

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

| | | | | | |
|----------------|---|------------------|--|-----------------|--|
| PATIENT | DISEASE Uterus endometrial adenocarcinoma papillary serous | PHYSICIAN | ORDERING PHYSICIAN Yeh, Yi-Chen | SPECIMEN | SPECIMEN SITE Uterus |
| | NAME Hsieh, Yu-Ying | | MEDICAL FACILITY Taipei Veterans General Hospital | | SPECIMEN ID S112-69102E (PF24003) |
| | DATE OF BIRTH 21 September 1963 | | ADDITIONAL RECIPIENT None | | SPECIMEN TYPE Slide Deck |
| | SEX Female | | MEDICAL FACILITY ID 205872 | | DATE OF COLLECTION 29 December 2023 |
| | MEDICAL RECORD # 33045679 | | PATHOLOGIST Not Provided | | SPECIMEN RECEIVED 09 January 2024 |

Biomarker Findings

Microsatellite status - MS-Stable
Tumor Mutational Burden - 7 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

KIT amplification - equivocal[†]
PDGFRA amplification
CCNE1 amplification
FBXW7 R479Q
MDM2 amplification
MYC amplification
BRAF D594N
FUBP1 deletion exons 2-11
KDR amplification - equivocal[†]
MED12 G44D
SPOE E46K
TP53 R248W

[†] See About the Test in appendix for details.

Report Highlights

- Targeted therapies with potential clinical benefit **approved in another tumor type**: Imatinib (p. 13), Nilotinib (p. 13), Sorafenib (p. 14), Sunitinib (p. 14)
- Evidence-matched **clinical trial options** based on this patient's genomic findings: (p. 15)
- Variants with **prognostic implications** for this tumor type that may impact treatment decisions: **Microsatellite status MS-Stable** (p. 3), **TP53 R248W** (p. 11)

BIOMARKER FINDINGS

Microsatellite status - MS-Stable

Tumor Mutational Burden - 7 Muts/Mb

GENOMIC FINDINGS

KIT - amplification - equivocal

10 Trials see p. 19

PDGFRA - amplification

1 Trial see p. 24

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. See Biomarker Findings section

No therapies or clinical trials. See Biomarker Findings section

| THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE) | THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE) |
|--|--|
| none | Imatinib |
| | Nilotinib |
| | Sorafenib |
| | Sunitinib |
| none | Imatinib |
| | |

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

| GENOMIC FINDINGS | THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE) | THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE) |
|------------------------------|--|--|
| CCNE1 - amplification | none | none |
| 7 Trials see p. 15 | | |
| FBXW7 - R479Q | none | none |
| 6 Trials see p. 17 | | |
| MDM2 - amplification | none | none |
| 2 Trials see p. 21 | | |
| MYC - amplification | none | none |
| 10 Trials see p. 22 | | |

GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

| | | | |
|--|------|---------------------|-------|
| BRAF - D594N | p. 8 | MED12 - G44D | p. 9 |
| FUBP1 - deletion exons 2-11 | p. 8 | SPOP - E46K | p. 10 |
| KDR - amplification - equivocal | p. 9 | TP53 - R248W | p. 11 |

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

BIOMARKER FINDINGS
BIOMARKER

Microsatellite status

RESULT

MS-Stable

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁻³, including approved therapies nivolumab and pembrolizumab⁴⁻⁵. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, $p=0.001$)⁶.

FREQUENCY & PROGNOSIS

MSS has been reported in 73-89% of endometrial cancers⁷⁻¹⁴. TP53 mutations in microsatellite stable/mismatch repair-proficient (MMRp) POLE-wildtype endometrial carcinoma are indicative of a ProMisE/TCGA molecularly classified subtype, "p53abn" (NCCN Uterine Neoplasms Guidelines, v1.2023) (^{7,15-21}), which is associated with high risk of recurrence²², yet also with the highest degree of benefit from adjuvant chemoradiotherapy over radiotherapy relative to the other subtypes²³. Additionally, classification as this molecular subtype refines FIGO disease stage to IIC (Stage "IICm-p53abn") for patients with histologically determined Stage I/II early endometrial cancer that is locally confined to the uterine corpus or with myometrial invasion²⁴. Data regarding the role of microsatellite instability on prognosis and survival in endometrial cancer are mixed^{8-10,12-13,25-29}, although these studies often evaluated endometrial cancers of all International Federation of

Gynecology and Obstetrics (FIGO) stages together.

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA MMR in the tumor³⁰. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2³⁰⁻³². This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers³³⁻³⁵. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{30,32,34-35}.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

BIOMARKER FINDINGS

BIOMARKER

Tumor Mutational Burden

RESULT

7 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1³⁶⁻³⁹, anti-PD-1 therapies³⁷⁻⁴¹, and combination nivolumab and ipilimumab⁴²⁻⁵⁰. In multiple pan-tumor studies, increased tissue tumor mutational burden (TMB) was associated with sensitivity to immune checkpoint inhibitors^{36-39,41,51-55}. In the KEYNOTE 158 trial of pembrolizumab monotherapy for patients with solid tumors, significant improvement in ORR was observed for patients with TMB ≥ 10 Muts/Mb (as measured by this assay) compared with those with TMB < 10 Muts/Mb in a large cohort that included multiple tumor types⁵¹; similar findings were observed in the KEYNOTE 028 and 012 trials⁴¹. At the same TMB cutpoint, retrospective analysis of patients with solid tumors treated with any checkpoint inhibitor identified that tissue TMB scores ≥ 10 Muts/Mb were associated with

prolonged time to treatment failure compared with scores < 10 muts/Mb (HR=0.68)⁵⁵. For patients with solid tumors treated with nivolumab plus ipilimumab in the CheckMate 848 trial, improved responses were observed in patients with a tissue TMB ≥ 10 Muts/Mb independent of blood TMB at any cutpoint in matched samples⁵⁶. However, support for higher TMB thresholds and efficacy was observed in the prospective Phase 2 MyPathway trial of atezolizumab for patients with pan-solid tumors, where improved ORR and DCR was seen in patients with TMB ≥ 16 Muts/Mb than those with TMB ≥ 10 and < 16 Muts/Mb⁵⁴. Similarly, analyses across several solid tumor types reported that patients with higher TMB (defined as $\geq 16-20$ Muts/Mb) achieved greater clinical benefit from PD-1 or PD-L1-targeting monotherapy compared with patients with higher TMB treated with chemotherapy³⁶ or those with lower TMB treated with PD-1 or PD-L1-targeting agents³⁸.

FREQUENCY & PROGNOSIS

A large-scale genomic analysis found that endometrial adenocarcinomas harbored a median TMB of 4.5 Muts/Mb, and 15% of cases had an elevated TMB of greater than 20 Muts/Mb⁵⁷. Another study evaluating TMB in endometrial adenocarcinoma reported that 24% of tumors had a mutational burden of greater than 10.4 Muts/Mb⁵⁸. Increased tumor mutational burden (TMB) in endometrial carcinoma has been correlated with POLE mutation and advanced high-grade

endometrioid subtypes^{7,14,59-60}. Ultramutated endometrial tumors (elevated TMB with POLE mutations) have also been associated with improved PFS⁷. The same study associated lower mutational burden, independent of PD-L1 status, in endometrial carcinomas with poorer prognosis⁷. For patients with advanced microsatellite-stable endometrial carcinoma not treated with immunotherapy, OS did not significantly differ between patients with TMB-high (≥ 10 Muts/Mb) and TMB-low (11.4 vs. 13.5 months, adjusted HR=1.15) in 1 study⁶¹.

FINDING SUMMARY

Tumor mutational burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitutions and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma⁶²⁻⁶³ and cigarette smoke in lung cancer⁶⁴⁻⁶⁵, treatment with temozolomide-based chemotherapy in glioma⁶⁶⁻⁶⁷, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes^{7,68-71}, and microsatellite instability^{7,70-71}. This sample harbors a TMB level associated with lower rates of clinical benefit from treatment with PD-1- or PD-L1-targeting immune checkpoint inhibitors compared with patients with tumors harboring higher TMB levels, based on several studies in multiple solid tumor types^{38-39,51}.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

GENOMIC FINDINGS

GENE
KIT

ALTERATION
amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence, primarily in gastrointestinal stromal tumor (GIST), melanoma, AML, and systemic mastocytosis, KIT activating alterations are associated with sensitivity to TKIs including imatinib, sunitinib, sorafenib, dasatinib, nilotinib, pazopanib, regorafenib, ponatinib, midostaurin, apatinib, avapritinib, and ripretinib⁷²⁻⁸⁶. The use of mTOR inhibitors as an alternative therapeutic strategy has demonstrated limited success in KIT-mutated, imatinib-resistant melanoma, with 1 PR and 3 SD observed for 4

patients treated with everolimus⁸⁷⁻⁸⁸. However, no responses were observed for 10 patients with mastocytosis following everolimus monotherapy, with 8/10 patients harboring the KIT D816V mutation⁸⁹. The role of KIT amplification as a biomarker for response to mTOR inhibitors has not been investigated (PubMed, Feb 2023). Clinical benefit has been observed for patients with KIT amplified or overexpressing tumors following treatment with imatinib⁹⁰⁻¹⁰⁰, nilotinib¹⁰¹, sorafenib¹⁰²⁻¹⁰⁵, and sunitinib¹⁰⁶⁻¹⁰⁷, suggesting that KIT amplification may be sensitive to these inhibitors. However, evidence demonstrating clinical benefit for regorafenib, dasatinib, pazopanib, or ponatinib in the context of KIT amplified or overexpressing tumors is limited.

FREQUENCY & PROGNOSIS

KIT amplifications are common in various solid tumors, most often seen in mucosal melanomas (8-10%), adenoid cystic carcinomas (4.0%), small

cell lung cancer (2.7%), and seminomas (2.1%)¹⁰⁸. KIT expression has been reported in 58% (42/72) of endometrial adenocarcinomas in one study¹⁰⁹. KIT expression was correlated with worse overall survival for patients with endometrial cancer¹⁰⁹. Published data investigating the prognostic implications of KIT alterations in endometrial cancer are limited (PubMed, Jun 2023).

FINDING SUMMARY

KIT (also called c-KIT) encodes a cell surface tyrosine kinase receptor that, upon ligand binding and dimerization, activates the PI3K-AKT and RAS-MAPK signaling pathways¹¹⁰. KIT aberrations, including point mutations, translocations, amplification, and overexpression, have been associated with various malignancies, and KIT is considered an oncoprotein¹¹¹. KIT has been reported to be amplified in cancer¹¹² and may be biologically relevant in this context¹¹³⁻¹¹⁴.

GENE
PDGFRA

ALTERATION
amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of limited evidence of clinical benefit for patients with increased PDGFR expression, PDGFRA amplification may be associated with sensitivity to imatinib¹¹⁵⁻¹¹⁶.

FREQUENCY & PROGNOSIS

In the Uterine Corpus Endometrial Carcinoma Provisional TCGA dataset, putative high-level amplification of PDGFRA has been reported in fewer than 1% of cases⁷. High expression of PDGFR-alpha has been reported in both primary and recurrent endometrioid and uterine papillary serous endometrial carcinomas, and has been shown to be increased in recurrent as compared to primary cases¹¹⁷. Increased PDGFR-alpha expression has been correlated with lower grade (grades 1 and 2) endometrial carcinomas¹¹⁷.

FINDING SUMMARY

PDGFRA encodes platelet-derived growth factor receptor alpha (PDGFR-alpha), a tyrosine kinase receptor that, upon binding of cognate ligands (PDGFA or PDGFB), activates several signaling pathways, including PI3K and MAPK¹¹⁸. PDGFR aberrations, including point mutations, translocations, amplification, and/or overexpression, have been associated with various malignancies¹¹¹. Amplification of PDGFRA, frequently occurring with amplification of the genes KDR and KIT, has been associated with increased PDGFRA expression¹¹⁹⁻¹²² and poor prognosis^{119,123-125} in some subtypes of glioma.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

GENOMIC FINDINGS

GENE
CCNE1

ALTERATION
amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no approved therapies that directly target CCNE1 alterations. Because amplification or overexpression of CCNE1 leads to increased genomic instability through the ATR-CHK1-WEE1 pathway¹²⁶⁻¹²⁷ and cyclin E1 promotes cell cycle progression in a complex with CDK2¹²⁸, clinical and preclinical studies have investigated inhibitors of ATR, CDK2, CHK1, HDAC, PKMYT1, and WEE1 as potential therapeutic approaches for tumors with CCNE1 activation. Clinical benefit has been reported for patients with recurrent high-grade serous ovarian carcinoma (HGSOC) with CCNE1 amplification or expression in response to treatment with the CHK1 inhibitor prexasertib¹²⁹. Studies of the WEE1 inhibitor adavosertib

observed PRs in patients with CCNE1-amplified HGSOC and ovarian cancer¹³⁰⁻¹³². Similarly, in a Phase 2 study of patients with CCNE1-amplified solid tumors, adavosertib elicited an ORR of 27% with PRs reported for patients with ovarian cancer, urothelial carcinoma, or melanoma; median PFS was 4.1 months and median OS was 9.9 months¹³²⁻¹³³. One study has reported a reduction in tumor CCNE1 levels in 4/6 lung and esophageal cancer cases following treatment with the HDAC inhibitor vorinostat¹³⁴. Preclinical studies have demonstrated that cell lines and murine models with CCNE1 amplification or overexpression were sensitive to inhibitors of ATR¹³⁵⁻¹³⁶, CDK2¹³⁷, PKMYT1¹³⁸⁻¹³⁹, or WEE1^{127,140}. However, other studies have shown that sensitivity of various cell lines to CDK2 inhibitors, including SNS-032, dinaciclib, and seliciclib, at clinically achievable doses, is largely independent of CCNE1 copy number or expression¹⁴¹⁻¹⁴⁴.

