

Package ‘batman’

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Title Bayesian AuTomated Metabolite Analyser for NMR spectra

Description BATMAN deconvolves resonance peaks from NMR spectra and obtain concentration estimates for the corresponding metabolites automatically.

Depends R (>= 2.15.0), doSNOW, foreach, iterators, snow, utils, plotrix

License GPL-2

LazyLoad yes

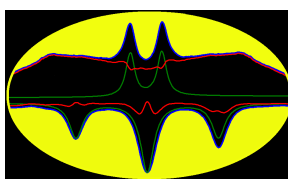
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batman-package	<i>Bayesian AuTomed Metabolite Analyser for NMR spectra (BAT-MAN)</i>
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Description

BATMAN deconvolves resonance peaks from NMR spectra of complex mixtures and obtains concentration estimates for the corresponding metabolites automatically. This is achieved through a database of spectral profiles for known metabolites and a Bayesian Markov Chain Monte Carlo algorithm. Users have the options to specify the multiplet ppm position, position shift range, peak width range and so on. Parallel processing is available if processing several spectra.



Details

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References

Astle W et al. (2012) A Bayesian Model of NMR Spectra for the Deconvolution and Quantification of Metabolites in Complex Biological Mixtures. Journal of the American Statistical Association, Vol. 107, Iss. 500, 2012 http://arxiv.org/PS_cache/arxiv/pdf/1105/1105.2204v1.pdf

Hao J et al. (2012) BATMAN-an R package for the automated quantification of metabolites from NMR spectra using a Bayesian Model. BIOINFORMATICS, 2012;28(15):2088-2090.

<http://arxiv.org/ftp/arxiv/papers/1112/1112.5794.pdf>

batman

*Perform BATMAN and Plot Analysis Result***Description**

The main function, it performs metabolite and wavelet fitting to input NMR spectra, plots fitting results, posterior distributions for relative concentrations and peak positions, and saves output. If the input `createDir = TRUE`, a folder name "runBATMAN" will be created in specified directory, within which, two folders "BatmanInput" and "BatmanOutput" are created. "BatmanInput" contains the input data files copied from installed package folder "extdata". The user only needs to modify files in this folder to change the settings for running batman. The batman output files are saved in "BatmanOutput" subfolders.

Usage

```
batman(BrukerDataDir, BrukerDataZipDir, txtFile, rData, createDir = TRUE,
       runBATMANDir = getwd(), overwriteDir = FALSE,
       figBatmanFit = TRUE, listMeta = FALSE,
       figRelCon = FALSE, figMetaFit = FALSE)
```

Arguments

- | | |
|------------------|---|
| BrukerDataDir | The directory of the folder containing 1D Bruker spectral data files. If not specified, spectral data will be read in from one of the following inputs prioritized in the order: BrukerDataZipDir, txtFile, rData and NMRdata.txt in "BatmanInput" folder. |
| BrukerDataZipDir | The directory of the folder containing zipped 1D Bruker spectral data files. If not specified, spectral data will be read in from one of the following inputs prioritized in the order: BrukerDataDir, txtFile, rData and NMRdata.txt in "BatmanInput" folder. |
| txtFile | The .txt file containing spectral data in the format of first column ppm, and the second column the real part of spectrum. If not specified, spectral data will be read in from one of the following inputs prioritized in the order: BrukerDataDir, BrukerDataZipDir, rData and NMRdata.txt in "BatmanInput" folder. |
| rData | The R data file containing spectral data in the format of first column ppm, and the second column the real part of spectrum. If not specified, spectral data will be read in from one of the following inputs prioritized in the order: BrukerDataDir, BrukerDataZipDir, txtFile and NMRdata.txt in "BatmanInput" folder. |
| createDir | If set TRUE, a new BATMAN work directory will be created specified by runBATMANDir. If set FALSE, batman input will be obtained from the "extdata" folder in batman package installation directory, and the batman output files will also be put within this folder. The default is TRUE. |
| runBATMANDir | User specified BATMAN work directory, the default is current work directory. It will only work when createDir is set TRUE. |
| overwriteDir | If folder "runBATMAN" exists, set TRUE to overwrite folder. The default is FALSE. |
| figBatmanFit | Plot metabolites and wavelets fit if set TRUE. The default is TRUE. |

listMeta	Individual metabolite fit will also be shown in the plot if set TRUE. The default is FALSE.
figRelCon	Plot posterior samples of the relative concentration for fitted metabolites with 95% credible interval if set TRUE. The default is FALSE.
figMetaFit	If set TRUE, plot the posterior mean of the metabolites fit with 95% credible interval. The default is FALSE.

Value

It returns a data list with the following objects:

specTitle	A matrix ($2 \times n$) containing the spectrum number in its first row and the corresponding title of the spectrum in its second row.
sFit	A matrix $t \times 5n$ of BATMAN fit results (down sampled). For 1 spectrum, it is a matrix with 5 columns: $[ppm, originalspectrum, metabolitesfit, waveletsfit, overallfit]$. The "overall fit" is the posterior mean of the BATMAN fit results after MCMC burn in iterations. Certain numbers of burn in iterations are used at the beginning of an MCMC run for finding a good starting point. n is the number of spectra, and t is the number of data points in each spectrum.
sFitHR	A matrix $t \times 3n$ of BATMAN fit results in the original resolution (without down sample). For 1 spectrum, it is a matrix with 3 columns: $[ppm, originalspectrum, metabolitesfit]$. n is the number of spectra, and t is the number of data points (without down sample) in each spectrum.
beta	A matrix ($m \times n$) containing the posterior means of relative concentrations for m fitted metabolites and n spectra after burn in.
betaSam	A matrix ($m \times (s * n)$) containing (for the first spectrum) s posterior samples of the relative concentrations in its rows. m is the number of fitted metabolites. n is the number of spectra analyzed. The subsequent columns contain the same format of data for the rest $n - 1$ spectra.
betaCI	A matrix ($m \times 2n$) containing the 95% credible interval of the relative concentrations for m fitted metabolites. Every pair of columns is for one spectrum.
metaTemp	A matrix ($t \times (m * n)$) containing the posterior means of m fitted metabolite templates in its columns (down sampled) after burn in. n is the number of spectra analyzed and t is the number of data points in each spectrum.
metaTempHR	A matrix ($t \times (m * n)$) containing the posterior means of m fitted metabolite templates in its columns (without down sample) after burn in. n is the number of spectra analyzed and t is the number of data points (without down sample) in each spectrum.
metaFitSam	A matrix ($t \times (s * n)$) containing s posterior samples of total metabolites fit during MCMC iterations in its columns. n is the number of spectra analyzed and t is the number of data points in each spectrum. The remaining $n - 1$ spectra metabolites fit results are saved in the same sequence in subsequent columns.
metaIndFitSam	A matrix ($t \times (m * s * n)$) containing s posterior samples of m individual metabolites fit during MCMC iterations in its columns. n is the number of spectra analyzed and t is the number of data points in each spectrum. The remaining $n - 1$ spectra results are saved in the same sequence in subsequent columns.

thetaSam	A matrix ($t \times (s \times n)$) containing s samples of wavelet fit during MCMC iterations in its columns. n is the number of spectra analyzed. The remaining $n - 1$ spectra wavelet fit results are saved in the same sequence in subsequent columns.
delta	A matrix ($M \times n$) containing posterior means of M multiplets ppm shift of fitted metabolites in its rows. M is the sum of all multiplets in the fitted metabolites. Each column of the matrix corresponds to one spectrum. If only 1 spectrum is analyzed, delta is a column vector.
deltaSam	A matrix ($s \times (M \times n)$) containing the posterior samples of multiplets ppm shift. Every M columns correspond the shift posterior samples of M multiplets for one spectrum. M is the sum of all multiplets in the fitted metabolites and n is the number of spectra analyzed.
outputDir	The directory of output folder with all the output result files.

See Also

[readBatmanOutput](#), [batmanrerun](#)

Examples

```
library(batman)
## Run BATMAN
if(interactive()) bm<-batman()
## This will create the folder "runBATMAN" in current working directory,
## within the folder "runBATMAN", a subfolder "BatmanInput" contains all the
## input files batman uses. Users can modify "metabolitesList.csv",
## "batmanOptions.txt" and so on to change the settings of batman.
## Please check "BatmanInput" for details on how to adjust input parameters.
#####
## The following is an example of what will be displayed in R
## and what value the user could input:
#####
## batman...
## Enter number of post-burn-in iterations (burn-in currently set to
## 400 iterations):
## 1: 100 ## user input
##
## Enter a number of choice from the menu below:
##
## 1: Include the default template of multiplets in multi_data.csv file only.
## 2: Include the user input template of multiplets in multi_data_user.csv file only.
## 3: Include both the above files.
##
## Selection: 1 ## user input
## Loading multi_data.csv...
## Percentage completed...
## | | 0%
## Size of each spectrum is 393.
## Size of metabolite list is 22.
## Constructing chain data structure...
## time used is 0 seconds.
## Running MCMC...
## |=====| 80%
## time used for burnin is 76 seconds.
## |=====| 100%
## time used is 95 seconds.
```

```

## saving posteriors...
##
## time elapsed
## 95.61
## second.
## Reading in saved data in folder
## ../user_specified_dir/runBATMAN/BatmanOutput/07_Dec_17_19_18
## Completed.
#####
## Alternatively if more than 1 spectrum are included without using fixed effect
## (in batmanOptions.txt file, set
## "Same concentration for all spectra (fixed effect) (1/0): 0"),
## user will be asked to input the following parameter:
#####
## How many parallel processes (multicores) do you want to run
## the multi-spectra analysis?
## (Enter 1 for running them sequentially.)
##
## Parallel processing of multi spectra currently cannot display
## progress bar (or any words), if you input is > 1, please be patient
## for the results :)
##
## 1: 2 ## user input
## time elapsed
## 78.79
## second.
## Reading in saved data in folder
## ../user_specified_dir/runBATMAN/BatmanOutput/07_Dec_17_35_53
## Completed.
#####

```

Batman-Input

BATMAN Input Files are Explained Here

Description

batman gets input parameters and metabolite templates information from the input files explained here. The input files are in either folder ".../runBATMAN/BatmanInput" or folder "extdata" depending on batman arguments. The user can modify the parameter values in the following input files (do not change the name of these files): batmanOptions.txt, metabolitesList.csv, multi_data.csv, multi_data_user.csv, NMRdata.txt.

Arguments

batmanOptions.txt

Option file to be used by batman. A copy of this file in the output directory is used for batmanrerun. The parameters in batmanOptions.txt file are explained here with example input values. The parameters have to be listed in the particular order given here, and do not leave empty lines in between except beginning with the comment character "%". Please note that for version 1.0.9 and later, one more input line,

"Use specified chemical shift for spectra (chemShiftperSpectra.csv) file (1/0): 0",

is added at the end of this file. For earlier version users updated to this version, running batman will add the above input line at the end of the file if missing.

