

Yeh, Hsiu-Chin

TUMOR TYPE
Peritoneum serous carcinoma
COUNTRY CODE
TW

REPORT DATE
18 Jan 2024

ORDERED TEST #

ORD-1795940-01

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT

DISEASE Peritoneum serous carcinoma
NAME Yeh, Hsiu-Chin
DATE OF BIRTH 12 February 1970
SEX Female

MEDICAL RECORD # 27711633

ORDERING PHYSICIAN Yeh, Yi-Chen

MEDICAL FACILITY Taipei Veterans General Hospital

ADDITIONAL RECIPIENT None

MEDICAL FACILITY ID 205872

PATHOLOGIST Not Provided

SPECIMEN SITE Omentum

SPECIMEN ID S113-00200U (PF24006)

SPECIMEN TYPE Slide Deck

DATE OF COLLECTION 03 January 2024

SPECIMEN RECEIVED 10 January 2024

Biomarker Findings

Loss of Heterozygosity score - 28.3% Homologous Recombination status - HRD Positive Microsatellite status - MS-Stable Tumor Mutational Burden - 7 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

TP53 R196* *CDKN2A/B* p16INK4a loss exon 1 *KMT2A (MLL)* amplification - equivocal[†]

2 Disease relevant genes with no reportable alterations: *BRCA1*, *BRCA2*

† See About the Test in appendix for details.

Report Highlights

- Targeted therapies with NCCN categories of evidence in this tumor type: Olaparib (p. 8), Niraparib (p. 7), Rucaparib (p. 9)
- Evidence-matched clinical trial options based on this patient's genomic findings: (p. 11)

BIOMARKER FINDINGS	THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)		
Loss of Heterozygosity score - 28.3%	Olaparib 1	Talazoparib		
	Niraparib 2A			
10 Trials see p. <u>11</u>	Rucaparib 2A			
Homologous Recombination status - HRD Positive	HRD Positive defined as presence of c LOH score ≥ 16% (Coleman et al., 2017	leleterious <i>BRCA1/2</i> alteration and/or 7; 28916367).		
Microsatellite status - MS-Stable	No therapies or clinical trials. See Biomarker Findings section			
Tumor Mutational Burden - 7 Muts/Mb	No therapies or clinical trials. See Biomarker Findings section			
GENOMIC FINDINGS	THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)		
TP53 - R196*	none	none		
2 Trials see p. <u>13</u>				
		NCCN category		

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.



Yeh, Hsiu-Chin

TUMOR TYPE
Peritoneum serous carcinoma
COUNTRY CODE
TW

REPORT DATE
18 Jan 2024

ORDERED TEST #

ORD-1795940-01

GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, includi implications, see the Genomic Findings section.	ng prognostic, diagnostic, germline, and potential chemosensitivity
<i>CDKN2A/B</i> - p16INK4a loss exon 1p. <u>6</u>	KMT2A (MLL) - amplification - equivocal p. 6

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

BIOMARKER FINDINGS

BIOMARKER

Loss of Heterozygosity score

RESULT 28.3%

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies —

On the basis of emerging clinical data in ovarian cancer, elevated genomic LOH may be associated with greater sensitivity to PARP inhibitors¹⁻³. In platinum-sensitive, BRCA1/2 wild-type ovarian, peritoneal, or Fallopian tube carcinoma, rucaparib elicited significantly longer median PFS (7.2 vs. 5.0 months, HR=0.51) and improved ORR (33.3% vs. 9.6%, p=0.0003) for patients with LOH score \geq 16%1. In the maintenance setting in platinum-sensitive, BRCA1/2 wild-type patients, rucaparib was superior to placebo in both the LOH score \geq 16% (median PFS, 9.7 vs. 5.4 months; HR=0.44) and LOH score < 16% (median PFS, 6.7 vs. 5.4 months; HR=0.58) cohorts². Similar results have been reported for maintenance treatment with niraparib

in ovarian cancer⁴ when using a different measure of HRD that includes genomic LOH⁵⁻⁶. Increased LOH has also been associated with improved sensitivity to platinum-containing chemotherapy regimens in patients with ovarian or breast cancer⁷⁻⁹.

FREQUENCY & PROGNOSIS

In a study of more than 4,000 ovarian, Fallopian tube, or peritoneal cancer samples, genomic LOH score ≥ 16% was identified in 24.2% of BRCA1/2 wild-type cases, deleterious BRCA1/2 mutation was identified in an additional 17.2% of cases, and the remaining 58.7% of cases had LOH score < 16%and were BRCA_{1/2} wild-type¹⁰. Among the histological subtypes, LOH score \geq 16% or BRCA_{1/2} mutation was reported in 42.4% of serous carcinomas, 37.6% of endometrioid carcinomas, 23.5% of carcinosarcomas, 20.6% of neuroendocrine carcinomas, 13.6% of clear cell carcinomas, and 8.1% of mucinous carcinomas; in BRCA_{1/2} wild-type samples, the median LOH score was significantly higher in serous as compared with non-serous cases¹⁰. In ovarian carcinoma, the median LOH score is significantly higher for BRCA1/2-mutated cases than BRCA1/2 wild-type cases (22.2% vs. 9.8%)10, and mutation or methylation of BRCA1, BRCA2, or RAD51C has been reported to be enriched in cases with increased genomic LOH^{7,11}. One study reported no association between LOH and either tumor stage or grade in ovarian serous carcinoma¹². In patients with high-grade serous ovarian carcinoma, the frequency of LOH has been reported to increase significantly with age¹³.

FINDING SUMMARY

The loss of heterozygosity (LOH) score is a profile of the percentage of the tumor genome that is under focal loss of one allele³; focal LOH events accumulate as genomic "scars" as a result of incorrect DNA double-strand break repair when the homologous recombination pathway is deficient (HRD)7,11,14-15. HRD and consequent genomic LOH occur as a result of genetic or epigenetic inactivation of one or more of the homologous recombination pathway proteins, including BRCA1, BRCA2, RAD51C, ATM, PALB2, and BRIP114-17. This sample harbors a genomic LOH score that has been shown to be associated with sensitivity to the PARP inhibitor rucaparib in platinum-sensitive, BRCA1/2 wild-type ovarian, peritoneal, or Fallopian tube carcinoma in both the treatment^{1,3} and maintenance² settings.

BIOMARKER

Microsatellite status

RESULT

MS-Stable

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁸⁻²⁰, including approved therapies nivolumab and pembrolizumab²¹⁻²². In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-

H and experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, p=0.001)²³.

FREQUENCY & PROGNOSIS

MSI-high (MSI-H) has been reported in 1.6-19.7% of ovarian cancer samples $^{24-25}$, including 3.8% (1/26) of ovarian endometrioid adenocarcinomas 26 , and 10.0% (3/30) of ovarian clear cell carcinomas (CCOCs) 27 . No association of MSI-H with stage or survival was found in patients with ovarian cancer 24,28 .

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive

amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA MMR in the tumor²⁹. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2²⁹⁻³¹. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers³²⁻³⁴. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{29,31,33-34}.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.



