STATS 3DA3

Homework Assignment 6

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1. Classification Problem Identification: Define and describe a classification problem based on the dataset.

```
import numpy as np
import pandas as pd
import matplotlib.pyplot as plt
import seaborn as sns
from ucimlrepo import fetch_ucirepo
import warnings
warnings.filterwarnings('ignore')

# fetch dataset
chronic_kidney_disease = fetch_ucirepo(id=336)

# data (as pandas dataframes)
X = chronic_kidney_disease.data.features
y = chronic_kidney_disease.data.targets
ckd = pd.concat([chronic_kidney_disease.data.features ,chronic_kidney_disease.data.targets],ax
print(chronic_kidney_disease.variables)
```

\	description	${\tt demographic}$	type	role	name	
	None	Age	Integer	Feature	age	0
	blood pressure	None	Integer	Feature	bp	1
	specific gravity	None	Categorical	Feature	sg	2
	albumin	None	Categorical	Feature	al	3
	sugar	None	Categorical	Feature	su	4
	red blood cells	None	Binary	Feature	rbc	5
	pus cell	None	Binary	Feature	рс	6
	pus cell clumps	None	Binary	Feature	рсс	7
	bacteria	None	Binary	Feature	ba	8
	blood glucose random	None	Integer	Feature	bgr	9

10	bu	Feature	Integer	None	blood urea
11	sc	Feature	Continuous	None	serum creatinine
12	sod	Feature	Integer	None	sodium
13	pot	Feature	Continuous	None	potassium
14	hemo	Feature	Continuous	None	hemoglobin
15	pcv	Feature	Integer	None	packed cell volume
16	wbcc	Feature	Integer	None	white blood cell count
17	rbcc	Feature	Continuous	None	red blood cell count
18	htn	Feature	Binary	None	hypertension
19	dm	Feature	Binary	None	diabetes mellitus
20	cad	Feature	Binary	None	coronary artery disease
21	appet	Feature	Binary	None	appetite
22	pe	Feature	Binary	None	pedal edema
23	ane	Feature	Binary	None	anemia
24	class	Target	Binary	None	ckd or not ckd

units missing_values

0	year	yes
1	mm/Hg	yes
2	None	yes
3	None	yes
4	None	yes
5	None	yes
6	None	yes
7	None	yes
8	None	yes
9	mgs/dl	yes
10	mgs/dl	yes
11	mgs/dl	yes
12	mEq/L	yes
13	mEq/L	yes
14	gms	yes

15	None	yes
16	cells/cmm	yes
17 n	millions/cmm	yes
18	None	yes
19	None	yes
20	None	yes
21	None	yes
22	None	yes
23	None	yes
24	None	no

ckd.head(10)

	age	bp	sg	al	su	rbc	pc	pcc	ba	bgr	•••	pcv	wbcc
0	48.0	80.0	1.020	1.0	0.0	NaN	normal	notpresent	notpresent	121.0		44.0	7800.0
1	7.0	50.0	1.020	4.0	0.0	NaN	normal	notpresent	notpresent	NaN		38.0	6000.0
2	62.0	80.0	1.010	2.0	3.0	normal	normal	notpresent	notpresent	423.0		31.0	7500.0
3	48.0	70.0	1.005	4.0	0.0	normal	abnormal	present	notpresent	117.0		32.0	6700.0
4	51.0	80.0	1.010	2.0	0.0	normal	normal	notpresent	notpresent	106.0		35.0	7300.0
5	60.0	90.0	1.015	3.0	0.0	NaN	NaN	notpresent	notpresent	74.0		39.0	7800.0
6	68.0	70.0	1.010	0.0	0.0	NaN	normal	notpresent	notpresent	100.0		36.0	NaN
7	24.0	NaN	1.015	2.0	4.0	normal	abnormal	notpresent	notpresent	410.0		44.0	6900.0
8	52.0	100.0	1.015	3.0	0.0	normal	abnormal	present	notpresent	138.0		33.0	9600.0
9	53.0	90.0	1.020	2.0	0.0	abnormal	abnormal	present	notpresent	70.0		29.0	12100.0

From the dataset we've been given, our main task is to determine whether someone has chronic kidney disease (CKD) or not. The dataset includes various measurements and test results which might help us identify whether a patient has CKD.

