# Capstone Project - Predicting Depression from plasma measurements

Zsombor Szoke-Kovacs

2022-04-22

## #Introduction

## #Project Background

#Methods and Data Analysis Workflow

**#DATA PREPARATION** First we load the data sheet downloaded from the Metabolomics website: https://www.metabolomicsworkbench.org/data/DRCCMetadata.php?Mode=Study&StudyID=ST000062&StudyType=MS&ResultType=1 I have edited the data set; some of the measured molecules have been removed from the list due to simplicity and also due to these did not have names, only ID numbers. A simplified data sheet is used to do the analysis.

```
library("readxl")
library("dplyr")
library("ggplot2")
library('corrplot')
library(purr)
# Here, I set the file to a path on my computer, but once this is saved to a
# different computer, the path will need to be updated. The simplified data sheet
# can also be downloaded from my git repository.
file_original <- "~/Desktop/HarvardX, EdX, Data Science/Capstone Project/Capstone Project - Chosen Proj
temp_file <- read_excel(file_original)
# Converting the temp file into a transposed data frame.
file_df <- as.data.frame(t(temp_file))
# removing unnecessary rows/columns
data <- file_df[c(-2,-3,-4),c(-1,-3,-4,-5,-7)]</pre>
```

To be able to work with the data sheet, new column names are introduced:

```
# Adding new column names for the molecules and the arm:
data[1,1] <- 'Samples'
data[1,2] <- 'Arm'
colnames(data) <-data[1,]
# Remove first row from the data frame:
data <- data[-1,]
# By investigating the data, we can see that measurements come from two groups:
# Group 1 (control) and Group 2 (patients diagnosed with depression):
data %>% group_by(Arm) %>% summarise(n = n())
```

```
## # A tibble: 2 x 2
## Arm n
```

```
## 1 Group 1 - Score 0 48
## 2 Group 2 - Score 50 49
```

To analyse the relationship between the different measurements between the two groups, I first separate the two arms and remove unnecessary columns:

```
# Separate the two arms from the data set:
group_1 <- data %>% group_by(Arm) %>% filter(Arm == "Group 1 - Score 0")
group_1_truncated <- group_1[, c(-1, -2)]
group_2 <- data %>% group_by(Arm) %>% filter(Arm == "Group 2 - Score 50")
group_2_truncated <- group_2[, c(-1, -2)]</pre>
```

#DATA ANALYSIS ##Data distribution To be able to compare the differences between the two groups, I next look at the distribution of the data in each measurement and in each arm. I generated and investigated the distribution of each measured parameter within the two groups using the codes below. However, due to these produce many plots (144 plots per arm), I commented these out in the .rmd file.

```
# Group 1:

#for (i in group_1_truncated){
# plot <- group_1_truncated %>% ggplot(aes(x = as.numeric(i))) +
# geom_density()
# print(plot)
#}

# Group 2

#for (i in group_2_truncated){
# plot <- group_2_truncated %>% ggplot(aes(x = as.numeric(i))) +
# geom_density()
# print(plot)
#}
```

Instead, I used Shapiro-Wilk's method (http://www.sthda.com/english/wiki/normality-test-in-r) to get a value of the normality for each measured parameter. The null hypothesis of this tests is that "the sample distribution is normal". So, if the p-value is >0.05, that implies that the distribution of the data is not significantly different from the normal distribution. In other words, if the p-value is >0.05 we can assume normality. First, I loop through the truncated and transposed list and generate Shapiro-Wilk's test for each column in the data set. I use the magicfor library to record p-values in a vector:

```
library(magicfor)
# Group 1:
magic_for(print)
for (c in group_1_truncated){
    # shap test for each col
    shap_test <- shapiro.test(as.numeric(c))
    output <- shap_test$p.value
    print(output)
}</pre>
```

```
## [1] 0.03075705
## [1] 0.08522262
```

