

Package ‘XMRF’

June 24, 2015

Type Package

Title Markov Random Fields for High-Throughput Genetics Data

Version 1.0

Date 2015-06-12

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Depends R (>= 3.0.2)

Imports igraph, glmnet, MASS, snowfall, parallel, Matrix

Description Fit Markov Networks to a wide range of high-throughput genomics data.

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URL <http://www.liuzlab.org/XMRF/>

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XMRF-package

A R Package to Fit Markov Networks to High-throughput Genomics Data

Description

An R package to learn and visualize the underlying relationships between genes from various types of high-throughput genomics data.

Details

Package: XMRF
 Type: Package
 Version: 1.0
 Date: 2015-06-12
 License: GPL-2

Technological advances have produced large amounts of high-throughput "omics" data that allow us to study the complicated interactions between genes, mutations, aberrations, and epi-genetic markers. Markov Random Fields (MRFs), or Markov Networks, enable us to estimate these genomics networks via sparse, high-dimensional undirected graphical models.

Here, we provide the community a convenient and useful tool to learn the complex genomics networks from various types of high-throughput genomics data. This package encodes the recently proposed parametric family of graphical models based on node-conditional univariate exponential family distributions (Yang *et. al*, 2012, 2013a). Specifically, our package has methods for estimating Gaussian graphical model (Meinshausen and Buhlmann, 2006), Ising model (Ravikumar *et. al*, 2010), and the Poisson family graphical models (Allen and Liu, 2012, 2013; Yang *et. al* 2013b). These models can be used to estimate genetic networks from a variety of data types:

Genomics Data	Type	XMRF Family
=====	=====	=====
RNA-Seq or miRNA-Seq	Counts	LPGM, SPGM
Microarray or Methylation array	Continuous	GGM
Mutation or CNVs	Binary	ISM

To estimate the network structures from different types of genomics data with this package, users simply need to specify the "method" in the main function, for example `XMRF(..., method="LPGM")` to fit LPGM to next-generation sequencing data.

In this package, we implement the neighborhood selection graph estimation technique by proximal or projected gradient descent using warm starts over the range of regularization parameters. This

technique allows estimation of the neighborhood for each node independently and thus can be done in parallel, thus speeding computation.

This package also implements two data-driven approaches to select the sparsity of a fitted network: a stability-based approach for a single regularization value over many bootstrap resamples (Meinshausen and Bühlmann, 2010), or computed over a range of regularization values with StARS (Liu *et. al.*, 2010).

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- Yang, E., Ravikumar, P.K., Allen, G.I., and Liu, Z. (2013a). On graphical models via univariate exponential family distributions. *arXiv preprint arXiv:1301.4183*.
- Yang, E., Ravikumar, P.K., Allen, G.I., and Liu, Z. (2013b). On Poisson graphical models. *NIPS*, pp.1718-1726.

See Also

[XMRF](#)

Examples

```
library(XMRF)

n = 100
p = 20
```

```

sim <- XMRF.Sim(n=n, p=p, model="LPGM", graph.type="scale-free")
simDat <- sim$X

# Compute the optimal lambda
lmax = lambdaMax(t(simDat))
lambda = 0.01* sqrt(log(p)/n) * lmax
# Run LPGM
lpgm.fit <- XMRF(simDat, method="LPGM", N=10, lambda.path=lambda, parallel=FALSE)

# Print the fitted Markov networks
lpgm.fit

m1 = plotNet(sim$B)
m1 = plot(lpgm.fit, mylayout=m1)

```

brcadat

RNA-Seq Data of BRCA Patients from TCGA

Description

A matrix of RNA-Seq read counts from BRCA patients.

Usage

```
data("brcadat")
```

Format

A matrix of level 3 RNA-Seq (UNC Illumina HiSeq RNASeqV2) data with 445 breast invasive carcinoma (BRCA) patients from the Cancer Genome Atlas (TCGA) project on 353 genes with somatic mutations listed in the Catalogue of Somatic Mutations in Cancer (COSMIC). The matrix is organized in dimension of *gene* x *sample*.

ggm.fit

Fitted Gaussian Graphical Models

Description

An example fitted Gaussian graphical model

Usage

```
data("ggm.fit")
```

Format

A [GMS](#) object.