Include ppm ranges for analysis: (1.2, 1.6) (2.1, 2.8)

- Put each set of ppm range in a pair of parentheses in the same line, separate start and end ppm values with a comma, separate each set of ppm range with space. Note that, very small number of spectra variables may cause error in wavelet analysis, do not give very narrow ppm ranges and also check the "Down sampling:" factor below, which used together, may also left very small number of spectra variables.

Spectra range to be included: 1-3, 5

- Integer, if no. > 1 and fixed effect (same concentration for all spectra) is 0, user will be asked to choose whether to parallelize fittings between spectra when running batman or rerunbatman.

Lower limit for spectrum intensity: -0.5

- Spectrum intensity smaller than the lower limit will be replaced by the lower limit.

Normalisation factor: 20000

- The whole spectrum will be divided by the normalisation factor.

Down sampling: 3

- Integer, number of spectra variable will be reduced by the factor of the input parameter, 3, in this case. For the example shown, the spectra variables with the index 1 : 3 : end will be used for analysis.

Save metabolites fit same as the original spectrum resolution (1/0): 1

- Whether to save the metabolites fitting result in the original resolution without down sampling. Input 1 for yes, and 0 for no.

Set seed for random number generation: 25

- Random number generation seed, integer.

Stop burn in at iteration: 4000

- Integer, this is the number of burn in iterations. The number of iterations after burn in will be asked when running batman. The posterior samples will be saved in the frequency specified by the next parameter. If changing the range of spectrum causing fitting results inconsistent, this indicates that the burn in stage hasn't found the best chemical shift. User may need to increase burn in iterations or reduce prior truncation on ppm shift for each multiple (adjust parameter "rdelta" below).

Save results in every ? iterations: 5

- Integer, save posterior samples for every 5 iterations.

Same concentration for all spectra (fixed effect) (1/0): 0

- Whether all the input spectra have the same metabolite concentrations (e.g. technical replicates). Input 1 for yes, and 0 for no.

Rerun iterations: 5000

- Integer, this is the number of iterations for batmanrerun. The rerun will use fixed multiplets positions obtained from running batman. There is no burnin for batman rerun.

Start temperature: 1000

- Sets the start temperature parameter of the likelihood of tempering. Higher temperature may need more burnin iterations to cool down.

Spectrometer frequency (MHz): 600

- Spectrometer used to collect the spectrum.

a: 0.00001

b: 0.000000001

- Hyper parameters for the global precision priors ($\lambda \sim \text{Gamma}(a, b/2)$) on wavelet coefficients.

Mean of the prior on mu: 0

Variance of the prior on mu: 0.1

Proposal variance for the Metropolis-Hastings sampler for mu: 0.002

Variance of each of the priors on the nu_m: 0.0025

Proposal variance for the Metropolis-Hastings sampler for each nu_m: 0.0001

- For peak width, γ , in Hz of metabolite m , the model for γ is $\ln(\gamma) = \mu + \nu_m$ where μ is the spectrum wide average log-peakwidth and ν_m is a random effect on metabolite deviation from μ . The mean of each prior on ν_m is 0. Set the variance of the prior on ν_m to 0 to turn off the random effect on peak width to keep peaks at the same width. The user can keep the proposal variance parameters unchanged for most of the case.

mean of the prior on tau: -0.01

- Hyper priors (τ) on negative wavelet coefficient (truncated normal). A more negative value means the wavelet fit will have more negative component.

steep: 2

- This parameter is inversely proportional to the variance of the prior on τ .

rdelta: 0.030

- Prior of the truncation on ppm shift for all multiplets, individual prior for each multiplet can be changed in the "multi_data.csv" file. Increase this parameter to allow multiplets to shift more. Please note, increasing this value may need more burn in iterations to find the best chemical shift for multiplets.

Use specified chemical shift for spectra (chemShiftperSpectra.csv) file (1/0): 0

- Input "1" to use file "chemShiftperSpectra.csv" to specify chemical shift per multiplet and per spectrum. Input "0" will not use that file. User can use the MATLAB tool "SplineFitBATMAN" provided to get more accurate chemical shift per spectra for each multiplet. This tool will save chemical shift information into "chemShiftperSpectra.csv".

metabolitesList.csv

List of metabolite names to be fitted. Put "%" in front of the metabolite name to comment out any metabolite for batman analysis.

multi_data.csv

Multiplet template parameters file, obtained from the online Human Metabolome Database (HMDB) version 2.5. The user can modify the parameters in the template file and specify ppm positions, and normal distribution truncation of ppm shift parameters (a positive value applied as +/- on the distribution).

The columns are:

Metabolite: The name of metabolite the multiplets belongs to.

pos_in_ppm: The ppm position of the multiplets.

couple_code: Coupling code. 0 = singlet, 1 = doublet, 2 = triplet, 3 = quartet, 4 = quintet, 5 = sextet, 6 = septet, 1,1 = doublet of doublets, 1,2 = doublet of triplets, 2,1 = triplet of doublets, 2,2 = triplet of triplets, 1,3 = doublet of quartets, 3,1 = quartet of doublets, 1,1,1 = doublet of doublet of doublets, 2,3 = triplet of quartets, 3,2 = quartet of triplets, 3,3 = quartet of quartets. If "-1" is inputted here, a user specified multiplet can be created. An example can be found in file "multi_data_user.csv". If "-2" is inputted here, a multiplet with range specified in ppm in the field "J_constant" is used. Examples can be found in file "multi_data_user.csv".