- ## [1] 2.431518e-08
- ## [1] 0.4683182
- ## [1] 0.003183624
- ## [1] 0.4751875
- ## [1] 0.237512
- ## [1] 0.00431784
- ## [1] 0.1507297
- ## [1] 0.03231325
- ## [1] 0.2276085
- ## [1] 0.02835112
- ## [1] 0.9503814
- ## [1] 0.5639228
- ## [1] 0.0001596303
- ## [1] 0.0001859755
- ## [1] 3.838608e-10
- ## [1] 0.5760497
- ## [1] 0.5129488
- ## [1] 0.001635735
- ## [1] 2.859378e-14
- ## [1] 5.950413e-06
- ## [1] 0.7951635
- ## [1] 0.7440938
- ## [1] 0.0001307812
- ## [1] 1.769107e-10
- ## [1] 5.592586e-07
- ## [1] 2.080603e-14
- ## [1] 7.373533e-08
- ## [1] 0.2458592
- ## [1] 0.1436593
- ## [1] 4.844873e-13
- ## [1] 2.119992e-07
- ## [1] 0.0008157663
- ## [1] 0.02371112
- ## [1] 0.006136389
- ## [1] 0.8390248
- ## [1] 6.586207e-12
- ## [1] 0.2145127
- ## [1] 0.001970347
- ## [1] 0.002844868
- ## [1] 0.5173994
- ## [1] 0.04001211
- ## [1] 0.0001117784
- ## [1] 0.1276793
- ## [1] 0.02324038
- ## [1] 0.005191888
- ## [1] 0.008093098
- ## [1] 6.88849e-05
- ## [1] 7.739744e-07
- ## [1] 0.0008360514
- ## [1] 0.8068052
- ## [1] 0.1150337
- ## [1] 0.01927027
- ## [1] 0.5908586
- ## [1] 0.005363307

- ## [1] 0.1046918
- ## [1] 0.08097101
- ## [1] 1.582163e-13
- ## [1] 3.709977e-07
- ## [1] 2.203858e-09
- ## [1] 0.04381657
- ## [1] 0.7613584
- ## [1] 0.8516382
- ## [1] 2.83857e-05
- ## [1] 1.156385e-08
- ## [1] 0.1914479
- ## [1] 0.0895071
- ## [1] 3.574687e-06
- ## [1] 4.420659e-07
- ## [1] 5.170312e-10
- ## [1] 7.584198e-06
- ## [1] 0.05041785
- ## [1] 2.896216e-07
- ## [1] 0.03230108
- ## [1] 0.0001333469
- ## [1] 7.273573e-10
- ## [1] 3.547011e-13
- ## [1] 0.008958414
- ## [1] 0.351065
- ## [1] 0.0004660933
- ## [1] 0.000318582
- ## [1] 4.474704e-08
- ## [1] 6.085384e-09
- ## [1] 0.3867491
- ## [1] 0.06902782
- ## [1] 0.5782418
- ## [1] 8.471097e-07
- ## [1] 0.007310657
- ## [1] 1.555231e-05
- ## [1] 3.457164e-08
- ## [1] 0.183637
- ## [1] 1.163639e-05
- ## [1] 3.332581e-07
- ## [1] 0.0002195566
- ## [1] 0.01156348
- ## [1] 0.08063735
- ## [1] 0.02113055
- ## [1] 1.603749e-08
- ## [1] 2.500726e-14
- ## [1] 6.602879e-05
- ## [1] 0.02840995
- ## [1] 0.5292255
- ## [1] 0.5413767
- ## [1] 0.008912426
- ## [1] 2.88464e-11
- ## [1] 0.02485436
- ## [1] 1.950487e-11
- ## [1] 0.07610434
- ## [1] 0.5720348

```
## [1] 2.000887e-06
## [1] 9.319506e-05
## [1] 0.6336717
## [1] 1.371973e-13
## [1] 0.001600674
## [1] 0.2434581
## [1] 0.007982874
## [1] 1.671043e-08
## [1] 0.008609122
## [1] 2.115987e-12
## [1] 0.0834015
## [1] 1.176368e-05
## [1] 0.004877992
## [1] 6.80456e-06
## [1] 0.003929345
## [1] 0.2785123
## [1] 2.755291e-07
## [1] 0.006870093
## [1] 6.047253e-12
## [1] 2.886682e-10
## [1] 3.685127e-08
## [1] 0.0002518801
## [1] 0.03174344
## [1] 0.1348054
## [1] 9.868894e-09
## [1] 3.383021e-08
## [1] 0.00286715
## [1] 0.01370772
## [1] 0.467026
## [1] 0.00181973
## [1] 2.310402e-07
## [1] 8.595669e-11
## [1] 0.2939691
# Saving printed p-values as a vector:
p_values_vector_group_1 <- magic_result_as_vector()</pre>
# Binding vector to the original data, so the last row is the p-value from the
# Shapiro-Wilk's test:
group_1_truncated_with_pvalues <- rbind(group_1_truncated,p_values_vector_group_1)</pre>
# Group 2:
magic_for(print)
# For loop for collecting all p-values and printing them to the console:
for (c in group_2_truncated){
  # shap test for each col
  shap_test <- shapiro.test(as.numeric(c))</pre>
  output <- shap_test$p.value
  print(output)
}
## [1] 0.01268071
## [1] 0.5320017
## [1] 1.046158e-14
## [1] 0.9271982
```

