

MATHEMATICAL MODELING OF RECOMBINANT DNA EXPRESSION

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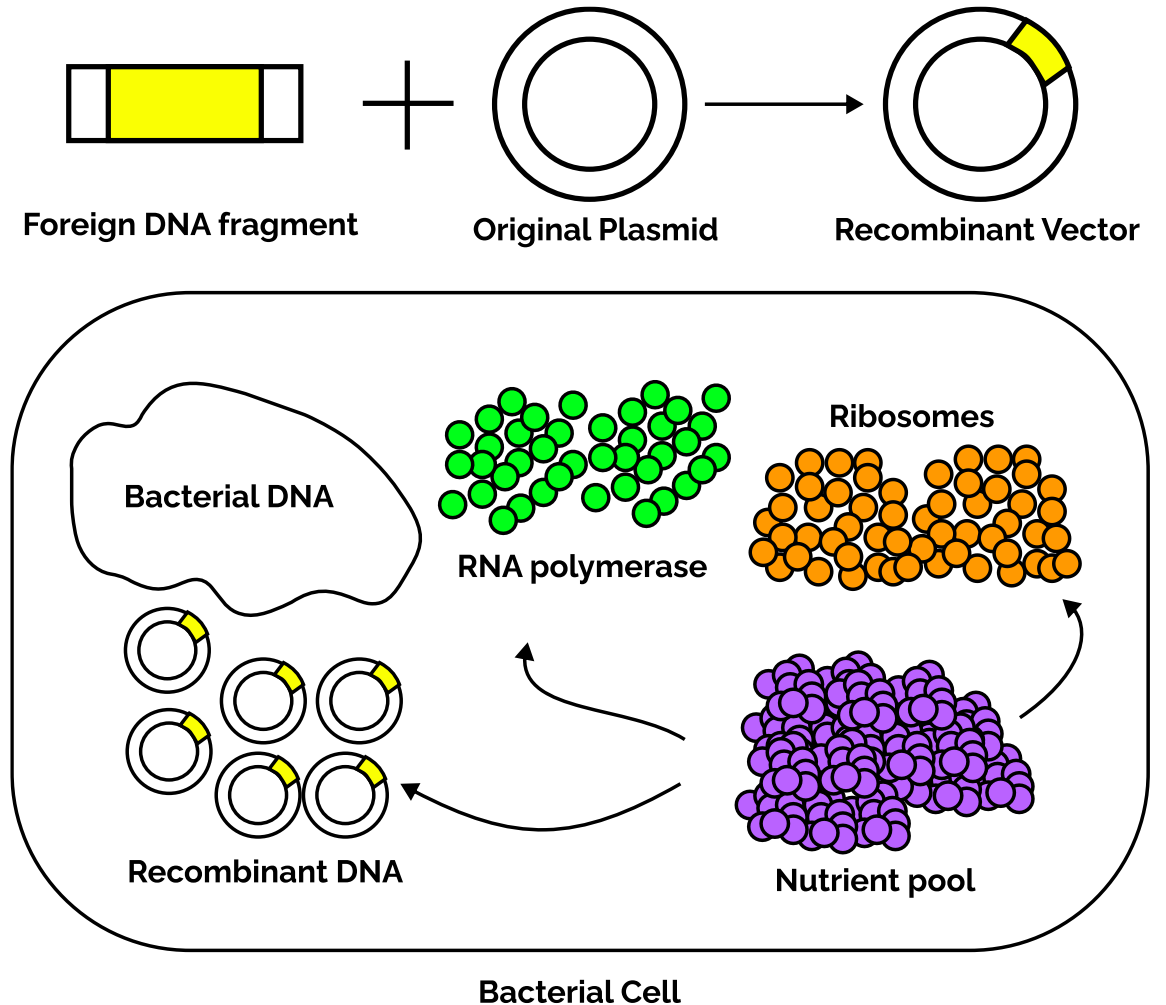


