Nuclear Magnetic Resonance (NMR) Spectroscopy Protocol

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Interpretation

For standard 1H NMR spectrum, there are three pieces of data we would like to see:

- (1) **Peak position**: Chemical shift (in ppm) of each peak
- (2) **Peak Area**: integration
- (3) **Peak "Splitting"**: coupling (e.g. doublet, J = 2.0 Hz)

Pre-NMR Steps (Organic Synthesis)

- 1. Obtain a clean, small 50 mL Round Bottom Flask (RBF), and weigh it **before** putting anything inside to determine the product yield. Be careful of weighing because of small yields.
- 2. After TLC, combine all fractions with same compounds into the RBF. When combining fractions, rinse the original container with small amounts of solvent and add the rinsing to the RBF.
- 3. Rotovap the combined fractions.
- 4. Weigh the RBF again to determine the yield for each product.
- 5. Dissolve sample in \sim 0.5 mL deuterated solvent (CDCl₃). Sample shouldn't be too much (diluted) or be too little; around three fingers in height in the tube is sufficient.
- 6. Carefully transfer sample to a clean, dry NMR tube. 3 fingers of liquid. Cap, <u>label</u>, and take down to the NMR room. Do not invert tube!
- 7. Before going to the NMR room, check SciFinder database for similar spectrum come prepared!

NMR Setup

- 1. Log in with jrscheerer, or jrsche, password: Chemi\$try42
- 2. **e** eject sample already in the NMR, then use kimwipe to touch turbines and transit samples you prepared.
- 3. **i** insert sample.
- 4. Enter sample name (initials, book number, page number, crude/pure, fractions in sample). Example: HH_kenone_frac22_23_10_13_2017. Put name under **start** tab. In the **sample name** space, and **comments box**.
- 5. Select solvent CDCl₃ or proper solvent being used.
- 6. Lock: go to lock, unclick the checked box next to lock, click **find z0**.
- 7. Shim gradient shim, the good number should be at least around 30-40.
- 8. Select experiment: proton NMR or carbon NMR. To load the experiment, drag the correct experiment from the left-hand options bar into the window. Set nt (number of transients) and bs (block size) nt=32 and bs = 4. (nt=3200 for carbon)
- 9. **ga** start the experiment.

Data Processing (H MNR)

Type commands: wft aph vsadj cz f
wft - fourier transform and view spectra
aph - autophase spectrum
vsadj - vertical scale adjustment
cz - clear zeros for integrals
f - full spectrum (-2 ppm to 15 ppm)

- 2. Spectrum reference: standard reference peak in CDCl₃ will be at 7.26. Click ~7.26 and type **nl** to select the nearest line, and then type **rl(7.26p)** to set the reference line. Reference TMS peak at 0 ppm also.
- 3. Integration: Full Spectrum icon turn on **integral** and use **cutting tool** to chop up into appropriate pieces. Choose a peak with known integration area, go to the integration tab, and enter the number (known area) where it says integral area.
- 4. Set the size of the spectra by typing sp=-0.5p wp=9.5p vsadj \rightarrow jrsstd

Print and Save

 Go to auto plot preview and print. Print parameters in horizontal box: click drop down menu by display parameters and select horizontal box. Or pl pscale ptext page pl – print line

pscale - print scale
ptext - print associated text
page - actually prints your spectra

- 2. svf('filename')
- 3. **e** (eject sample) and **i** (insert the standard CDCl₃).

Other

1. if NMR is not responding, and won't abort when type **aa**, go to **terminal** under **applications**, type **su acqproc** and retype it when something appears and type **exit**.

Post NMR

- 1. Use DCM to rinse glass pipet used to transfer sample back into vial or sample container
- 2. Pour contents of NMR tube back into sample container
- 3. Rinse NMR with DCM and deposit rinses in sample container
- 4. Rotovap sample back down
- 5. Rinse NMR tube at least 3x with acetone and put back in oven

Getting NMR Spectra Off the Computer

- 1. Open NMR software
- 2. Find and open your **saved.fid** file on the little window on the left side of the screen
- 3. At the top of the screen, click **Utilities** → **Printers** → **Plotter** drop down menu → Select "eps" → OK
- 4. Type: pl pscale page ('YourFileNameHere.ps')
- 5. If you want to get the chemical shifts too, you can type **pl pscale ppf page**...
- 6. Make sure you type the parentheses and apostrophes
- 7. Go to desktop and open the folder for dyoung
- 8. Find your .ps file, and copy/paste to your flash drive
- 9. At your computer, convert files from postscript files to pdf: http://ps2pdf.com/convert.htm