FREQUENCY & PROGNOSIS

In the Uterine Corpus Endometrial Carcinoma TCGA dataset, putative amplification of CCNE1 has been found in 6% of cases⁷. CCNE1

amplification has been observed in 41% (11/27) of endometrial intraepithelial carcinomas¹⁴⁵ and in a study of ovarian cancer, CCNE1 amplification was observed in 12 of 12 serous carcinoma samples and in 2 of 6 endometrioid carcinomas¹⁴⁶. Elevated cyclin E1 protein expression has been associated with poor prognosis in several cancer types, including endometrial and ovarian cancer¹⁴⁶⁻¹⁵⁰.

FINDING SUMMARY

CCNE1 encodes the protein cyclin E1, which plays a role in the regulated transition from the G1 to S phase by binding to and activating cyclin-dependent protein kinase 2 (CDK2). It also has a direct role in initiation of replication and the maintenance of genomic stability¹²⁸. Amplification of chromosomal region 19q12-q13 has been demonstrated in many types of cancer, and CCNE1 is a well-studied gene within this amplicon¹⁵¹⁻¹⁵². Increased copy number of CCNE1 is highly associated with overexpression of the cyclin E1 protein^{146,150}. Cyclin E1 overexpression can lead to cell transformation as a result of an increase in cyclin E1 activity^{128,153}.

GENE
FBXW7

ALTERATION
R479Q

HGVS VARIANT
NM_033632.3:c.1436G>A (p.R479Q)

VARIANT CHROMOSOMAL POSITION
chr4:153247366

VARIANT ALLELE FREQUENCY (% VAF)
16.5%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

FBXW7 inactivating alterations may indicate sensitivity to mTOR inhibitors¹⁵⁴⁻¹⁵⁵. Case series reported objective responses for 2 patients with FBXW7-mutated cervical squamous cell carcinoma treated with everolimus¹⁵⁶.

FREQUENCY & PROGNOSIS

FBXW7 mutations have been reported in up to 14% of endometrial cancers^{7,157-158}. In primary endometrial carcinomas, FBXW7 mutations correlated with lymph node involvement¹⁵⁹. Reduced FBXW7 expression has been associated with poor prognosis in some cancers such as

colorectal cancer, gastric cancer, esophageal SCC, cervical SCC, melanoma, non-small cell lung carcinoma, and osteosarcoma¹⁶⁰⁻¹⁶⁸.

FINDING SUMMARY

FBXW7 encodes the F-box protein subunit of the SCF ubiquitin ligase complex, which targets proteins for degradation¹⁶⁹. FBXW7 inactivation is associated with chromosomal instability and with stabilization of proto-oncogenes, such as mTOR, MYC, cyclin E, NOTCH, and JUN; FBXW7 is therefore considered a tumor suppressor¹⁶⁹⁻¹⁷⁰. Alterations such as seen here may disrupt FBXW7 function or expression¹⁷⁰⁻¹⁷⁷.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

GENOMIC FINDINGS

GENE

MDM2

ALTERATION
amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

MDM2 antagonists disrupt the MDM2-p53 interaction, thereby stabilizing p53¹⁷⁸. Preclinical studies have suggested that the amplification of MDM2, in the absence of concurrent TP53 mutations, may increase sensitivity to these agents¹⁷⁹⁻¹⁸⁰. Preliminary Phase 1 studies of the MDM2-p53 antagonist alrizomadlin (APG-115) reported a PR in a patient with liposarcoma harboring an MDM2 amplification and wildtype for TP53 and SD in 21%-38% (6/28 and 5/13, respectively) of patients in genomically unselected solid tumors¹⁸¹⁻¹⁸². A Phase 2 trial of alrizomadlin in combination with pembrolizumab reported a PR

in 1 of 3 patients with malignant peripheral nerve sheath tumor that had failed standard therapy, as well as PRs in patients with multiple types of solid tumors that had failed immunotherapy, including 1 out of 14 patients with non-small cell lung cancer; 1 out of 5 patients with urothelial carcinoma; and 2 out of 5, 1 out of 5, and 1 out of 11 patients with mucosal, uveal, and cutaneous melanoma, respectively¹⁸³. Phase 1b studies of the MDM2 inhibitor idasanutlin for refractory AML in combination with cytarabine or venetoclax reported anti-leukemic response rates of 33% (25/75) and 37% (11/30), respectively¹⁸⁴⁻¹⁸⁵; clinical benefit (58% ORR, 7/12) with idasanutlin monotherapy has been reported for patients with polycythemia vera¹⁸⁶. The dual MDM2/MDM4 inhibitor ALRN-6924 led to an ORR of 27% (4/15) for patients with TP53 wildtype peripheral T-cell lymphoma in a Phase 2 study¹⁸⁷; responses have also been observed in TP53 wildtype AML, MDS, Merkel cell carcinoma, colorectal cancer, and liposarcoma¹⁸⁸⁻¹⁸⁹.

FREQUENCY & PROGNOSIS

MDM2 amplification is common in various solid tumors and is often seen in soft tissue sarcomas (19%), bladder (7.8%), bone (7.5%), glioma (6.3%), non-small cell lung (5.4%), hepatobiliary (4.5%), esophagogastric (4.4%), breast (4.1%), and germ cell tumor (3.8%) cancer¹⁵⁷. Expression of MDM2 protein has been identified in 42-70% (15/36-19/27) of endometrioid endometrial carcinomas and 36-67% (9/25-8/12) of papillary serous endometrial carcinomas¹⁹⁰⁻¹⁹¹. One study observed that MDM2 protein expression was significantly associated with shorter overall survival (hazard ratio of 6) and an independent prognostic factor in high-risk endometrial cancer¹⁹⁰.

FINDING SUMMARY

MDM2 encodes an E3 ubiquitin protein ligase, which mediates the ubiquitination and subsequent degradation of p53, Rb1, and other proteins¹⁹²⁻¹⁹⁴. MDM2 has been reported to be amplified in cancer¹¹⁴ and may be biologically relevant in this context¹⁹⁵⁻¹⁹⁶.

GENE

MYC

ALTERATION
amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Limited clinical data indicates that MYC activation may predict sensitivity to the pan-MYC inhibitor OMO-103; a Phase 1 study for patients with solid tumors reported 7 SDs (n=18), including 8% tumor reduction in a patient with pancreas adenocarcinoma¹⁹⁷. Preclinical data indicate MYC overexpression may predict sensitivity to investigational agents targeting CDK1¹⁹⁸⁻¹⁹⁹, CDK2²⁰⁰, Aurora kinase A²⁰¹⁻²⁰⁹, Aurora kinase B²¹⁰⁻²¹³, glutaminase²¹⁴⁻²¹⁷, or BET bromodomain-containing proteins²¹⁸⁻²²¹, as well as agents

targeting both HDAC and PI3K²²²⁻²²⁴. Exploratory biomarker analysis in a Phase 2 study reported a PFS benefit associated with a combination of the Aurora A kinase inhibitor alisertib and paclitaxel as second-line therapy for patients with MYC-overexpressed small cell lung cancer, but not for patients without MYC overexpression²²⁵. A PR was reported for a patient with MYC-amplified invasive ductal breast carcinoma treated with an unspecified Aurora kinase inhibitor and taxol²²⁶.

— Nontargeted Approaches —

MYC amplification has also been suggested to predict response to chemotherapy in patients with breast cancer in some studies²²⁷⁻²²⁸. Preclinical evidence suggests that colon cancer cells with MYC amplification may be more sensitive to 5-fluorouracil and paclitaxel²²⁹⁻²³⁰.

FREQUENCY & PROGNOSIS

In the Uterine Corpus Endometrioid Carcinoma TCGA dataset, putative high-level amplification of MYC has been found in 8% of cases⁷. In the scientific literature, MYC amplification has been detected in 15-27% of endometrial cancer cases²³¹⁻²³⁴. In one study, MYC amplification has been associated with higher tumor grade in uterine corpus cancers²³⁴.

FINDING SUMMARY

MYC (c-MYC) encodes a transcription factor that regulates many genes related to cell cycle regulation and cell growth. It is an oncogene and may be activated in as many as 20% of cancers²³⁵. MYC dysregulation (amplification, overexpression, translocation) has been identified in a number of different cancer types²³⁶. MYC amplification has been significantly linked with increased mRNA and protein levels and results in the dysregulation of a large number of target genes^{235,237-238}.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

GENOMIC FINDINGS

GENE

BRAF

ALTERATION

D594N

HGVS VARIANT

NM_004333.4:c.1780G>A (p.D594N)

VARIANT CHROMOSOMAL POSITION

chr7:140453155

VARIANT ALLELE FREQUENCY (% VAF)

3.0%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Clinical outcomes for patients with activating BRAF alterations treated with BRAF and MEK inhibitors are most extensive at the V600 codon; outcomes are more limited for BRAF class-3 kinase-impaired or inactivating mutations such as 1 or more of the alterations seen here. A retrospective study of immunotherapies in non-small cell lung cancer (NSCLC) reported a 78% (7/9) DCR for patients with BRAF class-3

mutations²³⁹. MEK inhibitors alone or in combination with BRAF inhibitors may be efficacious in these alterations; a basket trial of the single-agent MEK inhibitor trametinib reported 1 PR, 8 SDs, and 9 PDs for these patients²⁴⁰, and combination therapies reported individual responses in other basket trials²⁴¹⁻²⁴². A retrospective analysis in melanoma with BRAF mutations reported PD as the best response in BRAF class-3 alterations for 2 patients treated with MEK inhibitors and 3 patients treated with RAF inhibitors²⁴³. The single-agent BRAF inhibitor vemurafenib was not effective in a Phase 2 trial in NSCLC, which reported no responses for 6 patients with class-3 BRAF alterations²⁴⁴; a basket trial of vemurafenib also observed no responses for these patients (n=3)²⁴⁵. Investigational BRAF inhibitors are also in development to target class-3 paradoxical activation²⁴⁶⁻²⁴⁷; a Phase 1a/1b trial of the pan-RAF inhibitor exarafenib reported 1 PR for a patient with a BRAF class-3 mutation²⁴⁶. A basket trial of the ERK inhibitor ulixertinib reported no responses and 3 SDs for patients across tumors with class-3 mutations²⁴⁸.

FREQUENCY & PROGNOSIS

BRAF mutations have been reported in 2.9% of endometrial carcinomas in the TCGA dataset⁷. In the literature, BRAF mutation has been generally been reported in 0-3% of endometrial carcinomas²⁴⁹⁻²⁵², with two reports of 11-21%²⁵³⁻²⁵⁴. Published data investigating the prognostic implications of BRAF alterations in endometrial carcinoma are limited (PubMed, Oct 2023).

FINDING SUMMARY

BRAF encodes a member of the RAF family of protein kinases, which includes ARAF, BRAF, and CRAF. These kinases function downstream of RAS as part of the MAPK (RAF-MEK-ERK) signaling cascade that facilitates cell proliferation, survival and transformation²⁵⁵⁻²⁵⁶. BRAF mutations have been reported in up to 20% of all cancers, with the majority of mutations occurring at the V600 position²⁵⁷⁻²⁵⁸. Alterations such as the class 3 mutation seen here have been shown to require concomitant upstream RAS activity in contrast with independently activating BRAF V600 or class 2 alterations²⁵⁹⁻²⁷², and may activate the MEK-ERK signaling pathway via CRAF^{259-261,268,273}.

GENE

FUBP1

ALTERATION

deletion exons 2-11

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Therapies targeting FUBP1 mutation directly or downstream effectors have not been tested preclinically or clinically in tumors that harbor FUBP1 mutations.

FREQUENCY & PROGNOSIS

FUBP1 alteration has been reported in 1.5% of samples analyzed in COSMIC, with the highest incidences reported in tumors of the endometrium (3.6%), central nervous system (3.2%), large intestine (3.0%), skin (2.8%), stomach (2.7%), and liver (2.6%) (COSMIC, Feb 2023)²⁷⁴. One study reported higher expression of FUBP1 in colorectal carcinoma tissues compared to adenoma and normal colon epithelial tissues²⁷⁵. A genetic signature defined by concomitant alterations in IDH1, CIC, and FUBP1 is associated with longer survival in patients with glioma²⁷⁶. FUBP1 has been shown to activate the expression of MYC²⁷⁷⁻²⁸⁰, activate p27KIP1²⁸¹, and regulate the

splicing of MDM2²⁸².

FINDING SUMMARY

FUBP1 encodes far upstream element binding protein 1 (also called FBP-1), a DNA-binding protein reported to have roles in transcriptional activation and splicing regulation of target genes. It is believed to act as an oncogene in some tumor types, such as hepatocellular carcinoma and non-small-cell lung cancer²⁸³⁻²⁸⁴, and as a tumor suppressor in others, particularly oligodendroglioma, for which mutations and/or loss of FUBP1 often co-occur with alterations in CIC or IDH1^{276,285-288}.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

GENOMIC FINDINGS

GENE

KDR

ALTERATION

amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

For patients with recurrent fibrosarcoma harboring unspecified KDR mutations, a retrospective study reported an ORR of 25% (7/28) and DCR of 82% (23/28) following apatinib monotherapy treatment²⁸⁹. On the basis of clinical benefit for patients with clear cell renal cell carcinoma (ccRCC)²⁹⁰⁻²⁹⁴ and a patient with breast angiosarcoma²⁹⁵, high VEGFR-2 expression has been associated with sensitivity to sunitinib. However, because data supporting concordance

between VEGFR-2 expression and KDR genomic biomarkers are limited, it is unclear whether these therapeutic strategies would be beneficial in this case. Limited preclinical data suggest that KDR activating mutations may increase sensitivity to VEGFR2-targeted agents such as linifanib²⁹⁶. On the basis of extensive clinical evidence across multiple tumor types, expression of plasma or tumor VEGFR-1 or VEGFR-2 has not been established as a reliable biomarker to predict response to the VEGFA-targeted agent bevacizumab²⁹⁷⁻³¹⁹.

