

# Package ‘PLNseq’

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**Type** Package

**Title** PLNseq: A multivariate Poisson lognormal distribution for  
high-throughput correlated RNA-sequencing read count data

**Version** 1.0

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**Depends** R (>= 2.10), MASS

**Description** PLNseq is an R package for identifying differentially expressed  
genes using RNA-sequencing read count data from correlated samples.

**License** GPL (>= 2)

**LazyLoad** yes

**URL** <http://github.com/zhanghfd/PLNseq>

**BugReports** <http://github.com/zhanghfd/PLNseq/issues>

**RoxygenNote** 5.0.1

## R topics documented:

PLNseq-package . . . . .	2
commonSigma . . . . .	2
correlationCoefficient . . . . .	3
genewiseSigma . . . . .	4
LRtest1 . . . . .	4
LRtest2 . . . . .	5
lung . . . . .	6
PLNobject . . . . .	7
sizeFactor . . . . .	8
<b>Index</b>	<b>9</b>

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 PLNseq-package

*Differential expression analysis using matched read count data*


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### Description

This R package conducts differential expression (DE) analysis using high throughput next-generation sequencing read count data generated from correlated samples. The marginal distribution of the read count is the compounding of the Poisson distribution and the lognormal distribution ('PLN' distribution for short), and the correlation between the read counts of each matched sample set is modeled by the multivariate lognormal distribution with correlation coefficient matrix that is assumed to be common for all genes. This package provides estimates of rho (correlation coefficient matrix in multivariate lognormal distribution) and its standard error, common or genewise sigma (standard deviation of lognormal distribution), fold change (defined as the difference between log-gene expression of matched samples) and p-value for detecting differentially expressed genes.

### Details

Package: PLNseq  
 Type: Package  
 Version: 1.0  
 Date: 2014-06-18  
 License: GPL (>= 3)

### Author(s)

Hong Zhang

Maintainer: Hong Zhang <zhanghd@fudan.edu.cn>

### References

Zhang, H., Xu, J., Jiang N., Hu, X., and Luo, Z. (2015). PLNseq: A multivariate Poisson lognormal distribution for high-throughput matched RNA-sequencing read count data. *Statistics in Medicine* 34: 1577-1589.

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 commonSigma

*Common sigma*


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### Description

Estimate 'mu' (mean parameter lognormal distribution) for each gene and condition and a common 'sigma' (standard deviation parameter of lognormal distribution).

### Usage

commonSigma(d)

**Arguments**

d This is a PLNseq object.

**Value**

d\$commonSigma A common 'sigma'

**Examples**

```
data(lung);
count = lung[,c(2:4,8:10)];
d = PLNobject(count,2);
d = sizeFactor(d,maxCount=2e3);
d = commonSigma(d);
```

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correlationCoefficient

*Correlation coefficient*

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**Description**

Estimate correlation coefficient parameter and its standard error in the multivariate lognormal distribution.

**Usage**

```
correlationCoefficient(d)
```

**Arguments**

d This is a PLNseq object.

**Value**

d\$rho Correlation coefficient 'rho' in the multivariate lognormal distribution  
d\$rho.se Standard error of estimated 'rho'

**Examples**

```
data(lung);
count = lung[,c(2:4,8:10)];
d = PLNobject(count,2);
d = sizeFactor(d,maxCount=2e3);
d = commonSigma(d);

## common correlation
## commonCorrelation = TRUE;
## d = correlationCoefficient(d);

## clustered correlation
```

```
## commonCorrelation = FALSE;
## J = nrow(count);
## J1 = round(J/2);
## d$cluster = c(rep(1,J1),rep(2,J-J1));
## d = correlationCoefficient(d);
```

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genewiseSigma	<i>Genewise sigma</i>
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### Description

Estimate genewise ‘sigma’ (standard deviation parameter of lognormal distribution).

### Usage

```
genewiseSigma(d,w=25)
```

### Arguments

d	This is a PLNseq object.
w	Shrinkage parameter.