J_constant: J constant. If "-1" is inputted in the previous field "couple_code", J_constant/f are the offset of peaks from the multiplet position (f is the magnet frequency). Note that the spectra are shown in reverse ppm axis, so a positive offset means peak at higher ppm value, and a negative offset is peak at lower ppm value. If "-2" is inputted in the previous field "couple_code", the field here requires a two values input separated by comma, which specifies the ppm range of the multiplet in the pure spectrum. Note in this case, the field "Metabolite" name will also be the .txt file name containing the pure spectrum (refer to [createPureSpectraTemplate](#)).

no_of_protons: Number of protons in each multiplet. If "-1" is inputted for "couple_code", one or more (corresponding to J_constant) values can be given here as peak weights. The sum of no_of_protons is the number of protons in this multiplet.

overwrite_pos: The default is "n" for not overwrite position, and in that case the value in "pos_in_ppm" is used for each multiplet. If user want to use a different value from "pos_in_ppm", it should be put in this column.

overwrite_truncation: The default is "n", and the default truncation value is obtained from the user input truncation on ppm shift (rdelta) in batmanOptions.txt. If the user wants to use different truncations for specific multiplets, it should be put in this column. This value will be used to calculate the ppm shift variance value (truncation/5) for the corresponding multiplets.

Include_multiplet: The default is "1" and all multiplets belong to the listed metabolites will be used. Set to "0" to exclude certain multiplet(s) from listed metabolite(s).

multi_data_user.csv

Metabolite template parameters file for user to add new metabolites in the same format as multi_data.csv.

NMRdata.txt

The file has ppm value as its first column, and real part of the NMR spectrum in each of the subsequent columns. This file will be used when none of the input data argument is given.

Batman-Output

BATMAN Outputs are Explained Here

Description

batman and batmanrerun return the results as a data list with the objects described in their individual function. They also put results in .txt format in a folder named after the start execution time (*date_month_hours_mins_seconds*) within either folder ".../runBATMAN/BatmanOutput" or folder "extdata" depending on batman input createDir settings.

Value

batman and batmanrerun save their results in the following files in the output folder:

beta_i_rr_j.txt

A column vector ($m \times 1$) containing the estimated posterior mean of relative concentrations for m fitted metabolites of spectrum i . For batman results, j is 0, and for batmanrerun results, j is 1.

beta_sam_i_rr_j.txt

A matrix ($m \times s$) with each row containing the s posterior samples of the relative concentrations for one fitted metabolite of spectrum i . m is the total number of fitted metabolites. For batman results, j is 0, and for batmanrerun results, j is 1.

delta_draw_mean_i.txt

A column vector ($M \times 1$) containing the posterior mean of M multiplets ppm shift from the pre-set ppm position value in multi_data.csv or multi_data_user.csv of spectrum i .

delta_sam_i.txt

A matrix ($s \times M$) containing the posterior samples of M multiplets ppm shift. Every column correspond the shift posterior samples of one multiplet for spectrum i . M is the sum of all multiplets in the fitted metabolites.

L_i.txt

A matrix ($t \times M$) with each column as the template of one fitted metabolite for spectrum i before fitting. t is the number of data points in each spectrum.

lambda_sam_i_rr_j.txt

A column vector ($s \times 1$) containing s posterior samples of λ (a scalar global precision parameter) for spectrum i . For batman results, j is 0, and for batmanrerun results, j is 1.

metabolitesListUsed.txt

A column vector ($m \times 1$) containing the m metabolite names which have multiplets in/near the ppm region specified in batmanOptions.txt and used in the fitting.

metaFit_sam_i_rr_j.txt

A matrix ($t \times s$) containing s posterior samples of total metabolites fit during MCMC iterations in its columns for spectrum i . t is the number of data points in each spectrum. For batman results, j is 0, and for batmanrerun results, j is 1.

metaIndFit_sam_i_rr_j.txt

A matrix ($t \times (m \times s)$) containing s posterior samples of m individual metabolites fit in its columns for spectrum i . t is the number of data points in each spectrum. Every m columns are the m individual metabolite fit samples for one posterior sample. For batman results, j is 0, and for batmanrerun results, j is 1.

metaTemp_i_rr_j.txt

A matrix $t \times m$) containing the posterior means of m fitted metabolite templates in its columns (down sampled) after burn in for spectra i . t is the number of data points in each spectrum. For batman results, j is 0, and for batmanrerun results, j is 1.

metaTempHR_i_rr_j.txt

A matrix ($t \times m$) containing the posterior means of m fitted metabolite templates in its columns (without down sample) after burn in for spectra i . t is the number of data points (without down sample) in each spectrum. For batman results, j is 0, and for batmanrerun results, j is 1.

MultipletsPpmShifts.txt	A table ($M \times n$) containing the posterior means of multiplets ppm shift for M multiplets as its rows. M is the sum of all multiplets in the fitted metabolites and n is the number of spectra analyzed.
NMRdata_mod_ i .txt	A matrix ($t \times 2$) containing the input spectrum i in its original resolution. The first column is ppm value, and the second column is the i th spectrum intensity.
RelCon.txt	A table ($m \times n$) of the posterior means of relative concentrations for m fitted metabolites and n spectra.
RelConCreInt.txt	A table ($m \times 2n$) containing the 95% credible intervals (2.5% and 97.5%) for the relative concentrations of m fitted metabolites for n spectra.
specFit_ i _rr_ j .txt	A matrix ($t \times 5$) of BATMAN fit results with five columns as: [ppm, Originalspectrum, Metabolitesfit, Waveletfit, Overallsum] of spectrum i . For batman results, j is 0, and for batmanrerun results, j is 1.
specFitHR_ i _rr_ j .txt	A column vector ($t \times 1$) of metabolite fit result in the original resolution for spectrum i . t is the number of data points (without down sample) in each spectrum. For batman results, j is 0, and for batmanrerun results, j is 1.
theta_sam_ i _rr_ j .txt	A matrix ($t \times s$) containing s samples of wavelet fit during MCMC iterations in its columns for spectrum i . For batman results, j is 0, and for batmanrerun results, j is 1.
batmanOptions.txt	The same file copied from batman input. This file will be used by batmanrerun.
metabolitesList.txt	The same file copied from batman input.
NMRdata.txt	The same file copied from batman input.

