

Perspective

# Management Pathways for Fragmented Populations: From Habitat Restoration to Genetic Intervention

Magdalene N. Ngeve <sup>1</sup>, Kyle E. Rufo <sup>2</sup> and Zachery D. Zbinden <sup>2,\*</sup>

<sup>1</sup> Horn Point Laboratory, University of Maryland Center for Environmental Science, Cambridge, MD 21613, USA; mngeve@umces.edu

<sup>2</sup> Appalachian Laboratory, University of Maryland Center for Environmental Science, Frostburg, MD 21532, USA; kyle.rufo@umces.edu

\* Correspondence: zach.zbinden@umces.edu

## Abstract

Habitat fragmentation is reshaping ecosystems worldwide, reducing connectivity, eroding genetic diversity, and limiting species' capacity to adapt to rapid environmental change. Conservation management responses to fragmentation generally follow three pathways: restoring habitats to rebuild connectivity, translocating individuals to bolster declining populations, and, more recently, directly managing adaptive genetic variation. We synthesize the ecological and genetic consequences of fragmentation and evaluate these management pathways along a continuum from landscape-scale interventions to genome-level strategies. Habitat restoration can reconnect patches and improve demographic stability, but its genetic outcomes remain uncertain without baseline and post-restoration monitoring. Translocation offers a more immediate means of restoring gene flow but introduces demographic risks, potential impacts on source populations, and uncertainties in establishment and long-term fitness. Emerging genomic technologies now support a third approach: Targeted Genetic Intervention (TGI), which aims to accelerate the spread of beneficial genetic variants or enhance adaptive potential directly. Although promising, TGI faces significant challenges, including polygenic trait architecture, risks to genome-wide diversity, and the need for robust ethical and governance frameworks. Across all pathways, genetic data are essential for prioritizing actions, diagnosing vulnerable species and populations, and restoring the evolutionary potential necessary for long-term persistence in increasingly fragmented landscapes.

**Keywords:** habitat fragmentation; genetic diversity; gene flow; translocation; restoration; genetic rescue; conservation genomics; targeted genetic intervention



Academic Editor: Mary M. Peacock

Received: 19 December 2025

Revised: 19 January 2026

Accepted: 27 January 2026

Published: 29 January 2026

**Copyright:** © 2026 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the [Creative Commons Attribution \(CC BY\) license](#).

## 1. Overview

Habitat alteration, including destruction and fragmentation, is now one of the dominant forces shaping global biodiversity [1]. It is widely recognized as a leading driver of species loss [2] and has contributed to declines of up to 75% in some regions [3]. Human activities have altered nearly 80% of terrestrial environments [4], 90% of oceans [5], and 50% of rivers [6].

Herein, habitat fragmentation refers to the division of once continuous habitat, not the total loss of habitat, although fragmentation can contribute to habitat loss. While the ecological consequences of fragmentation, including reduced habitat area, increased edge effects, and disrupted organismal movement, are well established [7,8], its genetic and evolutionary impacts remain inconsistently acknowledged in conservation planning [9–11].

Genetic diversity underpins a population's capacity to persist and adapt over time. Fragmentation restricts movement among populations [12], reducing gene flow and effective population sizes. In turn, this elevates inbreeding, genetic drift [13], and the stochastic loss of genetic variation [14]. Even populations that appear demographically stable may experience hidden erosion of their adaptive potential, increasing the risk of extirpation under environmental change [15]. Fragmentation can also prevent the spread of beneficial genetic variation, limiting isolated populations' ability to respond to changing environments.

Because fragmentation is largely human-driven, conservation management interventions seek to counter its ecological, demographic, and genetic consequences. We outline three primary pathways through which managers can maintain or restore genetic diversity in fragmented systems below (Table 1).

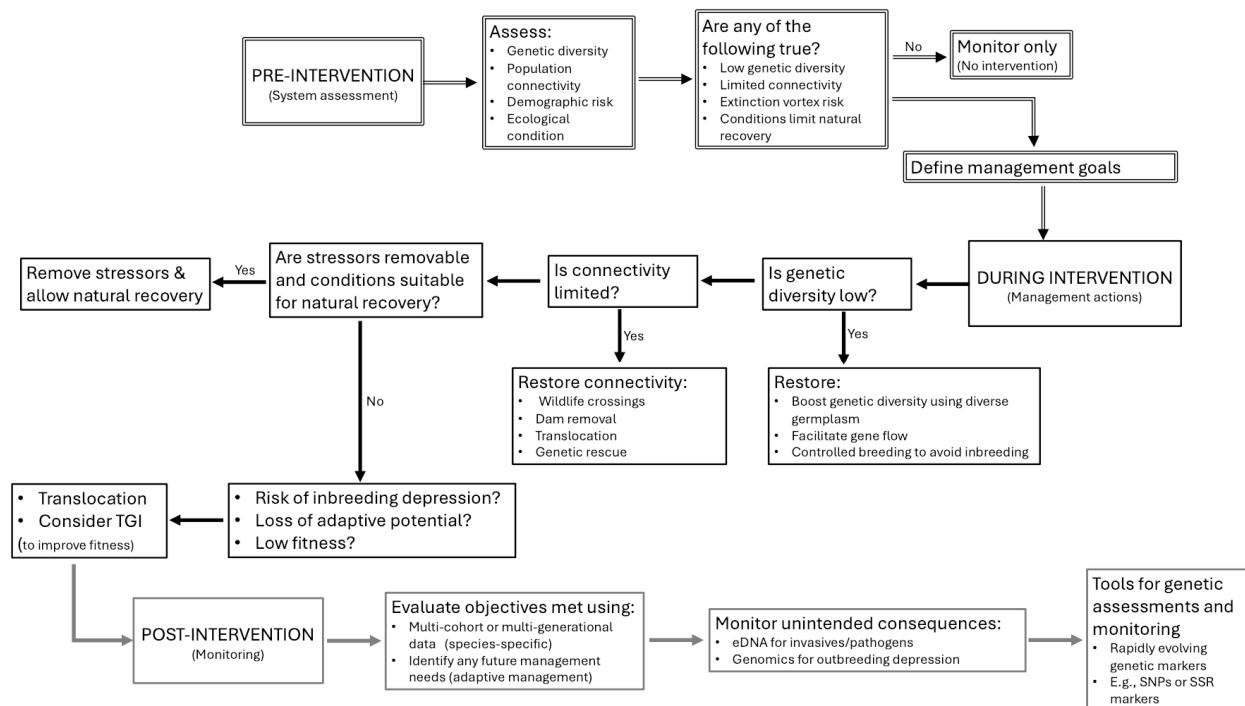
**Table 1.** Highlighting the main goals, challenges, and risks of the three management pathways for fragmented habitats. The essential genetic information required for each is indicated at the bottom of each column.

Feature	Habitat Restoration	Translocation	Targeted Genetic Intervention (TGI)
<b>Primary Goal</b>	Re-establish ecological connectivity to facilitate natural demographic recovery.	Deliberate movement of organisms to facilitate gene flow, provide demographic support, and genetic rescue of imperiled populations.	Directly alter allele frequencies through synthetic biology to increase fitness against specific threats or restore lost diversity.
<b>Spatial and temporal Scales</b>	Landscape level, long-term (decades to centuries).	Population level, medium-term (years to decades).	Individual/gene level, potentially rapid intergenerational change.
<b>Relative Cost</b>	Very high (land acquisition, earthworks, long-term maintenance).	Moderate to high (capture, transport, monitoring, disease screening).	High initial R&D, potentially lower long-term cost if self-sustaining.
<b>Key Feasibility Challenges</b>	Land ownership, political will, conflicts of interest, scale mismatch due to fragmentation, and a long time lag for genetic effects.	Finding suitable and sufficient source populations, logistical complexity of capture/transport.	Lack of genomic resources for non-model species, technical difficulty, regulatory hurdles, and public acceptance.
<b>Primary Risks</b>	Ineffective if populations are already genetically depauperate; may facilitate the spread of invasive species.	Outbreeding depression, disease transmission, genetic swamping, and demographic impact on the source population.	Off-target effects, unintended ecological consequences (pleiotropy), escape of modified genes, and ethical concerns.
<b>Essential Genetic Information</b>	Landscape genomics to prioritize corridors; pre- and post-monitoring of genetic diversity and connectivity.	Genomic assessment of divergence and local adaptation (GEA) to mitigate outbreeding depression risk.	Whole-genome sequencing, identification of adaptive loci (GEA), functional validation, and off-target analysis.

- I. Habitat restoration: rebuilding ecological connectivity to allow natural movement and gene flow to resume.
- II. Translocation: moving individuals to support demographic stability and bolster genetic variation.
- III. Targeted Genetic Intervention: directly facilitating the spread of genetic variants, including through emerging synthetic biology tools.

These strategies are not mutually exclusive and may be most effective when used in combination. In Figure 1, we present a concise decision tree to guide practitioners on

which strategies should be considered; see a complementary practitioner guide based on the Restoration Gene Pool concept for sourcing plant material [16]. In the sections that follow, we examine habitat fragmentation, its effects on genetic diversity and evolutionary potential, and highlight each management pathway in detail, emphasizing how explicit consideration of genetic diversity can improve restoration outcomes, guide translocation and genetic rescue efforts, and inform the emerging role of targeted genetic intervention in conservation management.



**Figure 1.** Concise decision tree for determining what management strategies may be considered by practitioners. Double-lined arrows connect steps to consider prior to any management intervention; simple solid black arrows connect steps to consider during the management phase; while simple solid gray-lined arrows connect steps to consider for monitoring, after management interventions are complete.

## 2. Habitat Fragmentation

Habitats in most ecosystems are naturally fragmented to some degree. This “patchiness” arises from spatial heterogeneity in resources, abiotic conditions, and species-specific niche requirements. Natural fragmentation structures biodiversity and promotes species coexistence [17]. Major landscape features such as mountains and rivers restrict dispersal and contribute to lineage diversification [18–20]. At the land–sea interface, shoreline geomorphology, coastal dynamics, and ocean currents generate natural patchiness in organisms such as mangroves [21–23]. Submerged aquatic vegetation similarly forms shifting mosaics of patches influenced by local environmental conditions in coastal and estuarine systems [24,25]. In riverine networks, hierarchical drainage structures naturally isolate fish populations across catchments [26,27]. In marine environments, biogeochemical provinces, nutrient dynamics, and other physical and biological processes similarly determine natural species patchiness and patch size [28–30].

In contrast, anthropogenic habitat fragmentation results from human activities that subdivide formerly continuous habitats into smaller, isolated patches. This process is typically more rapid and severe than natural fragmentation, driving habitat loss, degradation, and separation. In aquatic systems, structures such as weirs and dams disrupt

connectivity [27,31], while bridge construction and culverts can also impede dispersal [32]. Although less conspicuous resistance created by chemical pollution can create “soft” barriers that also reduce habitat continuity [33]. Terrestrial landscapes are further subdivided through deforestation, agriculture, urban expansion, roads, and railways. All species experience the effects of fragmentation, but those most at risk include short-range endemics [34], species with limited dispersal ability [35], and taxa already listed as critically endangered by the IUCN [36]. Across ecosystems, fragmentation affects biodiversity at multiple spatial scales, shaping ecological and evolutionary processes both within and among habitat patches.

### 2.1. Within-Patch Impacts of Habitat Fragmentation

By breaking large, contiguous habitats into smaller, isolated patches, fragmentation reduces the overall extent of hospitable space within a landscape. This loss and rearrangement of usable habitat lowers population carrying capacity by limiting resources, restricting movement, and forcing patches to function in isolation. The resulting within-patch effects consistently depress local population sizes, often with cascading consequences. For example, shrinking mangrove patches greatly diminish global carbon sequestration capacity [37], with millions of tons of carbon released annually as mangroves are destroyed. Fragmentation of dense aquatic vegetation beds leads to smaller, more structurally complex patches during recovery [38], and these changes occur alongside increasingly frequent extreme weather events that can eliminate entire beds [39–41].

