

```

library(DESeq2);load("CancerAndParacancer.RData")

## Loading required package: S4Vectors

## Loading required package: stats4

## Loading required package: BiocGenerics

## 
## Attaching package: 'BiocGenerics'

## The following objects are masked from 'package:stats':
##
##     IQR, mad, sd, var, xtabs

## The following objects are masked from 'package:base':
##
##     anyDuplicated, aperm, append, as.data.frame, basename, cbind,
##     colnames, dirname, do.call, duplicated, eval, evalq, Filter, Find,
##     get, grep, grepl, intersect, is.unsorted, lapply, Map, mapply,
##     match, mget, order, paste, pmax, pmax.int, pmin, pmin.int,
##     Position, rank, rbind, Reduce, rownames, sapply, saveRDS, setdiff,
##     table, tapply, union, unique, unsplit, which.max, which.min

## 
## Attaching package: 'S4Vectors'

## The following object is masked from 'package:utils':
##
##     findMatches

## The following objects are masked from 'package:base':
##
##     expand.grid, I, unname

## Loading required package: IRanges

## Loading required package: GenomicRanges

## Loading required package: GenomeInfoDb

## Loading required package: SummarizedExperiment

## Loading required package: MatrixGenerics

## Loading required package: matrixStats

## 
## Attaching package: 'MatrixGenerics'

## The following objects are masked from 'package:matrixStats':
##
##     colAlls, colAnyNAs, colAnys, colAvgsPerRowSet, colCollapse,
##     colCounts, colCummaxs, colCummins, colCumprods, colCumsums,
##     colDiffs, colIQRDiffs, colIQRs, colLogSumExps, colMadDiffs,
##     colMads, colMaxs, colMeans2, colMedians, colMins, colOrderStats,
##     colProds, colQuantiles, colRanges, colRanks, colSdDiffs, colSds,
##     colSums2, colTabulates, colVarDiffs, colVars, colWeightedMads,
##     colWeightedMeans, colWeightedMedians, colWeightedSds,
##     colWeightedVars, rowAlls, rowAnyNAs, rowAnys, rowAvgsPerColSet,
##     rowCollapse, rowCounts, rowCummaxs, rowCummins, rowCumprods,
##     rowCumsums, rowDiffs, rowIQRDiffs, rowIQRs, rowLogSumExps,
##     rowMadDiffs, rowMads, rowMaxs, rowMeans2, rowMedians, rowMins,
##     rowOrderStats, rowProds, rowQuantiles, rowRanges, rowRanks,
##     rowSdDiffs, rowSds, rowSums2, rowTabulates, rowVarDiffs, rowVars,
##     rowWeightedMads, rowWeightedMeans, rowWeightedMedians,
##     rowWeightedSds, rowWeightedVars

## Loading required package: Biobase

```

```

## Welcome to Bioconductor
##
##   Vignettes contain introductory material; view with
##   'browseVignettes()'. To cite Bioconductor, see
##   'citation("Biobase")', and for packages 'citation("pkgname")'.

##
## Attaching package: 'Biobase'

## The following object is masked from 'package:MatrixGenerics':
##
##     rowMedians

## The following objects are masked from 'package:matrixStats':
##
##     anyMissing, rowMedians

expr <- ExprMat; pdat <- pDataMat; feat <- FeatureMat
same_samples <- intersect(colnames(expr), rownames(pdat))
expr <- expr[, same_samples, drop = F]
pdat <- pdat[same_samples, , drop = F]
expr <- expr[rowSums(expr) >= 954, , drop = F]
expr <- expr[!duplicated(rownames(expr)), , drop = F]
feat <- feat[!duplicated(rownames(feat)), , drop = F]
new_col <- intersect(c("gene_symbol", "symbol", "chromosome", "description", "gene_biotype"), colnames(feat))
feat <- feat[, new_col, drop = F]
feat <- feat[rownames(expr), , drop = F]
colnames(pdat)[1] <- "condition"
pdat$condition <- factor(pdat$condition)
pdat$condition <- relevel(pdat$condition, ref = "paracancerous")
dds <- DESeqDataSetFromMatrix(countData = expr, colData = pdat, design = ~ condition)
dds <- DESeq(dds); res <- results(dds)

## estimating size factors

## estimating dispersions

## gene-wise dispersion estimates

## mean-dispersion relationship

## final dispersion estimates

## fitting model and testing

## -- replacing outliers and refitting for 1371 genes
## -- DESeq argument 'minReplicatesForReplace' = 7
## -- original counts are preserved in counts(dds)

## estimating dispersions

## fitting model and testing

res_ordered <- res[order(res$padj, na.last = NA), ]
des_order <- as.data.frame(res_ordered)
normal_des <- as.data.frame(counts(dds, normalized = T))
des_order$gene <- rownames(des_order)
top_genes <- rownames(des_order)[1:15]
top_genes <- top_genes[top_genes %in% rownames(FeatureMat)]
cutoff <- min(rowSums(expr)[top_genes]); cat("cutoff is:", cutoff, "\n")

## cutoff is: 9296

top <- cbind(GeneID = top_genes, FeatureMat[top_genes, , drop = F],
  des_order[top_genes, c("log2FoldChange", "pvalue", "padj", "stat", "lfcSE", "baseMean"), drop = F])
up_order <- order(des_order$log2FoldChange, decreasing = T)
up_genes <- rownames(des_order)[up_order][1:10]
up_genes <- up_genes[up_genes %in% rownames(FeatureMat)]
cutoff <- min(rowSums(expr)[up_genes]); cat("cutoff is:", cutoff, "\n")

## cutoff is: 1043