INTRODUCTION

INTRODUCTION TO DNA RECOMBINATION

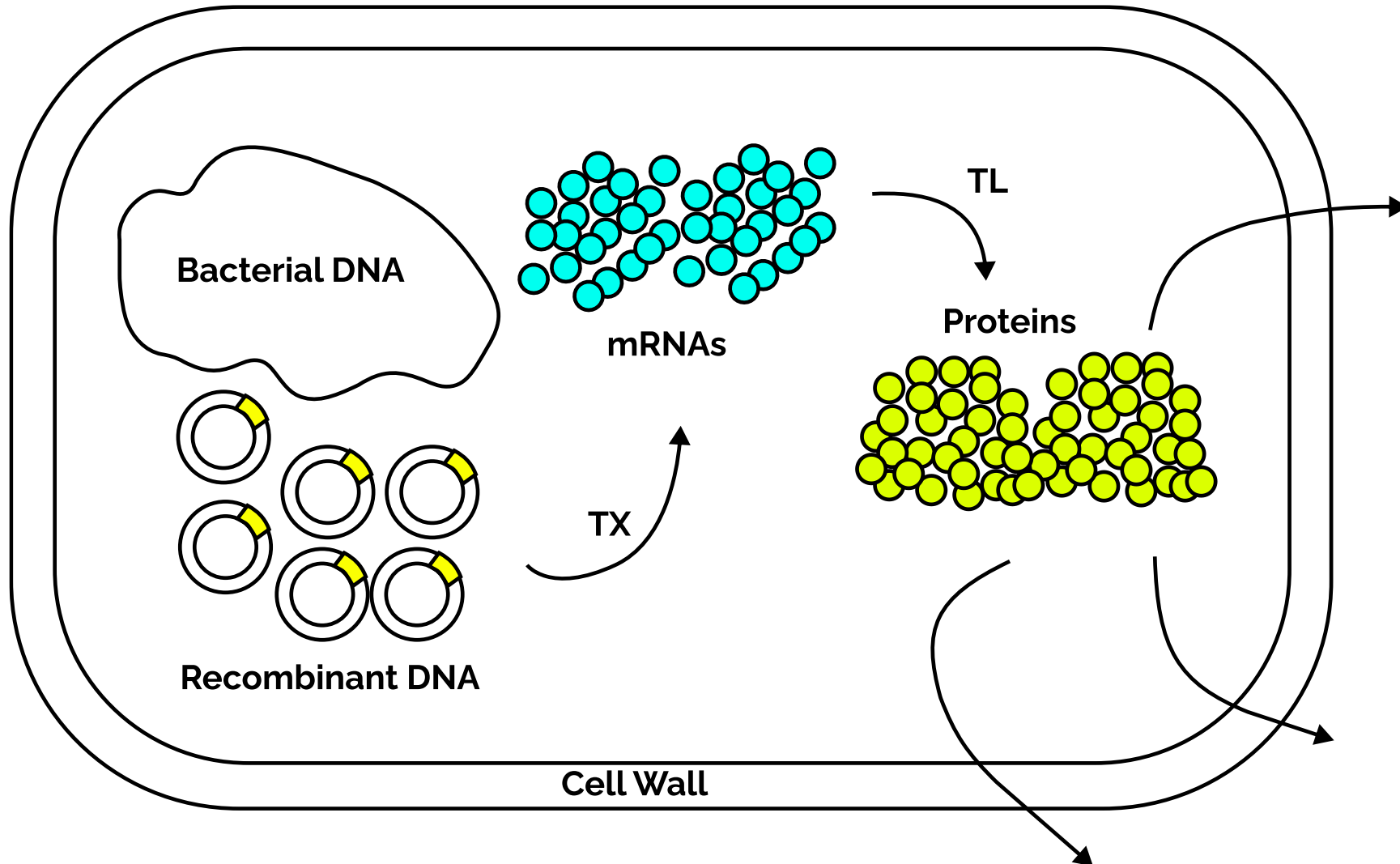
Genetic Engineering

- Most proteins are difficult to manufacture with available synthetic capabilities
- Genes that are involved in producing protein molecules can be assembled through *gene manipulation* to a *cistron*
- The cistron encodes necessary information and utilize intrinsic gene expression machinery to produce target proteins



INTRODUCTION TO DNA RECOMBINATION

Gene expression



OBJECTIVES

General Objective

- To develop a minimalistic mathematical model that grasps the basic mechanics of recombinant DNA expression

Specific Objectives

- To understand the crucial checkpoints of protein expression
- To understand the relationship between host history and protein production