FREQUENCY & PROGNOSIS

Amplification and mutation of KDR have been reported in fewer than 1% and 0.5-3.8% of endometrial cases, respectively^{7,158}. VEGFR-2 expression in endometrial carcinomas has been reported in 15-68% of cases in the scientific literature³²⁰⁻³²³. In addition, increased levels of

VEGF have been found in endometrial cancer tissue and correlated with increased expression and activity of VEGFR-2^{320-321,324}. VEGFR-2 expression has been associated with tumor grade, tumor stage, and poor prognosis in endometrial carcinoma^{320,322-323,325-327}. In one study, serum VEGF levels, but not tissue VEGFR-2 levels, correlated with tumor metastasis³²⁸.

FINDING SUMMARY

KDR encodes vascular endothelial growth factor receptor 2 (VEGFR2), a member of the vascular endothelial growth factor receptor (VEGFR) family. It is a receptor tyrosine kinase that transmits signals from VEGFA and is involved in both tumor angiogenesis and vasculogenesis during development³²⁹. KDR amplification has been reported in many tumor types and may be oncogenic³²⁹.

GENE

MED12

ALTERATION

G44D

HGVS VARIANT

NM_005120.2:c.131G>A (p.G44D)

VARIANT CHROMOSOMAL POSITION

chrX:70339254

VARIANT ALLELE FREQUENCY (% VAF)

31.1%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no targeted therapies available to address genomic alterations in MED12. Uterine leiomyomas with tumorigenic MED12 alterations have been reported to overexpress IGF2³³⁰. Also, in a preclinical study, BRD4 inhibition has been shown to release the mediator complex from select

genomic sites and to inhibit mediator-mediated transcription³³¹, but whether BET inhibitors and/or IGF2 pathway inhibitors would be beneficial for patients with oncogenic MED12 alterations is unknown.

FREQUENCY & PROGNOSIS

MED12 alterations, consisting predominantly of exon 2 mutations such as G44D, have been frequently reported in uterine leiomyomas (43-70%)³³²⁻³⁴⁰, smooth muscle tumors of uncertain malignant potential (STUMP; 11%)³⁴¹⁻³⁴², breast phyllodes tumors (PTs; 8-74%)^{340,343-344}, and breast fibroadenomas (46-59%)^{340,345}. Similar alterations have also been reported in uterine leiomyosarcomas (2-30%)^{342,346-347}, and chronic lymphocytic leukemia (CLL; 5-8.8%)³⁴⁸⁻³⁴⁹. In prostate cancer, MED12 mutations, localized to the central part of the protein rather than exon 2, have been reported in 1-5% of cases³⁵⁰⁻³⁵². MED12 alterations are rare in other cancer types³⁴⁶. In breast PTs, MED12 exon 2 mutation was diagnostic

of breast PT compared to other spindle breast neoplasms³⁴³, correlated with decreased pathological grade in some studies³⁵³⁻³⁵⁴ but were independent of grade in others^{344,355}, and, according to one study, correlated with improved progression-free survival but increased risk of recurrence³³⁴. In one study, expression of an oncogenic MED12 exon 2 mutation in mice led to the development of leiomyomas and increased chromosome instability³⁵⁶.

FINDING SUMMARY

MED12 encodes a subunit of the multi-protein mediator complex, an important transcriptional regulator³⁵⁷⁻³⁵⁸. MED12/mediator has been implicated in regulating WNT/beta-catenin^{350,359}, hedgehog/GLI³⁶⁰, and TGF-beta³⁶¹⁻³⁶² signaling, although the mechanisms by which MED12 mutations facilitate tumorigenesis are still poorly defined. Tumorigenic MED12 mutations have been shown to disrupt the composition and/or activity of the mediator complex^{350,363-364}.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

GENOMIC FINDINGS

GENE

SPOP

ALTERATION

E46K

HGVS VARIANT

NM_003563.3:c.136G>A (p.E46K)

VARIANT CHROMOSOMAL POSITION

chr17:47699372

VARIANT ALLELE FREQUENCY (% VAF)

19.7%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

SPOP inactivation leads to accumulation of its substrate proteins, including the androgen receptor (AR)³⁶⁵, and SPOP inactivating alterations in prostate cancer have been associated with increased clinical benefit from AR signaling inhibitors such as abiraterone, apalutamide, and

enzalutamide³⁶⁶⁻³⁷⁰. It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

FREQUENCY & PROGNOSIS

In the large MSK MetTropism genomic study, SPOP mutation was most frequently observed in prostate cancer (12.5%), endometrial cancer (3.3%), ovarian cancer (1.1%), small bowel cancer (1.1%), gastrointestinal neuroendocrine cancer (0.7%), and bladder and breast cancer (<0.5%)¹⁰⁸. Loss of SPOP protein expression is commonly observed and is reported most frequently in gastric (30%), colorectal (20%), and prostate (37%) tumors³⁷¹. Published data investigating the prognostic implications of SPOP alterations in solid tumors other than prostate cancers are limited (PubMed, Mar 2023).

FINDING SUMMARY

SPOP encodes the substrate adapter of a ubiquitin

ligase complex, which mediates the polyubiquitination and hence degradation of its substrate proteins such as the androgen, progesterone, and estrogen receptors, the bromodomain and extraterminal domain (BET) proteins BRD2, BRD3, and BRD4, and the DNA-damage response protein 53BP1^{365,372-374}. SPOP is reported to function as a tumor suppressor in prostate cancer; however, it may play an oncogenic role in renal cell carcinoma (RCC)³⁷⁵⁻³⁷⁶. Most SPOP mutations in cancer cluster within the substrate-binding MATH domain (aa 31-161), and inactivating alterations in this domain inhibit the degradation of oncogenic protein substrates of SPOP^{365,374,377-378}. Although alterations such as observed here have not been characterized and their functional effect is unknown, they have been reported as hotspot mutations in prostate and endometrial cancers³⁷⁹⁻³⁸⁰. Multiple mutations within the meprin and TRAF homology (MATH) MATH domain have been characterized and found to inactivate SPOP protein function^{365,381}.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

GENOMIC FINDINGS

GENE

TP53

ALTERATION

R248W

HGVS VARIANT

NM_000546.4:c.742C>T (p.R248W)

VARIANT CHROMOSOMAL POSITION

chr17:7577539

VARIANT ALLELE FREQUENCY (% VAF)

13.1%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Clinical and preclinical data suggest that solid tumors with TP53 mutations, such as R175H, Y220C, G245S, and R248W, may benefit from adoptive cell therapy targeting these specific TP53 mutations³⁸²⁻³⁸³. Clinical benefit has been reported for patients with breast cancer (2 PRs)³⁸³, ovarian cancer (1 PR)³⁸³, and colorectal cancer (CRC; 1 SD)³⁸² treated with tumor infiltrating lymphocyte-based or modified T-cell receptor-based adoptive cell therapy. There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor adavosertib³⁸⁴⁻³⁸⁷ or p53 gene therapy such as SGT53³⁸⁸⁻³⁹³. In a Phase 1 study, adavosertib in combination with gemcitabine, cisplatin, or carboplatin elicited PRs in 9.7% and SDs in 53% of patients with solid tumors; the response rate was 21% (4/19) for patients with TP53 mutations versus 12% (4/33) for patients who were TP53 wildtype³⁹⁴. Phase 2 studies of adavosertib in combination with chemotherapy reported ORRs of 32% (30/94) and 41% (12/29) for patients with platinum-refractory TP53-mutated ovarian, Fallopian tube, or peritoneal cancer³⁹⁵⁻³⁹⁶. For patients with platinum-sensitive TP53-mutated ovarian cancer, the combination of adavosertib with paclitaxel and carboplatin significantly increased PFS compared with paclitaxel and carboplatin alone (9.9 vs. 8.0 months)³⁹⁷. In the Phase 2 VIKTORY trial, patients with TP53-mutated metastatic and/or recurrent gastric cancer experienced a 24% (6/25) ORR with adavosertib combined with paclitaxel³⁹⁸. A Phase 1 trial of neoadjuvant adavosertib in combination with cisplatin and docetaxel for head and neck squamous cell carcinoma (HNSCC) elicited a 71% (5/7) response rate for patients with TP53 alterations³⁹⁹. The Phase 2 FOCUS4-C trial for

patients with TP53- and RAS-mutated colorectal cancer reported improvement in PFS (3.61 vs. 1.87 months, HR=0.35, p=0.0022), but not OS (14.0 vs 12.8 months, p=0.93), following adavosertib treatment compared with active monitoring⁴⁰⁰. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel for patients with solid tumors, 75% (9/12) of evaluable patients experienced clinical benefit, including 2 confirmed and 1 unconfirmed PRs and 2 instances of SD with significant tumor shrinkage³⁹³. Missense mutations leading to TP53 inactivation may be sensitive to therapies that reactivate mutated p53 such as eprenetapopt. In a Phase 1b trial for patients with p53-positive high-grade serous ovarian cancer, eprenetapopt combined with carboplatin and pegylated liposomal doxorubicin achieved a 52% (11/21) response rate and 100% DCR⁴⁰¹. A Phase 1 trial of eprenetapopt with pembrolizumab for patients with solid tumors reported an ORR of 10% (3/29)⁴⁰².

— Nontargeted Approaches —

Adjuvant chemo(radio)therapy improved outcomes over radiotherapy alone for patients with endometrial carcinoma (EC) molecularly classified as TP53 abnormal (p53abn; as defined by TP53 mutation or either p53 IHC nuclear or cytoplasmic overexpression or null expression) in retrospective analyses^{23,403}, and this approach is recommended by the ESGO-ESTRO-ESP guidelines for this EC subtype¹⁸. For patients with p53abn high-risk EC treated in the PORTEC-3 trial, adjuvant platinum-based chemotherapy plus external beam radiotherapy (EBRT) significantly improved 5-year relapse-free survival (RFS; 59% vs. 36%, HR=0.52) and 5-year OS rates (65% vs. 42%, HR=0.55) versus EBRT alone²³. In another retrospective study, patients with concurrent TP53 mutation and p53 overexpression experienced significantly improved RFS (median 12.8 vs. 7.4 months, HR=0.41) and OS (median 30.0 vs. 14.4 months, HR=0.28) from bevacizumab plus chemotherapy over temsirolimus plus chemotherapy²⁰.

FREQUENCY & PROGNOSIS

TP53 mutations are frequent in endometrial carcinomas, having been reported in 17-54% of cases across multiple studies^{7,158,404}. Among endometrial carcinoma subtypes, TP53 alteration is most common in endometrial serous carcinomas (88-94% of cases)^{7,158,404-405}. TP53 mutations in microsatellite stable/mismatch repair-proficient (MMRp) POLE-wildtype endometrial carcinoma

are indicative of a ProMisE/TCGA molecularly classified subtype, "p53abn" (NCCN Uterine Neoplasms Guidelines, v1.2023) (^{7,15-21}), which is associated with high risk of recurrence²², yet also with the highest degree of benefit from adjuvant chemoradiotherapy over radiotherapy relative to the other subtypes²³. Additionally, classification as this molecular subtype refines FIGO disease stage to IIC (Stage "IICm-p53abn") for patients with histologically determined Stage I/II early endometrial cancer that is locally confined to the uterine corpus or with myometrial invasion²⁴. In the context of endometrial carcinomas that are microsatellite stable (MSS)/mismatch repair-proficient (MMRp) and lack pathogenic POLE mutations, TP53 inactivating mutation or p53 nuclear expression is associated with poor prognosis (NCCN Uterine Neoplasms Guidelines, v1.2023)(^{7,15-22,406-407}).

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers¹⁹⁵. Alterations such as seen here may disrupt TP53 function or expression⁴⁰⁸⁻⁴¹².

POTENTIAL GERMLINE IMPLICATIONS

One or more of the TP53 variants observed here has been described in the ClinVar database as a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with Li-Fraumeni syndrome (ClinVar, Sep 2023)⁴¹³. Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. Germline mutations in TP53 are associated with the very rare autosomal dominant disorder Li-Fraumeni syndrome and the early onset of many cancers⁴¹⁴⁻⁴¹⁶, including sarcomas⁴¹⁷⁻⁴¹⁸. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000⁴¹⁹ to 1:20,000⁴¹⁸. For pathogenic TP53 mutations identified during tumor sequencing, the rate of germline mutations was 1% in the overall population and 6% in tumors arising before age 30⁴²⁰. In the appropriate clinical context, germline testing of TP53 is recommended.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # **ORD-1794439-01**
GENOMIC FINDINGS

expansion⁴²¹⁻⁴²⁶. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy⁴²¹⁻⁴²². Clinical management of patients with CH in this

gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease⁴²⁷. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH^{425,428-429}.

Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

THERAPIES WITH CLINICAL BENEFIT
IN OTHER TUMOR TYPE

Imatinib

Assay findings association
KIT
amplification - equivocal

PDGFRA
amplification

AREAS OF THERAPEUTIC USE

Imatinib targets the BCR-ABL fusion protein, PDGFR, and KIT. It is FDA approved for the treatment of KIT-positive gastrointestinal stromal tumors (GIST), Ph+ chronic myeloid leukemia (CML) and acute lymphoblastic leukemia (ALL), myelodysplastic syndrome/myeloproliferative syndrome (MDS/MPS), aggressive systemic mastocytosis without a D816V KIT mutation, hypereosinophilic syndrome and/or chronic eosinophilic leukemia, and dermatofibrosarcoma protuberans. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical and preclinical data in KIT-mutated^{91-92,430-431}, KIT-amplified⁹⁰⁻⁹³, or KIT-expressing tumors^{95-100,432-433}, KIT activating alterations may confer sensitivity to imatinib. PDGFRA amplification may predict sensitivity to TKIs such as imatinib; a patient with Merkel cell carcinoma expressing PDGFRA achieved a CR to imatinib¹¹⁵.

SUPPORTING DATA

A Phase 2 trial of imatinib for the treatment of uterine

carcinosarcoma reported that the therapy was generally well tolerated but showed minimal activity in the 23 evaluable patients; positive PDGFR-beta or KIT expression was not correlated with patient characteristics or outcome⁴³⁴. A Phase 2 trial of imatinib plus docetaxel for patients with head and neck squamous cell carcinoma (HNSCC) or non-small cell lung cancer (NSCLC) was stopped due to toxicity and lack of efficacy⁴³⁵. A Phase 2 study of pulse dose imatinib in combination with paclitaxel for elderly patients with NSCLC reported a response rate of 32% (11/34); however, PFS and OS was similar to that commonly seen for elderly patients with NSCLC treated with single-agent chemotherapy, and the combination was not recommended for further evaluation⁴³⁶. Phase 2 clinical trials of imatinib for unselected patients with breast cancer did not demonstrate significant efficacy⁴³⁷⁻⁴³⁸. In addition, a Phase 2 study for 13 patients with metastatic breast cancer with no c-KIT expression and overexpression of PDGFR-beta reported that imatinib as a single agent displayed no clinical activity⁴³⁹.

Nilotinib

Assay findings association
KIT
amplification - equivocal

AREAS OF THERAPEUTIC USE

Nilotinib targets tyrosine kinases such as ABL (including BCR-ABL), PDGFRs, KIT, CSF1R, DDR1, and DDR2. It is FDA approved to treat newly diagnosed pediatric or adult patients with Philadelphia chromosome-positive (Ph+) chronic myeloid leukemia (CML) in chronic phase, adults with Ph+ CML in chronic or accelerated phase with resistance or intolerance to prior therapy including imatinib, and pediatric patients with Ph+ CML in chronic phase with resistance or intolerance to prior tyrosine-kinase inhibitor therapy. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical and preclinical data in KIT-mutated^{101,440-443}, KIT-amplified¹⁰¹, or KIT-expressing tumors⁴⁴⁴, KIT activating alterations may confer sensitivity to nilotinib.

SUPPORTING DATA

Clinical data on the efficacy of nilotinib for the treatment

of endometrial cancer are limited (PubMed, Dec 2023). Nilotinib has been primarily investigated as a therapeutic option for the treatment of chronic myeloid leukemia (CML) or gastrointestinal stromal tumors (GIST). In the context of CML, a Phase 3 clinical trial for patients who are Philadelphia chromosome (Ph) positive treated with imatinib or nilotinib (300 or 400 mg) reported PFS rates of 93% and 97-98% and OS rates of 93% and 94-97%, respectively, at 4 years⁴⁴⁵. For Japanese patients with CML who are resistant to imatinib, a Phase 2 trial reported a 49% major medical response rate to treatment with nilotinib at 12 months⁴⁴⁶. A Phase 3 clinical trial of single-agent nilotinib in 240 patients with advanced GIST who failed prior treatment with imatinib or sunitinib reported no significant difference in PFS between nilotinib and the best supportive care but did report increased OS for patients treated with nilotinib⁴⁴⁷. A Phase 2 trial has shown that nilotinib was well tolerated and suggested it may be particularly useful for treating patients with GIST harboring mutations in KIT exon 17⁴⁴⁸.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Sorafenib

Assay findings association

KIT
amplification - equivocal

AREAS OF THERAPEUTIC USE

Sorafenib is a kinase inhibitor that targets the RAF kinases, KIT, FLT3, RET, VEGFRs, and PDGFRs. It is FDA approved for the treatment of unresectable hepatocellular carcinoma, advanced renal cell carcinoma, and recurrent or metastatic differentiated thyroid carcinoma. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical and preclinical data in KIT-mutated⁴⁴⁹⁻⁴⁵⁶ or KIT-expressing tumors¹⁰²⁻¹⁰⁵, KIT activating alterations may predict sensitivity to sorafenib.

SUPPORTING DATA

In a Phase 2 study of sorafenib for patients with advanced uterine carcinoma or carcinosarcoma, 5% (2/40) of patients with uterine carcinoma had a PR and 43% (17/40) had SD; no patients (0/16) with uterine carcinosarcoma had a PR and 25% (4/16) had SD⁴⁵⁷. Sorafenib was found to demonstrate modest clinical activity for patients with recurrent ovarian cancer or primary peritoneal carcinomatosis in a Phase 2 clinical trial; however, considerable toxicity was observed⁴⁵⁸. Another Phase 2 study of sorafenib as third-line treatment for patients with epithelial ovarian cancer or primary peritoneal cancer showed no benefit, with none of the 11 patients exhibiting a PR, CR, or sustained SD⁴⁵⁹.

Sunitinib

Assay findings association

KIT
amplification - equivocal

AREAS OF THERAPEUTIC USE

Sunitinib is a small-molecule tyrosine kinase inhibitor that targets PDGFRs, VEGFRs, KIT, FLT3, CSF-1R, and RET. It is FDA approved for the treatment of advanced or metastatic pancreatic neuroendocrine tumors, gastrointestinal stromal tumors (GISTs) in patients who have progressed on or are intolerant to imatinib, and advanced renal cell carcinoma (RCC) as well as for the adjuvant treatment of patients at high risk of recurrent RCC after nephrectomy. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical and preclinical data in KIT-mutated^{106,460-464} or KIT-expressing tumors¹⁰⁶⁻¹⁰⁷, KIT activating alterations may predict sensitivity to sunitinib.

SUPPORTING DATA

In a Phase 2 study, sunitinib elicited 18% (6/33) PR and 18% (6/33) SD rates in patients with endometrial carcinoma or carcinosarcoma, with median PFS and OS of 3 months and 19.4 months, respectively⁴⁶⁵.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually

updated and should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that

may require medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov. Or, visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

GENE

CCNE1

ALTERATION
amplification

RATIONALE

Strong preclinical and clinical data suggest that CCNE1 amplification may predict sensitivity to WEE1 inhibitors. Strong preclinical data suggest

that CCNE1 amplification may predict sensitivity to PKMYT1 inhibitors.

NCT04768868

PHASE 1

The Safety and Pharmacokinetics Preliminary Efficacy of IMP7068 in Patients With Advanced Solid Tumors

TARGETS
WEE1

LOCATIONS: Taipei (Taiwan), Taoyuan (Taiwan), Taichung (Taiwan), Tainan (Taiwan), Shanghai (China), Wuhan (China), Beijing (China), Chengdu (China), Kansas, Texas

NCT05605509

PHASE 2

RP-6306 in Patients With Advanced Cancer

TARGETS
PKMYT1, ERBB2

LOCATIONS: Vancouver (Canada), Ottawa (Canada), Toronto (Canada), Kingston (Canada), London (Canada)

NCT04814108

PHASE 2

A Study of ZN-c3 in Women With Recurrent or Persistent Uterine Serous Carcinoma

TARGETS
WEE1

LOCATIONS: Oregon, Nevada, California, Arizona, Missouri, Ohio, Texas, New York

NCT03968653

PHASE 1

Study of Oral Debio 0123 in Combination With Carboplatin in Participants With Advanced Solid Tumors

TARGETS
WEE1

LOCATIONS: Groningen (Netherlands), Nijmegen (Netherlands), Leiden (Netherlands), Barcelona (Spain)

NCT05109975

PHASE 1

A Study to Evaluate Safety and Preliminary Anti-tumor Activity of Debio 0123 as Monotherapy in Adult Participants With Advanced Solid Tumors

TARGETS
WEE1

LOCATIONS: Bellinzona (Switzerland), Zürich (Switzerland), Michigan, Texas

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

CLINICAL TRIALS

NCT04855656

PHASE 1

Study of RP-6306 Alone or in Combination With RP-3500 in Patients With Advanced Solid Tumors

TARGETS
PKMYT1, ATR

LOCATIONS: Copenhagen (Denmark), Utah, Toronto (Canada), Missouri, Massachusetts, Rhode Island, Connecticut, New York, Pennsylvania, Texas

NCT05147272

PHASE 1

Study of RP-6306 With Gemcitabine in Advanced Solid Tumors

TARGETS
PKMYT1

LOCATIONS: London (United Kingdom), California, Arizona, Minnesota, Michigan, Toronto (Canada), New York, Pennsylvania, Florida

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

CLINICAL TRIALS
GENE
FBXW7
ALTERATION
R479Q
RATIONALE

Loss or inactivation of FBXW7 may lead to increased mTOR activation and may predict sensitivity to mTOR inhibitors.

NCT03239015
PHASE 2

Efficacy and Safety of Targeted Precision Therapy in Refractory Tumor With Druggable Molecular Event

TARGETS

EGFR, ERBB4, ERBB2, PARP, mTOR, MET, ROS1, RET, VEGFRs, BRAF, CDK4, CDK6

LOCATIONS: Shanghai (China)

NCT04803318
PHASE 2

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

TARGETS

mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK

LOCATIONS: Guangzhou (China)

NCT05125523
PHASE 1

A Study of Sirolimus for Injection (Albumin Bound) in Patients With Advanced Solid Tumors

TARGETS

mTOR

LOCATIONS: Tianjin (China)

NCT03297606
PHASE 2

Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)

TARGETS

ALK, ROS1, AXL, TRKA, MET, TRKC, EGFR, PARP, CDK4, CDK6, mTOR, MEK, BRAF, SMO

LOCATIONS: Vancouver (Canada), Edmonton (Canada), Saskatoon (Canada), Regina (Canada), Ottawa (Canada), Montreal (Canada), Toronto (Canada), Kingston (Canada), London (Canada)

NCT03203525
PHASE 1

Combination Chemotherapy and Bevacizumab With the NovoTTF-100L(P) System in Treating Participants With Advanced, Recurrent, or Refractory Hepatic Metastatic Cancer

TARGETS

VEGFA, mTOR

LOCATIONS: Texas

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

CLINICAL TRIALS

NCT05036226

PHASE 1/2

COAST Therapy in Advanced Solid Tumors and Prostate Cancer

TARGETS
DDR2, ABL, SRC, KIT, mTOR

LOCATIONS: South Carolina

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

CLINICAL TRIALS
GENE
KIT
ALTERATION
amplification - equivocal

RATIONALE
KIT amplification or activating mutations may predict sensitivity to small molecule tyrosine kinase inhibitors. Also, because KIT activation

leads to activation of the PI3K-AKT-mTOR pathway, PI3K and mTOR pathway inhibitors may be relevant in a tumor with KIT activation.

NCT05007106
PHASE 2

MK-7684A With or Without Other Anticancer Therapies in Participants With Selected Solid Tumors (MK-7684A-005)

TARGETS
PD-1, KIT, VEGFRs, FGFRs, PDGFRA, RET, TIGIT

LOCATIONS: Taoyuan (Taiwan), Tainan (Taiwan), Taipei (Taiwan), Seoul (Korea, Republic of), Osaka (Japan), Nagoya (Japan), Tokyo (Japan), Kashiwa (Japan), Alaska, Adana (Turkey)

NCT04008797
PHASE 1

A Study of E7386 in Combination With Other Anticancer Drug in Participants With Solid Tumor

TARGETS
CBP, Beta-catenin, FGFRs, RET, PDGFRA, VEGFRs, KIT

LOCATIONS: Taipei (Taiwan), Taoyuan (Taiwan), Taichung (Taiwan), Tainan (Taiwan), Kurume (Japan), Matsuyama (Japan), Seongnam-si Bundang (Korea, Republic of), Songpa-gu (Korea, Republic of), Seoul (Korea, Republic of), Seodaemun (Korea, Republic of)

NCT05112991
PHASE 2

Study of Envafolelimab Alone or With Lenvatinib in Patients With Advanced Endometrial Cancer

TARGETS
PD-L1, FGFRs, RET, PDGFRA, VEGFRs, KIT

LOCATIONS: Fuzhou (China), Hangzhou (China), Ganzhou (China), Shanghai (China), Nanjing (China), Guangzhou (China), Changsha (China), Wuhan (China), Yueyang (China), Linyi (China)

NCT05024214
PHASE 1/2

Phase Ib/II Trial of Envafolelimab Plus Lenvatinib for Subjects With Solid Tumors

TARGETS
PD-L1, FGFRs, RET, PDGFRA, VEGFRs, KIT, FLT3, CSF1R

LOCATIONS: Hangzhou (China), Shanghai (China), Dongguan (China), Guangzhou (China), Zhuhai (China), Benbu (China), Zhengzhou (China), Jinan (China), Dalian (China), Tianjin (China)

NCT05098847
PHASE 2

Cryoablation Combined With Sintilimab Plus Lenvatinib In Previously Treated Unresectable Liver Metastasis From Solid Tumors

TARGETS
FGFRs, RET, PDGFRA, VEGFRs, KIT, PD-1

LOCATIONS: Shanghai (China)