BIOMARKER FINDINGS

BIOMARKER

Tumor Mutational Burden

RESULT 7 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L135-38, anti-PD-1 therapies36-40, and combination nivolumab and ipilimumab⁴¹⁻⁴⁹. In multiple pan-tumor studies, increased tissue tumor mutational burden (TMB) was associated with sensitivity to immune checkpoint inhibitors^{35-38,40,50-54}. In the KEYNOTE 158 trial of pembrolizumab monotherapy for patients with solid tumors, significant improvement in ORR was observed for patients with TMB ≥10 Muts/Mb (as measured by this assay) compared with those with TMB <10 Muts/Mb in a large cohort that included multiple tumor types⁵⁰; similar findings were observed in the KEYNOTE 028 and 012 trials $^{\!40}.$ At the same TMB cutpoint, retrospective analysis of

patients with solid tumors treated with any checkpoint inhibitor identified that tissue TMB scores ≥ 10 Muts/Mb were associated with prolonged time to treatment failure compared with scores <10 muts/Mb (HR=0.68)⁵⁴. For patients with solid tumors treated with nivolumab plus ipilimumab in the CheckMate 848 trial, improved responses were observed in patients with a tissue TMB ≥ 10 Muts/Mb independent of blood TMB at any cutpoint in matched samples⁵⁵. However, support for higher TMB thresholds and efficacy was observed in the prospective Phase 2 MyPathway trial of atezolizumab for patients with pan-solid tumors, where improved ORR and DCR was seen in patients with TMB ≥ 16 Muts/Mb than those with TMB \geq 10 and <16 Muts/Mb⁵³. Similarly, analyses across several solid tumor types reported that patients with higher TMB (defined as ≥16-20 Muts/Mb) achieved greater clinical benefit from PD-1 or PD-L1-targeting monotherapy compared with patients with higher TMB treated with chemotherapy³⁵ or those with lower TMB treated with PD-1 or PD-L1-targeting agents37.

FREQUENCY & PROGNOSIS

Ovarian carcinomas, including peritoneal and Fallopian tube carcinomas, harbor a median TMB of 2.7–3.6 Muts/Mb depending upon subtype, and up to 2.1% of cases have high TMB (>20 Muts/

Mb)⁵⁶. In a study of high grade serous ovarian cancer, homologous recombination (HR)-deficient tumors, which comprised ~50% of all samples, harbored a higher neoantigen load compared to HR-proficient tumors; higher neoantigen load was associated with longer OS but not disease free survival⁵⁷.

FINDING SUMMARY

Tumor mutational burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitutions and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma⁵⁸⁻⁵⁹ and cigarette smoke in lung cancer⁶⁰⁻⁶¹, treatment with temozolomide-based chemotherapy in glioma⁶²⁻⁶³, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes $^{64\text{-}68}$, and microsatellite instability^{64,67-68}. This sample harbors a TMB level associated with lower rates of clinical benefit from treatment with PD-1- or PD-L1-targeting immune checkpoint inhibitors compared with patients with tumors harboring higher TMB levels, based on several studies in multiple solid tumor types^{37-38,50}.



GENOMIC FINDINGS

GENE

TP53

ALTERATION

R196*

HGVS VARIANT NM_000546.4:c.586C>T (p.R196*)

VARIANT CHROMOSOMAL POSITION

chr17:7578263

VARIANT ALLELE FREQUENCY (% VAF) 62.0%

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor adavosertib⁶⁹⁻⁷² or p53 gene therapy such as SGT53⁷³⁻⁷⁸. In a Phase 1 study, adavosertib in combination with gemcitabine, cisplatin, or carboplatin elicited PRs in 9.7% and SDs in 53% of patients with solid tumors; the response rate was 21% (4/19) for patients with TP53 mutations versus 12% (4/33) for patients who were TP53 wildtype⁷⁹. Phase 2 studies of adavosertib in combination with chemotherapy reported ORRs of 32% (30/94) and 41% (12/29) for patients with platinum-refractory TP53-mutated ovarian, Fallopian tube, or peritoneal cancer⁸⁰⁻⁸¹. For patients with platinum-sensitive TP53-mutated ovarian cancer, the combination of adavosertib with paclitaxel and carboplatin significantly increased PFS compared with paclitaxel and carboplatin alone (9.9 vs. 8.0 months)82. In the Phase 2 VIKTORY trial, patients with TP53-mutated metastatic and/or recurrent gastric cancer experienced a 24% (6/25) ORR with adavosertib combined with paclitaxel⁸³. A Phase 1 trial of neoadiuvant adayosertib in combination with cisplatin and docetaxel for head and neck squamous cell carcinoma (HNSCC) elicited a 71% (5/7) response rate for patients with TP53 alterations 84 . The Phase 2 FOCUS₄-C trial for patients with TP53- and RAS-mutated colorectal cancer reported improvement in PFS (3.61 vs. 1.87 months, HR=0.35, p=0.0022), but not OS (14.0 vs 12.8 months, p=0.93), following adavosertib

treatment compared with active monitoring85. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel for patients with solid tumors, 75% (9/12) of evaluable patients experienced clinical benefit, including 2 confirmed and 1 unconfirmed PRs and 2 instances of SD with significant tumor shrinkage⁷⁸. Missense mutations leading to TP₅₃ inactivation may be sensitive to therapies that reactivate mutated p53 such as eprenetapopt. In a Phase 1b trial for patients with p53-positive highgrade serous ovarian cancer, eprenetapopt combined with carboplatin and pegylated liposomal doxorubicin achieved a 52% (11/21) response rate and 100% DCR86. A Phase 1 trial of eprenetapopt with pembrolizumab for patients with solid tumors reported an ORR of 10% (3/29)87.

FREQUENCY & PROGNOSIS

TP53 alterations have been reported in all subtypes of ovarian tumors; generally, high grade serous tumors are associated with greater frequency of TP53 alteration (>80%), while low grade serous and clear cell ovarian tumors are infrequently associated with TP53 alteration (<10%). The MSK-METropism study reported TP53 alterations in 96% of high grade serous ovarian cancers versus 1% in low grade serous and 15% of clear cell carcinomas, similar to trends reported across earlier studies^{16,88-95}. Similarly, multiple genomic studies of ovarian mucinous neoplasms reported TP53 mutations in 12-18% of benign or borderline tumors and in 52-64% of carcinomas 96-97. In endometrioid carcinomas, TP53 mutations were reported in 26% of cases in a genomic study98. Aberrant p53 expression has been associated with higher ovarian serous carcinoma grade (89-90% of high-grade vs. 6.6-9% of low-grade vs. 0% of benign)99-101. In one study, p53 expression was not prognostic in regard to outcome in patients with peritoneum serous carcinoma¹⁰². TP₅₃ mutations have been reported to be more frequent in advanced stage (63%, 55/87) and higher grade (65%, 42/64) than earlier stage (31%, 14/45) and lower grade (41%, 7/17) ovarian carcinomas93. Metaanalysis has suggested that TP53 expression was associated with poorer survival in ovarian epithelial cancers, although the effect was modest and considerable variability was observed between studies103.

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers¹⁰⁴. Alterations such as seen here may disrupt TP53 function or expression¹⁰⁵⁻¹⁰⁹.

POTENTIAL GERMLINE IMPLICATIONS

One or more of the TP53 variants observed here has been described in the ClinVar database as a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with Li-Fraumeni syndrome (ClinVar, Sep 2023)¹¹⁰. Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. Germline mutations in TP53 are associated with the very rare autosomal dominant disorder Li-Fraumeni syndrome and the early onset of many cancers¹¹¹⁻¹¹³, including sarcomas¹¹⁴⁻¹¹⁵. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000¹¹⁶ to 1:20,000¹¹⁵. For pathogenic TP53 mutations identified during tumor sequencing, the rate of germline mutations was 1% in the overall population and 6% in tumors arising before age 30¹¹⁷. In the appropriate clinical context, germline testing of TP53 is recommended.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion¹¹⁸⁻¹²³. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy¹¹⁸⁻¹¹⁹. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease¹²⁴. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH122,125-126. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.



GENOMIC FINDINGS

GENE

CDKN2A/B

ALTERATION

p16INK4a loss exon 1

POTENTIAL TREATMENT STRATEGIES

Targeted Therapies —

Preclinical data suggest that tumors with loss of p16INK4a function may be sensitive to CDK4/6 inhibitors, such as abemaciclib, ribociclib, and palbociclib¹²⁷⁻¹³⁰. Clinical data in mesothelioma, breast cancer, and uterine leiomyosarcoma indicate that CDKN2A loss may predict sensitivity to abemaciclib¹³¹ and palbociclib treatment¹³²⁻¹³³. However, multiple other clinical studies have shown no significant correlation between p16INK4a loss or inactivation and therapeutic benefit of these agents¹³⁴⁻¹⁴⁰; it is not known whether CDK4/6 inhibitors would be beneficial in this case.