Smaller, isolated patches also support fewer species interactions due to reduced resource availability and species diversity [42–44]. Because habitat size is strongly correlated with species richness, fragmentation-driven reductions in patch size can trigger “ecosystem decay” or the progressive decline in ecosystem health and the continued loss of species in small, isolated habitats [45]; although other mechanisms, such as distance-decay, environmental heterogeneity, and sampling effort, are also associated in fragmented landscapes [46]. In such populations, mate limitation becomes common, reducing sexual reproduction [47–49]. This isolation may favor selfing and increase inbreeding [50], reducing overall fitness and survival. Combined with limited resources and fewer species interactions, these demographic and ecological pressures increase the risk of extirpation [51,52]. Without intervention, these within-patch challenges often worsen or persist [35,53], with the smallest and most isolated fragments typically experiencing the most severe effects [3].

Beyond reducing patch size, fragmentation intensifies edge effects, thereby degrading resource availability and altering microclimates [53]. Such shifts can influence population processes (e.g., phenology) and community interactions [48,49,54], driving changes in selective pressures and compromising patch-level fitness [55]. Notably, approximately 70% of the world’s remaining forests lie within 1 km of an edge [3]. Although edge severity varies with forest age, affected forests tend to be drier, warmer, windier, more light-exposed, and more disturbance-prone [56]. These conditions contribute to an estimated 9% global loss of above-ground forest biomass due to edge effects [53]. In other systems, such as seagrass beds, edges show reduced shoot densities but elevated densities of invertebrates and benthic fishes [57], illustrating how edge effects can differentially influence community members.

### 2.2. Among Patch Impacts of Habitat Fragmentation

Habitat permeability and connectivity are central drivers of landscape-scale community dynamics. High permeability across space promotes functional diversity, facilitates species interactions, and supports long-term population persistence [58]. Fragmentation reduces this permeability by degrading habitat quality [33,59], increasing

movement costs [33,44], and altering individual fitness [60]. In plants, fragmentation disrupts pollination by altering wind regimes, reducing pollinator abundance, and weakening plant–pollinator interactions [49,61,62]. Seed dispersal is also restricted by fragmentation, reducing effective dispersal among patches [43]. These constraints reshape natural metapopulation dynamics as unsuitable habitat increasingly surrounds and isolates smaller suitable patches [35]. The surrounding habitat matrix, including agricultural land, roads, and urban development, can impede dispersal [63].

In aquatic systems, fragmentation caused by dams has variable impacts depending on species' dispersal abilities [64]. Yet, the consequences are broadly consistent: reduced habitat quality, lower reproductive success [65,66], and increased population isolation [27,36,66,67]. Altered movement pathways can also drive turnover in community composition, as observed in stream fish assemblages [68].

Although both natural and anthropogenic processes can isolate populations, anthropogenic fragmentation generally produces stronger and faster population divergence [69]. These heightened effects stem from the rapid pace of habitat alteration, the abrupt formation of barriers, and the broad spatial scale of human impacts. Anthropogenic fragmentation also imposes novel selection pressures [70], yet many species cannot adapt quickly enough to keep pace with these changes. Even when adaptive responses arise, they may exacerbate rather than mitigate the impacts of fragmentation because evolutionary adjustments rarely compensate for the magnitude of environmental change associated with human-driven habitat alteration [70].

### 3. Genetic Diversity and Evolutionary Potential

Gene flow, mediated by the movement of individuals, spores, or gametes among populations, is fundamental to maintaining genetic diversity [71,72]. Habitat fragmentation restricts dispersal and gene flow, isolating patches and creating small, evolutionarily independent populations [14]. These patches effectively function as habitat “islands” surrounded by a less favorable matrix of habitat [73]. When populations remain well connected, gene flow provides substantial genetic benefits within a metapopulation [74–76]. Gene flow counters the random fixation of alleles due to genetic drift [77,78] and increases genetic variation by replenishing rare alleles that might otherwise be lost [79]. In fragmented and isolated populations, these advantages diminish as dispersal and gene flow decline [80].

Isolation of mates and changes in habitat quality associated with fragmentation may alter fitness by lowering fecundity in birds [3] and by shifting reproductive strategy toward asexual reproduction in plants [70], thereby reducing seed production. Lower fecundity negatively affects populations by decreasing population size and the number of effective breeders, and by reducing total allelic diversity within the gene pool [81,82]. As inbreeding increases, deleterious recessive alleles become homozygous and accumulate [83,84]. Because drift is stronger in smaller populations, both census size ( $N_c$ ) and effective population size ( $N_e$ ) decline [85]. Long-term persistence typically requires  $N_e \geq 500$  to retain adaptive potential, while  $N_e \geq 50$  may suffice only in the short term [82]. Yet many disturbed or isolated populations fall well below these thresholds and suffer a disproportionately high stochastic loss of genetic variation each generation [34,41,86]. Low-diversity, inbred populations with fixed deleterious alleles exhibit reduced fitness and diminished resilience to environmental change [80,87]. In extreme cases, mate limitation can exacerbate inbreeding in self-compatible species or prevent reproduction entirely in dioecious species [88,89]. With fewer individuals, net mutation rates effectively decline, and new genetic variants arise more slowly than they are lost [90,91]. The Florida panther (*Puma concolor coryi*) illustrates these consequences: habitat loss isolated the population, driving

declines in genetic diversity and increases in inbreeding, resulting in reduced fitness and harmful phenotypes [92].

Together, the reduced gene flow, elevated drift, and increased inbreeding caused by fragmentation increase genetic divergence among patches while eroding standing genetic variation—the substrate on which natural selection acts [93,94]. Without gene flow, adaptive alleles cannot spread among populations, and genetic drift accelerates the loss of variation [87,95]. Populations subject to drift-driven allele loss (i.e., small isolated populations) may be less resistant or resilient to disease, environmental change, and other stressors [14,96,97]. Fragmentation also unfolds alongside other anthropogenic pressures and increasing extreme weather events [98,99], which can cause additional genetic bottlenecks and further reduce the diversity retained by surviving individuals [100]. Small, genetically depauperate populations respond less robustly to stochastic and environmental challenges than larger, more variable populations [101]. Collectively, these processes diminish evolutionary potential and the likelihood of population persistence.

#### 4. Habitat Restoration

Accelerated extinction rates in increasingly patchy habitats call for urgent efforts to mitigate landscape degradation and other human-driven pressures on biodiversity [102]. To counter fragmentation, managers have implemented a range of connectivity-focused interventions. Habitat corridors link isolated terrestrial patches [103], while dam removal and fish bypass structures restore movement pathways in river systems [67,104]. Additionally, constructing artificial habitats or refuge pools has been proposed to maintain in-stream connectivity during drought and reduce fragmentation impacts [65,105].

Most interventions occur at local scales, but ambitious large-scale restoration targets have been established across ecosystems. Examples include global initiatives for mangrove recovery [106], submerged aquatic vegetation [107–109], fish population restoration [110], and terrestrial reforestation efforts [111]. Many of these programs have been designated as United Nations World Restoration Flagships [112]. Ecological restoration encompasses diverse approaches, but combining protection of intact habitat with the restoration of degraded areas increases the likelihood of preventing biodiversity loss linked to fragmentation [107,108].

Despite global momentum, restoration outcomes remain mixed [109]. Many projects fail or fall short of expectations [110,113,114]. High failure rates often reflect an emphasis on the number or size of projects rather than on rigorous planning, design, implementation, and monitoring [106,115,116]. Scaling up restoration also requires broader adoption of technological innovations and meaningful involvement of local stakeholders [117]. Local engagement is essential, as resistance frequently arises when landowner priorities conflict with restoration objectives, limiting access to restorable areas [115]. Efforts to align habitat needs with societal benefits and foster community buy-in can improve long-term engagement and accessibility [118,119].

Large-scale restoration programs typically require extensive transdisciplinary collaboration and sustained political support [118]. However, political will remains a significant barrier, in part due to the complexity and high costs associated with restoration. Global estimates place median costs at up to USD3012/ha for terrestrial projects [120] and USD80,000/ha for marine coastal habitats [121]. Such costs can constrain restoration to smaller local initiatives [122], underscoring the challenge of implementing restoration at scales commensurate with global biodiversity loss.

#### 4.1. Realism Versus Scale in Habitat Restoration

Successful restoration requires overcoming numerous ecological, logistical, and social challenges. Although meeting ambitious restoration targets could yield benefits at local to global scales [122,123], the scale of restoration remains small compared to that of habitat loss and fragmentation. For example, Indonesia is planting 600,000 mangrove trees by 2024 [106], but the country has lost about 1 million hectares of mangroves since the 1800s [123], which implies that the restoration target represents roughly 0.06% of the total mangrove loss, assuming a planting density of 1000 trees per ha. Similarly, the Chesapeake Bay Program’s goal of restoring 130,000 acres of submerged aquatic vegetation by 2025 has not been met, and even if achieved, would remain well below historical estimates of 200,000–600,000 acres [124].

Shortfalls in meeting ambitious targets stem from multiple factors. A key challenge is the mismatch between restoration pragmatism and the scale of degradation. Project goals are often unrealistic, infeasible, or insufficiently defined [111,125]. Strategies that work locally may not scale to broader landscapes because site-specific conditions rarely translate across regions. Moreover, failures often arise from an inability to anticipate or plan for stochastic weather events, climatic variability, or other environmental uncertainties [123]. These uncertainties require flexible frameworks, making adaptive management essential [125,126].

The tension between realistic scale and ecological ambition, combined with broader restoration challenges, underscores the need to prioritize sites based on expected ecological and evolutionary outcomes. Genetic surveys can be used both to prioritize restoration sites and to measure the success of restoration activities [127]. Nevertheless, many restoration efforts still rely on feasibility and accessibility rather than scientific evidence. For instance, germplasm sourcing often emphasizes short-term practicality over long-term resilience [128]. Poor site selection driven by accessibility, as well as mismatches between species’ biology and selected sites, contribute substantially to mangrove restoration failures [129,130]. Ultimately, ecological restoration aims to establish systems capable of persisting under future environmental change without continual human intervention [115,131,132]. Achieving this goal requires accounting for species biology and for locally relevant within-species genetic variation, both of which are critical for building resilient restored landscapes [133]. Restoring habitat without restoring the inhabitants is bound to fail.

#### 4.2. The Value of Genetic Assessments in Ecological Restoration

Understanding the amount and spatial distribution of genetic diversity is fundamental to managing degraded landscapes, and although still underutilized, genetic diversity is increasingly considered in restoration research [127]. Yet for most species and regions, such data are still lacking. By quantifying levels and patterns of genetic variation, genetic assessments can identify vulnerable or at-risk populations that appear demographically stable but are genetically compromised, helping to determine the interventions needed to mitigate fragmentation impacts [134]. Assignment tests, mating-system analyses, and estimates of genetic connectivity can guide germplasm source population designation for minimizing risks associated with moving individuals or propagules [133,135]. Connectivity analyses can further highlight sites with strong potential for natural colonization [136], which may require only minimal intervention to support recruitment. These analyses also reveal dispersal barriers and population ancestries and can delineate management units or genetic neighborhoods [21,27,34].

Monitoring with genetic data can additionally illuminate the consequences of past restoration interventions and can refine future management strategies [134]. Such moni-

toring enables evaluation of whether restored populations retain diversity and integrate successfully into surrounding metapopulations [134]. Environmental DNA (eDNA) has expanded the capacity to monitor biodiversity non-invasively, providing crucial information on how fragmentation and other anthropogenic pressures reshape community composition in degraded habitats. eDNA can detect invasive species before they become problematic [137,138] and illuminate the status of rare or imperiled species, particularly where baseline community data from pre-disturbance conditions do not exist.

Genetic tools can also be paired with machine-learning approaches to optimize restoration design for landscape function, recovery, and resilience [139,140]. Despite their broad utility, genetic data remain underused in restoration planning—especially in low-resource settings [11]. Their omission often reflects the limited availability of baseline genetic information, making it difficult to distinguish natural patterns of variation from those driven by anthropogenic disturbance. This distinction is critical because genetic variation is the substrate on which natural selection acts and is fundamental to population viability.

Even when management aims to maximize genetic diversity for long-term resilience, restoration is not a rapid solution. Restored populations require time to establish, recruit, and meaningfully contribute to local gene pools. The timeline for genetic recovery depends on generation time, reproductive systems, and turnover rates. Locally sourced propagules are typically considered low-risk, high-reward for maintaining adaptation to local conditions [11,141]. However, in severely fragmented landscapes where local populations have experienced bottlenecks and are genetically depauperate, exclusive reliance on local germplasm may limit resilience [11,142]. In such cases, genetic rescue through assisted migration or targeted translocations may be necessary to restore viability [11,143,144].