- ## [1] 0.3338198
- ## [1] 0.002995611
- ## [1] 0.6509885
- ## [1] 0.3337033
- ## [1] 0.2176103
- ## [1] 0.2517892
- ## [1] 5.691081e-05
- ## [1] 0.008071441
- ## [1] 0.9496703
- ## [1] 0.003835506
- ## [1] 1.276141e-06
- ## [1] 4.014598e-06
- ## [1] 7.396027e-15
- ## [1] 1.000021C 10
- ## [1] 0.003291311 ## [1] 2.583564e-05
- ## [1] 2.000004e-00
- ## [1] 1.078828e-09
- ## [1] 6.213368e-12
- ## [1] 9.487599e-10
- ## [1] 0.4116826
- ## [1] 5.813022e-08
- ## [1] 0.004757193
- ## [1] 1.024567e-07
- ## [1] 5.331477e-08
- ## [1] 2.607211e-08
- ... [1] 2.00,2110 00
- ## [1] 2.121411e-14
- ## [1] 0.6218422
- ## [1] 0.3725114
- ## [1] 1.326105e-12
- ## [1] 3.895945e-09
- ## [1] 1.232802e-11
- ## [1] 0.09996526
- ## [1] 0.4281124
- ## [1] 0.214296
- ## [1] 2.509511e-10
- ## [1] 2.26814e-08
- ## [1] 8.840691e-12
- ## [1] 3.020783e-07
- ## [1] 0.0003244317
- ## [1] 0.3325172
- ## [1] 2.297722e-06
- ## [1] 0.1971648
- ## [1] 7.522532e-08
- ## [1] 0.1380847
- ## [1] 0.002045618
- ## [1] 2.969678e-10
- ## [1] 2.122113e-09
- ## [1] 0.0001097306
- ## [1] 0.2254222
- ## [1] 1.308554e-05
- ## [1] 0.00055218
- ## [1] 0.1044934
- ## [1] 0.2794276
- ## [1] 0.03188302
- ## [1] 0.1232555

