# SmartSeq2 quality control report

The data in this document is generated for plate CUB25DAY7, CUB25DAY0, CUB27DAY0, CUB27DAY7. Additional information regarding quality control can be found in the same folder as this report:

- QC metrics and outlier information per sample
- alignment percentages per sample
- plots used in this document

The kallisto quantifications for each of the samples and merged expression matrices (estimated counts by default) are in the neighbouring directory.

Tools used in the quality control pipeline are:

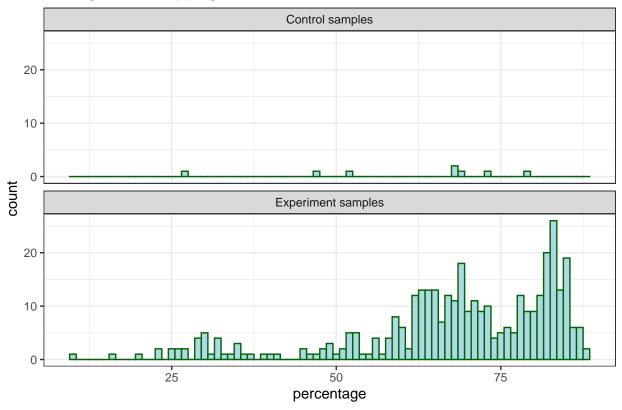
- kallisto 0.44.0
- R 4.1.2
- R markdown
- R packages: scater 1.22.0, ggplot2, dplyr, knitr, rjson
- nextflow 22.04.0
- singularity 3.8.7

#### Read mapping

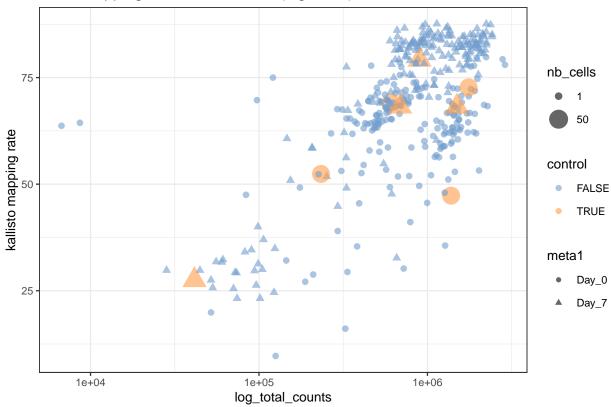
Reads were mapped with the kallisto pseudo-aligner to the reference genome.

Percentage of reads that map is shown for all the samples. Samples labelled as control are separated for comparison.

# Average read mapping rate of 67.89%



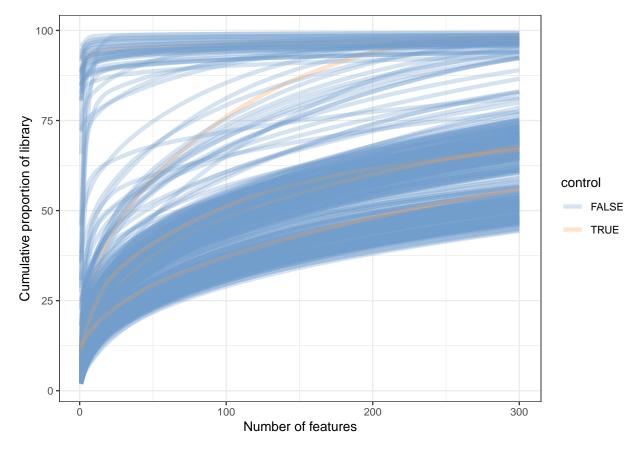
# Read mapping rate ~ total counts (log scale)



