## homework06

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### A. Improve the analysis code below.

```
df <- data.frame(a=1:10, b=seq(200,400,length=10),c=11:20,d=NA)
df$a <- (df$a - min(df$a)) / (max(df$a) - min(df$a))
df$b <- (df$b - min(df$a)) / (max(df$b) - min(df$b))
df$c <- (df$c - min(df$c)) / (max(df$c) - min(df$c))
df$d <- (df$d - min(df$d)) / (max(df$a) - min(df$d))</pre>
Write the function.

function_1 = function(x){
   (x - min(x)) / (max(x) - min(x))
}
Apply the function:
function_1(df$a)

[1] 0.0000000 0.1111111 0.2222222 0.3333333 0.4444444 0.5555556 0.6666667
[8] 0.7777778 0.8888889 1.0000000
```

B. Improve the below example code for the analysis of protein drug interactions.

```
library(bio3d)
s1 <- read.pdb("4AKE") # kinase with drug</pre>
```

```
Note: Accessing on-line PDB file

s2 <- read.pdb("1AKE") # kinase no drug

Note: Accessing on-line PDB file
PDB has ALT records, taking A only, rm.alt=TRUE

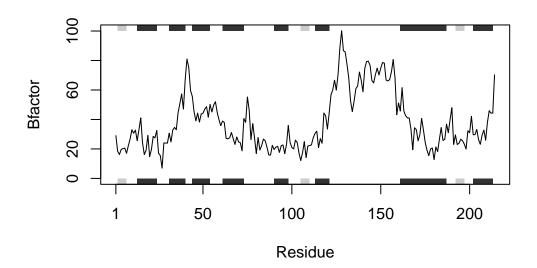
s3 <- read.pdb("1E4Y") # kinase with drug

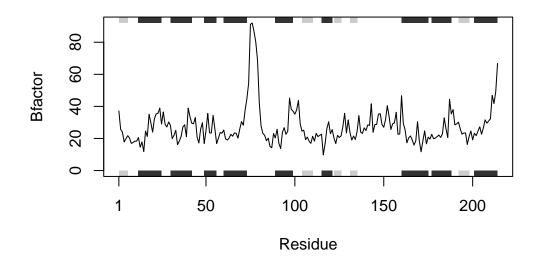
Note: Accessing on-line PDB file

s1.chainA <- trim.pdb(s1, chain="A", elety="CA")
s2.chainA <- trim.pdb(s2, chain="A", elety="CA")
s3.chainA <- trim.pdb(s1, chain="A", elety="CA")
s3.chainA <- trim.pdb(s1, chain="A", elety="CA")
s1.b <- s1.chainA$atom$b
s2.b <- s2.chainA$atom$b
```

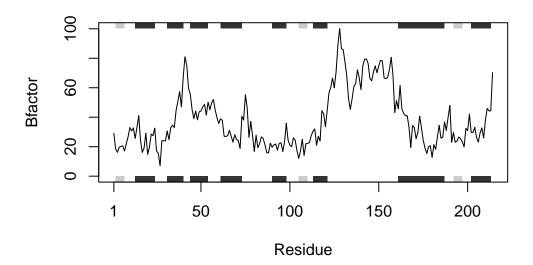
plotb3(s1.b, sse=s1.chainA, typ="l", ylab="Bfactor")

s3.b <- s3.chainA\$atom\$b





plotb3(s3.b, sse=s3.chainA, typ="1", ylab="Bfactor")



Write the function.

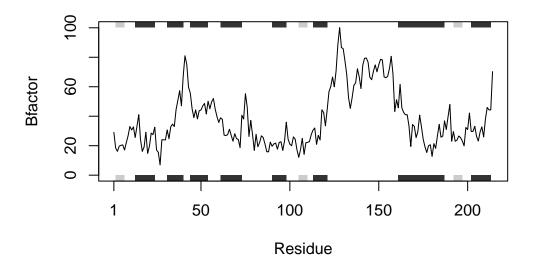
```
function_2 <- function(x){
    #input: PDB
    s <- read.pdb(x)
    s.chainA <- trim.pdb(s, chain="A", elety="CA")
    s.b <- s.chainA$atom$b
    plotb3(s.b, sse=s.chainA, typ="l", ylab="Bfactor")
    #output: a standard scatter plot with optional secondary structure in the marginal region
}</pre>
```

