BIMM-143: INTRODUCTION TO BIOINFORMATICS

The find-a-gene project assignment https://bioboot.github.io/bimm143_S20/Dr. Barry Grant

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Questions:

[Q1] Tell me the name of a protein you are interested in. Include the species and the accession number. This can be a human protein or a protein from any other species as long as it's function is known.

If you do not have a favorite protein, select human RBP4 or KIF11. Do not use beta globin as this is in the worked example report that I provide you with online.

Name: Kinesin family member 11 (KIF11)

Accession: NP_004514

Species: Homo sapiens

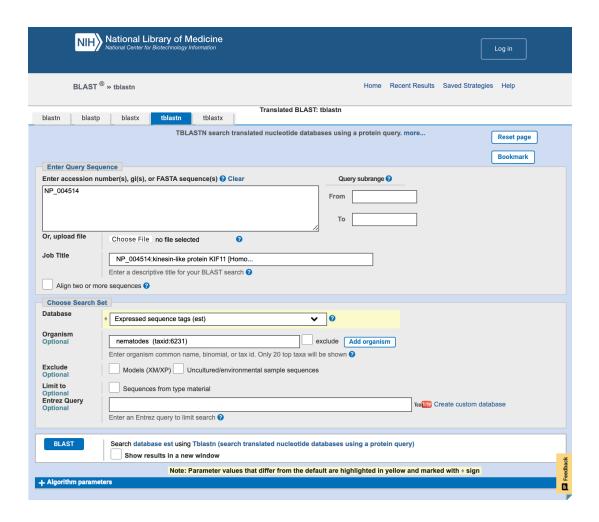
Function Known: Encoding a motor protein in which it belongs to the kinesin-like protein family. It is also known that it is involved in many kinds of the dynamics of spindle. Functions in chromosome positioning, centrosome separation and bipolar spindle establishment in mitosis of the cell.

[Q2] Perform a BLAST search against a DNA database, such as a database consisting of genomic DNA or ESTs. The BLAST server can be at NCBI or elsewhere. Include details of the BLAST method used, database searched and any limits applied (e.g. Organism).

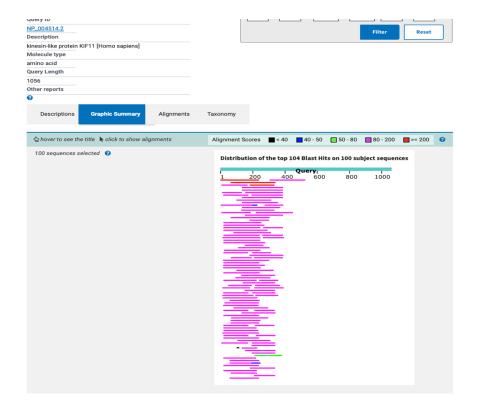
Method: TBLASTN (2.7.1) search against nematode ESTs

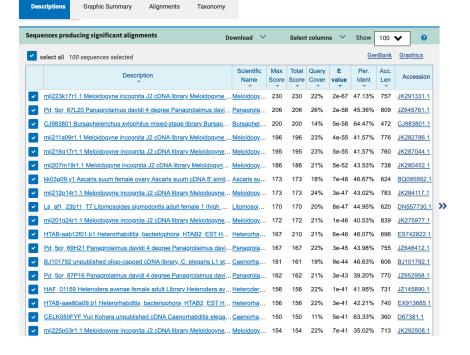
Database: Expressed Sequence Tags (est)

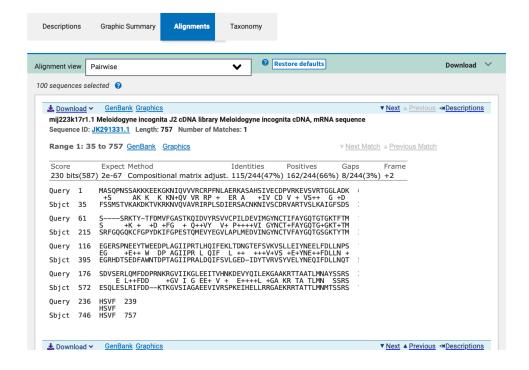
Organism: Nematodes (Taxid: 6231)



Chosen match: Accession JK291331.1, a 757 base pair clone from *Meloidogyne incognita*. See below for alignment details.







