PRFFECT User Manual

This program requires an R installation. Download R from: https://cran.r-project.org/ Or optionally RStudio from: https://www.rstudio.com/

RStudio is a user-friendly GUI for R.

Packages are installed via R/RStudio interface: install.packages("<package name>", dependencies = TRUE) Please always use 'dependencies = TRUE' when installing packages for PRFFECT.

What does PRFFECT do?

PRFFECT can be used to provide rapid pre-processing and classification results for many types of spectroscopy. There are several pre-processing options split into 4 main categories. These are binning, smoothing, normalisation, and baseline correction.

1.1 Binning

Binning is used when changing the overall resolution of the spectra is desired. It works by taking an average of the intensity value of every n points. It also effectively increases the wavenumber spacing of the spectra. This practice can sometimes increase the signal-to-noise ratio of spectra. In the implementation here, the parameter **bin_factor** is set in the user defined variables file. This factor dictates how many consecutive datapoints are averaged. For example, a **bin_factor** of 4 would provide a processed spectrum with 4 times fewer wavenumber columns and datapoints.

1.2 Smoothing

The choices for smoothing (set via the **smooth_choice** variable value) are four-fold; none (0), Savitzky-Golay filtering (1), wavelet denoising (2), and local polynomial fitting with Gaussian weighting (3).

These options also have parameters which can be set via the **smooth_par** value. This **smooth_par** becomes the parameter for the smoothing method set by **smooth_choice**. In the case of **smooth_choice** option 1 (S-G filter), the **smooth_par** value sets the filter order. For option 2 (wavelet denoising), **smooth_par** sets the length of the wavelet filter. If **smooth_choice** option 3 (local polynomial with Gaussian weighting) is selected, **smooth_par** sets the bandwidth of the Gaussians. See Table 5.1 for a clearer explanation of how this

works.

1.3 Normalisation

Normalisation choice is set by the **norm_choice** value. There are four options for normalisation in PRFFECT. These are none (0), min/max scaling (1), vector normalisation (2), and normalisation to Amide I band (3).

1.4 Baseline Correction

There are five options for baseline/background correction offered. These are set by the bg_choice value. The options are none (0), first derivative (1), second derivative (2), rubberband (3), and polynomial baseline correction (4). Options 3 and 4 have additional parameters which must be set. These are set by altering the value of the integer variable bg_par. If bg_choice is set to 3, the bg_par value controls the noise cut-off level used by the rubberband algorithm. An additional parameter RBp is provided to further control the noise-cutoff level for the rubberband method only. This RBp parameter acts as a multiplier for the bg_par value for rubberband. This is to allow for easier grid searching. When bg_choice is set to option 4 (polynomial baseline correction), bg_par controls the degree of the polynomial used.

Prerequisites and Data Format

The user should have a recent version of R, with the packages: "randomForest", "xtable", "caret", "prospectr", "signal", "rwt", "KernSmooth", "Peaks", and "hyperSpec" installed.

PRFFECT also expects the input spectroscopic data to be in the CSV (comma separated values) format. This is available in all modern spreadsheet programs. All of the data should be in one file, with **spectra in rows**. The labels for the data, eg **class** & **ID information**, **should be in columns at the front of the file**.

The column headings for the class and ID information can be text or numeric. The column headings for the actual intensity data should be numbers, (eg wavenumbers, wavelength etc.) but with "X_" appended. See table 2.1 below for an example acceptable layout.

Table 2.1: Example dataset format

Cancer/non	Met/brain/non	Origin	HGG/LGG	subtype	Patient_ID	X_898.65	X_900.58	X_902.51	X_904.44	X
1	1	6	0	0	13673	0.0116	0.0116	0.0115	0.0115	
1	1	6	0	0	13673	0.0115	0.0115	0.0115	0.0114	
1	1	6	0	0	13673	0.0116	0.0116	0.0115	0.0115	
1	1	6	0	0	13673	0.0114	0.0114	0.0113	0.0113	
1	1	6	0	0	13673	0.0116	0.0115	0.0115	0.0114	
1	1	6	0	0	13673	0.0117	0.0116	0.0116	0.0115	
1	1	6	0	0	13673	0.0116	0.0116	0.0116	0.0115	
1	1	6	0	0	13673	0.0117	0.0117	0.0116	0.0116	
1	1	6	0	0	13673	0.0117	0.0117	0.0116	0.0116	
1	1	3	0	0	13672	0.0116	0.0115	0.0115	0.0114	
1	1	3	0	0	13672	0.0115	0.0115	0.0114	0.0113	
1	1	3	0	0	13672	0.0116	0.0115	0.0115	0.0114	
1	1	3	0	0	13672	0.0115	0.0115	0.0114	0.0113	
1	1	3	0	0	13672	0.0115	0.0114	0.0113	0.0113	
1	1	3	0	0	13672	0.0115	0.0114	0.0114	0.0113	
1	1	3	0	0	13672	0.0117	0.0116	0.0115	0.0115	
1	1	3	0	0	13672	0.0117	0.0116	0.0116	0.0115	
1	1	3	0	0	13672	0.0119	0.0119	0.0118	0.0117	
1	1	6	0	0	13671	0.0119	0.0118	0.0118	0.0117	
1	1	6	0	0	13671	0.0121	0.0120	0.0119	0.0119	
1	1	6	0	0	13671	0.0121	0.0120	0.0120	0.0119	