2. Variable Transformation: Implement any transformations chosen or justify the absence of such modifications.

To prepare the data for analysis

- categorical variables should be translated into numerical variables. LableEncoder is used.
- Records with missing values is removed

```
from sklearn.preprocessing import LabelEncoder

# remove NaN values
ckd_transform = ckd.dropna()

# Applying label encoding to categorical variables
label_encoders = {}
categorical_col = list(ckd_transform.select_dtypes(include=['object']).columns)
le = LabelEncoder()
for col in categorical_col:
    if col == 'class':
        ckd_transform[col] = ckd_transform[col].apply(lambda x: 0 if 'not' in x else 1)
    else:
        ckd_transform[col] = le.fit_transform(ckd_transform[col].astype(str))
```

ckd_transform.head(10)

	age	bp	sg	al	su	rbc	pc	pcc	ba	bgr	 pcv	wbcc	rbcc	htn	dm	cad	ap
3	48.0	70.0	1.005	4.0	0.0	1	0	1	0	117.0	 32.0	6700.0	3.9	1	0	0	1
9	53.0	90.0	1.020	2.0	0.0	0	0	1	0	70.0	 29.0	12100.0	3.7	1	1	0	1
11	63.0	70.0	1.010	3.0	0.0	0	0	1	0	380.0	 32.0	4500.0	3.8	1	1	0	1
14	68.0	80.0	1.010	3.0	2.0	1	0	1	1	157.0	 16.0	11000.0	2.6	1	1	1	1
20	61.0	80.0	1.015	2.0	0.0	0	0	0	0	173.0	 24.0	9200.0	3.2	1	1	1	1
22	48.0	80.0	1.025	4.0	0.0	1	0	0	0	95.0	 32.0	6900.0	3.4	1	0	0	0
27	69.0	70.0	1.010	3.0	4.0	1	0	0	0	264.0	 37.0	9600.0	4.1	1	1	1	0
48	73.0	70.0	1.005	0.0	0.0	1	1	0	0	70.0	 29.0	18900.0	3.5	1	1	0	0
58	73.0	80.0	1.020	2.0	0.0	0	0	0	0	253.0	 33.0	7200.0	4.3	1	1	1	0
71	46.0	60.0	1.010	1.0	0.0	1	1	0	0	163.0	 28.0	14600.0	3.2	1	1	0	0

3. **Dataset Overview:** Provide a detailed description of the dataset, covering variables, summaries, observation counts, data types, and distributions (at least three statements).

print(ckd.shape)

(400, 25)

ckd.dtypes

age	float64
bp	float64
sg	float64
al	float64
su	float64
rbc	object
pc	object
pcc	object
ba	object
bgr	float64
bu	float64
sc	float64
sod	float64
pot	float64
hemo	float64
pcv	float64
wbcc	float64
rbcc	float64
htn	object
dm	object
cad	object
appet	object
pe	object

ane object class object

dtype: object

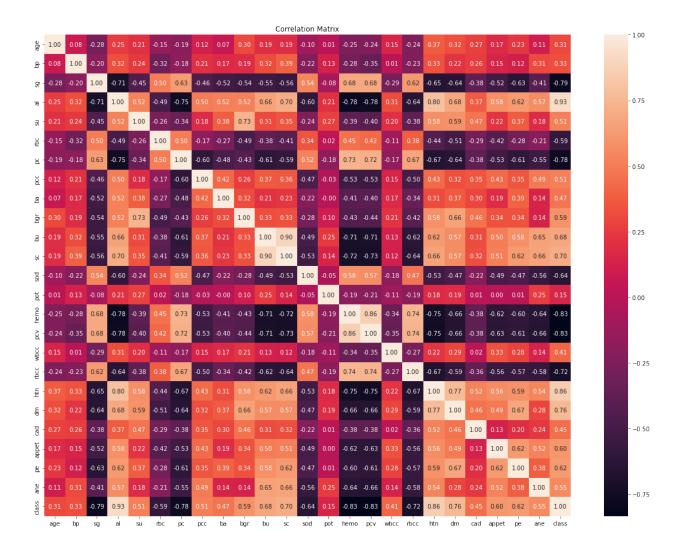
ckd.describe(include='all').transpose()