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

CLINICAL TRIALS
NCT04803318
PHASE 2

Trametinib Combined With Everolimus and Lenvatinib for Recurrent/Refractory Advanced Solid Tumors

TARGETS
mTOR, FGFRs, RET, PDGFRA, VEGFRs, KIT, MEK

LOCATIONS: Guangzhou (China)

NCT04977453
PHASE 1/2

GI-101 as a Single Agent or in Combination With Pembrolizumab, Lenvatinib or Local Radiotherapy in Advanced Solid Tumors

TARGETS
FGFRs, RET, PDGFRA, VEGFRs, KIT, PD-1, CTLA-4

LOCATIONS: Daejeon (Korea, Republic of), Suwon-si (Korea, Republic of), Seoul (Korea, Republic of), New York, North Carolina

NCT05740215
PHASE 1/2

Efficacy and Safety Study of F520 Combined With Lenvatinib in the Treatment of Patients With Advanced Solid Tumors

TARGETS
PD-1, FGFRs, RET, PDGFRA, VEGFRs, KIT

LOCATIONS: Chongqing (China)

NCT05554341
PHASE 2

Testing the Use of Nilotinib and Paclitaxel as a Treatment for Patients With Prior Taxane Treatment, A ComboMATCH Treatment Trial

TARGETS
ABL, KIT

LOCATIONS: Washington, Oregon, Idaho, Montana

NCT02693535
PHASE 2

TAPUR: Testing the Use of Food and Drug Administration (FDA) Approved Drugs That Target a Specific Abnormality in a Tumor Gene in People With Advanced Stage Cancer

TARGETS
CDK4, CDK6, FLT3, VEGFRs, CSF1R, KIT, RET, mTOR, ERBB2, MEK, BRAF, PARP, PD-1, CTLA-4, PD-L1, TRKB, ALK, TRKC, ROS1, TRKA, FGFRs

LOCATIONS: Washington, Oregon, California

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

CLINICAL TRIALS
GENE
MDM2
ALTERATION
amplification

RATIONALE
Inhibitors of the MDM2-p53 interaction are being tested in clinical trials. Overexpression or amplification of MDM2 may increase sensitivity to these agents, but more data are required.

NCT04785196
PHASE 1/2

APG-115 in Combination With PD-1 Inhibitor in Patients With Advanced Liposarcoma or Advanced Solid Tumors

TARGETS
PD-1, MDM2

LOCATIONS: Shanghai (China), Guangzhou (China)

NCT03611868
PHASE 1/2

A Study of APG-115 in Combination With Pembrolizumab in Patients With Metastatic Melanomas or Advanced Solid Tumors

TARGETS
MDM2, PD-1

LOCATIONS: Brisbane (Australia), South Brisbane (Australia), Bedford Park (Australia), Heidelberg (Australia), California, Arizona, Missouri, Arkansas, Ohio, Pennsylvania

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

CLINICAL TRIALS
GENE
MYC
ALTERATION
amplification

RATIONALE

MYC overexpression may predict sensitivity to inhibition of CDKs, especially CDK1 and CDK2, of Aurora kinases, including Aurora kinase A and B,

and of BET domain proteins, which are reported to downregulate MYC expression and MYC-dependent transcriptional programs.

NCT05253053
PHASE 1/2

Study to Evaluate the Efficacy and Safety of TT-00420 as Monotherapy and Combination Therapy in Patients With Advanced Solid Tumors

TARGETS

FGFR2, FGFR1, FGFR3, PD-L1

LOCATIONS: Shanghai (China), Wuhu (China), Nanjing (China), Changsha (China), Zhengzhou (China), Jinan (China), Beijing (China), Changchun (China)

NCT05110807
PHASE 1

A Clinical Study to Evaluate the Tolerability and Pharmacokinetics of TQB3617 Capsule in Patients With Advanced Malignant Tumors

TARGETS

BRD3, BRD4, BRD2, BRDT

LOCATIONS: Hangzhou (China), Guangzhou (China)

NCT04983810
PHASE 1/2

A Study to Investigate Fadraciclub (CYC065), in Subjects With Advanced Solid Tumors and Lymphoma

TARGETS

CDK2, CDK9

LOCATIONS: Seoul (Korea, Republic of), Barcelona (Spain), California, Texas

NCT04686682
PHASE 1/2

A First-in-Human, JAB-8263 in Adult Patients With Advanced Tumors

TARGETS

BRD2, BRD3, BRD4, BRDT

LOCATIONS: Tianjin (China)

NCT05252390
PHASE 1/2

NUV-868 as Monotherapy and in Combination With Olaparib or Enzalutamide in Adult Patients With Advanced Solid Tumors

TARGETS

BRD4, PARP, AR

LOCATIONS: Nedlands (Australia), Waratah (Australia), North Ryde (Australia), Melbourne (Australia), Malvern (Australia), Montana, California, Colorado

NCT05327010
PHASE 2

Testing the Combination of the Anti-cancer Drugs ZEN003694 (ZEN-3694) and Talazoparib in Patients With Advanced Solid Tumors, The ComBET Trial

TARGETS

PARP, BRD4, BRDT, BRD2, BRD3

LOCATIONS: California, Colorado, Illinois, Pennsylvania, Kentucky, Virginia, Texas

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

CLINICAL TRIALS
NCT05372640
PHASE 1

Testing the Safety and Efficacy of the Combination of Two Anti-cancer Drugs, ZEN003694 and Abemaciclib, for Adult and Pediatric Patients (12-17 Years) With Metastatic or Unresectable NUT Carcinoma and Other Solid Tumors

TARGETS
CDK4, CDK6, BRD3, BRD4, BRD2, BRDT

LOCATIONS: California, Pennsylvania, Massachusetts, Texas

NCT05053971
PHASE 1/2

Testing A New Anti-cancer Drug Combination, Entinostat and ZEN003694, for Advanced and Refractory Solid Tumors and Lymphomas

TARGETS
BRD3, BRD4, BRD2, BRDT, HDAC

LOCATIONS: Oklahoma, Connecticut, Florida

NCT04840589
PHASE 1

Testing the Combination of ZEN003694 and Nivolumab With or Without Ipilimumab in Solid Tumors

TARGETS
PD-1, CTLA-4, BRD4, BRDT, BRD2, BRD3

LOCATIONS: Ohio, Pennsylvania, New York, Maryland

NCT04587479
PHASE 1

A First-in-Human Study of JAB-8263 in Adult Patients With Advanced Solid Tumors

TARGETS
BRD2, BRD3, BRD4, BRDT

LOCATIONS: Colorado, Tennessee, Florida

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

CLINICAL TRIALS

GENE

PDGFRA

ALTERATION

amplification

RATIONALE

PDGFRA amplification may predict sensitivity to imatinib and to anti-PDGFRA antibodies.

NCT04817956

PHASE 2

Improving Public Cancer Care by Implementing Precision Medicine in Norway

TARGETS

PD-L1, VEGFA, ERBB2, ALK, RET, PARP, SMO, TRKB, TRKC, ROS1, TRKA, MEK, BRAF, PI3K-alpha, FGFR1, FGFR2, FGFR3, MET, KIT, ABL

LOCATIONS: Tromsø (Norway), Bodø (Norway), Hamar (Norway), Oslo (Norway), Fredrikstad (Norway), Drammen (Norway), Trondheim (Norway), Skien (Norway), Førde (Norway), Bergen (Norway)

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

APPENDIX
Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

ASXL1

 NM_015338.5: c.1495C>T
(p.R499C)
chr20:31021496

ATM

 NM_000051.3: c.691C>A
(p.H231N)
chr11:108115543

BCORL1

 NM_021946.4: c.823C>G
(p.L275V)
chrX:129147571

CIC

 NM_015125.4: c.4081G>C
(p.E1361Q)
chr19:42798127 and
NM_015125.4: c.3987G>C
(p.K1329N)
chr19:42797935

DAXX

 NM_001350.4: c.1629A>G
(p.I543M)
chr6:33287468

DIS3

 NM_001128226.1: c.1481C>T
(p.A494V)
chr13:73345967

KDR

rearrangement

KMT2D (MLL2)

 NM_003482.4: c.10024C>T
(p.R3342C)
chr12:49431115

SPEN

 NM_015001.2: c.2227C>G
(p.Q743E)
chr1:16254962 and
NM_015001.2: c.3455C>T
(p.S1152L)
chr1:16256190

TBX3

amplification

TNFAIP3

 NM_006290.2: c.1398C>G
(p.S466R)
chr6:138199980

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

 Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

 Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

APPENDIX

Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

| | | | | | | | | |
|--------------|-----------------|-----------------------|----------------|---------|-----------------|----------------|------------------------|------------------|
| ABL1 | ACVR1B | AKT1 | AKT2 | AKT3 | ALK | ALOX12B | AMER1 (FAM123B or WTX) | |
| APC | AR | ARAF | ARFRP1 | ARID1A | ASXL1 | ATM | ATR | ATRX |
| AURKA | AURKB | AXIN1 | AXL | BAP1 | BARD1 | BCL2 | BCL2L1 | BCL2L2 |
| BCL6 | BCOR | BCORL1 | BRAF | BRCA1 | BRCA2 | BRD4 | BRIP1 | BTG1 |
| BTG2 | BTK | CALR | CARD11 | CASP8 | CBFB | CBL | CCND1 | CCND2 |
| CCND3 | CCNE1 | CD22 | CD274 (PD-L1) | CD70 | CD79A | CD79B | CDC73 | CDH1 |
| CDK12 | CDK4 | CDK6 | CDK8 | CDKN1A | CDKN1B | CDKN2A | CDKN2B | CDKN2C |
| CEBPA | CHEK1 | CHEK2 | CIC | CREBBP | CRKL | CSF1R | CSF3R | CTCF |
| CTNNA1 | CTNNB1 | CUL3 | CUL4A | CXCR4 | CYP17A1 | DAXX | DDR1 | DDR2 |
| DIS3 | DNMT3A | DOT1L | EED | EGFR | EMSY (C11orf30) | EP300 | EPHA3 | EPHB1 |
| EPHB4 | ERBB2 | ERBB3 | ERBB4 | ERCC4 | ERG | ERRF1 | ESR1 | EZH2 |
| FANCA | FANCC | FANCG | FANCL | FAS | FBXW7 | FGF10 | FGF12 | FGF14 |
| FGF19 | FGF23 | FGF3 | FGF4 | FGF6 | FGFR1 | FGFR2 | FGFR3 | FGFR4 |
| FH | FLCN | FLT1 | FLT3 | FOXL2 | FUBP1 | GABRA6 | GATA3 | GATA4 |
| GATA6 | GID4 (C17orf39) | GNA11 | GNA13 | GNAS | GNAS | GRM3 | GSK3B | H3-3A (H3F3A) |
| HDAC1 | HGF | HNFA1 | HRAS | HSD3B1 | ID3 | IDH1 | IDH2 | IGF1R |
| IKBKE | IKZF1 | INPP4B | IRF2 | IRF4 | IRS2 | JAK1 | JAK2 | JAK3 |
| JUN | KDM5A | KDM5C | KDM6A | KDR | KEAP1 | KEL | KIT | KLHL6 |
| KMT2A (MLL) | KMT2D (MLL2) | KRAS | LTK | LYN | MAF | MAP2K1 (MEK1) | MAP2K2 (MEK2) | MAP2K4 |
| MAP3K1 | MAP3K13 | MAPK1 | MCL1 | MDM2 | MDM4 | MED12 | MEF2B | MEN1 |
| MERTK | MET | MITF | MKNK1 | MLH1 | MPL | MRE11 (MRE11A) | MSH2 | MSH3 |
| MSH6 | MST1R | MTAP | MTOR | MUTYH | MYC | MYCL (MYCL1) | MYCN | MYD88 |
| NBN | NF1 | NF2 | NFE2L2 | NFKBIA | NKX2-1 | NOTCH1 | NOTCH2 | NOTCH3 |
| NPM1 | NRAS | NSD2 (WHSC1 or MMSET) | NSD3 (WHSC1L1) | NT5C2 | NTRK1 | NTRK2 | NTRK3 | NTRK3 |
| P2RY8 | PALB2 | PARP1 | PARP2 | PARP3 | PAX5 | PBRM1 | PDCD1 (PD-1) | PDCD1LG2 (PD-L2) |
| PDGFRA | PDGFRB | PDK1 | PIK3C2B | PIK3C2G | PIK3CA | PIK3CB | PIK3R1 | PIM1 |
| PMS2 | POLD1 | POLE | PPARG | PPP2R1A | PPP2R2A | PRDM1 | PRKAR1A | PRKCI |
| PRKN (PARK2) | PTCH1 | PTEN | PTPN11 | PTPRO | QKI | RAC1 | RAD21 | RAD51 |
| RAD51B | RAD51C | RAD51D | RAD52 | RAD54L | RAF1 | RARA | RB1 | RBM10 |
| REL | RET | RICTOR | RNF43 | ROS1 | RPTOR | SDHA | SDHB | SDHC |
| SDHD | SETD2 | SF3B1 | SGK1 | SMAD2 | SMAD4 | SMARCA4 | SMARCB1 | SMO |
| SNCAIP | SOC1 | SOX2 | SOX9 | SPEN | SPOP | SRC | STAG2 | STAT3 |
| STK11 | SUFU | SYK | TBX3 | TEK | TENT5C (FAM46C) | TET2 | TET2 | TGFBP2 |
| TIPARP | TNFAIP3 | TNFRSF14 | TP53 | TSC1 | TSC2 | TYRO3 | U2AF1 | VEGFA |
| VHL | WT1 | XPO1 | XRCC2 | ZNF217 | ZNF703 | | | |

DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

| | | | | | | | | |
|------|------|-------|--------|-------|---------|-------|--------|-------------|
| ALK | BCL2 | BCR | BRAF | BRCA1 | BRCA2 | CD74 | EGFR | ETV4 |
| ETV5 | ETV6 | EWSR1 | EZR | FGFR1 | FGFR2 | FGFR3 | KIT | KMT2A (MLL) |
| MSH2 | MYB | MYC | NOTCH2 | NTRK1 | NTRK2 | NUTM1 | PDGFRA | RAF1 |
| RARA | RET | ROS1 | RSP02 | SDC4 | SLC34A2 | TERC* | TERT** | TPR2SS2 |

*TERC is an NCRNA

**Promoter region of TERT is interrogated

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Homologous Recombination status
Loss of Heterozygosity (LOH) score
Microsatellite (MS) status
Tumor Mutational Burden (TMB)

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.


Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531

ORDERED TEST # ORD-1794439-01

APPENDIX

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Ciplstraat 3, 2440 Geel, Belgium. 

ABOUT FOUNDATIONONE CDx

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:
www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLE

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes

detection of alterations in a total of 324 genes.

Using an Illumina® Sequencing platform (HiSeq 4000 or NovaSeq 6000), hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI), and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for *ERBB2* and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is

one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials

Ranking of Therapies in Summary Table
Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2023. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

1. In the fraction-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

- MSS cut-off thresholds were determined by analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as “MS-Stable” with median exon coverage <300X, “MS-Equivocal,” or “Cannot Be Determined” should receive confirmatory testing using a validated orthogonal (alternative) method.
- TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
 - Homologous Recombination status may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas (Coleman et al., 2017; 28916367). Samples with deleterious *BRCA1/2* alteration and/or Loss of Heterozygosity (LOH) score ≥ 16% will be reported as “HRD Positive” and samples with absence of these findings will be reported as “HRD Not Detected,” agnostic of potential secondary *BRCA1/2* reversion alterations. Certain potentially deleterious missense or small in-frame deletions in *BRCA1/2* may not be classified as deleterious and, in the absence of an elevated LOH profile, samples with such mutations may be classified as “HRD Not Detected.” A result of “HRD Not Detected” does not rule out the presence of a *BRCA1/2* alteration or an elevated LOH profile outside the assay performance characteristic limitations.
 - The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the

genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as “Cannot Be Determined” if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.

- Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
- Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine whether the patient is a candidate for biopsy.
- Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. *HER2* overexpression occurs in 18-20% of breast cancers (Owens et al. 2004 [PMID: 15140287]; Salmon et al. 1987 [PMID: 3798106]; Yaziji et al. 2004 [PMID: 15113815]). Based on the F1CDx *HER2* CDx concordance study, approximately 10% of *HER2* amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic,

nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

| BASE SUBSTITUTIONS | %CV* |
|--------------------|--------------|
| Repeatability | 5.11 - 10.40 |
| Reproducibility | 5.95 - 12.31 |
| INDELS | %CV* |
| Repeatability | 6.29 - 10.00 |
| Reproducibility | 7.33 - 11.71 |

*Interquartile Range = 1st Quartile to 3rd Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an “official / formal solution” and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

ORDERED TEST # ORD-1794439-01

APPENDIX
About FoundationOne®CDx

distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D (MLL2)*, *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent

medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

| ABBREVIATION | DEFINITION |
|--------------|-----------------------------|
| CR | Complete response |
| DCR | Disease control rate |
| DNMT | DNA methyltransferase |
| HR | Hazard ratio |
| ITD | Internal tandem duplication |
| MMR | Mismatch repair |
| mut/Mb | Mutations per megabase |
| NOS | Not otherwise specified |
| ORR | Objective response rate |
| OS | Overall survival |
| PD | Progressive disease |
| PFS | Progression-free survival |
| PR | Partial response |
| SD | Stable disease |
| TKI | Tyrosine kinase inhibitor |

REFERENCE SEQUENCE INFORMATION

Sequence data is mapped to the human genome, Genome Reference Consortium Human Build 37 (GRCh37), also known as hg19.

SOFTWARE VERSION INFORMATION

MR Suite Version (RG) 7.15.0
MR Reporting Config Version Config 49
Analysis Pipeline Version v3.29.0
Computational Biology Suite Version 6.29.0

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of the data. The median exon coverage for this sample is 905x. The suitability of use.

ORDERED TEST # ORD-1794439-01

APPENDIX
References

1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) PMID: 25392179
2. Kroemer G, et al. Oncoimmunology (2015) PMID: 26140250
3. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
4. Overman et al., 2016; ASCO Abstract 3501
5. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
6. Ayers et al., 2016; ASCO-SITC Abstract P60
7. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
8. Black D, et al. J. Clin. Oncol. (2006) PMID: 16549821
9. Mackay HJ, et al. Eur. J. Cancer (2010) PMID: 20304627
10. Kanopienė D, et al. Medicina (Kaunas) (2014) PMID: 25458958
11. Hampel H, et al. Cancer Res. (2006) PMID: 16885385
12. Steinbakk A, et al. Cell Oncol (Dordr) (2011) PMID: 21547578
13. Bilbao C, et al. Int. J. Radiat. Oncol. Biol. Phys. (2010) PMID: 20005452
14. Church DN, et al. Hum. Mol. Genet. (2013) PMID: 23528559
15. McCluggage WG, et al. Histopathology (2022) PMID: 34996131
16. Oaknin A, et al. Ann Oncol (2022) PMID: 35690222
17. Talhouk A, et al. Br J Cancer (2015) PMID: 26172027
18. Concin N, et al. Virchows Arch (2021) PMID: 33604759
19. Wu Q, et al. BMC Cancer (2022) PMID: 36357827
20. Thiel KW, et al. J Clin Oncol (2022) PMID: 35658479
21. Irshaid L, et al. Mod Pathol (2023) PMID: 36857998
22. Leon-Castillo A, et al. Gynecol Oncol (2022) PMID: 35078648
23. León-Castillo A, et al. J Clin Oncol (2020) PMID: 32749941
24. Berek JS, et al. Int J Gynaecol Obstet (2023) PMID: 37337978
25. Zigelboim I, et al. J. Clin. Oncol. (2007) PMID: 17513808
26. Bilbao-Sieyro C, et al. Oncotarget (2014) PMID: 25026289
27. Arabi H, et al. Gynecol. Oncol. (2009) PMID: 19275958
28. Stelloo E, et al. Clin. Cancer Res. (2016) PMID: 27006490
29. Nout RA, et al. Gynecol. Oncol. (2012) PMID: 22609107
30. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
31. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
32. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
33. Boland CR, et al. Cancer Res. (1998) PMID: 9823339
34. Pawlik TM, et al. Dis. Markers (2004) PMID: 15528785
35. Boland CR, et al. Gastroenterology (2010) PMID: 20420947
36. Legrand et al., 2018; ASCO Abstract 12000
37. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
38. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
39. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
40. Marabelle et al., 2019; ESMO Abstract 11920
41. Cristescu R, et al. Science (2018) PMID: 30309915
42. Rizvi et al., 2017; WCLC Abstract 1106
43. Hodi et al., 2019; AACR abstract CT037
44. Lee et al., 2019; ASCO Abstract 641
45. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
46. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
47. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
48. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
49. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
50. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
51. Marabelle A, et al. Lancet Oncol. (2020) PMID: 32919526
52. Ott PA, et al. J. Clin. Oncol. (2019) PMID: 30557521
53. Cristescu R, et al. J Immunother Cancer (2022) PMID: 35101941
54. Friedmann CF, et al. Cancer Discov (2022) PMID: 34876409
55. Sturgill EG, et al. Oncologist (2022) PMID: 35274716
56. Schenker at al., 2022; AACR Abstract 7845
57. Chalmers ZR, et al. Genome Med (2017) PMID: 28420421
58. Santin et al., 2016; ASCO Abstract 5591
59. Mehnert JM, et al. J. Clin. Invest. (2016) PMID: 27159395
60. Hussein YR, et al. Mod. Pathol. (2015) PMID: 25394778
61. Shao C, et al. JAMA Netw Open (2020) PMID: 33119110
62. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
63. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
64. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
65. Rizvi NA, et al. Science (2015) PMID: 25765070
66. Johnson BE, et al. Science (2014) PMID: 24336570
67. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
68. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
69. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
70. Nature (2012) PMID: 22810696
71. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
72. Heinrich et al., 2014; ASCO Abstract 10506
73. Heinrich et al., 2017; ASCO Abstract 11011
74. DeAngelo et al., 2017; ASH Abstract 2
75. George et al., 2018; ASCO Abstract 11511
76. Kim et al., 2022; ENA Abstract 237
77. Evans EK, et al. Sci Transl Med (2017) PMID: 29093181
78. Abbaspour Babaei M, et al. Drug Des Devel Ther (2016) PMID: 27536065
79. Ramaswamy A, et al. J Gastrointest Oncol (2016) PMID: 27634456
80. Demetri GD, et al. Lancet (2013) PMID: 23177515
81. Gotlib J, et al. N. Engl. J. Med. (2016) PMID: 27355533
82. Jawhar M, et al. Blood (2017) PMID: 28424161
83. Xu X, et al. Int J Clin Exp Pathol (2014) PMID: 25031773
84. Gotlib J, et al. Blood (2005) PMID: 15972446
85. Luo C, et al. Onco Targets Ther (2017) PMID: 29066909
86. Janku F, et al. ESMO Open (2022) PMID: 35753087
87. Si et al., 2012; ASCO Abstract 8562
88. Si L, et al. J. Clin. Oncol. (2012) PMID: 22162580
89. Parikh SA, et al. Leuk Lymphoma (2010) PMID: 20038218
90. Wei X, et al. Oncol. Res. (2019) PMID: 30075827
91. Hodi FS, et al. J. Clin. Oncol. (2013) PMID: 23775962
92. Carvajal RD, et al. JAMA (2011) PMID: 21642685
93. Guo J, et al. J. Clin. Oncol. (2011) PMID: 21690468
94. Debiec-Rychter M, et al. Gastroenterology (2005) PMID: 15685537
95. Dematteo RP, et al. Lancet (2009) PMID: 19303137
96. Faivre S, et al. J. Clin. Oncol. (2005) PMID: 16135502
97. Hotte SJ, et al. J. Clin. Oncol. (2005) PMID: 15659505
98. Alcedo JC, et al. Head Neck (2004) PMID: 15350030
99. Brandwein JM, et al. Leukemia (2011) PMID: 21403650
100. Reardon DA, et al. Br. J. Cancer (2009) PMID: 19904263
101. Lee SJ, et al. Oncologist (2015) PMID: 26424760
102. Llovet JM, et al. Clin. Cancer Res. (2012) PMID: 22374331
103. Zhang HL, et al. Clin Genitourin Cancer (2013) PMID: 23058498
104. Seino S, et al. Gastroenterology (2014) PMID: 25450081
105. Li XF, et al. Med. Oncol. (2009) PMID: 18846437
106. Minor DR, et al. Clin. Cancer Res. (2012) PMID: 22261812
107. Mahipal A, et al. Melanoma Res. (2012) PMID: 23114504
108. Nguyen B, et al. Cell (2022) PMID: 35120664
109. Scobie JV, et al. Int. J. Gynecol. Pathol. (2003) PMID: 12649669
110. Int. J. Biochem. Cell Biol. (1999) PMID: 10582339
111. Semin. Oncol. (2004) PMID: 15175998
112. Gao J, et al. Sci Signal (2013) PMID: 23550210
113. Zack TI, et al. Nat. Genet. (2013) PMID: 24071852
114. Beroukhi R, et al. Nature (2010) PMID: 20164920
115. Frenard C, et al. JAAD Case Rep (2016) PMID: 27051816
116. Goebler B, et al. Eur J Cancer (2009) PMID: 19362466
117. Slomovitz BM, et al. Gynecol. Oncol. (2004) PMID: 15385107
118. Andrae J, et al. Genes Dev. (2008) PMID: 18483217
119. Burford A, et al. PLoS ONE (2013) PMID: 23990986
120. Flavahan WA, et al. Nature (2016) PMID: 26700815
121. Roszik J, et al. Sci Rep (2016) PMID: 26787600
122. Verhaak RG, et al. Cancer Cell (2010) PMID: 20129251
123. Koschmann C, et al. Oncotarget (2016) PMID: 27582545
124. Phillips JJ, et al. Brain Pathol. (2013) PMID: 23438035
125. Puget S, et al. PLoS ONE (2012) PMID: 22389665
126. Lin AB, et al. Clin. Cancer Res. (2017) PMID: 28331049
127. Chen X, et al. Clin Cancer Res (2018) PMID: 30181387
128. Mörry T, et al. Int. J. Biochem. Cell Biol. (2004) PMID: 15147722
129. Lee JM, et al. Lancet Oncol. (2018) PMID: 29361470
130. Lheureux S, et al. Lancet (2021) PMID: 33485453
131. Oza AM, et al. Clin Cancer Res (2020) PMID: 32611648
132. Fu S, et al. J Clin Oncol (2023) PMID: 36469840
133. Fu et al., 2021; AACR abstract 974
134. Ma T, et al. Mol. Cancer Ther. (2013) PMID: 23686769
135. Toledo LI, et al. Nat. Struct. Mol. Biol. (2011) PMID: 21552262
136. Buisson R, et al. Mol. Cell (2015) PMID: 26365377
137. Yang L, et al. Oncotarget (2015) PMID: 26204491
138. Gallo D, et al. Nature (2022) PMID: 35444283
139. Szychowski J, et al. J Med Chem (2022) PMID: 35880755
140. Kok YP, et al. Oncogenesis (2020) PMID: 33028815
141. Taylor-Harding B, et al. Oncotarget (2015) PMID: 25557169
142. Etemadmoghadam D, et al. Clin. Cancer Res. (2013) PMID: 24004674
143. Scaltriti M, et al. Proc. Natl. Acad. Sci. U.S.A. (2011) PMID: 21321214
144. Nanos-Webb A, et al. Breast Cancer Res. Treat. (2012) PMID: 21695458
145. Kuhn E, et al. Mod. Pathol. (2014) PMID: 24309323
146. Mayr D, et al. Am. J. Clin. Pathol. (2006) PMID: 16753589
147. Keyomarsi K, et al. N. Engl. J. Med. (2002) PMID: 12432043
148. Potemski P, et al. Med. Sci. Monit. (2009) PMID: 19179977
149. Callagy G, et al. J. Pathol. (2005) PMID: 15682439
150. Nakayama N, et al. Cancer (2010) PMID: 20336784
151. Leung SY, et al. Mod. Pathol. (2006) PMID: 16575401
152. Lin L, et al. Cancer Res. (2000) PMID: 11156406
153. Stamatakis M, et al. World J Surg Oncol (2010) PMID: 21176227