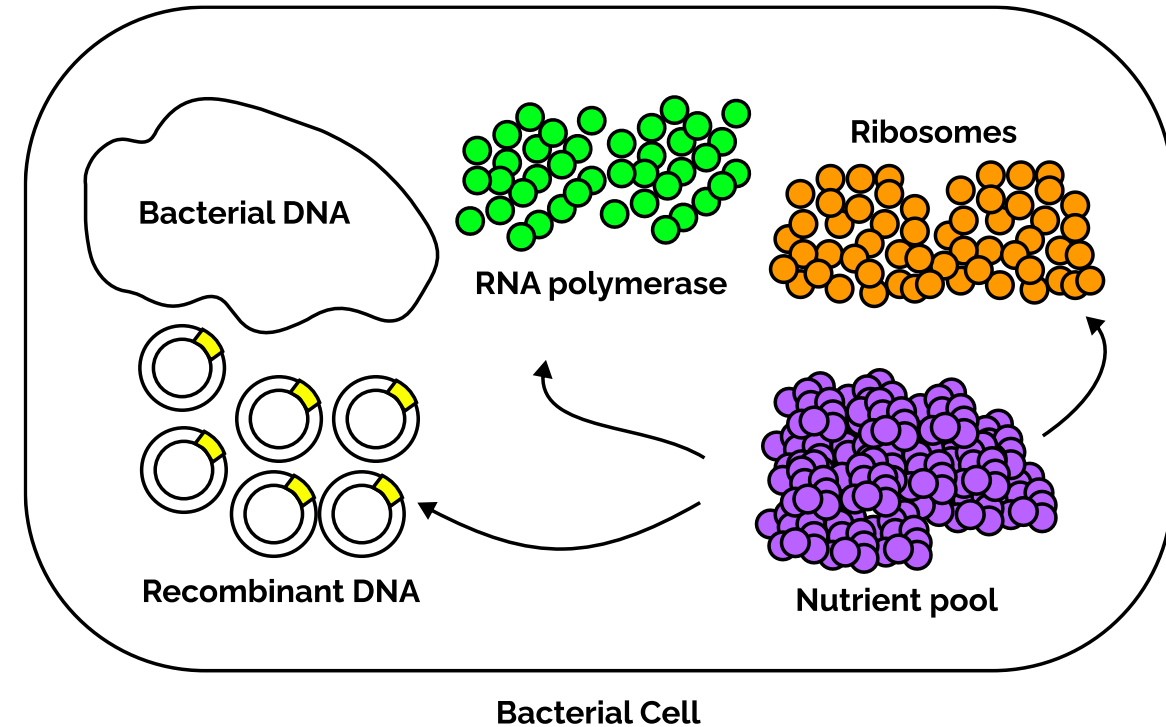
MODEL DEVELOPMENT

MODEL DEVELOPMENT

STEP 1

Assumptions

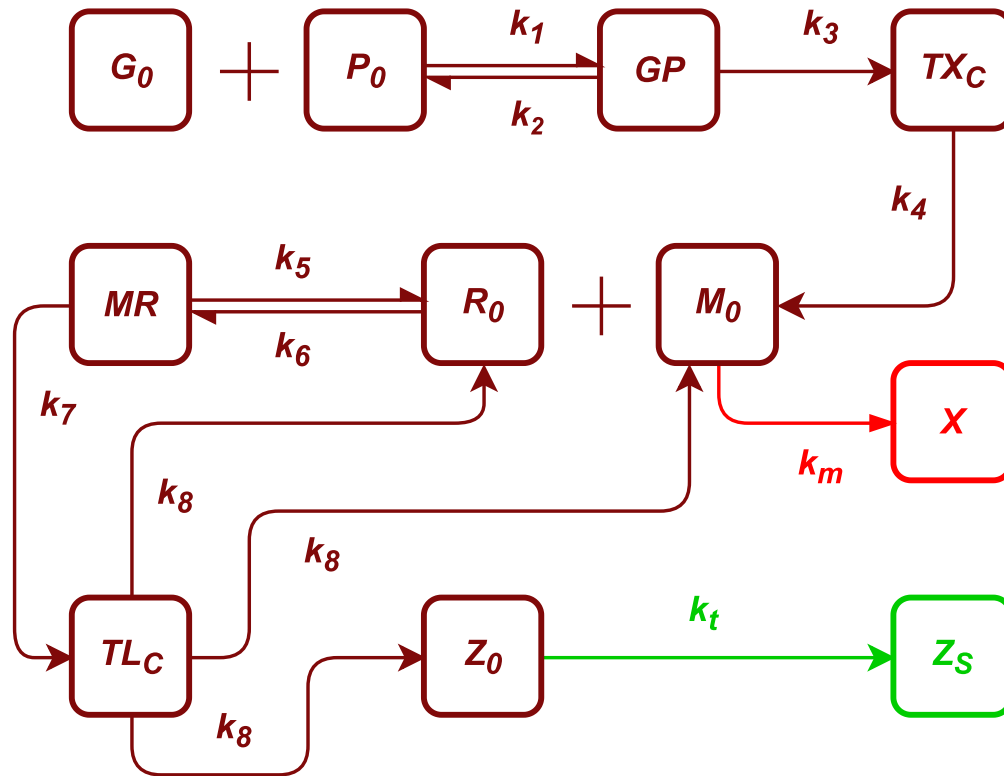
1. The bacterial cytoplasm is assumed to be homogenous and posses a fixed volume
2. The biochemical nutrient pool is large and the changes during consumption is negligible
3. No replication occurs within the simulation time/observation time
4. The resources available for expression is assumed to be in high numbers and continuum hypothesis and mass action-based formalisms become applicable
5. The synthesized proteins are assumed to be transported outside (i.e., to culture supernatant)



MODEL DEVELOPMENT

STEP 2

Understanding the biochemical interactions and developing a chemical reaction network



G_0 – plasmid

P_0 – RNA polymerase (RNAP)