- ## [1] 1.348212e-11
- ## [1] 6.331095e-14
- ## [1] 4.309545e-12
- ## [1] 0.2554861
- ## [1] 5.045201e-08
- ## [1] 0.01483991
- ## [1] 0.03036964
- ## [1] 4.80947e-12
- ## [1] 0.2054814
- ## [1] 0.0001040936
- ## [1] 3.371332e-09
- ## [1] 0.006638151
- ## [1] 1.565909e-08
- ## [1] 0.002225898
- ## [1] 0.2305581
- ## [1] 2.791335e-11
- ## [1] 0.04784196
- ## [1] 1.563023e-09
- ## [1] 8.730435e-06
- ## [1] 8.412952e-11
- ## [1] 0.6350672
- ## [1] 0.2121618
- ## [1] 1.692299e-05
- ## [1] 1.901465e-05
- ## [1] 2.076581e-09
- ## [1] 1.23874e-07
- ## [1] 2.796222e-07
- ## [1] 0.9728612
- ## [1] 0.6713909
- ## [1] 2.197602e-08
- ## [1] 7.510541e-06
- ## [1] 8.175529e-06
- ## [1] 0.000740605
- ## [1] 0.3345837
- ## [1] 0.0009565749
- ## [1] 6.451756e-12
- ## [1] 3.312299e-05
- ## [1] 0.7167231
- ## [1] 4.774386e-12
- ## [1] 0.003379862
- ## [1] 6.348476e-09
- ## [1] 5.033355e-14
- ## [1] 0.0001545562
- ## [1] 0.0001110415
- ## [1] 0.02478078
- ## [1] 0.02641833
- ## [1] 0.1739291
- ## [1] 2.784913e-11
- ## [1] 1.167685e-05
- ## [1] 3.381197e-10
- ## [1] 0.01268761
- ## [1] 0.06897864
- ## [1] 4.75765e-07
- ## [1] 4.527211e-05

```
## [1] 5.633739e-11
## [1] 4.312549e-15
## [1] 1.72198e-06
## [1] 0.1555038
## [1] 0.003044546
## [1] 0.4012447
## [1] 0.0002663313
## [1] 3.665618e-14
## [1] 6.023239e-05
## [1] 1.736724e-06
## [1] 0.05283555
## [1] 8.825776e-07
## [1] 6.194916e-13
## [1] 0.1922048
## [1] 6.154861e-15
## [1] 0.08027211
## [1] 9.050325e-13
## [1] 2.338873e-06
## [1] 2.232031e-08
## [1] 0.05178879
## [1] 1.921007e-05
## [1] 0.0005122628
## [1] 0.0004112289
## [1] 0.0005876738
## [1] 1.251389e-06
## [1] 2.130454e-07
## [1] 0.0724654
## [1] 0.002222042
## [1] 1.046887e-09
## [1] 1.936056e-10
## [1] 6.314063e-05
# Saving printed p-values as a vector:
p_values_vector_group_2 <- magic_result_as_vector()</pre>
# Binding vector to the original data, so the last row is the p-value from the
# Shapiro-Wilk's test:
group_2_truncated_with_pvalues <- rbind(group_2_truncated,p_values_vector_group_2)</pre>
# Remove magicalization:
magic free()
# The last row in these two data frames are the Shapiro-Wilk's p-values:
group_1_truncated_with_pvalues %>%
  summarise(Arm = 'Group 2',
            nrow = dim(group_1_truncated_with_pvalues)[1],
            ncol = dim(group_1_truncated_with_pvalues)[2])
## # A tibble: 1 x 3
     Arm
              nrow ncol
             <int> <int>
     <chr>>
## 1 Group 2
                49
                     143
group_2_truncated_with_pvalues %>%
  summarise(Arm = 'Group 2',
            nrow = dim(group_2_truncated_with_pvalues)[1],
            ncol = dim(group_2_truncated_with_pvalues)[2])
```

```
## # A tibble: 1 x 3
## Arm nrow ncol
## <chr> <int> <int> <int> 143
```

Now that I have the p-values for the Shapiro-Wilk's test for the measured parameters from each arm, I transposevthe data frames, so the p-values are in a separate column and the data frame in tidy format:

```
## Arm nrow ncol
## 1 Group 1 143 49
```

```
## Arm nrow ncol
## 1 Group 2 143 50
```

