

Announcements

- Problem set 2 due today
- Problem set 3 out later today
 - HMMs: Viterbi, Forward-backward, ...

Today's lecture

- · Finish log probabilities
- Continuing Hidden Markov models (HMMs)
 - One form of unsupervised learning
- Application of HMMs to genetics:
 - Background on genetic assays
 - Inferring haplotypes in families
 - Background on inferring haplotypes in unrelated samples

Numerical stability in evaluating HMMs

Issue: computers store numbers in finite space

- To analyze HMMs, we multiply many numbers < 1
 - Can produce underflow: a number too small for the computer to store
- Example:
 - Want to analyze a sequence of length 10,000 bases
 - $\ a_{kl} = .1 \ \text{for all} \ k, l \qquad \qquad e_k(b) = 1 \ \text{for some} \ k, b \ \text{pair}$
 - Viterbi path score: 10^{-10,000}
 - > Way too small for standard floating point representations

Underflow example

• Underflow example in Python:

```
>>> prob = 1
>>> for i in range(0,10000):
... prob *= .1
...
>>> prob
0.0
```

- · Solution: log probabilities
 - Effective way to compute using very small numbers
 - Have $\log(x^y) = y \log(x)$ much easier number to represent for large positive/negative exponent y

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Example: no underflow using log probabilities

```
>>> from math import log
>>> logProb = 0
>>> logTransitionProb = log(.1)
>>> for i in range(0,10000):
... logProb += logTransitionProb
...
>>> logProb
-23025.850929942502
>>> logProb / log(.1)
```

10000.00000000089

Notes:

- log() is expensive
 Should precompute probabilities of HMM parameters
- Log probabilities are often more efficient because we sum instead of multiply

Issue: need to be able to add probabilities

- Forward & backward algorithms sum probabilities
 Inconvenient: log(x) + log(y) ≠ log(x + y)
- · Can try converting to normal space
- Approach 1:

 log(exp(a) + exp(b)) ←

 (Here a, b are log probabilities)

Mathematically correct, but

• Can underflow:

exp(-1000) == 0.0Expensive: two exp() and one log() calls

Improved sum of log probabilities

• Let a, b be log probabilities, then

```
\begin{split} \exp(a) + \exp(b) &= \exp(a) \cdot \left(1 + \frac{\exp(b)}{\exp(a)}\right) = \exp(a) \cdot (1 + \exp(b - a)) \\ \text{So:} \quad \log(\exp(a) + \exp(b)) &= \boxed{a + \log(1 + \exp(b - a))} \end{split}
```

- Better numerical stability:
 - Worst case for approach 1: $\exp(a) + \exp(b) == 0.0$, so get $\log(0.0)$: $-\infty$ or undefined
 - Worst case for above approach: exp(b-a) == 0.0, so get a + log(1.0 + 0.0) == a ← much better
- Better computation:
 - Now only one $\texttt{exp}\left(\right)$ and one $\texttt{log}\left(\right)$ call

Even better sum of log probabilities

- If $\exp(b-a)$ is very small (e.g., 10^{-20}), can have $1+\exp(b-a) == 1.0$, and thus $\log(1) == 0$
- Solution: log1p(x)
 - Computes log(1+x) at higher precision:

```
>>> log1p(1e-20)
9.99999999999995e-21
>>> log(1+1e-20)
0.0
```

- This is related to the lower.tail=FALSE option in R:
 - One-tailed p-value definition: p=1-F(x), F the CDF: $P(X \le x)$ pnorm(7) == 1.0 \Rightarrow 1-pnorm(7) == 0.0 pnorm(7, lower.tail=FALSE) == 1.279813e-12

Python implementation to sum log probabilities

```
from numpy import log1p
from math import exp
def sumLogProb(a, b):
    if a > b: return a + log1p(exp(b - a))
    else:        return b + log1p(exp(a - b))
```

- If statement:
 - Why do we prefer max of a and b?
 - Trying to sum the corresponding probabilities: is $\geq \max(a, b)$
- For efficiency, Durbin et al. suggest generating a table for log(1 + exp(b-a))
 - Idea: b-a often close to 0, so don't need large table
 - Use linear interpolation between table values

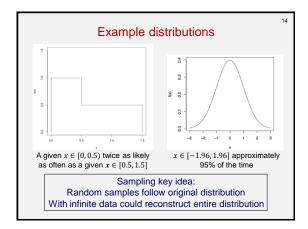
Hidden Markov models How do we sample from P(z|x)?

Use forward probabilities

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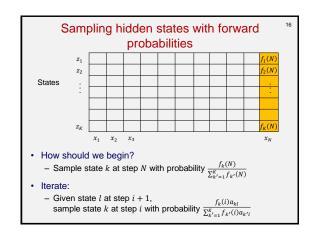
Background: what is random sampling?

- Sampling: choosing a subset of a population in order to identify characteristics of the full population
 - Example (simple random sampling):
 Given a population of 10,000 students, select 500 at random with equal probability
- · Can sample from a given distribution
 - Want sampling to give a value x with probability proportional to its density (i.e., pdf) f(x)



Why sample state path?