```

```
top_up <- cbind(GeneID = up_genes, FeatureMat[up_genes, , drop = F],
  des_order[up_genes, c("log2FoldChange", "pvalue", "padj", "stat", "lfcSE", "baseMean"), drop = F])
down_order <- order(des_order$log2FoldChange, decreasing = F)
down_genes <- rownames(des_order)[down_order][1:10]
down_genes <- down_genes[down_genes %in% rownames(FeatureMat)]
cutoff <- min(rowSums(expr)[down_genes]);cat("cutoff is:", cutoff, "\n")
```

```
## cutoff is: 954
```

```
top_down <- cbind(GeneID = down_genes, FeatureMat[down_genes, , drop = F],
  des_order[down_genes, c("log2FoldChange", "pvalue", "padj", "stat", "lfcSE", "baseMean"), drop = F])
summary(top);knitr::kable(top)
```

```
##      GeneID          GeneID        Symbol       Description
##  Length:15      Min.   : 1589  Length:15      Length:15
##  Class :character 1st Qu.: 4898  Class :character  Class :character
##  Mode  :character Median : 9340  Mode  :character  Mode  :character
##                           Mean   : 79939
##                           3rd Qu.:182952
##                           Max.  :286133
##      Synonyms          GeneType      EnsemblGeneID      Status
##  Length:15      Length:15      Length:15      Length:15
##  Class :character Class :character  Class :character  Class :character
##  Mode  :character  Mode  :character  Mode  :character  Mode  :character
##
##      ChrAcc          ChrStart      ChrStop       Orientation
##  Length:15      Length:15      Length:15      Length:15
##  Class :character Class :character  Class :character  Class :character
##  Mode  :character  Mode  :character  Mode  :character  Mode  :character
##
##      Length     GOFunctionID      GOProcessID      GOComponentID
##  Min.   : 947  Length:15      Length:15      Length:15
##  1st Qu.: 2186  Class :character  Class :character  Class :character
##  Median : 2429  Mode  :character  Mode  :character  Mode  :character
##  Mean   : 4340
##  3rd Qu.: 5370
##  Max.   :13899
##      GOFunction      GOProcess      GOComponent      log2FoldChange
##  Length:15      Length:15      Length:15      Min.   :-7.681
##  Class :character Class :character  Class :character  1st Qu.:-4.934
##  Mode  :character  Mode  :character  Mode  :character  Median :-4.777
##                           Mean   :-4.722
##                           3rd Qu.:-4.088
##                           Max.   :-3.357
##      pvalue          padj          stat          lfcSE
##  Min.   :0.000e+00  Min.   :0.000e+00  Min.   :-19.64  Min.   :0.2385
##  1st Qu.:0.000e+00  1st Qu.:0.000e+00  1st Qu.:-15.38  1st Qu.:0.2853
##  Median :2.520e-45  Median :6.770e-42  Median :-14.13  Median :0.3125
##  Mean   :4.419e-42  Mean   :6.613e-39  Mean   :-14.87  Mean   :0.3156
##  3rd Qu.:6.347e-43  3rd Qu.:1.164e-39  3rd Qu.:-13.75  3rd Qu.:0.3529
##  Max.   :3.319e-41  Max.   :4.766e-38  Max.   :-13.44  Max.   :0.3911
##      baseMean
##  Min.   :136.3
##  1st Qu.:217.3
##  Median :435.4
##  Mean   :1081.2
##  3rd Qu.:1428.7
##  Max.   :4452.2
```

GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID	Status	ChrAcc
221476	221476	221476	PI16	peptidase inhibitor 16	CD364 CRISP9 MSMBBP PSPBP	protein-coding	ENSG00000164530	active NC_000
1675	1675	1675	CFD	complement factor D	ADIPSIN ADN DF PFD	protein-coding	ENSG00000197766	active NC_000
286133	286133	286133	SCARA5	scavenger receptor class A member 5	NET33 Tesr	protein-coding	ENSG00000168079	active NC_000

GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID	Status	ChrAcc
7123	7123	7123 CLEC3B	C-type lectin domain family 3 member B	MCDR4 TN TNA	protein-coding	ENSG00000163815	active	NC_000
9340	9340	9340 GLP2R	glucagon like peptide 2 receptor		protein-coding	ENSG0000065325	active	NC_000
11170	11170	11170 FAM107A	family with sequence similarity 107 member A	DRR1 TU3A	protein-coding	ENSG00000168309	active	NC_000
219348	219348	219348 PLAC9	placenta associated 9		protein-coding	ENSG00000189129	active	NC_000
146556	146556	146556 C16orf89	chromosome 16 open reading frame 89		protein-coding	ENSG00000153446	active	NC_000
221091	221091	221091 LRRN4CL	LRRN4 C-terminal like		protein-coding	ENSG00000177363	active	NC_000
1589	1589	1589 CYP21A2	cytochrome P450 family 21 subfamily A member 2	CA21H CAH1 CPS1 CYP21 CYP21B P450c21B	protein-coding	ENSG00000231852	active	NC_000
2674	2674	2674 GFRA1	GDNF family receptor alpha 1	GDNFR GDNFR-alpha-1 GDNFRA GFR-ALPHA-1 GFRalpha-1 RET1L RETL1 RHD4 TRNR1	protein-coding	ENSG00000151892	active	NC_000
1590	1590	1590 CYP21A1P	cytochrome P450 family 21 subfamily A member 1, pseudogene	CYP21A CYP21P P450c21A	pseudo	ENSG00000290788	active	NC_000
7146	7146	7146 TNXA	tenascin XA (pseudogene)	D6S103E HXBL TNX XA	pseudo		active	NC_000

GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID	Status	ChrAcc
55022	55022	55022 PID1	phosphotyrosine interaction domain containing 1	HMFN2073 NYGGF4 P-CLI1 PCLI1	protein-coding	ENSG00000153823	active	NC_000
7148	7148	7148 TNXB	tenascin XB	EDS3 EDSCLL EDSCLL1 HXBL TENX TN-X TNX TNXB1 TNXB2 TNXBS VUR8 XB XBS	protein-coding	ENSG00000168477	active	NC_000

```
summary(top_up);knitr::kable(top_up)
```

```

##   GeneID          GeneID          Symbol          Description
##   Length:10      Min. : 1472  Length:10      Length:10
##   Class :character 1st Qu.: 19677  Class :character  Class :character
##   Mode :character Median : 109906  Mode :character  Mode :character
##   Mean : 41924921
##   3rd Qu.:101929231
##   Max. :109729169
##   Synonyms        GeneType        EnsemblGeneID    Status
##   Length:10      Length:10      Length:10      Length:10
##   Class :character Class :character  Class :character  Class :character
##   Mode :character Mode :character  Mode :character  Mode :character
##
##
##
##   ChrAcc          ChrStart        ChrStop         Orientation
##   Length:10      Length:10      Length:10      Length:10
##   Class :character Class :character  Class :character  Class :character
##   Mode :character Mode :character  Mode :character  Mode :character
##
##
##
##   Length     GOFunctionID      GOProcessID      GOComponentID
##   Min. : 679  Length:10      Length:10      Length:10
##   1st Qu.:1236  Class :character  Class :character  Class :character
##   Median :1519  Mode :character  Mode :character  Mode :character
##   Mean : 1804
##   3rd Qu.:2160
##   Max. : 4158
##   GOFunction      GOProcess       GOComponent      log2FoldChange
##   Length:10      Length:10      Length:10      Min. :5.448
##   Class :character Class :character  Class :character  1st Qu.:5.788
##   Mode :character Mode :character  Mode :character  Median :5.931
##   Mean : 6.141
##   3rd Qu.:6.124
##   Max. : 7.555
##   pvalue          padj           stat           lfcSE
##   Min. : 0.000e+00  Min. :0.000e+00  Min. : 3.806  Min. : 0.6162
##   1st Qu.: 0.000e+00 1st Qu.:0.000e+00  1st Qu.:5.354  1st Qu.: 0.7731
##   Median :9.300e-10  Median :1.230e-08  Median : 6.512  Median : 0.9580
##   Mean : 1.763e-05  Mean : 6.626e-05  Mean : 6.558  Mean : 1.0348
##   3rd Qu.:1.525e-07 3rd Qu.:1.219e-06  3rd Qu.: 7.230  3rd Qu.: 1.1248
##   Max. : 1.410e-04  Max. : 5.105e-04  Max. : 9.996  Max. : 1.9849
##   baseMean
##   Min. : 10.84
##   1st Qu.: 21.86
##   Median : 33.03
##   Mean : 41.44
##   3rd Qu.: 45.57
##   Max. :132.55