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

ORDERED TEST # **ORD-1794439-01**
APPENDIX
References

154. Mao JH, et al. Science (2008) PMID: 18787170
155. Yang H, et al. Oncotarget (2015) PMID: 25749036
156. Kulkarni et al., 2020; SGO Abstract 356
157. Zehir A, et al. Nat. Med. (2017) PMID: 28481359
158. Soumerai TE, et al. Clin. Cancer Res. (2018) PMID: 30068706
159. Garcia-Dios DA, et al. Gynecol. Oncol. (2013) PMID: 23219661
160. Tu K, et al. Hepatol. Res. (2012) PMID: 22548670
161. Iwatsuki M, et al. Int. J. Cancer (2010) PMID: 19739118
162. Yokobori T, et al. Int. J. Oncol. (2012) PMID: 22576686
163. Yokobori T, et al. Cancer Res. (2009) PMID: 19366810
164. Yokobori T, et al. Mol. Cancer Res. (2014) PMID: 24165483
165. Rajagopalan H, et al. Nature (2004) PMID: 14999283
166. Cheng Y, et al. J. Invest. Dermatol. (2013) PMID: 23381582
167. Xu Y, et al. Biomarkers (2016) PMID: 26954701
168. Li Z, et al. Int J Mol Sci (2015) PMID: 25622249
169. Welcker M, et al. Nat. Rev. Cancer (2008) PMID: 18094723
170. Akhoondi S, et al. Cancer Res. (2007) PMID: 17909001
171. Welcker M, et al. Genes Dev. (2013) PMID: 24298052
172. Welcker M, et al. Cell Div (2007) PMID: 17298674
173. Strohmaier H, et al. Nature (2001) PMID: 11565034
174. Pashkova N, et al. Mol. Cell (2010) PMID: 21070969
175. O'Neil J, et al. J. Exp. Med. (2007) PMID: 17646409
176. Malyyukova A, et al. Leukemia (2013) PMID: 23228967
177. Thompson BJ, et al. J. Exp. Med. (2007) PMID: 17646408
178. Cheok CF, et al. Nat Rev Clin Oncol (2011) PMID: 20975744
179. Ohnstad HO, et al. Cancer (2013) PMID: 23165797
180. Gamble LD, et al. Oncogene (2012) PMID: 21725357
181. Zhang et al., 2019; ASCO Abstract 3124
182. Rasco et al., 2019; ASCO Abstract 3126
183. Tolcher et al., 2021; ASCO Abstract 2506
184. Martinelli et al., 2016; EHA21 Abstract S504
185. Daver et al., 2018; ASH Abstract 767
186. Mascarenhas et al., 2019; ASH Abstract 134
187. Shustov et al., 2018; ASH Abstract 1623
188. Sallman et al., 2018; ASH Abstract 4066
189. Meric-Bernstam et al., 2017; ASCO Abstract 2505
190. Edmondson RJ, et al. Gynecol. Oncol. (2017) PMID: 28511869
191. Ambros RA, et al. Mod. Pathol. (1996) PMID: 8972476
192. Sdek P, et al. Mol. Cell (2005) PMID: 16337594
193. Brady M, et al. Mol. Cell. Biol. (2005) PMID: 15632057
194. Li M, et al. Mol. Cell (2004) PMID: 15053880
195. Brown CJ, et al. Nat. Rev. Cancer (2009) PMID: 19935675
196. Cordon-Cardo C, et al. Cancer Res. (1994) PMID: 8306343
197. Garralda et al., 2022; ENA Abstract 7
198. Horiuchi D, et al. J. Exp. Med. (2012) PMID: 22430491
199. Goga A, et al. Nat. Med. (2007) PMID: 17589519
200. Molenaar JJ, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) PMID: 19525400
201. Dammert MA, et al. Nat Commun (2019) PMID: 31375684
202. Mollaoglu G, et al. Cancer Cell (2017) PMID: 28089889
203. Cardnell RJ, et al. Oncotarget (2017) PMID: 29088717
204. Wang L, et al. Mol Oncol (2017) PMID: 28417568
205. Takahashi Y, et al. Ann. Oncol. (2015) PMID: 25632068
206. Li Y, et al. Thyroid (2018) PMID: 30226440
207. Mahadevan D, et al. PLoS ONE (2014) PMID: 24893165
208. Park SI, et al. Target Oncol (2019) PMID: 31429028
209. Kong et al., 2017; DOI: 10.21037/tcr.2017.06.41
210. Helfrich BA, et al. Mol. Cancer Ther. (2016) PMID: 27496133
211. Hook KE, et al. Mol. Cancer Ther. (2012) PMID: 22222631
212. Yang D, et al. Proc. Natl. Acad. Sci. U.S.A. (2010) PMID: 20643922
213. He J, et al. Anticancer Drugs (2019) PMID: 30540594
214. Shroff EH, et al. Proc. Natl. Acad. Sci. U.S.A. (2015) PMID: 25964345
215. Effenberger M, et al. Oncotarget (2017) PMID: 29156762
216. Qu X, et al. Biochem. Biophys. Res. Commun. (2018) PMID: 30103944
217. Xiang Y, et al. J. Clin. Invest. (2015) PMID: 25915584
218. Delmore JE, et al. Cell (2011) PMID: 21889194
219. Bandopadhyay P, et al. Clin. Cancer Res. (2014) PMID: 24297863
220. Lovén J, et al. Cell (2013) PMID: 23582323
221. Otto C, et al. Neoplasia (2019) PMID: 31734632
222. Dong LH, et al. J Hematol Oncol (2013) PMID: 23866964
223. Pei Y, et al. Cancer Cell (2016) PMID: 26977882
224. Fu XH, et al. Acta Pharmacol. Sin. (2019) PMID: 30224636
225. Bandopadhyay TK, et al. J Thorac Oncol (2020) PMID: 31655296
226. Ganesan P, et al. Mol. Cancer Ther. (2014) PMID: 25253784
227. Pereira CB, et al. PLoS ONE (2013) PMID: 23555992
228. Yasojima H, et al. Eur. J. Cancer (2011) PMID: 21741827
229. Arango D, et al. Cancer Res. (2001) PMID: 11406570
230. Bottone MG, et al. Exp. Cell Res. (2003) PMID: 14516787
231. Konopka B, et al. J. Cancer Res. Clin. Oncol. (2004) PMID: 14663583
232. Williams JA, et al. Exp. Mol. Pathol. (1999) PMID: 10600396
233. Schraml P, et al. Clin. Cancer Res. (1999) PMID: 10473073
234. Monk BJ, et al. Am. J. Obstet. Gynecol. (1994) PMID: 7977518
235. Dang CV, et al. Semin. Cancer Biol. (2006) PMID: 16904903
236. Nesbit CE, et al. Oncogene (1999) PMID: 10378696
237. Blancato J, et al. Br. J. Cancer (2004) PMID: 15083194
238. Fromont G, et al. Hum. Pathol. (2013) PMID: 23574779
239. Marin-Acevedo et al., 2021; ASCO Abstract e21016
240. Johnson DB, et al. Clin Cancer Res (2020) PMID: 31924734
241. Nikanjam M, et al. Mol Cancer Ther (2021) PMID: 33722853
242. Nebhan CA, et al. Oncologist (2021) PMID: 33861486
243. Kim DW, et al. Cancer (2017) PMID: 27911979
244. Mazieres J, et al. Ann. Oncol. (2020) PMID: 31959346
245. Hainsworth JD, et al. J. Clin. Oncol. (2018) PMID: 29320312
246. Spira et al., 2023; AACR Abstract CT032
247. Janku et al., 2021; AACR Abstract CT212
248. Sullivan RJ, et al. Cancer Discov (2018) PMID: 29247021
249. Mizumoto Y, et al. Cancer Sci. (2007) PMID: 17388789
250. Pappa KI, et al. Gynecol. Oncol. (2006) PMID: 16256179
251. Mutch DG, et al. Am. J. Obstet. Gynecol. (2004) PMID: 15118616
252. Salvesen HB, et al. Int. J. Cancer (2005) PMID: 15723290
253. Feng YZ, et al. Clin. Cancer Res. (2005) PMID: 16144912
254. He M, et al. Diagn. Mol. Pathol. (2013) PMID: 23370429
255. Holderfield M, et al. Nat. Rev. Cancer (2014) PMID: 24957944
256. Burotto M, et al. Cancer (2014) PMID: 24948110
257. Davies H, et al. Nature (2002) PMID: 12068308
258. Kandath C, et al. Nature (2013) PMID: 24132290
259. Wan PT, et al. Cell (2004) PMID: 15035987
260. Noeparast A, et al. Oncotarget (2017) PMID: 28947956
261. Smalley KS, et al. Oncogene (2009) PMID: 18794803
262. Smalley KS, et al. Cancer Res. (2009) PMID: 19351826
263. Ikenoue T, et al. Cancer Res. (2004) PMID: 15150094
264. Sen B, et al. Sci Transl Med (2012) PMID: 22649091
265. Kamata T, et al. Cancer Res. (2010) PMID: 20978199
266. Cardarella S, et al. Clin. Cancer Res. (2013) PMID: 23833300
267. Niihori T, et al. Nat. Genet. (2006) PMID: 16474404
268. Moretti S, et al. Biochim. Biophys. Acta (2009) PMID: 19735675
269. Rodriguez-Viciana P, et al. Meth. Enzymol. (2008) PMID: 18413255
270. Anastasaki C, et al. Dis Model Mech (2012) PMID: 22301711
271. Yao Z, et al. Nature (2017) PMID: 28783719
272. Sheikine Y, et al. JCO Precis Oncol (2018) PMID: 32913992
273. Garnett MJ, et al. Mol. Cell (2005) PMID: 16364920
274. Tate JG, et al. Nucleic Acids Res. (2019) PMID: 30371878
275. Jia MY, et al. Neoplasia (2014) PMID: 25030436
276. Jiao Y, et al. Oncotarget (2012) PMID: 22869205
277. Duncan R, et al. Genes Dev. (1994) PMID: 8125259
278. Kim MJ, et al. Nat. Genet. (2003) PMID: 12819782
279. Weber A, et al. BMC Cancer (2008) PMID: 19087307
280. Jang M, et al. Oncogene (2009) PMID: 19219071
281. Zheng Y, et al. Int. J. Biochem. Cell Biol. (2011) PMID: 21855647
282. Jacob AG, et al. J. Biol. Chem. (2014) PMID: 24798327
283. Rabenhorst U, et al. Hepatology (2009) PMID: 19637194
284. Singer S, et al. Cancer Res. (2009) PMID: 19258502
285. Sahn F, et al. Acta Neuropathol. (2012) PMID: 22588899
286. Bettgeowda C, et al. Science (2011) PMID: 21817013
287. Baumgarten P, et al. Neuropathol. Appl. Neurobiol. (2014) PMID: 24117486
288. Chan AK, et al. Mod. Pathol. (2014) PMID: 24030748
289. Ma H, et al. Onco Targets Ther (2020) PMID: 32158235
290. Beuselinck B, et al. Acta Oncol (2018) PMID: 29095068
291. Song Y, et al. Chin. Med. J. (2015) PMID: 26228213
292. Dornbusch J, et al. PLoS ONE (2013) PMID: 24086736
293. Terakawa T, et al. Urol. Oncol. (2013) PMID: 21478036
294. You D, et al. World J Urol (2015) PMID: 24710685
295. Silva E, et al. Breast J () PMID: 25639617
296. Grillo E, et al. Cancer Lett (2021) PMID: 33035615
297. Nishikawa et al., 2013; ASCO Abstract 2023
298. Wick et al., 2013; WCN Abstract 1207
299. Van Cutsem et al., 2011; EMCC Abstract 803
300. Baumgarten P, et al. Neuro-oncology (2016) PMID: 26627848
301. Sathornsumetee S, et al. J. Clin. Oncol. (2008) PMID: 18182667
302. Olafson LR, et al. J Clin Neurosci (2019) PMID: 31582283
303. Duda DG, et al. Oncologist (2010) PMID: 20484123
304. Stremtitz S, et al. Mol. Cancer Ther. (2016) PMID: 27535973
305. Weickhardt AJ, et al. Br. J. Cancer (2015) PMID: 26125443
306. Kopetz S, et al. J. Clin. Oncol. (2010) PMID: 20008624
307. Miles DW, et al. Br. J. Cancer (2013) PMID: 23422754
308. Fountzilas G, et al. Anticancer Res. (2011) PMID: 21868552
309. Gianni L, et al. J. Clin. Oncol. (2013) PMID: 23569311