	count	unique	top	freq	mean	std	min	25%	50%	75%	max
age	391.0	NaN	NaN	NaN	51.483376	17.169714	2.0	42.0	55.0	64.5	90.0
bp	388.0	NaN	NaN	NaN	76.469072	13.683637	50.0	70.0	80.0	80.0	180.0
sg	353.0	NaN	NaN	NaN	1.017408	0.005717	1.005	1.01	1.02	1.02	1.02
al	354.0	NaN	NaN	NaN	1.016949	1.352679	0.0	0.0	0.0	2.0	5.0
su	351.0	NaN	NaN	NaN	0.450142	1.099191	0.0	0.0	0.0	0.0	5.0
rbc	248	2	normal	201	NaN	NaN	NaN	NaN	NaN	NaN	NaN
pc	335	2	normal	259	NaN	NaN	NaN	NaN	NaN	NaN	NaN
pcc	396	2	notpresent	354	NaN	NaN	NaN	NaN	NaN	NaN	NaN
ba	396	2	notpresent	374	NaN	NaN	NaN	NaN	NaN	NaN	NaN
bgr	356.0	NaN	NaN	NaN	148.036517	79.281714	22.0	99.0	121.0	163.0	490.0
bu	381.0	NaN	NaN	NaN	57.425722	50.503006	1.5	27.0	42.0	66.0	391.
sc	383.0	NaN	NaN	NaN	3.072454	5.741126	0.4	0.9	1.3	2.8	76.0
sod	313.0	NaN	NaN	NaN	137.528754	10.408752	4.5	135.0	138.0	142.0	163.0
pot	312.0	NaN	NaN	NaN	4.627244	3.193904	2.5	3.8	4.4	4.9	47.0
hemo	348.0	NaN	NaN	NaN	12.526437	2.912587	3.1	10.3	12.65	15.0	17.8
pcv	329.0	NaN	NaN	NaN	38.884498	8.990105	9.0	32.0	40.0	45.0	54.0
wbcc	294.0	NaN	NaN	NaN	8406.122449	2944.47419	2200.0	6500.0	8000.0	9800.0	2640
rbcc	269.0	NaN	NaN	NaN	4.707435	1.025323	2.1	3.9	4.8	5.4	8.0
htn	398	2	no	251	NaN	NaN	NaN	NaN	NaN	NaN	NaN
dm	398	3	no	260	NaN	NaN	NaN	NaN	NaN	NaN	NaN
cad	398	2	no	364	NaN	NaN	NaN	NaN	NaN	NaN	NaN
appet	399	2	good	317	NaN	NaN	NaN	NaN	NaN	NaN	NaN
pe	399	2	no	323	NaN	NaN	NaN	NaN	NaN	NaN	NaN
ane	399	2	no	339	NaN	NaN	NaN	NaN	NaN	NaN	NaN
class	400	3	ckd	248	NaN	NaN	NaN	NaN	NaN	NaN	NaN

count unique top freq mean std min 25% 50% 75% max

- Observation Count and Variables: The dataset contains 158 rows and 25 columns
- There are missing values (NaN) in many columns. For example, several key variables such as sg (specific gravity), al (albumin), and su (sugar) showed substantial missing data in observations. This is critical because they are important for diagnosing kidney diseases.
- Data Types: Most variables related to medical measurements like blood pressure (bp), glucose levels (bgr), and others are floating point numbers. Variables like rbc (red blood cells) and pc (pus cell) are categorical, initially with text values which we converted to numerical codes.
- 4. **Association Between Variables:** Analyze variable relationships and their implications for feature selection or extraction (at least three statements).

```
correlation_matrix = ckd_transform.corr()
plt.figure(figsize=(20,15))
sns.heatmap(correlation_matrix, annot=True, fmt=".2f")
plt.title('Correlation Matrix')
plt.show()
```



