



Project Proposal: Process Control for OCT-In-The-Loop Bioprinting

Zane Bates, Department of Biomedical Engineering, CMU

2025-08-29

Department of Biomedical Engineering
Carnegie Mellon University

Contents

Contents	2
1 Project Proposal: Process Control for OCT-In-The-Loop Bioprinting	3
1.1 Objective	3
1.2 Background & Prior Work	3
1.3 Specific Aims	3
1.4 Approach	4
1.4.1 OCT-in-the-loop control	4
1.5 Risks & Mitigations	4
1.6 Timeline & Milestones	4
1.7 Questions for Dr. Feinberg	5
1.8 Future Investigations	6
1.9 Other Projects of High Interest	6
References	7

1 Project Proposal: Process Control for OCT-In-The-Loop Bio-printing

TL;DR. Investigating process innovations in extrusion bioprinting, with emphasis on closed-loop control using OCT imaging to dynamically tune construct mechanics and fidelity.

1.1 Objective

Develop and demonstrate a **closed-loop control system** that uses OCT feedback to actively tune the crosslinking, degradation, or chemical stiffening of FRESH/volumetric bioprinted constructs during print.

1.2 Background & Prior Work

- Process innovations are highlighted in [1] under *2.2 Engineering advances* and *2.3 Process Control and improvements*.
- Embedded OCT has been shown for monitoring in FRESH [2], but no fully integrated feedback-control loop yet.
- Photodegradable hydrogels (embedded nitrobenzyl ether into the hydrogel) can dynamically soften under specific light exposure [3].
- Embedded Extrusion-Volumetric Printing (EmVP) demonstrates multi-material constructs using photo-crosslinkable micro-resins, GelMA-based μ Resins [4].
- Microfluidics approaches using EDC/NHS as a cross-linker to increase stiffness (highly aligned fibrillar structure) [6]. However, the exposure time will need to be flushed out as literature include times as between 0.5hr to 48hr to achieve optimal results and crosslinking [7]. Cytotoxicity isn't a concern as long as standard protocols are follows [8]. Additionally, a correct ratio will likely need to be experimented for with FRESH, but other applications found a optimum NHS/EDC molar ratio was 0.5 [8].

1.3 Specific Aims

Aim 1. Develop computer vision ML algorithm to determine interventions.

Aim 2. Integrate OCT into the print path for real-time error detection.

Aim 3. Demonstrate corrections using light (crosslink/degrade) or microfluidics (EDC/NHS).

1.4 Approach

1.4.1 OCT-in-the-loop control

- OCT captures **layer height, pore geometry, void space**.
- Controller: PID/MPC based on OCT → mechanics calibration curve or CV algorithm → informed tuning of control knobs.
- Control knobs: light dose (405 nm ↑ stiffness), degrade dose (365 nm → soften), EDC/NHS microfluidics-assisted delivery (↑ stiffness), extrusion parameters.

1.5 Risks & Mitigations

- **Nitrobenzyl polymerization toxicity** → cytocompatibility of PEG monoacrylate and free radical/ROS generation [3]
- **Spectral crosstalk** → orthogonality tests with 405 nm 1 W diode laser for EmVP and 365 nm diode laser for photodegradable nitrobenzyl polymer [3]. For reference, at 10 mW/cm², nitrobenzyl hydrogels fully degrade in ~10 min at 365 nm ($t = 280$ s), in ~5 min at 20 mW/cm² ($t = 140$ s), but takes ~15–16 min at 405 nm ($t = 930$ s), meaning 405 nm works but is ~3–4× slower and less efficient than 365 nm [3].
- **Cell viability under near-UV/UV** → antioxidant additives to combat free radicals and ROS from photopolymerize or photodegrade, dose windows
- **EDC/NHS chemical crosslinking time** → exposure time for EDC/NHS, HEPES buffer solution to achieve desirable properties, optimal EDC/NHS molar ratio
- **OCT computer vision explainability** → SNR, false negatives + positives, print path adjustments

1.6 Timeline & Milestones

Quarter	Deliverables
Q1	Spectral/dose orthogonality test + exposure times for EDC/NHS chemical crosslinker
Q2	OCT–mechanics calibration + model fit
Q3	OCT-induced print planning edits

Quarter	Deliverables
Q4	FRESH with NB- μ Resin, volumetric editing with OCT feedback

1.7 Questions for Dr. Feinberg

1. Do you have competitors in extrusion bioprinting?

It seems like people use extrusion-based bioprinting and FRESH bioprinting interchangeably.

2. Does the needle geometry need to be tuned? Or is that a solved problem?

I read that the 6-DOF helps with reducing the damage done by needles running through the construct.

3. Is there a need for post-processing bioreactors

In the talks in this paper [1] about an integrated bioreactor setup, my understanding is that there would be an automated cell culture system that would then be sent into a bioink.

