabba (Australia), Darlinghurst (Australia), Randwick (Australia), Melbourne (Australia), Haifa (Israel)

**NCT03239015**
**PHASE 2**

Efficacy and Safety of Targeted Precision Therapy in Refractory Tumor With Druggable Molecular Event

**TARGETS**  
 EGFR, ERBB2, ERBB4, PARP, mTOR, MET, RET, ROS1, VEGFRs, BRAF, CDK4, CDK6

**LOCATIONS:** Shanghai (China)

**NCT04337463**
**PHASE NULL**

ATG-008 Combined With Toripalimab in Advanced Solid Tumors

**TARGETS**  
 mTORC1, mTORC2, PD-1

**LOCATIONS:** Chongqing (China), Chengdu (China)

**NCT04803318**
**PHASE 2**

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

**TARGETS**  
 mTOR, FGFRs, KIT, PDGFRA, RET, VEGFRs, MEK

**LOCATIONS:** Guangzhou (China)

**NCT03772561**
**PHASE 1**

Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies

**TARGETS**  
 PARP, AKTs, PD-L1

**LOCATIONS:** Singapore (Singapore)

© 2021 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Tyler Janovitz, MD, PhD | 29 September 2021  
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Shakti Ramkissoon, M.D., Ph.D., M.M. Sc, Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531  
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531  
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1190754-01

**CLINICAL TRIALS**
**NCT04632992**
**PHASE 2**

A Study Evaluating Targeted Therapies in Participants Who Have Advanced Solid Tumors With Genomic Alterations or Protein Expression Patterns Predictive of Response

**TARGETS**

ALK, ROS1, TRKA, TRKB, TRKC, PD-L1, ERBB2, ERBB3, PI3K-alpha, RET, AKTs

**LOCATIONS:** Alaska, Washington, Oregon, California, Montana

**NCT03994796**
**PHASE 2**

Genetic Testing in Guiding Treatment for Patients With Brain Metastases

**TARGETS**

ALK, ROS1, TRKA, TRKB, TRKC, CDK4, CDK6, PI3K, mTOR

**LOCATIONS:** Alaska, Washington

**NCT02693535**
**PHASE 2**

TAPUR: Testing the Use of Food and Drug Administration (FDA) Approved Drugs That Target a Specific Abnormality in a Tumor Gene in People With Advanced Stage Cancer

**TARGETS**

VEGFRs, ABL, SRC, ALK, AXL, MET, ROS1, TRKA, TRKC, CDK4, CDK6, CSF1R, FLT3, KIT, RET, mTOR, EGFR, ERBB2, ERBB3, MEK, BRAF, SMO, DDR2, PARP, PD-1, CTLA-4, ERBB4

**LOCATIONS:** Hawaii, Washington, Oregon, California

**NCT03297606**
**PHASE 2**

Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)

**TARGETS**

VEGFRs, ABL, SRC, ALK, AXL, MET, ROS1, TRKA, TRKC, DDR2, KIT, EGFR, PD-1, CTLA-4, PARP, CDK4, CDK6, CSF1R, FLT3, RET, mTOR, ERBB2, ERBB3, MEK, BRAF, SMO

**LOCATIONS:** Vancouver (Canada), Edmonton (Canada), Saskatoon (Canada), Regina (Canada), Ottawa (Canada), Montreal (Canada), Toronto (Canada), Kingston (Canada), London (Canada)

**NCT03673787**
**PHASE 1/2**

A Trial of Ipatasertib in Combination With Atezolizumab

**TARGETS**

AKTs, PD-L1

**LOCATIONS:** Sutton (United Kingdom)

ORDERED TEST # ORD-1190754-01

**CLINICAL TRIALS**
**GENE**
**ARID1A**
**RATIONALE**

ARID1A loss or inactivation may predict

sensitivity to ATR inhibitors.