FREQUENCY & PROGNOSIS

CDKN2A mutation has been observed in <1% of peritoneal tumors analyzed in the COSMIC database (Jan 2023)¹⁴¹. In the Ovarian Serous Cystadenocarcinoma TCGA dataset, CDKN2A/B

loss has been reported in 2.2% of cases, whereas mutations in these genes have not been detected¹⁶. Inactivation of CDKN2A/B by promoter methylation, leading to loss of p16INK4a and p15INK4b expression, has been reported in ovarian cancer samples¹⁴²⁻¹⁴³. Reduced expression of both CDKN2A mRNA and p16INK4a protein levels has been reported in 30% of epithelial ovarian tumors, but gene deletion accounted for very few to none of these cases¹⁴⁴⁻¹⁴⁵. Among low-grade tumors, loss of p16INK4a has been associated with the progression from borderline to low-grade invasive ovarian cancer, but p16INK4a expression in highgrade ovarian cancer has been reported to be variable¹⁴⁶. Although earlier studies reported conflicting results on the association of p16INK4a expression and ovarian cancer prognosis¹⁴⁷⁻¹⁵², a large study of high-grade serous ovarian carcinoma concluded that homogeneous (high) p16INK4a expression and concurrent Rb positivity predict shorter patient survival¹⁵³.

FINDING SUMMARY

CDKN2A encodes two different, unrelated tumor suppressor proteins, p16INK4a and p14ARF, whereas CDKN2B encodes the tumor suppressor p15INK4b¹⁵⁴⁻¹⁵⁵. Both p15INK4b and p16INK4a bind to and inhibit CDK4 and CDK6, thereby maintaining the growth-suppressive activity of the

Rb tumor suppressor; loss or inactivation of either p15INK4b or p16INK4a contributes to dysregulation of the CDK4/6-cyclin-Rb pathway and loss of cell cycle control¹⁵⁶⁻¹⁵⁷. The tumor suppressive functions of p14ARF involve stabilization and activation of p53, via a mechanism of MDM2 inhibition¹⁵⁸⁻¹⁵⁹. One or more alterations observed here are predicted to result in p16INK4a loss of function¹⁶⁰⁻¹⁸¹.

POTENTIAL GERMLINE IMPLICATIONS

Germline CDKN2A mutation is associated with melanoma-pancreatic cancer syndrome, a condition marked by increased risk of developing malignant melanoma and/or pancreatic cancer¹⁸². Mutation carriers within families may develop either or both types of cancer, and melanoma cases may be referred to as familial or hereditary melanoma¹⁸³⁻¹⁸⁴. CDKN₂A is the most implicated gene in familial melanoma, with germline mutations present in 16% to 20% of familial melanoma cases¹⁸⁵⁻¹⁸⁷. CDKN₂A alteration has also been implicated in familial melanoma-astrocytoma syndrome, an extremely rare tumor association characterized by dual predisposition to melanoma and nervous system tumors 188-190. In the appropriate clinical context, germline testing of CDKN2A is recommended.

GENE

KMT2A (MLL)

ALTERATION

amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

- Targeted Therapies -

There are no targeted therapies available to address

genomic mutations in or amplification of MLL.

FREQUENCY & PROGNOSIS

MLL amplification is rare in most solid tumors, occurring in <2% of cases in most tumor types (cBioPortal, Apr 2023)¹⁹¹⁻¹⁹². Published data investigating the prognostic implications of MLL alterations in ovarian, fallopian and peritoneal carcinomas are limited (PubMed, May 2023). Published data investigating the prognostic implications of KMT2A (MLL) alterations in solid

tumors are limited (PubMed, Nov 2023).

FINDING SUMMARY

MLL (also known as KMT2A) encodes a histone methyltransferase, an enzyme involved in the modification of histones. It is involved in the positive regulation of transcription, particularly during development, although it is also expressed in most adult tissues¹⁹³⁻¹⁹⁴. MLL has been reported to be amplified in cancer, and may be biologically relevant in this context¹⁹⁵⁻¹⁹⁷.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.



THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

Niraparib

Assay findings association

Loss of Heterozygosity score 28.3%

AREAS OF THERAPEUTIC USE

The PARP inhibitor niraparib is FDA approved to treat patients with epithelial ovarian, fallopian tube, or primary peritoneal cancer, with or without homologous recombination deficiency (HRD)-positive status. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of emerging clinical data in ovarian cancer, elevated genomic LOH may be associated with greater sensitivity to PARP inhibitors $^{1-3}$. In platinum-sensitive, BRCA1/2 wild-type ovarian, peritoneal, or Fallopian tube carcinoma, the PARP inhibitor rucaparib elicited significantly longer median PFS and improved ORR for patients with LOH score $\geq 16\%$.

SUPPORTING DATA

For patients with platinum-sensitive, recurrent ovarian cancer (OC), the Phase 3 ENGOT-OV16/NOVA study showed that niraparib maintenance therapy significantly increased median PFS (mPFS) relative to placebo, regardless of germline BRCA (gBRCA) mutation (21.0 vs. 5.5 months, HR=0.27 for patients with gBRCA mutations; 9.3 vs. 3.9 months, HR=0.45 for patients without gBRCA mutations) or homologous recombination deficiency (HDR) status (12.9 vs. 3.8 months, HR=0.38)^{4,198}. Similarly, the Phase 3 PRIMA trial for patients with newly diagnosed advanced OC reported an extended mPFS from

niraparib maintenance therapy after response to first-line platinum chemotherapy compared with placebo (13.8 vs. 8.2 months, HR=0.62); subgroup analysis showed that patients with HRD-positive OC experienced the longest mPFS, which was irrespective of BRCA mutational status (21.9 vs. 10.4 months, HR=0.43)¹⁹⁹. The Phase 2 QUADRA study evaluating niraparib monotherapy as late-line treatment for patients with relapsed high-grade serous epithelial ovarian cancer (HGSOC) reported a median OS (mOS) of 12.2 months for all HGSOC patients, with highest mOS seen for those with BRCA1/2-mutated and HRD-positive tumors (26.0 and 19.0 months, respectively)200. A Phase 2 trial for patients with platinum-sensitive HGSOC and endometrioid recurrent ovarian cancer reported significantly improved mPFS from the addition of bevacizumab to niraparib compared with niraparib alone (11.9 vs. 5.5 months, HR=0.35)²⁰¹. In a Phase 1/2 study of niraparib in combination with pembrolizumab for patients with recurrent platinumresistant OC, patients experienced an ORR of 19% (11/59) and mPFS of 3.4 months; no significant differences in ORR were noted among analyzed subgroups: 14% (3/21) versus 19% (6/32) for patients with HRD-positive versus HRD-negative tumors, 18% (2/11) versus 19% (9/47) for patients with BRCA-mutated versus BRCA-wildtype tumors, and 21% (7/33) versus 10% (2/21) for patients with PD-L1-positive versus PD-L1-negative tumors²⁰².

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

Olaparib

Assay findings association

Loss of Heterozygosity score 28.3%

AREAS OF THERAPEUTIC USE

The PARP inhibitor olaparib is FDA approved to treat patients with epithelial ovarian, Fallopian tube, or primary peritoneal cancer, patients with deleterious or suspected deleterious gBRCA-mutated pancreatic adenocarcinoma or HER2-negative breast cancer, and patients with prostate cancer and mutations in homologous recombination repair genes. Olaparib is also approved in combination with bevacizumab to treat patients with ovarian, Fallopian tube, or primary peritoneal cancer with deleterious or suspected deleterious somatic or gBRCA mutation and/or genomic instability. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of emerging clinical data in ovarian cancer, elevated genomic LOH may be associated with greater sensitivity to PARP inhibitors $^{1-3}$. In platinum-sensitive, BRCA1/2 wild-type ovarian, peritoneal, or Fallopian tube carcinoma, the PARP inhibitor rucaparib elicited significantly longer median PFS and improved ORR for patients with LOH score $\geq 16\%$ $^{1.3}$.

