

**ABOUT THE TEST** FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

<b>PATIENT</b>	<b>DISEASE</b> Brain glioblastoma (GBM)	<b>PHYSICIAN</b>	<b>ORDERING PHYSICIAN</b> Yeh, Yi-Chen	<b>SPECIMEN</b>	<b>SPECIMEN SITE</b> Brain
	<b>NAME</b> Huang, Chin I		<b>MEDICAL FACILITY</b> Taipei Veterans General Hospital		<b>SPECIMEN ID</b> S112-18016A (PF23080)
	<b>DATE OF BIRTH</b> 11 December 1961		<b>ADDITIONAL RECIPIENT</b> None		<b>SPECIMEN TYPE</b> Slide Deck
	<b>SEX</b> Male		<b>MEDICAL FACILITY ID</b> 205872		<b>DATE OF COLLECTION</b> 25 April 2023
	<b>MEDICAL RECORD #</b> 49450933		<b>PATHOLOGIST</b> Not Provided		<b>SPECIMEN RECEIVED</b> 14 June 2023

## Biomarker Findings

**Microsatellite status** - MS-Stable  
**Tumor Mutational Burden** - 0 Muts/Mb

## Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

**EGFR** S229C, R222C - subclonal, amplification, rearrangement intron 24<sup>†</sup>

**MTAP** loss

**CDKN2A/B** CDKN2A loss, CDKN2B loss

**RB1** Q436\*, E237\*

**TERT** promoter -124C>T

2 Disease relevant genes with no reportable alterations: **IDH1**, **PDGFRA**

<sup>†</sup> See About the Test in appendix for details.

## Report Highlights

- Variants with **diagnostic implications** that may indicate a specific cancer type: **EGFR** amplification (p. 4), **TERT** promoter -124C>T (p. 7)
- Targeted therapies with potential clinical benefit **approved in another tumor type**: Cetuximab (p. 8), Erlotinib (p. 8), Gefitinib (p. 9), Osimertinib (p. 10), Panitumumab (p. 11)
- Evidence-matched **clinical trial options** based on this patient's genomic findings: (p. 12)
- Variants with **prognostic implications** for this tumor type that may impact treatment decisions: **TERT** promoter -124C>T (p. 7)

### BIOMARKER FINDINGS

**Microsatellite status** - MS-Stable

**Tumor Mutational Burden** - 0 Muts/Mb

### GENOMIC FINDINGS

**EGFR** - S229C, R222C - subclonal, amplification, rearrangement intron 24

6 Trials see p. 12

**MTAP** - loss

1 Trial see p. 14

### THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. See Biomarker Findings section

No therapies or clinical trials. See Biomarker Findings section

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)
none	Cetuximab
	Erlotinib
	Gefitinib
	Osimertinib
	Panitumumab
none	none

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**GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS**

*For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.*

**CDKN2A/B** - CDKN2A loss, CDKN2B loss ..... [p. 6](#)    **TERT** - promoter -124C>T ..... [p. 7](#)  
**RB1** - Q436\*, E237\* ..... [p. 7](#)

**NOTE** Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

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## BIOMARKER FINDINGS

## BIOMARKER

## Microsatellite status

## RESULT

MS-Stable

### POTENTIAL TREATMENT STRATEGIES

#### — Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors<sup>1-3</sup>, including approved therapies nivolumab and pembrolizumab<sup>4</sup>. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and

experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%,  $p=0.001$ )<sup>5</sup>.

### FREQUENCY & PROGNOSIS

Low-level MSI has been reported in 5-9% of glioblastoma (GBM) samples<sup>6-8</sup>. A large-scale study did not find high-level microsatellite instability (MSI-H) in any of 129 GBM samples<sup>6</sup>, although a small-scale study reported MSI-H in 4 of 15 pediatric GBMs and 1 of 12 adult GBMs<sup>9</sup>. The frequency of MSI has been reported to be increased in relapsed compared to primary GBM<sup>6</sup>, in GBMs with a previous lower grade astrocytoma<sup>7</sup>, and in giant cell GBM compared to classic GBM<sup>8</sup>.

### FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor<sup>10</sup>. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2<sup>10-12</sup>. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers<sup>13-15</sup>. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins<sup>10,12,14-15</sup>.

## BIOMARKER

## Tumor Mutational Burden

## RESULT

0 Muts/Mb

### POTENTIAL TREATMENT STRATEGIES

#### — Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1<sup>16-18</sup>, anti-PD-1 therapies<sup>16-19</sup>, and combination nivolumab and ipilimumab<sup>20-25</sup>. In glioma, a lack of association between TMB and clinical benefit from immune checkpoint inhibitors has been reported<sup>16,26-27</sup>. However, multiple case studies have reported that patients with ultramutated gliomas driven by POLE mutations

have benefited from treatment with anti-PD-1<sup>28-29</sup> or anti-PD-L1<sup>30</sup> therapies. Therefore, although increased TMB alone may not be a strong biomarker for PD-1 or PD-L1 inhibitors in this cancer type, these agents may have efficacy for patients with glioma harboring both high TMB and POLE mutation.

### FREQUENCY & PROGNOSIS

Glioblastoma (GBM) harbors a median TMB of 2.7 mutations per megabase (mut/Mb), and 4.2% of cases have high TMB (>20 muts/Mb)<sup>31</sup>. For pediatric patients, high TMB has been reported in a subset of high-grade gliomas, frequently in association with mutations in mismatch repair or proofreading genes and in TP53, whereas BRAF alterations or other oncogene fusions were observed more frequently in brain tumors harboring low TMB<sup>32-33</sup>. Increased TMB has been reported to correlate with higher tumor grade in glioma<sup>34</sup> and glioblastoma (GBM) tissue samples with biallelic mismatch repair deficiency

(bMMRD)<sup>28</sup>, as well as with shorter OS of patients with diffuse glioma<sup>35</sup>.

### FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma<sup>36-37</sup> and cigarette smoke in lung cancer<sup>38-39</sup>, treatment with temozolomide-based chemotherapy in glioma<sup>40-41</sup>, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes<sup>42-46</sup>, and microsatellite instability (MSI)<sup>42,45-46</sup>. This sample harbors a TMB below levels that would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents<sup>16,26-30</sup>.

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## GENOMIC FINDINGS

## GENE

## EGFR

## ALTERATION

S229C, R222C - subclonal, amplification, rearrangement intron 24

## HGVS VARIANT

NM\_005228.3:c.685A>T (p.S229C),  
NM\_005228.3:c.664C>T (p.R222C)

## VARIANT CHROMOSOMAL POSITION

chr7:55220295, chr7:55220274

## VARIANT ALLELE FREQUENCY (% VAF)

57.2%, 4.4%

## POTENTIAL TREATMENT STRATEGIES

## — Targeted Therapies —

In multiple glioblastoma (GBM) studies, the presence of EGFRvIII has not predicted clinical benefit from first-generation EGFR TKIs such as erlotinib<sup>47-52</sup> or gefitinib<sup>50,53</sup>. However, case reports have described patients with EGFRvIII-positive GBM responding to erlotinib<sup>54-57</sup>. In a retrospective study of patients with GBM treated with erlotinib or gefitinib, co-expression of EGFRvIII with PTEN protein was the strongest predictor of response ( $P < 0.001$ )<sup>58</sup>, suggesting that activity in this setting is dependent on PTEN status<sup>59-60</sup>. However, a prospective Phase 2 trial testing erlotinib monotherapy for patients with EGFRvIII and PTEN-positive recurrent glioblastoma reported minimal efficacy and was terminated<sup>52</sup>. The second-generation EGFR TKIs afatinib and dacomitinib have shown minimal efficacy for patients with EGFRvIII glioblastoma (GBM)<sup>61-64</sup>. A Phase 1/2 study of afatinib, temozolomide, or the combination for patients with GBM reported clinical benefit, including for patients with EGFRvIII; however, temozolomide alone and in combination exhibited better responses than afatinib monotherapy<sup>61-62</sup>. A Phase 2 trial of dacomitinib for patients with EGFR-amplified GBM reported a DCR of 26% (5/19) among patients with EGFR amplification and EGFRvIII; however, the trial failed to meet its primary endpoint of 6-month PFS<sup>63</sup>. A retrospective biomarker analysis of another Phase 2 study of dacomitinib for patients with GBM found no association between EGFRvIII and clinical benefit<sup>64</sup>. Patients with glioma and co-occurring EGFR amplification and EGFRvIII have reported responses to osimertinib<sup>65</sup>. However, a patient with multiple glioblastoma (GBM) tumors, one of which harbored EGFRvIII,