Details

This fitted Gaussian graphical model is included as the example model for demonstrating the usage of other functions. The model was fitted with a simulated multivariate Gaussian data of scale-free graph structure, 30 nodes, and 200 observations. StARS stability selection of 100 iterations was used to identify the optimal network over the regularization path of 10 parameters.

See Also

[GMS](#), [plot.GMS](#)

GMS

Graphical Models Object

Description

This class of objects is returned by the XMRF function in this package, to represent the fitted Markov Network(s). Objects of this class have the `print` method to display the core information of the fitted models and `plot` method to plot the optimal Markov Network.

Arguments

<code>v</code>	vector of (<code>n_lams</code> length) network variability measured for each regularization level.
<code>D</code>	a list of $p \times p$ matrices of stability scores of inferred edges of each network along the regularization path.
<code>lambda.path</code>	numeric vector used for regularization path.
<code>opt.lambda</code>	lambda value that gives the optimal network (network with maximum variability).
<code>network</code>	a list of $p \times p$ matrices of fitted networks along the regularization path.
<code>opt.index</code>	index of the regularization value that gives the optimal network.

See Also

[XMRF](#), [plot.GMS](#)

lambdaMax	<i>Maximum lambda</i>
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Description

Compute the maximum lambda

Usage

```
lambdaMax(X)
```

Arguments

X a *n* × *p* data matrix.

Details

Compute the largest value for regularization (maximum lambda) that gives the null model. The maximum lambda is computed based on the input data matrix, and is the maximum element from column-wise multiplication of data matrix normalized by the number of observations.

Value

a numeric value

plot.GMS	<i>Plot GMS Object</i>
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Description

Default function to plot the network of a GMS object.

Usage

```
## S3 method for class 'GMS'
plot(x, fn = "", th = 1e-06, i = NULL, mylayout = NULL, vars = NULL, ...)
```

Arguments

x	a GMS object.
fn	file name to save the network plot; default to be an empty string, so the network is plotted to the standard output (screen). NOTE: if a file name is specified, it should be file name for PDF file.
th	numeric value, default to 1e-06. To specify the threshold if the estimated coefficient between two variables is to be considered connected.

<code>i</code>	index of the network (along the regularization path) to be plotted. Default to NULL to plot the optimal network.
<code>mylayout</code>	graph layout to draw the network, default to NULL.
<code>vars</code>	vector of variable names, default to NULL.
<code>...</code>	other generic arguments for plot method.

Details

This is the default plotting function for GMS objects (Markov Networks inferred over a regularization path). Refer to [GMS](#) for details on GMS object. This function will plot the optimal network on the screen by default. However, given a file name, the plot will be saved to a PDF file. Also, given a specific index corresponding to the index of `lambda.path`, the associated network will be plotted.

The network will be plotted in force-directed layout (`layout.fruchterman.reingold` with default parameters implemented in `igraph` package).

Value

Returns the layout object from `igraph` package - numeric matrix of two columns and the rows with the same number as the number of vertices.

See Also

[GMS](#)

Examples

```
library(XMRF)
data('ggm.fit')
plot(ggm.fit, fn="ggm.fit.net.pdf")
```

plotGML

Plot Network in GML

Description

Plot the network in graph modeling language (GML).

Usage

```
plotGML(x, fn = "", th = 1e-06, i = NULL, weight = FALSE, vars = NULL)
```

Arguments

x	a GMS object.
fn	file name to save the GML file.
th	numeric value, default to 1e-06. To specify the threshold if the estimated coefficient between two variables is to be considered connected.
i	index of the network (along the regularization path) to be plotted. Default to NULL for optimal network.
weight	boolean value to indicate if writing the stability on the inferred edges, default to FALSE.
vars	vector of variable names, default to NULL.

plotNet

Plot Network

Description

Plot a network with specific layout.

