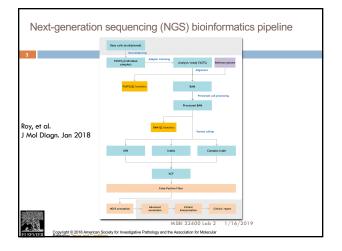
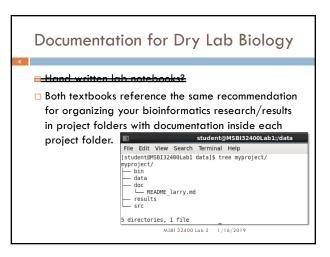
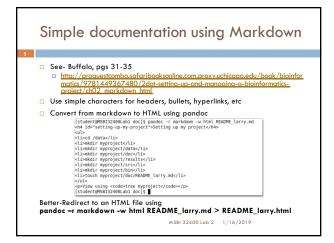
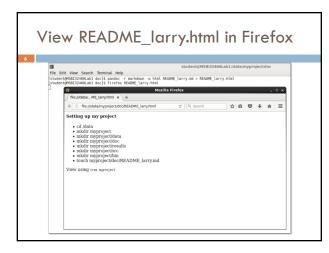


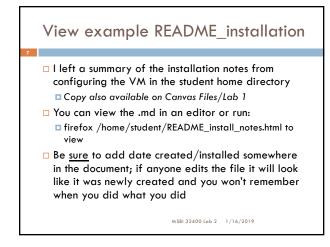
# Why document? □ Titus Brown's manifesto on reproducibility: "How we make our papers replicable", July 15, 2014 □ http://ivory.idyll.org/blog/2014-our-paper-process.html □ Cited by MacArthur Lab when they released code to reproduce all the figures in their ExAC paper □ https://macarthurlab.org/2016/03/17/reproduce-all-the-figures-g-users-guide-to-exac-part-2/ ➤ CAP, CLIA requirements to document bioinformatics workflow as a part of NGS analysis □ Roy, et al. "Standards and Guidelines for Validating Next-Generation Sequencing Bioinformatics Pipelines: A Joint Recommendation of the Association for Molecular Pathology and the College of American Pathologists." J Mol Diagn. 2018 Jan;20(1):4-27. doi: 10.1016/j.jmoldx.2017.11.003. Epub 2017 Nov 21. PMID: 29154853