- · Posterior decoding isn't always a valid state path
- · Viterbi decoding is the most likely path
 - But only one path: could be many other likely paths
- Can sample z from P(z|x)
 - Is a distribution over vectors z (typically high dimensional)
 - Can sample z multiple times as alternate to expectation maximization (EM – to be discussed later)
 - · More randomized: possibly less prone to find local maxima
- · How is this different from generating random data?
 - Generating samples from P(x, z) gives both x and z
 - Want a random \mathbf{z} sampled from $P(\mathbf{z}|\mathbf{x})$: fixed observed data



Randomized training algorithm

- 1. Initialize $\theta^{(0)} = \left(a_{kl}^{(0)}, e_k^{(0)}(b)\right)$ for all k, l, b
- 2. Iterate:
 - 1. Run forward algorithm: compute $f_k(i)$ for all k, i
 - 2. Sample n hidden state vectors \mathbf{z} from $P(\mathbf{z}|\mathbf{x}, \boldsymbol{\theta}^{(t)})$
 - 3. Calculate $A_{kl}^{(t)}, E_k^{(t)}(b)$ using all n paths \mathbf{z} [as in supervised]
 - 4. Calculate new $\boldsymbol{\theta}^{(t+1)} = (a_{kl}^{(t+1)}, e_k^{(t+1)}(b))$ for all k, l, b [MLE]
- 3. Terminate when $\Delta P(x|\theta)$ small or fixed # iterations
- Notes:
 - May not converge as quickly as EM: randomized
 - But stochastic so may yield higher $P(x|\theta)$

Desired uses of HMMs (highlighted done)

- Evaluation:
 - **Given**: observed \emph{x} and HMM specification

Question: what is the joint probability of *x* and a given *z*? **Question:** what is the likelihood of *x* based on the HMM?

- · Decoding:
 - **Given**: observed x and HMM

Question: what sequence of hidden states produced x?

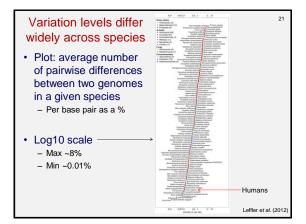
- Viterbi decoding: most likely hidden state sequence
- <u>Posterior probability of hidden states</u>: probability of each state z_i producing each x_i
- · Learning:
 - Given: observed x and HMM without complete probabilities
 Question: what emission, transition probabilities produced x?

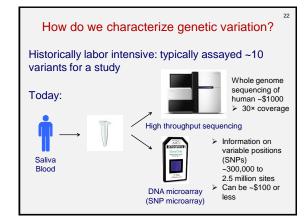
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Applying HMMs to genetic data Background: genetic assays

Genetic variants of interest for many analyses

- Genomes of any two humans are > 99.9% identical
- · However, variation exists:
 - Approximately $.001 \times 3 \times 10^9 = 3 \times 10^6$ variants between a pair of human genomes
- · Why do we care about variation?
 - Genetic variation drives biological variation
 - > Want to characterize it to understand biological impacts of variants
 - Genetic variation reveals population genetics
 - Must account for/model these dynamics to avoid false discoveries related to biology
 - > Of direct interest to studying evolutionary & demographic history
 - · Will see examples of both the above later





SNP microarrays

• Microarray: flat substrate (often glass or silicone) with numerous probes

- Many kinds of microarrays: protein, antibody, DNA, etc.

1. Sample DNA is labeled with florescent dye

2. DNA allowed to hybridize with probes on microarray

3. Lasers used to detect binding to probe sequences

SNP microarrays

• To design probes, need:

- Sequence flanking SNP

- Ideally whole genome sequence of species to ensure probe sequence is unique

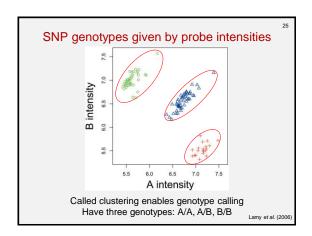
ACTTTCAGTGTCGCAT ACTTTCATTGTCGCAT Probe target

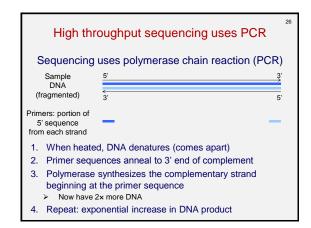
• Probe sequences:

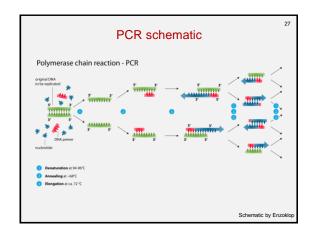
- Are complementary to flanking sequence

• Example: ATGCGACAA (recall: pair is reverse complement) ATGCGACACC

- Are relatively long (often 50 bp)







High throughput sequencing

• Sequencing details complicated, but at a high level:

- Sequencer performs PCR

- Nucleotides added during elongation are florescently labeled; colors differ between A, C, G, T

- Machine detects color of each incorporated nucleotide

• Above is done across millions of DNA fragments

• Produces "reads" of specific lengths

- Often 100 or 150 bp

• For humans, 30x coverage, 100 bp reads ≈ 960 million reads!

Sequencing next step: read mapping

• SNP microarrays have probes at known locations

- Probes chosen to be unique (based on human reference genome)

• Sequencing reads can come from anywhere

- This leads to the problem of read mapping / alignment

> Must determine where in the genome a given 100 bp sequence comes from

- May discuss methods for this later this term

Final step: genotype calling

• After read mapping: many reads overlap a position

- Reads are mapped from both homologs (blue, orange)

- Homologs will have differences (variants) at some positions

Genotype calling: method for deciding genotypes based on some input (SNP array intensity plot or mapped reads)

> Won't discuss, but interesting; can view as hypothesis testing

> Some packages (e.g., GATK) use machine learning techniques

Applying HMMs to genetic data
Inferring haplotypes in family data