Now that I have the data for the two arms, together with the p-values for normal distribution, I filter the data to keep the measured parameters, where the distribution was approximately normal. In other words, I keep all measured parameters, where the p-value was >0.05:

```
group_1_tidy <- group_1_tidy %>% filter(`Shapiro-Wilk's p-values`>0.05)
group_2_tidy <- group_2_tidy %>% filter(`Shapiro-Wilk's p-values`>0.05)
# There are 97 and 112 measured parameters where the p-value is >0.05 in Group 1
# and Group 2, respectively. Group 1 has 48 patients, whereas Group 2 has 49. The extra
# column in each data frame is the Shapiro-Wilk's p-value.
group_1_tidy %>% summarise(Arm = 'Group 1', nrow = dim(group_1_tidy)[1],
                            ncol = dim(group_1_tidy)[2])
##
         Arm nrow ncol
## 1 Group 1
               97
group_2_tidy %>% summarise(Arm = 'Group 2', nrow = dim(group_2_tidy)[1],
                            ncol = dim(group_2_tidy)[2])
         Arm nrow ncol
## 1 Group 2 112
Due to the number of the normally distributed measured parameters are different in the two groups, I will
work with the list from the control group (Group 1 - baseline), where the normally distributed parameters
were 97 (as opposed to Group 2 where it was 112). I use semi join to keep only the records from Group 2,
that have a match in Group 1.
# Adding row names as an extra column, so I can use semi_join:
group_1_tidy <- cbind(group_1_tidy, rownames = rownames(group_1_tidy))</pre>
group_2_tidy <- cbind(group_2_tidy, rownames = rownames(group_2_tidy))</pre>
# we should have one extra column in each data frame:
group_1_tidy %>% summarise(Arm = 'Group 1', nrow = dim(group_1_tidy)[1],
                            ncol = dim(group 1 tidy)[2])
##
         Arm nrow ncol
## 1 Group 1
               97
group_2_tidy %>% summarise(Arm = 'Group 2', nrow = dim(group_2_tidy)[1],
                            ncol = dim(group_2_tidy)[2])
##
         Arm nrow ncol
## 1 Group 2 112
# Keep everything from Group 1 with a match in Group 2:
group_1_tidy <- semi_join(group_1_tidy, group_2_tidy, by = "rownames")</pre>
# Keep everything from Group 2 with a match in Group 1:
group_2_tidy <- semi_join(group_2_tidy, group_1_tidy, by = "rownames")</pre>
# Investigating the dimensions of the two newly generated data frames, we can see, that
# both arms have 78 measured parameters, as well as 48 and 49 sample count (plus the two columns
# with p-values and row names), respectively.
group_1_tidy %>% summarise(Arm = 'Group 1', nrow = dim(group_1_tidy)[1],
                            ncol = dim(group_1_tidy)[2])
         Arm nrow ncol
## 1 Group 1
               78
```

```
group_2_tidy %>% summarise(Arm = 'Group 2', nrow = dim(group_2_tidy)[1],
                           ncol = dim(group_2_tidy)[2])
```

```
Arm nrow ncol
## 1 Group 2
               78
                     51
```

##Two-sample t-test In this next section, I will calculate two-sample t-tests for the selected parameters, so I can see if there is a significant difference in any parameters between the two groups. First, I transpose the data frame generated above and remove unnecessary rows.

```
# Transpose tidy data, so I can loop through the columns: Group 1
group_1_tidy_t <- as.data.frame(t(group_1_tidy))</pre>
# Removing last two rows with p-values and row names:
group_1_tidy_t \leftarrow group_1_tidy_t[c(-49,-50),]
# Transpose tidy data, so I can loop through the columns: Group 2
group_2_tidy_t <- as.data.frame(t(group_2_tidy))</pre>
# Removing last two rows with p-values and row names:
group_2_tidy_t \leftarrow group_2_tidy_t[c(-50,-51),]
# The two data set has 78 measured parameters and 48 and 49 samples, respectively:
group_1_tidy_t %>% summarise(Arm = 'Group 1', nrow = dim(group_1_tidy_t)[1],
                              ncol = dim(group_1_tidy_t)[2])
##
         Arm nrow ncol
## 1 Group 1
               48
group_2_tidy_t %>% summarise(Arm = 'Group 2', nrow = dim(group_2_tidy_t)[1],
                              ncol = dim(group_2_tidy_t)[2])
##
         Arm nrow ncol
## 1 Group 2
```

Now, that I have the two data frames with the same measured parameters in both, and all of the measurements show approximately normal distribution, I can test the vectors for significant differences:

```
# Two-sample t-test by looping through the columns:
magic_for(print)
for (j in seq(ncol(group_1_tidy_t))){
  testresults <- t.test(as.numeric(group_1_tidy_t[,j]), as.numeric(group_2_tidy_t[,j]))</pre>
  print(testresults$p.value)
}
```

```
## [1] 0.901493
## [1] 0.1944979
## [1] 0.5386687
## [1] 0.3329934
## [1] 0.9450576
## [1] 0.9846481
## [1] 0.5639318
## [1] 0.1921548
## [1] 4.942357e-05
## [1] 0.04571605
```