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

ORDERED TEST # **ORD-1794439-01**
APPENDIX References

310. Sánchez-Rovira P, et al. Clin Transl Oncol (2013) PMID: 23397155
311. Cameron D, et al. Lancet Oncol. (2013) PMID: 23932548
312. Mok T, et al. J Thorac Oncol (2014) PMID: 24807156
313. An SJ, et al. Cancer Gene Ther. (2014) PMID: 24577128
314. Bais C, et al. J. Natl. Cancer Inst. (2017) PMID: 29059426
315. Cohen EE, et al. Lancet Oncol. (2009) PMID: 19201650
316. Van Cutsem E, et al. J. Clin. Oncol. (2012) PMID: 22565005
317. Lee EQ, et al. Clin. Cancer Res. (2018) PMID: 29941486
318. Xu L, et al. Cancer Res. (2009) PMID: 19826039
319. Heist RS, et al. Proc. Natl. Acad. Sci. U.S.A. (2015) PMID: 25605928
320. Giatromanolaki A, et al. Mod. Pathol. (2006) PMID: 16557278
321. Talvensaari-Mattila A, et al. Tumour Biol. () PMID: 15867479
322. Dobrzycka B, et al. Ginekol. Pol. (2010) PMID: 20695190
323. Yokoyama Y, et al. Gynecol. Oncol. (2000) PMID: 10831352
324. Koukourakis MI, et al. Cytokine (2011) PMID: 21208810
325. Piastowska-Ciesielska AW, et al. Cytokine (2013) PMID: 23273598
326. Anticancer Res. () PMID: 11908694
327. Giatromanolaki A, et al. Cancer (2001) PMID: 11745191
328. Saarelainen SK, et al. Tumour Biol. (2014) PMID: 24420153
329. Biol. Pharm. Bull. (2011) PMID: 22130231
330. Di Tommaso S, et al. Mol. Hum. Reprod. (2014) PMID: 25015674
331. Bhagwat AS, et al. Cell Rep (2016) PMID: 27068464
332. Mäkinen N, et al. Science (2011) PMID: 21868628
333. Mäkinen N, et al. Int. J. Cancer (2014) PMID: 23913526
334. Ng CC, et al. J. Clin. Pathol. (2015) PMID: 26018969
335. Kämpjärvi K, et al. Br. J. Cancer (2016) PMID: 27187686
336. Shahbazi S, et al. Am J Cancer Res (2015) PMID: 26396919
337. Halder SK, et al. Mol. Genet. Genomics (2015) PMID: 25325994
338. Markowski DN, et al. Int. J. Gynecol. Pathol. (2014) PMID: 25272295
339. Piscuoglio S, et al. J. Pathol. (2016) PMID: 26832993
340. Mishima C, et al. Breast Cancer Res. Treat. (2015) PMID: 26093648
341. Holzmann C, et al. Anticancer Res. (2015) PMID: 26637855
342. Pérot G, et al. PLoS ONE (2012) PMID: 22768200
343. Lien HC, et al. APMIS (2016) PMID: 26860948
344. Cani AK, et al. Mol. Cancer Res. (2015) PMID: 25593300
345. Lim WK, et al. Nat. Genet. (2014) PMID: 25038752
346. Croce S, et al. Eur. J. Cancer (2015) PMID: 26037152
347. Ravegnini G, et al. Mod. Pathol. (2013) PMID: 23222489
348. Guizèze R, et al. Blood (2015) PMID: 26316624
349. Kämpjärvi K, et al. Oncotarget (2015) PMID: 25595892
350. Kämpjärvi K, et al. Prostate (2016) PMID: 26383637
351. Barbieri CE, et al. Nat. Genet. (2012) PMID: 22610119
352. Yoon N, et al. Asian J. Androl. () PMID: 26924278
353. Yoon N, et al. Genes Chromosomes Cancer (2016) PMID: 26856273
354. Piscuoglio S, et al. Histopathology (2015) PMID: 25855048
355. Yoshida M, et al. Br. J. Cancer (2015) PMID: 25839987
356. Mittal P, et al. J. Clin. Invest. (2015) PMID: 26193636
357. Clark AD, et al. Crit. Rev. Biochem. Mol. Biol. (2015) PMID: 26182352
358. Ding N, et al. Mol. Cell (2008) PMID: 18691967
359. Kim S, et al. J. Biol. Chem. (2006) PMID: 16565090
360. Zhou H, et al. Mol. Cell. Biol. (2006) PMID: 17000779
361. Huang S, et al. Cell (2012) PMID: 23178117
362. Shaikhbrahim Z, et al. Endocr. Relat. Cancer (2014) PMID: 24938407
363. Turunen M, et al. Cell Rep (2014) PMID: 24746821
364. Kämpjärvi K, et al. Hum. Mutat. (2014) PMID: 24980722
365. An J, et al. Cell Rep (2014) PMID: 24508459
366. Stangl et al., 2021; ASCO Abstract 160
367. Boysen G, et al. Clin Cancer Res (2018) PMID: 30068710
368. Swami U, et al. Clin Cancer Res (2022) PMID: 36088616
369. McKay RR, et al. Clin Cancer Res (2021) PMID: 33849963
370. Abida W, et al. Proc. Natl. Acad. Sci. U.S.A. (2019) PMID: 31061129
371. Kim MS, et al. APMIS (2013) PMID: 23216165
372. Cuneo MJ, et al. Mol Cell (2023) PMID: 36693379
373. Zhang H, et al. Cancer Res (2023) PMID: 36512624
374. Zhang P, et al. Cell Death Dis (2015) PMID: 25766326
375. Wang Z, et al. Nat Rev Urol (2020) PMID: 32355326
376. Song Y, et al. Mol Cancer (2020) PMID: 31901237
377. Zhuang M, et al. Mol. Cell (2009) PMID: 19818708
378. Blattner M, et al. Neoplasia (2014) PMID: 24563616
379. Drug Discov. Today (2014) PMID: 25058385
380. Ostertag MS, et al. J Mol Biol (2019) PMID: 31026449
381. Bernasocchi T, et al. Cancer Lett (2022) PMID: 34974131
382. Morelli et al., 2023; ASCO Abstract 2547
383. Kim SP, et al. Cancer Immunol Res (2022) PMID: 35749374
384. Hirai H, et al. Cancer Biol. Ther. (2010) PMID: 20107315
385. Bridges KA, et al. Clin. Cancer Res. (2011) PMID: 21799033
386. Rajeshkumar NV, et al. Clin. Cancer Res. (2011) PMID: 21389100
387. Osman AA, et al. Mol. Cancer Ther. (2015) PMID: 25504633
388. Leung et al., 2021; ASCO Abstract 4139
389. Xu L, et al. Mol. Cancer Ther. (2002) PMID: 12489850
390. Xu L, et al. Mol. Med. (2001) PMID: 11713371
391. Camp ER, et al. Cancer Gene Ther. (2013) PMID: 23470564
392. Kim SS, et al. Nanomedicine (2015) PMID: 25240597
393. Pirolo KF, et al. Mol. Ther. (2016) PMID: 27357628
394. Leijen S, et al. J. Clin. Oncol. (2016) PMID: 27601554
395. Moore et al., 2019; ASCO Abstract 5513
396. Embaby A, et al. Gynecol Oncol (2023) PMID: 37236033
397. Oza et al., 2015; ASCO Abstract 5506
398. Lee J, et al. Cancer Discov (2019) PMID: 31315834
399. Méndez E, et al. Clin. Cancer Res. (2018) PMID: 29535125
400. Seligmann JF, et al. J Clin Oncol (2021) PMID: 34538072
401. Gourley et al., 2016; ASCO Abstract 5571
402. Park H, et al. ESMO Open (2022) PMID: 36084396
403. Jamieson et al., 2022; SGO Abstract 086
404. Dou Y, et al. Cell (2020) PMID: 32059776
405. Lin DI, et al. Gynecol Oncol (2022) PMID: 34998597
406. Wild PJ, et al. EMBO Mol Med (2012) PMID: 22678923
407. Lee EJ, et al. Gynecol. Oncol. (2010) PMID: 20006376
408. Joergers AC, et al. Annu. Rev. Biochem. (2008) PMID: 18410249
409. Kato S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) PMID: 12826609
410. Kamada R, et al. J. Biol. Chem. (2011) PMID: 20978130
411. Zerdoumi Y, et al. Hum. Mol. Genet. (2017) PMID: 28472496
412. Yamada H, et al. Carcinogenesis (2007) PMID: 17690113
413. Landrum MJ, et al. Nucleic Acids Res. (2018) PMID: 29165669
414. Bougeard G, et al. J. Clin. Oncol. (2015) PMID: 26014290
415. Sorrell AD, et al. Mol Diagn Ther (2013) PMID: 23355100
416. Nichols KE, et al. Cancer Epidemiol. Biomarkers Prev. (2001) PMID: 11219776
417. Kleihues P, et al. Am. J. Pathol. (1997) PMID: 9006316
418. Gonzalez KD, et al. J. Clin. Oncol. (2009) PMID: 19204208
419. Lalloo F, et al. Lancet (2003) PMID: 12672316
420. Mandelker D, et al. Ann. Oncol. (2019) PMID: 31050713
421. Jaiswal S, et al. N. Engl. J. Med. (2014) PMID: 25426837
422. Genovese G, et al. N. Engl. J. Med. (2014) PMID: 25426838
423. Xie M, et al. Nat. Med. (2014) PMID: 25326804
424. Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) PMID: 28669404
425. Severson EA, et al. Blood (2018) PMID: 29678827
426. Fuster JJ, et al. Circ. Res. (2018) PMID: 29420212
427. Hematology Am Soc Hematol Educ Program (2018) PMID: 30504320
428. Chabon JJ, et al. Nature (2020) PMID: 32269342
429. Razavi P, et al. Nat. Med. (2019) PMID: 31768066
430. Debiec-Rychter M, et al. Eur. J. Cancer (2006) PMID: 16624552
431. Heinrich MC, et al. J. Clin. Oncol. (2003) PMID: 14645423
432. Kamenz T, et al. World J. Gastroenterol. (2006) PMID: 16570351
433. Wang YY, et al. Proc. Natl. Acad. Sci. U.S.A. (2005) PMID: 15650049
434. Huh WK, et al. Gynecol. Oncol. (2010) PMID: 20189232
435. Tsao AS, et al. J Thorac Oncol (2011) PMID: 21892101
436. Bauman JE, et al. BMC Cancer (2012) PMID: 23033932
437. Yardley DA, et al. Clin. Breast Cancer (2009) PMID: 19933079
438. Chew HK, et al. Clin. Breast Cancer (2008) PMID: 19073506
439. Cristofanilli M, et al. Ann. Oncol. (2008) PMID: 18515258
440. Carvajal RD, et al. Clin. Cancer Res. (2015) PMID: 25695690
441. Hochhaus A, et al. J. Cancer Res. Clin. Oncol. (2015) PMID: 26002753
442. Blay JY, et al. Lancet Oncol. (2015) PMID: 25882987
443. Kajimoto N, et al. Int J Clin Exp Pathol (2015) PMID: 26722383
444. Sako H, et al. PLoS ONE (2014) PMID: 25221952
445. Hughes TP, et al. Blood (2014) PMID: 24335106
446. Takahashi N, et al. Biomark Res (2014) PMID: 24650752
447. Reichardt P, et al. Ann. Oncol. (2012) PMID: 22357255
448. Cauchi C, et al. Cancer Chemother. Pharmacol. (2012) PMID: 22119758
449. Quintás-Cardama A, et al. Nat Clin Pract Oncol (2008) PMID: 18936790
450. Bisagni G, et al. J Thorac Oncol (2009) PMID: 19461405
451. Handolias D, et al. Br. J. Cancer (2010) PMID: 20372153
452. Dişel U, et al. Lung Cancer (2011) PMID: 20970876
453. Park SH, et al. Invest New Drugs (2012) PMID: 22270258
454. Catania C, et al. Onco Targets Ther (2014) PMID: 24855380
455. Guo T, et al. Clin. Cancer Res. (2007) PMID: 17699867
456. Hu S, et al. Mol. Cancer Ther. (2008) PMID: 18483300
457. Nimeiri HS, et al. Gynecol. Oncol. (2010) PMID: 2017828
458. Matei D, et al. J. Clin. Oncol. (2011) PMID: 21098323
459. Bodnar L, et al. Gynecol. Oncol. (2011) PMID: 21723597

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

ORDERED TEST # **ORD-1794439-01**
APPENDIX
References

- | | | |
|--|---|--|
| <p>460. Heinrich MC, et al. J. Clin. Oncol. (2008) pmid: 18955458</p> <p>461. Buchbinder EI, et al. Cancer (2015) pmid: 26264378</p> | <p>462. Reichardt P, et al. BMC Cancer (2016) pmid: 26772734</p> <p>463. Hirai F, et al. Mol Clin Oncol (2016) pmid: 27073655</p> <p>464. Goemans BF, et al. Leuk. Res. (2010) pmid: 20435347</p> | <p>465. Castonguay V, et al. Gynecol. Oncol. (2014) pmid: 24882554</p> |
|--|---|--|

Disclaimer: Foundation Medicine Inc. only provides PDF report as an official issuance of the test result. Any other transformed format is not an "official / formal solution" and not guarantee the accuracy of this conversion. It is suggested the hospital to verify the outputs and validate the suitability of use.

© 2024 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 16 January 2024
 Lauren L. Ritterhouse Casariego, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | www.rochefoundationmedicine.com

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531