SUPPORTING DATA

Olaparib has been studied primarily to treat patients with ovarian cancer harboring BRCA1/2 mutations. Numerous Phase 2 studies have demonstrated significant clinical activity for patients with BRCA-mutated ovarian cancer, with response rates often significantly higher for patients with BRCA mutations than for those without 203-204 and for patients with platinum-sensitive (vs. platinum resistant) cancer²⁰⁴⁻²⁰⁷. As maintenance therapy for patients with newly diagnosed or platinum-sensitive relapsed ovarian cancer, olaparib monotherapy demonstrated significantly improved OS and median PFS (mPFS) compared with placebo in the Phase 3 SOLO-1 study²⁰⁸ and multiple later-phase studies²⁰⁹⁻²¹³. At the 7-year follow-up of SOLO-1, olaparib continued to improve mPFS compared with placebo for patients with ovarian cancer²¹⁴, and mOS was not reached for olaparib compared with 75.2 months for placebo (HR=0.55)²¹⁵. In the first study of PARP inhibitor rechallenge for patients with ovarian cancer, the Phase 3 OReO/ENGOT Ov-38 study of maintenance olaparib compared with placebo reported statistically improved mPFS for both patients with BRCA-mutated (4.3 vs. 2.8 months, HR=0.57) and BRCA-unmutated (5.3 vs. 2.8 months, HR=0.43) cancer; in an exploratory analysis of the BRCA-unmutated cohort, improved mPFS was reported regardless of homologous recombination deficiency status²¹⁶. The Phase 4 ORZORA trial confirmed the efficacy of olaparib as maintenance therapy for platinum-sensitive relapsed ovarian cancer after ≥2 lines of treatment, with an mPFS of 16.6 months

for patients with somatic BRCA mutations (n=35) and of 19.3 months for those with germline BRCA mutations (n=52)²¹⁷. Olaparib has been evaluated in combination with other therapies. A statistically superior median PFS (mPFS) from treatment with olaparib in combination with the VEGF inhibitor bevacizumab compared with bevacizumab monotherapy was reported in the Phase 3 PAOLA-1 study for patients with newly diagnosed advanced ovarian cancer in the intent-to-treat population (22.1 vs. 16.6 months, HR=0.59), in the population with BRCA1/2 mutations (37.2 vs. 21.7 months, HR=0.31), and in the population with wildtype BRCA1/2 harboring homologous recombination deficiency (HRD)-positive status (28.1 vs. 16.6 months, HR=0.43); however, no significant difference in mPFS was reported in the population with mutations in non-BRCA genes involved in homologous recombination repair (HRR) (16.8-22.1 vs. 16.6-27.0 months, HR=0.95-1.83)²¹⁸. The final OS analysis for this trial reported benefit from the maintenance treatment using olaparib plus bevacizumab versus bevacizumab alone for patients with HRD-positive status (65.5 vs. 48.4 months), patients with HRD-positive status and BRCA1/2 mutations (73.2 vs. 53.8 months), and patients with HRD-positive status without BRCA1/2 mutations (54.7 vs. 44.2 months)²¹⁹. For patients with platinum-sensitive recurrent ovarian cancer who previously progressed on chemotherapy, statistically increased mPFS was reported in a Phase 2 study of olaparib in combination with chemotherapy (12.2 months) compared with chemotherapy alone (9.6 months)220, as well as from treatment with the VEGFR inhibitor cediranib compared with olaparib monotherapy in a Phase 1/2 trial²²¹. Combination treatment with cediranib and olaparib also demonstrated numerically longer mPFS compared with chemotherapy alone for patients with BRCA₁/₂-mutated platinum-sensitive recurrent ovarian cancer (10.5 vs. 18 months); however, improved clinical benefit was not observed for patients without BRCA1/2 mutation (mPFS of 9.7 vs. 8.9 months) or for the overall intent-to-treat population (mPFS of 10.3 vs. 10.4 months, ORR of 71% vs. 69%)222. The Phase 2 CAPRI study for PARP inhibitor-resistant patients with HRD platinumsensitive high-grade ovarian cancer treated with combination therapy of olaparib and the ATR inhibitor ceralasertib reported an ORR of 46% (n=13, 6 PRs) and a PFS of 7.5 months²²³. For patients with PARP-resistant ovarian cancer, the combination of olaparib and the WEE1 inhibitor adavosertib elicited improved clinical benefit (ORR 29%, DCR 89%) compared with adavosertib alone (ORR 23%, DCR 63%); however, in the BRCA-mutated cohort, no significant difference in clinical benefit was observed between the combination (ORR 19%) and monotherapy (ORR 20%) treatments²²⁴.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.



THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

Rucaparib

Assay findings association

Loss of Heterozygosity score 28.3%

AREAS OF THERAPEUTIC USE

The PARP inhibitor rucaparib is FDA approved to treat patients with metastatic castration-resistant prostate cancer (mCRPC) and deleterious somatic or germline BRCA mutations. Rucaparib is also approved as a maintenance treatment of patients with recurrent epithelial ovarian, Fallopian tube, or primary peritoneal cancer. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of emerging clinical data in ovarian cancer, elevated genomic LOH may be associated with greater sensitivity to PARP inhibitors $^{1-3}$. In platinum-sensitive, BRCA1/2 wild-type ovarian, peritoneal, or Fallopian tube carcinoma, the PARP inhibitor rucaparib elicited significantly longer median PFS and improved ORR for patients with LOH score $\geq 16\%$.

SUPPORTING DATA

As first-line maintenance therapy evaluated in the Phase 3 ATHENA study, rucaparib monotherapy significantly improved median PFS (mPFS) compared with placebo for patients with ovarian cancer in both the homologous

recombination deficient (HRD)-positive population (BRCA mutated or BRCA wildtype and high genomic loss of heterozygosity [LOH] score defined as ≥16%; mPFS of 28.7 vs. 11.3 months, HR=0.47) and the overall population (mPFS of 20.2 vs. 9.2 months, HR=0.52); exploratory subgroup analysis also reported benefit for the HRDnegative group (BRCA wildtype and low genomic LOH score defined as <16%; mPFS of 12.1 vs. 9.1 months, HR=0.65)²²⁵. In the Phase 3 ARIEL3 study of rucaparib maintenance treatment for patients with recurrent platinum-sensitive high-grade serous or endometrioid ovarian, primary peritoneal, or fallopian tube carcinoma, mPFS was significantly improved with rucaparib compared with placebo for patients, irrespective of HRD or BRCA status, with benefit observed for both the LOHhigh subgroup (13.6 vs. 5.4 months, HR=0.32) and the overall population (10.8 vs. 5.4 months, HR=0.36), with CR rates of 12% for the HRD-positive group and 7% for the overall population; PFS benefit was also observed for the BRCA-wildtype, LOH-low group (HR=0.58)2. In the Phase 2 ARIEL2 trial for patients with recurrent platinumsensitive ovarian, peritoneal, or fallopian tube carcinoma, mPFS on rucaparib was 5.7 months for patients with high LOH and 5.2 months for patients with low LOH3.



REPORT DATE 18 Jan 2024



ORDERED TEST # ORD-1795940-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Talazoparib

Assay findings association

Loss of Heterozygosity score 28.3%

AREAS OF THERAPEUTIC USE

The PARP inhibitor talazoparib is FDA approved to treat HER2-negative locally advanced or metastatic breast cancer with deleterious or suspected deleterious germline BRCA mutations. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of emerging clinical data in ovarian cancer, elevated genomic LOH may be associated with greater sensitivity to PARP inhibitors¹⁻³. In platinum-sensitive, BRCA1/2 wild-type ovarian, peritoneal, or Fallopian tube

carcinoma, the PARP inhibitor rucaparib elicited significantly longer median PFS and improved ORR for patients with LOH score $\geq 16\%^{1,3}$.

