14-0421 PAH/BIOMARKERs QA/QC Summary

Project:	ANIMIDA III		
Parameters:	PAH and Biomarkers		
Laboratory:	Battelle, Norwell, MA		
Matrix:	Tissue		
Data Set:	DP-14-0587		
Analytical SOP:	5-157		
Method Reference:	Modified EPA Method 8270D		
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	Receipt Date	Temp (°C)	
Sample Custody	8/14/2014	4.0	
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Corrective Actions	None.		
Sample Storage	The samples were stored in an access-limited freezer until sample preparation		
	could begin.		
	METHOD SUMMARIES		
Sample	Tissue samples were homogenized with titanium blades and split for metals		
Preparation	analysis at Sequim and FIT.		
	The tissue samples were extracted following a modified EPA Method 3510C.		
	Samples were prepared for analysis by w	veighing approximately 20 grams of	
	sample material into a pre-cleaned extraction vessel and dried using sodium		
	sulfate. Each sample was spiked with PA	AH, Biomarker and SHC surrogates and	
	extracted 3 times using methylene chlori	ide by tissuemizer. The combined	
	extracts were dried over sodium sulfate and concentrated by Kuderna-Danish		
	(KD) and nitrogen evaporation techniques. Sample clean-up was performed on		
	the extracts using alumina columns. Extr	acts were further cleaned up and	
	fractionated using silica gel columns. The F1 fraction was collected and split for		
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	TPH/SHC and biomarker analyses. The F2 fraction was collected for PAH and		
	alkyated PAH analysis. The extracts were concentrated and spiked with IS for		
	analysis.	policy to complex MESES MESES	
	The batch average dry weight is being applied to samples M5856, M5857,		
	M5859,M5860, M5862, M5863, M5865, M5866, M5867, M5869, M5870,		
	M5871, M5880, M5903 and M5904 as there was not enough material		
Prep comments	to perform separate dry weights.		
	Another NSC for MS was added to batch because original NSC was made with		
	ID61. 1 mL of IC44 used to make NSC.		
Analysis	PAH, alkylated PAH (F2 fraction) and Biomarkers (F1 fraction) were measured by		
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	gas chromatography-mass spectrometry (GC/MS) in the selected ion mode (SIM).		
	An initial calibration consisting of target analytes was analyzed prior to analysis		
	to demonstrate the linear range of analysis. Calibration verification was		
	performed every 24 hours in which samples were analyzed. Concentrations of		
	target compounds were calculated versus internal standards. Target PAH were		
	guantified using the average response factors (DE) generated from the initial		

quantified using the average response factors (RF) generated from the initial

14-0421 PAH/BIOMARKERs QA/QC Summary

	calibration. The alkyl homologue PAH series were assigned the RF of the parent PAH. Biomarkers used RFs from the single individual biomarkers within the calibration standard curve. All reported data (except NSC) is corrected based on surrogate recoveries. All data is reported on dry weight basis except the SRM (wet weight) and NSC and CO (oil weight).	
Analysis comments	None.	
Holding Times	Extraction Date(s)	Analysis Date(s)
	10/17, 29/2014 & 11/19, 20/2014	11/7-9, 19, 21-24/2014

Procedural Blank (PB)	A PB was prepared with this analytical batch to ensure the sample
	extraction and analysis methods are free of contamination.
PB <5 X MDL	Many exceedences noted.
Samples must be >5x PB	Comments: There were thirty-three exceedences for analytes
	detected in samples at less than five times the blank concentration.
	Reanalysis of the PB confirmed results. These were mostly
	Naphthalene, C1-Naphthalenes, and C2-Naphthalenes.

Laboratory Control Spike (LCS)	A LCS was prepared with this analytical batch. The percent	
	recoveries of target analytes were calculated to measure accuracy.	
Recovery of 70-130%	No exceedences noted.	
	Comments: None.	
Surrogate Recovery	Surrogate compounds were added prior to extraction. The surrogate	
	recoveries are calculated to measure extraction efficiency.	
Recovery of 40-120%	Three exceedences noted.	
	Comments: Surrogate recoveries for 5B(H)-Cholane were high in the	
	CO and in samples M5856 and M5880. Reanalysis on another	
	instrument confirmed the results. Prep records and integrations	
	were verified. All other surrogate recoveries were acceptable.	
Sample Duplicate (QADUP)	A QADUP was prepared with this analytical batch. The RPD of target	
	analytes were calculated to measure data quality in terms of	
	accuracy.	
Relative Percent Difference	No exceedences noted.	
(RPD) < 30%	Comments: None.	
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Standard Reference Material	An SRM was prepared with this batch to assess accuracy of the	
(SRM)	analytical procedures.	
< 30 PD from target concentration	One exceedence noted.	
and the 95% confidence level	Comments: Benzo(a)anthracene was recovered low in the SRM.	
analyte concentration must be > 5x the MDL. Concentration must	Results were confirmed by reanalysis on another instrument. Prep	
be certified and >5x the MDL for	records and integrations were verified. Recoveries for this analyte	
be certified and >3x the MDE for	were acceptable in all CCVs as well as the LCS for this batch. No	

14-0421 PAH/BIOMARKERs QA/QC Summary

MQO to apply	further action was taken.
North Slope Crude (NSC)	A NSC Reference Oil was prepared with this batch to evaluate the instrumental accuracy and also provide petroleum pattern information, aiding in the qualitative identification of target analytes.
	The control oil also run in this batch has no associated target values and is not evaluated.
< 30% RPD for 90% of analytes	No exceedences noted.
·	Comments: None.
Initial Calibration (ICAL)	The GC/MS is calibrated with a minimum 5 level curve for all compounds.
Individual RSD ≤25%; Mean	No exceedences noted.
RSD ≤15%	Comments: None.
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Independent Calibration Check (ICC)	The independent check was run after each initial calibration to verify the calibration. This standard is from a different source than the ICAL.
Individual and Mean PD <25%	No exceedences noted.
	Comments: None.
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Continuing Calibration Verification (CCV)	Continuing calibration standards were run every 24 hours to ensure that initial calibration is still valid.
Individual RSD ≤25%; Mean	No exceedences noted.
RSD ≤15%	Comments: CCVs Q9609, method MQ0378C, had a mean PD above 15%. Individual PDs were acceptable and no further action was taken.