experienced progression of the EGFRvIII-positive tumor during treatment with osimertinib<sup>66</sup>. On the basis of preclinical data, osimertinib inhibits EGFRvIII-driven tumor growth in vitro<sup>67-68</sup>. For patients with non-small cell lung cancer (NSCLC), EGFR activating mutations may predict sensitivity to EGFR-TKIs, including erlotinib<sup>69</sup>, gefitinib<sup>70-73</sup>, afatinib<sup>74-77</sup>, dacomitinib<sup>78</sup>, and osimertinib<sup>75,79</sup>; however, the data for patients with other tumor types are limited<sup>64,80-84</sup>. Patients with EGFR-mutated bithalamic glioma have reported responses to osimertinib<sup>66,85</sup>. In a case series of 11 patients with bithalamic gliomas with EGFR mutations, EGFR inhibitors, including osimertinib, showed improved survival; however, it showed a lack of significant clinical responses<sup>83</sup>. On the basis of preclinical data, EGFR mutations confer sensitivity to EGFR inhibitors, including osimertinib<sup>83</sup>. Clinical studies of the second-generation EGFR TKIs afatinib and dacomitinib for patients with EGFR-amplified gliomas have shown limited efficacy<sup>61,63-64,86-87</sup>; however, a small subset of patients has experienced clinical benefit<sup>63-64,86</sup>. Multiple studies have failed to find a positive association between increased EGFR expression and clinical benefit from erlotinib or gefitinib for patients with glioblastoma<sup>58,88-90</sup>. There are conflicting data on the efficacy of anti-EGFR antibodies for the treatment of EGFR-amplified tumors. A meta-analysis of colorectal cancer patients treated with second-line or higher cetuximab or panitumumab observed an association between EGFR copy number gain and increased OS and PFS<sup>91</sup>. However, studies of patients with head and neck squamous cell carcinoma or gastric cancer found either no association or a negative association between EGFR copy number gain and survival after treatment with first-line cetuximab or panitumumab in combination with chemotherapy<sup>92-93</sup>. The Phase 3 INTELLANCE trial of depatuxizumab mafodotin (ABT-414), an EGFR-targeted antibody-drug conjugate with a toxic payload, in patients with EGFR-amplified glioblastoma (GBM) was stopped for futility. Interim analysis demonstrated improved median PFS (mPFS) of ABT-414 monotherapy compared with placebo (HR=0.84); however, no OS benefit was observed (HR=1.01). Improved mPFS was also observed in patients harboring EGFRvIII (HR=0.73) but without an OS improvement (HR=0.95)<sup>94</sup>. The Phase 2 INTELLANCE trial demonstrated clinical benefit for EGFR-amplified GBM for the combination of ABT-414, temozolomide, and radiotherapy (HR=0.66,  $p=0.017$ ), but there was no

evidence of efficacy for ABT-414 monotherapy (HR=1.04,  $p=0.83$ )<sup>95</sup>. A single-center Phase 2 pilot study for EGFR-amplified solid tumors ( $n=13$ ) reported 1 PR and 2 SDs, including 1 with a 9-month duration, for the 8 evaluable patients with glioblastoma<sup>96</sup>.

## FREQUENCY &amp; PROGNOSIS

Across several genomic studies of CNS tumors, EGFR amplification has been reported in 16.9% of anaplastic astrocytomas, and 39.7% of glioblastoma multiformes (GBMs)<sup>97-100</sup>. EGFR alterations have been reported in 13.2% of anaplastic astrocytomas, 5.3-15.9% of glioblastoma multiformes (GBMs), and 0% of pilocytic astrocytomas in several genomic studies of CNS tumors<sup>97-100</sup>. In GBMs, Missense mutations in the EGFR extracellular domain have been found in 10-15% of cases and approximately half have a low-level amplification of the mutated allele<sup>101-102</sup>. In a study of IDH-wildtype GBM samples, EGFR alterations were detected in 50% (117/232) of IDH-wildtype GBM samples analyzed, including 41% (95/232) with a co-occurring EGFR amplification and mutation, 26% (61/232) with an EGFR domain truncation event, such as EGFRvIII, and 2.2% (5/232) with an EGFR fusion event<sup>103</sup>. No definitive correlation has been identified between EGFR amplification and length of survival in patients with GBM<sup>104-105</sup>; however, EGFR amplification has been associated with prolonged survival in patients over the age of 60 with GBM<sup>106</sup>.

## FINDING SUMMARY

EGFR encodes the epidermal growth factor receptor, which belongs to a class of proteins called receptor tyrosine kinases. In response to signals from the environment, EGFR passes biochemical messages to the cell that stimulate it to grow and divide<sup>107</sup>. Amplification of EGFR has been associated with increased expression of EGFR mRNA and protein in several cancer types<sup>108-110</sup>. Disruption of the EGFR C-terminal region through deletion<sup>111-113</sup>, truncation<sup>111-112,114</sup>, splicing errors<sup>111,114</sup>, or gene fusion<sup>115-116</sup>, has been demonstrated to be activating and is likely to be oncogenic. These mutations have been shown to cause cellular transformation and tumor formation and to be sensitive to EGFR-targeting therapies, including erlotinib, lapatinib, and cetuximab<sup>111-113,115-116</sup>. One or more of the alterations observed here are predicted to be activating. EGFR mutations that have been characterized in biochemical assays to be activating, as observed here, are predicted to confer sensitivity

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GENOMIC FINDINGS

to EGFR-targeted therapies<sup>101,117-133</sup>. Although alterations such as seen here have not been fully characterized and are of unknown functional significance, similar alterations have been previously reported in the context of cancer, which

may indicate biological relevance.

**POTENTIAL DIAGNOSTIC IMPLICATIONS**

The presence of EGFR gene amplification or TERT

promoter mutations are indicative of diffuse astrocytic glioma with molecular features of glioblastoma, WHO grade 4 in IDH1/2-wildtype tumors (NCCN CNS Cancers Guidelines, v2.2022)<sup>134</sup>.

GENE

**MTAP**

ALTERATION

loss

**POTENTIAL TREATMENT STRATEGIES**

— Targeted Therapies —

MTAP inactivation produces specific metabolic vulnerabilities that may be sensitive to MAT2A<sup>135-136</sup> or PRMT5 inhibition<sup>136-138</sup>. A Phase 1 trial of MAT2A inhibitor AG-270 reported 1 PR and 2 SDs lasting longer than 6 months for patients with advanced solid tumors displaying MTAP loss<sup>139</sup>. Preclinical data suggest that MTAP loss sensitizes cells to S-adenosyl-L-methionine (SAM)-competitive PRMT5 inhibitors<sup>140</sup>, dual PRMT1 and PRMT5 inhibitors<sup>141-143</sup>, and PRMT5 inhibitors that selectively bind the PRMT5 when complexed with S-methyl-5'-thioadenosine (MTA), such as MRTX1719, TNG908, and AMG193<sup>144</sup>. In preclinical models, MTAP inactivation showed

increased sensitivity to inhibitors of purine synthesis or purine analogs, especially upon addition of exogenous MTA<sup>145-155</sup>. A Phase 2 study of L-alanosine, an inhibitor of adenine synthesis, as a monotherapy for 65 patients with MTAP-deficient cancers reported no responses and SD for 24% (13/55) of patients<sup>156</sup>. Preclinical and limited clinical evidence suggest MTAP deficiency may confer sensitivity to pemetrexed<sup>157</sup>.

**FREQUENCY & PROGNOSIS**

MTAP loss/homozygous deletion as well as loss of expression has been reported in a wide variety of solid tumors and hematologic cancers<sup>158-159</sup>; such events have been correlated with poor prognosis in a variety of cancer types, including hepatocellular carcinoma<sup>160</sup>, gastrointestinal stromal tumors<sup>161</sup>, mantle cell lymphoma (MCL)<sup>162</sup>, melanoma<sup>163-164</sup>, gastric cancer<sup>165</sup>, myxofibrosarcoma<sup>166</sup>, nasopharyngeal carcinoma<sup>167</sup>, ovarian carcinoma<sup>158</sup> and non-small cell lung cancer<sup>168</sup>. MTAP loss was not prognostic in pediatric B-cell acute lymphocytic leukemia<sup>169</sup> or in astrocytoma<sup>170</sup>. However, MTAP has also been reported to be

overexpressed in colorectal cancer (CRC) samples<sup>171</sup>, and MTAP retention is thought to be important for prostate cancer growth due to continuous supply of SAM<sup>172</sup>. Germline SNPs in MTAP have been correlated with the development of cutaneous melanoma<sup>173-174</sup>, esophageal cancer<sup>175-176</sup>, osteosarcoma<sup>177</sup>, and CRC<sup>178</sup>.

**FINDING SUMMARY**

MTAP encodes S-methyl-5'-thioadenosine (MTA) phosphorylase, a tumor suppressor involved in polyamine metabolism and methionine synthesis, although its enzymatic function is dispensable for its tumor suppressor activity<sup>179-180</sup>. Decreased expression of MTAP leads to MTA accumulation within tumor cells and their microenvironment<sup>160,181-182</sup>, thereby reducing intracellular arginine methylation<sup>136-138</sup> and altering cell signaling<sup>182-183</sup>. MTAP is located at 9p21, adjacent to CDKN2A and CDKN2B, with which it is frequently co-deleted in various cancers. Other alterations in MTAP are rare and have not been extensively characterized.

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GENOMIC FINDINGS

GENE  
**CDKN2A/B**

ALTERATION  
CDKN2A loss, CDKN2B loss

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Preclinical data suggest that tumors with loss of p16INK4a function may be sensitive to CDK4/6 inhibitors, such as abemaciclib, ribociclib, and palbociclib<sup>184-187</sup>. Clinical data in mesothelioma, breast cancer, and uterine leiomyosarcoma indicate that CDKN2A loss may predict sensitivity to abemaciclib<sup>188</sup> and palbociclib treatment<sup>189-190</sup>. However, multiple other clinical studies have shown no significant correlation between p16INK4a loss or inactivation and therapeutic benefit of these agents<sup>191-197</sup>; it is not known whether CDK4/6 inhibitors would be beneficial in this case. The p15INK4b protein encoded by CDKN2B is known to inhibit CDK4, and although concomitant loss of CDKN2A and CDKN2B may predict sensitivity to CDK4/6 inhibitors, such as ribociclib, abemaciclib, and palbociclib<sup>194-195,198-199</sup>, direct supporting data for CDKN2B alteration as a predictive biomarker for these therapies are limited<sup>200-201</sup>. Although preclinical studies have suggested that loss of p14ARF function may be associated with reduced sensitivity to MDM2 inhibitors<sup>202-203</sup>, the clinical relevance of p14ARF as a predictive biomarker is not clear.