**ALTERATION**

rearrangement intron 2, Y1369\*

**NCT02264678**
**PHASE 1/2**

Ascending Doses of AZD6738 in Combination With Chemotherapy and/or Novel Anti Cancer Agents

**TARGETS**

ATR, PARP, PD-L1

**LOCATIONS:** Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Cambridge (United Kingdom), Withington (United Kingdom), London (United Kingdom), Sutton (United Kingdom), Villejuif (France), Saint Herblain (France), California

**NCT02630199**
**PHASE 1**

Study of AZD6738, DNA Damage Repair/Novel Anti-cancer Agent, in Combination With Paclitaxel, in Refractory Cancer

**TARGETS**

ATR

**LOCATIONS:** Seoul (Korea, Republic of)

**NCT02595931**
**PHASE 1**

ATR Kinase Inhibitor VX-970 and Irinotecan Hydrochloride in Treating Patients With Solid Tumors That Are Metastatic or Cannot Be Removed by Surgery

**TARGETS**

ATR

**LOCATIONS:** California, Missouri, Pennsylvania, Massachusetts, Connecticut, Tennessee, Florida

**NCT03641547**
**PHASE 1**

M6620 Plus Standard Treatment in Oesophageal and Other Cancer

**TARGETS**

ATR

**LOCATIONS:** Glasgow (United Kingdom), Manchester (United Kingdom), Oxford (United Kingdom), Cardiff (United Kingdom)

**NCT03669601**
**PHASE 1**

AZD6738 &amp; Gemcitabine as Combination Therapy

**TARGETS**

ATR

**LOCATIONS:** Cambridge (United Kingdom)

ORDERED TEST # ORD-1190754-01

CLINICAL TRIALS

 GENE  
**CTNNB1**

 RATIONALE  
Based on clinical and preclinical evidence, tumors sensitive to mTOR inhibitors.  
with activating CTNNB1 alterations may be

 ALTERATION  
S33Y

**NCT04337463**

PHASE NULL

ATG-008 Combined With Toripalimab in Advanced Solid Tumors

 TARGETS  
mTORC1, mTORC2, PD-1

LOCATIONS: Chongqing (China), Chengdu (China)

**NCT04803318**

PHASE 2

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

 TARGETS  
mTOR, FGFRs, KIT, PDGFRA, RET, VEGFRs, MEK

LOCATIONS: Guangzhou (China)

**NCT03008408**

PHASE 2

Phase II Ribociclib, Everolimus and Letrozole in Endometrial Cancer

 TARGETS  
Aromatase, mTOR, CDK4, CDK6

LOCATIONS: Texas

**NCT03065062**

PHASE 1

Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head &amp; Neck and Other Solid Tumors

 TARGETS  
PI3K-alpha, PI3K-gamma, mTORC1, mTORC2, CDK4, CDK6

LOCATIONS: Massachusetts

**NCT01582191**

PHASE 1

A Phase 1 Trial of Vandetanib (a Multi-kinase Inhibitor of EGFR, VEGFR and RET Inhibitor) in Combination With Everolimus (an mTOR Inhibitor) in Advanced Cancer

 TARGETS  
mTOR, EGFR, RET, SRC, VEGFRs

LOCATIONS: Texas

**NCT02159989**

PHASE 1

Sapanisertib and Ziv-Aflibercept in Treating Patients With Recurrent Solid Tumors That Are Metastatic or Cannot Be Removed by Surgery

 TARGETS  
PIGF, VEGFA, VEGFB, mTORC1, mTORC2

LOCATIONS: Texas

ORDERED TEST # ORD-1190754-01

CLINICAL TRIALS

**NCT02321501**
**PHASE 1**

Phase I/Ib Dose Escalation &amp; Biomarker Study of Ceritinib (LDK378) + Everolimus for Locally Advanced or Metastatic Solid Tumors With an Expansion in Non-Small Cell Lung Cancer (NSCLC) Characterized by Abnormalities in Anaplastic Lymphoma Kinase (ALK) Expression

**TARGETS**  
 ROS1, ALK, mTOR

**LOCATIONS:** Texas

**NCT03017833**
**PHASE 1**

Sapanisertib and Metformin in Treating Patients With Advanced or Metastatic Relapsed or Refractory Cancers

**TARGETS**  
 mTORC1, mTORC2

**LOCATIONS:** Texas

**NCT03430882**
**PHASE 1**

Sapanisertib, Carboplatin, and Paclitaxel in Treating Patients With Recurrent or Refractory Malignant Solid Tumors

**TARGETS**  
 mTORC1, mTORC2

**LOCATIONS:** Texas

ORDERED TEST # ORD-1190754-01

**CLINICAL TRIALS**
**GENE**  
**PIK3CA**
**ALTERATION**  
 H419\_C420del

**RATIONALE**  
 PIK3CA activating mutations may lead to activation of the PI3K-AKT-mTOR pathway and may therefore indicate sensitivity to inhibitors of this pathway. Strong clinical data support sensitivity of PIK3CA-mutated solid tumors to the