49

- ## [1] 0.01102079
- ## [1] 0.781581
- ## [1] 0.1682779
- ## [1] 0.0419918
- ## [1] 0.3691553
- ## [1] 0.3899988
- ## [1] 0.270009
- ## [1] 0.001947676
- ## [1] 0.09498654
- ## [1] 0.1854972
- ## [1] 0.3880945
- ## [1] 0.4855856
- ... [1] 0.100000
- ## [1] 0.4947039
- ## [1] 0.1916466
- ## [1] 0.1236281 ## [1] 0.0005894853
- ## [1] 0.08959645
- ## [1] 0.2790141
- ## [1] 0.3967777
- ## [1] 0.00106033
- ## [1] 0.02861927
- ## [1] 0.1148812
- ## [1] 0.0752034
- ## [1] 0.275934
- ## [1] 0.002000258
- ## [1] 0.5798523
- ## [1] 0.8924322
- ## [1] 0.1573088
- ## [1] 0.05893773
- ## [1] 0.3321228
- ## [1] 0.1155188
- ## [1] 0.6678818
- ## [1] 0.1238112
- ## [1] 0.38568
- ## [1] 0.5241215
- ## [1] 0.06437688
- ## [1] 0.0001549329
- ## [1] 0.2873975
- ## [1] 0.6738781
- ## [1] 0.4753776
- ## [1] 0.09426011
- ## [1] 0.04067713
- ## [1] 0.01122825
- ## [1] 0.1402982
- ## [1] 0.5078799
- ## [1] 0.535678
- ## [1] 0.5192594
- ## [1] 0.3900095
- ## [1] 0.02823496
- ## [1] 0.5166175
- ## [1] 0.5375167
- ## [1] 0.08410382
- ## [1] 0.3515712
- ## [1] 0.9093331

```
## [1] 0.9311662
## [1] 0.2831985
## [1] 0.086737
## [1] 0.1851199
## [1] 0.9373875
## [1] 0.2306857
## [1] 0.3310121
## [1] 0.9455772
## [1] 0.2934011
# Saving p-values from the two-sample t-test into a dataframe, and adding the
twosample_ttest <- magic_result_as_dataframe()</pre>
magic_free()
# Adding the names of the measured parameters to the p-values:
colnames(twosample_ttest)[1] <- 'rownames'</pre>
twosample_ttest$`rownames` <- colnames(group_1_tidy_t)</pre>
# Filtering out measured parameters that showed significant differences between
# the two groups:
twosample_ttest_significant <- twosample_ttest %>% filter(`testresults$p.value` <= 0.05)
# There are 15 measured parameters that show normal distribution, and there is a significant difference
# between the two groups:
twosample_ttest_significant
```

```
##
                 rownames testresults$p.value
## 1
                                  4.942357e-05
             stearic acid
## 2
                 sorbitol
                                  4.571605e-02
## 3
            shikimic acid
                                  1.102079e-02
## 4
                  ribitol
                                  4.199180e-02
## 5
           pseudo uridine
                                  1.947676e-03
                                  5.894853e-04
## 6
             nicotinamide
## 7
             myo-inositol
                                  1.060330e-03
## 8
                                  2.861927e-02
                  mannose
## 9
                  lyxitol
                                  2.000258e-03
## 10
      heptadecanoic acid
                                  1.549329e-04
## 11
            glutaric acid
                                  4.067713e-02
## 12
            glutamic acid
                                  1.122825e-02
## 13
                                  2.823496e-02
              citric acid
## 14
             behenic acid
                                  9.529080e-04
## 15 alpha-ketoglutarate
                                  1.629588e-03
```

## [1] 0.000952908 ## [1] 0.1326068 ## [1] 0.1060869 ## [1] 0.5360112 ## [1] 0.001629588

##Correlation analysis From the previous section, I have a set of measured parameters that show significant difference of the mean between the two arms. To see the actual relationship between the two groups, I will use correlation analysis for the 15 parameters. Initially, I will subset the two dataframes group\_1\_tidy and group 2 tidy, to only consist of the 15 parameters of interest.

```
group_1_final <- semi_join(group_1_tidy, twosample_ttest_significant, by = 'rownames')
group_1_final <- group_1_final[,c(-49,-50)]</pre>
```