FREQUENCY & PROGNOSIS

Concurrent putative homozygous deletion of CDKN2A and CDKN2B has been reported in 35% of patients with gliomas<sup>99</sup> and detected more frequently in patients with glioblastoma multiforme (GBM; 58%)<sup>98</sup> than in those with lower grade gliomas (6%)<sup>97</sup>. In other studies, loss of CDKN2A/B by deletion has been reported in up to 78% of astrocytomas (including anaplastic astrocytomas and GBM)<sup>204-206</sup>. A study found homozygous deletion of both p16INK4a and p14ARF in 26% (13/50) of glioblastomas (GBMs); 18% (9/50) of cases showed homozygous deletion of the p14ARF-encoding locus alone<sup>207</sup>. One study detected CDKN2A/B loss in 69% (161/232) and mutation in 2.6% (6/232) of IDH-wildtype GBM samples analyzed<sup>103</sup>. Decreased p14ARF and p16INK4a expression levels were found to be tightly associated in a study of glioma samples<sup>208</sup>. Homozygous deletion of the genomic region including CDKN2A and CDKN2B has been found to be associated with poor prognosis in glioblastoma (GBM) and likely serves as an early event in GBM progression<sup>205,209</sup>. In addition, expression of p16INK4a has been found to be lower in patients with high grade malignant gliomas compared with patients with low grade gliomas, and loss of p16INK4a expression has been associated with shorter OS in pilocytic astrocytomas<sup>210-211</sup>.

FINDING SUMMARY

CDKN2A encodes two different, unrelated tumor suppressor proteins, p16INK4a and p14ARF, whereas CDKN2B encodes the tumor suppressor

p15INK4b<sup>212-213</sup>. Both p15INK4b and p16INK4a bind to and inhibit CDK4 and CDK6, thereby maintaining the growth-suppressive activity of the Rb tumor suppressor; loss or inactivation of either p15INK4b or p16INK4a contributes to dysregulation of the CDK4/6-cyclin-Rb pathway and loss of cell cycle control<sup>214-215</sup>. The tumor suppressive functions of p14ARF involve stabilization and activation of p53, via a mechanism of MDM2 inhibition<sup>216-217</sup>. One or more alterations observed here are predicted to result in p16INK4a loss of function<sup>218-239</sup>. One or more alterations seen here are predicted to result in p14ARF loss of function<sup>222,239-242</sup>. CDKN2B alterations such as seen here are predicted to inactivate p15INK4b<sup>243</sup>.

POTENTIAL GERMLINE IMPLICATIONS

Germline CDKN2A mutation is associated with melanoma-pancreatic cancer syndrome, a condition marked by increased risk of developing malignant melanoma and/or pancreatic cancer<sup>244</sup>. Mutation carriers within families may develop either or both types of cancer, and melanoma cases may be referred to as familial or hereditary melanoma<sup>245-246</sup>. CDKN2A is the most implicated gene in familial melanoma, with germline mutations present in 16% to 20% of familial melanoma cases<sup>247-249</sup>. CDKN2A alteration has also been implicated in familial melanoma-astrocytoma syndrome, an extremely rare tumor association characterized by dual predisposition to melanoma and nervous system tumors<sup>250-252</sup>. In the appropriate clinical context, germline testing of CDKN2A is recommended.

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## GENOMIC FINDINGS

### GENE **RB1**

**ALTERATION**  
Q436\*, E237\*

**HGVS VARIANT**  
NM\_000321.2:c.1306C>T (p.Q436\*),  
NM\_000321.2:c.709G>T (p.E237\*)

**VARIANT CHROMOSOMAL POSITION**  
chr13:48951144, chr13:48934254

**VARIANT ALLELE FREQUENCY (% VAF)**  
23.0%, 23.6%

#### POTENTIAL TREATMENT STRATEGIES

##### — Targeted Therapies —

On the basis of limited clinical data<sup>253</sup> and strong preclinical data<sup>254-257</sup>, RB1 inactivation may be associated with sensitivity to inhibitors of Aurora kinase A, particularly in small cell lung cancer (SCLC). A clinical study evaluating the Aurora

kinase A inhibitor alisertib for patients with prostate cancer did not find an association between RB1 deletion and clinical benefit<sup>258</sup>. Other approaches to target RB1 inactivation under investigation in preclinical studies include inhibitors of BCL-2 family members<sup>259</sup> and activation of the NOTCH pathway<sup>260</sup>.

#### FREQUENCY & PROGNOSIS

In the TCGA datasets, RB1 mutation or homozygous deletion was observed in 9% of glioblastomas<sup>98</sup> and 2.5% of lower grade glioma cases<sup>261</sup>. In one study, loss of RB1 transcript expression was observed in 10.6% of glioblastomas and occurred more frequently in the proneural subtype<sup>262</sup>. The reports studying the association of RB1 mutations with survival are contrasting. One study reported that mutation of RB1 is correlated with shorter survival in glioblastoma patients<sup>263</sup>, whereas another study reported that the presence of RB1 mutations correlated with prolonged survival in glioblastoma patients<sup>264</sup>. Several studies

suggest that RB1, PTEN, and/or TP53 mutations are early events in the development of glioblastoma<sup>265-267</sup>.

#### FINDING SUMMARY

RB1 encodes the retinoblastoma protein (Rb), a tumor suppressor and negative regulator of the cell cycle<sup>268-269</sup>. Alterations such as seen here may disrupt RB1 function or expression<sup>270-276</sup>.

#### POTENTIAL GERMLINE IMPLICATIONS

Mutations in RB1 underlie the development of retinoblastoma (RB), a rare tumor that arises at a rate of approximately 1:20,000 live births, with nearly 5,000 new cases worldwide per year<sup>277</sup>. Germline mutations in RB1 account for approximately 40% of RB tumors<sup>278</sup> and are associated with an increased risk of developing secondary malignancies that include soft tissue and bone sarcoma and malignant melanoma<sup>279-280</sup>. In the appropriate clinical context, germline testing of RB1 is recommended.

### GENE **TERT**

**ALTERATION**  
promoter -124C>T

**HGVS VARIANT**  
NM\_198253.2:c.-124C>T

**VARIANT CHROMOSOMAL POSITION**  
chr5:1295228

**VARIANT ALLELE FREQUENCY (% VAF)**  
28.2%

#### POTENTIAL TREATMENT STRATEGIES

##### — Targeted Therapies —

Therapeutic options for targeting tumors with TERT mutations are limited, although a variety of approaches have been investigated, including immunotherapies using TERT as a tumor-associated antigen and antisense oligonucleotide- or peptide-based therapies. TERT peptide vaccines showed limited anticancer efficacy in clinical trials<sup>281</sup>; however, in one preclinical study, the combination of a TERT peptide vaccine and anti-CTLA-4 therapy suppressed tumor growth<sup>282</sup>. A Phase 2 study of the TERT inhibitor imetelstat for patients with advanced non-small cell lung cancer

reported no improvement in PFS or OS<sup>283</sup>.

#### FREQUENCY & PROGNOSIS

TERT promoter mutations have been reported in 51-59% of gliomas<sup>284-285</sup>, most frequently in glioblastoma (GBM, 54-84%), gliosarcoma (81%), oligodendroglioma (78%), and historically in oligoastrocytomas (25-31%) but less frequently in lower grade astrocytomas (10-18%) and in only 1% of ependymomas<sup>284-288</sup>. In patients with glioblastoma (GBM), the prevalence of TERT promoter mutation is lower in pediatric primary GBM (11%) and adult secondary GBM (28%) compared with adult primary GBM (58-83%)<sup>284,286</sup>. One study detected TERT promoter mutations in 78% (181/232) of IDH-wildtype GBM samples analyzed<sup>103</sup>. TERT promoter mutation has been shown to be significantly associated with increased TERT gene expression in astrocytoma, oligodendroglioma, and GBM<sup>289</sup>. TERT promoter mutations significantly associate with poor prognosis in patients with GBM, although this correlation may be due to the association with primary GBM as opposed to IDH-positive secondary GBM<sup>284,286,289-290</sup>. In the context of IDH-wildtype glioma, TERT mutations are associated with reduced OS (NCCN CNS Cancers Guidelines, v2.2022).

#### FINDING SUMMARY

Telomerase reverse transcriptase (TERT, or hTERT) is a catalytic subunit of the telomerase complex, which is required to maintain appropriate chromosomal length<sup>291</sup>. Activation of TERT is a hallmark of cancer, being detected in up to 80-90% of malignancies and absent in quiescent cells<sup>292-294</sup>. Mutations within the promoter region of TERT that confer enhanced TERT promoter activity have been reported in two hotspots, located at -124 bp and -146 bp upstream of the transcriptional start site (also termed C228T and C250T, respectively)<sup>295-297</sup>, as well as tandem mutations at positions -124/-125 bp and -138/-139 bp<sup>295</sup>.

#### POTENTIAL DIAGNOSTIC IMPLICATIONS

TERT mutations are associated with 1p/19q co-deletion in oligodendrogliomas, and are highly recurrent in IDH/ATRX-wildtype glioblastoma (GBM) (NCCN CNS Cancers Guidelines, v2.2022)<sup>298</sup>. The presence of EGFR gene amplification or TERT promoter mutations are indicative of diffuse astrocytic glioma with molecular features of glioblastoma, WHO grade 4 in IDH1/2-wildtype tumors (NCCN CNS Cancers Guidelines, v2.2022)<sup>134</sup>.

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**THERAPIES WITH CLINICAL BENEFIT**
**IN OTHER TUMOR TYPE**

## Cetuximab

*Assay findings association*

### EGFR

S229C, R222C - subclonal, amplification, rearrangement intron 24

### AREAS OF THERAPEUTIC USE

Cetuximab is a monoclonal antibody that targets EGFR. It is FDA approved for the treatment of head and neck squamous cell carcinoma (HNSCC) and KRAS-wild-type, EGFR-expressing metastatic colorectal cancer (CRC). Please see the drug label for full prescribing information.