## 5. Translocation

The human-mediated movement of individuals from one site to another via translocation can serve as an artificial means of facilitating gene flow in populations threatened by fragmentation and low genetic diversity. Translocations may support assisted colonization into suitable environments under global change, recolonize extirpated sites, or demographically and genetically bolster declining populations [145–147]. Effective use of translocation requires consideration of the source and number of individuals, the frequency and timing of releases, and the criteria used to evaluate success [146]. Individuals may be sourced from other wild populations or from captive-breeding and hatchery programs [148–150]. The number of individuals matters, as moving 50 versus 100 can have substantially different demographic and genetic consequences for the recipient population [151–153]. Frequency of translocation is also essential, as single versus repeated introductions may differ in their ability to meet project goals [148,151]. Success further depends on seasonal and phenological timing, release location, and the ability of individuals to disperse among viable habitat patches [145,146]. Ultimately, outcomes hinge on the interplay between translocated individuals and the biotic and abiotic conditions of the recipient environment [146,154].

The ecological results of translocation can be varied and occasionally unintended. First, introduced individuals must recover from transport stress and survive release [155,156]. Beyond survival, successful establishment requires that translocated individuals mature, enter the effective breeding population, and reproduce [154,157]. They must interbreed not only among themselves (assortative mating) but also with local individuals [154,158]. Concerns include increased competition for resources and the introduction of novel pathogens that could reduce fitness in recipient populations [159,160].

Nevertheless, when implemented in biological restoration and conservation contexts, translocations are generally successful and rarely result in adverse ecosystem-wide effects [161]. Beyond concerns about the recipient population, the effects of removing

individuals from source populations are often left unassessed, including changes in demographic stability over time [149]. However, the demographic consequences for source populations are frequently overlooked, and these populations may experience long-term declines after donating individuals—an especially critical issue for vulnerable or critically endangered species [162,163].

When translocations result in interbreeding between introduced and local populations, genetic benefits can be substantial. Facilitated or assisted gene flow can reverse genetic erosion, temporarily increase connectivity during periods when natural dispersal is unlikely, and restore genetic variation lost to drift [155,157]. Through genetic rescue [164], translocated individuals can enhance adaptive potential by replenishing the genetic variation available to natural selection [164,165]. Although outbreeding depression resulting in low fitness of offspring from adaptively divergent parents remains a concern [166], recent evidence suggests it is less common than previously thought [1,167]. Nonetheless, understanding population structure, divergence, and local adaptation remains essential when selecting donor populations to both preserve genetic identity and spread beneficial variation across landscapes [34,154,168]. It is also important to note that genetic benefits often unfold over many generations, potentially limiting the utility of translocation under rapid climate or land-use change [169].

#### *Genetic Surveys to Guide Translocation*

Facilitating gene flow through translocation to achieve genetic rescue requires a detailed understanding of population structure, standing genetic diversity, and patterns of adaptive differentiation across the landscape [170]. Genetic surveys provide the foundation for identifying the number of biologically meaningful populations, their boundaries, and the magnitude and direction of migration among them [27,34,63]. Population structure analyses, therefore, allow managers to move beyond arbitrary geographic units and instead define management units that reflect underlying evolutionary and demographic processes [170]. Quantifying within-population genetic diversity, inbreeding, and effective population size further enables managers to prioritize populations for intervention [34]. Populations with large effective sizes and high diversity may serve as robust donor sources. In contrast, small, isolated, or highly inbred populations can be identified as recipients most likely to benefit from assisted gene flow [164].

Although outbreeding depression appears to be less common than once feared [1,167], it remains prudent to evaluate adaptive differentiation among populations, particularly when translocations involve environmentally heterogeneous landscapes [170]. Genetic surveys can identify signals of local adaptation using genotype–environment association analyses, outlier scans, and clustering or assignment tests based on putatively adaptive genomic regions [34,171]. These approaches help managers avoid mixing individuals that are strongly adapted to contrasting environments, while still allowing movement among populations that share similar selective regimes. Models of contemporary landscape-level adaptation can also be projected into the future to estimate the potential maladaptation of populations and to prioritize donor and recipient populations [140]. Together, genetic surveys transform translocation from a largely demographic exercise into a genetically informed management strategy. By explicitly linking donor–recipient selection to population structure, diversity, and adaptive context, managers can maximize the likelihood that translocations enhance genetic variation and adaptive potential while minimizing the risks of maladaptation or unintended genetic consequences.

## 6. Targeted Genetic Intervention

The progression from managing landscapes (restoration) to managing populations (translocation) culminates in an emerging frontier: directly managing adaptive genetic variation within genomes [172,173]. Targeted Genetic Intervention (TGI) is broadly defined as a suite of approaches that seek to “change adaptive allele frequencies to increase population fitness,” using tools such as artificial selection, gene editing, synthetic biology, and targeted gene flow [174]. This definition encompasses a range of related concepts—including “assisted evolution” [173], “facilitated adaptation” [175], and “gene-targeted conservation” [176]—all aimed at promoting adaptation in species for which traditional conservation measures are inadequate [174].

Historically, conservation genetics emphasized minimizing genetic erosion and inbreeding to preserve existing evolutionary potential [177]. TGI departs from this preservation-oriented paradigm. For challenges such as infectious diseases, climate change, and toxic pollutants—where eliminating the threat is often not feasible—long-term persistence may depend on the species’ ability to adapt. TGI seeks to accelerate this adaptive process, shifting conservation from maintaining genetic variation to actively managing the genes themselves [174]. For species already reliant on intensive management (e.g., Conservation Breeding Programs), TGI offers a potential shift from maintaining perpetual “ark” populations to establishing resilient, self-sustaining wild populations.

Conservation breeding programs, often viewed as a last resort for critically endangered species, are in fact the indispensable platform for developing and implementing TGI. Controlled, ex situ environments provide access to individuals for physiological studies, controlled breeding, gamete collection, and application of advanced reproductive technologies, which are prerequisites for nearly all genetic interventions. However, captivity introduces significant genetic challenges, including relaxed selection, adaptation to captive conditions, and reduced fitness [178]. This generates a well-documented “demography–genetics trade-off” [179]: longer periods in captivity allow greater demographic growth and more individuals for release, but also increase the accumulation of genetic load and reduce post-release fitness. TGI provides a compelling solution by targeting and preserving—or even enhancing—wild-type adaptive traits, potentially breaking the link between time in captivity and fitness decline.

Gene editing approaches such as CRISPR–Cas9 [180] can introduce advantageous variants directly into genomes to accelerate adaptation [175,181], enabling “synthetically assisted conservation translocations” [182]. Genetic modification has long been used in crops to circumvent reproductive barriers in plant breeding [183]. Although the application of such technologies to wildlife has progressed more slowly [184], several promising examples are emerging. Synthetic methods are being explored to enhance thermal tolerance in branching corals [185] and combat chytridiomycosis in amphibians [186]. The American Chestnut Research and Restoration Project has used transgenesis to insert a wheat gene conferring blight tolerance into the American chestnut genome [187]. The resulting strain, “Darling-54,” is currently under review for wider planting by the USDA, FDA, and EPA, though The American Chestnut Foundation withdrew support in 2023 [188].

### 6.1. Facilitating the Spread of Adaptive Traits with TGI

TGI can be used to intentionally spread advantageous genetic variants that already exist in some populations but are absent in others that would benefit from them [181]. This approach avoids transgenesis and the introduction of ‘foreign’ DNA into species and may not be strictly regulated as genetic modification [189]. Implementation begins with a comprehensive genomic survey—ideally using low-coverage whole-genome sequencing [190]. Once genomic variation is characterized at scale, candidate adaptive loci can be identified

through genotype–environment association analyses [34,171], followed by functional validation using genome annotation and gene ontology [34], and, when possible, expression or physiological evidence linking loci to traits of interest. Experimental validation can then confirm that manipulating these genes yields meaningful phenotypic improvements, such as enhanced tolerance to warming or disease [191].

After identifying and validating target variants, CRISPR-based editing [180] can be integrated into ex situ conservation breeding programs to introduce these alleles into fertilized embryos from populations that lack them. Before release, edited individuals must be evaluated in contained ex situ environments to assess performance, fitness, and potential unintended effects. Initial applications should focus on species that are already highly fragmented and at high risk of extinction.

TGI offers several advantages over conventional translocation-based genetic rescue. It reduces demographic pressure on source populations, avoids introducing linked deleterious alleles that may contribute to outbreeding depression, and preserves unique variation in recipient populations. It also eliminates the risk of pathogen transfer associated with moving whole individuals. Most importantly, for species that are genetically depauperate, inbred, and facing increasingly limited conventional conservation options, TGI may represent the only viable path to restoring adaptive potential and preventing population collapse.

## 6.2. Challenges and Promises for TGI

Despite its theoretical appeal, TGI is not without criticism [176,192]. Kardos and Shafer [176] caution that such approaches may be ineffective at best and counterproductive at worst. A central challenge is that most fitness-related traits of conservation interest, such as thermal tolerance, disease resistance, growth, and reproduction, are highly polygenic, influenced by hundreds or thousands of loci of small effect [176,193,194]. Identifying a small number of “target genes” that meaningfully shift such traits may be unlikely, and even the best-supported candidates typically explain only a small fraction of the underlying genetic variance [193].

Focusing on a narrow set of beneficial alleles also risks eroding genome-wide genetic diversity, which remains the primary buffer against unpredictable future environments [176]. By promoting specific alleles and narrowing the breeding pool, TGI may reduce effective population size and accelerate genetic drift, potentially worsening long-term outcomes. In addition, strong selection on individual loci can have unintended effects through pleiotropy or linkage, enabling deleterious alleles to “hitchhike” to high frequency [195].

Even if these biological and technical hurdles are addressed, bringing TGI into practice will require more than scientific feasibility. It demands a governance model that is ethically grounded, publicly legitimate, and internationally coherent [196]. Because TGI involves heritable changes to wild populations—shared public goods rather than private entities—its implementation must be guided by robust ethical frameworks, ongoing and inclusive stakeholder engagement, and regulatory systems capable of assessing ecological risks over landscape and generational scales [197]. The recent IUCN policy [198] on synthetic biology offers an initial foundation, emphasizing precaution, transparency, and the requirement of Free, Prior, and Informed Consent from affected communities. Yet regulatory pathways remain underdeveloped, public trust cannot be assumed, and governance must be proactively designed rather than adapted from agricultural or biomedical precedents [196]. Ultimately, even if TGI is technically feasible, its viability will depend on whether society is willing to accept and carefully steward its use.

## 7. Conclusions

Across fragmented landscapes, conservation efforts increasingly require a coordinated strategy that integrates habitat restoration, population management, and genetic insight [127]. A central message emerging from this perspective is that genetic work does not detract from traditional conservation approaches; rather, it strengthens them. The cost of generating genetic data is modest compared to the immense financial and logistical investments required for large-scale restoration, and these data can dramatically increase the efficiency and impact of those investments [128].

Genetic information provides a powerful basis for prioritizing restoration sites by identifying populations most at risk and landscapes where connectivity can deliver the greatest ecological and evolutionary benefits [127]. Restoration alone, however, rarely “fixes” genetically depauperate populations. Without renewed connectivity to genetically diverse sources—or direct genetic intervention—recovery of diversity can take many generations. Critically, the genetic outcomes of restoration cannot be inferred without baseline data and follow-up monitoring, underscoring the need to incorporate genetic assessments as standard practice [11], and to broaden population genomics to the community-level [27].

Practitioners should incorporate genomic data into their monitoring toolkit. Several reasonable data types for manager use exist depending on the scope of the project, ranging from microsatellites and single nucleotide polymorphisms (SNPs) to low-coverage and whole-genome sequencing, allowing for a variety of survey tools of sampled individuals’ genomes to assess success of the project [199]. Sampled individuals should be from a range of age classes, to ensure multiple generations (pre- and post-intervention) are included. This data can be used in post-intervention monitoring, which will, at minimum, require two timepoints: baseline (collected before or at the time of intervention) and short-term (collected after 1–2 generations). This schedule will help determine if gene flow is occurring between connected or introduced populations, with the potential for annual and long-term (collected several generations after intervention) monitoring. However, specific monitoring timelines will rely heavily on the species of interest, considering generation time, dispersal, reproductive system, and intervention type [200,201].

