

REPORT SUMMARY

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PATIENT AND SAMPLE INFORMATION

PATIENT

Name: 陳昭綺
Gender: Female
Date of Birth: May 03, 1965
Patient ID: 19749225
Diagnosis: Gastric adenocarcinoma

SPECIMEN

Type: FFPE tissue
Date received: Oct 28, 2021
Collection site: Colon
Specimen ID: S11030322
Lab ID: AA-21-04836
D/ID: NA

ORDERING PHYSICIAN

Name: 陳明晃醫師
Facility: 臺北榮總
Tel: 886-228712121
Address: 臺北市北投區石牌路二段 201 號

VARIANT(S) WITH CLINICAL RELEVANCE

Only variant(s) with clinical significance are listed. See the "DETAILED TEST RESULTS" section for full details.

SINGLE NUCLEOTIDE AND SMALL INDEL VARIANTS

Not detected.

COPY NUMBER VARIANTS (CNVS)

Loss of heterozygosity (LOH) information was used to infer tumor cellularity. Copy number alteration in the tumor was determined based on **31%** tumor purity.

Amplification (Copy number ≥ 8)

| Chr | Gene | Copy Number |
|-----|------|-------------|
| ND | ND | ND |

Homozygous deletion (Copy number=0)

| Chr | Gene |
|-----|------|
| ND | ND |

Heterozygous deletion (Copy number=1)

| Chr | Gene |
|-------|------------|
| chr5 | RAD50 |
| chr11 | ATM |
| chr22 | CHEK2, NF2 |

ND, Not Detected

TUMOR MUTATIONAL BURDEN (TMB)

1.9 muts/Mb

Muts/Mb, mutations per megabase

MICROSATELLITE INSTABILITY (MSI)

Microsatellite stable (MSS)

Note:

TMB was calculated by using the sequenced regions of ACTOnco®+ to estimate the number of somatic nonsynonymous mutations per megabase of all protein-coding genes (whole exome). The threshold for high mutation load is set at ≥ 7.5 mutations per megabase. TMB, microsatellite status and gene copy number deletion cannot be determined if calculated tumor purity is < 30%.

Variant Analysis:

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Sign Off

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THERAPEUTIC IMPLICATIONS

TARGETED THERAPIES

| Genomic Alterations | Therapies | Effect |
|------------------------------------|---|------------------|
| Level 3B | | |
| ATM Heterozygous deletion | Niraparib, Olaparib, Rucaparib, Talazoparib | sensitive |
| CHEK2 Heterozygous deletion | Niraparib, Rucaparib | sensitive |
| Level 4 | | |
| CHEK2 Heterozygous deletion | Olaparib | sensitive |
| RAD50 Heterozygous deletion | Olaparib, Rucaparib | sensitive |
| NF2 Heterozygous deletion | Everolimus | sensitive |

† Refer to "ONGOING CLINICAL TRIALS" section for detailed trial information.

Note: Therapies associated with benefit or lack of benefit are based on biomarkers detected in this tumor and published evidence.

| Level | Description |
|-------|---|
| 1 | FDA-recognized biomarker predictive of response to an FDA approved drug in this indication |
| 2 | Standard care biomarker (recommended as standard care by the NCCN or other expert panels) predictive of response to an FDA approved drug in this indication |
| 3 | A Biomarkers that predict response or resistance to therapies approved by the FDA or professional societies for a different type of tumor |
| | B Biomarkers that serve as inclusion criteria for clinical trials |
| 4 | Biomarkers that show plausible therapeutic significance based on small studies, few case reports or preclinical studies |

IMMUNE CHECKPOINT INHIBITORS (ICI) THERAPIES

Genomic markers and alterations that are associated with response to ICI therapies

| Positive Biomarker | Negative Biomarker |
|----------------------------------|-----------------------------------|
| TMB-H: ND | EGFR aberration: ND |
| MSI-H: ND | MDM2/MDM4 amplification: ND |
| MMR biallelic inactivation: ND | STK11 biallelic inactivation: ND |
| PBRM1 biallelic inactivation: ND | PTEN biallelic inactivation: ND |
| SERPINB3/SERPINB4 mutation: ND | B2M biallelic inactivation: ND |
| | JAK1/2 biallelic inactivation: ND |

MMR, mismatch repair; ND, not detected

Note: Tumor non-genomic factors, such as patient germline genetics, PDL1 expression, tumor microenvironment, epigenetic alterations or other factors not provided by this test may affect ICI response.

CHEMOTHERAPIES

No genomic alterations detected in this tumor predicted to confer sensitivity or lack of benefit to chemotherapies.

HORMONAL THERAPIES

No genomic alterations detected in this tumor predicted to confer sensitivity or lack of benefit to hormonal therapies.

OTHERS

No genomic alterations detected in this tumor predicted to confer sensitivity or lack of benefit to other therapies.

Note:

Therapeutic implications provided in the test are based solely on the panel of 440 genes sequenced. Therefore, alterations in genes not covered in this panel, epigenetic and post-transcriptional and post-translational factors may also determine a patient's response to therapies. In addition, several other patient-associated clinical factors, including but not limited to, prior lines of therapies received, dosage and combinations with other therapeutic agents, patient's cancer types, sub-types, and/or stages, may also determine the patient's clinical response to therapies.

VARIANT INTERPRETATION

ATM Heterozygous deletion

Biological Impact

The ataxia-telangiectasia mutated protein kinase (ATM) gene encodes a PI3K-related serine/threonine protein kinase involved in genomic integrity maintenance and plays central roles in DNA double-strand break (DSB) repair, which can be induced by ionizing radiation, chemotherapy drugs, or oxidative stress^[1]. ATM is a well-characterized tumor suppressor gene, hereditary mutations and haploinsufficiency of ATM result in markedly increased susceptibility to a variety of cancer types^{[2][3][4][5][6]}. Results from a case-cohort study of colorectal cancer and cancer-free control individuals suggested that germline pathogenic mutations in ATM and PALB2 should be added to established CRC risk genes as part of standard tumor genetic testing panels^[7]. ATM is among the most commonly aberrant genes in sporadic cancers. Somatic ATM aberrations are frequently observed in hematologic malignancies^{[8][9][10][11]} and a broad range of tumors such as prostate cancer^[12], head and neck squamous cell carcinoma (HNSCC)^[13], pancreatic cancer^[14], lung adenocarcinoma^[15], breast cancer^[16], and ovarian cancer^[3].

Therapeutic and prognostic relevance

In May 2020, the U.S. FDA approved olaparib for the treatment of adult patients with metastatic castration-resistant prostate cancer (mCRPC) who carry mutations in homologous recombination repair (HRR) genes, including BRCA1, BRCA2, ATM, BARD1, BRIP1, CDK12, CHEK1, CHEK2, FANCL, PALB2, RAD51B, RAD51C, RAD51D, RAD54L, and progressed following prior treatment with enzalutamide or abiraterone acetate^[17].

In addition, ATM has been determined as an inclusion criterion for the trials evaluating rucaparib efficacy in ovarian cancer^[18] or prostate cancer^[19], niraparib efficacy in pancreatic cancer (NCT03553004), prostate cancer (NCT02854436), and any malignancy, except prostate (NCT03207347), and talazoparib efficacy in advanced or metastatic cancer (NCT02286687), HER2-negative breast cancer (NCT02401347), prostate cancer (NCT03148795), and lung cancer (NCT03377556), respectively.

Besides, another randomized, double-blind Phase II trial in patients with metastatic gastric cancer has shown that addition of olaparib to paclitaxel significantly increased the overall survival in both the overall population and patients with low or undetectable ATM protein expression^[20]. Also, a prospective study in muscle-invasive bladder cancer patients suggested that genomic alterations in the DNA repair genes ATMs, RB1 and FANCC could be recognized as biomarkers predictive of response to cisplatin-based neoadjuvant chemotherapy^[21]. However, loss-of-function of the ATM-CHEK2-TP53 cascade is associated with resistance to anthracycline/mitomycin-containing chemotherapy in patients with breast cancer^[22].