### GENE ASSOCIATION

For patients with metastatic CRC receiving cetuximab or panitumumab as mono- or combination therapy, increased EGFR copy number associated with improved OS (HR=0.62) in a meta-analysis, although increased survival was not seen in populations that received first-line treatment with EGFR antibodies<sup>91</sup>.

### SUPPORTING DATA

A Phase 2 trial of cetuximab with the anti-VEGF monoclonal antibody bevacizumab for patients with glioblastoma (GBM) did not show improved efficacy compared with bevacizumab alone<sup>299</sup>. However, another Phase 2 study demonstrated that for patients with GBM harboring EGFR amplification but lacking expression of the EGFRvIII variant, treatment with cetuximab resulted in significantly longer PFS and numerical (although not statistically significant) improvement in OS<sup>300</sup>. In addition, a case report for an EGFR-amplified patient with GBM treated with cetuximab using intraarterial cerebral infusion (SIACI) in combination with chemotherapy reported a stable response without recurrence at 6 months<sup>301</sup>.

## Erlotinib

*Assay findings association*

### EGFR

S229C, R222C - subclonal, amplification, rearrangement intron 24

### AREAS OF THERAPEUTIC USE

Erlotinib is a small-molecule inhibitor of EGFR. It is FDA approved as a monotherapy or in combination with ramucirumab for patients with metastatic non-small cell lung cancer (NSCLC) harboring EGFR exon 19 deletions or exon 21 (L858R) mutations. Erlotinib is also FDA approved in combination with gemcitabine as a first-line treatment for advanced pancreatic cancer. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

Amplification or activation of EGFR may predict sensitivity to therapies such as erlotinib. For patients with activating mutations in EGFR, treatment with erlotinib has been associated with improved response and lengthened time to progression<sup>69,302-304</sup>. For patients with esophageal or biliary cancer treated with erlotinib or gefitinib, elevated EGFR copy number or amplification is associated with clinical responses and longer survival<sup>305-309</sup>. Responses to erlotinib have been reported for patients with EGFR rearrangements<sup>116,310-313</sup>.

### SUPPORTING DATA

In the MyPathway Phase 2a basket study for advanced

solid tumors, 1 of 9 patients with EGFR activating mutations responded to erlotinib monotherapy; the responding patient had urethral adenocarcinoma<sup>314</sup>. A patient with EGFR-mutated metastatic lacrimal gland adenoid cystic carcinoma experienced clinical benefit from erlotinib treatment that was ongoing at 14 months<sup>315</sup>. A clinical study of patients with glioblastoma (GBM) treated with gefitinib or erlotinib found that 9/49 (18%) had tumor shrinkage of 25% or more; in this study, the extracellular domain EGFRvIII mutation was correlated with response<sup>58</sup>. In a Phase 2 study of 65 patients with GBM or gliosarcoma, treatment with erlotinib, temozolomide, and radiotherapy resulted in longer PFS relative to a historical control study utilizing a regimen of temozolomide and radiotherapy alone (19.3 months vs. 14.1 months)<sup>316</sup>. However, in a Phase 1/2 trial of erlotinib monotherapy in 11 patients with relapsed or refractory GBM or anaplastic astrocytoma, all patients showed disease progression and the drug showed significant toxicity<sup>317</sup>. In addition, a Phase 2 trial of patients with recurrent or progressive GBM treated with erlotinib and sorafenib did not meet its objective of a 30% increase in OS compared with historical controls; sorafenib was found to increase erlotinib clearance<sup>318</sup>.

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THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

## Gefitinib

### Assay findings association

#### EGFR

S229C, R222C - subclonal, amplification, rearrangement intron 24

### AREAS OF THERAPEUTIC USE

Gefitinib targets the tyrosine kinase EGFR and is FDA approved to treat non-small cell lung cancer (NSCLC) harboring exon 19 deletions or exon 21 (L858R) substitution mutations in EGFR. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

Activation of EGFR may predict sensitivity to therapies such as gefitinib. Clinical studies have consistently shown significant improvement in response rates and PFS for patients with EGFR-mutated non-small cell lung cancer (NSCLC) treated with gefitinib compared with chemotherapy<sup>304,319-324</sup>, and responses have been reported for patients with EGFR-rearranged NSCLC<sup>311-312</sup>. For patients with esophageal or biliary cancer treated with erlotinib or gefitinib, elevated EGFR copy number or amplification is associated with clinical responses and longer survival<sup>305-309</sup>. Patients with refractory advanced esophageal carcinoma and EGFR amplification derived significant OS benefit from gefitinib compared with placebo (HR=0.21)<sup>305,325</sup>.

### SUPPORTING DATA

In a clinical study for patients with glioblastoma (GBM) treated with gefitinib or erlotinib, 24% (4/17) of patients treated with gefitinib experienced tumor shrinkage of 44-87%; furthermore, the extracellular domain EGFRvIII mutation was correlated with response<sup>58</sup>. Phase 2 studies of gefitinib for patients with high-grade glioma (HGG; including GBM, anaplastic astrocytoma, and oligodendroglioma)<sup>88</sup> or solely GBM<sup>326</sup> reported disease stabilization for 18% (5/28) and 42% (22/52) of patients, respectively; however, these and other Phase 1/2 studies reported that efficacy did not correlate with EGFR expression<sup>53,88,326-327</sup>. Similarly, a Phase 1/2 study of gefitinib plus radiotherapy for 178 patients with GBM reported no OS benefit of added gefitinib, and EGFR expression was found to be of no prognostic value<sup>89</sup>. However, Phase 1 and Phase 2 trials for HGG have reported clinical benefit from combining gefitinib with other modalities, including the mTOR inhibitors sirolimus (n=34; 2 PRs [5.9%], 13 SDs [38%])<sup>328</sup> and everolimus (n=22; 3 PRs [14%], 8 SDs [36%])<sup>327</sup> and the VEGF inhibitor cediranib (n=19; 8 PRs [42%])<sup>329</sup>.

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THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

## Osimertinib

### Assay findings association

#### EGFR

S229C, R222C - subclonal, amplification, rearrangement intron 24

### AREAS OF THERAPEUTIC USE

Osimertinib is an irreversible EGFR TKI that is selective for EGFR TKI-sensitizing mutations and the EGFR T790M mutation. It is FDA approved in various treatment settings for patients with non-small cell lung cancer (NSCLC) whose tumors have EGFR exon 19 deletions, exon 21 L858R mutations, or T790M mutations. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

EGFR TKI-sensitizing mutations or rearrangements and/or the EGFR T790M mutation may predict sensitivity to osimertinib in non-small cell lung cancer<sup>79,311,330-332</sup>.

EGFR mutations may confer sensitivity to osimertinib on the basis of clinical responses to the third-generation TKI osimertinib for patients with EGFR-mutated glioma<sup>66,85</sup> and additional clinical studies suggesting clinical benefit for these patients<sup>83,333</sup>. On the basis of clinical responses to the third-generation TKI osimertinib for patients with EGFR-rearranged glioma, EGFRvIII and activating rearrangements may confer sensitivity to osimertinib<sup>65,333</sup>. However, a case study of a patient with multiple glioblastoma tumors reported that the tumor harboring EGFRvIII and EGFR amplification did not respond to osimertinib<sup>66</sup>.

### SUPPORTING DATA

Clinical benefit from osimertinib has been observed for cases of pediatric and adult patients with EGFR-altered glioma<sup>65-66,83,85,333</sup>. Osimertinib has been studied primarily for the treatment of EGFR-mutated NSCLC. A Phase 2 study of osimertinib for EGFR-TKI-naïve patients with metastatic or recurrent NSCLC and uncommon EGFR mutations reported a 50% (18/36) ORR and an 89% (32/36) DCR with a median PFS of 8.2 months and a median duration of response of 11.2 months; patients harboring L861Q, G719X, or S768I mutations had ORRs of 78% (7/9), 53% (10/19), and 38% (3/8), respectively<sup>334</sup>. The Phase 3 FLAURA study reported that, relative to

erlotinib or gefitinib, first-line osimertinib significantly increased both median PFS (mPFS; 18.9 vs. 10.2 months, HR=0.46) and median OS (38.6 vs. 31.8 months; HR=0.80) for patients with advanced non-small cell lung cancer (NSCLC) and activating, sensitizing EGFR mutations (specifically, exon 19 deletion or L858R)<sup>330,335</sup>. In the Phase 3 ADAURA study, patients with early-stage (IB/II/IIIA) EGFR-mutated NSCLC experienced longer disease-free survival on osimertinib compared with placebo in the adjuvant setting (65.8 vs. 28.1 months, HR=0.27)<sup>336</sup>. A Phase 1 study reported that T790M-negative patients with acquired EGFR TKI resistance experienced an ORR of 21% and a median PFS of 2.8 months<sup>79</sup>. A Phase 1b/2 study evaluating osimertinib in combination with the CD73 inhibitor oleclumab for patients with advanced EGFR-mutated, T790M-negative NSCLC reported an ORR of 19% (4/21), a DCR of 81%, and mPFS of 11 months (Kim et al., 2021 AACR Abstract CT163). A Phase 2 trial of osimertinib in combination with bevacizumab versus osimertinib monotherapy for patients with untreated advanced non-small cell lung cancer (NSCLC) harboring EGFR del19 or L858R reported no difference in ORR (82% vs 86%) and median PFS (22.1 vs 20.2 months, HR 0.862 p=0.213)<sup>337</sup>. The Phase 2 BOOSTER study of osimertinib in combination with bevacizumab versus osimertinib monotherapy for patients with advanced NSCLC with EGFR-sensitizing mutations (exon 19 del or L858R) and L790M at progression on prior EGFR TKI reported no difference in ORR (55% vs 55%), median OS (24.0 vs 24.3 months, HR 1.03 p=0.91), or median PFS (15.4 vs 12.3 months, HR 0.96 p=0.83), although improved PFS was observed for the combination in the subgroup of current or former smokers (16.5 vs 8.4, HR 0.52) while nonsmokers had no benefit (HR 1.47)<sup>338</sup>. The Phase 1b TATTON study of osimertinib in combination with selumetinib, savolitinib, or durvalumab for patients with previously treated EGFR-mutated NSCLC reported ORRs of 42% (15/36), 44% (8/18), and 44% (10/23), respectively<sup>339</sup>.