In this case, there are several class columns and a patient ID column. The wavenumber columns are denoted by the X_{-} prefix. The content of the label and ID columns are numeric identifiers in this case, but they could be text also. The content of the X_{-} prefix columns are the recorded absorbencies at each wavenumber, forming a whole spectrum per row. If a column contains intensity/absorbance data, it **must be prefixed with X_{-} as shown in the table**. An easy way to do this is to use the CONCATENATE command in Excel. This command allows a user to add a prefix to every sell in a selected range.

Setting up and splitting datasets

For the RF classifier, a splitting of the data into a training and test set is required. Cross validation is performed on the training set, and the learned patterns from this are used to predict the content of the test set. In the Github repository, there is a small R script provided called "select_training_and_test_sets.r". This script will split a spectroscopic dataset into two files, a training set and a test set. There are **four variables a user must set in order to used this script**. These are the name of the dataset, the column header denoting the ID column, the yvar column and the desired training set size.

Figure 3.1: Interface of the splitting script

```
f<-read.table("coffee_BRS.csv", header=TRUE, sep=",")
patients<-"sample_number"

yvar="coffee_type"
trprop <- 0.66</pre>
```

The first line should be changed to reflect the filename of the dataset under study. The second line should be changed to signify the column heading containing patient ID or sample number. This is to allow for datasets containing more than one spectrum per sample. It ensures that spectra are kept together, and that multi-spectrum samples are not split between the training and test

sets. yvar should be set to a class column. This is the column which the script "takes aim at" making sure an even split of classes occurs. trprop is the proportion of the data which ends up being in the training set. The rest goes to the test set. At the setting shown, a training set would be two thirds of the data, and the test set would be the remaining third.

After running this script, the original dataset will still be present and two new files will be created containing the training and test sets. By default, these are named "input_training_dataset.ssv" and "input_testing_dataset.ssv". These files are in the format of semicolon-separated values.

Data can of course be split manually, but care must be taken to ensure the two resulting files have the same number of columns and column headers.

The user-defined input file

This file is how the user interacts with the program, and where the pre-processing and output choices are made.

The first two variables, **infile_tr & infile_te** are for telling the program the names of the training and testing input datasets.

The variable ${\bf k}$ sets the number of folds of the k-fold cross validation.

The logicals **se_and_ci** & **se_and_ci2** set whether standard errors and confidence intervals for each statistic are reported the the results; this is for text and tabular result files respectively.

The variable **bin_factor** sets the degree of wavenumber resolution decrease. A bin_factor setting of 2 would signify that the effective spectral resolution is being decreased by a factor of 2.

smooth_choice allows for the selection of the smoothing method. The variable **smooth_par** allows adjustment of the settings for certain smoothing methods. See the section "Available Pre-processing Options" for specific settings of both smooth_choice and smooth_par.

Normalisation choice is set by selecting the appropriate number for **norm_choice**. Baseline/background correction methods and settings are chosen by adjusting **bg_choice** & **bg_par** respectively.

RBp is a setting for the rubberband baseline correction method. It specifies the level of the estimated noise floor of the spectrum. It is used in conjunction with **bg_par** when using the rubberband method. It is simply a multiplier of the **bg_par** only when rubberband baseline correction is used. We recommend that it is set to the same order of magnitude as the overall lowest y-axis point of the spectra. This allows a simplification of a grid search, allowing bg_par only to be adjusted.

Next comes the section of the input file which dictates the types of graphs desired. These are all set by logical variables. Graphs of spectra only, average spectra (per class), Gini importance & spectra, and average spectra & Gini importance can all be requested. The plot of Gini importance only is always plotted.

The **yvariables** input should contain the column headers with which to do the classification. This can be either one or multiple columns.

