

**A. Discovery of New Genes related to
Leishmania Pathogenesis
and as Biomarkers of Attenuation Using
a Genomic Microarray**

**B. Multiplex PCR Microarray Assay to
Detect Pathogens in
Blood**

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Site Visit Presentation

Meeting the Challenge of Leishmaniasis: harvesting the benefits of the genomic era

Our knowledge of the *Leishmania* genome:

8333	Open Reading Frames
307 (4%)	Experimentally characterized
2618 (31%)	Inferred from homology
4673 (56%)	Conserved hypothetical

A method to rapidly identify virulence related genes: The Microarray

Goals:

- Genetic mechanisms of *Leishmania* pathogenesis
- Genetic characterization of live attenuated vaccine candidates
- Better diagnostics based on genetic technology
- Biomarkers of vaccine safety

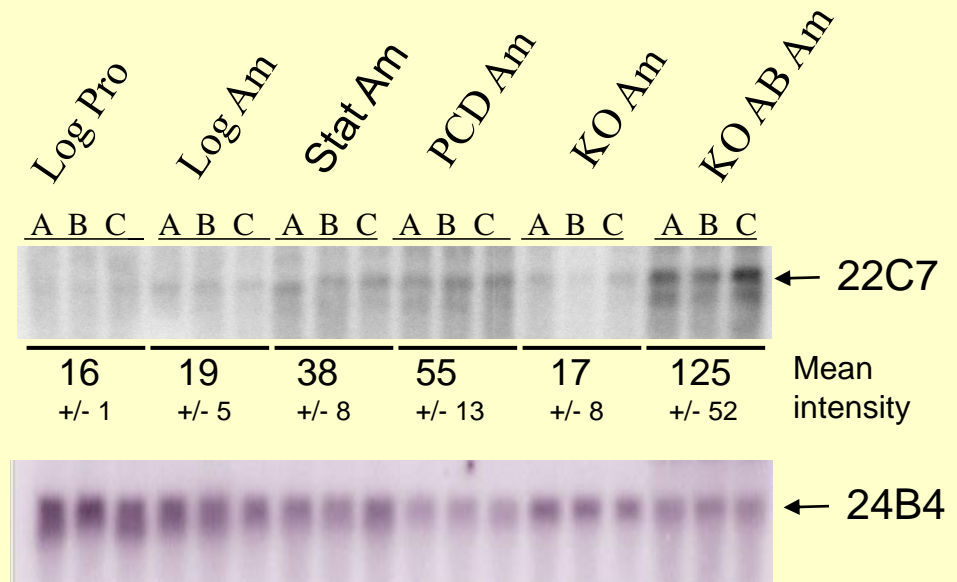
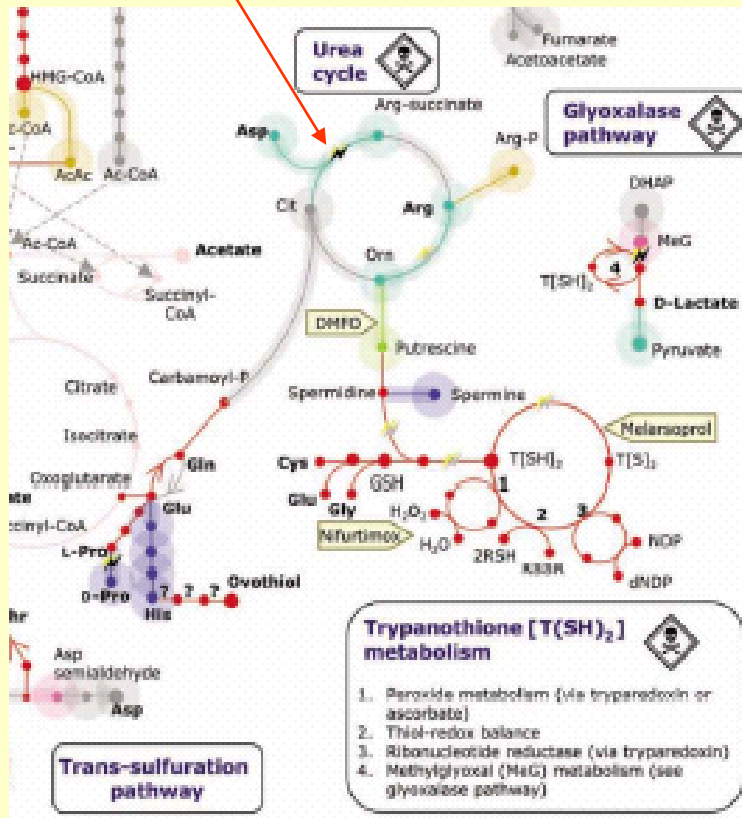
Rationale for microarray characterization of the centrin-deleted cell line