A Retrospective study of the VICTOR clinical trial in patients with colorectal cancer showed that loss of expression of ATM is associated with worse prognosis^[23].

CHEK2 Heterozygous deletion

Biological Impact

The checkpoint kinase 2 (CHEK2 or CHK2) gene encodes a serine/threonine protein kinase involved in transducing DNA damage signals that are required for both the intra-S phase and G2/M checkpoints^[24]. CHEK2 heterozygosity has been shown to cause haploinsufficient phenotypes that can contribute to tumorigenesis through inappropriate S phase entry, accumulation of DNA damage during replication, and failure to restrain mitotic entry^{[25][26]}. CHEK2 aberrations are associated with glioblastoma, breast, ovarian, prostate, colorectal, gastric, thyroid, and lung cancers^{[27][28][29][30][31]}.

Therapeutic and prognostic relevance

In May 2020, the U.S. FDA approved olaparib for the treatment of adult patients with metastatic castration-resistant prostate cancer (mCRPC) who carry mutations in homologous recombination repair (HRR) genes, including BRCA1, BRCA2, ATM, BARD1, BRIP1, CDK12, CHEK1, CHEK2, FANCL, PALB2, RAD51B, RAD51C, RAD51D, RAD54L, and progressed following prior treatment with enzalutamide or abiraterone acetate^[17].

In addition, CHEK2 has been determined as an inclusion criterion for the trials evaluating rucaparib efficacy in ovarian cancer^[18] or prostate cancer^[19] (NCT03533946), niraparib efficacy in melanoma (NCT03925350), pancreatic cancer (NCT03553004), prostate cancer (NCT02854436), and any malignancy, except prostate (NCT03207347), and talazoparib efficacy in HER2-negative breast cancer (NCT02401347), prostate cancer (NCT03148795), and lung cancer (NCT03377556), respectively.

In a phase 2 trial, two prostate cancer patients harboring CHEK2 homozygous deletion was enrolled. One of the two patients had a response to olaparib^[32].

NF2 Heterozygous deletion

Biological Impact

The neurofibromin (NF2) gene encodes the protein Merlin, a tumor suppressor that functions as a negative regulator of the PI3K/AKT/mTOR pathway^{[33][34][35]}. NF2 is a haploinsufficient tumor suppressor gene with one copy loss may lead to weak protein expression and is insufficient to execute its original physiological functions^[36]. Inactivation germline mutations in the NF2 are associated with the hereditary neurofibromatosis type 2, a disorder characterized by the growth of noncancerous tumors in the nervous system^{[33][37]}. Somatic mutations or deletion of NF2 are frequently observed in human cancers, including 20-50% of pleural mesotheliomas^[38], 6% papillary renal cell carcinoma, 5% pancreas cancer, and 4% melanoma (cbioPortal; June 2015), and less frequently in other cancers^[39].

Therapeutic and prognostic relevance

Genomic alterations with activating effects on the mTOR signaling pathway have been identified to confer sensitivity to everolimus across multiple cancer types^{[40][41][42][43]}. There are at least two case studies indicating the

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clinical efficacy of everolimus in bladder cancer^[44] and urothelial carcinoma^[45], both harboring NF2 truncating mutations. Preclinical evidence has shown the efficacy of MEK1/2 inhibitor selumetinib in KRAS-mutant thyroid cancer model with NF2 loss^[46].

Analysis of afatinib-plus-cetuximab-resistant biopsy specimens revealed a loss-of-function alteration in genes that modulate mTOR signaling pathway, including NF2 and TSC1^[47].

RAD50 Heterozygous deletion

Biological Impact

The RAD50 gene encodes a highly-conserved DNA double-strand break (DSB) repair factor. It forms MRN complex with NBS1 and MRE11 protein and is involved in sensing and early processing of DSB, cell cycle checkpoints, DNA recombination and maintenance of telomeres^{[48][49]}. Mutations in the components of the MRN complex could increase susceptibility to familial breast cancer^{[50][51]}, gastric cancer^[52], colorectal cancer^[53], and urothelial cancer^[54]. RAD50 has been implicated as a haploinsufficient gene with one copy loss may lead to weak protein expression and is insufficient to execute its original physiological functions^[55]. Besides, RAD50 deletion was also suggested as a marker of BRCAness, a phenotype shared between non-BRCA1/2-mutated ovarian cancers and BRCA1/2-mutated ovarian cancers^[56].

Therapeutic and prognostic relevance

Preclinical data showed that knockdown of the RAD50 gene in ovarian cancer cell lines was significantly associated with better responses to two PARP inhibitors, olaparib and rucaparib^[56].

US FDA-APPROVED DRUG(S)

Everolimus (AFINITOR)

Everolimus, a derivative of sirolimus, works as an inhibitor of mammalian target of rapamycin complex 1 (mTORC1) and blocks mTORC1-mediated downstream signals for cell growth, proliferation, and survival. Everolimus is developed and marketed by Novartis under the trade name AFINITOR.

FDA Approval Summary of Everolimus (AFINITOR)

| | |
|---|---|
| RADIANT-4 ^[57] NCT01524783 | Lung or gastrointestinal neuroendocrine tumor (Approved on 2016/02/26) |
| | - |
| | Everolimus vs. Placebo [PFS(M): 11 vs. 3.9] |
| BOLERO-2 ^[58] NCT00863655 | Breast cancer (Approved on 2012/07/20) |
| | ER+/HER2- |
| | Everolimus + exemestane vs. Placebo + exemestane [PFS(M): 7.8 vs. 3.2] |
| RADIANT-3 ^[59] NCT00510068 | Pancreatic neuroendocrine tumor (Approved on 2011/05/05) |
| | - |
| | Everolimus vs. Placebo [PFS(M): 11 vs. 4.6] |
| EXIST-1 ^[60] NCT00789828 | Subependymal giant cell astrocytoma (Approved on 2010/10/29) |
| | - |
| | Everolimus vs. Placebo [ORR(%): 35.0] |
| RECORD-1 ^[61] NCT00410124 | Renal cell carcinoma (Approved on 2009/05/30) |
| | - |
| | Everolimus vs. Placebo [PFS(M): 4.9 vs. 1.9] |

Niraparib (ZEJULA)

Niraparib is an oral, small molecule inhibitor of the DNA repair enzyme poly (ADP-ribose) polymerase-1 and -2 (PARP-1, -2). Niraparib is developed and marketed by Tesaro under the trade name ZEJULA.

FDA Approval Summary of Niraparib (ZEJULA)

| | |
|---|--|
| QUADRA^[62] NCT02354586 | Ovarian cancer (Approved on 2019/10/23) |
| | HRD-positive (defined by either a deleterious or suspected deleterious BRCA mutation, and/or genomic instability) |
| | Niraparib [ORR(%): 24.0, DOR(M): 8.3] |
| NOVA^[63] NCT01847274 | Ovarian cancer, Fallopian tube cancer, Peritoneal carcinoma (Approved on 2017/03/27) |
| | gBRCA+ CR/PR to platinum-based chemotherapy |
| | Niraparib vs. Placebo [PFS(M): 21 vs. 5.5] |
| NOVA^[63] NCT01847274 | Ovarian cancer, Fallopian tube cancer, Peritoneal carcinoma (Approved on 2017/03/27) |
| | gBRCA- CR/PR to platinum-based chemotherapy |
| | Niraparib vs. Placebo [PFS(M): 9.3 vs. 3.9] |

Olaparib (LYNPARZA)

Olaparib is an oral, small molecule inhibitor of poly (ADP-ribose) polymerase-1, -2, and -3 (PARP-1, -2, -3). Olaparib is developed by KuDOS Pharmaceuticals and marketed by AstraZeneca under the trade name LYNPARZA.