Usage

```
plotNet(net, fn = "", th = 1e-06, mylayout = NULL)
```

Arguments

net	a square adjacency matrix of the network to be plotted.
fn	file name to save the network plot. Default to be an empty string, so the network is plotted to the standard output (screen). NOTE: if a file name is specified, it should be file name for PDF file.
th	numeric value, default to 1e-06. To specify the threshold if the estimated coefficient between two variables is to be considered connected.
mylayout	graph layout to draw the network, default to NULL.

Details

This function serves as the alternative plotting function to allow users to plot a specific network with specific layout, such as plotting the simulated network.

Value

Returns the layout object from igraph package - numeric matrix of two columns and the rows with the same number as the number of vertices.

Examples

```
library(XMRF)
n = 200
p = 30
sim <- XMRF.Sim(n=n, p=p, model="LPGM", graph.type="scale-free")
m1 = plotNet(sim$B)
```

processSeq

*Process Sequencing Data for Poisson-based MRFs***Description**

Process and normalize RNA-Sequencing count data into a distribution appropriate for Poisson MRFs.

Usage

```
processSeq(X, quanNorm = 0.75, nLowCount = 20, percentLowCount = 0.95, NumGenes = 500,
PercentGenes = 0.1)
```

Arguments

X	<i>n</i> × <i>p</i> data matrix.
quanNorm	an optional parameter controlling the quantile for sample normalization, default to 0.75.
nLowCount	minimum read count to decide if to filter a gene, default to 20.
percentLowCount	filter out a gene if it has this percentage of samples less than nLowCount, default to 0.95.
NumGenes	number of genes to retain in the final data set, default to 500.
PercentGenes	percentage of genes to retain, default to 0.1.

Details

To process the next-generation sequencing count data into proper distribution (with dispersion removed), the following steps are taken in this function:

1. Quantile normalization for the samples.
2. Filter out genes with all low counts.
3. Filter genes by maximal variance (if specified).
4. Transform the data to be closer to the Poisson distribution. A log or power transform is considered and selected based upon the Kolmogorov-Smirnov goodness of fit test.

Value

a *n* × NumGenes or PercentGenes processed data matrix.

Examples

```
library(XMRF)
data('brcadat')
brca = t(processSeq(t(brcadat), PercentGenes=1))
```

XMRF

Markov Random Fields for Exponential Family Distributions

Description

Infer networks from genomics data using Markov Random Fields specified by node-conditional univariate exponential family distributions.

Usage

```
XMRF(X, method = "LPGM", stability = "bootstrap", N = 100, beta = 0.01, lmin = 0.01,
      nlams = 20, lambda.path = NULL, parallel = TRUE, nCpus = 4, sym = TRUE, th = 0.01,
      sth = 0.95, R = max(X), R0 = 0)
```

Arguments

X	a $p \times n$ data matrix.
method	specification of the type of MRF model, default to "LPGM" for log-linear Poisson-based graphical model. Other allowed methods are "PGM" for regular Poisson, "TPGM" for truncated Poisson, "SPGM" for sublinear Poisson, "GGM" for Gaussian graphical models, and "ISM" for Ising model.
stability	specification of the stability method, default to "bootstrap". Another accepted value is "star" for Stability Approach to Regularization Selection (StARS).
N	number of iterations for stability selection, default to 100.
beta	threshold value on sparsity of the network, default to 0.01.
lmin	ratio of minimum lambda value from the maximum lambda value, default to 0.01.
nlams	number of lambda for regularization, default to 20.
lambda.path	vector lambda used for regularization, default ot NULL.
parallel	logical value to indicate if the process should be run parallely in multiple threads, default to TRUE.
nCpus	number of (maximum) cores to use for parallel execution, default to 4.
sym	logical value to indicate if symmetry is enforced on the inferred edges, default to TRUE.
th	threshold value for the estimated edge coefficient, default to 0.005.
sth	an inferred edge is retained only if its stability score is greater than sth, default to 0.9.
R	truncation level for classes "TPGM" and "SPGM". The value has to be positive. Default to the maximum value of the input data matrix.
R0	lower-bound truncation level for "SPGM", default to 0.

Details

This is the main function of the package that fits exponential family Markov Networks to genomics data. To estimate the network structures using native distribution of the genomics data, specify the MRF family types in the "method" parameter. For genomic networks based on next-generation sequencing data, we recommend using the LPGM family. The table at the beginning of the document lists the family type recommended for each of the genomic data platforms.