### Cumulative distribution of expression

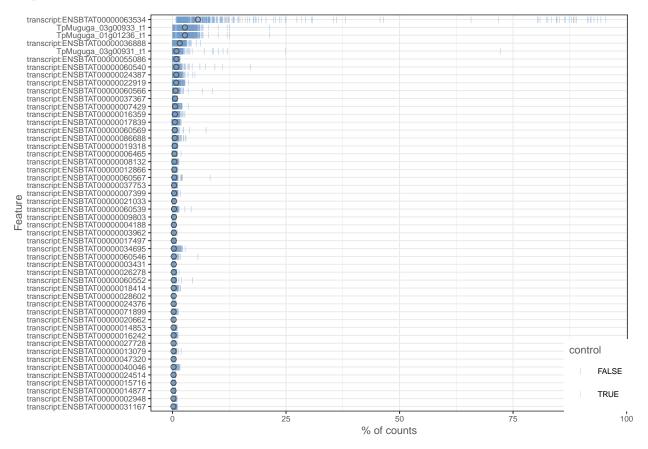
The following plot shows the proportion of the library (y-axis) covered by the top 300 most expressed transcripts (x-axis) for the given sample.

Distributions which rise high quickly are samples which are dominated by low number of transcripts while the ones having a less steep rise have counts more evenly distributed. Wells with less diverse count distribution are more similar to empty wells and might indicate lower quality or damaged cells.



### Most highly expressed transcripts

Shown are top 50 features by the proportion of counts they take in all samples. Feature names (as Ensembl transcripts or annotated control features) denote samples on the y-axis with the percentage of counts they capture on the x-axis. A circle shows the mean proportion across all samples with proportion for each of samples shown on the same line.

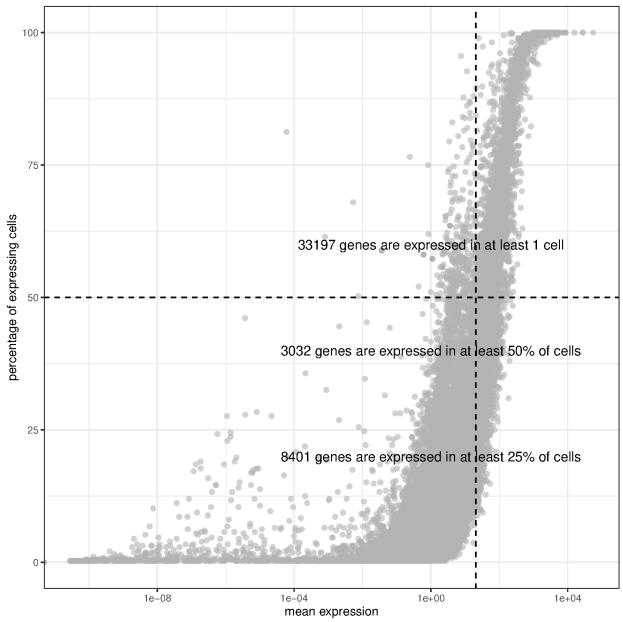


### Expression frequency - mean distribution of transcripts

For all of the transcripts (features), their frequency of expression (percentage of samples expressing the specific transcript) is shown against their mean value of expression

The relationship between the two variables is typically sigmoidal looking.

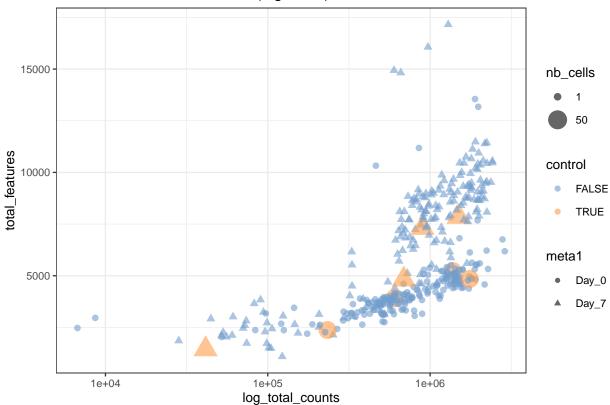
The vertical dashed line is the median of expression levels across the samples. The horizontal dashed line is at 50% of expression presence - dots above are transcripts which are expressed in more than 50% of the samples.