If any plotting is performed, pdf files of the figure will be saved. For details, please refer to each plotting functions.

batmanrerun	<i>Perform BATMAN with Fixed (Previously Estimated) Multiplet Positions</i>
-------------	---

Description

This performs metabolite and wavelet fitting to input NMR spectra with fixed multiplet position obtained from running batman, and also plots fitting results. The user should modify parameters in the copy file "batmanOptions.txt" in batman output folder to change the rerun settings.

Usage

```
batmanrerun(BM, figBatmanFit = TRUE, listMeta = FALSE,
            figRelCon = FALSE, figMetaFit = FALSE)
```

Arguments

BM	batman output data frame.
figBatmanFit	Plot metabolites and wavelets fit if set TRUE. The default is TRUE.
listMeta	Individual metabolite fit will also be shown in the plot if set TRUE. The default is FALSE.
figRelCon	Plot posterior samples of the relative concentration for listed metabolites with 95% credible interval if set TRUE. The default is FALSE.
figMetaFit	If set TRUE, plot the posterior mean of the metabolites fit with 95% credible interval. The default is FALSE.

Value

When batmanrerun is called with multiplet ppm shifts fixed from the batman results, the following objects are added to the batman result:

sFitRerun	A matrix $t \times 5n$ of BATMAN rerun fit results (down sampled). For 1 spectrum, it is a matrix with 5 columns: $[ppm, originalspectrum, metabolitesfit, waveletsfit, overallfit]$. n is the number of spectra, and t is the number of data points in each spectrum.
sFitRerunHR	A matrix $t \times 3n$ of BATMAN rerun fit results in the original resolution (without down sample). For 1 spectrum, it is a matrix with 3 columns: $[ppm, originalspectrum, metabolitesfit]$. n is the number of spectra, and t is the number of data points (without down sample) in each spectrum.
betaRerun	For batman rerun, a matrix $(m \times n)$ containing the posterior means of relative concentrations for m fitted metabolites and n spectra.
betaSamRerun	For batman rerun, a matrix $(m \times (s * n))$ containing (for the first spectrum) s posterior samples of the relative concentrations in its rows. m is the number of fitted metabolites. n is the number of spectra analyzed. The subsequent columns contain the same data format for the rest $n - 1$ spectra.
betaCIRerun	For batman rerun, a matrix $(m \times 2n)$ containing the 95% credible interval of the relative concentrations for m fitted metabolites. Every pair of columns is for one spectrum.
metaTempRerun	For batman rerun, a matrix $(t \times (m * n))$ containing the posterior means of m fitted metabolite templates in its columns (down sampled). n is the number of spectra analyzed and t is the number of data points in each spectrum.
metaTempRerunHR	For batman rerun, a matrix $(t \times (m * n))$ containing the posterior means of m fitted metabolite templates in its columns (without down sample). n is the number of spectra analyzed and t is the number of data points (without down sample) in each spectrum.
metaFitSamRerun	For batman rerun, a matrix $(t \times (s * n))$ containing s posterior samples of total metabolites fit in its columns. n is the number of spectra analyzed and t is the number of data points in each spectrum. The remaining $n - 1$ spectra metabolites fit results are saved in the same sequence in subsequent columns.

metaIndFitSamRerun

For batman rerun, a matrix ($t \times (m * s * n)$) containing s posterior samples of m individual metabolites fit in its columns. n is the number of spectra analyzed and t is the number of data points in each spectrum. The remaining $n - 1$ spectra results are saved in the same sequence in subsequent columns.

thetaSamRerun For batman rerun, a matrix ($t \times (s * n)$) containing s samples of wavelet fit in its columns. n is the number of spectra analyzed. The remaining $n - 1$ spectra wavelet fit results are saved in the same sequence in subsequent columns.

outputDir The directory of output folder with all the output result files.

See Also

[batman](#), [readBatmanOutput](#)

Examples

```
library(batman)
## Run batman
if(interactive())
{
  bm<-batman()
  ## then call batmanrerun
  bm<-batmanrerun(bm)
}
#####
## The following is an example of what will be displayed in R
## and what value the user could input:
#####
## Rerunning batman for 500 iterations.
## percentage completed...
## | 0%
## Size of each spectrum is 382.
## Size of metabolite list is 22.
## Constructing chain data structure...
## time used is 1 seconds.
## Running MCMC...
## |=====| 100%
## time used is 65 seconds.
## saving posteriors...
##
## For rerun, time elapsed
## 65.96 seconds.
## Reading in saved data in folder
## ../user_specified_dir/runBATMAN/BatmanOutput/07_Dec_17_35_53
## Completed.
#####
## Alternatively if more than 1 spectrum are included without using fixed
## effect, user will be asked to input whether to parallelize the analysis
## between spectra.
#####
## How many parallel processes (multicores) do you want to run the
## multi-spectra analysis?
## (Enter 1 for running them sequentially.)
##
## Parallel processing of multi spectra currently cannot display
## the progress bar (or any words), if you input is > 1, please be patient
```

```
## for the results :)
##
## 1: 2 ## user input
##
## For rerun, time elapsed
## 64.4
## Reading in saved data in folder
## ../user_specified_dir/runBATMAN/BatmanOutput/07_Dec_17_35_53
## Completed.
#####
```

checkBatmanOptions	<i>Check batmanOptions.txt file for version later than 1.0.4</i>
--------------------	--

Description

Check batmanOptions.txt file and add a new line input at the end of the file.