GP – RNAP-plasmid complex

TX_c – escape committed complex

M_0 – mRNA

R_0 – ribosome

MR – mRNA-ribosome complex

TL_c – translation complex

Z_0 – reporter protein

Z_s – secreted protein

k_1 – RNAP-promoter association

k_2 – RNAP-promoter dissociation

k_3 – transcription commitment

k_4 – mRNA production

k_5 – mRNA-ribosome association

k_6 – mRNA-ribosome dissociation

k_7 – translation commitment

k_8 – protein production

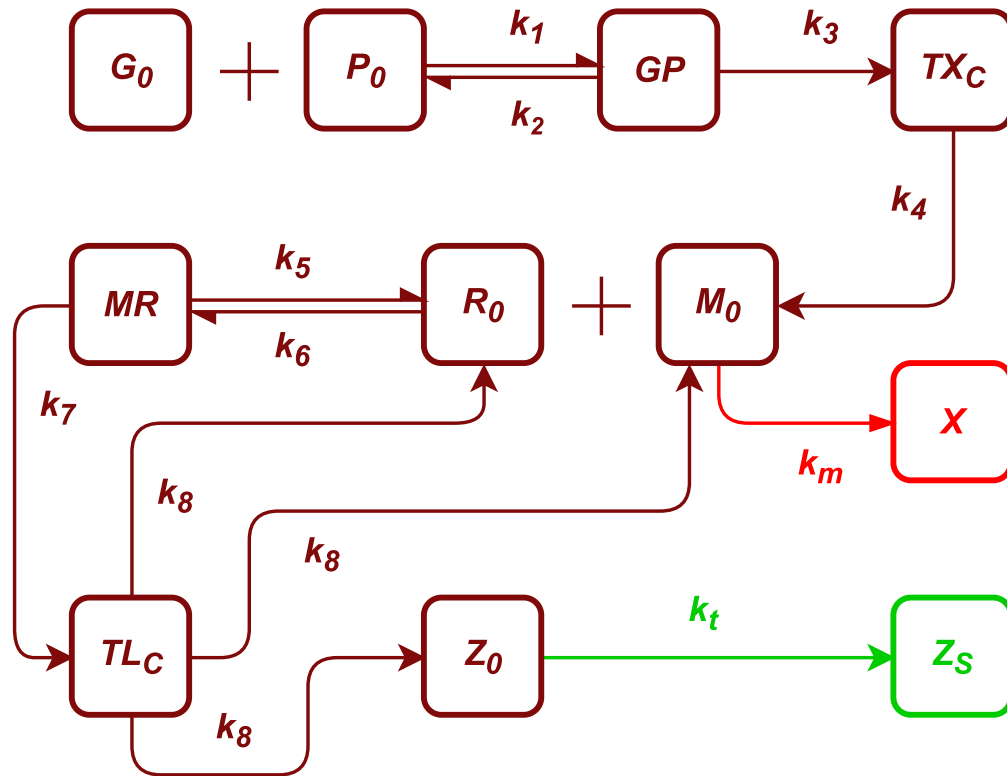
k_m – mRNA degradation

k_t – protein transport

MODEL DEVELOPMENT

STEP 3

Develop differential equations for the formulated reaction network



Differential Equations

$$dG_0 = -k_1 G_0 P_0 + k_2 GP + k_4 TX_C$$

$$dP_0 = -k_1 G_0 P_0 + k_2 GP + k_4 TX_C$$

$$dGP = k_1 G_0 P_0 - k_2 GP - k_3 GP$$

$$dTX_C = k_3 GP - k_4 TX_C$$

$$dM_0 = k_4 TX_C - k_m M_0 - k_5 M_0 R_0 + k_6 MR + k_8 TL_C$$

$$dR_0 = -k_5 M_0 R_0 + k_6 MR + k_8 TL_C$$

$$dMR = k_5 M_0 R_0 - k_6 MR - k_7 MR$$

$$dTL_C = k_7 MR - k_8 TL_C$$

$$dZ_0 = k_8 TL_C - k_t Z_0$$

$$dZ_s = k_t Z_0$$

MODEL DEVELOPMENT

STEP 4

Carry out simulation for a collection of time points using a computational software (*R* and the package *deSolve*)

Initial Concentrations	
Go – 0.5 nM	TL _C – 0 nM
Po – 0.5 nM	Zo – 0 nM
GP – 0 nM	Zs – 0 nM
TX _C – 0 nM	Mo – 0 nM
Ro – 0.5 nM	MR – 0 nM

Rate Constants	
$k_1 - 6 \times 10^9 \text{ M}^{-1}\text{min}^{-1}$	$k_7 - 30 \text{ min}^{-1}$
$k_2 - 6 \times 10^9 \text{ M}^{-1}\text{min}^{-1}$	$k_8 - 0.9 \text{ min}^{-1}$
$k_3 - 60 \text{ min}^{-1}$	$k_t - 60 \text{ min}^{-1}$
$k_4 - 0.9 \text{ min}^{-1}$	$k_m - 18 \text{ min}^{-1}$
$k_5 - 6 \times 10^9 \text{ M}^{-1}\text{min}^{-1}$	$k_6 - 135 \text{ min}^{-1}$