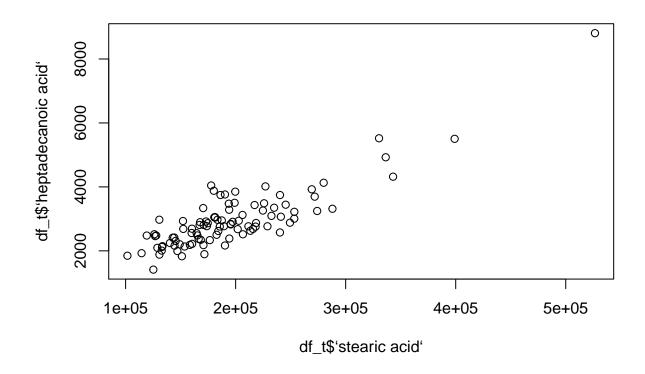
```
group_2_final <- semi_join(group_2_tidy, twosample_ttest_significant, by = 'rownames')</pre>
group_2_final \leftarrow group_2_final[,c(-50,-51)]
# Here I have two dataframes from the two arms, one control and one diagnosed with depression,
# where the parameters of interest are included only. The dataframes consist of 48 and 49
# patients, respectively:
group_1_final %>% summarise(Arm = 'Group 1', nrow = dim(group_1_final)[1],
                             ncol = dim(group 1 final)[2])
##
         Arm nrow ncol
## 1 Group 1
               15
group_2_final %>% summarise(Arm = 'Group 2', nrow = dim(group_2_final)[1],
                             ncol = dim(group_2_final)[2])
##
         Arm nrow ncol
## 1 Group 2
               15
```

Now that I have the two dataframes with the 15 measured parameters that showed approximately normal distribution and significant differences between the two groups, I will merge the two arms, and will generate a new dataframe with all of the subjects and the 15 measured parameters. I will use this dataframe for my further work:

```
# first, I create a new column in both dataframes, so I can merge these with
# left_join()
group_1_final <- cbind(group_1_final, rownames = rownames(group_1_final))
group_2_final <- cbind(group_2_final, rownames = rownames(group_2_final))
# Merging the two dataframes by rownames:
df <- left_join(group_1_final, group_2_final, by = "rownames")
# Adding rownames based on the rownames column
rownames(df) <- df$rownames
# Removing rownames column:
df <- subset(df, select = -rownames)</pre>
```

Testing for correlation:

```
# This is my dataset with all 15 parameters and the entire cohort.
# I now transpose it and will do a correlation analysis to see if any of these
# parameters are correlated:
df_t <- as.data.frame(t(df))
# A quick plotting of the data shows that there is a potential correlation between
# stearic acid and heptadecanoic acid: commented out so, otherwise many lots will be printed.
# plot(df_t)
plot(df_t$`stearic acid`, df_t$`heptadecanoic acid`)</pre>
```



```
# A correlation analysis between the two parameters shows a strong positive correlation
# with a value of 0.862:
cor.test(as.numeric(df_t$`stearic acid`), as.numeric(df_t$`heptadecanoic acid`))
##
##
   Pearson's product-moment correlation
## data: as.numeric(df_t$'stearic acid') and as.numeric(df_t$'heptadecanoic acid')
## t = 16.583, df = 95, p-value < 2.2e-16
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
   0.8002618 0.9058056
## sample estimates:
##
         cor
## 0.8621075
# Here, I convert all df_t to numeric, so I can do a correlation analysis:
df_num <-as.data.frame(sapply(df_t, as.numeric))</pre>
# This also shows, that the only correlation is between stearic acid and heptadecanoic acid:
cor_15_param <- as.data.frame(cor(df_num))</pre>
cor_15_param %>% filter(cor_15_param >= 0.7)
##
                      stearic acid sorbitol shikimic acid
## stearic acid
                         1.0000000 0.2083188
                                                  0.2492042 0.15060595
## heptadecanoic acid
                         0.8621075 0.1501909
                                                  0.3128487 0.09976116
```

```
##
                      pseudo uridine nicotinamide myo-inositol
## stearic acid
                         -0.19706050
                                        0.3878188
                                                    0.04917488 0.1807085
## heptadecanoic acid
                         -0.05231672
                                        0.2501691
                                                    0.13140503 0.2167668
##
                         lyxitol heptadecanoic acid glutaric acid glutamic acid
## stearic acid
                      0.02011295
                                          0.8621075 -0.034593242
                                                                     0.05591120
                                          1.0000000 -0.005699254
## heptadecanoic acid 0.01543889
                                                                    -0.04747537
                      citric acid behenic acid alpha-ketoglutarate
## stearic acid
                       0.08475043
                                     0.4184708
                                                        0.02048958
## heptadecanoic acid 0.17991391
                                     0.3845567
                                                       -0.01077332
```

