

Self-contained RNA inhibition with trans-cleaving ribozymes

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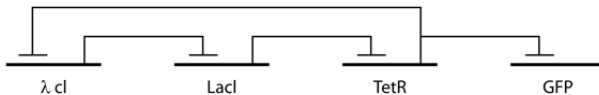
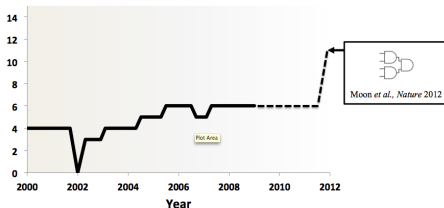
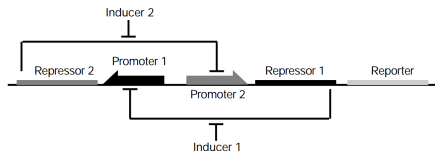
Promise of Synthetic Biology

Complexity of eukaryotes \approx # of protein coding genes		
Oryza sativa (rice)	470 million	51,00
Gallus gallus (chicken)	1 billion	20,000-23,00
Canis familiaris (dog)	2.4 billion	19,00
Mus musculus (mouse)	2.5 billion	30,00
Homo sapiens	2.9 billion	20,000-25,000

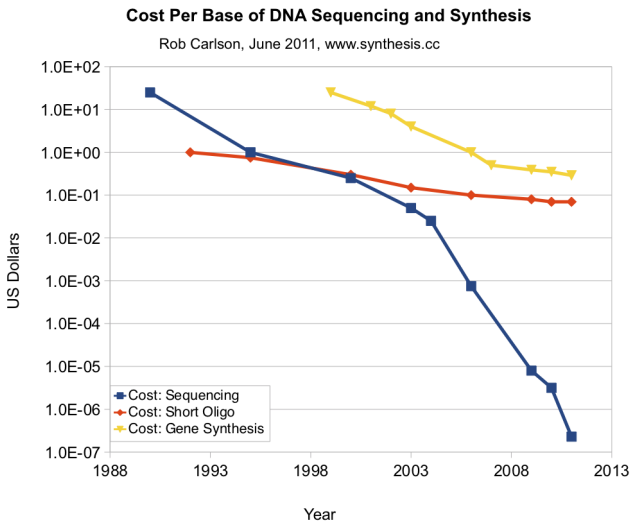
The root of biological complexity is believed to be genetic regulation. However, the creation of novel protein regulatory elements is too difficult. Re-writers RNA-world may be the key to getting a handle on regulation.

Motivation

Toggle switch and repressilator in 2000, to now.



Exponential decrease in the cost of enabling technologies should result in exponential growth of circuit complexity.



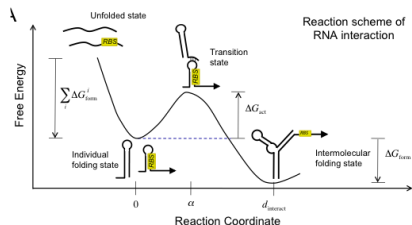
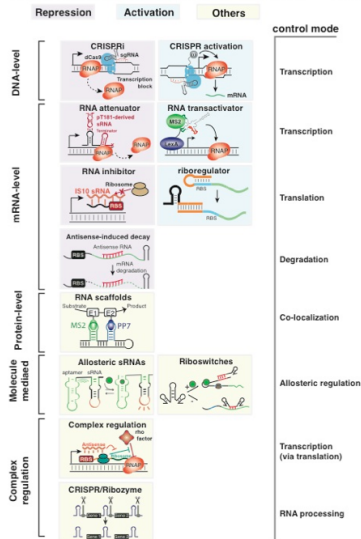
Pitfalls in current promoter-repressor pair design

- Orthogonal - Limited number of repressors (until very recently)
- Predictable - Gene circuit evolves away
- Safe - shRNA toxicity in gene therapy
- Reliable - 40 hour toggle switch breakdown
- Designable - Protein structure prediction too difficult
- Composable - Unpredictable behavior when juxtaposed

Signal Transduction with RNA, possible tools

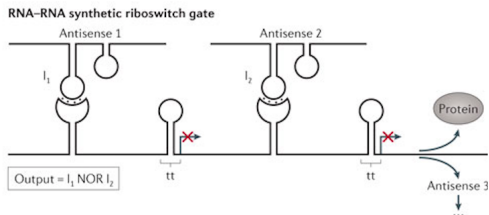
A summary of noncoding RNA tools for genetic engineering

Qi & Arkin, Figure 1



Choice of trans-cleaving hammerhead ribozymes

Self-contained mechanism of RNA degradation
Composable and functionally complete: NOR gate



Base-pairing rather than aptamer coupling for ease of rational design
Watson Crick base pairing dominates free energy minimization

General riboregulation model

Stochastic model - Gillespie algorithm

Algorithm 1 Gillespie

Inputs:

Set of M reactions; $R_i = (c_i) i \in 1, \dots, M$

Initial population sizes, $endtime$

Output: Catalog of Molecular events

while $\tau < endtime$ **do**

$a_0 = 0$

for $i = 1$ to M **do**

$a_i = h_i c_i$, where h_i is the amount of reactant

$a_0 += a_i$

end for

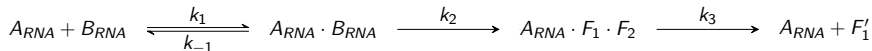
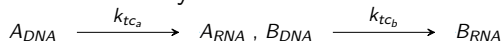
$(\tau, \mu) \leftarrow P(\tau, \mu)$

 updatePopulation(R_μ)

end while

trans-cleaving ribozyme model and kinetics

From Samarsky *et al.*



All species undergo degradation at some rate k_{deg} .