The options min_wavenumber & max_wavenumber choose the region of the spectrum to be used in the RF classification. This also affects the region should on the graphs.

The variable **patients** is where the name of the column containing patient IDs/sample numbers should be put.

The input file is read automatically upon running the main program.

Available Pre-processing Options

Table 5.1: Key to results

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Setting	Explanation	Parameter	Explanation
bin_factor	Binning factor		
1	No binning		
2	delta*2		
4	delta*4		
8	delta*8		
16	delta*16		
32	delta*32		
n	delta*n		
smooth_choice	Smoothing choice	smooth_par	Smoothing parameter
0	none	-	-
1	S-G filter	1,2,3,4,5,6,	Filter order
2	Wavelet denoising	2*(2,3,4,5,6,)	Filter length
3	Local polynomial fit with Gaussian	1,2,3,4,5,6,	Width of Gaussian
		, , , , , ,	
norm_choice	Normalisation choice		
0	none		
1	min/max to 0-1 scaling		
2	Vector normalisation		
3	Normalise to Amide I band		
bg_choice	Baseline/background Correction choice	bg_par	Baseline parameter
0	none	-	-
1	First derivative	-	-
2	Second derivative	_	-
3	Rubberband baseline correction	RBp*(1,2,3,4,5,6,)	Noise cut-off level
4	Polynomial baseline correction	1,2,3,4,5,6,	Polynomial degree

```
Figure 5.1: User Defined Variables
infile_tr<-"input_training_dataset.ssv"
infile_te<-"input_testing_dataset.ssv"
```

```
<<-5
se_and_ci<-FALSE
se_and_ci2<-FALSE
bin factor <- 2
smooth_choice <- 0
smooth_par <- 0
norm_choice <- 0
bg_choice <- 1
bg_par <- 0
RBp < -0.1
spectra <- TRUE
average_spectra <- FALSE
imp_and_all_spectra <-FALSE</pre>
avg_and_gini <- FALSE
yvariables<-c("coffee_type")</pre>
```

min_wavenumber<-800
max wavenumber<-1900

patients<-"sample_number"

Example Workflow - General Use

6.1 Input

An example workflow for a single run of the program is shown in this section. The user wishes to study a dataset called "olive_oil_BRS.csv" containing FTIR ATR spectra of various olive oil samples from different countries. The dataset contains 2 columns at the start of the file called "sample_number" and "country". The remainder of the file contains wavenumber columns with absorbency values, with the X_ prefix, as described above.

The user would first used the script "select_training_and_test_sets.r" and set desired parameters in that. In this case, **trprop** will be set to 0.75 for 75% of data as the training set. A simple R command to run the script is shown:

source "select_training_and_test_sets.r"

Screen captures of the script and the resulting log are shown below.

Figure 6.1: Example Workflow - general : Split script setup

```
f<-read.table("olive_oil_BRS.csv",

patients<-"sample_number"

yvar="country"
trprop <- 0.75</pre>
```

After this step, 3 new files have been generated: **select_training_and_test_sets.log**, input_training_dataset.ssv & input_testing_dataset.ssv.

Now comes the inputting of user-defined variables in the **user_defined_input.R** file. In this case, the user wished to bin the data by a factor of 2, perform S-G filtering of order 2, vector normalisation and first derivative processing. A screen capture of these options is shown below. Other settings were a wavenumber range of 800 - 1800 cm $^{-1}$, setting all graphical output to TRUE, and specifying the relevant columns.

```
Figure 6.3: Example Workflow - general: User defined input
         bin_factor <- 2
         smooth_choice <- 1</pre>
         smooth_par <- 2</pre>
         norm_choice <- 2
         bg_choice <- 1
         bg_par <- 0
         RBp <- ∅
          spectra <- TRUE
          average_spectra <- TRUE
          imp_and_all_spectra <-TRUE
         avg_and_gini <- TRUE</pre>
         yvariables<-c("country")</pre>
         min_wavenumber<-800
         max_wavenumber<-1800
         patients<-"sample_number"</pre>
```

Then all that remains is to run the program. A command in R or Rstudio to do this is:

source "PRFFECTv1.r"

6.2 Output

Several files are generated by running the program. These are listed and described below.

country_random_forest_importance_MeanDecreaseGini.txt This contains two columns, wavenumber and Gini importance. It shows how important each wavenumber column was to the classification. It is formatted in order of importance, with the most important wavenumbers at the end of the file.

country_results_ALL.html This file contains a table of statistical results for the classification. It can be opened in a web browser for quick viewing.

country_results_ALL.txt Contains the same data as the html file, but in a standard text format suitable for viewing in spreadsheet programs etc.

seed.txt This contains the current random seed in R at the time of running the program. It is only useful if you wish to perform EXACTLY the same random forest classifier again in the future.

country_rf.RData This is captured image of the internal R workspace. It can be opened and browsed should one wish to see details of the internal process of the classification for example.