FDA Approval Summary of Olaparib (LYNPARZA)

| | |
|---|--|
| PROfound^[17] NCT02987543 | Prostate cancer (Approved on 2020/05/19) |
| | ATMm, BRCA1m, BRCA2m, BARD1m, BRIP1m, CDK12m, CHEK1m, CHEK2m, FANCLm, PALB2m, RAD51Bm, RAD51Cm, RAD51Dm, RAD54Lm |
| | Olaparib vs. Enzalutamide or abiraterone acetate [PFS(M): 5.8 vs. 3.5] |
| PAOLA-1^[64] NCT02477644 | Ovarian cancer (Approved on 2020/05/08) |
| | HRD-positive (defined by either a deleterious or suspected deleterious BRCA mutation, and/or genomic instability) |
| | Olaparib + bevacizumab vs. Placebo + bevacizumab [PFS(M): 37.2 vs. 17.7] |

| | |
|--|---|
| POLO^[65] NCT02184195 | Pancreatic adenocarcinoma (Approved on 2019/12/27) |
| | Germline BRCA mutation (deleterious/suspected deleterious) Olaparib vs. Placebo [ORR(%): 23.0 vs. 12.0, PFS(M): 7.4 vs. 3.8] |
| SOLO-1^[66] NCT01844986 | Ovarian cancer, Fallopian tube cancer, Peritoneal carcinoma (Approved on 2018/12/19) |
| | Germline or somatic BRCA-mutated (gBRCAm or sBRCAm) Olaparib vs. Placebo [PFS(M): NR vs. 13.8] |
| OlympiAD^[67] NCT02000622 | Breast cancer (Approved on 2018/02/06) |
| | Germline BRCA mutation (deleterious/suspected deleterious) HER2-negative Olaparib vs. Chemotherapy [PFS(M): 7 vs. 4.2] |
| SOLO-2/ENGOT-Ov21^[68] NCT01874353 | Ovarian cancer, Fallopian tube cancer, Peritoneal carcinoma (Approved on 2017/08/17) |
| | gBRCA+ Olaparib vs. Placebo [PFS(M): 19.1 vs. 5.5] |
| Study19^[69] NCT00753545 | Ovarian cancer, Fallopian tube cancer, Peritoneal carcinoma (Approved on 2017/08/17) |
| | - Olaparib vs. Placebo [PFS(M): 8.4 vs. 4.8] |
| Study 42^[70] NCT01078662 | Ovarian cancer (Approved on 2014/12/19) |
| | Germline BRCA mutation (deleterious/suspected deleterious) Olaparib [ORR(%): 34.0, DOR(M): 7.9] |

Rucaparib (RUBRACA)

Rucaparib is an inhibitor of the DNA repair enzyme poly (ADP-ribose) polymerase-1, -2 and -3 (PARP-1, -2, -3). Rucaparib is developed and marketed by Clovis Oncology under the trade name RUBRACA.

FDA Approval Summary of Rucaparib (RUBRACA)

| | |
|-------------------------------|---|
| TRITON2 NCT02952534 | Prostate cancer (Approved on 2020/05/15) |
| | gBRCA+, sBRCA Rucaparib [ORR(%): 44.0, DOR(M): NE] |

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| | |
|--|---|
| ARIEL3^[18] NCT01968213 | Ovarian cancer, Fallopian tube cancer, Peritoneal carcinoma (Approved on 2018/04/06) |
| | All HRD tBRCA |
| | Rucaparib vs. Placebo [PFS (All)(M): 10.8 vs. 5.4, PFS (HRD)(M): 13.6 vs. 5.4, PFS (tBRCA)(M): 16.6 vs. 5.4] |
| | Ovarian cancer (Approved on 2016/12/19) |
| ARIEL2^[71] NCT01482715, NCT01891344 | Germline and/or somatic BRCA mutation |
| | Rucaparib |
| | [ORR(%): 54.0] |

Talazoparib (TALZENNA)

Talazoparib is an inhibitor of poly (ADP-ribose) polymerase (PARP) enzymes, including PARP1 and PARP2. Talazoparib is developed and marketed by Pfizer under the trade name TALZENNA.

FDA Approval Summary of Talazoparib (TALZENNA)

| | |
|--|---|
| EMBRACA^[72] NCT01945775 | Breast cancer (Approved on 2018/10/16) |
| | Germline BRCA mutation (deleterious/suspected deleterious) HER2-negative |
| | Talazoparib vs. Chemotherapy [PFS(M): 8.6 vs. 5.6] |

d=day; w=week; m=month

ONGOING CLINICAL TRIALS

Clinical trials shown below were selected by applying filters: study status, patient's diagnosis, intervention, location and/or biomarker(s). Please visit <https://clinicaltrials.gov> to search and view for a complete list of open available and updated matched trials.

No trial has been found.

DETAILED TEST RESULTS

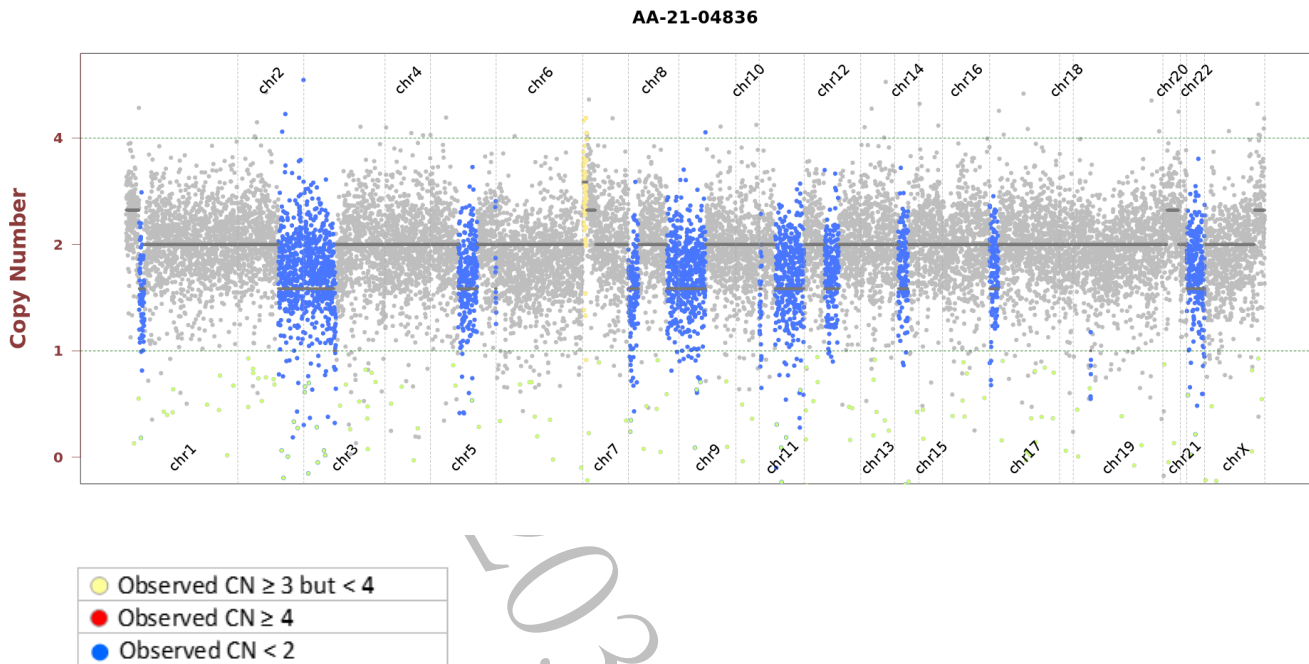
SINGLE NUCLEOTIDE AND SMALL INDEL VARIANTS