Translocation offers a more direct mechanism for restoring genetic diversity: moving individuals is ultimately moving genes, assuming subsequent breeding. When implemented carefully, assisted gene flow and genetic rescue can counteract drift, replenish adaptive variation, and bolster long-term persistence [164]. Yet translocation also carries demographic, ecological, and logistical constraints that limit its utility for many species.

Connectivity between patches, achieved through habitat restoration or facilitated by translocation, does come with associated risks to the persistence of the intended conservation species. Re-establishing connectivity can provide pathways for invasive species spread and the introduction of novel pathogens or even increase the abundance of predators/competitors, whereas translocations are more commonly associated with disease spread and intraspecific competition pressure [159,160,202]. While increasing connectivity may introduce or increase risks to ecosystem health and stability, the demographic and genetic benefits gained may outweigh these costs and are likely not achieved using other means.

Looking ahead, the future of genetic rescue may lie in Targeted Genetic Intervention (TGI). TGI seeks to accelerate the spread of advantageous genetic variants already present within a species, effectively mimicking—and in some cases surpassing—the goals of traditional translocation while reducing pressure on donor populations and minimizing unintended consequences [174]. Although TGI remains in early development and faces scientific, ethical, and governance challenges, its potential to restore adaptive capacity in severely fragmented or rapidly declining species is substantial.

Collectively, restoration, translocation, and emerging genetic interventions should not be viewed as competing pathways but as complementary tools within a unified management framework. Integrating genomic data throughout these efforts will be essential for ensuring that conservation actions not only rebuild habitat and populations but also restore the evolutionary potential required for species to persist in an increasingly dynamic world.

**Author Contributions:** Z.D.Z. conceived and outlined the research with input from all authors. All authors contributed to the drafting and critical revision of the manuscript. All authors have read and agreed to the published version of the manuscript.

**Funding:** Funding and support were provided by the University of Maryland Center for Environmental Science.

**Institutional Review Board Statement:** Not applicable.

**Data Availability Statement:** No new data were created or analyzed in this study. Data sharing is not applicable to this article.

**Acknowledgments:** We thank the excellent administrative staff who make our work possible.

**Conflicts of Interest:** The auhtors declare no conflicts of interest.

## References

1. Ralls, K.; Ballou, J.D.; Dudash, M.R.; Eldridge, M.D.B.; Fenster, C.B.; Lacy, R.C.; Sunnucks, P.; Frankham, R. Call for a Paradigm Shift in the Genetic Management of Fragmented Populations. *Conserv. Lett.* **2018**, *11*, e12412. [[CrossRef](#)]
2. Hogue, A.S.; Breon, K. The Greatest Threats to Species. *Conserv. Sci. Pract.* **2022**, *4*, e12670. [[CrossRef](#)]
3. Haddad, N.M.; Brudvig, L.A.; Clobert, J.; Davies, K.F.; Gonzalez, A.; Holt, R.D.; Lovejoy, T.E.; Sexton, J.O.; Austin, M.P.; Collins, C.D.; et al. Habitat Fragmentation and Its Lasting Impact on Earth's Ecosystems. *Sci. Adv.* **2015**, *1*, e1500052. [[CrossRef](#)] [[PubMed](#)]
4. Kuipers, K.J.J.; Hilbers, J.P.; Garcia-Ulloa, J.; Graae, B.J.; May, R.; Verones, F.; Huijbregts, M.A.J.; Schipper, A.M. Habitat Fragmentation Amplifies Threats from Habitat Loss to Mammal Diversity across the World's Terrestrial Ecoregions. *One Earth* **2021**, *4*, 1505–1513. [[CrossRef](#)]
5. Jones, K.R.; Klein, C.J.; Halpern, B.S.; Venter, O.; Grantham, H.; Kuempel, C.D.; Shumway, N.; Friedlander, A.M.; Possingham, H.P.; Watson, J.E.M. The Location and Protection Status of Earth's Diminishing Marine Wilderness. *Curr. Biol.* **2018**, *28*, 2506–2512.e3. [[CrossRef](#)] [[PubMed](#)]
6. Su, G.; Logez, M.; Xu, J.; Tao, S.; Villéger, S.; Brosse, S. Human Impacts on Global Freshwater Fish Biodiversity. *Science* **2021**, *371*, 835–838. [[CrossRef](#)]
7. Didham, R.K. Ecological Consequences of Habitat Fragmentation. In *Encyclopedia of Life Sciences*; John Wiley & Sons, Ltd.: Hoboken, NJ, USA, 2010; ISBN 978-0-470-01590-2.
8. Saunders, D.A.; Hobbs, R.J.; Margules, C.R. Biological Consequences of Ecosystem Fragmentation: A Review. *Conserv. Biol.* **1991**, *5*, 18–32. [[CrossRef](#)]
9. Hoban, S.; Campbell, C.D.; da Silva, J.M.; Ekblom, R.; Funk, W.C.; Garner, B.A.; Godoy, J.A.; Kershaw, F.; MacDonald, A.J.; Mergeay, J.; et al. Genetic Diversity Is Considered Important but Interpreted Narrowly in Country Reports to the Convention on Biological Diversity: Current Actions and Indicators Are Insufficient. *Biol. Conserv.* **2021**, *261*, 109233. [[CrossRef](#)]
10. Laikre, L.; Allendorf, F.W.; Aroner, L.C.; Baker, C.S.; Gregovich, D.P.; Hansen, M.M.; Jackson, J.A.; Kendall, K.C.; McKelvey, K.; Neel, M.C.; et al. Neglect of Genetic Diversity in Implementation of the Convention of Biological Diversity. *Conserv. Biol.* **2010**, *24*, 86–88. [[CrossRef](#)]
11. Ngeve, M.N. Genetic Diversity Must Be Explicitly Recognized in Ecological Restoration. *Nat. Clim. Change* **2025**, *15*, 908–909. [[CrossRef](#)]
12. Vranckx, G.; Jacquemyn, H.; Muys, B.; Honnay, O. Meta-Analysis of Susceptibility of Woody Plants to Loss of Genetic Diversity through Habitat Fragmentation. *Conserv. Biol.* **2012**, *26*, 228–237. [[CrossRef](#)]
13. Lienert, J. Habitat Fragmentation Effects on Fitness of Plant Populations—A Review. *J. Nat. Conserv.* **2004**, *12*, 53–72. [[CrossRef](#)]
14. Keyghobadi, N. The Genetic Implications of Habitat Fragmentation for Animals. *Can. J. Zool.* **2007**, *85*, 1049–1064. [[CrossRef](#)]
15. Jamieson, I.G.; Allendorf, F.W. How Does the 50/500 Rule Apply to MVPs? *Trends Ecol. Evol.* **2012**, *27*, 578–584. [[CrossRef](#)] [[PubMed](#)]
16. Jones, T.A.; Monaco, T.A. A Restoration Practitioner's Guide to the Restoration Gene Pool Concept. *Ecol. Restor.* **2007**, *25*, 12–19. [[CrossRef](#)]

17. Johnson, M.P. Rocky Intertidal Invertebrates: The Potential for Metapopulations within and among Shores. In *Marine Metapopulations*; Academic Press: Cambridge, MA, USA, 2006; pp. 247–270.
18. Bacon, C.D.; Mora, A.; Wagner, W.L.; Jaramillo, C.A. Testing Geological Models of Evolution of the Isthmus of Panama in a Phylogenetic Framework. *Bot. J. Linn. Soc.* **2013**, *171*, 287–300. [[CrossRef](#)]
19. Moonlight, P.W.; Richardson, J.E.; Tebbitt, M.C.; Thomas, D.C.; Hollands, R.; Peng, C.-I.; Hughes, M. Continental-Scale Diversification Patterns in a Megadiverse Genus: The Biogeography of Neotropical Begonia. *J. Biogeogr.* **2015**, *42*, 1137–1149. [[CrossRef](#)]
20. Rana, H.K.; Luo, D.; Rana, S.K.; Sun, H. Geological and Climatic Factors Affect the Population Genetic Connectivity in *Mirabilis Himalaica* (Nyctaginaceae): Insight from Phylogeography and Dispersal Corridors in the Himalaya-Hengduan Biodiversity Hotspot. *Front. Plant Sci.* **2020**, *10*, 1721. [[CrossRef](#)]
21. Ngeve, M.N.; der Stocken, T.V.; Menemenlis, D.; Koedam, N.; Triest, L. Contrasting Effects of Historical Sea Level Rise and Contemporary Ocean Currents on Regional Gene Flow of *Rhizophora Racemosa* in Eastern Atlantic Mangroves. *PLoS ONE* **2016**, *11*, e0150950. [[CrossRef](#)]
22. Raw, J.L.; Van der Stocken, T.; Carroll, D.; Harris, L.R.; Rajkaran, A.; Van Niekerk, L.; Adams, J.B. Dispersal and Coastal Geomorphology Limit Potential for Mangrove Range Expansion under Climate Change. *J. Ecol.* **2023**, *111*, 139–155. [[CrossRef](#)]
23. Singh, A.R.; Thirumurugan, V.; Bhomia, R.K.; Prabakaran, N. Mangrove Vegetation Response to Alteration in Coastal Geomorphology after an Earthquake in Andaman Islands, India. *Reg. Stud. Mar. Sci.* **2024**, *76*, 103583. [[CrossRef](#)]
24. Arnold, T.M.; Zimmerman, R.C.; Engelhardt, K.A.M.; Stevenson, J.C. Twenty-First Century Climate Change and Submerged Aquatic Vegetation in a Temperate Estuary: The Case of Chesapeake Bay. *Ecosyst. Health Sustain.* **2017**, *3*, 1353283. [[CrossRef](#)]
25. Bernard, A.; Alleaume, S.; Fortuny, X.; Munoz, F.; Pitard, E. Environmental Factors Governing Spatio-Temporal Series of Aquatic Vegetation in the Bagnas Lagoon. *Estuar. Coast. Shelf Sci.* **2025**, *319*, 109261. [[CrossRef](#)]
26. Felin, S.; Belliard, J.; Grenouillet, G.; Moatar, F.; Le Pichon, C.; Thieu, V.; Thirel, G.; Jeliazkov, A. The Role of River Connectivity in the Distribution of Fish in an Anthropized Watershed. *Sci. Total Environ.* **2025**, *959*, 178204. [[CrossRef](#)]
27. Zbinden, Z.D.; Douglas, M.R.; Chafin, T.K.; Douglas, M.E. Riverscape Community Genomics: A Comparative Analytical Approach to Identify Common Drivers of Spatial Structure. *Mol. Ecol.* **2023**, *32*, 6743–6765. [[CrossRef](#)]
28. Benoit-Bird, K.J. Resource Patchiness as a Resolution to the Food Paradox in the Sea. *Am. Nat.* **2024**, *203*, 1–13. [[CrossRef](#)] [[PubMed](#)]
29. Gray, P.C.; Boss, E.; Bourdin, G.; Lehahn, Y. Emergent Patterns of Patchiness Differ between Physical and Planktonic Properties in the Ocean. *Nat. Commun.* **2025**, *16*, 1808. [[CrossRef](#)]
30. Robinson, K.L.; Sponaugle, S.; Luo, J.Y.; Gleiber, M.R.; Cowen, R.K. Big or Small, Patchy All: Resolution of Marine Plankton Patch Structure at Micro- to Submesoscales for 36 Taxa. *Sci. Adv.* **2021**, *7*, eabk2904. [[CrossRef](#)]
31. Bai, X.; Zhang, P.; Xiong, L.; Yang, Z.; Wang, S.; Dong, X.; Sun, H.; Li, W.; Chang, J. River Network Connectivity Reductions Dominate Declines in the Richness of Plateau Fish Species Under Climate Change in the Upper Yangtze River Basin. *Water Resour. Res.* **2025**, *61*, e2024WR037557. [[CrossRef](#)]
32. Hieb, E.E.; Eniang, E.A.; Keith-Diagne, L.W.; Carmichael, R.H. In-Water Bridge Construction Effects on Manatees with Implications for Marine Megafauna Species. *J. Wildl. Manag.* **2021**, *85*, 674–685. [[CrossRef](#)]
33. Salvatierra, D.; González, M.P.; Blasco, J.; Krull, M.; Araújo, C.V.M. Habitat Loss and Discontinuity as Drivers of Habitat Fragmentation: The Role of Contamination and Connectivity of Habitats. *Environ. Res.* **2025**, *266*, 120609. [[CrossRef](#)]
34. Zbinden, Z.D.; Chafin, T.K.; Tiemann, J.S.; Edds, D.R.; Martin, B.T.; Hofmeier, J.; Douglas, M.E.; Douglas, M.R. Historic and Contemporary Selection Define Conservation Units for a Short-Range Endemic within an Anthropogenically-Altered Riverscape. *Glob. Ecol. Conserv.* **2025**, *62*, e03783. [[CrossRef](#)]
35. Schlaepfer, D.R.; Braschler, B.; Rusterholz, H.-P.; Baur, B. Genetic Effects of Anthropogenic Habitat Fragmentation on Remnant Animal and Plant Populations: A Meta-Analysis. *Ecosphere* **2018**, *9*, e02488. [[CrossRef](#)]
36. Sun, J.; Baldan, D.; Lucas, M.C.; Wang, J.; Rodeles, A.A.; Galib, S.M.; Tao, J.; Li, M.; He, D.; Ding, C. Widespread and Strong Impacts of River Fragmentation by Anthropogenic Barriers on Fishes in the Mekong River Basin. *Commun. Earth Environ.* **2025**, *6*, 534. [[CrossRef](#)]
37. Wang, G.; Zheng, R.; He, Y.; Wei, L.; Guan, D.; Motelica-Heino, M.; Xiao, L. Community Structure and Carbon Storage of Mangrove Forests in Hainan Island, China Affected by Their Patch Characteristics. *Reg. Stud. Mar. Sci.* **2025**, *90*, 104388. [[CrossRef](#)]
38. Rodemann, J.R.; James, W.R.; Rehage, J.S.; Furman, B.T.; Pittman, S.J.; Santos, R.O. Response of Submerged Aquatic Vegetation Seascapes to a Large-Scale Seagrass Die-off: A Case Study in Florida Bay. *Estuar. Coast. Shelf Sci.* **2025**, *318*, 109221. [[CrossRef](#)]
39. Hamberg, J.; Findlay, S.E.G.; Limburg, K.E.; Diemont, S.A.W. Post-Storm Sediment Burial and Herbivory of *Vallisneria Americana* in the Hudson River Estuary: Mechanisms of Loss and Implications for Restoration. *Restor. Ecol.* **2017**, *25*, 629–639. [[CrossRef](#)]
40. Michelena, T.M.; Boylen, C.W.; Nierwicki-Bauer, S.A. Impacts of Hurricane Irene and Tropical Storm Lee on the Ecology of the Hudson River Estuary. *Int. J. River Basin Manag.* **2019**, *17*, 403–410. [[CrossRef](#)]