SUPPORTING DATA

An ORR of 42% (5/12) was reported for patients with BRCA1/2-mutated ovarian cancer treated with talazoparib in a Phase 1 study²²⁶. In a Phase 2 study of talazoparib in advanced solid tumors, 1 patient with BRIP1-mutated ovarian carcinoma with wildtype BRCA status experienced a prolonged SD²²⁷.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.



CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial > Geographical proximity > Later trial phase. Clinical trials listed here may have additional enrollment criteria that

may require medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov. Or, visit https://www.foundationmedicine.com/genomictesting#support-services.

Loss of Heterozygosity

RATIONALE

On the basis of emerging clinical data in ovarian cancer, elevated genomic LOH may be associated with greater sensitivity to PARP inhibitors.

RESULT 28.3%

> NCT05489211 PHASE 2 Study of Dato-Dxd as Monotherapy and in Combination With Anti-cancer Agents in Patients With **TARGETS** Advanced Solid Tumours (TROPION-PanTumor03) TROP2, PD-L1, PARP1, PD-1

LOCATIONS: Taipei (Taiwan), Taoyuan (Taiwan), Liou Ying Township (Taiwan), Hangzhou (China), Shanghai (China), Zhengzhou (China), Seoul (Korea, Republic of), Chongqing (China), Suita-shi (Japan), Nagoya-shi (Japan)

NCT04434482 **PHASE 1/2** IMP4297 in Combination With Temozolomide in Patients With Advanced Solid Tumors and Small Cell **TARGETS** Lung Cancer **PARP**

LOCATIONS: Taipei (Taiwan), Taoyuan (Taiwan), Taichung (Taiwan), Tainan (Taiwan), Wuhan (China), Gyeonggi-do (Korea, Republic of), Seoul (Korea, Republic of), Cheongju-si (Korea, Republic of), Beijing (China), Jilin (China)

NCT05797168 PHASE 1/2 Phase I/IIa Study for AZD5335 as Monotherapy and in Combination With Anti-cancer Agents in **TARGETS** PARP1

LOCATIONS: Tainan City (Taiwan), Tokyo (Japan), Melbourne (Australia), Haifa (Israel), Ramat Gan (Israel), California, Texas

NCT04884360 PHASE 3 D9319C00001-1L OC Mono Global RCT **TARGETS PARP**

LOCATIONS: Wenzhou (China), Jiaxing (China), Shanghai (China), Suzhou (China), Wuxi (China), Nanjing (China), Hefei (China), Beijing (China), Changchun (China), Changsha (China)

NCT04518501 PHASE 1/2 **TARGETS** Fuzuloparib Arsenic Trioxide Platinum Resistance Relapsed Ovarian Cancer RARA, PARP LOCATIONS: Zhejiang (China)

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy



REPORT DATE 18 Jan 2024

FOUNDATIONONE®CDx

ORDERED TEST # ORD-1795940-01

CLINICAL TRIALS

NCT05489926	PHASE 2		
A Study to Explore Pamiparib Treatment in Epithelial Ovarian Cancer After Prior PARP Inhibitor Exposure	TARGETS PARP		
LOCATIONS: Hangzhou (China)			
NCT03983226	PHASE 2		
Surgery and Niraparib in Secondary Recurrent Ovarian Cancer (SOC-3 Trial)	TARGETS PARP		
LOCATIONS: Shanghai (China)			
NCT04586335	PHASE 1		
Study of CYH33 in Combination With Olaparib an Oral PARP Inhibitor in Patients With Advanced Solid Tumors.	TARGETS PARP, PI3K-alpha		
LOCATIONS: Shanghai (China)			
NCT03742895	PHASE 2		
Efficacy and Safety of Olaparib (MK-7339) in Participants With Previously Treated, Homologous Recombination Repair Mutation (HRRm) or Homologous Recombination Deficiency (HRD) Positive Advanced Cancer (MK-7339-002 / LYNK-002)	TARGETS PARP		
LOCATIONS: Seongnam-si (Korea, Republic of), Seoul (Korea, Republic of), Darlinghurst (Australia), Ac Ramat Gan (Israel), Istanbul (Turkey), Antalya (Turkey), Edirne (Turkey)	dana (Turkey), Jerusalem (Israel), Konya (Turkey),		
NCT02264678	PHASE 1/2		
Ascending Doses of AZD6738 in Combination With Chemotherapy and/or Novel Anti Cancer Agents	TARGETS ATR, PARP, PD-L1		
LOCATIONS: Seoul (Korea, Republic of), Goyang-si (Korea, Republic of), Cambridge (United Kingdom), Kingdom), London (United Kingdom), Coventry (United Kingdom), Sutton (United Kingdom), Oxford (U			



REPORT DATE 18 Jan 2024

FOUNDATIONONE®CDx

ORDERED TEST # ORD-1795940-01

CLINICAL TRIALS

TP53

RATIONALE

TP53 loss of function alterations may predict sensitivity to WEE1 inhibitors.

ALTERATION R196*

NCT04516447	PHASE 1
A Study of ZN-c3 in Patients With Platinum-Resistant Ovarian Cancer	TARGETS WEE1

LOCATIONS: Busan (Korea, Republic of), Seoul (Korea, Republic of), Nedlands (Australia), Sunshine Coast (Australia), South Brisbane (Australia), Melbourne (Australia), Belgrade (Serbia), Tuzla (Bosnia and Herzegovina), Banja Luka (Bosnia and Herzegovina), Colorado

NCT03968653	PHASE 1			
Study of Oral Debio 0123 in Combination With Carboplatin in Participants With Advanced Solid Tumors	TARGETS WEE1			
LOCATIONS: Groningen (Netherlands), Nijmegen (Netherlands), Leiden (Netherlands), Barcelona (Spain)				



report date 18 Jan 2024



ORDERED TEST # ORD-1795940-01

APPENDIX

Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

ARAF

NM_001654.3: c.1734G>C (p.E578D) chrX:47430769

EGFR

NM_005228.3: c.-38_26del (p.M1?) chr7:55086932-55086996

KMT2D (MLL2)

NM_003482.4: c.12143C>T (p.P4048L) chr12:49426345 **AURKB**

NM_004217.2: c.241C>T (p.R81C) chr17:8110651

KDM5C

NM_004187.3: c.4094C>T (p.P1365L) chrX:53222978

NTRK1

NM_002529.3: c.1697T>C (p.M566T) chr1:156846256 BRD4

NM_014299.2: c.682G>A (p.V228I) chr19:15376332

KDM6A

NM_021140.2: c.1736G>A (p.C579Y) chrX:44922875 CHEK1

amplification

KMT2A (MLL)

NM_005933.3: c.9742_9752del (p.V3248Yfs*5) chr11:118376357-118376368

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1795940-01

APPENDIX

Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY **NUMBER ALTERATIONS**

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B	or WTX)
APC	AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX
AURKA	AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2
BCL6	BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1
BTG2	BTK	CALR	CARD11	CASP8	CBFB	CBL	CCND1	CCND2
CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73	CDH1
CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B	CDKN2C
CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R	CTCF
CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1	DDR2
DIS3	DNMT3A	DOT1L	EED	EGFR	EMSY (C11orf30)	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRFI1	ESR1	EZH2
FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12	FGF14
FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3	FGFR4
FGF 19 FH	FGF23 FLCN	FLT1	FGF4 FLT3	FOXL2	FUBP1	GABRA6	GATA3	GATA4
					GNAS		GSK3B	
GATA6	GID4 (C17orf39)	GNA11	GNA13	GNAQ		GRM3		H3-3A (H3F3A)
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDM5A	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11 (MRE11A)	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2	<i>NOTCH3</i>
NPM1	NRAS	NSD2 (WHSC1 or I	•	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3
P2RY8	PALB2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)
PDGFRA	PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1
PMS2	POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI
PRKN (PARK2)	PTCH1	PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51
RAD51B	RAD51C	RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10
REL	RET	RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC
SDHD	SETD2	SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO
SNCAIP	SOCS1	SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3
STK11	SUFU	SYK	TBX3	TEK	TENT5C (FAM46C)	TET2	TGFBR2
TIPARP	TNFAIP3	TNFRSF14	TP53	TSC1	TSC2	TYRO3	U2AF1	VEGFA
VHL	WT1	XPO1	XRCC2	ZNF217	ZNF703			
DNA GENE LIS	ST: FOR THE D	ETECTION OF	SELECT REAR	RANGEMENTS				
ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSPO2	SDC4	SLC34A2	TERC*	TERT**	TMPRSS2