PI3K-alpha inhibitor alpelisib. It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

**NCT04589845**
**PHASE 2**

Tumor-Agnostic Precision Immuno-Oncology and Somatic Targeting Rational for You (TAPISTRY) Platform Study

**TARGETS**  
 ALK, ROS1, TRKA, TRKB, TRKC, RET, PD-L1, AKTs, ERBB2, MDM2, PI3K-alpha

**LOCATIONS:** Zhongzheng Dist. (Taiwan), Taipei City (Taiwan), Tainan (Taiwan), Seoul (Korea, Republic of), Beijing (China), Woolloongabba (Australia), Darlinghurst (Australia), Randwick (Australia), Melbourne (Australia), Haifa (Israel)

**NCT03239015**
**PHASE 2**

Efficacy and Safety of Targeted Precision Therapy in Refractory Tumor With Druggable Molecular Event

**TARGETS**  
 EGFR, ERBB2, ERBB4, PARP, mTOR, MET, RET, ROS1, VEGFRs, BRAF, CDK4, CDK6

**LOCATIONS:** Shanghai (China)

**NCT04337463**
**PHASE NULL**

ATG-008 Combined With Toripalimab in Advanced Solid Tumors

**TARGETS**  
 mTORC1, mTORC2, PD-1

**LOCATIONS:** Chongqing (China), Chengdu (China)

**NCT04803318**
**PHASE 2**

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

**TARGETS**  
 mTOR, FGFRs, KIT, PDGFRA, RET, VEGFRs, MEK

**LOCATIONS:** Guangzhou (China)

**NCT03772561**
**PHASE 1**

Phase I Study of AZD5363 + Olaparib + Durvalumab in Patients With Advanced or Metastatic Solid Tumor Malignancies

**TARGETS**  
 PARP, AKTs, PD-L1

**LOCATIONS:** Singapore (Singapore)



ORDERED TEST # ORD-1190754-01

**CLINICAL TRIALS**
**NCT04801966**
**PHASE NULL**

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

**TARGETS**  
CDK4, CDK6, PI3K-alpha, PD-L1, MEK,  
PARP, PD-1, BRAF

**LOCATIONS:** Melbourne (Australia)

**NCT04632992**
**PHASE 2**

A Study Evaluating Targeted Therapies in Participants Who Have Advanced Solid Tumors With Genomic Alterations or Protein Expression Patterns Predictive of Response

**TARGETS**  
ALK, ROS1, TRKA, TRKB, TRKC, PD-L1,  
ERBB2, ERBB3, PI3K-alpha, RET, AKTs

**LOCATIONS:** Alaska, Washington, Oregon, California, Montana

**NCT03994796**
**PHASE 2**

Genetic Testing in Guiding Treatment for Patients With Brain Metastases

**TARGETS**  
ALK, ROS1, TRKA, TRKB, TRKC, CDK4,  
CDK6, PI3K, mTOR

**LOCATIONS:** Alaska, Washington

**NCT03297606**
**PHASE 2**

Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)

**TARGETS**  
VEGFRs, ABL, SRC, ALK, AXL, MET,  
ROS1, TRKA, TRKC, DDR2, KIT, EGFR,  
PD-1, CTLA-4, PARP, CDK4, CDK6,  
CSF1R, FLT3, RET, mTOR, ERBB2,  
ERBB3, MEK, BRAF, SMO

**LOCATIONS:** Vancouver (Canada), Edmonton (Canada), Saskatoon (Canada), Regina (Canada), Ottawa (Canada), Montreal (Canada), Toronto (Canada), Kingston (Canada), London (Canada)

**NCT03006172**
**PHASE 1**

To Evaluate the Safety, Tolerability, and Pharmacokinetics of GDC-0077 Single Agent in Participants With Solid Tumors and in Combination With Endocrine and Targeted Therapies in Participants With Breast Cancer

**TARGETS**  
PI3K-alpha, Aromatase, ER, CDK4,  
CDK6

**LOCATIONS:** London (United Kingdom), Surrey (United Kingdom), Bordeaux (France), Barcelona (Spain), Valencia (Spain), Toronto (Canada), Massachusetts, New York, Tennessee