- Urgent need for a vaccine against leishmaniasis—live, attenuated approach
- Vaccine candidate must be safe—genetically stable to avoid reversion
- Global gene expression and identified biomarkers to measure genetic stability
- Focus CBER research to make a unique scientific contribution

L. d. Argininosuccinate synthase (22C7)

Argininosuccinate synthase

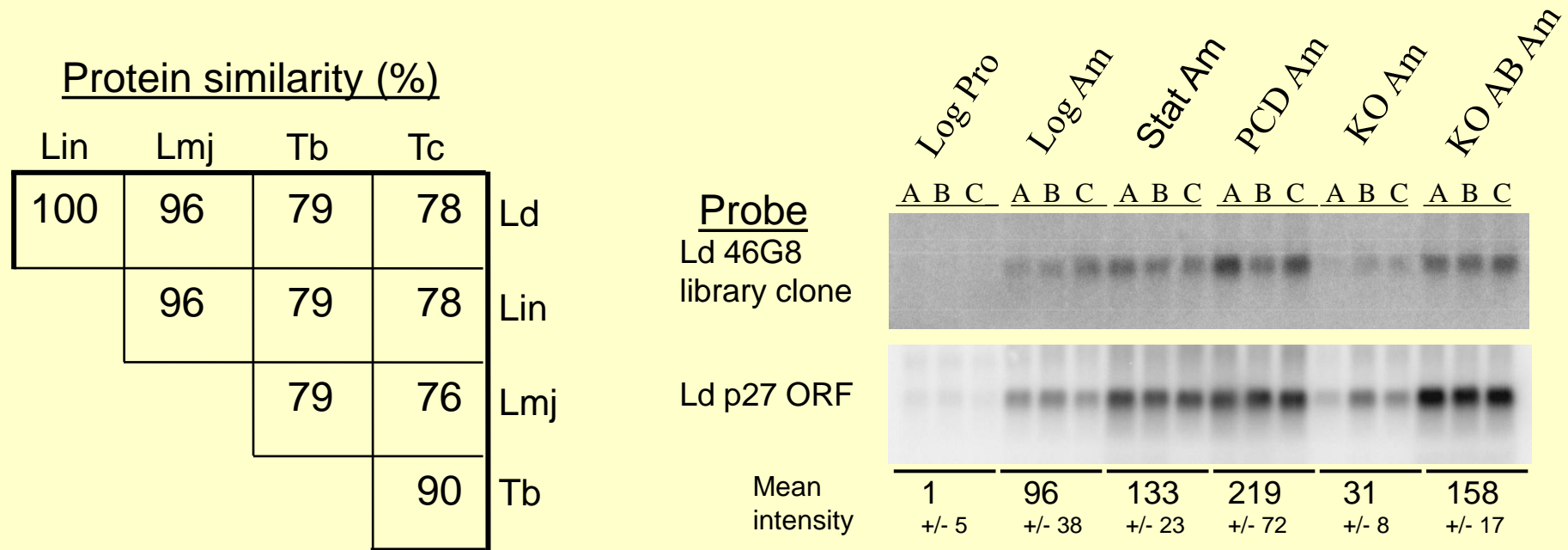
Science 7/15/05



Placement on a critical metabolic pathway and reproducible pattern of expression make *L. d.* argininosuccinate synthase a potential biomarker of attenuation

The hypothetical conserved, 27.6kD protein

(46G8) *L. d.* homologue of LmjF28.0980



Trypanosomatid restriction and high level of conservation suggest a critical function unique to the flagellated parasite physiology and the reproducible pattern of expression make *L.d.* p27 a potential biomarker of attenuation