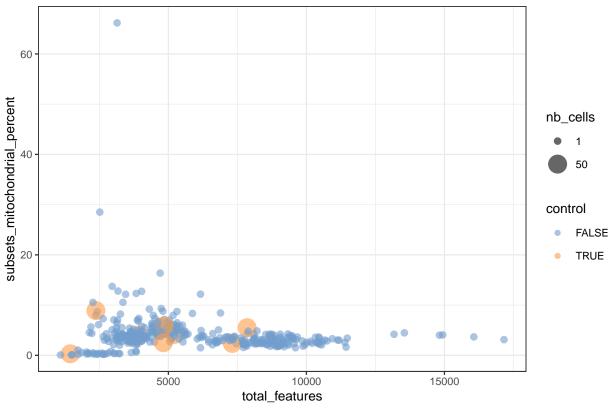
### Scatterplots

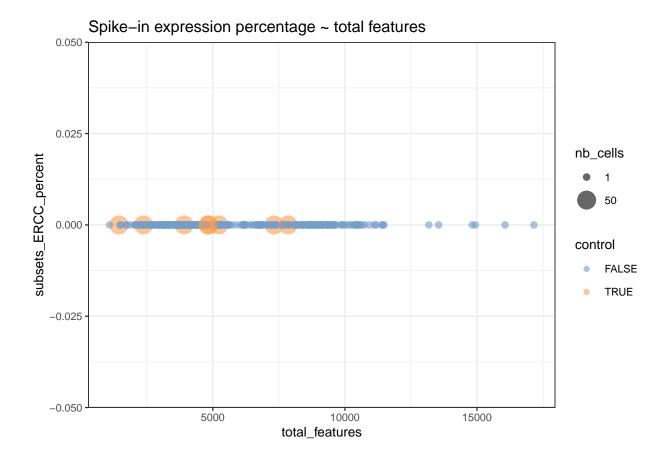
Following scatterplots show, for all of the samples and their total counts, their transcript number, proportion of mitochondrial transcript and proportion of ERCC transcript.