41. Ngeve, M.N.; Engelhardt, K.A.M.; Gray, M.; Neel, M.C. Calm after the Storm? Similar Patterns of Genetic Variation in a Riverine Foundation Species before and after Severe Disturbance. *Ecol. Evol.* **2023**, *13*, e10670. [[CrossRef](#)]
42. Bovo, A.A.A.; Ferraz, K.M.P.M.B.; Magioli, M.; Alexandrino, E.R.; Hasui, É.; Ribeiro, M.C.; Tobias, J.A. Habitat Fragmentation Narrows the Distribution of Avian Functional Traits Associated with Seed Dispersal in Tropical Forest. *Perspect. Ecol. Conserv.* **2018**, *16*, 90–96. [[CrossRef](#)]
43. Browne, L.; Karubian, J. Habitat Loss and Fragmentation Reduce Effective Gene Flow by Disrupting Seed Dispersal in a Neotropical Palm. *Mol. Ecol.* **2018**, *27*, 3055–3069. [[CrossRef](#)]
44. Crowther, M.S.; Rus, A.I.; Mella, V.S.A.; Krockenberger, M.B.; Lindsay, J.; Moore, B.D.; McArthur, C. Patch Quality and Habitat Fragmentation Shape the Foraging Patterns of a Specialist Folivore. *Behav. Ecol.* **2022**, *33*, 1007–1017. [[CrossRef](#)]
45. Chase, J.M.; Blowes, S.A.; Knight, T.M.; Gerstner, K.; May, F. Ecosystem Decay Exacerbates Biodiversity Loss with Habitat Loss. *Nature* **2020**, *584*, 238–243. [[CrossRef](#)]
46. Riva, F.; Pierre, E.; Guisan, A. On the Emergence of Ecosystem Decay: A Critical Assessment of Patch Area Effects across Spatial Scales. *Biol. Conserv.* **2024**, *296*, 110674. [[CrossRef](#)]
47. Engelhardt, K.A.M.; Lloyd, M.W.; Neel, M.C. Effects of Genetic Diversity on Conservation and Restoration Potential at Individual, Population, and Regional Scales. *Biol. Conserv.* **2014**, *179*, 6–16. [[CrossRef](#)]
48. Ingala, M.R.; Becker, D.J.; Bak Holm, J.; Kristiansen, K.; Simmons, N.B. Habitat Fragmentation Is Associated with Dietary Shifts and Microbiota Variability in Common Vampire Bats. *Ecol. Evol.* **2019**, *9*, 6508–6523. [[CrossRef](#)] [[PubMed](#)]
49. Xiao, Y.; Li, X.; Cao, Y.; Dong, M. The Diverse Effects of Habitat Fragmentation on Plant–Pollinator Interactions. *Plant Ecol.* **2016**, *217*, 857–868. [[CrossRef](#)]
50. Jump, A.S.; Peñuelas, J. Genetic Effects of Chronic Habitat Fragmentation in a Wind-Pollinated Tree. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 8096–8100. [[CrossRef](#)]
51. Brauer, C.J.; Beheregaray, L.B. Recent and Rapid Anthropogenic Habitat Fragmentation Increases Extinction Risk for Freshwater Biodiversity. *Evol. Appl.* **2020**, *13*, 2857–2869. [[CrossRef](#)]
52. Crooks, K.R.; Burdett, C.L.; Theobald, D.M.; King, S.R.B.; Di Marco, M.; Rondinini, C.; Boitani, L. Quantification of Habitat Fragmentation Reveals Extinction Risk in Terrestrial Mammals. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 7635–7640. [[CrossRef](#)] [[PubMed](#)]
53. Yang, G.; Crowther, T.W.; Lauber, T.; Zohner, C.M.; Smith, G.R. A Globally Consistent Negative Effect of Edge on Aboveground Forest Biomass. *Nat. Ecol. Evol.* **2025**, *9*, 2036–2045. [[CrossRef](#)]
54. Stangler, E.S.; Hanson, P.E.; Steffan-Dewenter, I. Interactive Effects of Habitat Fragmentation and Microclimate on Trap-Nesting Hymenoptera and Their Trophic Interactions in Small Secondary Rainforest Remnants. *Biodivers. Conserv.* **2015**, *24*, 563–577. [[CrossRef](#)]
55. Fontúrbel, F.E.; Murúa, M.M. Microevolutionary Effects of Habitat Fragmentation on Plant-Animal Interactions. *Adv. Ecol.* **2014**, *2014*, 379267. [[CrossRef](#)]
56. Banbury Morgan, R.; Jucker, T. A Unifying Framework for Understanding How Edge Effects Reshape the Structure, Composition and Function of Forests. *New Phytol.* **2025**, *248*, 529–541. [[CrossRef](#)]
57. Yarnall, A.H.; Byers, J.E.; Yeager, L.A.; Fodrie, F.J. Comparing Edge and Fragmentation Effects within Seagrass Communities: A Meta-Analysis. *Ecology* **2022**, *103*, e3603. [[CrossRef](#)]
58. da Silva, W.K.; de Matos, A.C.L.; Zenni, R.D. Habitat Permeability Drives Community Metrics, Functional Traits, and Herbivory in Neotropical Spontaneous Urban Flora. *Flora* **2024**, *319*, 152581. [[CrossRef](#)]
59. Xu, J.; Ling, Y.; Sun, Y.; Jiang, Y.; Shen, R.; Wang, Y. How Do Different Processes of Habitat Fragmentation Affect Habitat Quality?—Evidence from China. *Ecol. Indic.* **2024**, *160*, 111880. [[CrossRef](#)]
60. Hulting, K.A.; Brudvig, L.A.; Damschen, E.I.; Levey, D.J.; Resasco, J.; Tewksbury, J.J.; Haddad, N.M. Habitat Edges Decrease Plant Reproductive Output in Fragmented Landscapes. *J. Ecol.* **2025**, *113*, 531–541. [[CrossRef](#)]
61. Aguirre, A.; Dirzo, R. Effects of Fragmentation on Pollinator Abundance and Fruit Set of an Abundant Understory Palm in a Mexican Tropical Forest. *Biol. Conserv.* **2008**, *141*, 375–384. [[CrossRef](#)]
62. Damschen, E.I.; Baker, D.V.; Bohrer, G.; Nathan, R.; Orrock, J.L.; Turner, J.R.; Brudvig, L.A.; Haddad, N.M.; Levey, D.J.; Tewksbury, J.J. How Fragmentation and Corridors Affect Wind Dynamics and Seed Dispersal in Open Habitats. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 3484–3489. [[CrossRef](#)]
63. Chafin, T.K.; Zbinden, Z.D.; Douglas, M.R.; Martin, B.T.; Middaugh, C.R.; Gray, M.C.; Ballard, J.R.; Douglas, M.E. Spatial Population Genetics in Heavily Managed Species: Separating Patterns of Historical Translocation from Contemporary Gene Flow in White-Tailed Deer. *Evol. Appl.* **2021**, *14*, 1673–1689. [[CrossRef](#)]
64. Rodeles, A.A.; Galicia, D.; Miranda, R. A Simple Method to Assess the Fragmentation of Freshwater Fish Meta-Populations: Implications for River Management and Conservation. *Ecol. Indic.* **2021**, *125*, 107557. [[CrossRef](#)]
65. Acre, M.R.; Grabowski, T.B.; Leavitt, D.J.; Smith, N.G.; Pease, A.A.; Pease, J.E. Blue Sucker Habitat Use in a Regulated Texas River: Implications for Conservation and Restoration. *Environ. Biol. Fishes* **2021**, *104*, 501–516. [[CrossRef](#)]