Code Availability

<https://github.com/zachariah-ibrahim/recombinant-DNA-expression>

Parameter availability

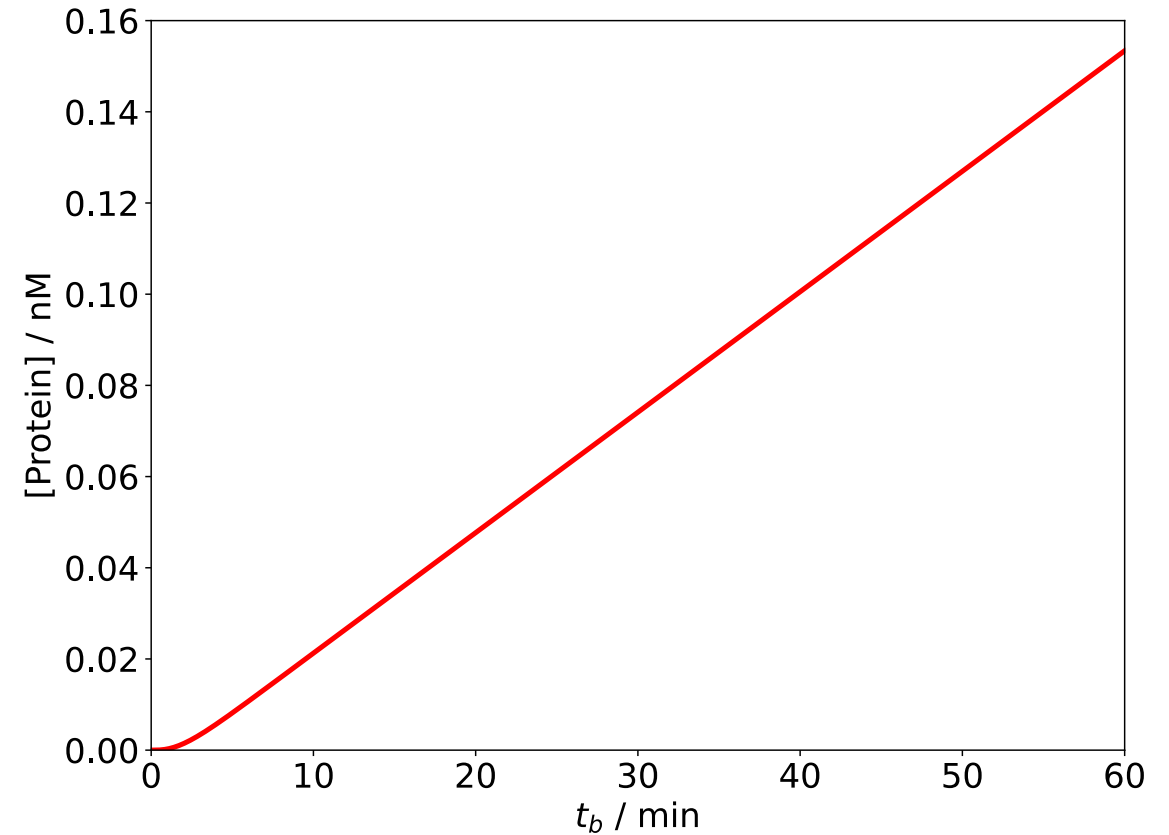
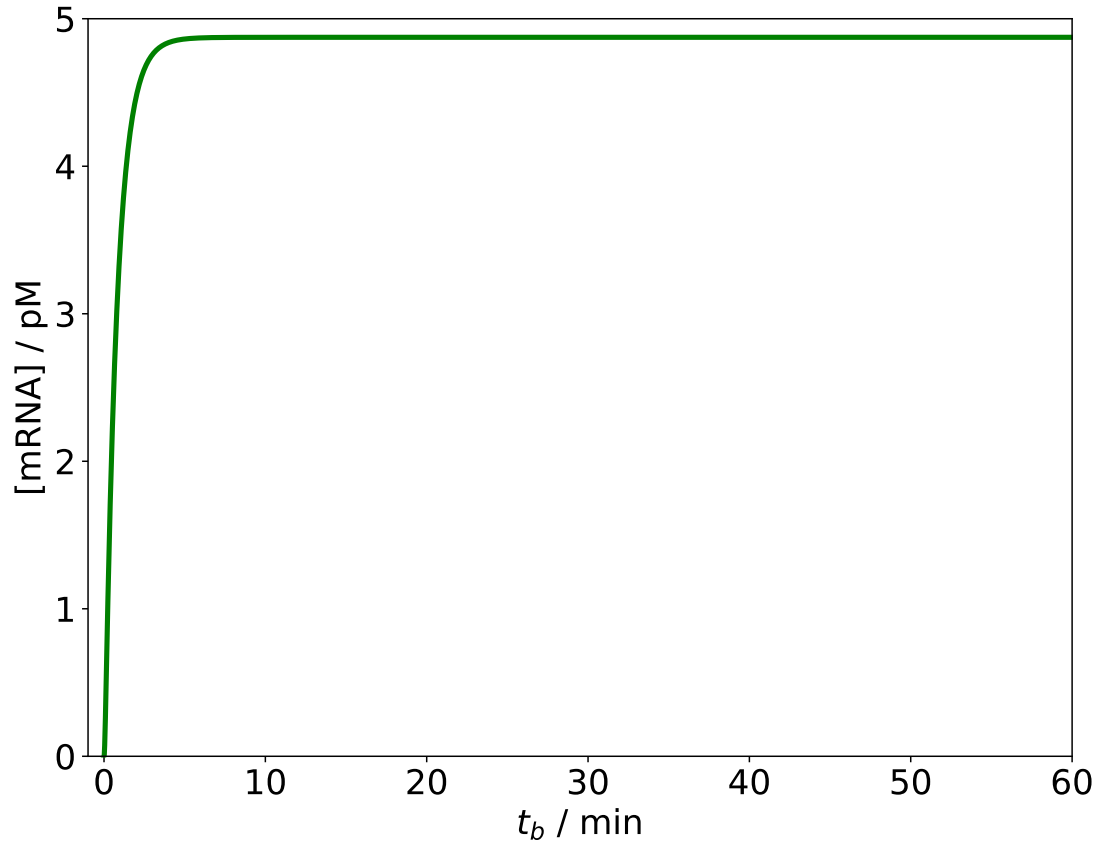
Kierzek, A. M., Zaim, J., & Zielenkiewicz, P. (2001). The effect of transcription and translation initiation frequencies on the stochastic fluctuations in prokaryotic gene expression. The Journal of Biological Chemistry, 276(11), 8165–8172. doi:10.1074/jbc.M006264200