ORDERED TEST # ORD-1190754-01

**APPENDIX**
**Variants of Unknown Significance**

**NOTE** One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

**CTCF**  
G47V

**ERBB3**  
A1030T

**FANCG**  
H140Q

**RB1**  
L819V

**RICTOR**  
L177F and P670S

**TBX3**  
A562V

**TNFAIP3**  
P180S

ORDERED TEST # ORD-1190754-01

**APPENDIX**
**Genes Assayed in FoundationOne®CDx**

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

**DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS**

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTB	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL	CCND1
CCND2	CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2	DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FAM46C	FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3
FGFR4	FH	FLCN	FLT1	FLT3	FOXO2	FUBP1	GABRA6	GATA3
GATA4	GATA6	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3F3A
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDMSA	KDMS5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKB1A	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3	P2RY8	PALB2
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA
PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET
RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2
SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOC3
SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU
SYK	TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53
TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1	WT1	XPO1
XRCC2	ZNF217	ZNF703						

**DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS**

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSP02	SDC4	SLC34A2	TERC*	TERT**	TPMRS2

\*TERC is an NCRNA

\*\*Promoter region of TERT is interrogated

**ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS**

Loss of Heterozygosity (LOH) score  
Microsatellite (MS) status  
Tumor Mutational Burden (TMB)

ORDERED TEST # ORD-1190754-01

**APPENDIX**

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.


**ABOUT FOUNDATIONONE CDx**

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:  
[www.rochefoundationmedicine.com/f1cdxtech](http://www.rochefoundationmedicine.com/f1cdxtech).

**INTENDED USE**

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

**TEST PRINCIPLES**

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

**THE REPORT**

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

**Diagnostic Significance**

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

**Qualified Alteration Calls (Equivocal and Subclonal)**

An alteration denoted as "amplification – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

**Ranking of Therapies and Clinical Trials**
**Ranking of Therapies in Summary Table**

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

**Ranking of Clinical Trials**

Pediatric trial qualification → Geographical proximity → Later trial phase.

**NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION**

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) ([www.nccn.org](http://www.nccn.org)). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2021. All rights reserved. To view the most recent and complete version of the guidelines, go online to [NCCN.org](http://NCCN.org). NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

**Limitations**

1. The MSI-H/MSS designation by FMI F1CDx test is based on genome wide analysis of 95 microsatellite loci and not based on the 5 or 7 MSI loci described in current clinical practice guidelines. The threshold for MSI-H/MSS was determined by analytical concordance to comparator assays (IHC and PCR) using uterine, cecum and colorectal cancer FFPE tissue. The clinical validity of the qualitative MSI designation has not been established. For Microsatellite Instability (MSI) results, confirmatory testing using a validated orthogonal method should be considered.
2. TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics

© 2021 Foundation Medicine, Inc. All rights reserved.

ORDERED TEST # ORD-1190754-01

APPENDIX

About FoundationOne®CDx

of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation [https://www.accessdata.fda.gov/cdrh\\_docs/pdf17/P170019B.pdf](https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf). The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.

- The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
- Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
- Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine whether the patient is a candidate for biopsy.
- Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive,

and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of HER2 overexpression as 20% in breast cancer. Based on the F1CDx HER2 CDx concordance study, approximately 10% of HER2 amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

### VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

#### Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

\*Interquartile Range = 1<sup>st</sup> Quartile to 3<sup>rd</sup> Quartile

### VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*,

*RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

### VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

### LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

### NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

### NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

### TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating

ORDERED TEST # ORD-1190754-01

APPENDIX

About FoundationOne®CDx

physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

#### SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mutS/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

MR Suite Version 5.0.0

The median exon coverage for this sample is 962x

© 2021 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Tyler Janovitz, MD, PhD | 29 September 2021  
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Shakti Ramkissoon, M.D., Ph.D., M.M. Sc, Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531