Alignment details:

```
>qb|JK291331.1| mij223k17r1.1 Meloidogyne incognita J2 cDNA library
Meloidogyne incognita
cDNA, mRNA sequence.
Length=757
 Score = 230 bits (587), Expect = 2e-67, Method: Compositional matrix
adjust.
 Identities = 115/244(47%, Positives = 162/244(66%, Gaps = 8/244(3%)
Frame = +2
Query 1
           MASQPNSSAKKKEEKGKNIQVVVRCRPFNLAERKASAHSIVECDPVRKEVSVRTGGLADK
                    AK K KN+QV VR RP + ER A
            +S
                                                 +IV CD V + VS++ G +D
Sbjct
            FSSMSTVKAKDKTVKRKNVQVAVRIRPLSDIERSACNKNIVSCDRVARTVSLKAIGFSDS
                                                                          214
            S----SRKTY-TFDMVFGASTKQIDVYRSVVCPILDEVIMGYNCTIFAYGQTGTGKTFTM
                                                                          115
       61
Ouerv
            S
                  +K + +D +FG + Q++VY V+ P++++VI GYNCT+FAYGQTG+GKT+TM
Sbjct
       215
           SRFGQGQKCFGPYDKIFGPESTQMEVYEGVLAPLMEDVINGYNCTVFAYGQTGSGKTYTM
                                                                          394
           EGERSPNEEYTWEEDPLAGIIPRTLHQIFEKLTDNGTEFSVKVSLLEIYNEELFDLLNPS
                                                                          175
Query
      116
                  +E++ W DP AGIIPR L QIF L ++ +++V+VS +E+YNE++FDLLN +
           EGRHDTSEDFAWNTDPTAGIIPRALDQIFSVLGED-IDYTVRVSYVELYNEQIFDLLNOT
Sbict
       395
                                                                          571
Query
       176
            SDVSERLQMFDDPRNKRGVIIKGLEEITVHNKDEVYQILEKGAAKRTTAATLMNAYSSRS
                                                                          235
                E L++FDD
                           +GV I G EE+ V + E+++L +GA KR TA TLMN SSRS
Sbjct
           ESQLESLRIFDD--KTKGVSIAGAEEVIVRSPKEIHELLRRGAEKRRTATTLMNMTSSRS
                                                                          745
       236
           HSVF 239
Query
            HSVF
           HSVF
Sbjct
      746
                 757
```

[Q3] Gather information about this "novel" <u>protein</u>. At a minimum, show me the protein sequence of the "novel" protein as displayed in your BLAST results from [Q2] as FASTA format (you can copy and paste the aligned sequence subject lines from your BLAST result page if necessary) or translate your novel DNA sequence using a tool called EMBOSS Transeq at the EBI. Don't forget to translate all six reading frames; the ORF (open reading frame) is likely to be the longest sequence without a stop codon. It may not start with a methionine if you don't have the complete coding region. Make sure the sequence you provide includes a header/subject line and is in traditional FASTA format.

Chosen sequence:

>M. incognita protein (sequence taken from BLAST result)

FSSMSTVKAKDKTVKRKNVQVAVRIRPLSDIERSACNKNIVSCDRVARTVSLKAIGFSDS SRFGQGQKCFGPYDKIFGPESTQMEVYEGVLAPLMEDVINGYNCTVFAYGQTGSGKTYTM EGRHDTSEDFAWNTDPTAGIIPRALDQIFSVLGEDIDYTVRVSYVELYNEQIFDLLNQT ESQLESLRIFDDKTKGVSIAGAEEVIVRSPKEIHELLRRGAEKRRTATTLMNMTSSRS HSVF

Here, tell me the name of the novel protein, and the species from which it derives. It is very unlikely (but still definitely possible) that you will find a novel gene from an organism such as *S. cerevisiae*, human or mouse, because those genomes have already been thoroughly annotated. It is more likely that you will discover a new gene in a genome that is currently being sequenced, such as bacteria or plants or protozoa.

Name: Meloidogyne incognita

Species: Meloidogyne incognita

Eukaryota; Metazoa; Ecdysozoa; Nematoda; Chromadorea; Rhabditida;

Tylenchina; Tylenchomorpha; Tylenchoidea; Meloidogynidae; Meloidogyninae; Meloidogyne; Meloidogyne incognita group.