### Value

d\$genewiseSigma	Genewise ‘sigma’
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### Examples

```
data(lung);
count = lung[,c(2:4,8:10)];
d = PLNobject(count,2);
d = sizeFactor(d,maxCount=2e3);

d = genewiseSigma(d);
```

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LRtest1	<i>Likelihood ratio test for differential expression analysis with common correlation.</i>
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### Description

This function calculates log-fold changes, likelihood ratio test statistics, and p-values for a list of genes. This function should be called after a common correlation matrix is returned by ‘correlationCoefficient’.

### Usage

```
LRtest1(d,z,use.commonSigma,id)
```

**Arguments**

d	This is a PLNseq object.
z	J independent samples (a matrix of dimension J by R) drawn from multivariate normal distribution with expectations 0, variances 1, and a common correlation coefficient matrix estimated by 'correlationCoefficient'.
use.commonSigma	Use common 'sigma' (TRUE) or genewise 'sigma' (FALSE), with default value 'FALSE'.
id	A vector consisting of a subset of 1,...,J, with default value 1:J.

**Value**

LR	Estimation and test results: 'log-FC', 'LR statistic', 'p value'.
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**Examples**

```
data(lung);
count = lung[,c(2:4,8:10)];
d = PLNobject(count,conditionNumber=2);
d = sizeFactor(d,maxCount=2e3);

## Not run:
## d = commonSigma(d);
## d$commonCorrelation = TRUE;
## d = correlationCoefficient(d);
## d = genewiseSigma(d);
## library(MASS);
## z = mvrnorm(n=1e5,mu=rep(0,2),Sigma=d$rho);
## d = LRtest1(d,z,use.commonSigma=FALSE,id=1:100);
```

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LRtest2	<i>Likelihood ratio test for differential expression analysis with cluster-specific correlations.</i>
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**Description**

This function calculates log-fold changes, likelihood ratio test statistics, and p-values for a list of genes. This function should be called after cluster-specific correlations are returned by 'correlationCoefficient'.

**Usage**

```
LRtest2(d,M,use.commonSigma,id)
```

**Arguments**

d	This is a PLNseq object.
M	The number of simulations used in Monte-Carlo method for calculating likelihood ratio test statistics.

`use.commonSigma` Use common 'sigma' (TRUE) or genewise 'sigma' (FALSE), with default value 'FALSE'.

`id` A vector consisting of a subset of 1,...,J, with default value 1:J.

### Value

LR Estimation and test results: 'log-FC', 'LR statistic', 'p value'.

### Examples

```
data(lung);
count = lung[,c(2:4,8:10)];
d = PLNobject(count,conditionNumber=2);
d = sizeFactor(d,maxCount=2e3);

## Not run:
## d = commonSigma(d);
## J = nrow(count);
## J1 = round(J/2);
## d$commonCorrelation = FALSE;
## d$cluster = c(rep(1,J1),rep(2,J-J1));
## d = correlationCoefficient(d);
## d = genewiseSigma(d);
## d = LRtest2(d,M=3e4,use.commonSigma=FALSE,id=1:100);
```

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lung	<i>Lung cancer data</i>
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### Description

The data are from a study of the lung cancer. Six patients provided tissue samples and normal samples besides the lung tissues. The read counts were summarized by RefSeq transcript, and only those transcripts with at least 50 aligned reads for at least one tissue in each condition were provided in the table. RefSeq identifiers were mapped to the latest official gene symbols by following the user guide of the Bioconductor package 'edgeR' using the Bioconductor annotation package 'org.Hs.eg.db' (version 2.7.1). Those RefSeq identifiers not in the database were discarded, and each gene was represented by the RefSeq transcript with the greatest number of exons and the other transcripts were removed. Altogether 11,597 transcripts (genes) were kept.

### Usage

```
data(lung)
```

### Format

A data frame with 11,597 observations on the following 13 variables.