| Gene | Chr | Exon | Accession Number | cDNA Change | Amino Acid Change | Coverage | Allele Frequency | COSMIC ID |
|-------|-----|------|------------------|--------------|-------------------|----------|------------------|-------------|
| ETV4 | 17 | 13 | NM_001079675 | c.1298C>T | P433L | 488 | 51.8% | - |
| IL7R | 5 | 8 | NM_002185 | c.1134C>G | D378E | 870 | 47.6% | - |
| IL7R | 5 | 3 | NM_002185 | c.314G>A | S105N | 1225 | 47.3% | - |
| INSR | 19 | 22 | NM_000208 | c.4115G>A | R1372Q | 641 | 48.0% | - |
| KAT6A | 8 | - | NM_006766 | c.600+5G>T | Splice region | 814 | 54.8% | - |
| KDR | 4 | 8 | NM_002253 | c.1055C>T | A352V | 496 | 19.4% | - |
| LRP1B | 2 | 53 | NM_018557 | c.8413T>C | C2805R | 569 | 15.5% | COSM9312136 |
| MUC16 | 19 | 3 | NM_024690 | c.10543C>T | H3515Y | 539 | 48.8% | COSM371362 |
| MUC16 | 19 | 19 | NM_024690 | c.37181T>A | F12394Y | 1327 | 46.9% | - |
| NBN | 8 | 5 | NM_002485 | c.505C>T | R169C | 521 | 50.1% | - |
| PAX8 | 2 | 7 | NM_003466 | c.670C>T | R224C | 618 | 49.2% | - |
| POLD1 | 19 | 16 | NM_001256849 | c.1932C>G | D644E | 324 | 42.3% | - |
| PTEN | 10 | 8 | NM_000314 | c.974_976dup | L325_D326insV | 1079 | 18.7% | - |
| PTPRT | 20 | 31 | NM_007050 | c.4289A>G | Y1430C | 232 | 14.2% | - |

Mutations with clinical relevance are highlighted in red.

COPY NUMBER VARIANTS (CNVs)

Observed copy number (CN) for each evaluated position is shown on the y-axis. Regions referred to as amplification or deletion are shown in color. Regions without significant changes are represented in gray.



HOTSPOT GENOTYPES

Listed variants are biomarkers or hotspots that are recommended as standard care by the NCCN or other expert panels and not necessarily FDA-recognized for a particular indication. The genotypes have been manually checked to ensure sufficient coverage for each hotspot of the target gene.

| Gene | Variant | Genotype Detected |
|---------------|---|-------------------|
| <i>BRAF</i> | V600X | Not detected |
| <i>EGFR</i> | A763_Y764insFQEA, E709K, E709_T710delinsD, Exon 19 deletion, Exon 19 insertion, Exon 20 insertion, G719A/C/D/S, L747P, L833V, L858R, L861Q/R, S768I, T790M | Not detected |
| <i>IDH2</i> | R140Q, R172G/K/M/S | Not detected |
| <i>KIT</i> | A502_Y503dup, D419del, D579del, D816F/V/Y, D820A/E/G/Y, E554_I571del, E554_K558del, E554_V559del, Exon 11 mutation, F522C, H697Y, I563_L576del, I653T, K550_W557del, K558N, K558_E562del, K558_V559del, K558delinsNP, K642E, M552_W557del, N505I, N564_Y578del, N822H/I/K/Y, P551_M552del, P573_D579del, P577_D579del, P577_W582delinsPYD, P838L, Q556_K558del, T417_D419delinsI, T417_D419delinsRG, T574_Q575insTQLPYD, V530I, V555_L576del, V555_V559del, V559A/C/D/G, V559_V560del, V559del, V560D/G, V560del, V569_L576del, V654A, W557G/R, W557_K558del, Y553N, Y553_K558del, Y570H, Y578C | Not detected |
| <i>KRAS</i> | A146T/V/P, G12X, G13X, Q61X | Not detected |
| <i>MET</i> | D1028H/N/Y | Not detected |
| <i>NRAS</i> | G12X, G13X, Q61X | Not detected |
| <i>PDGFRA</i> | A633T, C450_K451insMIEWMI, C456_N468del, C456_R481del, D568N, D842I/V, D842_H845del, D842_M844del, D846Y, E311_K312del, G853D, H650Q, H845Y, H845_N848delinsP, I843del, N659K/R/S, N848K, P577S, Q579R, R560_V561insER, R748G, R841K, S566_E571delinsR, S584L, V469A, V536E, V544_L545insAVLVLLVIVISLI, V561A/D, V561_I562insER, V658A, W559_R560del, Y375_K455del, Y555C, Y849C/S | Not detected |
| <i>PIK3CA</i> | C420R, E542K/V, E545A/D/G/K, H1047X, Q546E/R | Not detected |

V600X= any mutation in the valine (V) at amino acid 600 being replaced by a different amino acid.

G12X = any mutation in the glycine (G) at amino acid 12 being replaced by a different amino acid.

G13X= any mutation in the glycine (G) at amino acid 13 being replaced by a different amino acid.

Q61X = any mutation in the glutamine (Q) at amino acid 61 being replaced by a different amino acid.

H1047X = any mutation in the histidine (H) at amino acid 1047 being replaced by a different amino acid.

| Gene | Copy Number Detected |
|--------------|----------------------|
| <i>CDK4</i> | 2 |
| <i>EGFR</i> | 3 |
| <i>ERBB2</i> | 2 |
| <i>MET</i> | 2 |

Copy number ≥ 8 is considered amplification

Other known alterations that are associated with sensitivity, resistance, and toxicity to therapies.

| Gene | Variant | Genotype Detected |
|--------|---|-------------------|
| AKT1 | E17K | Not detected |
| ALK | C1156Y, D1203N, G1202R, L1152R, S1206Y, T1151_L1152insT | Not detected |
| BRAF | K601E, L597V/Q/R/S | Not detected |
| DPYD | D949V, I560S, splice-site mutation | Not detected |
| EGFR | A750P, C797S/Y, S492R | Not detected |
| ERBB2 | V659E | Not detected |
| ESR1 | D538G, E380Q, L469V, L536H/P/Q/R, S432L, S463P, V422del, V534E, Y537C/N/S | Not detected |
| FGFR3 | G370C, G380R, K650E/N/R/M/T/Q, R248C, S249C, S371C, Y373C | Not detected |
| IDH1 | R132C/G/H/L/Q/S | Not detected |
| MAP2K1 | D67N, E203K, F53L, K57E/N, P124S, Q56P, Q56_V60del, R47Q, R49L, S222D | Not detected |
| PTEN | R130*/fs/G/L/P/Q | Not detected |
| TPMT | A154T, Y240C | Not detected |

| Gene | Copy Number Detected |
|-------|----------------------|
| FGFR1 | 2 |
| MDM2 | 2 |
| MDM4 | 2 |

Copy number ≥ 8 is considered amplification

TEST DETAILS

ABOUT ACTOnco®+

The test is a next-generation sequencing (NGS)-based assay developed for efficient and comprehensive genomic profiling of cancers. This test interrogates coding regions of 440 genes associated with cancer treatment, prognosis and diagnosis. Genetic mutations detected by this test include small-scale mutations like single nucleotide variants (SNVs), small insertions and deletions (INDELs) (≤ 15 nucleotides) and large-scale genomic alterations like copy number variations (CNVs).

See ACTOnco®+ Gene List' Section for details of gene sequenced.

DATABASE USED

- Reference genome: human genome sequence hg19
- COSMIC v.92
- Genome Aggregation database r2.1.1
- ClinVar (version 20210208)
- ACT Genomics in-house database

NEXT-GENERATION SEQUENCING (NGS) METHODS

Extracted genomic DNA was amplified using four pools of primer pairs targeting coding exons of analyzed genes. Amplicons were ligated with barcoded adaptors. Quality and quantity of amplified library were determined using the fragment analyzer (AATI) and Qubit (Invitrogen). Barcoded libraries were subsequently conjugated with sequencing beads by emulsion PCR and enriched using Ion Chef system (Thermo Fisher Scientific) according to the Ion PI Hi-Q Chef Kit protocol (Thermo Fisher Scientific) or Ion 540 Kit-Chef protocol (Thermo Fisher Scientific). Sequencing was performed on the Ion Proton or Ion S5 sequencer (Thermo Fisher Scientific).