Usage

```
checkBatmanOptions(dir)
```

Arguments

dir	The directory of batmanOptions.txt file.
-----	--

Examples

```
library(batman)
## createfolder "runBATMAN" in current working directory
batmanDir = newDir(runBATMANDir = getwd(), overwriteFile = TRUE)
checkBatmanOptions(dir = paste(batmanDir[2], "/batmanOptions.txt", sep = ""))
```

createChemShiftPerSpec	<i>Creating the file chemShiftPerSpec.csv which contains chemical shift parameters for all multiplets and spectra.</i>
------------------------	--

Description

This function creates a file called chemShiftPerSpec.csv, so user can specify chemical shift parameter for each spectrum and multiplet. The first column is multiplet names in the same order as the template inputs in multi_data.csv and/or multi_data_user.csv (depending on user choice of using one or both of them) file(s). The second column is the default chemical shift value (pos_in_ppm) for the corresponding multiplet. From the third column forward is the chemical shift value for each spectrum in the same order as they read in by BATMAN, if 'n' is present in the field, the default chemical shift value (or overwrite_pos value if given) will be used.

Usage

```
createChemShiftPerSpec(templateOption, dirIP)
```

Arguments

templateOption Choose template file(s). templateOption = 1 for multi_data.csv, templateOption = 2 for multi_data_user.csv, and templateOption = 3 for both files.

dirIP The input directory of BATMAN. This is the path ending with '/BatmanInput' if runBATMAN directory is created.

See Also

[batman](#)

Examples

```
library(batman)
## createfolder "runBATMAN" in current working directory
batmanDir = newDir(runBATMANDir = getwd(), overwriteFile = TRUE)
## create chemShiftPerSpec.csv
createChemShiftPerSpec(templateOption = 1, dirIP = batmanDir[2])
```

```
createPureSpectraTemplate
```

Creating a folder called 'PureSpectraTemplate' in the specified input directory. The folder contains pure metabolite spectrum template in .txt file with metabolite name as the file name.

Description

This function will read in pure metabolites spectra in Bruker format and save them in .txt format in folder "PureSpectraTemplate". The .txt file name is the same as the input to "metaNames". The "PureSpectraTemplate" folder will be used if "couple_code" value in multi_data.csv and/or multi_data_user.csv is set to '-2'.

Usage

```
createPureSpectraTemplate(dirPureSpec, metaNames, dirIP)
```

Arguments

dirPureSpec A vector containing the directories of Bruker pure metabolite spectra files.

metaNames The vector of metabolites names in the same order as the spectra directories in dirPureSpec.

dirIP The input directory of BATMAN. This is the path ending with '/runBATMAN/BatmanInput' if runBATMAN directory is created.

Examples

```
library(batman)
## createfolder "runBATMAN" in current working directory
batmanDir = newDir(runBATMANDir = getwd(), overwriteFile = TRUE)
## create chemShiftPerSpec.csv
createPureSpectraTemplate(dirPureSpec = c(paste(batmanDir[2], "/testBrukerPureSpec/testPure1", sep = ""),
paste(batmanDir[2], "/testBrukerPureSpec/testPure2", sep = "")), metaNames = c("testPure1", "testPure2"),
dirIP = batmanDir[2])
```

plotBatmanFit

*Plot Batman Metabolite Fit of NMR Spectra (With Down Sampling)***Description**

This function plots the BATMAN fit results, and saves the figure to pdf file in specified directory. For multiple spectra analysis, the file name is in the format of "specFit_itoj_metaName.pdf", where i and j are the range numbers of spectra in the figure and the metabolite name will be shown in place of *metaName* if supplied. Maximum of 2 spectra will be shown in each figure. The figure file will not be overwritten if it already exists. A prefix can be added to the file name for new saves.

Usage

```
plotBatmanFit(BM, xfrom, xto, yfrom, yto, listMeta = FALSE,
              metaName, saveFig = TRUE, saveFigDir = BM$outputDir,
              prefixFig, rerun = FALSE, placeLegend,
              plotColour, overwriteFig = FALSE)
```

Arguments

BM	batman output data frame.
xfrom	The start ppm value to plot. Default is set to the start ppm value of the whole processed range.
xto	The end ppm value to plot. Default is set to the end ppm value of the whole processed range.
yfrom	The start value of vertical axis to plot. Default is set to 0.
yto	The end value of vertical axis to plot. Default is set to the maximum value of the spectrum point in display.
listMeta	Individual metabolite fit will also be shown in the plot if set TRUE. The default is FALSE.
metaName	One or more specified metabolite fits will be shown in the plot. If no name was given and listMeta = TRUE, all the individual metabolite fit will be shown.
saveFig	Save figure(s) to pdf file(s) if set TRUE. The default is TRUE.
saveFigDir	Save figure(s) in this directory. The default is output directory of BM.
prefixFig	Add prefix to each saved figure name. The default is no prefix.
rerun	Set to FALSE to plot batman result, and TRUE to plot batmanrerun result.
placeLegend	Where to place the legend in figure. For example, placeLegend = "topright".
plotColour	User can specify colours for each metabolite if listMeta = TRUE. If not, a set of randomly generated colours will be used.
overwriteFig	Overwrite the saved figure file in pdf format if overwriteFig = TRUE. The default is FALSE.