##Linearity between the two arms The below code looks at whether the data is linearly separable between the two arms. Values from each measured parameters are plotted on y and the arm is plotted on x.

```
# for (v in data[,c(-1,-2)]){
# plot(as.numeric(v), col = as.factor(data$Arm))
# }
```

Based on the plots generated by the above code, the values do not show a linear association between the two arms, therefore, when applying SVM, although the model performs well on the training data, on the test data, there is a significant drop in the model performance. See this in the subsequent sections.

##Creating training and testing datasets

```
# Creating a training and test set from the above dataframe df:
# install.packages("caTools")
# install.packages("e1071")
library(caTools)
library(e1071)
index <- sample.split(data$Arm, SplitRatio = .7)
tr_set <- subset(data, index == TRUE)
final_val_set <- subset(data, index == FALSE)
# Splitting the training set into further training and test sets:
index_train <- sample.split(tr_set$Arm, SplitRatio = .5)
tr_set_train <- subset(tr_set, index_train == TRUE)
tr_set_test <- subset(tr_set, index_train == FALSE)</pre>
```

#MODEL FITTING ##Support Vector Machine Now, that I have the train\_set, I will train the SVM on the training set. Here I train the SVM to predict the Arm, based on the most significant variables from the df dataset; these are stearic acid, nicotinamide, heptadecanoic acid and behenic acid.

## [1] 0.7647059

```
# Getting the mean of the correctly predicted arm on the test data set:
predict_test_linear <- predict(svm_model_linear, tr_set_test)</pre>
mean linear test <- mean(predict test linear == tr set test$Arm)</pre>
mean linear test
## [1] 0.7647059
# SVM model with radial kernel:
svm_model_radial <- svm(as.factor(Arm)~</pre>
                           as.numeric(tr_set_train$`stearic acid`) +
                           as.numeric(tr_set_train$nicotinamide) +
                           as.numeric(tr_set_train$`heptadecanoic acid`) +
                           as.numeric(tr_set_train$`behenic acid`),
                 data = tr_set_train, method = "C-classification", kernel = "radial", gamma = 1,
                 cost = 1)
# Getting the mean of the correctly predicted arm on the train data set:
predict_tr_radial <- predict(svm_model_radial, tr_set_train)</pre>
mean_radial_train <- mean(predict_tr_radial == tr_set_train$Arm)</pre>
mean_radial_train
## [1] 0.8823529
# Getting the mean of the correctly predicted arm on the test data set:
predict_test_radial <- predict(svm_model_radial, tr_set_test)</pre>
mean_radial_test <- mean(predict_test_radial == tr_set_test$Arm)</pre>
mean_radial_test
## [1] 0.8823529
#VALIDATION ##SVM Validating the SVM model (linear and radial)
predict_valid_linear <- predict(svm_model_linear, final_val_set)</pre>
mean_linear_valid <- mean(predict_valid_linear == final_val_set$Arm)</pre>
## Warning in '==.default'(predict_valid_linear, final_val_set$Arm): longer object
## length is not a multiple of shorter object length
## Warning in is.na(e1) | is.na(e2): longer object length is not a multiple of
## shorter object length
mean linear valid
## [1] 0.7647059
# Radial
predict_valid_radial <- predict(svm_model_radial, final_val_set)</pre>
mean_radial_valid <- mean(predict_valid_radial == final_val_set$Arm)</pre>
```

```
## Warning in '==.default'(predict_valid_radial, final_val_set$Arm): longer object
## length is not a multiple of shorter object length

## Warning in '==.default'(predict_valid_radial, final_val_set$Arm): longer object
## length is not a multiple of shorter object length

mean_radial_valid
```

## [1] 0.7647059

## #RESULTS AND CONCLUSION

## **#FUTURE PERSPECTIVES**