ORDERED TEST # ORD-1190754-01

**APPENDIX**
**References**

1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) PMID: 25392179
2. Kroemer G, et al. Oncoimmunology (2015) PMID: 26140250
3. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
4. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
5. Ayers et al., 2016; ASCO-SITC Abstract P60
6. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
7. Black D, et al. J. Clin. Oncol. (2006) PMID: 16549821
8. Mackay HJ, et al. Eur. J. Cancer (2010) PMID: 20304627
9. Kanopienė D, et al. Medicina (Kaunas) (2014) PMID: 25458958
10. Hampel H, et al. Cancer Res. (2006) PMID: 16885385
11. Steinbakk A, et al. Cell Oncol (Dordr) (2011) PMID: 21547578
12. Bilbao C, et al. Int. J. Radiat. Oncol. Biol. Phys. (2010) PMID: 20005452
13. Church DN, et al. Hum. Mol. Genet. (2013) PMID: 23528559
14. Zigelboim I, et al. J. Clin. Oncol. (2007) PMID: 17513808
15. Bilbao-Sieyro C, et al. Oncotarget (2014) PMID: 25026289
16. Arabi H, et al. Gynecol. Oncol. (2009) PMID: 19275958
17. Stelloo E, et al. Clin. Cancer Res. (2016) PMID: 27006490
18. Nout RA, et al. Gynecol. Oncol. (2012) PMID: 22609107
19. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
20. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
21. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
22. Boland CR, et al. Cancer Res. (1998) PMID: 9823339
23. Pawlik TM, et al. Dis. Markers (2004) PMID: 15528785
24. Boland CR, et al. Gastroenterology (2010) PMID: 20420947
25. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
26. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
27. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
28. Cristescu R, et al. Science (2018) PMID: 30309915
29. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
30. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
31. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
32. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
33. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
34. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
35. Marabelle A, et al. Lancet Oncol. (2020) PMID: 32919526
36. Legrand et al., 2018; ASCO Abstract 12000
37. Chalmers ZR, et al. Genome Med (2017) PMID: 28420421
38. Santin et al., 2016; ASCO Abstract 5591
39. Mehnert JM, et al. J. Clin. Invest. (2016) PMID: 27159395
40. Hussein YR, et al. Mod. Pathol. (2015) PMID: 25394778
41. Shao C, et al. JAMA Netw Open (2020) PMID: 33119110
42. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
43. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
44. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
45. Rizvi NA, et al. Science (2015) PMID: 25765070
46. Johnson BE, et al. Science (2014) PMID: 24336570
47. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
48. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
49. Heitz E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
50. Nature (2012) PMID: 22810696
51. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
52. Smyth LM, et al. Clin Cancer Res (2020) PMID: 32312891
53. Hyman DM, et al. J. Clin. Oncol. (2017) PMID: 28489509
54. Kalinsky K, et al. JAMA Oncol (2020) PMID: 33377972
55. Coleman et al., 2021; ASCO Abstract 3017
56. Trédan O, et al. Target Oncol (2013) PMID: 23238879
57. Aghajanian C, et al. Gynecol Oncol (2018) PMID: 29804638
58. Schneider TC, et al. J Clin Endocrinol Metab (2017) PMID: 27870581
59. Bryce AH, et al. Oncotarget (2017) PMID: 28423702
60. Wheler J, et al. Oncotarget (2013) PMID: 23765114
61. Lim B, et al. Clin Cancer Res (2021) PMID: 33820779
62. Smyth LM, et al. Cancer Discov (2020) PMID: 31924700
63. Cohen Y, et al. Gynecol. Oncol. (2010) PMID: 19853286
64. Shoji K, et al. Br. J. Cancer (2009) PMID: 19491896
65. Dutt A, et al. Br. J. Cancer (2009) PMID: 19738612
66. Wahl H, et al. Gynecol. Oncol. (2010) PMID: 19878980
67. Bland AE, et al. Int. J. Gynecol. Cancer (2009) PMID: 19396006
68. Vivanco I, et al. Nat. Rev. Cancer (2002) PMID: 12094235
69. Bessière L, et al. EBioMedicine (2015) PMID: 26137586
70. Auguste A, et al. Hum. Mol. Genet. (2015) PMID: 26362254
71. Chang MT, et al. Cancer Discov (2018) PMID: 29247016
72. Yeh YC, et al. Mod. Pathol. (2019) PMID: 31527710
73. Parikh C, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) PMID: 23134728
74. Calleja V, et al. PLoS Biol. (2009) PMID: 19166270
75. Askham JM, et al. Oncogene (2010) PMID: 19802009
76. Fritsch C, et al. Mol. Cancer Ther. (2014) PMID: 24608574