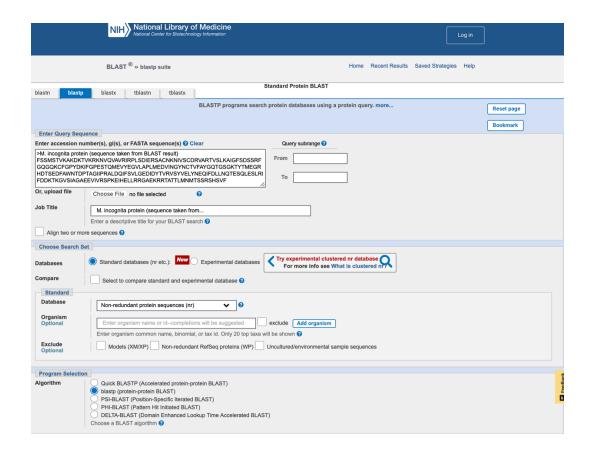
[Q4] Prove that this gene, and its corresponding protein, are novel. For the purposes of this project, "novel" is defined as follows. Take the protein sequence (your answer to [Q3]), and use it as a query in a blastp search of the nr database at NCBI.

- If there is a match with 100% amino acid identity to a protein in the database, from the same species, then your protein is NOT novel (even if the match is to a protein with a name such as "unknown"). Someone has already found and annotated this sequence, and assigned it an accession number.
- If the top match reported has less than 100% identity, then it is likely that your protein is novel, and you have succeeded.
- If there is a match with 100% identity, but to a different species than the one you started with, then you have likely succeeded in finding a novel gene.
- If there are no database matches to the original query from [Q1], this indicates that you have partially succeeded: yes, you may have found a new gene, but no, it is not actually homologous to the original query. You should probably start over.

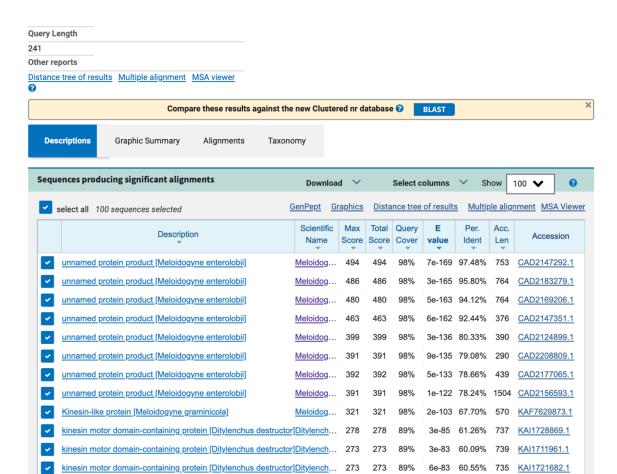
Details:

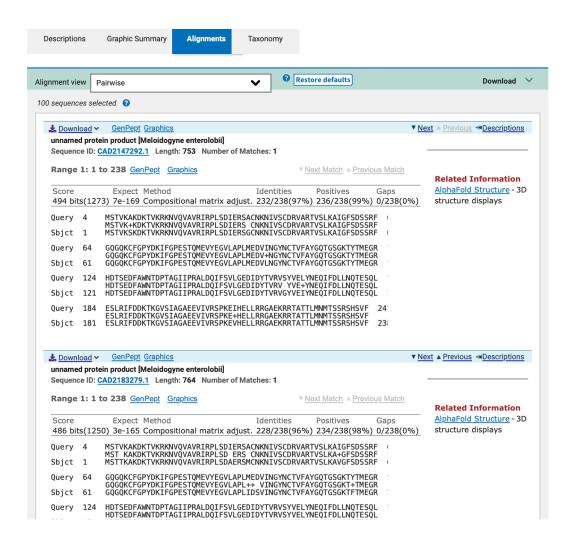
A BLASTP search against NR database (see setup in first screen-shot below) yielded a top hit result is to a protein from *Meloidogyne enterolobii* (nematodes).

See additional screen shots below for top hits and selected alignment details:



The top result is to a protein from *Meloidogyne enterolobii* (nematodes)see second screen shot below for alignment details:





[Q5] Generate a multiple sequence alignment with your novel protein, your original query protein, and a group of other members of this family from different species. A typical number of proteins to use in a multiple sequence alignment for this assignment purpose is a minimum of 5 and a maximum of 20 - although the exact number is up to you. Include the multiple sequence alignment in your report. Use Courier font with a size appropriate to fit page width.