Summary: characterization of gene expression in the centrin-deleted cell line

- Differentially expressed genes identified and validated
- Selected genes with potential as biomarkers of attenuation further characterized
- Characterized genes reveal physiological correlates of centrin deletion
- Characterization of newly described gene function may lead to better understanding of *Leishmania* pathogenesis

Meeting the Challenge of Blood Safety: harvesting the benefits of new technologies

- Transfusion blood safety has improved with pathogen testing
- Increasing number of known potential infectious agents and emerging threats, including bioterrorism increases the burden of testing
- Urgent need for methods to streamline and consolidate testing: nucleic acid tests (NAT), real-time PCR, microarrays, nanotechnology
- Multiplex potential of a pathogen detection microarray assay

Microarray for Detection of Blood-borne and BT Pathogens

Bacteria, and Parasites

Ba: *Bacillus anthracis* (**anthrax**)

Ft: *Francisella tularensis* (**tularemia**)

LT: *Leishmania* /*Trypanosoma*

Yp: *Yersinia pestis* and *pseudotuberculosis* (**plague**)

Bioterror Viruses

POX: Pox viruses

VAC: Vaccinia

VAR: Variola (**Smallpox**)

MPV: Monkeypox Viruses

CPV: Cowpox Viruses

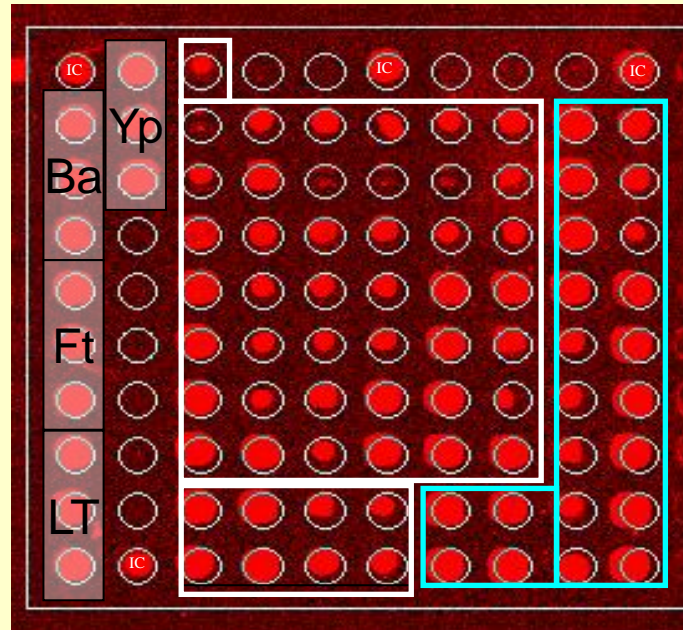
NOVAC: All Pox viruses but Vaccinia

EBO: Ebola Viruses

VE: Venezuelan Equine Enceph. Virus

VETD: VE Trinidad Donkey

MBG: Marburg Viruses



IC 4 internal control probes (Human rRNA gene)

Blood Borne Viruses

WNV: West Nile Viruses

HCV: Hepatitis C Viruses

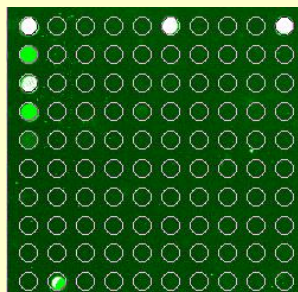
HBV: Hepatitis B Viruses

HIV: Human Immunodeficiency Viruses

HTLV: Human T-cell Leukemia Viruses

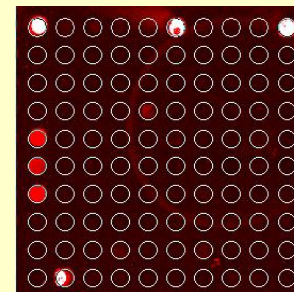
Results of detection in pathogen-spiked blood – 50 cells/ml

Bacillus anthracis



livestock
vaccine
strain

Francisella tularensis



Live
Vaccine
Strain

Yersinia pseudotub.