66. Wegscheider, B.; Waldock, C.; Calegari, B.B.; Josi, D.; Brodersen, J.; Seehausen, O. Neglecting Biodiversity Baselines in Longitudinal River Connectivity Restoration Impacts Priority Setting. *Sci. Total Environ.* **2024**, *954*, 175167. [[CrossRef](#)]
67. Barbarossa, V.; Schmitt, R.J.P.; Huijbregts, M.A.J.; Zarfl, C.; King, H.; Schipper, A.M. Impacts of Current and Future Large Dams on the Geographic Range Connectivity of Freshwater Fish Worldwide. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 3648–3655. [[CrossRef](#)]
68. Edge, C.B.; Fortin, M.-J.; Jackson, D.A.; Lawrie, D.; Stanfield, L.; Shrestha, N. Habitat Alteration and Habitat Fragmentation Differentially Affect Beta Diversity of Stream Fish Communities. *Landscape Ecol.* **2017**, *32*, 647–662. [[CrossRef](#)]
69. Valenzuela-Aguayo, F.; McCracken, G.R.; Manosalva, A.; Habit, E.; Ruzzante, D.E. Human-Induced Habitat Fragmentation Effects on Connectivity, Diversity, and Population Persistence of an Endemic Fish, *Percilia Irwini*, in the Biobío River Basin (Chile). *Evol. Appl.* **2020**, *13*, 794–807. [[CrossRef](#)] [[PubMed](#)]
70. Cheptou, P.-O.; Hargreaves, A.L.; Bonte, D.; Jacquemyn, H. Adaptation to Fragmentation: Evolutionary Dynamics Driven by Human Influences. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2017**, *372*, 20160037. [[CrossRef](#)] [[PubMed](#)]
71. Bohonak, A.J. Dispersal, Gene Flow, and Population Structure. *Q. Rev. Biol.* **1999**, *74*, 21–45. [[CrossRef](#)]
72. Lowe, W.H.; Allendorf, F.W. What Can Genetics Tell Us about Population Connectivity? *Mol. Ecol.* **2010**, *19*, 3038–3051. [[CrossRef](#)] [[PubMed](#)]
73. MacArthur, R.H.; Wilson, E.O. *The Theory of Island Biogeography*; REV-Revised.; Princeton University Press: Princeton, NJ, USA, 1967; ISBN 978-0-691-08836-5.
74. Hanski, I. *Metapopulation Ecology*; Oxford Series in Ecology and Evolution; Oxford University Press: Oxford, NY, USA, 1999; ISBN 978-0-19-854065-6.
75. Levins, R. Some Demographic and Genetic Consequences of Environmental Heterogeneity for Biological Control. *Bull. Entomol. Soc. Am.* **1969**, *15*, 237–240. [[CrossRef](#)]
76. Saatoglu, D.; Lundregan, S.L.; Fetterplace, E.; Goedert, D.; Husby, A.; Niskanen, A.K.; Muff, S.; Jensen, H. The Genetic Basis of Dispersal in a Vertebrate Metapopulation. *Mol. Ecol.* **2024**, *33*, e17295. [[CrossRef](#)]
77. Chiu, M.-C.; Nukazawa, K.; Resh, V.H.; Watanabe, K. Environmental Effects, Gene Flow and Genetic Drift: Unequal Influences on Genetic Structure across Landscapes. *J. Biogeogr.* **2023**, *50*, 352–364. [[CrossRef](#)]
78. Wright, S. Evolution in Mendelian Populations. *Genetics* **1931**, *16*, 97–159. [[CrossRef](#)]
79. Slatkin, M. Rare Alleles as Indicators of Gene Flow. *Evolution* **1985**, *39*, 53–65. [[CrossRef](#)] [[PubMed](#)]
80. Pinto, A.V.; Hansson, B.; Patramanis, I.; Morales, H.E.; van Oosterhout, C. The Impact of Habitat Loss and Population Fragmentation on Genomic Erosion. *Conserv. Genet.* **2024**, *25*, 49–57. [[CrossRef](#)]
81. Bitume, E.V.; Bonte, D.; Ronce, O.; Bach, F.; Flaven, E.; Olivieri, I.; Nieberding, C.M. Density and Genetic Relatedness Increase Dispersal Distance in a Subsocial Organism. *Ecol. Lett.* **2013**, *16*, 430–437. [[CrossRef](#)]
82. Waples, R.S. The Idiot's Guide to Effective Population Size. *Mol. Ecol.* **2025**, *34*, e17670. [[CrossRef](#)]
83. Barton, N.H.; Charlesworth, B. Genetic Revolutions, Founder Effects, and Speciation. *Annu. Rev. Ecol. Syst.* **1984**, *15*, 133–164. [[CrossRef](#)]
84. Bosse, M.; Megens, H.-J.; Derkx, M.F.L.; de Cara, Á.M.R.; Groenen, M.A.M. Deleterious Alleles in the Context of Domestication, Inbreeding, and Selection. *Evol. Appl.* **2019**, *12*, 6–17. [[CrossRef](#)]
85. Mathur, S.; DeWoody, J.A. Genetic Load Has Potential in Large Populations but Is Realized in Small Inbred Populations. *Evol. Appl.* **2021**, *14*, 1540–1557. [[CrossRef](#)] [[PubMed](#)]
86. Turnock, M.F.; Teisberg, J.E.; Kasworm, W.F.; Falcy, M.R.; Proctor, M.F.; Waits, L.P. Gene Flow Prevents Genetic Diversity Loss despite Small Effective Population Size in Fragmented Grizzly Bear (*Ursus Arctos*) Populations. *Conserv. Genet.* **2025**, *26*, 279–291. [[CrossRef](#)]
87. Schmidt, C.; Hoban, S.; Hunter, M.; Paz-Vinas, I.; Garroway, C.J. Genetic Diversity and IUCN Red List Status. *Conserv. Biol.* **2023**, *37*, e14064. [[CrossRef](#)]
88. Cristóbal-Pérez, E.J.; Fuchs, E.J.; Martén-Rodríguez, S.; Quesada, M. Habitat Fragmentation Negatively Affects Effective Gene Flow via Pollen, and Male and Female Fitness in the Dioecious Tree, *Spondias purpurea* (Anacardiaceae). *Biol. Conserv.* **2021**, *256*, 109007. [[CrossRef](#)]
89. Giuntini, S.; Pedruzzi, L. Sex and the Patch: The Influence of Habitat Fragmentation on Terrestrial Vertebrates' Mating Strategies. *Ethol. Ecol. Evol.* **2023**, *35*, 269–298. [[CrossRef](#)]
90. Lynch, M. Evolution of the Mutation Rate. *Trends Genet.* **2010**, *26*, 345–352. [[CrossRef](#)]
91. Sniegowski, P.D.; Gerrish, P.J.; Johnson, T.; Shaver, A. The Evolution of Mutation Rates: Separating Causes from Consequences. *BioEssays* **2000**, *22*, 1057–1066. [[CrossRef](#)] [[PubMed](#)]
92. Hedrick, P.W. Gene Flow and Genetic Restoration: The Florida Panther as a Case Study. *Conserv. Biol.* **1995**, *9*, 996–1007.
93. Hoffmann, A.A.; Sgrò, C.M.; Kristensen, T.N. Revisiting Adaptive Potential, Population Size, and Conservation. *Trends Ecol. Evol.* **2017**, *32*, 506–517. [[CrossRef](#)]
94. Swindell, W.R.; Bouzat, J.L. Modeling the adaptive potential of isolated populations: Experimental simulations using drosophila. *Evolution* **2005**, *59*, 2159–2169. [[CrossRef](#)]

95. With, K.A.; Gardner, R.H.; Turner, M.G. Landscape Connectivity and Population Distributions in Heterogeneous Environments. *Oikos* **1997**, *78*, 151–169. [CrossRef]
96. Chevalet, C.; De Rochambeau, H. Predicting the Genetic Drift in Small Populations. *Livest. Prod. Sci.* **1985**, *13*, 207–218. [CrossRef]
97. Pardo, L.M.; MacKay, I.; Oostra, B.; van Duijn, C.M.; Aulchenko, Y.S. The Effect of Genetic Drift in a Young Genetically Isolated Population. *Ann. Hum. Genet.* **2005**, *69*, 288–295. [CrossRef] [PubMed]
98. Beita, C.M.; Murillo, L.F.S.; Alvarado, L.D.A. Ecological Corridors in Costa Rica: An Evaluation Applying Landscape Structure, Fragmentation-Connectivity Process, and Climate Adaptation. *Conserv. Sci. Pract.* **2021**, *3*, e475. [CrossRef]
99. Weeks, T.L.; Betts, M.G.; Pfeifer, M.; Wolf, C.; Banks-Leite, C.; Barbaro, L.; Barlow, J.; Cerezo, A.; Kennedy, C.M.; Kormann, U.G.; et al. Climate-Driven Variation in Dispersal Ability Predicts Responses to Forest Fragmentation in Birds. *Nat. Ecol. Evol.* **2023**, *7*, 1079–1091. [CrossRef]
100. Bouzat, J.L. Conservation Genetics of Population Bottlenecks: The Role of Chance, Selection, and History. *Conserv. Genet.* **2010**, *11*, 463–478. [CrossRef]
101. Vásquez, C.; Quiñones, R.A.; Brante, A.; Hernández-Miranda, E.; Vásquez, C.; Quiñones, R.A.; Brante, A.; Hernández-Miranda, E. Genetic Diversity and Resilience in Benthic Marine Populations. *Rev. Chil. Hist. Nat.* **2023**, *96*, 4. [CrossRef]
102. Ceballos, G.; Ehrlich, P.R.; Barnosky, A.D.; García, A.; Pringle, R.M.; Palmer, T.M. Accelerated Modern Human-Induced Species Losses: Entering the Sixth Mass Extinction. *Sci. Adv.* **2015**, *1*, e1400253. [CrossRef]
103. Christie, M.R.; Knowles, L.L. Habitat Corridors Facilitate Genetic Resilience Irrespective of Species Dispersal Abilities or Population Sizes. *Evol. Appl.* **2015**, *8*, 454–463. [CrossRef]
104. Sun, J.; Tummers, J.S.; Galib, S.M.; Lucas, M.C. Fish Community and Abundance Response to Improved Connectivity and More Natural Hydromorphology in a Post-Industrial Subcatchment. *Sci. Total Environ.* **2022**, *802*, 149720. [CrossRef]
105. Kuo, P.-H.; Shih, S.-S.; Otte, M.L. Restoration Recommendations for Mitigating Habitat Fragmentation of a River Corridor. *J. Environ. Manag.* **2021**, *296*, 113197. [CrossRef]
106. Friess, D.A.; Gatt, Y.M.; Ahmad, R.; Brown, B.M.; Sidik, F.; Wodehouse, D. Achieving Ambitious Mangrove Restoration Targets Will Need a Transdisciplinary and Evidence-Informed Approach. *One Earth* **2022**, *5*, 456–460. [CrossRef]
107. Banks-Leite, C.; Ewers, R.M.; Folkard-Tapp, H.; Fraser, A. Countering the Effects of Habitat Loss, Fragmentation, and Degradation through Habitat Restoration. *One Earth* **2020**, *3*, 672–676. [CrossRef]
108. Buelow, C.A.; Connolly, R.M.; Turschwell, M.P.; Adame, M.F.; Ahmadia, G.N.; Andradi-Brown, D.A.; Bunting, P.; Canty, S.W.J.; Dunic, J.C.; Friess, D.A.; et al. Ambitious Global Targets for Mangrove and Seagrass Recovery. *Curr. Biol.* **2022**, *32*, 1641–1649.e3. [CrossRef] [PubMed]
109. Hilderbrand, R.H.; Watts, A.C.; Randle, A.M. The Myths of Restoration Ecology. *Ecol. Soc.* **2005**, *10*, 1–11. [CrossRef]
110. Coleman, E.A.; Schultz, B.; Ramprasad, V.; Fischer, H.; Rana, P.; Filippi, A.M.; Güneralp, B.; Ma, A.; Rodriguez Solorzano, C.; Guleria, V.; et al. Limited Effects of Tree Planting on Forest Canopy Cover and Rural Livelihoods in Northern India. *Nat. Sustain.* **2021**, *4*, 997–1004. [CrossRef]
111. Waltham, N.J.; Elliott, M.; Lee, S.Y.; Lovelock, C.; Duarte, C.M.; Buelow, C.; Simenstad, C.; Nagelkerken, I.; Claassens, L.; Wen, C.K.-C.; et al. UN Decade on Ecosystem Restoration 2021–2030—What Chance for Success in Restoring Coastal Ecosystems? *Front. Mar. Sci.* **2020**, *7*, 00071. [CrossRef]
112. United Nations Decade on Ecosystem Restoration World Restoration Flagships. Available online: <https://www.decadeonrestoration.org/world-restoration-flagships> (accessed on 6 November 2025).
113. Hemraj, D.A.; Bishop, M.J.; Hancock, B.; Minuti, J.J.; Thurstan, R.H.; Zu Ermgassen, P.S.E.; Russell, B.D. Oyster Reef Restoration Fails to Recoup Global Historic Ecosystem Losses despite Substantial Biodiversity Gain. *Sci. Adv.* **2022**, *8*, eabp8747. [CrossRef] [PubMed]
114. Kodikara, K.A.S.; Mukherjee, N.; Jayatissa, L.P.; Dahdouh-Guebas, F.; Koedam, N. Have Mangrove Restoration Projects Worked? An in-Depth Study in Sri Lanka. *Restor. Ecol.* **2017**, *25*, 705–716. [CrossRef]
115. Palmer, A.M.; Stewart, A.G. Ecosystem Restoration Is Risky . . . but We Can Change That. *One Earth* **2020**, *3*, 661–664. [CrossRef]
116. Sasmito, S.D.; Basyuni, M.; Kridalaksana, A.; Saragi-Sasmito, M.F.; Lovelock, C.E.; Murdiyarso, D. Challenges and Opportunities for Achieving Sustainable Development Goals through Restoration of Indonesia’s Mangroves. *Nat. Ecol. Evol.* **2023**, *7*, 62–70. [CrossRef]
117. Waring, B.G. Grand Challenges in Ecosystem Restoration. *Front. Environ. Sci.* **2024**, *11*, 1353829. [CrossRef]
118. Samuel, E.M.; Mitchell, R.M.; Winkler, D.E. Perspectives on Challenges and Opportunities at the Restoration-Policy Interface in the U.S.A. *Restor. Ecol.* **2023**, *31*, e13823. [CrossRef]
119. Smith, C.S.; DeMattia, E.A.; Albright, E.; Bromberger, A.F.; Hayward, O.G.; Mackinson, I.J.; Mantell, S.A.; McAdoo, B.G.; McAfee, D.; McCollum, A.; et al. Beyond Despair: Leveraging Ecosystem Restoration for Psychosocial Resilience. *Proc. Natl. Acad. Sci. USA* **2025**, *122*, e2307082121. [CrossRef] [PubMed]
120. Verhoeven, D.; Berkhouwt, E.; Sewell, A.; van der Esch, S. The Global Cost of International Commitments on Land Restoration. *Land Degrad. Dev.* **2024**, *35*, 4864–4874. [CrossRef]