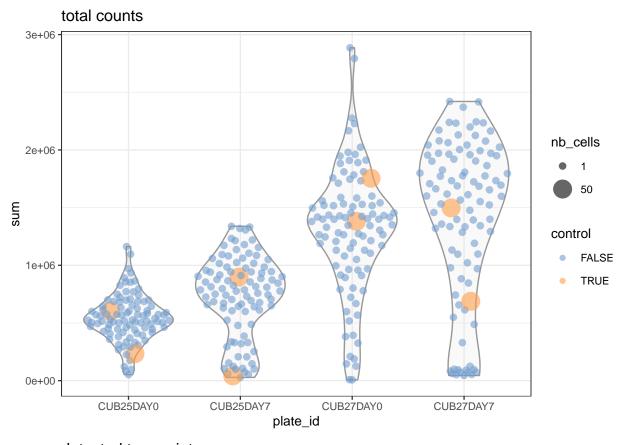


Out of 48113 transcripts, Mitochondrial expression percentage ~ total features

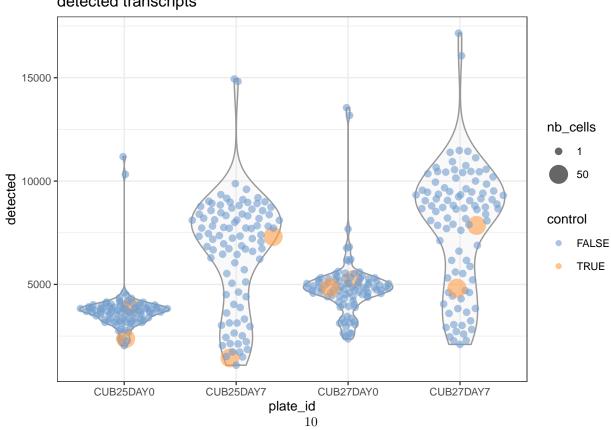


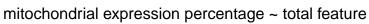


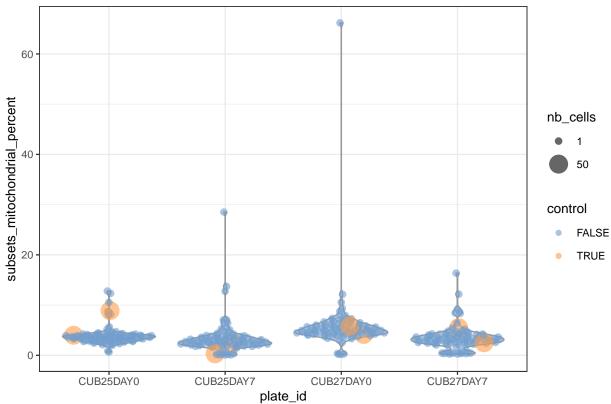
### Violin plots

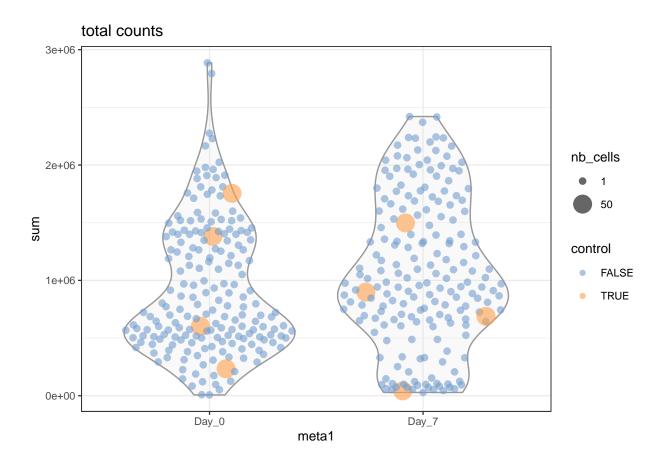


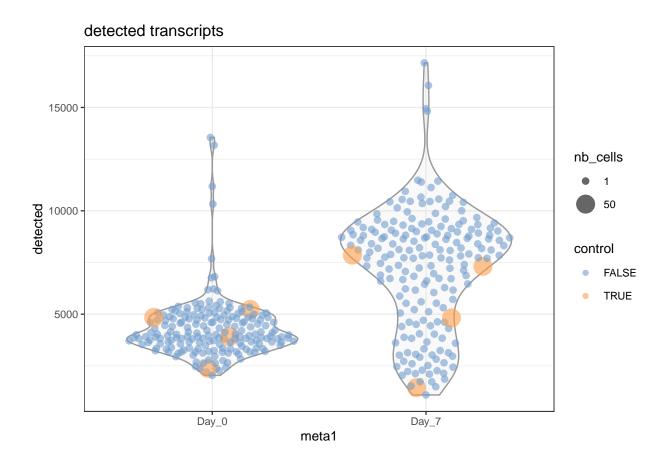
# detected transcripts

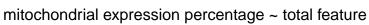


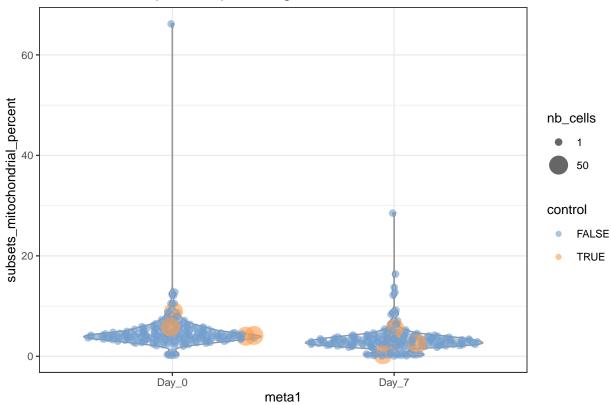












#### Dimensionality reductions

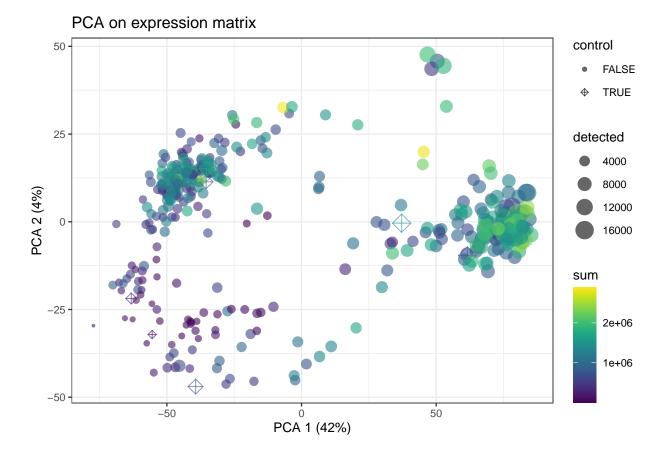
Dimensionality reductions summarise the large feature space with a smaller number of dimensions aiming to capture most of the variance in the data.

Principal component analysis (PCA) and t-SNE are one of the most common methods. PCA makes new dimensions by trying to capture as much as variance as possible in the top ones, while t-SNE tries to place the samples in 2 dimensions in a way which best captures the similarities and differences of the samples.

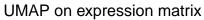
PCA/t-SNE dimensions are derived from the expression profiles of all the transcripts and visualised in the following scatterplots.

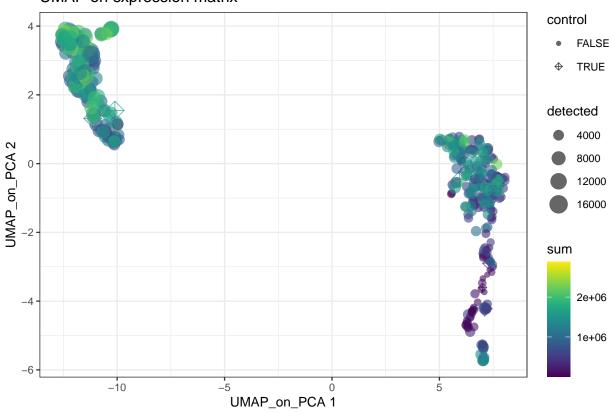
Samples similar in expression profile will be plotted closer together.