### Apply function:

```
function_2("4AKE")
```

Note: Accessing on-line PDB file

Warning in get.pdb(file, path = tempdir(), verbose = FALSE):
C:\Users\Lenovo\AppData\Local\Temp\RtmpYnJmt3/4AKE.pdb exists. Skipping
download



#### Q1. What type of object is returned from the read.pdb() function?

class(s1)

[1] "pdb" "sse"

The type of object returned from the read.pdb() function is large pdb.

#### Q2. What does the trim.pdb() function do?

The function is used to produce a new smaller PDB object, containing a subset of atoms, from a given larger PDB object.

Q3. What input parameter would turn off the marginal black and grey rectangles in the plots and what do they represent in this case?

```
s1 <- read.pdb("4AKE")</pre>
```

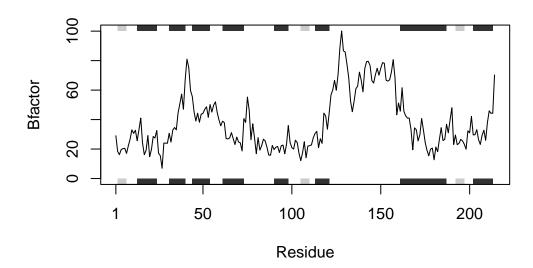
Note: Accessing on-line PDB file

Warning in get.pdb(file, path = tempdir(), verbose = FALSE):
C:\Users\Lenovo\AppData\Local\Temp\RtmpYnJmt3/4AKE.pdb exists. Skipping
download

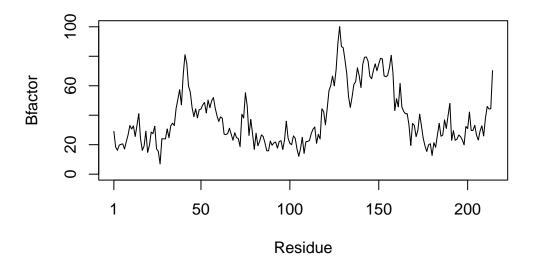
```
s1.chainA <- trim.pdb(s1, chain="A", elety="CA")
function_2("4AKE")</pre>
```

Note: Accessing on-line PDB file

Warning in get.pdb(file, path = tempdir(), verbose = FALSE):
C:\Users\Lenovo\AppData\Local\Temp\RtmpYnJmt3/4AKE.pdb exists. Skipping
download



plotb3(s1.b, sse=s1.chainA, typ="l", ylab="Bfactor", top=FALSE, bot=FALSE)



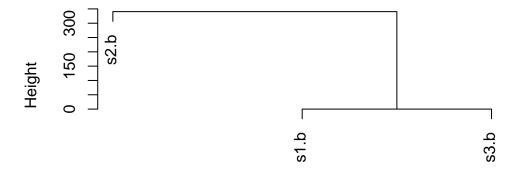
The input parameter "top=FALSE" and "bot=FALSE" would turn off the marginal black and grep rectangles in the plot.

In this case, the black ones represent alpha helices, and the grey ones represent beta strands.

#### Q4. What would be a better plot to compare across the different proteins?

```
hc <- hclust(dist(rbind(s1.b, s2.b, s3.b)))
plot(hc)</pre>
```

# **Cluster Dendrogram**



The difference distance matrix can be a better plot to compare across the different proteins.

Q5. Which proteins are more similar to each other in their B-factor trends. How could you quantify this?

```
hc <- hclust(dist(rbind(s1.b, s2.b, s3.b)))
plot(hc)</pre>
```

## **Cluster Dendrogram**



Q6. How would you generalize the original code above to work with any set of input protein structures?

```
function_2 <- function(x){
   s <- read.pdb(x)
   s.chainA <- trim.pdb(s, chain="A", elety="CA")
   s.b <- s.chainA$atom$b
   plotb3(s.b, sse=s.chainA, typ="l", ylab="Bfactor")
}</pre>
```