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THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

## Panitumumab

*Assay findings association*

### EGFR

S229C, R222C - subclonal, amplification, rearrangement intron 24

### AREAS OF THERAPEUTIC USE

Panitumumab is a monoclonal antibody that targets EGFR. It is FDA approved to treat KRAS wild-type and NRAS wild-type metastatic colorectal cancer (CRC) combined with chemotherapy or as monotherapy for patients who have progressed on prior chemotherapy. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

For patients with metastatic CRC receiving cetuximab or panitumumab as mono- or combination therapy, increased EGFR copy number associated with improved OS (HR=0.62) in a meta-analysis, although increased survival was not seen in populations that received first-line treatment with EGFR antibodies<sup>91</sup>.

### SUPPORTING DATA

A Phase 1 trial of EnGeneIC delivery vehicle (EDV) targeting EGFR with panitumumab in combination with

doxorubicin for 14 patients with glioblastoma (GBM) reported no responses and 28% (4/14) SDs<sup>340</sup>.

Panitumumab has shown efficacy as monotherapy or in combination with chemotherapy for patients with KRAS-wildtype colorectal cancer<sup>341-343</sup> and has been investigated in a variety of other tumor types. For patients with head and neck squamous cell carcinoma (HNSCC), data are conflicting; some trials of panitumumab in various lines and with different chemotherapy combinations have shown modest benefit<sup>344-346</sup> and others have reported no benefit<sup>347-349</sup>. A Phase 3 study of chemotherapy with or without panitumumab for patients with advanced gastroesophageal cancer was terminated for futility<sup>350</sup>. Trials in a variety of tumor types have failed to show significant benefit for patients, including non-small cell lung cancer (NSCLC)<sup>351-352</sup>; biliary tract cancers, including cholangiocarcinoma<sup>353-354</sup>; and renal cell carcinoma (RCC)<sup>355</sup>.

**NOTE** Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

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CLINICAL TRIALS

**NOTE** Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually

updated and should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that

may require medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://clinicaltrials.gov). Or, visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

## GENE EGFR

### ALTERATION

S229C, R222C - subclonal, amplification, rearrangement intron 24

### RATIONALE

EGFR activating mutations, rearrangements, or amplification may predict sensitivity to EGFR-targeted therapies. Strategies to overcome

resistance to current agents include next-generation EGFR inhibitors and combination therapies.

## NCT03239015

### PHASE 2

Efficacy and Safety of Targeted Precision Therapy in Refractory Tumor With Druggable Molecular Event

### TARGETS

EGFR, ERBB4, ERBB2, PARP, mTOR, MET, ROS1, RET, VEGFRs, BRAF, CDK4, CDK6

**LOCATIONS:** Shanghai (China)

## NCT04946968

### PHASE 2

Phase-2 Dacomitinib Study on Patients With EGFR-Driven Advanced Solid Tumours With Low EGFR-AS1 lncRNA Expr or Other Novel Emerging Biomarkers

### TARGETS

ERBB4, EGFR, ERBB2

**LOCATIONS:** Singapore (Singapore)

## NCT04720976

### PHASE 1/2

JAB-3312 Activity in Adult Patients With Advanced Solid Tumors

### TARGETS

MEK, SHP2, PD-1, EGFR, KRAS

**LOCATIONS:** Utah, California, Arizona, Minnesota, Illinois, Michigan, Oklahoma, Missouri, Indiana, Connecticut

## NCT04670679

### PHASE 1

A Dose Escalation/Expansion Study of ERAS-601 in Patients With Advanced or Metastatic Solid Tumors

### TARGETS

SHP2, EGFR

**LOCATIONS:** Perth (Australia), Melbourne (Australia), Nevada, California, Missouri, Texas, Massachusetts, New York, Pennsylvania, Tennessee

## NCT02800486

### PHASE 2

Super Selective Intra-arterial Repeated Infusion of Cetuximab (Erbix) With Reirradiation for Treatment of Relapsed/Refractory GBM, AA, and AOA

### TARGETS

EGFR

**LOCATIONS:** New York

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CLINICAL TRIALS

**NCT02861898**

**PHASE 1/2**

Super-selective Intra-arterial Repeated Infusion of Cetuximab for the Treatment of Newly Diagnosed Glioblastoma

**TARGETS**  
**EGFR**

**LOCATIONS:** New York

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**ORDERED TEST #** ORD-1651461-01

**CLINICAL TRIALS**
**GENE**
**MTAP**
**ALTERATION**

loss

**RATIONALE**

MTAP loss may predict sensitivity to MAT2A inhibitors, or to inhibitors that target PRMT5 when in complex with MTA.

**NCT05245500**
**PHASE 1/2**

Phase 1/2 Study of MRTX1719 in Solid Tumors With MTAP Deletion

**TARGETS**

PRMT5-MTA

**LOCATIONS:** Colorado, Arizona, Minnesota, Massachusetts, New York, Tennessee, Texas, Florida

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**APPENDIX**
**Variants of Unknown Significance**

**NOTE** One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

**BRD4**

 NM\_058243.2: c.3035C>T  
(p.P1012L)  
chr19:15353845

**GNAS**

 NM\_080425.2: c.11G>A  
(p.R4H)  
chr20:57428331

**SYK**

 NM\_003177.5: c.793C>G  
(p.Q265E)  
chr9:93626946

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ORDERED TEST # ORD-1651461-01

**APPENDIX**
**Genes Assayed in FoundationOne®CDx**

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

**DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS**

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B or WTX)	
APC	AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX
AURKA	AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2
BCL6	BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1
BTG2	BTK	CALR	CARD11	CASP8	CBFB	CBL	CCND1	CCND2
CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73	CDH1
CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B	CDKN2C
CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R	CTCF
CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1	DDR2
DIS3	DNMT3A	DOT1L	EED	EGFR	EMSY (C11orf30)	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12	FGF14
FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3	FGFR4
FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3	GATA4
GATA6	GID4 (C17orf39)	GNA11	GNA13	GNAS	GNAS	GRM3	GSK3B	H3-3A (H3F3A)
HDAC1	HGF	HNFA1	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDM5A	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11 (MRE11A)	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD2 (WHSC1 or MMSET)	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3	NTRK3
P2RY8	PALB2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)
PDGFRA	PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1
PMS2	POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI
PRKN (PARK2)	PTCH1	PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51
RAD51B	RAD51C	RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10
REL	RET	RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC
SDHD	SETD2	SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO
SNCAIP	SOC1	SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3
STK11	SUFU	SYK	TBX3	TEK	TENT5C (FAM46C)	TET2	TET2	TGFB2
TIPARP	TNFAIP3	TNFRSF14	TP53	TSC1	TSC2	TYRO3	U2AF1	VEGFA
VHL	WT1	XPO1	XRCC2	ZNF217	ZNF703			

**DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS**

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSP02	SDC4	SLC34A2	TERC*	TERT**	TPR2S2

\*TERC is an NCRNA

\*\*Promoter region of TERT is interrogated

**ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS**


Homologous Recombination status  
Loss of Heterozygosity (LOH) score  
Microsatellite (MS) status  
Tumor Mutational Burden (TMB)

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## APPENDIX

## About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a., Ciplstraat 3, 2440 Geel, Belgium. 

## ABOUT FOUNDATIONONE CDx

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:  
[www.rochefoundationmedicine.com/f1cdxtech](http://www.rochefoundationmedicine.com/f1cdxtech).

## INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

## TEST PRINCIPLE

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes

detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

## THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

### Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

### Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for *ERBB2* and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical

methodology has identified as being present in <10% of the assayed tumor DNA.

## Ranking of Therapies and Clinical Trials

*Ranking of Therapies in Summary Table*  
Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

### Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

## NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) ([www.nccn.org](http://www.nccn.org)). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2023. All rights reserved. To view the most recent and complete version of the guidelines, go online to [NCCN.org](http://NCCN.org). NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

## Limitations

1. In the fraction-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by

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About FoundationOne®CDx

- analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.
- TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation [https://www.accessdata.fda.gov/cdrh\\_docs/pdf17/P170019B.pdf](https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf). The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
  - Homologous Recombination status may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas (Coleman et al., 2017; 28916367). Samples with deleterious *BRCA1/2* alteration and/or Loss of Heterozygosity (LOH) score  $\geq 16$  will be reported as "HRD Positive" and samples with absence of these findings will be reported as "HRD Not Detected," agnostic of potential secondary *BRCA1/2* reversion alterations. Certain potentially deleterious missense or small in-frame deletions in *BRCA1/2* may not be classified as deleterious and, in the absence of an elevated LOH profile, samples with such mutations may be classified as "HRD Not Detected." A result of "HRD Not Detected" does not rule out the presence of a *BRCA1/2* alteration or an elevated LOH profile outside the assay performance characteristic limitations.
  - The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and

extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.

- Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
- Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine whether the patient is a candidate for biopsy.
- Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of HER2 overexpression as 20% in breast cancer. Based on the F1CDx HER2 CDx concordance study, approximately 10% of HER2 amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

## REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal

hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

## VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

\*Interquartile Range = 1st Quartile to 3rd Quartile

## VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's

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**APPENDIX**

About FoundationOne®CDx

tumor sequencing is germline or somatic.  
Interpretation should be based on clinical context.

**VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS**

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D (MLL2)*, *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

**LEVEL OF EVIDENCE NOT PROVIDED**

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

**NO GUARANTEE OF CLINICAL BENEFIT**

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

**NO GUARANTEE OF REIMBURSEMENT**

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

**TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN**

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking

into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

**SELECT ABBREVIATIONS**

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

**REFERENCE SEQUENCE INFORMATION**

Sequence data is mapped to the human genome, Genome Reference Consortium Human Build 37 (GRCh37), also known as hg19.

MR Suite Version (RG) 7.9.0

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**APPENDIX** **References**

1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) PMID: 25392179
2. Kroemer G, et al. Oncoimmunology (2015) PMID: 26140250
3. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
4. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
5. Ayers et al., 2016; ASCO-SITC Abstract P60
6. Martinez R, et al. Oncology (2004) PMID: 15331927
7. Martinez R, et al. J. Cancer Res. Clin. Oncol. (2005) PMID: 15672285
8. Martinez R, et al. Cancer Genet. Cytogenet. (2007) PMID: 17498554
9. Szybka M, et al. Clin. Neuropathol. (2010) PMID: 12908754
10. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
11. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
12. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
13. Boland CR, et al. Cancer Res. (1998) PMID: 9823339
14. Pawlik TM, et al. Dis. Markers (2004) PMID: 15528785
15. Boland CR, et al. Gastroenterology (2010) PMID: 20420947
16. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
17. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
18. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
19. Cristescu R, et al. Science (2018) PMID: 30309915
20. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
21. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
22. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
23. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
24. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
25. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
26. Zhao J, et al. Nat. Med. (2019) PMID: 30742119
27. Touat M, et al. Nature (2020) PMID: 32322066
28. Bouffett E, et al. J. Clin. Oncol. (2016) PMID: 27001570
29. Johanns TM, et al. Cancer Discov (2016) PMID: 27683556
30. Lukas RV, et al. J. Neurooncol. (2018) PMID: 30073642
31. Chalmers ZR, et al. Genome Med (2017) PMID: 28420421
32. Patel RR, et al. Pediatr Blood Cancer (2020) PMID: 32386112
33. Johnson A, et al. Oncologist (2017) PMID: 28912153
34. Draaisma K, et al. Acta Neuropathol Commun (2015) PMID: 26699864
35. Wang L, et al. BMC Cancer (2020) PMID: 32164609
36. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
37. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
38. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
39. Rizvi NA, et al. Science (2015) PMID: 25765070
40. Johnson BE, et al. Science (2014) PMID: 24336570
41. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
42. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
43. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
44. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
45. Nature (2012) PMID: 22810696
46. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
47. van den Bent MJ, et al. J Clin Oncol (2009) PMID: 19204207
48. Haas-Kogan DA, et al. J Natl Cancer Inst (2005) PMID: 15956649
49. Brown PD, et al. J Clin Oncol (2008) PMID: 18955445
50. Preusser M, et al. J Neurooncol (2008) PMID: 18458820
51. Wen PY, et al. Neuro-oncology (2014) PMID: 24470557
52. Gallego O, et al. J Neurooncol (2014) PMID: 24352766
53. Uhm JH, et al. Int J Radiat Oncol Biol Phys (2011) PMID: 20510539
54. Doyle SP, et al. Oxf Med Case Reports (2018) PMID: 30410775
55. D'Alessandris QG, et al. Acta Neurochir (Wien) (2013) PMID: 23132371
56. Custodio A, et al. Clin Transl Oncol (2010) PMID: 20462843
57. D'Alessandris QG, et al. Acta Neurochir (Wien) (2018) PMID: 30306271
58. Mellinghoff IK, et al. N. Engl. J. Med. (2005) PMID: 16282176
59. Mellinghoff IK, et al. Clin Cancer Res (2007) PMID: 17255257
60. Arif SH, et al. Asian J Neurosurg (2019) PMID: 29492119
61. Reardon DA, et al. Neuro-oncology (2015) PMID: 25140039
62. Alshami J, et al. Oncotarget (2015) PMID: 26423602
63. Sepúlveda-Sánchez JM, et al. Neuro-oncology (2017) PMID: 28575464
64. Chi AS, et al. JCO Precis Oncol (2020) PMID: 32923886
65. Cardona AF, et al. J Neurooncol (2021) PMID: 34498213
66. Makhlin I, et al. CNS Oncol (2019) PMID: 31769726
67. Chagoya G, et al. Oncotarget (2020) PMID: 32547705
68. Gao M, et al. Cancer Res (2021) PMID: 33910930
69. Rosell R, et al. Lancet Oncol. (2012) PMID: 22285168
70. Douillard JY, et al. Br. J. Cancer (2014) PMID: 24263064
71. Hayashi T, et al. Hum Pathol (2020) PMID: 32673682
72. Cao L, et al. Onco Targets Ther (2018) PMID: 29780256
73. Yang TY, et al. J. Clin. Oncol. (2011) PMID: 21422421
74. Sequist LV, et al. J. Clin. Oncol. (2013) PMID: 23816960
75. Qin BD, et al. Onco Targets Ther (2018) PMID: 30127622
76. Frega S, et al. J Thorac Oncol (2016) PMID: 27131295
77. Long X, et al. Onco Targets Ther (2020) PMID: 33116645
78. Mok TS, et al. J. Clin. Oncol. (2018) PMID: 29864379
79. Jänne PA, et al. N. Engl. J. Med. (2015) PMID: 25923549
80. Hong MH, et al. Cancer (2020) PMID: 32749686
81. Kim HS, et al. Oncotarget (2015) PMID: 26462025
82. Kim HS, et al. Clin. Cancer Res. (2015) PMID: 25424851
83. Mondal G, et al. Acta Neuropathol (2020) PMID: 32303840
84. Cavalieri S, et al. Eur. J. Cancer (2018) PMID: 29734047
85. Goyal A, et al. World Neurosurg (2021) PMID: 33940677
86. Tanaka S, et al. Sci Rep (2019) PMID: 30644426
87. Blumenthal DT, et al. J. Neurooncol. (2016) PMID: 27531351
88. Franceschi E, et al. Br. J. Cancer (2007) PMID: 17353924
89. Chakravarti A, et al. Int. J. Radiat. Oncol. Biol. Phys. (2013) PMID: 23182702
90. Hegi ME, et al. Mol. Cancer Ther. (2011) PMID: 21471286
91. Jiang Z, et al. PLoS ONE (2013) PMID: 23441167
92. Licitra L, et al. Ann. Oncol. (2011) PMID: 21048039
93. Smyth EC, et al. Gut (2021) PMID: 33199443
94. Lassman et al., 2019; Neuro-Oncology Abstract ACTR-21
95. Van Den Bent M, et al. Neuro Oncol (2020) PMID: 31747009
96. Byeon S, et al. Target Oncol (2020) PMID: 32107712
97. Jonsson P, et al. Clin. Cancer Res. (2019) PMID: 31263031
98. Brennan CW, et al. Cell (2013) PMID: 24120142
99. Ceccarelli M, et al. Cell (2016) PMID: 26824661
100. Thomas AA, et al. Neuro-oncology (2017) PMID: 28472509
101. Lee JC, et al. PLoS Med. (2006) PMID: 17177598
102. Vivanco I, et al. Cancer Discov (2012) PMID: 22588883
103. Yan et al. 2020; DOI:10.1200/PO.19.00385
104. Srividya MR, et al. J. Clin. Pathol. (2010) PMID: 20702468
105. Das P, et al. J Clin Neurosci (2011) PMID: 20888234
106. Smith JS, et al. J. Natl. Cancer Inst. (2001) PMID: 11504770
107. Ciardiello F, et al. N. Engl. J. Med. (2008) PMID: 18337605
108. Liang Z, et al. BMC Cancer (2010) PMID: 20637128
109. Bhargava R, et al. Mod. Pathol. (2005) PMID: 15920544
110. Yang YL, et al. Chin. Med. J. (2012) PMID: 22490401
111. Cho J, et al. Cancer Res. (2011) PMID: 22001862
112. Imielinski M, et al. Cell (2012) PMID: 22980975
113. Pines G, et al. Oncogene (2010) PMID: 20676128
114. Wang Q, et al. Exp. Cell Res. (2007) PMID: 17643422
115. Frattini V, et al. Nat. Genet. (2013) PMID: 23917401
116. Konduri K, et al. Cancer Discov (2016) PMID: 27102076
117. Foster JM, et al. World J Surg Oncol (2010) PMID: 20942962
118. Cai CQ, et al. Oncogene (2008) PMID: 18193092
119. Stabile LP, et al. Cancer Res. (2005) PMID: 15735034
120. Zhang JW, et al. J Thorac Oncol (2006) PMID: 17409930
121. Siegfried JM, et al. J Thorac Oncol (2012) PMID: 22258476
122. U M, et al. PLoS Comput. Biol. (2014) PMID: 24743239
123. Cho J, et al. Mol. Cancer (2014) PMID: 24894453
124. Hama T, et al. Oncologist (2009) PMID: 19726454
125. Tam IY, et al. Mol. Cancer Ther. (2009) PMID: 19671738
126. Kancha RK, et al. Clin. Cancer Res. (2009) PMID: 19147750
127. Chen YR, et al. Oncogene (2006) PMID: 16205628
128. Ymer SI, et al. Cancers (Basel) (2011) PMID: 24212795
129. Razis E, et al. Clin. Cancer Res. (2009) PMID: 19789313
130. Wang H, et al. Neoplasia (2011) PMID: 21532887
131. Kim N, et al. Int. J. Cancer (2019) PMID: 31290142
132. Sueaengoen N, et al. Cell Biosci (2020) PMID: 32190291
133. Lundby A, et al. Cell (2019) PMID: 31585087
134. Louis DN, et al. Neuro Oncol (2021) PMID: 34185076
135. Kalev P, et al. Cancer Cell (2021) PMID: 33450196
136. Marjon K, et al. Cell Rep (2016) PMID: 27068473
137. Mavrikis KJ, et al. Science (2016) PMID: 26912361
138. Kryukov GV, et al. Science (2016) PMID: 26912360
139. Heist et al., 2019; AACR-NCI-EORTC Abstract B116
140. Guccione E, et al. Nat. Rev. Mol. Cell Biol. (2019) PMID: 31350521
141. Fedoriv A, et al. Cancer Cell (2019) PMID: 31257072
142. Srour N, et al. Cancer Cell (2019) PMID: 31287990
143. Gao G, et al. Nucleic Acids Res. (2019) PMID: 30916320
144. Smith CR, et al. J Med Chem (2022) PMID: 35041419
145. Hansen LJ, et al. Cancer Res. (2019) PMID: 31040154
146. Tang B, et al. Cancer Res. (2018) PMID: 29844120
147. Munshi PN, et al. Oncologist (2014) PMID: 24928612
148. de Oliveira SF, et al. PLoS ONE (2016) PMID: 26751376
149. Lubin M, et al. PLoS ONE (2009) PMID: 19478948
150. Tang B, et al. Cancer Biol. Ther. (2012) PMID: 22825330
151. Collins CC, et al. Mol. Cancer Ther. (2012) PMID: 22252602
152. Bertino JR, et al. Cancer Biol. Ther. (2011) PMID: 21301207
153. Coulthard SA, et al. Mol. Cancer Ther. (2011) PMID: 21282358