#### PCA

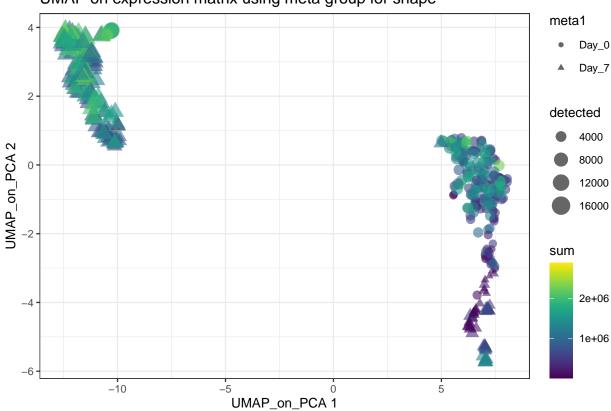


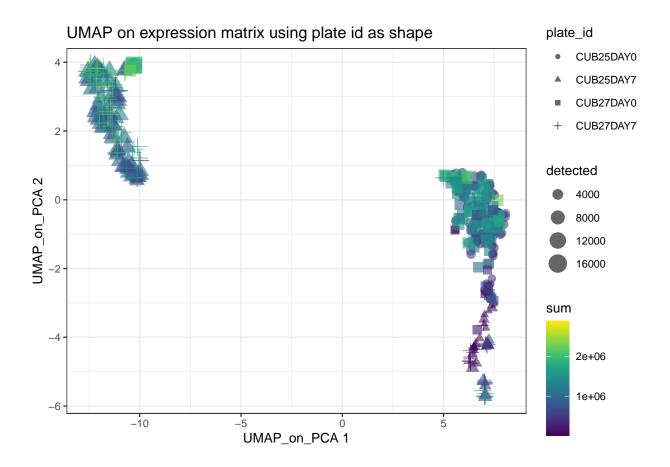
#### **UMAP**



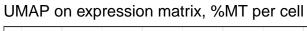


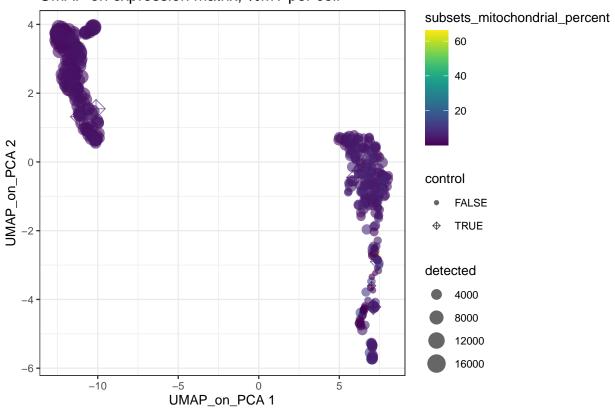
### UMAP on expression matrix using meta group for shape



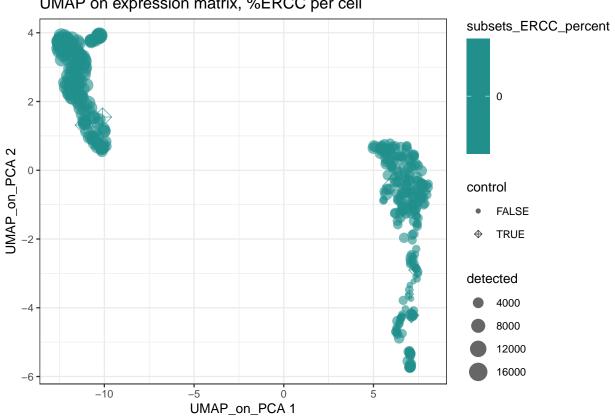


### UMAP on MT/ERCC





## UMAP on expression matrix, %ERCC per cell



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#### Outliers and key metrics per sample

The accompanying spreadsheet (Per\_sample\_key\_metrics.tsv) contains the value for some of the most important metrics, position on the plate, control information and whether or not the sample is considered an outlier in the population when it comes to mapping rate, and percentage of counts which are mitochondrial.

If a sample's average read mapping rate is below 50%, or the mitochondrial percentage is over 10%, the sample is consider as an outlier.

Being an outlier on one metric is unlikely to be of much concern, but if the same sample is an outlier in both metrics, it could be a sign of an unviable sample.

The following samples have been marked as **outliers**.

For 2 outlier metrics:

 $R0851-S0001\_A67379\_CUB25DAY7D8, R0851-S0001\_A67379\_CUB27DAY0B12, R0851-S0001\_A67379\_CUB27DAY0F10$ 

Exact information for the metrics used in the generation of this document can be found in the accompanying table - Per\_sample\_key\_metrics.tsv

By default, it contains the following information:

- plate\_id
- fastqname
- plate\_position
- outlier\_hits the number of times that sample has been labeled as an outlier
- outlier\_pct\_MT TRUE if a sample meets the outlier metric on mitochondrial percentage
- outlier\_mapping reate TRUE if a sample meets the outlier metric on average read mapping rate
- sum total counts
- detected total expressed features (transcripts)
- subsets\_mitochondrial\_percent Percentage of counts belonging to mitochondrial transcripts
- subsets\_ERCC\_percent Percentage of counts belonging to spike-ins
- $\bullet \;\; \text{map\_rate} \; \text{-} \; \text{average read mapping rate}$
- $\bullet$  is\_cell\_control