4. How is printing resolution lower than the bidirectional error of the stepper motor in this paper [9]?

I read *Development of a high-performance open-source 3D bioprinter* [9] and I was wondering about the printing resolution. When the accuracy of the stepper motor was assessed, it was found that the bidirectional error was anywhere from 20 μ m to 50 μ m depending on the axis. However, then later it is said that the bioink printing resolution is approaching 20 μ m.

5. Can you print the support bath in a nonplanar setup?

In this paper [2]:

To date, FRESH and other embedded 3D bioprinting have used pre-filled containers of support material, whether the gelatin microparticle support bath or some other support bath such as alginate microparticles. However, for prints larger than \sim 5 mm tall the deflection of the fine 34-gauge, 6.35 mm long needle tips limits resolution. We have previously overcome this by building custom needle tips with long, larger diameter and rigid tips terminated in a smaller diameter tip. However, printing into deep dishes of gelatin microparticle support bath

presents other challenges such as dehydration and skinning of the upper layer of the gelatin microparticle support bath. Here we developed a new alternative approach by printing the gelatin microparticle support bath itself in order to minimize the deflection of the needle tip and without a limitation on construct height.

1.8 Future Investigations

- Investigate **2P degradation control** (740–800 nm) for spatially selective softening.
- Feed mapping into **PID/MPC** controller for automated correction.

1.9 Other Projects of High Interest

1. Closed-loop bioink rheology system

- Suggested by Annie, though Charlie noted shear damage may be less critical.
- ML/AI algorithms could be applied to rheology datasets [1], but challenges remain in generating sufficient sample sizes for biological contexts.
- Promising precedents exist in direct ink writing, where ML models have been trained to optimize ink rheology [10].

2. Print planning software

- Current nonplanar solutions are mostly custom and lack universality [1].
- There is a strong need for “automated trajectory generation from medical imaging data, including fiber orientation information [1]” to streamline planning and execution.

References

- [1] A. S. Bakirci E Asghari Adib A, “Advancing extrusion-based embedded 3D bioprinting via scientific, engineering, and process innovations,” *Biofabrication*, 2025, doi: [10.1088/1758-5090/adb7c3](https://doi.org/10.1088/1758-5090/adb7c3).
- [2] C. B. Tashman JW Shiawski DJ, “In situ volumetric imaging and analysis of FRESH 3D bioprinted constructs using optical coherence tomography,” *Biofabrication*, 2022, doi: [10.1088/1758-5090/ac975e](https://doi.org/10.1088/1758-5090/ac975e).
- [3] S. C. Kloxin AM Kasko AM, “Photodegradable hydrogels for dynamic tuning of physical and chemical properties,” *Science*, 2009, doi: [10.1126/science.1169494](https://doi.org/10.1126/science.1169494).
- [4] S. F. D. Ribezzi M. Gueye, “Shaping synthetic multicellular and complex multimaterial tissues via embedded extrusion-volumetric printing of microgels,” *Adv. Mater.*, 2023, doi: [10.1002/adma.202301673](https://doi.org/10.1002/adma.202301673).
- [5] G. S. Shepherd DV Shepherd JH, “The process of EDC-NHS cross-linking of reconstituted collagen fibres increases collagen fibrillar order and alignment,” *APL Mater.*, 2015, doi: [10.1063/1.4900887](https://doi.org/10.1063/1.4900887).
- [6] F. M. Omobono MA Zhao X, “Enhancing the stiffness of collagen hydrogels for delivery of encapsulated chondrocytes to articular lesions for cartilage regeneration,” *J Biomed Mater Res Part A*, 2015, doi: [10.1002/jbm.a.35266](https://doi.org/10.1002/jbm.a.35266).
- [7] A. C. Alavarce, E. C. G. Frachini, R. L. C. G. da Silva, V. H. Lima, A. Shavandi, and D. F. S. Petri, “Crosslinkers for polysaccharides and proteins: Synthesis conditions, mechanisms, and crosslinking efficiency, a review,” *International Journal of Biological Macromolecules*, vol. 202, pp. 558–596, 2022, doi: [10.1016/j.ijbiomac.2022.01.029](https://doi.org/10.1016/j.ijbiomac.2022.01.029).
- [8] J.-Y. Lai, “Corneal stromal cell growth on gelatin/chondroitin sulfate scaffolds modified at different NHS/EDC molar ratios,” *Int. J. Mol. Sci.*, 2013, doi: [10.3390/ijms14012036](https://doi.org/10.3390/ijms14012036).
- [9] S. Tashman J. W., “Development of a high-performance open-source 3D bioprinter,” *Sci Rep*, 2022, doi: [10.1038/s41598-022-26809-4](https://doi.org/10.1038/s41598-022-26809-4).
- [10] R. Weeks R. D. and J. O. Hardin, “In-situ rheology measurements via machine-learning enhanced direct-ink-writing,” *Adv. Intell. Syst.*, 2025, doi: [10.1002/aisy.202400293](https://doi.org/10.1002/aisy.202400293).