See Also

[batman](#), [batmanrerun](#)

Examples

```
library(batman)
## Run BATMAN
if(interactive())
{
  bm<-batman()
  ## then plot results
  plotBatmanFit(bm)
}
```

plotBatmanFitHR	<i>Plot BATMAN Metabolite Fit of NMR Spectra in Original Resolution (Without Down Sampling)</i>
-----------------	---

Description

This function plots a high resolution BATMAN fit results (without down sampling), and save figure to pdf file in user specified directory. For multiple spectra analysis, the file name is in the format of "specFitHR_*i*_metaName.pdf", where *i* is the spectrum number in the figure and the metabolite name will be shown in place of *metaName* if supplied. The figure file will not be overwritten if it already exists. A prefix can be given to the file name for new saves.

Usage

```
plotBatmanFitHR(BM, xfrom, xto, yfrom, yto, metaName, saveFig = TRUE,
  saveFigDir = BM$outputDir, prefixFig, rerun = FALSE,
  overwriteFig = FALSE)
```

Arguments

BM	batman output data frame.
xfrom	The start ppm value to plot. Default is set to the start ppm value of the whole processed range.
xto	The end ppm value to plot. Default is set to the end ppm value of the whole processed range.
yfrom	The start value of vertical axis to plot. Default is set to 0.
yto	The end value of vertical axis to plot. Default is set to the maximum value of the spectrum point in display.
metaName	Individual metabolite fit will also be shown in the plot if a metabolite name is given. Only one metabolite name can be given, if missing from input all metabolites will be plotted.
saveFig	Save figure(s) to pdf file(s) if set TRUE. The default is TRUE.
saveFigDir	Save figure(s) in this directory. The default is the output directory of BM.
prefixFig	Add prefix to each saved figure name. The default is no prefix.
rerun	Set to FALSE to plot batman result, and TRUE to plot batmanrerun result.
overwriteFig	Overwrite the saved figure file in pdf format if overwriteFig = TRUE. The default is FALSE.

See Also

[batman](#), [batmanrerun](#)

Examples

```
library(batman)
## Run BATMAN fit
if(interactive())
{
  bm<-batman()
  ## Plot batman Fit in its original resolution if the option parameter
  ## is set to 1 for "Save metabolites fit same as the original spectrum
  ## resolution (1/0)" in "batmanOptions.txt", .
  plotBatmanFitHR(bm)
}
```

plotMetaFit

Plot Posterior Means of Metabolites Fit with 95% Credible Interval

Description

This function plots posterior means of the metabolite fit with 95% credible interval , and saves the figure to pdf file in specified directory. For multiple metabolites, the file name is in the format of "spec_itoj_mFitSam.pdf", where *i* and *j* are range numbers of spectra in the figure. A maximum of 2 spectra will be shown in each figure. Figure file will not be overwritten if it already exists. Prefix can be added to the file name for new saves.

Usage

```
plotMetaFit(BM, from, to, metaName, saveFig = TRUE,
             saveFigDir = BM$outputDir, prefixFig,
             rerun = FALSE, overwriteFig = FALSE)
```

Arguments

BM	batman output data frame.
from	The start ppm value to plot. Default is set to the start ppm value of the whole processed range.
to	The end ppm value to plot. Default is set to the end ppm value of the whole processed range.
metaName	Only multiplets belonging to the named Metabolite will be shown. Only one metabolite name can be given. If missing, all metabolites will be plotted.
saveFig	Save figure to pdf file if set TRUE. The default is TRUE.
saveFigDir	Save figure in this directory. The default is current working directory.
prefixFig	Add prefix to each saved figure name. The default is no prefix.
rerun	Set to FALSE to plot batman result, and TRUE to plot batmanrerun result.
overwriteFig	Overwrite the saved figure file in pdf format if overwriteFig = TRUE. The default is FALSE.

See Also

[batman](#), [batmanrerun](#)

Examples

```
library(batman)
## Run BATMAN fit, then plot metabolite fit
if(interactive())
{
  bm<-batman()
  ## Plot metabolites Fit.
  plotMetaFit(bm)
}
```

plotRelCon

Boxplot or Histogram of Posterior distributions of Relative Concentrations for Listed Metabolites with 95% Credible Interval

Description

This function plots the posterior distributions of relative concentrations, and saves the figure to pdf file. The file name is in the format of "spec_*i*_RelCon_*j1*to*j2*.pdf", where *i* are the spectrum numbers and *j1* and *j2* are the order numbers of fitted metabolites in the order of their input in file metaboliteList.csv. The figure file will not be overwritten if it already exists. A prefix can be added to file name for new saves.