Raw reads generated by the sequencer were mapped to the hg19 reference genome using the Ion Torrent Suite (version 5.10). Coverage depth was calculated using Torrent Coverage Analysis plug-in. Single nucleotide variants (SNVs) and short insertions/deletions (INDELs) were identified using the Torrent Variant Caller plug-in (version 5.10). The coverage was down-sampled to 4000. VEP (Variant Effect Predictor) (version 100) was used to annotate every variant using databases from Clinvar (version 20210208), COSMIC v.92 and Genome Aggregation database r2.1.1. Variants with coverage ≥ 25 , allele frequency $\geq 5\%$ and actionable variants with allele frequency $\geq 2\%$ were retained.

This test provides uniform coverage of the targeted regions, enabling target base coverage at $100\times \geq 85\%$ with a mean coverage $\geq 500\times$.

Variants reported in Genome Aggregation database r2.1.1 with $> 1\%$ minor allele frequency (MAF) were

considered as polymorphisms. ACT Genomics in-house database was used to determine technical errors. Clinically actionable and biologically significant variants were determined based on the published medical literature.

The copy number variations (CNVs) were predicted as described below:

Amplicons with read counts in the lowest 5th percentile of all detectable amplicons and amplicons with a coefficient of variation ≥ 0.3 were removed. The remaining amplicons were normalized to correct the pool design bias. ONCOCNV (an established method for calculating copy number aberrations in amplicon sequencing data by Boeva et al., 2014) was applied for the normalization of total amplicon number, amplicon GC content, amplicon length, and technology-related biases, followed by segmenting the sample with a gene-aware model. The method was used as well for establishing the baseline of copy number variations from samples in ACT Genomics in-house database.

Tumor mutational burden (TMB) was calculated by using the sequenced regions of ACT Onco®+ to estimate the number of somatic nonsynonymous mutations per megabase of all protein-coding genes (whole exome). The TMB calculation predicted somatic variants and applied a machine learning model with a cancer hotspot correction. TMB may be reported as “TMB-High”, “TMB-Low” or “Cannot Be Determined”. TMB-High corresponds to ≥ 7.5 mutations per megabase (Muts/Mb); TMB-Low corresponds to < 7.5 Muts/Mb. TMB is reported as “Cannot Be Determined” if the tumor purity of the sample is $< 30\%$.

Classification of microsatellite instability (MSI) status is determined by a machine learning prediction algorithm. The change of a number of repeats of different lengths from a pooled microsatellite stable (MSS) baseline in > 400 genomic loci are used as the features for the algorithm. The final output of the results is either microsatellite Stable (MSS) or microsatellite instability high (MSI-H).

STANDARD OPERATING PROCEDURES (SOPS)

Standard operating procedures (SOPs) are shown below:

- AG2-QP-15 Specimen Management Procedure
- AG3-QP16-03 SOP of Cancer Cell DNA and RNA Extraction
- AG3-QP16-07 SOP of Nucleic Acid Extraction with QIAasympyphony SP
- AG3-QP16-08 SOP of FFPE Nucleic Acid Extraction
- AG3-QP16-10 SOP of HE Staining
- AG3-QP16-13 SOP of Library Construction and Preparation
- AG3-QP16-17 SOP of DNA Quantification with Qubit Fluorometer
- AG3-QP16-20 SOP of CE-Fragment Analysis
- AG3-QP16-22 SOP of Variant Calling
- AG3-QP16-24 SOP of Ion Torrent System Sequencing Reaction
- AG3-QP16-26 SOP of Ion Chef Preparation

行動基因僅提供技術檢測服務及檢測報告，檢測結果之臨床解釋及相關醫療處置，請諮詢專業醫師。報告結果僅對此試驗件有效。

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- AG3-QP16-35 SOP of Variant Annotation
- AG3-QP16-96 SOP of Manual Inspection for SNV/Indel Variant
- AG3-QP16-95 SOP of Manual Inspection for Copy Number Variant
- AG3-QP40-08 (02) Standard protocol for variant interpretation, curation and classification
- AG3-QP16-41 SOP of The user manual for clinical report system (CRS)

LIMITATIONS

This test does not provide information of variant causality and does not detect variants in non-coding regions that could affect gene expression. This report does not report polymorphisms and we do not classify whether a mutation is germline or somatic. Variants identified by this assay were not subject to validation by Sanger or other technologies.

NOTES

We do not exclude the possibility that pathogenic variants may not be reported by one or more of the tools and the parameters used.

PATHOLOGY EVALUATION

- H&E-stained section No.: S11030322
- Collection site: Colon
- Examined by: Dr. Yeh-Han Wang
- Estimated neoplastic nuclei (whole sample): The percentage of viable tumor cells in total cells in the whole slide (%): 20%
The percentage of viable tumor cells in total cells in the encircled areas in the whole slide (%): 30%
The percentage of necrotic cells (including necrotic tumor cells) in total cells in the whole slide (%): 0%
The percentage of necrotic cells (including necrotic tumor cells) in total cells in the encircled areas in the whole slide (%): 0%
Additional comment: NA
- Manual macrodissection: Performed on the highlighted region



The outline highlights the area of malignant neoplasm annotated by a pathologist.

SPECIMEN PHOTO(S)