^{*}TERC is an NCRNA

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Homologous Recombination status Loss of Heterozygosity (LOH) score Microsatellite (MS) status Tumor Mutational Burden (TMB)

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy

^{**}Promoter region of TERT is interrogated



APPENDIX

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a,

Cipalstraat 3, 2440 Geel, Belgium. C €

ABOUT FOUNDATIONONE CDX

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details: www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffinembedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLE

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes

detection of alterations in a total of 324 genes.

Using an Illumina® Sequencing platform (HiSeq 4000 or NovaSeq 6000), hybrid capture–selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI), and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for *ERBB2* and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is

one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials

Ranking of Therapies in Summary Table
Therapies are ranked based on the following
criteria: Therapies with clinical benefit (ranked
alphabetically within each evidence category),
followed by therapies associated with resistance
(when applicable).

Ranking of Clinical Trials
Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK* (NCCN*) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2023. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

1. In the fraction-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

APPENDIX

About FoundationOne®CDx

- MSS cut-off thresholds were determined by analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.
- 2. TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh_docs/ pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
- 3. Homologous Recombination status may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas (Coleman et al., 2017; 28016367). Samples with deleterious BRCA1/2 alteration and/or Loss of Heterozygosity (LOH) score ≥ 16% will be reported as "HRD Positive" and samples with absence of these findings will be reported as "HRD Not Detected," agnostic of potential secondary BRCA1/2 reversion alterations. Certain potentially deleterious missense or small in-frame deletions in BRCA1/2 may not be classified as deleterious and, in the absence of an elevated LOH profile, samples with such mutations may be classified as "HRD Not Detected." A result of "HRD Not Detected" does not rule out the presence of a BRCA1/2 alteration or an elevated LOH profile outside the assay performance characteristic limitations.
- **4.** The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the

- genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding armand chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
- 5. Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
- 6. Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine whether the patient is a candidate for biopsy.
- 7. Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an ERBB2 amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of ERBB2 copy number 4 in breast cancer is estimated to be approximately 2%. HER2 overexpression occurs in 18-20% of breast cancers (Owens et al. 2004 [PMID: 15140287]; Salmon et al. 1987 [PMID: 3798106]; Yaziji et al. 2004 [PMID: 15113815]). Based on the F1CDx HER2 CDx concordance study, approximately 10% of HER2 amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic,

nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
INDELS Repeatability	%CV*

^{*}Interquartile Range = 1st Quartile to 3rd Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of followup germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are ATM, BAP1, BRCA1, BRCA2, BRIP1, CHEK2, FH, FLCN, MLH1, MSH2, MSH6, MUTYH, PALB2, PMS2, POLE, RAD51C, RAD51D, RET, SDHA, SDHB, SDHC, SDHD, TSC2, and VHL, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.



APPENDIX

About FoundationOne®CDx

distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are ASXL1, CBL, DNMT3A, IDH2, JAK2, KMT2D (MLL2), MPL, MYD88, SF3B1, TET2, and U2AF1 and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent

medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
muts/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
os	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

REFERENCE SEQUENCE INFORMATION

Sequence data is mapped to the human genome, Genome Reference Consortium Human Build 37 (GRCh37), also known as hg19.

SOFTWARE VERSION INFORMATION

MR Suite Version (RG) 7.15.0 MR Reporting Config Version Config 49 Analysis Pipeline Version v3.29.0 Computational Biology Suite Version 6.29.0

The median exon coverage for this sample is 836x

APPENDIX

References

ORDERED TEST # ORD-1795940-01

- 1. Coleman et al., 2016; ASCO Abstract 5540
- 2. Coleman RL, et al. Lancet (2017) pmid: 28916367
- 3. Swisher EM, et al. Lancet Oncol. (2017) pmid: 27908594 4. Mirza MR, et al. N. Engl. J. Med. (2016) pmid: 27717299
- 5. Telli ML, et al. Clin. Cancer Res. (2016) pmid: 26957554
- 6. Timms KM, et al. Breast Cancer Res. (2014) pmid: 25475740
- 7. Wang ZC, et al. Clin. Cancer Res. (2012) pmid: 22912389
- 8. Telli ML, et al. J. Clin. Oncol. (2015) pmid: 25847929
- 9. Isakoff SJ, et al. J. Clin. Oncol. (2015) pmid: 25847936
- 10. Elvin et al., 2017; ASCO Abstract 5512
- 11. Abkevich V, et al. Br. J. Cancer (2012) pmid: 23047548
- 12. Marquard AM, et al. Biomark Res (2015) pmid: 26015868
- 13. Pedersen BS, et al. Genes Chromosomes Cancer (2013) pmid: 23716468
- 14. Watkins JA, et al. Breast Cancer Res. (2014) pmid: 25093514
- Vanderstichele A, et al. Eur. J. Cancer (2017) pmid: 28950147
- 16. Nature (2011) pmid: 21720365
- 17. N. Engl. J. Med. (2003) pmid: 12736286
- Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. 18. (2014) pmid: 25392179
- 19. Kroemer G, et al. Oncoimmunology (2015) pmid: 26140250
- 20. Lal N, et al. Oncoimmunology (2015) pmid: 25949894
- 21. Overman et al., 2016; ASCO Abstract 3501
- 22. Le DT, et al. N. Engl. J. Med. (2015) pmid: 26028255
- 23. Ayers et al., 2016; ASCO-SITC Abstract P60
- **24.** Segev Y, et al. Eur. J. Gynaecol. Oncol. (2015) pmid: 26775351
- 25. Plisiecka-Hałasa J, et al. Anticancer Res. () pmid: 18507046
- 26. Huang HN, et al. Histopathology (2015) pmid: 25195947
- 27. Strickland et al., 2016; ASCO Abstract 5514
- 28. Aysal A, et al. Am. J. Surg. Pathol. (2012) pmid: 22189970
- Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) pmid: 26337942
- **30.** You JF, et al. Br. J. Cancer (2010) pmid: 21081928
- 31. Bairwa NK, et al. Methods Mol. Biol. (2014) pmid: 24623249
- 32. Boland CR, et al. Cancer Res. (1998) pmid: 9823339
- 33. Pawlik TM, et al. Dis. Markers (2004) pmid: 15528785
- Boland CR, et al. Gastroenterology (2010) pmid: 20420947
- 35. Legrand et al., 2018; ASCO Abstract 12000
- 36. Samstein RM, et al. Nat. Genet. (2019) pmid: 30643254
- 37. Goodman AM, et al. Mol. Cancer Ther. (2017) pmid:
- Goodman AM, et al. Cancer Immunol Res (2019) pmid: 38. 31405947
- 39. Marabelle et al., 2019; ESMO Abstract 11920
- 40. Cristescu R. et al. Science (2018) pmid: 30309915
- 41. Rizvi et al., 2017; WCLC Abstract 1106
- 42. Hodi et al., 2019; AACR abstract CT037
- 43. Lee et al., 2019; ASCO Abstract 641
- 44. Ready N, et al. J. Clin. Oncol. (2019) pmid: 30785829
- 45. Hellmann MD, et al. N. Engl. J. Med. (2018) pmid: 29658845
- Hellmann MD, et al. Cancer Cell (2018) pmid: 29657128
- 47. Hellmann MD, et al. Cancer Cell (2018) pmid: 29731394
- 48. Rozeman EA, et al. Nat Med (2021) pmid: 33558721 49. Sharma P, et al. Cancer Cell (2020) pmid: 32916128
- 50. Marabelle A, et al. Lancet Oncol. (2020) pmid:

- 32919526
- 51. Ott PA, et al. J. Clin. Oncol. (2019) pmid: 30557521
- 52. Cristescu R, et al. J Immunother Cancer (2022) pmid:
- Friedman CF, et al. Cancer Discov (2022) pmid: 34876409
- 54. Sturgill EG, et al. Oncologist (2022) pmid: 35274716
- 55. Schenker at al., 2022; AACR Abstract 7845
- 56. Chalmers ZR, et al. Genome Med (2017) pmid: 28420421
- 57. Strickland KC, et al. Oncotarget (2016) pmid: 26871470
- 58. Pfeifer GP, et al. Mutat. Res. (2005) pmid: 15748635
- 59. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) pmid: 23875803
- **60.** Pfeifer GP, et al. Oncogene (2002) pmid: 12379884
- 61. Rizvi NA, et al. Science (2015) pmid: 25765070
- 62. Johnson BE, et al. Science (2014) pmid: 24336570
- 63. Choi S, et al. Neuro-oncology (2018) pmid: 29452419
- Cancer Genome Atlas Research Network, et al. Nature (2013) pmid: 23636398
- 65. Briggs S, et al. J. Pathol. (2013) pmid: 23447401
- Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) pmid: 24583393
- 67. Nature (2012) pmid: 22810696
- Roberts SA, et al. Nat. Rev. Cancer (2014) pmid: 25568919
- 69. Hirai H, et al. Cancer Biol. Ther. (2010) pmid: 20107315
- 70. Bridges KA, et al. Clin. Cancer Res. (2011) pmid: 21799033
- Rajeshkumar NV, et al. Clin. Cancer Res. (2011) pmid: 21389100
- 72. Osman AA, et al. Mol. Cancer Ther. (2015) pmid:
- 73. Leung et al., 2021; ASCO Abstract 4139
- 74. Xu L, et al. Mol. Cancer Ther. (2002) pmid: 12489850
- 75. Xu L, et al. Mol. Med. (2001) pmid: 11713371
- 76. Camp ER, et al. Cancer Gene Ther. (2013) pmid: 23470564
- 77. Kim SS, et al. Nanomedicine (2015) pmid: 25240597
- 78. Pirollo KF, et al. Mol. Ther. (2016) pmid: 27357628
- 79. Leijen S, et al. J. Clin. Oncol. (2016) pmid: 27601554
- 80. Moore et al., 2019; ASCO Abstract 5513
- 81. Embaby A, et al. Gynecol Oncol (2023) pmid: 37236033
- 82. Oza et al., 2015; ASCO Abstract 5506
- 83. Lee J, et al. Cancer Discov (2019) pmid: 31315834
- 84. Méndez E, et al. Clin. Cancer Res. (2018) pmid: 29535125
- 85. Seligmann JF, et al. J Clin Oncol (2021) pmid: 34538072
- 86. Gourley et al., 2016; ASCO Abstract 5571
- 87. Park H, et al. ESMO Open (2022) pmid: 36084396
- 88. Nguyen B, et al. Cell (2022) pmid: 35120664
- 89. Ahmed AA, et al. J. Pathol. (2010) pmid: 20229506
- 90. Wojnarowicz PM, et al. PLoS ONE (2012) pmid: 23029043
- 91. Karst AM, et al. J Oncol (2010) pmid: 19746182
- 92. Gadducci A, et al. Gynecol. Endocrinol. (2012) pmid: 22304686
- 93. Rechsteiner M, et al. Exp. Mol. Pathol. (2013) pmid: 23965232
- 94. McConechy MK, et al. Mod. Pathol. (2014) pmid: 23765252
- Mira Navarro J, et al. An Esp Pediatr (1988) pmid: 3178070
- 96. Ryland GL, et al. Genome Med (2015) pmid: 26257827 97. Cheasley D, et al. Nat Commun (2019) pmid: 31477716
- 98. Hollis RL, et al. Nat Commun (2020) pmid: 33020491

- 99. Altman AD, et al. Mod. Pathol. (2013) pmid: 23558569
- Giurgea LN, et al. Rom J Morphol Embryol (2012) pmid: 23303020
- Rajesh NG, et al. Indian J Pathol Microbiol (2007) pmid: 101. 17883046
- 102. Lee S, et al. Int. J. Gynecol. Pathol. (2013) pmid: 24071870
- 103. de Graeff P, et al. Br. J. Cancer (2009) pmid: 19513073
- 104. Brown CJ, et al. Nat. Rev. Cancer (2009) pmid: 19935675
- Joerger AC, et al. Annu. Rev. Biochem. (2008) pmid: 18410249
- 106. Kato S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) pmid:
- 107. Kamada R, et al. J. Biol. Chem. (2011) pmid: 20978130
- Zerdoumi Y, et al. Hum. Mol. Genet. (2017) pmid: 108. 28472496
- 109. Yamada H, et al. Carcinogenesis (2007) pmid: 17690113 Landrum MJ, et al. Nucleic Acids Res. (2018) pmid:
- 29165669
- 111. Bougeard G, et al. J. Clin. Oncol. (2015) pmid: 26014290 112. Sorrell AD, et al. Mol Diagn Ther (2013) pmid: 23355100
- Nichols KE, et al. Cancer Epidemiol. Biomarkers Prev. (2001) pmid: 11219776
- 114. Kleihues P, et al. Am. J. Pathol. (1997) pmid: 9006316
- Gonzalez KD, et al. J. Clin. Oncol. (2009) pmid: 19204208
- 116. Lalloo F, et al. Lancet (2003) pmid: 12672316
- 117. Mandelker D, et al. Ann. Oncol. (2019) pmid: 31050713 118. Jaiswal S, et al. N. Engl. J. Med. (2014) pmid: 25426837
- Genovese G, et al. N. Engl. J. Med. (2014) pmid: 25426838
- 120. Xie M, et al. Nat. Med. (2014) pmid: 25326804
- Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) pmid: 28669404
- Severson EA, et al. Blood (2018) pmid: 29678827
- 123. Fuster JJ, et al. Circ. Res. (2018) pmid: 29420212 124. Hematology Am Soc Hematol Educ Program (2018) pmid: 30504320
- 125. Chabon JJ, et al. Nature (2020) pmid: 32269342
- Razavi P, et al. Nat. Med. (2019) pmid: 31768066 Konecny GE, et al. Clin. Cancer Res. (2011) pmid:
- 21278246 Katsumi Y, et al. Biochem. Biophys. Res. Commun. (2011) pmid: 21871868 128
- 129. Cen L, et al. Neuro-oncology (2012) pmid: 22711607
- 130. Logan JE, et al. Anticancer Res. (2013) pmid: 23898052
- 131. Fennell DA, et al. Lancet Oncol (2022) pmid: 35157829
- 132. Elvin JA, et al. Oncologist (2017) pmid: 28283584
- 133. Gao J, et al. Curr Oncol (2015) pmid: 26715889 134. Gopalan et al., 2014; ASCO Abstract 8077
- 135. Peguero et al., 2016; ASCO Abstract 2528
- Konecny et al., 2016; ASCO Abstract 5557 136. DeMichele A, et al. Clin. Cancer Res. (2015) pmid: 137.
- 25501126
- Finn RS, et al. Lancet Oncol. (2015) pmid: 25524798 138 Infante JR, et al. Clin. Cancer Res. (2016) pmid:
- 27542767 Johnson DB, et al. Oncologist (2014) pmid: 24797823
- 141. Tate JG, et al. Nucleic Acids Res. (2019) pmid: 30371878
- 142. Ozdemir F, et al. Exp Ther Med (2012) pmid: 23226780 Abou-Zeid AA, et al. Scand. J. Clin. Lab. Invest. (2011)
- pmid: 21728901 144. Fujita M. et al. Int. J. Cancer (1997) pmid: 9133447
- Niederacher D, et al. Br. J. Cancer (1999) pmid:
- 146. Schlosshauer PW, et al. Int. J. Gynecol. Pathol. (2011) pmid: 21131838