121. Bayraktarov, E.; Saunders, M.I.; Abdullah, S.; Mills, M.; Beher, J.; Possingham, H.P.; Mumby, P.J.; Lovelock, C.E. The Cost and Feasibility of Marine Coastal Restoration. *Ecol. Appl.* **2016**, *26*, 1055–1074. [CrossRef] [PubMed]
122. Mappin, B.; Ward, A.; Hughes, L.; Watson, J.E.M.; Cosier, P.; Possingham, H.P. The Costs and Benefits of Restoring a Continent’s Terrestrial Ecosystems. *J. Appl. Ecol.* **2022**, *59*, 408–419. [CrossRef]
123. Hobbs, R.J. Setting Effective and Realistic Restoration Goals: Key Directions for Research. *Restor. Ecol.* **2007**, *15*, 354–357. [CrossRef]
124. Chesapeake Progress Submerged Aquatic Vegetation (SAV). Available online: <https://marylandmatters.org/2025/11/03/sav-planting-chesapeake-bay-new-agreement/> (accessed on 18 January 2025).
125. Grayson, J.E.; Chapman, M.G.; Underwood, A.J. The Assessment of Restoration of Habitat in Urban Wetlands. *Landsc. Urban Plan.* **1999**, *43*, 227–236. [CrossRef]
126. Ellison, A.M.; Felson, A.J.; Friess, D.A. Mangrove Rehabilitation and Restoration as Experimental Adaptive Management. *Front. Mar. Sci.* **2020**, *7*, 00327. [CrossRef]
127. Mijangos, J.L.; Pacioni, C.; Spencer, P.B.S.; Craig, M.D. Contribution of Genetics to Ecological Restoration. *Mol. Ecol.* **2015**, *24*, 22–37. [CrossRef]
128. Nef, D.P.; Gotor, E.; Wiederkehr Guerra, G.; Zumwald, M.; Kettle, C.J. Initial Investment in Diversity Is the Efficient Thing to Do for Resilient Forest Landscape Restoration. *Front. For. Glob. Change* **2021**, *3*, 615682. [CrossRef]
129. Lovelock, C.E.; Barbier, E.; Duarte, C.M. Tackling the Mangrove Restoration Challenge. *PLoS Biol.* **2022**, *20*, e3001836. [CrossRef] [PubMed]
130. Ouyang, X.; Guo, F.; Lee, S.Y.; Yang, Z. Mangrove Restoration in China’s Tidal Ecosystems. *Science* **2024**, *385*, 836. [CrossRef] [PubMed]
131. Broadhurst, L.M.; Lowe, A.; Coates, D.J.; Cunningham, S.A.; McDonald, M.; Vesk, P.A.; Yates, C. Seed Supply for Broadscale Restoration: Maximizing Evolutionary Potential. *Evol. Appl.* **2008**, *1*, 587–597. [CrossRef] [PubMed]
132. Procaccini, G.; Piazzesi, L. Genetic Polymorphism and Transplantation Success in the Mediterranean Seagrass *Posidonia Oceanica*. *Restor. Ecol.* **2001**, *9*, 332–338. [CrossRef]
133. Marsden, B.W.; Ngeve, M.N.; Engelhardt, K.A.M.; Neel, M.C. Assessing the Potential to Extrapolate Genetic-Based Restoration Strategies Between Ecologically Similar but Geographically Separate Locations of the Foundation Species *Vallisneria Americana* Michx. *Estuaries Coasts* **2022**, *45*, 1656–1673. [CrossRef]
134. Ngeve, M.N.; Triest, L. Planted Mangroves Reflect Low Genetic Diversity of Natural Stands in Southern Cameroon. *For. Ecol. Manag.* **2026**, *601*, 123318. [CrossRef]
135. Zeng, X.; Fischer, G.A. Using Multiple Seedlots in Restoration Planting Enhances Genetic Diversity Compared to Natural Regeneration in Fragmented Tropical Forests. *For. Ecol. Manag.* **2021**, *482*, 118819. [CrossRef]
136. Ngeve, M.N.; Koedam, N.; Triest, L. Genotypes of *Rhizophora* Propagules from a Non-Mangrove Beach Provide Evidence of Recent Long-Distance Dispersal. *Front. Conserv. Sci.* **2021**, *2*, 65. [CrossRef]
137. Jeunen, G.-J.; Lipinskaya, T.; Gajduchenko, H.; Golovenchik, V.; Moroz, M.; Rizevsky, V.; Semenchenko, V.; Gemmell, N.J. Environmental DNA (eDNA) Metabarcoding Surveys Show Evidence of Non-Indigenous Freshwater Species Invasion to New Parts of Eastern Europe. *Metabarcoding Metagenom.* **2022**, *6*, e68575. [CrossRef]
138. Wiggins, J.J.; Tobias, V.D.; Holcombe, E.F.; Karpenko, K.; Huber, E.R.; Goodman, A.C. Leveraging Environmental DNA (eDNA) to Optimize Targeted Removal of Invasive Fishes. *J. Freshw. Ecol.* **2024**, *39*, 2378841. [CrossRef]
139. Gabur, I.; Simioniu, D.P.; Snowdon, R.J.; Cristea, D. Machine Learning Applied to the Search for Nonlinear Features in Breeding Populations. *Front. Artif. Intell.* **2022**, *5*, 876578. [CrossRef] [PubMed]
140. Lachmuth, S.; Capblancq, T.; Prakash, A.; Keller, S.R.; Fitzpatrick, M.C. Novel Genomic Offset Metrics Integrate Local Adaptation into Habitat Suitability Forecasts and Inform Assisted Migration. *Ecol. Monogr.* **2024**, *94*, e1593. [CrossRef]
141. Marsden, B.W.; Engelhardt, K.A.M.; Neel, M.C. Genetic Rescue versus Outbreeding Depression in *Vallisneria Americana*: Implications for Mixing Seed Sources for Restoration. *Biol. Conserv.* **2013**, *167*, 203–214. [CrossRef]
142. Jones, T.A. When Local Isn’t Best. *Evol. Appl.* **2013**, *6*, 1109–1118. [CrossRef]
143. Madsen, T.; Stille, B.; Shine, R. Inbreeding Depression in an Isolated Population of Adders *Vipera berus*. *Biol. Conserv.* **1996**, *75*, 113–118. [CrossRef]
144. Madsen, T.; Loman, J.; Anderberg, L.; Anderberg, H.; Georges, A.; Ujvari, B. Genetic Rescue Restores Long-Term Viability of an Isolated Population of Adders (*Vipera berus*). *Curr. Biol.* **2020**, *30*, R1297–R1299. [CrossRef] [PubMed]
145. Griffith, B.; Scott, J.M.; Carpenter, J.W.; Reed, C. Translocation as a Species Conservation Tool: Status and Strategy. *Science* **1989**, *245*, 477–480. [CrossRef]
146. Morris, S.D.; Brook, B.W.; Moseby, K.E.; Johnson, C.N. Factors Affecting Success of Conservation Translocations of Terrestrial Vertebrates: A Global Systematic Review. *Glob. Ecol. Conserv.* **2021**, *28*, e01630. [CrossRef]
147. Sherman, J.; Menon, V.; Kock, R.; King, T.; Luz, S.; Ashraf, N.V.K.; Soorae, P.S.; Moehrenschlager, A. *IUCN Guidelines on Responsible Translocation of Displaced Organisms*; IUCN: Gland, Switzerland, 2025; ISBN 978-2-8317-2333-4.