Answer:

Hemoglobin and Packed Cell Volume (PCV): There is a very strong positive correlation between 'hemo' (hemoglobin) and 'pcv' (packed cell volume). Since both are indicators of the blood's ability to carry oxygen, they typically increase or decrease together.

Red Blood Cell Count (RBCC) and Hemoglobin (Hemo): A strong positive correlation exists between 'rbcc' and 'hemo'. These measures are linked to the blood's oxygen-carrying capacity, and typically, a lower hemoglobin level corresponds with a reduced red blood cell count.

Albumin (Al) and Blood Urea (Bu): A positive correlation is observed between these variables, suggesting that higher albumin levels in the urine, which may indicate reduced kidney function, are associated with increased blood urea levels due to the kidney's diminished ability to filter urea.

5. Missing Value Analysis and Handling: Implement your strategy for identifying and addressing missing values in the dataset, or provide reasons for not addressing them.

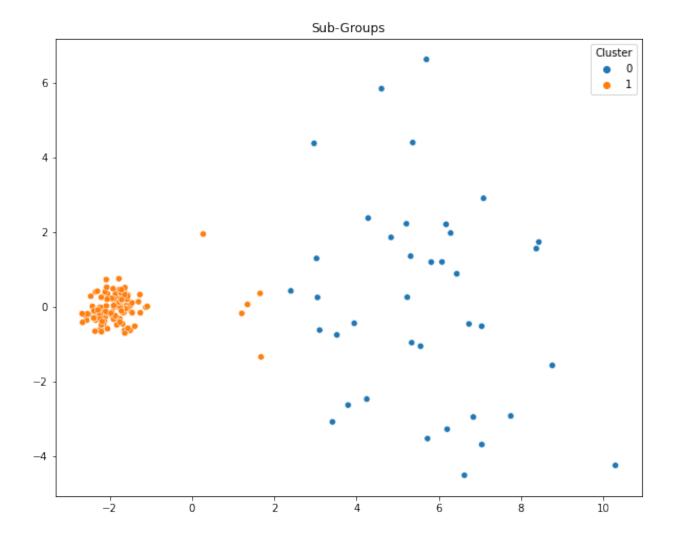
We should remove all the records that has missing values, instad of using imputed values, for the following reasons:

- Imputation introduces estimated values based on assumptions or patterns observed in other data. In health contexts, where every variable can be crucial and highly individual-specific, these estimates might not accurately represent the true state or condition of a patient. This could lead to incorrect conclusions or treatment decisions.
- Imputation methods themselves may introduce bias, particularly if the missing value is not random but related to certain unseen factors. If the method or the data used for imputation does not account for these factors accurately, it can skew the model's outputs.
- By removing incomplete records, it ensures that the model only learns from the most reliable, fully observed datasets. This is particularly important in clinical trials or scenarios where data quality is paramount for accurate predictions.
- 6. **Outlier Analysis:** Implement your approach for identifying and managing outliers, or provide reasons for not addressing them.

We should keep the outlier, because:

- Outliers might signify rare diseases or unique complications that are not common in the general population. Removing these outliers could prevent the model from learning how to identify and treat these rare conditions.
- A predictive model trained on a dataset that includes outliers might develop a more robust understanding of the full spectrum of possible medical scenarios.
- 7. **Sub-group Analysis:** Explore potential sub-groups within the data, employing appropriate data science methods to find the sub-groups of patients and visualize the sub-groups. The sub-group analysis must not include the labels (for CKD patients and healthy controls).

```
import matplotlib.pyplot as plt
import seaborn as sns
from sklearn.preprocessing import StandardScaler
from sklearn.cluster import KMeans
from sklearn.decomposition import PCA
X = ckd_transform.drop(columns='class')
y = ckd_transform['class']
norm_transformer = StandardScaler()
X_normalized = norm_transformer.fit_transform(X)
# Configure and apply K-Means clustering
cluster_model = KMeans(n_clusters=2, random_state=1)
X_clusters = cluster_model.fit_predict(X_normalized)
# Setup and apply PCA
pca_transformer = PCA(n_components=2)
pca_results = pca_transformer.fit_transform(X_normalized)
# Plot the clustered data
plt.figure(figsize=(10, 8))
sns.scatterplot(x=pca_results[:, 0], y=pca_results[:, 1], hue=X_clusters)
plt.title('Sub-Groups')
plt.legend(title='Cluster')
plt.show()
```



Use the K-means and PCA, it can be seen that there are 1 clear subgroup of the data as labeled orange in the chart.

8. **Data Splitting:** Segregate 30% of the data for testing, using a random seed of 1. Use the remaining 70% for training and model selection.

```
from sklearn.model_selection import train_test_split

X = ckd_transform.drop('class', axis=1)

y = ckd_transform['class']

X_train, X_test, y_train, y_test = train_test_split(X, y, test_size=0.3, random_state=100, strain)
```

9. Classifier Choices: Identify the two classifiers you have chosen and justify your selections.

- Logistic Regression: Logistic regression provides coefficients that can be directly interpreted in terms of odds ratios for understanding the impact of each feature.
- Random Forest: Random forests can capture complex interactions between features without the need for feature engineering, as they are non-linear models.
- 10. **Performance Metrics:** Outline the two metrics for comparing the performance of the classifiers.
 - AUC (Area Under the Receiver Operating Characteristic Curve): Measures the entire area under the ROC curve, which evaluates the trade-off between true positive rate and false positive rate across all classification thresholds, offering a robust assessment especially beneficial for imbalanced datasets.
 - Accuracy Socre: Calculates the ratio of correct predictions (true positives and true negatives) to total observations, providing an easy-to-understand metric that's most effective for balanced datasets but can be misleading if the data is imbalanced.
- 11. **Feature Selection/Extraction:** Implement methods to enhance the performance of at least one classifier in (9). The answer for this question can be included in (12).

We can normalize the numerical predictors.

12. Classifier Comparison: Utilize the selected metrics to compare the classifiers based on the test set. Discuss your findings (at least two statements).

```
from sklearn.linear_model import LogisticRegression
from sklearn.ensemble import RandomForestClassifier
from sklearn.metrics import roc_auc_score, accuracy_score

# Initialize the two classifiers
log_reg = LogisticRegression()
random_forest = RandomForestClassifier(random_state=1)

# Train the logistic regression classifier
```

```
log_reg.fit(X_train, y_train)
# Train the random forest classifier
random forest.fit(X train, y train)
# Make predictions with both classifiers
y_pred_lr = log_reg.predict(X_test)
y_pred_rf = random_forest.predict(X_test)
y_pred_proba_lr = log_reg.predict_proba(X_test)[:, 1]
y_pred_proba_rf = random_forest.predict_proba(X_test)[:, 1]
# Calculate ROC AUC
roc_auc_lr = roc_auc_score(y_test, y_pred_proba_lr)
roc_auc_rf = roc_auc_score(y_test, y_pred_proba_rf)
# Calculate F1 Scores
acc_score_lr = accuracy_score(y_test, y_pred_lr)
acc_score_rf = accuracy_score(y_test, y_pred_rf)
print(f'Logistic Regression ROC score: {roc_auc_lr:.4f}')
print(f'Random Forest ROC score: {roc_auc_rf:.4f}')
print(f'Logistic Regression accuracy score: {acc_score_lr:.4f}')
print(f'Random Forest accuracy score: {acc score rf:.4f}')
```

```
Logistic Regression ROC score: 0.9978
Random Forest ROC score: 1.0000
```