Usage

```
plotRelCon(BM, metaName, plotHist = FALSE, breaks,
            saveFig = TRUE, saveFigDir = BM$outputDir,
            prefixFig, rerun = FALSE, overwriteFig = FALSE)
```

Arguments

BM	batman output data frame.
metaName	Only multiplets belonging to the named Metabolite will be shown. Only one metabolite name can be given. If missing, all metabolites will be plotted.
plotHist	If plotHist = TRUE, the ppm shift posteriors will be displayed as histogram. The default is FALSE.
breaks	A single number to set the number of bins for the histogram. If missing from the input, it is set to the data length divided by 3.
saveFig	Save figure(s) to pdf file(s) if set TRUE. The default is TRUE.
saveFigDir	Save figure(s) in this directory. The default is output directory of BM.
prefixFig	Add prefix to each saved figure name. The default is no prefix.
rerun	Set to FALSE to plot batman result, and TRUE to plot batmanrerun result.
overwriteFig	Overwrite the saved figure file in pdf format if overwriteFig = TRUE. The default is FALSE.

See Also

[batman](#), [batmanrerun](#)

Examples

```
library(batman)
## Run BATMAN and then plot relative concentration
if(interactive())
{
  bm<-batman()
  ## Plot relative concentrations
  plotRelCon(bm)
}
```

plotShift

Boxplot or Histogram of ppm Shift Posterior distributions for Multiplets of Named Metabolite

Description

This function provides boxplots or histograms of the ppm shift posterior distributions of multiplets, and saves the figure to pdf file in specified directory. The file name is in the format of "spec_*i*_metaName_ppmShift.pdf", where *i* is the spectrum number and "metaName" is the input metabolite name if given. The figure file will not be overwritten if it already exists. A prefix can be given to the file name for new saves.

Usage

```
plotShift(BM, metaName, plotHist = FALSE, breaks, perMult = FALSE,
          saveFig = TRUE, saveFigDir = BM$outputDir, prefixFig,
          overwriteFig = FALSE))
```

Arguments

BM	batman output data frame.
metaName	Only multiplets belonging to the named Metabolite will be shown. Only one metabolite name can be given. If missing, all metabolites will be plotted.
plotHist	If plotHist = TRUE, the ppm shift posteriors will be displayed as histogram. The default is FALSE.
breaks	A single number to set the number of bins for the histogram. If missing from the input, it is set to the data length divided by 3.
perMult	If set TRUE plot the shifts per multiplet, otherwise, plot the shifts per spectrum.
saveFig	Save figure to pdf file if set TRUE. The default is TRUE.
saveFigDir	Save pdf file in this directory. The default is the output directory of BM.
prefixFig	Add prefix to each saved figure name. The default is no prefix.
overwriteFig	Overwrite the saved figure file in pdf format if overwriteFig = TRUE. The default is FALSE.

See Also[batman](#), [batmanrerun](#)**Examples**

```
library(batman)
## Run BATMAN
if(interactive())
{
  bm<-batman()
  ## Plot ppm shift for each multiplet.
  plotShift(bm)
}
```

readBatmanOutput	<i>Reads in BATMAN Output Data Files</i>
------------------	--

Description

Reads in output data files from batman in specified folder.

Usage

```
readBatmanOutput(dirOP,dirIP)
```

Arguments

dirOP	The folder with batman output files.
dirIP	The folder with batman input files.

Value

It returns a data list with the objects described in [batman](#).

See Also[batman](#), [batmanrerun](#)**Examples**

```
library(batman)
## Run BATMAN
if(interactive())
{
  bm<-batman()
  ## Read in output files in saved directory.
  bmread<-readBatmanOutput(bm$outputDir,bm$inputDir)
}
```

readBruker

Read Raw Binary Bruker NMR Spectra

Description

Read in multiple raw binary Bruker NMR spectra (1D) from a specified folder, and return a matrix with columns:

$$[ppm, spectrum1, spectrum2, \dots]$$

Interpolation may be performed if spectra have different ppm scales.

Usage

```
readBruker(BrukerDataDir)
```

Arguments

BrukerDataDir The directory of the folder containing 1D Bruker spectral data files. Recursively finds all the "1r" files in datapath and read in.

Value

It returns a matrix with columns:

$$[ppm, spectrum1, spectrum2, \dots]$$

Examples

```
library(batman)
## Read in all Bruker NMR spectra files, replace "your\data\path\here" with the
## directory of the data files you want to read.
## brukerdata<-readBruker("your\data\path\here")
batmanDir = newDir(runBATMANDir = getwd(), overwriteFile = TRUE)
## The testBrukerPureSpec folder contains Bruker spectra files for testing
## purpose.
brukerdata<-readBruker(BrukerDataDir = batmanDir[5])
```

readBrukerZipped

Read Raw Binary Bruker NMR Spectra in Zipped format

Description

Read in multiple raw binary Bruker NMR spectra (1D), with spectrum data in a zipped format, from a specified folder, and return a matrix with columns:

$$[ppm, spectrum1, spectrum2, \dots]$$

Interpolation may be performed if spectra have different ppm scales.

Usage

```
readBrukerZipped(BrukerDataZipDir)
```

Arguments

BrukerDataZipDir

The directory of the folder containing zipped 1D Bruker spectral data files. Recursively finds all the "*.zip" files in datapath, unzipped them in the same folder, call "*.zip" files to read in spectra, and delete the unzipped folders. If no "*.zip" file was found, it works the same as "*.zip" files.

Value

It returns a matrix same as readBruker, with columns:

[ppm, spectrum1, spectrum2, ...].

Examples

```
library(batman)
## Read in all Bruker NMR spectra files, replace "your\data\path\here" with the
## directory of the data files you want to read.
## brukerdata<-readBruker("your\data\path\here")
batmanDir = newDir(runBATMANDir = getwd(), overwriteFile = TRUE)
## The testBrukerPureSpec folder contains Bruker spectra files for testing
## purpose.
brukerdata<-readBrukerZipped(BrukerDataZipDir = batmanDir[5])
```

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