`nameOfGene` Gene name

`N4` Read count for normal sample of patient 4

`T4` Read count for normal sample of patient 4

N12 Read count for normal sample of patient 12  
 T12 Read count for tumor sample of patient 12  
 N13 Read count for normal sample of patient 13  
 T13 Read count for tumor sample of patient 13  
 N14 Read count for normal sample of patient 14  
 T14 Read count for tumor sample of patient 14  
 N15 Read count for normal sample of patient 15  
 T15 Read count for tumor sample of patient 15  
 N16 Read count for normal sample of patient 16  
 T16 Read count for tumor sample of patient 16

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 PLNobject

*PLN object*


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## Description

Create a PLN object, a list containing a read count matrix 'count' and sample description matrix 'sample'.

## Usage

```
PLNobject(count, conditionNumber)
```

## Arguments

count	This is a matrix containing the read counts of $R \times I$ samples at $J$ genes ( $R$ is the number of conditions in each matched sample set and $I$ is the number of sample sets). Here columns 1 through $I$ are for $I$ independent samples from condition 1, columns $I+1$ through $2I$ are for $I$ samples from condition 2 matched by samples 1 through $I$ , ... , columns $(R-1) \times I + 1$ through $R \times I$ are for $I$ samples from condition $R$ matched by samples 1 through $I$ .
conditionNumber	Number of conditions.

## Value

d\$count	Original read count matrix
d\$conditionNumber	The number of conditions
d\$sample	A matrix of sample information: 'SampleName', 'TotalCount', 'MedianCount'

## Examples

```

data(lung);
count = lung[,c(2:4,8:10)];
d = PLNobject(count,2);

```

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sizeFactor	<i>Estimate size factor for each sample.</i>
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**Description**

Estimate size factor for each sample using median normalization method.

**Usage**

```
sizeFactor(d,maxCount)
```

**Arguments**

d	This is a PLNseq object.
maxCount	The maximal count after shrinkage, with a default value NA (no shrinkage).

**Value**

```
d$sample$sizeFactor
```

Estimated size factors

**Examples**

```
data(lung);  
count = lung[,c(2:4,8:10)];  
d = PLNobject(count,conditionNumber=2);  
d = sizeFactor(d,maxCount=2e3);
```



# Index

- \*Topic **RNA-seq read count**
  - PLNobject, [7](#)
- \*Topic **cluster-specific correlation**
  - LRtest2, [5](#)
- \*Topic **common correlation**
  - LRtest1, [4](#)
- \*Topic **correlated read count**
  - LRtest1, [4](#)
  - LRtest2, [5](#)
- \*Topic **correlation coefficient**
  - correlationCoefficient, [3](#)
- \*Topic **datasets**
  - lung, [6](#)
- \*Topic **differential expression**
  - LRtest1, [4](#)
  - LRtest2, [5](#)
- \*Topic **likelihood ratio test**
  - LRtest1, [4](#)
  - LRtest2, [5](#)
- \*Topic **lognormal distribution**
  - commonSigma, [2](#)
  - genewiseSigma, [4](#)
- \*Topic **matched sample**
  - PLNobject, [7](#)
- \*Topic **mean**
  - commonSigma, [2](#)
  - genewiseSigma, [4](#)
- \*Topic **median normalization**
  - sizeFactor, [8](#)
- \*Topic **package**
  - PLNseq-package, [2](#)
- \*Topic **size factor**
  - sizeFactor, [8](#)
- \*Topic **standard deviation**
  - commonSigma, [2](#)
  - genewiseSigma, [4](#)

commonSigma, [2](#)  
correlationCoefficient, [3](#)  
  
genewiseSigma, [4](#)  
  
LRtest1, [4](#)  
LRtest2, [5](#)  
  
lung, [6](#)  
  
PLNobject, [7](#)  
PLNseq-package, [2](#)  
  
sizeFactor, [8](#)