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy

APPENDIX

References

- ORDERED TEST # ORD-1795940-01
- 147. Bilyk OO, et al. Exp. Oncol. (2011) pmid: 21956468
 148. Giordano G, et al. Pathol. Res. Pract. (2008) pmid: 18180113
- Khouja MH, et al. Int. J. Gynecol. Pathol. (2007) pmid: 17885492
- **150.** Surowiak P, et al. Histol. Histopathol. (2008) pmid: 18283637
- **151.** Lee YH, et al. Int. J. Gynecol. Pathol. (2011) pmid: 21464733
- **152.** Bali A, et al. Clin. Cancer Res. (2004) pmid: 15297421
- 153. Milea A. et al. Mod. Pathol. (2014) pmid: 24336157
- **154.** Quelle DE, et al. Cell (1995) pmid: 8521522
- 155. Mutat. Res. (2005) pmid: 15878778
- 156. Gazzeri S, et al. Oncogene (1998) pmid: 9484839
- 157. Oncogene (1999) pmid: 10498883
- 158. Sherr CJ, et al. Cold Spring Harb. Symp. Quant. Biol. (2005) pmid: 16869746
- 159. Ozenne P, et al. Int. J. Cancer (2010) pmid: 20549699
- 160. Ruas M, et al. Oncogene (1999) pmid: 10498896
- **161.** Jones R, et al. Cancer Res. (2007) pmid: 17909018
- **162.** Haferkamp S, et al. Aging Cell (2008) pmid: 18843795
- **163.** Huot TJ, et al. Mol. Cell. Biol. (2002) pmid: 12417717 **164.** Rizos H, et al. J. Biol. Chem. (2001) pmid: 11518711
- 104. KIZOS II, et al. J. Biol. Chem. (2001) piniu. 11518/11
- 165. Gombart AF, et al. Leukemia (1997) pmid: 9324288
- 166. Yang R, et al. Cancer Res. (1995) pmid: 7780957167. Parry D, et al. Mol. Cell. Biol. (1996) pmid: 8668202
- 168. Greenblatt MS, et al. Oncogene (2003) pmid: 12606942
- Yarbrough WG, et al. J. Natl. Cancer Inst. (1999) pmid: 10491434
- 170. Poi MJ, et al. Mol. Carcinog. (2001) pmid: 11255261
- 171. Byeon IJ, et al. Mol. Cell (1998) pmid: 9660926
- Kannengiesser C, et al. Hum. Mutat. (2009) pmid: 19260062
- **173.** Lal G, et al. Genes Chromosomes Cancer (2000) pmid: 10719365

- 174. Koh J, et al. Nature (1995) pmid: 7777061
- 175. McKenzie HA, et al. Hum. Mutat. (2010) pmid: 20340136
- 176. Miller PJ, et al. Hum. Mutat. (2011) pmid: 21462282
- 177. Kutscher CL, et al. Physiol. Behav. (1977) pmid: 905385
- 178. Scaini MC, et al. Hum. Mutat. (2014) pmid: 24659262
- 179. Jenkins NC, et al. J. Invest. Dermatol. (2013) pmid: 23190892
- 180. Walker GJ, et al. Int. J. Cancer (1999) pmid: 10389768
- 181. Rutter JL, et al. Oncogene (2003) pmid: 12853981
- 182. Whelan AJ, et al. N Engl J Med (1995) pmid: 7666917
- 183. Adv Exp Med Biol (2010) pmid: 20687502
- **184.** Hogg D, et al. J Cutan Med Surg (1998) pmid: 9479083
- **185.** De Unamuno B, et al. Melanoma Res (2018) pmid: 29543703
- Soura E, et al. J Am Acad Dermatol (2016) pmid: 26892650
- Huerta C, et al. Acta Derm Venereol (2018) pmid: 29405243
- 188. Kaufman DK, et al. Neurology (1993) pmid: 8414022
- 189. Bahuau M, et al. Cancer Res (1998) pmid: 9622062
- 190. Chan AK, et al. Clin Neuropathol () pmid: 28699883
- 191. Cerami E, et al. Cancer Discov (2012) pmid: 22588877
- 192. Gao J, et al. Sci Signal (2013) pmid: 23550210
- 193. Br. J. Haematol. (2011) pmid: 21118195
- **194.** Muntean AG, et al. Annu Rev Pathol (2012) pmid: 22017583
- 195. Tang G, et al. Hum. Pathol. (2015) pmid: 25387813
- 196. Andersen MK, et al. Genes Chromosomes Cancer (2001) pmid: 11284033
- Dolan M, et al. Cancer Genet. Cytogenet. (2002) pmid: 12034519
- 198. Fabbro M, et al. Gynecol. Oncol. (2019) pmid: 30638768
- 199. González-Martín A, et al. N. Engl. J. Med. (2019) pmid: 31562799

- 200. Moore KN, et al. Lancet Oncol. (2019) pmid: 30948273
- 201. Mirza MR, et al. Lancet Oncol. (2019) pmid: 31474354
- 202. Konstantinopoulos PA, et al. JAMA Oncol (2019) pmid: 31194228
- 203. Fong PC, et al. N. Engl. J. Med. (2009) pmid: 19553641
- 204. Gelmon KA, et al. Lancet Oncol. (2011) pmid: 21862407
- 205. Domchek SM, et al. Gynecol. Oncol. (2016) pmid: 26723501
- 206. Matulonis UA, et al. Ann. Oncol. (2016) pmid: 26961146
- 207. Fong PC, et al. J. Clin. Oncol. (2010) pmid: 20406929
- 208. Moore K, et al. N. Engl. J. Med. (2018) pmid: 30345884
- 209. Poveda A, et al. Lancet Oncol (2021) pmid: 33743851 210. Pujade-Lauraine E, et al. Lancet Oncol. (2017) pmid:
- 28754483
- 211. Ledermann JA, et al. Lancet Oncol. (2016) pmid: 27617661
- **212.** Ledermann J, et al. N. Engl. J. Med. (2012) pmid: 22452356
- 213. Ledermann J, et al. Lancet Oncol. (2014) pmid: 24882434
- 214. DiSilvestro P, et al. J Clin Oncol (2022) pmid: 36082969
- 215. DiSilvestro et al., 2022; ESMO Abstract 5170
- 216. Pujade-Lauraine et al., 2021; ESMO Abstract LBA33
- 217. Pignata S, et al. Gynecol Oncol (2023) pmid: 37030280
- **218.** Ray-Coquard I, et al. N Engl J Med (2019) pmid: 31851799
- 219. Ray-Coquard et al., 2022; ESMO Abstract LBA29
- 220. Oza AM, et al. Lancet Oncol. (2015) pmid: 25481791
- **221.** Liu JF, et al. Lancet Oncol. (2014) pmid: 25218906
- 222. Liu JF, et al. J Clin Oncol (2022) pmid: 35290101
- 223. Wethington et al., 2021; ASCO Abstract 5516
- **224.** Westin et al., 2021; ASCO Abstract 5505
- 225. Monk BL et al. I Clin Oncol (2022) pmid: 35658487
- **226.** de Bono J, et al. Cancer Discov (2017) pmid: 28242752
- 227. Piha Paul et al., 2018; AACR abstract A096