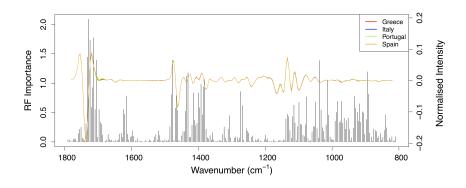
PDF files, *5 These contain the requested plots for viewing the spectra and/or the Gini importance.

Some files are shown as images below.

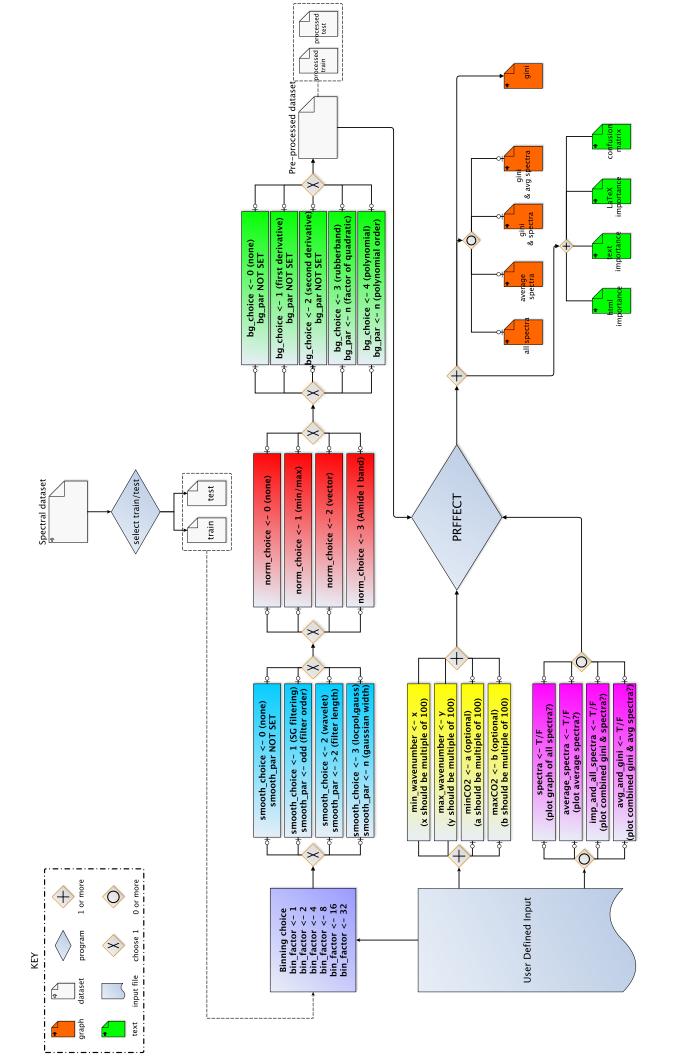
Figure 6.4: Example Workflow - general : html table

Class	pacCV	mccCV	sensCV	specCV	pprCV	nprCV	pacTS	mccTS	sensTS	specTS	pprTS	nprTS
Greece	1	1	1	1	1	1	0.964	0.876	0.8	1	1	0.958
Italy	0.978	0.949	0.929	1	1	0.97	0.893	0.786	0.727	1	1	0.85
Portugal	0.967	0.862	0.846	0.987	0.917	0.975	0.929	0.679	1	0.923	0.5	1
Spain	0.946	0.889	0.971	0.93	0.895	0.981	0.929	0.861	1	0.889	0.833	1

Figure 6.5: Example Workflow - general : Average spectra and Gini



The next page features a full map of the output and input to PRFFECT.



Troubleshooting and Known Bugs

Incorrect wavenumber scale on graphs - this can sometimes happen if the min_wavenumber & max_wavenumber are not set to a multiple of 100. It is purely a graphical issue, the underlying classification will still work. Occasionally, graphs will be produced with a "-" symbol in front of the wavenumbers on the x-axis. This is again just a graphical issue. It can sometimes appear when the input file has the wavenumber columns in reverse numerical order.

Some pre-processing combinations don't work . This issue can sometimes be found with certain combinations. It is due to mathematics and the incompatibility of certain combinations with some dataset sizes. It is very difficult to predict which methods will produce an error with a particular dataset.