RESULTS & DISCUSSION

RESULTS & DISCUSSION

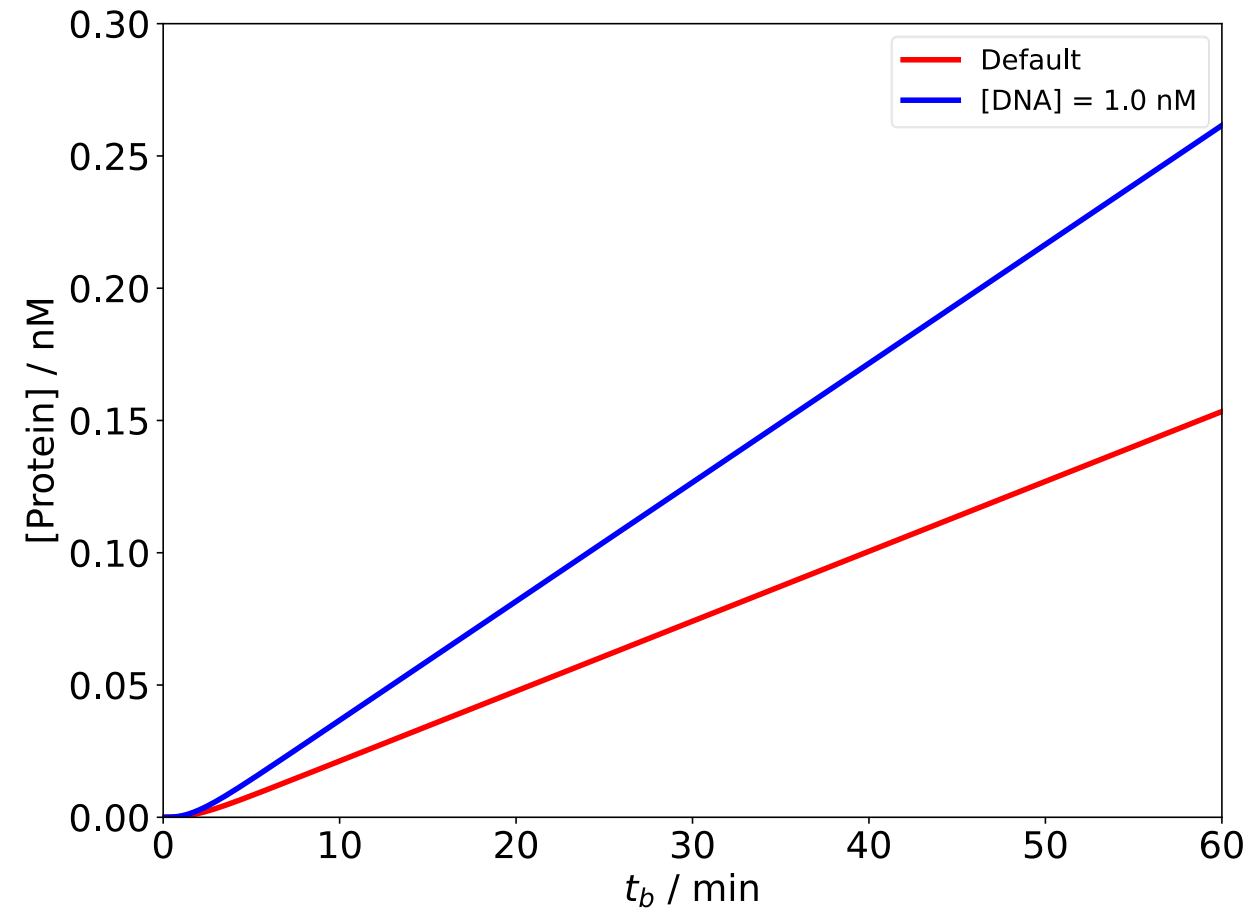