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ORDERED TEST # ORD-1651461-01

**APPENDIX**
**References**

154. Miyazaki S, et al. *Int. J. Oncol.* (2007) PMID: 17912432
155. Efferth T, et al. *Blood Cells Mol. Dis.* ( ) PMID: 11987241
156. Kindler HL, et al. *Invest New Drugs* (2009) PMID: 18618081
157. Alhalabi O, et al. *Nat Commun* (2022) PMID: 35379845
158. Wei R, et al. *Sci Rep* (2016) PMID: 27929028
159. Zhao M, et al. *BMC Genomics* (2016) PMID: 27556634
160. Kirovski G, et al. *Am. J. Pathol.* (2011) PMID: 21356366
161. Huang HY, et al. *Clin. Cancer Res.* (2009) PMID: 19887491
162. Marcé S, et al. *Clin. Cancer Res.* (2006) PMID: 16778103
163. Meyer S, et al. *Exp. Dermatol.* (2010) PMID: 20500769
164. Wild PJ, et al. *Arch Dermatol* (2006) PMID: 16618867
165. Kim J, et al. *Genes Chromosomes Cancer* (2011) PMID: 21412930
166. Li CF, et al. *Oncotarget* (2014) PMID: 25426549
167. He HL, et al. *Medicine (Baltimore)* (2015) PMID: 26656376
168. Su CY, et al. *Eur J Surg Oncol* (2014) PMID: 24969958
169. Mirebeau D, et al. *Haematologica* (2006) PMID: 16818274
170. Becker AP, et al. *Pathobiology* (2015) PMID: 26088413
171. Snezhkina AV, et al. *Oxid Med Cell Longev* (2016) PMID: 27433286
172. Bistulfi G, et al. *Oncotarget* (2016) PMID: 26910893
173. Antonopoulou K, et al. *J. Invest. Dermatol.* (2015) PMID: 25407435
174. Maccioni L, et al. *BMC Cancer* (2013) PMID: 23816148
175. Hyland PL, et al. *Int J Epidemiol* (2016) PMID: 26635288
176. Lin X, et al. *Cancer Sci.* (2017) PMID: 27960044
177. Zhi L, et al. *J Cancer* (2016) PMID: 27994653
178. Gu F, et al. *Br. J. Cancer* (2013) PMID: 23361049
179. Limm K, et al. *PLoS ONE* (2016) PMID: 27479139
180. Tang B, et al. *G3 (Bethesda)* (2014) PMID: 25387827
181. Limm K, et al. *Eur. J. Cancer* (2013) PMID: 23265702
182. Stevens AP, et al. *J. Cell. Biochem.* (2009) PMID: 19097084
183. Limm K, et al. *Eur. J. Cancer* (2014) PMID: 25087184
184. Konecny GE, et al. *Clin. Cancer Res.* (2011) PMID: 21278246
185. Katsumi Y, et al. *Biochem. Biophys. Res. Commun.* (2011) PMID: 21871868
186. Cen L, et al. *Neuro-oncology* (2012) PMID: 22711607
187. Logan JE, et al. *Anticancer Res.* (2013) PMID: 23898052
188. Fennell DA, et al. *Lancet Oncol* (2022) PMID: 35157829
189. Elvin JA, et al. *Oncologist* (2017) PMID: 28283584
190. Gao J, et al. *Curr Oncol* (2015) PMID: 26715889
191. Gopalan et al., 2016; ASCO Abstract 8077
192. Peguero et al., 2016; ASCO Abstract 2528
193. Konecny et al., 2016; ASCO Abstract 5557
194. DeMichele A, et al. *Clin. Cancer Res.* (2015) PMID: 25501126
195. Finn RS, et al. *Lancet Oncol.* (2015) PMID: 25524798
196. Infante JR, et al. *Clin. Cancer Res.* (2016) PMID: 27542767
197. Johnson DB, et al. *Oncologist* (2014) PMID: 24797823
198. Flaherty KT, et al. *Clin. Cancer Res.* (2012) PMID: 22090362
199. Dickson MA, et al. *J. Clin. Oncol.* (2013) PMID: 23569312
200. Su D, et al. *Nat Commun* (2019) PMID: 31700061
201. Tramontana TF, et al. *JCO Precis Oncol* (2020) PMID: 32923894
202. Van Maerken T, et al. *Mol. Cancer Ther.* (2011) PMID: 21460101
203. Gamble LD, et al. *Oncogene* (2012) PMID: 21725357
204. Verhaak RG, et al. *Cancer Cell* (2010) PMID: 20129251
205. Sottoriva A, et al. *Proc. Natl. Acad. Sci. U.S.A.* (2013) PMID: 23412337
206. Weber RG, et al. *Oncogene* (2007) PMID: 16909113
207. Nakamura M, et al. *Brain Pathol.* (2001) PMID: 11303791
208. Chakravarti A, et al. *Clin. Cancer Res.* (2001) PMID: 11489817
209. Feng J, et al. *Cancer* (2012) PMID: 21713760
210. Raabe EH, et al. *Clin. Cancer Res.* (2011) PMID: 21636552
211. Liu W, et al. *J. Exp. Clin. Cancer Res.* (2011) PMID: 21843312
212. Quelle DE, et al. *Cell* (1995) PMID: 8521522
213. Mutat. Res. (2005) PMID: 15878778
214. Gazzeri S, et al. *Oncogene* (1998) PMID: 9484839
215. *Oncogene* (1999) PMID: 10498883
216. Sherr CJ, et al. *Cold Spring Harb. Symp. Quant. Biol.* (2005) PMID: 16869746
217. Ozenne P, et al. *Int. J. Cancer* (2010) PMID: 20549699
218. Ruas M, et al. *Oncogene* (1999) PMID: 10498896
219. Jones R, et al. *Cancer Res.* (2007) PMID: 17909018
220. Haferkamp S, et al. *Aging Cell* (2008) PMID: 18843795
221. Huot TJ, et al. *Mol. Cell. Biol.* (2002) PMID: 12417717
222. Rizo H, et al. *J. Biol. Chem.* (2001) PMID: 11518711
223. Gombart AF, et al. *Leukemia* (1997) PMID: 9324288
224. Yang R, et al. *Cancer Res.* (1995) PMID: 7780957
225. Parry D, et al. *Mol. Cell. Biol.* (1996) PMID: 8668202
226. Greenblatt MS, et al. *Oncogene* (2003) PMID: 12606942
227. Yarbrough WG, et al. *J. Natl. Cancer Inst.* (1999) PMID: 10491434
228. Poi MJ, et al. *Mol. Carcinog.* (2001) PMID: 11255261
229. Byeon IJ, et al. *Mol. Cell* (1998) PMID: 9660926
230. Kannengiesser C, et al. *Hum. Mutat.* (2009) PMID: 19260062
231. Lal G, et al. *Genes Chromosomes Cancer* (2000) PMID: 10719365
232. Koh J, et al. *Nature* (1995) PMID: 7777061
233. McKenzie HA, et al. *Hum. Mutat.* (2010) PMID: 20340136
234. Miller PJ, et al. *Hum. Mutat.* (2011) PMID: 21462282
235. Kutscher CL, et al. *Physiol. Behav.* (1977) PMID: 905385
236. Scaini MC, et al. *Hum. Mutat.* (2014) PMID: 24659262
237. Jenkins NC, et al. *J. Invest. Dermatol.* (2013) PMID: 23190892
238. Walker GJ, et al. *Int. J. Cancer* (1999) PMID: 10389768
239. Rutter JL, et al. *Oncogene* (2003) PMID: 12853981
240. Itahana K, et al. *Cancer Cell* (2008) PMID: 18538737
241. Zhang Y, et al. *Mol. Cell* (1999) PMID: 10360174
242. Zhang Y, et al. *Cell* (1998) PMID: 9529249
243. Jafri M, et al. *Cancer Discov* (2015) PMID: 25873077
244. Whelan AJ, et al. *N Engl J Med* (1995) PMID: 7666917
245. *Adv Exp Med Biol* (2010) PMID: 20687502
246. Hogg D, et al. *J Cutan Med Surg* (1998) PMID: 9479083
247. De Unamuno B, et al. *Melanoma Res* (2018) PMID: 29543703
248. Soura E, et al. *J Am Acad Dermatol* (2016) PMID: 26892650
249. Huerta C, et al. *Acta Derm Venereol* (2018) PMID: 29405243
250. Kaufman DK, et al. *Neurology* (1993) PMID: 8414022
251. Bahua M, et al. *Cancer Res* (1998) PMID: 9622062
252. Chan AK, et al. *Clin Neuropathol* ( ) PMID: 28699883
253. Owonikoko et al., 2016; ESMO Abstract 14230
254. Hook KE, et al. *Mol. Cancer Ther.* (2012) PMID: 22222631
255. Gong X, et al. *Cancer Discov* (2019) PMID: 30373917
256. Oser MG, et al. *Cancer Discov* (2019) PMID: 30373918
257. Yang W, et al. *Kaohsiung J Med Sci* (2022) PMID: 34741392
258. Beltran H, et al. *Clin. Cancer Res.* (2019) PMID: 30232224
259. Allaman-Pillet N, et al. *Ophthalmic Genet.* ( ) PMID: 21955141
260. Viatour P, et al. *J. Exp. Med.* (2011) PMID: 21875955
261. Cancer Genome Atlas Research Network, et al. *N. Engl. J. Med.* (2015) PMID: 26061751
262. Goldhoff P, et al. *J. Neuropathol. Exp. Neurol.* (2012) PMID: 22157621
263. Bäcklund LM, et al. *Clin. Cancer Res.* (2003) PMID: 14519639
264. Dono A, et al. *Neurooncol Adv* (2021) PMID: 34131647
265. Chow LM, et al. *Cancer Cell* (2011) PMID: 21397855
266. Nobusawa S, et al. *Brain Pathol.* (2010) PMID: 20406234
267. Deighton RF, et al. *Brain Pathol.* (2010) PMID: 20175778
268. Burkhardt DL, et al. *Nat. Rev. Cancer* (2008) PMID: 18650841
269. Knudsen ES, et al. *Nat. Rev. Cancer* (2008) PMID: 19143056
270. Berge EO, et al. *Mol. Cancer* (2010) PMID: 20594292
271. Giacinti C, et al. *Oncogene* (2006) PMID: 16936740
272. Otterson GA, et al. *Proc. Natl. Acad. Sci. U.S.A.* (1997) PMID: 9342358
273. Otterson GA, et al. *Am. J. Hum. Genet.* (1999) PMID: 10486322
274. Qin XQ, et al. *Genes Dev.* (1992) PMID: 1534305
275. Rubin SM, et al. *Cell* (2005) PMID: 16360038
276. Sun H, et al. *Mol. Cell. Biol.* (2006) PMID: 16449662
277. Chen Z, et al. *Hum. Mutat.* (2014) PMID: 24282159
278. Yun J, et al. *Int J Ophthalmol* (2011) PMID: 22553621
279. Houston SK, et al. *Int Ophthalmol Clin* (2011) PMID: 21139478
280. Ng AK, et al. *Semin Radiat Oncol* (2010) PMID: 19959033
281. *Nat Rev Clin Oncol* (2017) PMID: 27245281
282. Duperret EK, et al. *Mol Ther* (2018) PMID: 29249395
283. Chiappori AA, et al. *Ann Oncol* (2015) PMID: 25467017
284. Killela PJ, et al. *Proc. Natl. Acad. Sci. U.S.A.* (2013) PMID: 23530248
285. Killela PJ, et al. *Oncotarget* (2014) PMID: 24722048
286. Nonoguchi N, et al. *Acta Neuropathol.* (2013) PMID: 23955565
287. Liu X, et al. *Cell Cycle* (2013) PMID: 23603989
288. Koelsche C, et al. *Acta Neuropathol.* (2013) PMID: 24154961
289. Arita H, et al. *Acta Neuropathol.* (2013) PMID: 23764841
290. Reitman ZJ, et al. *Acta Neuropathol.* (2013) PMID: 24217890
291. Shay JW, et al. *Semin. Cancer Biol.* (2011) PMID: 22015685
292. Shay JW, et al. *Eur. J. Cancer* (1997) PMID: 9282118
293. Kim NW, et al. *Science* (1994) PMID: 7605428
294. Hanahan D, et al. *Cell* (2000) PMID: 10647931
295. Horn S, et al. *Science* (2013) PMID: 23348503
296. Huang FW, et al. *Science* (2013) PMID: 23348506
297. Vinagre J, et al. *Nat Commun* (2013) PMID: 23887589
298. Weller M, et al. *Nat Rev Clin Oncol* (2021) PMID: 33293629
299. Hasselbalch B, et al. *Neuro-oncology* (2010) PMID: 20406901
300. Lv S, et al. *Int. J. Oncol.* (2012) PMID: 22752145
301. Kulason KO, et al. *J Exp Ther Oncol* (2018) PMID: 29790314
302. Cappuzzo F, et al. *Lancet Oncol.* (2010) PMID: 20493771
303. Zhong WZ, et al. *J. Clin. Oncol.* (2019) PMID: 31194613
304. Petrelli F, et al. *Clin Lung Cancer* (2012) PMID: 222027531