77. Juric D, et al. J. Clin. Oncol. (2018) PMID: 29401002
78. Gallant JN, et al. NPJ Precis Oncol (2019) PMID: 30793038
79. André F, et al. N. Engl. J. Med. (2019) PMID: 31091374
80. Smyth LM, et al. NPJ Breast Cancer (2021) PMID: 33863913
81. Park HS, et al. PLoS ONE (2016) PMID: 27105424
82. Lim SM, et al. Oncotarget (2016) PMID: 26859683
83. Hou MM, et al. Oncotarget (2014) PMID: 25426553
84. Varnier R, et al. Eur J Cancer (2019) PMID: 31351267
85. Janku F, et al. Cell Rep (2014) PMID: 24440717
86. Moroney J, et al. Clin. Cancer Res. (2012) PMID: 22927482
87. Basho RK, et al. JAMA Oncol (2017) PMID: 27893038
88. Moroney JW, et al. Clin. Cancer Res. (2011) PMID: 21890452
89. Krop et al., 2018; ASCO Abstract 101
90. Dolly SO, et al. Clin. Cancer Res. (2016) PMID: 26787751
91. Aust Fam Physician (1986) PMID: 2941002
92. Santin AD, et al. Gynecol Oncol Rep (2020) PMID: 31934607
93. Patnaik A, et al. Ann. Oncol. (2016) PMID: 27672108
94. Esteve FJ, et al. Am. J. Pathol. (2010) PMID: 20813970
95. Baselga J, et al. J. Clin. Oncol. (2014) PMID: 25332247
96. Chakrabarty A, et al. Oncogene (2010) PMID: 20581867
97. Kataoka Y, et al. Ann. Oncol. (2010) PMID: 19633047
98. Wang L, et al. BMC Cancer (2011) PMID: 21676217
99. Garcia-Dios DA, et al. Gynecol. Oncol. (2013) PMID: 23219661
100. Rudd ML, et al. Clin. Cancer Res. (2011) PMID: 21266528
101. Oda K, et al. Cancer Res. (2005) PMID: 16322209
102. Oda K, et al. Cancer Res. (2008) PMID: 18829572
103. Akiyama-Abe A, et al. Br. J. Cancer (2013) PMID: 23949151
104. McIntyre JB, et al. Gynecol. Oncol. (2014) PMID: 24262879
105. Samuels Y, et al. Cancer Cell (2005) PMID: 15950905
106. Nat. Rev. Cancer (2009) PMID: 19629070
107. Tate JG, et al. Nucleic Acids Res. (2019) PMID: 30371878
108. Rivière JB, et al. Nat. Genet. (2012) PMID: 22729224
109. Akgumus G, et al. J Mol Diagn (2017) PMID: 28502730
110. Slomovitz BM, et al. J. Clin. Oncol. (2015) PMID: 25624430
111. Al-Rohil RN, et al. Cancer (2016) PMID: 26479420
112. Cerami E, et al. Cancer Discov (2012) PMID: 22588877
113. Gao J, et al. Sci Signal (2013) PMID: 23550210
114. Thomas A, et al. J. Clin. Oncol. (2018) PMID: 29252124
115. Williamson CT, et al. Nat Commun (2016) PMID: 27958275
116. Bitler BG, et al. Nat. Med. (2015) PMID: 25686104
117. Kim KH, et al. Nat. Med. (2015) PMID: 26552009
118. Wiegand KC, et al. BMC Cancer (2014) PMID: 24559118
119. Huang HN, et al. Mod. Pathol. (2014) PMID: 24336158
120. Samartzis EP, et al. Oncotarget (2014) PMID: 24979463
121. Yokoyama Y, et al. J Gynecol Oncol (2014) PMID: 24459582
122. Katagiri A, et al. Mod. Pathol. (2012) PMID: 22101352
123. Xie C, et al. Tumour Biol. (2014) PMID: 24833095
124. Wu RC, et al. Cancer Biol. Ther. (2014) PMID: 24618703
125. Jones S, et al. Hum. Mutat. (2012) PMID: 22009941
126. Dulak AM, et al. Nat. Genet. (2013) PMID: 23525077
127. Streppe MM, et al. Oncogene (2014) PMID: 23318448
128. Jiao Y, et al. J. Pathol. (2014) PMID: 24293293
129. Ross JS, et al. Oncologist (2014) PMID: 24563076
130. Huang HN, et al. Histopathology (2015) PMID: 25195947
131. Bosse T, et al. Mod. Pathol. (2013) PMID: 23702729
132. Allo G, et al. Mod. Pathol. (2014) PMID: 23887303
133. Okamura R, et al. J Immunother Cancer (2020) PMID: 32111729
134. Chou A, et al. Hum. Pathol. (2014) PMID: 24925223
135. Ye J, et al. Hum. Pathol. (2014) PMID: 25311944
136. Wei XL, et al. World J. Gastroenterol. (2014) PMID: 25561809
137. Chen K, et al. Proc. Natl. Acad. Sci. U.S.A. (2015) PMID: 25583476
138. Wang K, et al. Nat. Genet. (2011) PMID: 22037554
139. Abe H, et al. Virchows Arch. (2012) PMID: 22915242
140. Wang DD, et al. PLoS ONE (2012) PMID: 22808142
141. Wiegand KC, et al. Hum. Pathol. (2014) PMID: 24767857
142. Rahman M, et al. Hum. Pathol. (2013) PMID: 22939958
143. Maeda D, et al. Int J Mol Sci (2010) PMID: 21614196
144. Lowery WJ, et al. Int. J. Gynecol. Cancer (2012) PMID: 22193641
145. Fadare O, et al. Mod. Pathol. (2013) PMID: 23524907
146. Mao TL, et al. Am. J. Surg. Pathol. (2013) PMID: 24076775
147. Guan B, et al. Cancer Res. (2011) PMID: 21900401
148. Wiegand KC, et al. N. Engl. J. Med. (2010) PMID: 20942669
149. Jones S, et al. Science (2010) PMID: 20826764
150. Yan HB, et al. Carcinogenesis (2014) PMID: 24293408
151. Huang J, et al. Nat. Genet. (2012) PMID: 22922871
152. Chan-On W, et al. Nat. Genet. (2013) PMID: 24185513
153. Mamo A, et al. Oncogene (2012) PMID: 21892209