- Collection date: Oct 2021
- Facility retrieved: 臺北榮總

RUN QC

- Panel: ACTOnco[®]+
- Mean Depth: 595x
- Target Base Coverage at 100x: 93%

ACTOnco® + GENE LIST

| | | | | | | | | | | | | | |
|----------|---------|--------|----------|--------|--------|-----------|--------|---------|----------|---------|----------|----------|----------|
| ABC1* | AURKB | CBL | CDKN2B | E2F3 | FAT1 | GRIN2A | JAK2 | MED12 | NOTCH4 | PMS1 | RAD51D | SLCO1B3* | TNFRSF14 |
| ABC2* | AXIN1 | CCNA1 | CDKN2C | EGFR | FBXW7 | GSK3B | JAK3 | MEF2B | NPM1 | PMS2 | RAD52 | SMAD2 | TNFSF11 |
| ABC2* | AXIN2 | CCNA2 | CEBPA* | EP300 | FCGR2B | GSTP1* | JUN* | MEN1 | NQO1* | POLB | RAD54L | SMAD3 | TOP1 |
| ABL1 | AXL | CCNB1 | CHEK1 | EPCAM | FGF1* | GSTT1* | KAT6A | MET | NRAS | POLD1 | RAF1 | SMAD4 | TP53 |
| ABL2 | B2M | CCNB2 | CHEK2 | EPHA2 | FGF10 | HGF | KDM5A | MITF | NSD1 | POLE | RARA | SMARCA4 | TPMT* |
| ADAMTS1 | BAP1 | CCNB3 | CIC | EPHA3 | FGF14 | HIF1A | KDM5C | MLH1 | NTRK1 | PPARG | RB1 | SMARCB1 | TSC1 |
| ADAMTS13 | BARD1 | CCND1 | CREBBP | EPHA5 | FGF19* | HIST1H1C* | KDM6A | MPL | NTRK2 | PPP2R1A | RBM10 | SMO | TSC2 |
| ADAMTS15 | BCL10 | CCND2 | CRKL | EPHA7 | FGF23 | HIST1H1E* | KDR | MRE11 | NTRK3 | PRDM1 | RECQL4 | SOC1* | TSHR |
| ADAMTS16 | BCL2* | CCND3 | CRLF2 | EPHB1 | FGF3 | HNF1A | KEAP1 | MSH2 | PAK3 | PRKAR1A | REL | SOX2* | TYMS |
| ADAMTS18 | BCL2L1 | CCNE1 | CSF1R | ERBB2 | FGF4* | HR | KIT | MSH6 | PALB2 | PRKCA | RET | SOX9 | U2AF1 |
| ADAMTS6 | BCL2L2* | CCNE2 | CTCF | ERBB3 | FGF6 | HRAS* | KMT2A | MTHFR* | PARP1 | PRKCB | RHOA | SPEN | UBE2A* |
| ADAMTS9 | BCL6 | CCNH | CTLA4 | ERBB4 | FGFR1 | HSP90AA1 | KMT2C | MTOR | PAX5 | PRKCG | RICTOR | SPOP | UBE2K |
| ADAMTSL1 | BCL9 | CD19 | CTNNA1 | ERCC1 | FGFR2 | HSP90AB1 | KMT2D | MUC16 | PAX8 | PRKCI | RNF43 | SRC | UBR5 |
| ADGRA2 | BCOR | CD274 | CTNNB1 | ERCC2 | FGFR3 | HSPA4 | KRAS | MUC4 | PBRM1 | PRKCQ | ROS1 | STAG2 | UGT1A1* |
| ADH1C* | BIRC2 | CD58 | CUL3 | ERCC3 | FGFR4 | HSPA5 | LCK | MUC6 | PDCD1 | PRKDC | RPPH1 | STAT3 | USH2A |
| AKT1 | BIRC3 | CD70* | CYLD | ERCC4 | FH | IDH1 | LIG1 | MUTYH | PDCD1LG2 | PRKN | RPTOR | STK11 | VDR* |
| AKT2 | BLM | CD79A | CYP1A1* | ERCC5 | FLCN | IDH2 | LIG3 | MYC | PDGFRA | PSMB8 | RUNX1 | SUFU | VEGFA |
| AKT3 | BMPR1A | CD79B | CYP2B6* | ERG | FLT1 | IFNL3* | LMO1 | MYCL | PDGFRB | PSMB9 | RUNX1T1 | SYK | VEGFB |
| ALDH1A1* | BRAF | CDC73 | CYP2C19* | ESR1 | FLT3 | IGF1 | LRP1B | MYCN | PDIA3 | PSME1 | RXRA | SYNE1 | VHL |
| ALK | BRCA1 | CDH1 | CYP2C8* | ESR2 | FLT4 | IGF1R | LYN | MYD88 | PGF | PSME2 | SDHA | TAF1 | WT1 |
| AMER1 | BRCA2 | CDK1 | CYP2D6 | ETV1 | FOXL2* | IGF2 | MALT1 | NAT2* | PHOX2B* | PSME3 | SDHB | TAP1 | XIAP |
| APC | BRD4 | CDK12 | CYP2E1* | ETV4 | FOXP1 | IKBKB | MAP2K1 | NBN | PIK3C2B | PTCH1 | SDHC | TAP2 | XPO1 |
| AR | BRIP1 | CDK2 | CYP3A4* | EZH2 | FRG1 | IKBKE | MAP2K2 | NEFH | PIK3C2G | PTEN | SDHD | TAPBP | XRCC2 |
| ARAF | BTG1* | CDK4 | CYP3A5* | FAM46C | FUBP1 | IKZF1 | MAP2K4 | NF1 | PIK3C3 | PTGS2 | SERPINB3 | TBX3 | ZNF217 |
| ARID1A | BTG2* | CDK5 | DAXX | FANCA | GATA1 | IL6 | MAP3K1 | NF2 | PIK3CA | PTPN11 | SERPINB4 | TEK | |
| ARID1B | BTK | CDK6 | DCUN1D1 | FANCC | GATA2 | IL7R | MAP3K7 | NFE2L2 | PIK3CB | PTPRD | SETD2 | TERT | |
| ARID2 | BUB1B | CDK7 | DDR2 | FANCD2 | GATA3 | INPP4B | MAPK1 | NFKB1 | PIK3CD | PTPRT | SF3B1 | TET1 | |
| ASXL1 | CALR | CDK8 | DICER1 | FANCE | GNA11 | INSR | MAPK3 | NFKBIA | PIK3CG | RAC1 | SGK1 | TET2 | |
| ATM | CANX | CDK9 | DNMT3A | FANCF | GNA13 | IRF4 | MAX | NKX2-1* | PIK3R1 | RAD50 | SH2D1A* | TGFBR2 | |
| ATR | CARD11 | CDKN1A | DOT1L | FANCG | GNAQ | IRS1 | MCL1 | NOTCH1 | PIK3R2 | RAD51 | SLC19A1* | TMSB4X* | |
| ATRX | CASP8 | CDKN1B | DPYD | FANCL | GNAS | IRS2* | MDM2 | NOTCH2 | PIK3R3 | RAD51B | SLC22A2* | TNF | |
| AURKA | CBFB | CDKN2A | DTX1 | FAS | GREM1 | JAK1 | MDM4 | NOTCH3 | PIM1 | RAD51C | SLCO1B1* | TNFAIP3 | |

*Analysis of copy number alteration not available.

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In this report, neither any biomarker alteration nor any drug associated with a potential clinical benefit (or potential lack of clinical benefit), are ranked in order of potential or predicted efficacy.

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任何治療與用藥需經由醫師在考慮病患所有健康狀況相關資訊包含健檢、其他檢測報告和病患意願後，依照該地區醫療照護標準由醫師獨立判斷。醫師不應僅依據單一報告結果(例如本檢測或本報告書內容)做決策。

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本報告中列出之生物標記變異與藥物資訊並非依照潛在治療有效性排序。