Logistic Regression accuracy score: 0.9583

Random Forest accuracy score: 1.0000

Compare the result, Random Forest is better than logestic regression. However, given the 100% accurate rate, it indicatest that random forest could be over-fitted.

```
# normalize X
norm_transformer = StandardScaler()
X_normalized = norm_transformer.fit_transform(X)
X_train, X_test, y_train, y_test = train_test_split(X_normalized, y, test_size=0.3, random_sta
# redo logistic regression
# Initialize the two classifiers
log_reg = LogisticRegression()
log_reg.fit(X_train, y_train)
y_pred_lr = log_reg.predict(X_test)
y_pred_proba_lr = log_reg.predict_proba(X_test)[:, 1]
# Calculate ROC AUC
roc_auc_lr = roc_auc_score(y_test, y_pred_proba_lr)
# Calculate F1 Scores
acc_score_lr = accuracy_score(y_test, y_pred_lr)
print(f'Logistic Regression ROC score: {roc_auc_lr:.4f}')
print(f'Logistic Regression accuracy score: {acc_score_lr:.4f}')
```

Logistic Regression ROC score: 1.0000
Logistic Regression accuracy score: 1.0000

Normalization improved the logistic regression.

13. **Interpretable Classifier Insight:** After re-training the interpretable classifier with all available data, analyze and interpret the significance of predictor variables in the context of the data and the challenge (at least two statements).

```
feature_importance = pd.DataFrame(log_reg.coef_[0], index=X_train.columns, columns=['Coefficient']
print(feature_importance.sort_values(by='Coefficient', ascending=False))
```

	Coefficient
bu	0.262789
bgr	0.138596
al	0.103505
sc	0.068797
bp	0.052720
ane	0.022546
рсс	0.022447
htn	0.020422
pe	0.020112
appet	0.019206
age	0.018603
su	0.003776
ba	0.003511
sg	0.001972
wbcc	0.001658
dm	0.001412
cad	0.000149
rbc	-0.001143
rbcc	-0.007020
pc	-0.020369
pot	-0.040813
hemo	-0.052248
pcv	-0.212403
sod	-0.325618

From the coefficient parameters, we can observe that: * The negative coefficient for sodium levels (sod) suggests that higher serum sodium concentrations are associated with a lower likelihood of having CKD. This could indicate that lower sodium levels, possibly reflecting issues with the kidneys' ability to balance minerals and electrolytes, are a marker of kidney dysfunction.

• Similarly, a negative coefficient for packed cell volume implies that lower pcv values are linked with a higher risk of CKD. This relationship might be due to anemia, which is common in

kidney disease because of decreased erythropoietin production by the kidneys.

• The positive coefficient for blood glucose random levels (bgr) indicates that higher glucose

levels are a predictor of CKD. This aligns with the understanding that diabetes, a leading

cause of CKD, is characterized by high blood sugar levels, which over time, can damage the

kidneys.

15. Team Contributions: Document each team member's specific contributions related to the

questions above.

• Question 1-8: Zhen Ye

• Question 9-16: Jianghan Sun

16. Link to the public GitHub repository.

https://github.com/zhenye4003/STATS3DA-A6

Grading scheme

1.	Answer [1]
2.	Codes [2]
	OR answer [2]
3.	Codes [3] and answer [3]
4.	Codes [2] and answer [3]
5.	Codes [2]
	OR answer [2]
6.	Codes [2]
	OR answer [2]
7.	Codes [3] and Plot [1]
8.	Codes [1]
9.	Answers [2]
10.	Describe the two metrics [2]
11.	Codes [2]
	these codes can be included in (12)
12.	Codes (two classifiers training,
	model selection for each classifier,
	classifiers comparisons) $[5]$ and answer $[2]$
13.	Codes [1] and answers [2]
14.	Codes and comparison will
	give bonus 2 points for the final grade.

The maximum point for this assignment is 39. We will convert this to 100%.

All group members will receive the same grade if they contribute to the same.