Approximated by MM kinetics

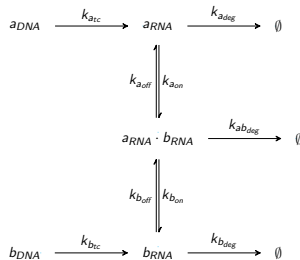
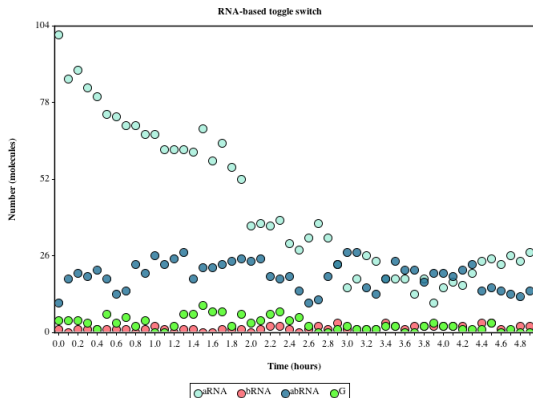
Fraction of substrate/time: $k_{obs} = k_2 \times [A_{RNA} \cdot B_{RNA}] / [B_{RNA}_0]$

$k_1 \gg k_{-1}$, and tunable.

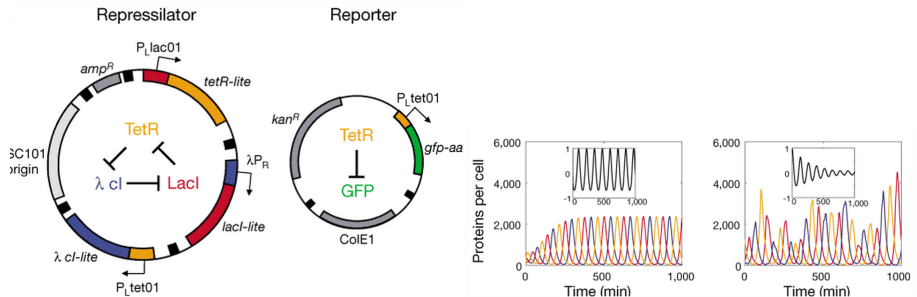
$k_3 \gg k_2 \gg k_{deg}$ by single orders of magnitude.

Toggle switch model riboregulation

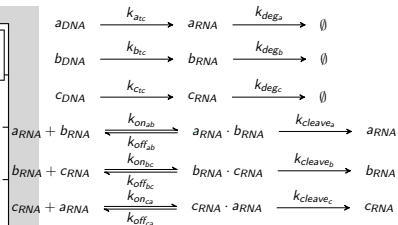
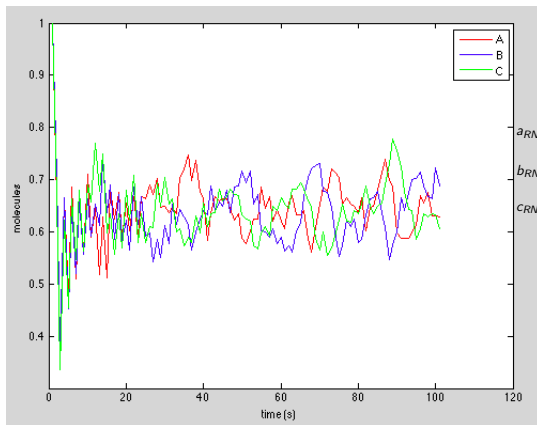
No possible bistable point. First limitation.



Classic Repressilator



Riboregulatory Repressilator



Repressilator Comparison

Rate of oscillation

Tunable:

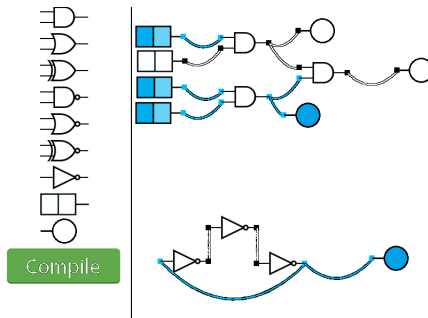
- promoter strength - well characterized process
- strength of binding - determined by W-C Base-pairing
- cleavage rate - fairly fixed
- degradation rate - introduction of cleavage sites

Character of trans-cleaving ribozymes

- Orthogonal - determined by WC Base-pairing
- Predictable - less stress on cells, cleaving reaction well characterized
- Safe - mRNA is less persistent, and less chemically reactive
- Reliable - still possible for cell to mutate
- Designable - minimization of free energy based on WC Base-pairing
- Composable - Unpredictable behavior when juxtaposed

Further work

Biocircuit-Design Automation, Biocompiler



Thanks!

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Sources

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