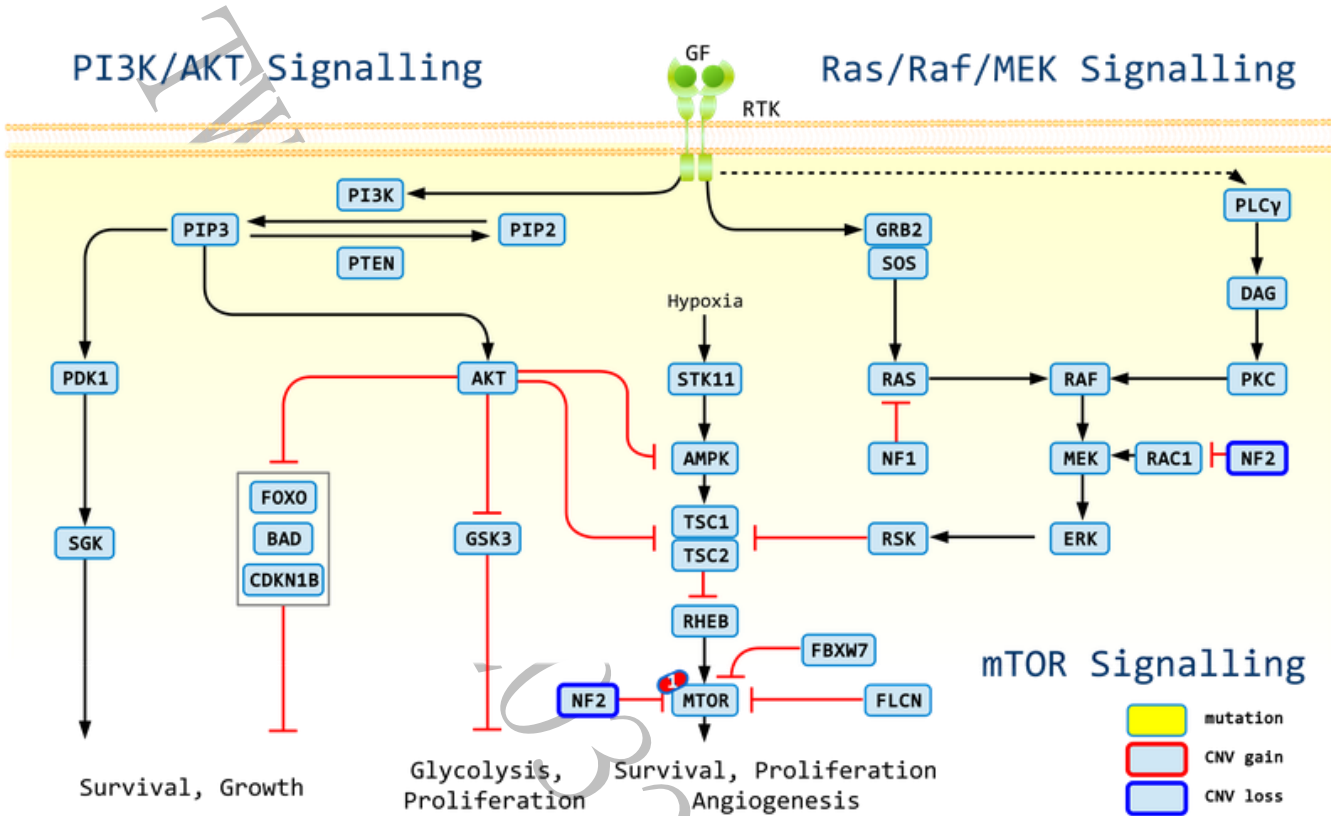
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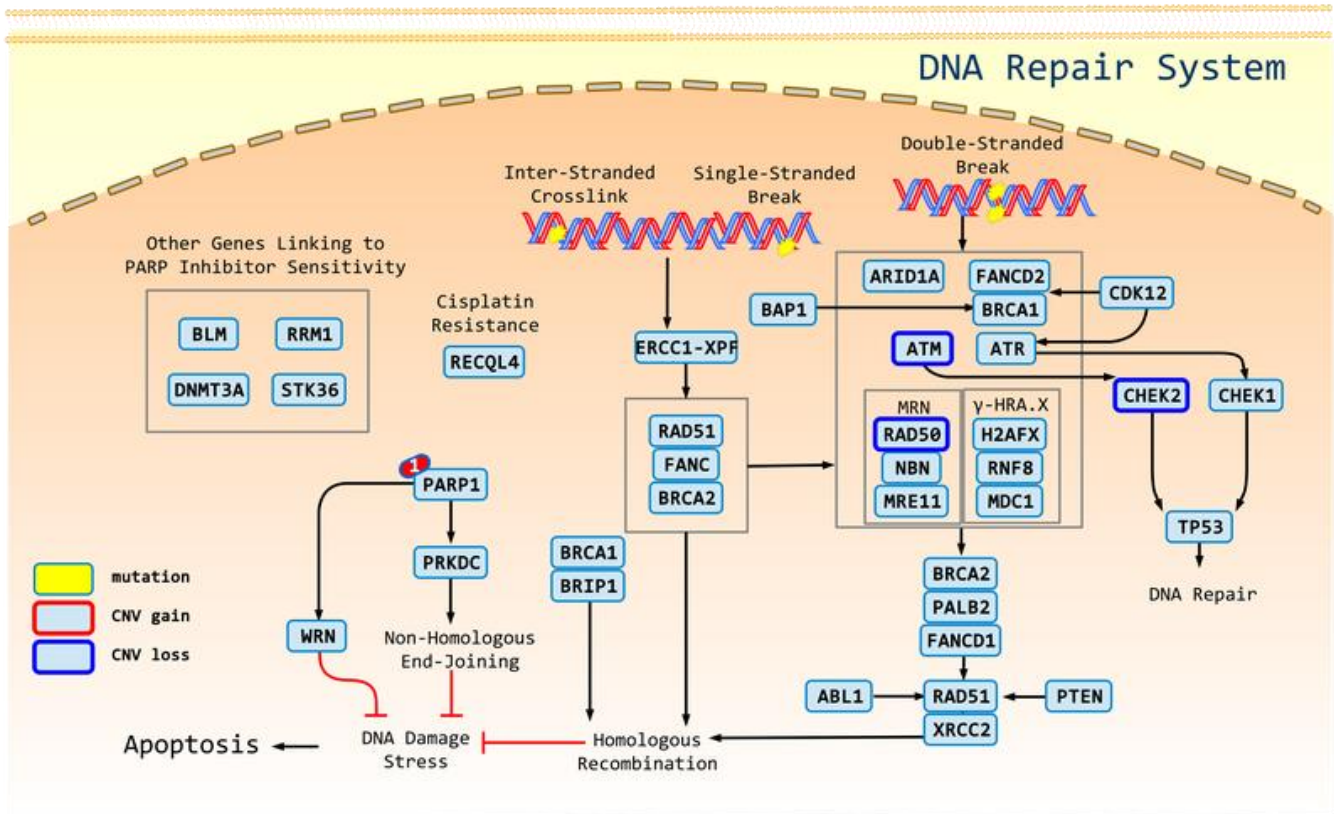
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SIGNALING PATHWAYS AND MOLECULAR-TARGETED AGENTS



1: Everolimus



1: Olaparib, Niraparib, Rucaparib, Talazoparib

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| PATIENT | SPECIMEN | ORDERING PHYSICIAN |
|-----------------------------------|-----------------------------|----------------------------|
| Name: 陳昭綺 | Type: FFPE tissue | Name: 陳明晃醫師 |
| Gender: Female | Date received: Oct 28, 2021 | Facility: 臺北榮總 |
| Date of Birth: May 03, 1965 | Collection site: Colon | Tel: 886-228712121 |
| Patient ID: 19749225 | Specimen ID: S11030322 | Address: 臺北市北投區石牌路二段 201 號 |
| Diagnosis: Gastric adenocarcinoma | Lab ID: AA-21-04836 | |
| | D/ID: NA | |

ABOUT ACTFusion™

The test is a next-generation sequencing (NGS) based in vitro diagnostic assay to detect fusion transcripts of 13 genes, including *ALK*, *BRAF*, *EGFR*, *FGFR1*, *FGFR2*, *FGFR3*, *MET*, *NRG1*, *NTRK1*, *NTRK2*, *NTRK3*, *RET*, and *ROS1*.

VARIANT(S) WITH CLINICAL RELEVANCE
FUSION RESULTS

No fusion gene detected in this sample.

Variant Analysis:

醫檢師陳韻仔 博士
 Yun-Yu Chen Ph.D.
 檢字第 015647 號

Yun Yu Chen

Sign Off

醫檢師陳韻仔 博士
 Yun-Yu Chen Ph.D.
 檢字第 015647 號

Yun Yu Chen

THERAPEUTIC IMPLICATIONS**TARGETED THERAPIES**

Not Applicable.

VARIANT INTERPRETATION

Not Applicable.

US FDA-APPROVED DRUG(S)

Not Applicable.

ONGOING CLINICAL TRIAL(S)

Clinical trials shown below were selected by applying filters: study status, patient's diagnosis, intervention, location and/or biomarker(s). Please visit <https://clinicaltrials.gov> to search and view for a complete list of open available and updated matched trials.

No trial has been found.