© 2021 Foundation Medicine, Inc. All rights reserved.

 Electronically signed by Tyler Janovitz, MD, PhD | 29 September 2021  
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531  
 Shakti Ramkissoon, M.D., Ph.D., M.M. Sc, Laboratory Director CLIA: 34D2044309  
 Foundation Medicine, Inc. | 1.888.988.3639

 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531  
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1190754-01

**APPENDIX**
**References**

154. Zang ZJ, et al. Nat. Genet. (2012) pmid: 22484628
155. Tanwar PS, et al. Biol. Reprod. (2009) pmid: 19403928
156. Tanwar PS, et al. PLoS ONE (2011) pmid: 21695255
157. Fujishita T, et al. Proc. Natl. Acad. Sci. U.S.A. (2008) pmid: 18768809
158. Bhoori S, et al. J. Hepatol. (2010) pmid: 20347502
159. Janku F, et al. Oncotarget (2014) pmid: 24931142
160. Niida A, et al. Oncogene (2004) pmid: 15378020
161. Chamorro MN, et al. EMBO J. (2005) pmid: 15592430
162. Kagey MH, et al. Br. J. Pharmacol. (2017) pmid: 28574171
163. Kagey et al., 2017; AACR Abstract 369
164. Kwon C, et al. Nat. Cell Biol. (2011) pmid: 21841793
165. Arcaroli JJ, et al. Br. J. Cancer (2013) pmid: 23868008
166. Shang H, et al. Cancer (2015) pmid: 26349011
167. Kode A, et al. Nature (2014) pmid: 24429522
168. Kummur et al., 2015; ASCO Abstract 10563
169. Messersmith WA, et al. Clin. Cancer Res. (2015) pmid: 25231399
170. Zhu J, et al. Carcinogenesis (2012) pmid: 22964660
171. Kogan Y, et al. Biochem. J. (2012) pmid: 22356261
172. Lachenmayer A, et al. Clin. Cancer Res. (2012) pmid: 22811581
173. Dellinger TH, et al. Expert Rev Anticancer Ther (2012) pmid: 22149432
174. Saegusa M, et al. Br. J. Cancer (2001) pmid: 11161379
175. McConechy MK, et al. Mod. Pathol. (2014) pmid: 23765252
176. Huang M, et al. Cancer (2013) pmid: 23760948
177. Yeramian A, et al. Oncogene (2013) pmid: 22430211
178. Machin P, et al. Hum. Pathol. (2002) pmid: 11957146
179. Peiró G, et al. Hum. Pathol. (2013) pmid: 22955108
180. Kurnit KC, et al. Mod. Pathol. (2017) pmid: 28281553
181. Liu Y, et al. J. Natl. Cancer Inst. (2014) pmid: 25214561
182. Rosen DG, et al. Mod. Pathol. (2010) pmid: 19820688
183. Athanassiadou P, et al. Int. J. Gynecol. Cancer ( ) pmid: 17504383
184. Luke JJ, et al. Clin Cancer Res (2019) pmid: 30635339
185. Biochem. Biophys. Res. Commun. (2000) pmid: 10679188
186. Anastas JN, et al. Nat. Rev. Cancer (2013) pmid: 23258168
187. Fukuchi T, et al. Cancer Res. (1998) pmid: 9721853
188. Cancer Sci. (2003) pmid: 12824913
189. Takahashi Y, et al. Virchows Arch. (2006) pmid: 16523258
190. Tanaka Y, et al. Cancer Res. (2001) pmid: 11731417
191. Abraham SC, et al. Am. J. Pathol. (2002) pmid: 11943721
192. Austinat M, et al. Mol. Cancer (2008) pmid: 18282277