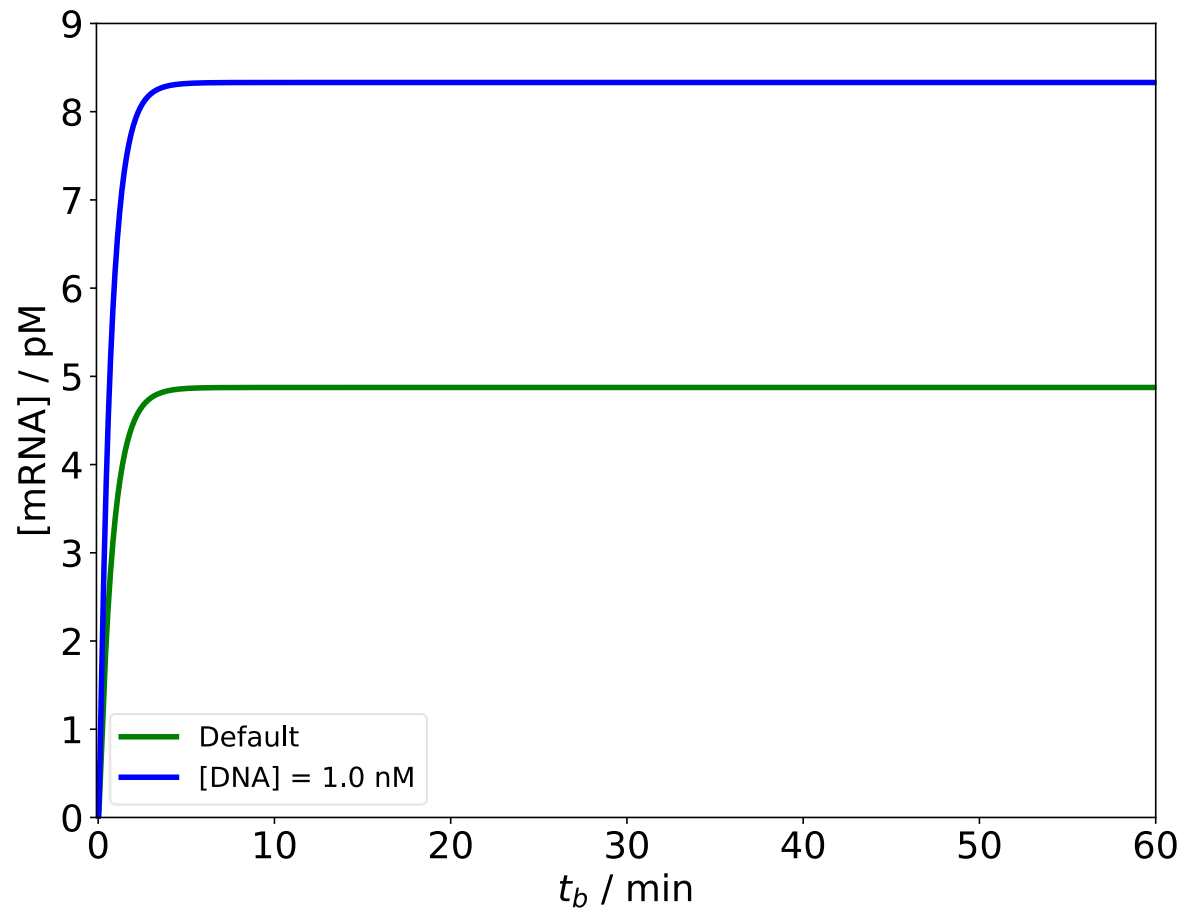
Simulation results (Default Settings)