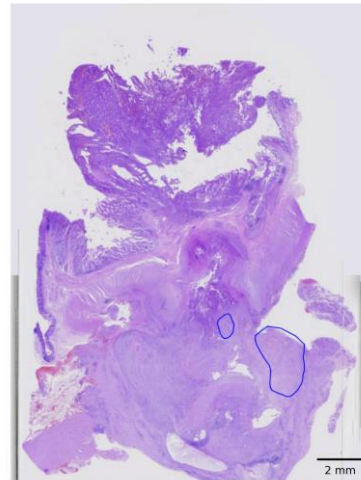
ACTFusion™ Report

ACTFusion™ GENE LIST

| ALK | BRAF | EGFR | FGFR1 | FGFR2 | FGFR3 | MET | NRG1 |
|-------|-------|-------|-------|-------|-------|-----|------|
| NTRK1 | NTRK2 | NTRK3 | RET | ROS1 | - | - | - |

TEST DETAILS

SPECIMEN RECEIVED



- H&E-stained section No.: S11030322
- Collection date: Oct 2021
- Collection site: Colon
- Facility retrieved: 臺北榮總
- Examined by: Dr. Yeh-Han Wang
- Estimated neoplastic nuclei (whole sample): The percentage of viable tumor cells in total cells in the whole slide (%): 20%
The percentage of viable tumor cells in total cells in the encircled areas in the whole slide (%): 30%
The percentage of necrotic cells (including necrotic tumor cells) in total cells in the whole slide (%): 0%
The percentage of necrotic cells (including necrotic tumor cells) in total cells in the encircled areas in the whole slide (%): 0%
Additional comment: NA
- Manual macrodissection: Performed on the highlighted region

The outline highlights the area of malignant neoplasm annotated by a pathologist.

NEXT-GENERATION SEQUENCING (NGS) METHODS

The extracted RNA was reverse-transcribed and subjected to library construction. The quality and quantity of the amplified library was determined using the fragment analyzer (AATI) and Qubit (Invitrogen). Sequencing was performed on the Ion 540™ Chip/ Ion 550™ Chip / Ion P1™ Chip and Ion GeneStudio™ S5 Prime System / Ion Proton™ System (Life Technologies). All assays were performed in accordance with ACT Genomics testing SOPs.

Data processing and statistical analysis for the identification of relevant fusions was performed using in-house fusion calling pipeline with default parameter setting. The four internal controls for the purpose of monitoring the overall sequencing quality of the sample were built into the assay, including CHMP2A, RABA7A, GPI, and VCP. Amplification of these genes using gene specific primers was performed, and the sequencing results were applied to the analysis pipeline to assess RNA quality. The inability of the software to detect these genes was considered a run failure. To ensure optimal sequencing quality for variant analysis, all samples had to meet the following sample quality control (QC) criteria: 1) Average unique RNA Start Sites (SS) per control Gene Specific Primer 2 (GSP 2) ≥ 10 (default), and 2) Total reads after sequencing $\geq 500,000$ (recommended).

Samples passed the sample QC would be subjected to the fusion analysis pipeline for fusion transcript calling. Briefly, the analysis pipeline aligned sequenced reads to a reference genome, identified regions that map to noncontiguous regions of the genome, and applied filters to exclude probable false-positive events and annotate previously characterized fusion events. A minimum of 5 reads with 3 unique sequencing start sites that cross the breakpoints was set as the cutoff value to indicate strong evidence of fusions. RNA fusions would need to be in frame in order to generate productive transcripts. In addition, databases with details for documented fusions were used to authenticate the fusion sequence identified. Known fusions were queried using Quiver Gene Fusion Database, a curated database owned and maintained by ArcherDX. In summary, samples with detectable fusions had to meet the following criteria: 1) Number of unique start sites (SS) for the GSP2 ≥ 3 . 2) Number of supporting reads spanning the fusion junction ≥ 5 . 3) Percentage of supporting reads spanning the fusion junction $\geq 10\%$. 4) Fusions annotated in Quiver Gene Fusion Database.

DATABASE USED

Quiver Gene Fusion Database version 5.1.18

LIMITATIONS

This test has been designed to detect fusions in 13 genes sequenced. Therefore, fusion in genes not covered by this test would not be reported. For novel fusions detected in this test, Sanger sequencing confirmation is recommended if residue specimen is available.

STANDARD OPERATING PROCEDURES (SOPs)

Standard operating procedures (SOPs) are shown below:

- AG2-QP-15 Specimen Management Procedure
- AG3-QP16-08 SOP of FFPE Nucleic Acid Extraction
- AG3-QP16-10 SOP of HE Staining
- AG3-QP16-17 SOP of DNA Quantification with Qubit Fluorometer
- AG3-QP16-20 SOP of CE-Fragment Analysis
- AG3-QP16-24 SOP of Ion Torrent System Sequencing Reaction
- AG3-QP16-26 SOP of Ion Chef Preparation
- AG3-QP40-08 (02) Standard protocol for variant interpretation, curation and classification
- AG3-QP16-94 (01) SOP of ACTFusion v3 Library Construction and Preparation
- AG3-QP16-36(02) SOP of Fusion Gene Detection
- AG3-QP16-41 SOP of The user manual for clinical report system (CRS)

RUN QC

- Panel: ACTFusion™
- Total reads: 684374
- Average unique RNA Start Sites per control GSP2: 158

DISCLAIMER**Legal Statement**

This test was developed by ACT Genomics and its performing characteristics were determined by ACT Genomics. This test result is to be used for clinical consultative purposes only and is not intended as a substitute for a clinical guidance of your doctor or another qualified medical practitioner. It should not be regarded as investigational or used for research.

The detection of genomic alterations does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; the detection of no genomic alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Treatment Decisions are the Responsibility of the Physician

Decisions on clinical care and treatment should be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, including physical examinations, information from other diagnostics tests and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this test, or the information contained in this report.

In terms of consulting a different treating physician, the patient must file an application and fulfill the listed criteria for ACT Genomics to provide the patient's report to the assigned physician. The report may not be copied or reproduced except in its totality.

Genetic Alterations and Drugs Not Presented in Ranked Order

In this report, neither any biomarker alteration nor any drug associated with a potential clinical benefit (or potential lack of clinical benefit), are ranked in order of potential or predicted efficacy.

Level of Evidence Provided

Drugs with a potential clinical benefit (or potential lack of clinical benefit) are evaluated for level of published evidence with at least one clinical efficacy case report or preclinical study. We endeavor to keep the information in the report up to date. However, customers must be aware that scientific understanding and technologies change over time, and we make no warranty as to the accuracy, suitability or currency of information provided in this report at any time.

No Guarantee of Clinical Benefit

This report makes no promises or guarantees about the effectiveness of a particular drug or any treatment procedure in any disease or in any patient. This report also makes no promises or guarantees that a drug without an association of reportable genomic alteration will, in fact, provide no clinical benefit.

Liability

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免責聲明

法律聲明

本檢驗報告僅提供專業醫療參考，結果需經專業醫師解釋及判讀。基因突變資訊非必具備藥物或治療有效性指標，反之亦然。本檢驗報告提供之用藥指引不聲明或保證其臨床有效性，反之亦然。本基因檢測方法係由本公司研究開發，已經過有效性測試。

本檢驗報告非經本公司許可，不得私自變造、塗改，或以任何方式作為廣告及其他宣傳之用途。

本公司於提供檢驗報告後，即已完成本次契約義務，後續之報告解釋、判讀及用藥、治療，應自行尋求相關專業醫師協助，若需將報告移件其他醫師，本人應取得該醫師同意並填寫移件申請書，主動告知行動基因，行動基因僅能配合該醫師意願與時間提供醫師解說。

醫療決策需由醫師決定

任何治療與用藥需經由醫師在考慮病患所有健康狀況相關資訊包含健檢、其他檢測報告和病患意願後，依照該地區醫療照護標準由醫師獨立判斷。醫師不應僅依據單一報告結果(例如本檢測或本報告書內容)做決策。

基因突變與用藥資訊並非依照有效性排序

本報告中列出之生物標記變異與藥物資訊並非依照潛在治療有效性排序。

證據等級

藥物潛在臨床效益(或缺乏潛在臨床效益)的實證證據是依據至少一篇臨床療效個案報告或臨床前試驗做為評估。本公司盡力提供適時及準確之資料，但由於醫學科技之發展日新月異，本公司不就本報告提供的資料是否為準確、適宜或最新作保證。

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REFERENCES

Not Applicable.