193. Wu G, et al. Mol. Cell (2003) pmid: 12820959
194. Provost E, et al. Oncogene (2005) pmid: 15829978
195. Curr. Opin. Genet. Dev. (1999) pmid: 10072352
196. Segditsas S, et al. Oncogene (2006) pmid: 17143297
197. Barth AI, et al. J. Cell Biol. (1997) pmid: 9024698
198. Harada N, et al. EMBO J. (1999) pmid: 10545105
199. Hsu SC, et al. Mol. Cell. Biol. (1998) pmid: 9671490
200. Breuhahn K, et al. J. Pathol. (2008) pmid: 18491352
201. Soon PS, et al. Oncologist (2008) pmid: 18515740
202. Tacon LJ, et al. Oncologist (2011) pmid: 21212436
203. Simon DP, et al. Mol. Cell. Endocrinol. (2012) pmid: 22266195
204. Hirotsu Y, et al. Hepatol. Res. (2016) pmid: 26850916
205. Zehir A, et al. Nat. Med. (2017) pmid: 28481359
206. Bailey MH, et al. Cell (2018) pmid: 29625053
207. Bolton KL, et al. Nat Genet (2020) pmid: 33106634
208. Scheuermann JC, et al. Nature (2010) pmid: 20436459
209. Cho YS, et al. J. Biol. Chem. (2006) pmid: 16606617
210. Park UH, et al. J. Biol. Chem. (2011) pmid: 21047783
211. Inoue D, et al. J. Clin. Invest. (2013) pmid: 24216483
212. Abdel-Wahab O, et al. Cancer Cell (2012) pmid: 22897849
213. Br. J. Cancer (2013) pmid: 23736028
214. Jaiswal S, et al. N. Engl. J. Med. (2014) pmid: 25426837
215. Genovese G, et al. N. Engl. J. Med. (2014) pmid: 25426838
216. Xie M, et al. Nat. Med. (2014) pmid: 25326804
217. Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) pmid: 28669404
218. Severson EA, et al. Blood (2018) pmid: 29678827
219. Fuster JJ, et al. Circ. Res. (2018) pmid: 29420212
220. Hematology Am Soc Hematol Educ Program (2018) pmid: 30504320
221. Chabon JJ, et al. Nature (2020) pmid: 32269342
222. Razavi P, et al. Nat. Med. (2019) pmid: 31768066
223. Janku F, et al. Cancer Res. (2013) pmid: 23066039
224. Janku F, et al. J. Clin. Oncol. (2012) pmid: 22271473
225. Janku F, et al. Mol. Cancer Ther. (2011) pmid: 21216929
226. Moulder S, et al. Ann. Oncol. (2015) pmid: 25878190
227. Byeon et al., 2020; doi: 10.21037/tcr.2020.04.07
228. Myers AP, et al. Gynecol. Oncol. (2016) pmid: 27016228
229. Slomovitz BM, et al. Cancer (2010) pmid: 20681032
230. Soliman et al., 2016; ASCO Abstract 5506
231. Wheler JJ, et al. Oncotarget (2014) pmid: 24912489
232. Noriega-Iriondo MF, et al. Hered Cancer Clin Pract (2015) pmid: 25649062
233. Tolcher AW, et al. Ann. Oncol. (2015) pmid: 25344362
234. Patterson et al., 2018; AACR Abstract 3891
235. Mackay HJ, et al. Cancer (2014) pmid: 24166148
236. Dhami J, et al. Cold Spring Harb Mol Case Stud (2018) pmid: 29588307
237. Oza AM, et al. J. Clin. Oncol. (2011) pmid: 21788564
238. Myers et al., 2015; ASCO Annual Meeting Abstract 5592
239. Kollmannsberger C, et al. Ann. Oncol. (2012) pmid: 21447615
240. Alvarez EA, et al. Gynecol. Oncol. (2013) pmid: 23262204