- mRNA within the bacterial cell shows saturation behavior
- The protein production show a quasi-linear behavior

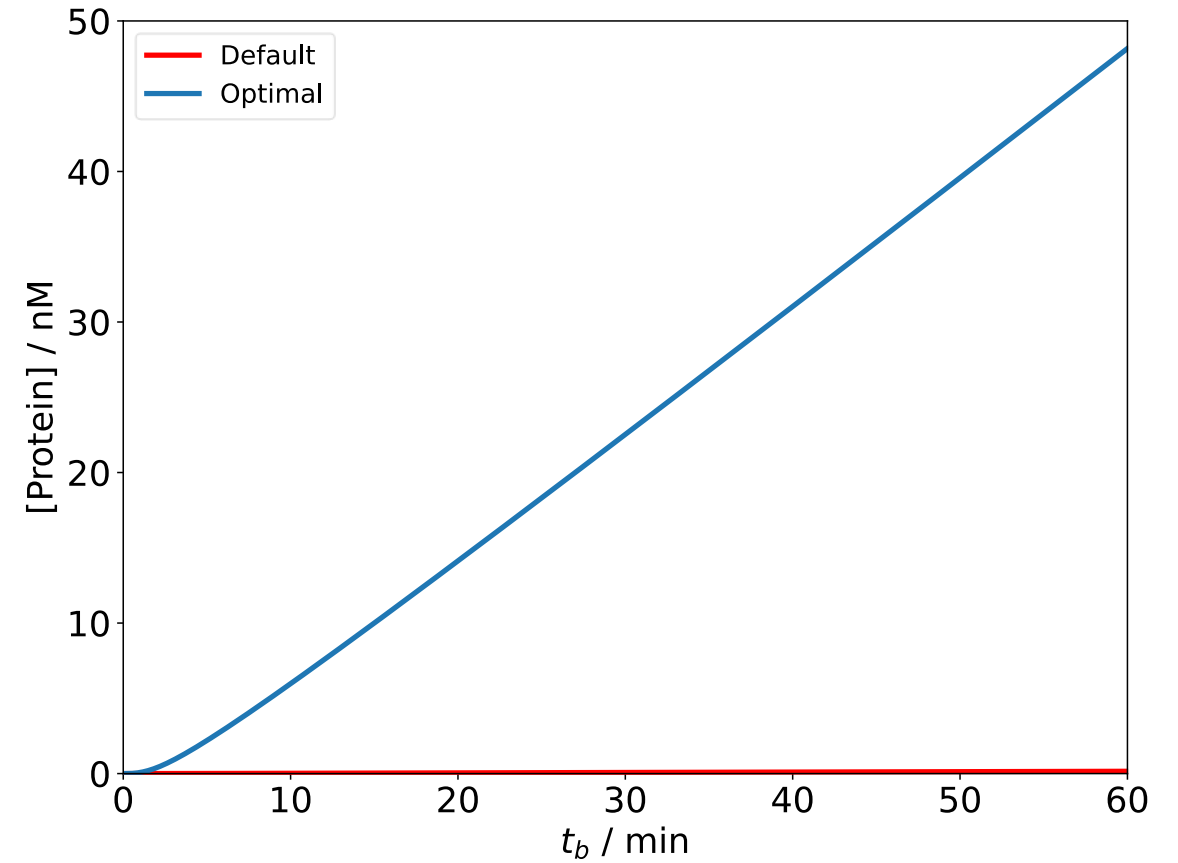
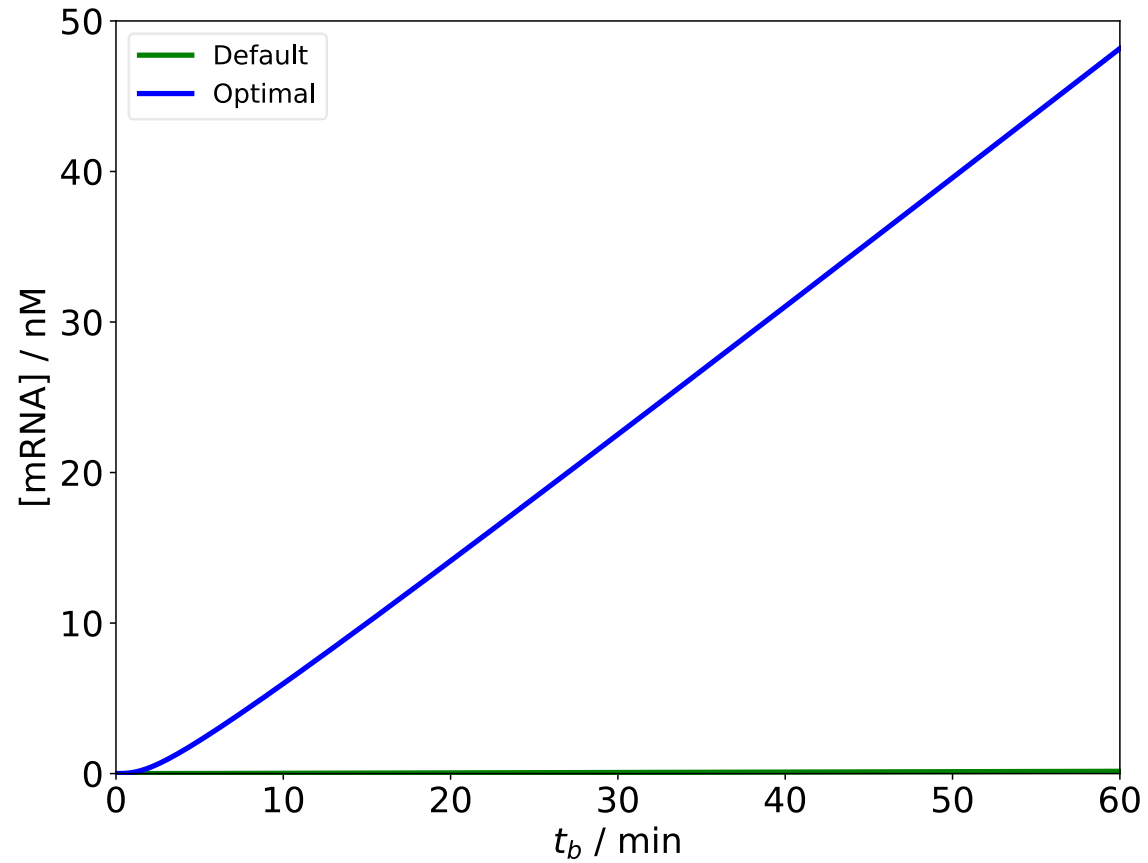
RESULTS & DISCUSSION

Simulation results (High copy number)



RESULTS & DISCUSSION

Simulation results (High copy number + no mRNA degradation + increased nutrient pool)



CONCLUSION

- **Positive modifications can improve the production yield significantly**
- **Choosing a better candidate/host is also a promising approach to improve yield**

THANK YOU