Side-note: Indicate your sequence in the alignment by choosing an appropriate name for each sequence in the input unaligned sequence file (i.e. edit the sequence file so that the species, or short common, names (rather than accession numbers) display in the output alignment and in the subsequent answers below). The goal in this step is to create an interesting an alignment for building a phylogenetic tree that illustrates species divergence.

[Q6] Create a phylogenetic tree, using either a parsimony or distance-based approach. Bootstrapping and tree rooting are optional. Use "simple phylogeny" online from the EBI or any respected phylogeny program (such as MEGA, PAUP, or Phylip). Paste an image of your Cladogram or tree output in your report.

[Q7] Generate a sequence identity based **heatmap** of your aligned sequences using R. If necessary convert your sequence alignment to the ubiquitous FASTA format (Seaview can read in clustal format and "Save as" FASTA format for example). Read this FASTA format alignment into R with the help of functions in the **Bio3D package**. Calculate a sequence identity matrix (again using a function within the Bio3D package). Then generate a heatmap plot and add to your report. Do make sure your labels are visible and not cut at the figure margins.

[Q8] Using R/Bio3D (or an online blast server if you prefer), search the main protein structure database for the most similar atomic resolution structures to your aligned sequences.

List the top 3 *unique* hits (i.e. not hits representing different chains from the same structure) along with their Evalue and sequence identity to your query. Please also add annotation details of these structures. For example include the annotation terms PDB identifier (structureId), Method used to solve the structure (experimentalTechnique), resolution (resolution), and source organism (source).

HINT: You can use a single sequence from your alignment or generate a consensus sequence from your alignment using the Bio3D function consensus(). The Bio3D functions blast.pdb(), plot.blast() and pdb.annotate() are likely to be of most relevance for completing this task. Note that the results of blast.pdb() contain the hits PDB identifier (or pdb.id) as well as Evalue and identity. The results of pdb.annotate() contain the other annotation terms noted above.

Note that if your consensus sequence has lots of gap positions then it will be better to use an original sequence from the alignment for your search of the PDB. In this case you could chose the sequence with the highest identity to all others in your alignment by calculating the row-wise maximum from your sequence identity matrix.

[Q9] Generate a molecular figure of one of your identified PDB structures using the NGL viewer online (or VMD/PyMol). You can optionally highlight conserved residues that are

likely to be functional. Please use a white or transparent background for your figure (i.e. not the default black).

Based on sequence similarity. How likely is this structure to be similar to your "novel" protein?

[Q10] Perform a "Target" search of ChEMBEL (https://www.ebi.ac.uk/chembl/) with your novel sequence. Are there any **Target Associated Assays** and **ligand efficiency data** reported that may be useful starting points for exploring potential inhibition of your novel protein?

Scoring Rubric:

[45 total points available]

Q1 (4 points) Protein name	1
	' 1
Species	•
Accession number	1
Function known	1
Q2 (6 points)	
Blast method	1
Database searched	1
Limits applied	1
Search output list (top hits)	1
Alignment of choice	1
Evalue and other alignment stats	1
Q3 (3 points)	
Protein sequence of choice matches Subject above	1
Name in header	1

Species	1
Q4 (3 point)	
Blastp output list with identities & Evalue	1
Top alignment shown with alignment statistics	1
Results indicates a "novel" gene found	1
Q5 (3 points)	
MSA labeled with useful names	1
MSA trimmed appropriately (i.e. no gap overhangs)	1
Pasted MSA fits report page width (i.e. font, format)	1
Q6 (1 point)	
Figure illustrates sequence clustering pattern	1
Q7 (10 points)	
Heatmap figure included in report	5
Heatmap is legible (i.e. no labels obscured)	5
Q8 (10 points)	
PDB identifiers from multiple species reported	5
Annotation of PDB source, resolution and technique 4 Annotation of Evalue and Sequence Identity	1
Q9 (4 points)	
Structure figure provided	2
Uses white background for molecular figure	1
Figure of high resolution (i.e. not just snapshot)	1
Q10 (1 point)	
Evidence of ChEMBEL searches	1