148. Hogg, C.J.; McLennan, E.A.; Wise, P.; Lee, A.V.; Pemberton, D.; Fox, S.; Belov, K.; Grueber, C.E. Preserving the Demographic and Genetic Integrity of a Single Source Population during Multiple Translocations. *Biol. Conserv.* **2020**, *241*, 108318. [[CrossRef](#)]
149. Mitchell, W.F.; Boulton, R.L.; Sunnucks, P.; Clarke, R.H. Are We Adequately Assessing the Demographic Impacts of Harvesting for Wild-Sourced Conservation Translocations? *Conserv. Sci. Pract.* **2022**, *4*, e569. [[CrossRef](#)]
150. Tenhumberg, B.; Tyre, A.J.; Shea, K.; Possingham, H.P. Linking Wild and Captive Populations to Maximize Species Persistence: Optimal Translocation Strategies. *Conserv. Biol.* **2004**, *18*, 1304–1314. [[CrossRef](#)]
151. Lubow, B.C. Optimal Translocation Strategies for Enhancing Stochastic Metapopulation Viability. *Ecol. Appl.* **1996**, *6*, 1268–1280. [[CrossRef](#)]
152. Pacioni, C.; Wayne, A.F.; Page, M. Guidelines for Genetic Management in Mammal Translocation Programs. *Biol. Conserv.* **2019**, *237*, 105–113. [[CrossRef](#)]
153. Rout, T.M.; Hauser, C.E.; Possingham, H.P. Minimise Long-Term Loss or Maximise Short-Term Gain? Optimal Translocation Strategies for Threatened Species. *Ecol. Model.* **2007**, *201*, 67–74. [[CrossRef](#)]
154. Menges, E.S. Restoration Demography and Genetics of Plants: When Is a Translocation Successful? *Aust. J. Bot.* **2008**, *56*, 187–196. [[CrossRef](#)]
155. Lewis, J.C.; Jenkins, K.J.; Happe, P.J.; Manson, D.J.; Griffin, P.C. Post-Release Survival of Translocated Fishers: Implications for Translocation Success. *J. Wildl. Manag.* **2022**, *86*, e22192. [[CrossRef](#)]
156. Sheller, F.J.; Fagan, W.F.; Unmack, P.J. Using Survival Analysis to Study Translocation Success in the Gila Topminnow (*Poeciliopsis occidentalis*). *Ecol. Appl.* **2006**, *16*, 1771–1784. [[CrossRef](#)]
157. Meyerpetter, M.B.; Coates, P.S.; Mathews, S.R.; Lazenby, K.D.; Prochazka, B.G.; Dahlgren, D.K.; Delehanty, D.J. Brood Translocation Increases Post-Release Recruitment and Promotes Population Restoration of *Centrocercus Urophasianus* (*Greater sage-grouse*). *Ornithol. Appl.* **2024**, *126*, duae013. [[CrossRef](#)]
158. Van Rossum, F.; Le Pajolec, S.; Godé, C. Assessing Spatial Mating Patterns in Translocated Populations of *Campanula glomerata*. *Glob. Ecol. Conserv.* **2023**, *46*, e02548. [[CrossRef](#)]
159. Shier, D.M.; Fischer, J.N.; King, S.N.D.; Greggor, A.L.; Grether, G.F. Pre-Release Experience with a Heterospecific Competitor Increases Fitness of a Translocated Endangered Species. *Biol. Conserv.* **2025**, *307*, 111193. [[CrossRef](#)]
160. Warne, R.K.; Chaber, A.-L. Assessing Disease Risks in Wildlife Translocation Projects: A Comprehensive Review of Disease Incidents. *Animals* **2023**, *13*, 3379. [[CrossRef](#)]
161. Novak, B.J.; Phelan, R.; Weber, M.U.S. Conservation Translocations: Over a Century of Intended Consequences. *Conserv. Sci. Pract.* **2021**, *3*, e394. [[CrossRef](#)]
162. Van Rossum, F.; Destombes, A.; Raspé, O. Are Large Census-Sized Populations Always the Best Sources for Plant Translocations? *Restor. Ecol.* **2021**, *29*, e13316. [[CrossRef](#)]
163. Yackulic, C.B.; Van Haverbeke, D.R.; Dzul, M.; Bair, L.; Young, K.L. Assessing the Population Impacts and Cost-Effectiveness of a Conservation Translocation. *J. Appl. Ecol.* **2021**, *58*, 1602–1612. [[CrossRef](#)]
164. Whiteley, A.R.; Fitzpatrick, S.W.; Funk, W.C.; Tallmon, D.A. Genetic Rescue to the Rescue. *Trends Ecol. Evol.* **2015**, *30*, 42–49. [[CrossRef](#)]
165. Onorato, D.P.; Cunningham, M.W.; Lotz, M.; Criffield, M.; Shindle, D.; Johnson, A.; Clemons, B.C.F.; Shea, C.P.; Roelke-Parker, M.E.; Johnson, W.E.; et al. Multi-Generational Benefits of Genetic Rescue. *Sci. Rep.* **2024**, *14*, 17519. [[CrossRef](#)]
166. Frankham, R.; Ballou, J.D.; Eldridge, M.D.B.; Lacy, R.C.; Ralls, K.; Dudash, M.R.; Fenster, C.B. Predicting the Probability of Outbreeding Depression. *Conserv. Biol.* **2011**, *25*, 465–475. [[CrossRef](#)]
167. Kelly, E.; Jolly, C.J.; Indigo, N.; Smart, A.; Webb, J.; Phillips, B. Kenbi Traditional Owners and Rangers No Outbreeding Depression in a Trial of Targeted Gene Flow in an Endangered Australian Marsupial. *Conserv. Genet.* **2021**, *22*, 23–33. [[CrossRef](#)]
168. Walsh, G.; McMahon, B.J.; Thörn, F.; Rödin-Mörch, P.; Irestedt, M.; Höglund, J. Inbreeding or Outbreeding Depression? How to Manage an Endangered and Locally Adapted Population of Red Grouse *Lagopus scotica*. *bioRxiv* **2023**, bioRxiv:2023.08.14.552414. [[CrossRef](#)]
169. Grummer, J.A.; Booker, T.R.; Matthey-Doret, R.; Nietlisbach, P.; Thomaz, A.T.; Whitlock, M.C. The Immediate Costs and Long-Term Benefits of Assisted Gene Flow in Large Populations. *Conserv. Biol.* **2022**, *36*, e13911. [[CrossRef](#)] [[PubMed](#)]
170. Funk, W.C.; McKay, J.K.; Hohenlohe, P.A.; Allendorf, F.W. Harnessing Genomics for Delineating Conservation Units. *Trends Ecol. Evol.* **2012**, *27*, 489–496. [[CrossRef](#)]
171. Forester, B.R.; Lasky, J.R.; Wagner, H.H.; Urban, D.L. Comparing Methods for Detecting Multilocus Adaptation with Multivariate Genotype–Environment Associations. *Mol. Ecol.* **2018**, *27*, 2215–2233. [[CrossRef](#)]
172. Piaggio, A.J.; Segelbacher, G.; Seddon, P.J.; Alphey, L.; Bennett, E.L.; Carlson, R.H.; Friedman, R.M.; Kanavy, D.; Phelan, R.; Redford, K.H.; et al. Is It Time for Synthetic Biodiversity Conservation? *Trends Ecol. Evol.* **2017**, *32*, 97–107. [[CrossRef](#)]
173. van Oppen, M.J.H.; Oliver, J.K.; Putnam, H.M.; Gates, R.D. Building Coral Reef Resilience through Assisted Evolution. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, 2307–2313. [[CrossRef](#)] [[PubMed](#)]

174. Kosch, T.A.; Waddle, A.W.; Cooper, C.A.; Zenger, K.R.; Garrick, D.J.; Berger, L.; Skerratt, L.F. Genetic Approaches for Increasing Fitness in Endangered Species. *Trends Ecol. Evol.* **2022**, *37*, 332–345. [CrossRef] [PubMed]
175. Samuel, M.D.; Liao, W.; Atkinson, C.T.; LaPointe, D.A. Facilitated Adaptation for Conservation—Can Gene Editing Save Hawaii’s Endangered Birds from Climate Driven Avian Malaria? *Biol. Conserv.* **2020**, *241*, 108390. [CrossRef]
176. Kardos, M.; Shafer, A.B.A. The Peril of Gene-Targeted Conservation. *Trends Ecol. Evol.* **2018**, *33*, 827–839. [CrossRef] [PubMed]
177. Hedrick, P.W. Conservation Genetics: Where Are We Now? *Trends Ecol. Evol.* **2001**, *16*, 629–636. [CrossRef]
178. Williams, S.E.; Hoffman, E.A. Minimizing Genetic Adaptation in Captive Breeding Programs: A Review. *Biol. Conserv.* **2009**, *142*, 2388–2400. [CrossRef]
179. Robert, A. Captive Breeding Genetics and Reintroduction Success. *Biol. Conserv.* **2009**, *142*, 2915–2922. [CrossRef]
180. Doudna, J.A.; Charpentier, E. The New Frontier of Genome Engineering with CRISPR-Cas9. *Science* **2014**, *346*, 1258096. [CrossRef]
181. Meek, M.H.; Beever, E.A.; Barbosa, S.; Fitzpatrick, S.W.; Fletcher, N.K.; Mittan-Moreau, C.S.; Reid, B.N.; Campbell-Staton, S.C.; Green, N.F.; Hellmann, J.J. Understanding Local Adaptation to Prepare Populations for Climate Change. *BioScience* **2023**, *73*, 36–47. [CrossRef]
182. Brodie, J.F.; Emmel, A.; Wiedenheft, B.; Sandler, R.L.; Redford, K.H.; Schultz, C.A.; Moehrenschlager, A.; Mark-Shabbolt, M.; Kamau, W.S.; Helm, J.E.; et al. Synthetically Assisted Conservation and the Application of Emerging Biological Technologies for the Protection of Biodiversity. *Conserv. Lett.* **2025**, *18*, e13114. [CrossRef]
183. Kumar, K.; Gambhir, G.; Dass, A.; Tripathi, A.K.; Singh, A.; Jha, A.K.; Yadava, P.; Choudhary, M.; Rakshit, S. Genetically Modified Crops: Current Status and Future Prospects. *Planta* **2020**, *251*, 91. [CrossRef] [PubMed]
184. Redford, K.H.; Adams, W.; Carlson, R.; Mace, G.M.; Ceccarelli, B. Synthetic Biology and the Conservation of Biodiversity. *Oryx* **2014**, *48*, 330–336. [CrossRef]
185. Cleves, P.A.; Tinoco, A.I.; Bradford, J.; Perrin, D.; Bay, L.K.; Pringle, J.R. Reduced Thermal Tolerance in a Coral Carrying CRISPR-Induced Mutations in the Gene for a Heat-Shock Transcription Factor. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 28899–28905. [CrossRef]
186. Kosch, T.A.; Silva, C.N.S.; Brannelly, L.A.; Roberts, A.A.; Lau, Q.; Marantelli, G.; Berger, L.; Skerratt, L.F. Genetic Potential for Disease Resistance in Critically Endangered Amphibians Decimated by Chytridiomycosis. *Anim. Conserv.* **2019**, *22*, 238–250. [CrossRef]
187. Newhouse, A.E.; Powell, W.A. Intentional Introgression of a Blight Tolerance Transgene to Rescue the Remnant Population of American Chestnut. *Conserv. Sci. Pract.* **2021**, *3*, e348. [CrossRef]
188. The American Chestnut Foundation. Available online: <https://tafc.org/darling-58/> (accessed on 9 December 2025).
189. Waltz, E. Gene-Edited CRISPR Mushroom Escapes US Regulation. *Nature* **2016**, *532*, 293. [CrossRef] [PubMed]
190. Lou, R.N.; Jacobs, A.; Wilder, A.P.; Therkildsen, N.O. A Beginner’s Guide to Low-Coverage Whole Genome Sequencing for Population Genomics. *Mol. Ecol.* **2021**, *30*, 5966–5993. [CrossRef] [PubMed]
191. Verrico, B.M.; Capblancq, T.; Fitzpatrick, M.C.; Keller, S.R. Reciprocal Evaluation of Genomic Offset Predictions of Climate Maladaptation with Independent Empirical Datasets. *Am. Nat.* **2026**. [CrossRef]
192. Pearse, D.E. Saving the Spandrels? Adaptive Genomic Variation in Conservation and Fisheries Management. *J. Fish Biol.* **2016**, *89*, 2697–2716. [CrossRef]
193. Kardos, M.; Husby, A.; McFarlane, S.E.; Qvarnström, A.; Ellegren, H. Whole-Genome Resequencing of Extreme Phenotypes in Collared Flycatchers Highlights the Difficulty of Detecting Quantitative Trait Loci in Natural Populations. *Mol. Ecol. Resour.* **2016**, *16*, 727–741. [CrossRef]
194. Csilléry, K.; Rodríguez-Verdugo, A.; Rellstab, C.; Guillaume, F. Detecting the Genomic Signal of Polygenic Adaptation and the Role of Epistasis in Evolution. *Mol. Ecol.* **2018**, *27*, 606–612. [CrossRef]
195. Hartfield, M.; Glémén, S. Hitchhiking of Deleterious Alleles and the Cost of Adaptation in Partially Selfing Species. *Genetics* **2014**, *196*, 281–293. [CrossRef] [PubMed]
196. Schwartz, M.K.; Dunn, S.L.; Gendron, W.A.C.; Helm, J.E.; Kamau, W.S.; Mark-Shabbolt, M.; Moehrenschlager, A.; Redford, K.H.; Russell, G.; Sandler, R.L.; et al. Principles for Introducing New Genes and Species for Conservation. *Trends Ecol. Evol.* **2025**, *40*, 296–307. [CrossRef]
197. Redford, K.; Brooks, T.; Macfarlane, N.B.W.; Adams, J.S.; Alphey, L.; Bennet, E.; Delborne, J.; Eggermont, H.; Esvelt, K.; KinGirl, A.; et al. *Genetic Frontiers for Conservation: An Assessment of Synthetic Biology and Biodiversity Conservation*; Technical Assessment; IUCN: Gland, Switzerland, 2019; p. 166.
198. IUCN Agrees First Global Policy on Synthetic Biology—News | IUCN. Available online: <https://iucn.org/news/202510/iucn-agrees-first-global-policy-synthetic-biology> (accessed on 11 December 2025).
199. Hoban, S.; Archer, F.I.; Bertola, L.D.; Bragg, J.G.; Breed, M.F.; Bruford, M.W.; Coleman, M.A.; Ekblom, R.; Funk, W.C.; Grueber, C.E.; et al. Global Genetic Diversity Status and Trends: Towards a Suite of Essential Biodiversity Variables (EBVs) for Genetic Composition. *Biol. Rev.* **2022**, *97*, 1511–1538. [CrossRef]

200. Flanagan, S.P.; Forester, B.R.; Latch, E.K.; Aitken, S.N.; Hoban, S. Guidelines for Planning Genomic Assessment and Monitoring of Locally Adaptive Variation to Inform Species Conservation. *Evol. Appl.* **2018**, *11*, 1035–1052. [[CrossRef](#)]
201. Stem, C.; Margoluis, R.; Salafsky, N.; Brown, M. Monitoring and Evaluation in Conservation: A Review of Trends and Approaches. *Conserv. Biol.* **2005**, *19*, 295–309. [[CrossRef](#)]
202. Milt, A.W.; Diebel, M.W.; Doran, P.J.; Ferris, M.C.; Herbert, M.; Khoury, M.L.; Moody, A.T.; Neeson, T.M.; Ross, J.; Treska, T.; et al. Minimizing Opportunity Costs to Aquatic Connectivity Restoration While Controlling an Invasive Species. *Conserv. Biol.* **2018**, *32*, 894–904. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher’s Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.