Value

An object of class `GMS` will be returned representing the inferred Markov networks over the regularization path. See [GMS](#) for details.

References

- Allen, G.I., and Liu, Z. (2012). A Log-Linear graphical model for inferring genetic networks from high-throughput sequencing data. *The IEEE International Conference on Bioinformatics and Biomedicine (BIBM 2012)*.
- Allen, G. I., and Liu, Z. (2013). A Local Poisson Graphical Model for Inferring Genetic Networks from Next Generation Sequencing Data. *IEEE Transactions on NanoBioscience*, **12**(3), pp.1-10
- Liu, H., Roeder, K., and Wasserman, L. (2010). Stability approach to regularization selection (stars) for high dimensional graphical models. *NIPS* 23, pp.1432-1440.
- Meinshausen, N. and Bühlmann, P. (2006). High-dimensional graphs and variable selection with the lasso. *The Annals of Statistics*, **34**(3), pp.1436-1462.
- Meinshausen, N. and Bühlmann, P. (2010). Stability selection. *Journal of the Royal Statistical Society: Series B (Statistical Methodology)*, **72**(4), pp.417-473.
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- Yang, E., Ravikumar, P.K., Allen, G.I., and Liu, Z. (2013a). On graphical models via univariate exponential family distributions. *arXiv preprint arXiv:1301.4183*.
- Yang, E., Ravikumar, P.K., Allen, G.I., and Liu, Z. (2013b). On Poisson graphical models. *NIPS*, pp.1718-1726.

See Also

[XMRF-package](#), [GMS](#), [plot.GMS](#)

Examples

```
# Example for LPGM
# Refer to the package's introduction for identical example
## Not run: n = 100
## Not run: p = 20
## Not run: sim <- XMRF.Sim(n=n, p=p, model="LPGM", graph.type="scale-free")
## Not run: simDat <- sim$X
## Not run: # Compute the optimal lambda
```

```
## Not run: lmax = lambdaMax(t(simDat))
## Not run: lambda = 0.01* sqrt(log(p)/n) * lmax
## Not run: # Run LPGM
## Not run: lpgm.fit <- XMRF(simDat, method="LPGM", N=10, lambda.path=lambda)
## Not run: m1 = plotNet(sim$B, fn="simDat.netPlot.pdf")
## Not run: m1 = plot(lpgm.fit, fn="lpgm.netPlot_1.pdf", i=1, mylayout=m1)
## Not run: plot(lpgm.fit, fn="lpgm.fit.net.pdf")
```

XMRF.Sim

Generate simulated data from XMRF models

Description

Generate data from different multivariate distributions with different network structures.

Usage

```
XMRF.Sim(n = 100, p = 50, model = "LPGM", graph.type = "scale-free")
```

Arguments

n	number of samples, default to 100.
p	number of variables, default to 50.
model	Markov Network models to indicate the distribution family of the data to be generated, default to "LPGM". Other model options include "PGM", "TPGM", "SPGM", "GGM" and "ISM".
graph.type	graph structure with 3 options:"scale-free", "hub", and "lattice". Default to "scale-free".

Details

This function will first generate a graph of the specified graph structure; then based on the generated network, it simulates a multivariate data matrix that follows distribution for the Markov Random Fields model specified.

Value

A list of two elements:

B	pxp adjacency matrix of the generated graph.
X	pxn data matrix.

Examples

```
library(XMRF)

# simulate scale-free network and data of multivariate Poisson for LPGM
sim <- XMRF.Sim(n=100, p=20, model="LPGM", graph.type="scale-free")
hist(sim$X)
plotNet(sim$B)

# simulate hub network and data of multivariate Gaussian for GGM
sim <- XMRF.Sim(n=100, p=20, model="GGM", graph.type="hub")
hist(sim$X)
plotNet(sim$B)

# simulate hub network and data of multivariate binomial for ISM
sim <- XMRF.Sim(n=100, p=15, model="ISM", graph.type="hub")
hist(sim$X)
plotNet(sim$B)
```

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