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ORDERED TEST # ORD-1651461-01

**APPENDIX**
**References**

- 22056888
305. Petty RD, et al. J. Clin. Oncol. (2017) PMID: 28537764
306. Philip PA, et al. J. Clin. Oncol. (2006) PMID: 16809731
307. Xie C, et al. Br J Cancer (2020) PMID: 32958820
308. Luo H, et al. JAMA Netw Open (2020) PMID: 33026449
309. Lee J, et al. Lancet Oncol. (2012) PMID: 22192731
310. Zhu YC, et al. Lung Cancer (2018) PMID: 29290255
311. Wang J, et al. Int. J. Cancer (2019) PMID: 30255937
312. Baik CS, et al. J Thorac Oncol (2015) PMID: 26398831
313. Xu et al., 2018; ASCO Abstract e13538
314. Hainsworth JD, et al. J. Clin. Oncol. (2018) PMID: 29320312
315. Nie KK, et al. Chin Med J (Engl) (2018) PMID: 29998897
316. Prados MD, et al. J. Clin. Oncol. (2009) PMID: 19075262
317. Kesavabhotla K, et al. J. Exp. Ther. Oncol. (2012) PMID: 22946346
318. Peereboom DM, et al. Neuro-oncology (2013) PMID: 23328813
319. Han JY, et al. J. Clin. Oncol. (2012) PMID: 22370314
320. Maemondo M, et al. N. Engl. J. Med. (2010) PMID: 20573926
321. Mitsudomi T, et al. Lancet Oncol. (2010) PMID: 20022809
322. Mok TS, et al. N. Engl. J. Med. (2009) PMID: 19692680
323. Qi WX, et al. Curr Med Res Opin (2015) PMID: 25329826
324. Zhao H, et al. J Thorac Oncol (2015) PMID: 25546556
325. Dutton SJ, et al. Lancet Oncol. (2014) PMID: 24950987
326. Rich JN, et al. J Clin Oncol (2004) PMID: 14638850
327. Kreisl TN, et al. J. Neurooncol. (2009) PMID: 19018475
328. Reardon DA, et al. Clin Cancer Res (2006) PMID: 16467100
329. Brown N, et al. PLoS One (2016) PMID: 27232884
330. Soria JC, et al. N. Engl. J. Med. (2018) PMID: 29151359
331. Alanazi A, et al. Lung Cancer Manag (2020) PMID: 33318755
332. Kim et al., 2021; DOI: 10.1200/PO.20.00296
333. Abousand et al., 2021; DOI: 10.26502/jcsct.5079114
334. Cho JH, et al. J. Clin. Oncol. (2019) PMID: 31825714
335. Ramalingam SS, et al. N. Engl. J. Med. (2019) PMID: 31751012
336. Tsuboi et al., 2022; ESMO Abstract LBA47
337. Kenmotsu H, et al. J Thorac Oncol (2022) PMID: 35636696
338. Soo et al., 2021; ESMO Abstract VP3-2021
339. Oxnard GR, et al. Ann. Oncol. (2020) PMID: 32139298
340. Whittle JR, et al. J Clin Neurosci (2015) PMID: 26279503
341. Douillard JY, et al. Ann. Oncol. (2014) PMID: 24718886
342. Price TJ, et al. Lancet Oncol. (2014) PMID: 24739896
343. Van Cutsem E, et al. J. Clin. Oncol. (2007) PMID: 17470858
344. Vermorken JB, et al. Lancet Oncol (2013) PMID: 23746666
345. Wirth LJ, et al. Ann. Oncol. (2010) PMID: 19892746
346. Siano M, et al. Oncologist (2017) PMID: 28592616
347. Mesia R, et al. Lancet Oncol (2015) PMID: 25596660
348. Giral J, et al. Lancet Oncol (2015) PMID: 25596659
349. Siu LL, et al. JAMA Oncol (2016) PMID: 27930762
350. Waddell T, et al. Lancet Oncol. (2013) PMID: 23594787
351. Crawford J, et al. J Thorac Oncol (2013) PMID: 24389433
352. Schuette W, et al. Clin Lung Cancer (2015) PMID: 26094080
353. Leone F, et al. Cancer (2016) PMID: 26540314
354. Vogel A, et al. Eur J Cancer (2018) PMID: 29413685
355. Rowinsky EK, et al. J. Clin. Oncol. (2004) PMID: 15210739

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