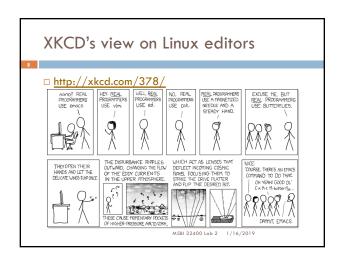


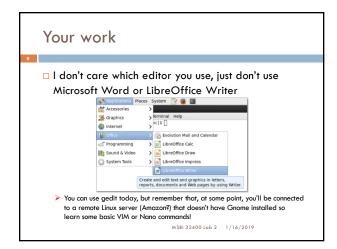


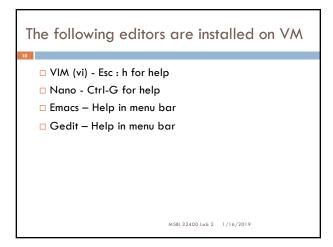




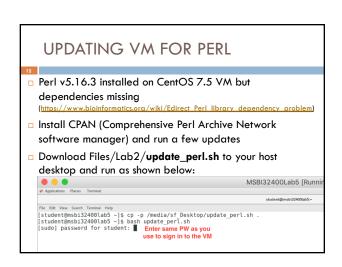


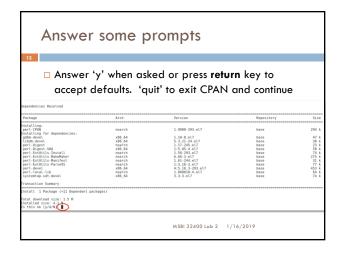


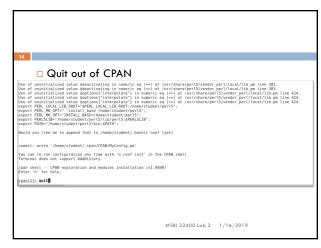


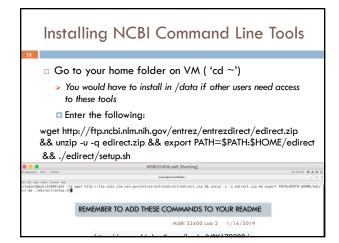


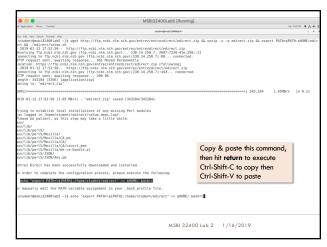
# Capturing Linux commands is simple Linux keeps a command history Scroll back through commands with up/down cursor Type 'history' to view most recent ~1000 commands Can cut and paste from console Better, use 'echo' Scroll back to a command, then add echo and quotes echo 'mkdir myproject/doc' >> README\_larry.md NB-Single > creates a new file (ERASING what was there!) so use >> to append Can type 'history >> README\_larry.md' then edit lines you don't want Don't assume the history will "be there the next time"....











### Try a few of the examples using tools

- □ Create a new directory: mkdir –p /data/Lab2/doc
  - □ From your /data/Lab2 folder (should work if your PATH is right; if not, add ~/esearch/<command>):
    - esearch –db pubmed –query "helseth dl AND collagen" | efetch –format pubmed > doc/example1.txt
    - esearch -db pubmed -query "bioinformatics [MAJR] AND software [TIAB]" | efetch -format xml | xtract -pattern PubmedArticle -block Author -sep " " -tab "\n" -element LastName,Initials | sort-uniq-count-rank > doc/bioinformatics\_authors.txt
    - esearch -db protein -query 'NP\_000509.1' | efetch -format fasta > doc/hbb.fasta

**CAUTION**-Copy & paste from **Microsoft PowerPoint** substitutes the wrong kind of dash (-- instead of -) so check carefully.

REMEMBER TO ADD THESE COMMANDS TO YOUR README

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### Sickle Cell Disease - 1 SNP

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- https://www.nhlbi.nih.gov/health/health-topics/topics/sca
- Single mutation in HBB subunit causes the hemoglobin tetramer to aggregate when deoxygenated, forming strands within the red blood cells.
- A Glutamic acid ('E') is changed to a Valine ('V'), altering the way hemoglobin molecules interact

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### Viewing HBB in 1000 Genome Data

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- □ YRI Trio (Yoruban) Trio from Ibadan, Nigeria
- NA19238, NA19239 and NA19240 are the YRI trio: mother, father, & daughter respectively
  - http://www.internationalgenome.org/data-portal/sample/NA19240
- 1000 Genome data, and reference links like dbSNP, are available to add "From server" in IGV

A Auton et al. Nature 526, 68-74 (2015) doi:10.1038/nature15393

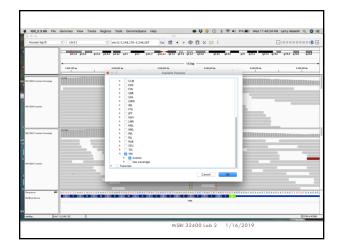
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### 1000 Genome & dbSNP in IGV

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- Click File/Load from server then expand 1000 Genome
- Expand Alignments, scroll down and expand YRI
- □ Expand **exome**
- Scroll down and select NA19238, NA19239 and NA19240 then click OK
- □ Enter **HBB** in IGV then zoom in on exon 1
- Add dbSNP with File/Load from server, expand Annotations, expand Variations and Repeats, select dbSNP 1.4.7 and click OK

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### Include in your write-up Who has variant at E7? Expand the dbSNP track. Which rsID(s) are associated with this variant at the E7 position?

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### Using NCBI tools to visualize HBSc

- This will <u>not</u> work on the Virtual Machine so you'll need to use your laptop.
- Go to OMIM.org and search for "sickle cell disease", then click on the second link (+ 141900. HEMOGLOBIN--BETA LOCUS; HBB)
- Click on the "Table View" in the left menu, then search the web page (Command-F on Mac, Ctrl-F Win) for "sickle", and click on the left link ('.0243')
- Note the first rs#. Open dbSNP (https://www.ncbi.nlm.nih.gov/snp/) and search for that rs#

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### Visualizing HBSc (cont)

- □ Click on the 'Protein 3D' link below the sequence coordinates and HGVS entries
- Install Cn3D if it's not already installed, then click the "View Structure and Alignment in Cn3D" button to view the single HBB chain.
- □ Highlight Glu 6 ("E"), and note where it appears on the surface (spin molecule if needed). Use File/Export PNG to save to your Desktop (<username>\_hbb.png)

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