```

	GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID	Status	ChrAcc
51350	51350	51350	KRT76	keratin 76	HUMCYT2A KRT2B KRT2P	protein-coding	ENSG00000185069	active	NC_00001:
9119	9119	9119	KRT75	keratin 75	CK-75 K6HF K75 KB18 PFB hK6hf	protein-coding	ENSG00000170454	active	NC_00001:
1472	1472	1472	CST4	cystatin S		protein-coding	ENSG00000101441	active	NC_000021:
101929412	101929412	101929412	LINC02212	long intergenic non-protein coding RNA 2212		ncRNA	ENSG00000249396	active	NC_00000:
101928687	101928687	101928687	LNCAROD	lncRNA activating regulator of DKK1	A-ROD LINC01468 lnc-MBL2-4	ncRNA	ENSG00000231131	active	NC_000010:

GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID	Status	ChrAcc
6704	6704	6704 SPRR2E	small proline rich protein 2E		protein-coding	ENSG00000203785	active	NC_00000:
56033	56033	56033 BARX1	BARX homeobox 1		protein-coding	ENSG00000131668	active	NC_00000:
163778	163778	163778 SPRR4	small proline rich protein 4		protein-coding	ENSG00000184148	active	NC_00000:
105373485	105373485	105373485 LOC105373485	uncharacterized LOC105373485		ncRNA		active	NC_00000:
109729169	109729169	109729169 LINC02154	long intergenic non-protein coding RNA 2154		ncRNA	ENSG00000235385	active	NC_00002:

```
summary(top_down);knitr::kable(top_down)
```

```

##   GeneID          GeneID          Symbol          Description
##   Length:10      Min. : 70    Length:10      Length:10
##   Class :character 1st Qu.: 4770  Class :character  Class :character
##   Mode :character Median : 7276  Mode :character  Mode :character
##               Mean : 10283197
##               3rd Qu.: 187198
##               Max. : 101927499
##   Synonyms        GeneType        EnsemblGeneID     Status
##   Length:10      Length:10      Length:10      Length:10
##   Class :character Class :character Class :character  Class :character
##   Mode :character Mode :character Mode :character  Mode :character
##
##
##
##   ChrAcc          ChrStart        ChrStop        Orientation
##   Length:10      Length:10      Length:10      Length:10
##   Class :character Class :character Class :character  Class :character
##   Mode :character Mode :character Mode :character  Mode :character
##
##
##
##   Length       GOFunctionID      GOProcessID      GOComponentID
##   Min. : 382.0  Length:10      Length:10      Length:10
##   1st Qu.: 767.5 Class :character Class :character  Class :character
##   Median :1741.0 Mode :character Mode :character  Mode :character
##   Mean   :2201.6
##   3rd Qu.:2241.2
##   Max.   :6769.0
##   GOFunction      GOProcess      GOComponent      log2FoldChange
##   Length:10      Length:10      Length:10      Min.   :-7.681
##   Class :character Class :character Class :character  1st Qu.:-6.584
##   Mode :character Mode :character Mode :character  Median :-6.087
##               Mean   :-6.317
##               3rd Qu.:-5.924
##               Max.   :-5.751
##   pvalue          padj           stat          lfcSE
##   Min.   :0.000e+00  Min.   :0.000e+00  Min.   :-19.638  Min.   :0.3872
##   1st Qu.:0.000e+00  1st Qu.:0.000e+00  1st Qu.:-12.127  1st Qu.:0.5233
##   Median :0.000e+00  Median :0.000e+00  Median :-11.327  Median :0.5599
##   Mean   :5.015e-17  Mean   :2.762e-15   Mean   :-11.969  Mean   :0.5514
##   3rd Qu.:1.000e-22  3rd Qu.:1.900e-20  3rd Qu.:-9.968  3rd Qu.:0.6075
##   Max.   :5.015e-16  Max.   :2.762e-14   Max.   :-8.111   Max.   :0.7293
##   baseMean
##   Min.   : 19.44
##   1st Qu.: 145.78
##   Median : 242.39
##   Mean   : 4194.07
##   3rd Qu.: 1797.90
##   Max.   :34495.23

```

	GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID
221476	221476	221476	PI16	peptidase inhibitor 16	CD364 CRISP9 MSMBBP PSPBP	protein-coding	ENSG00000164530

GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID
4653	4653	4653 MYOC	myocilin	GLC1A GPOA JOAG JOAG1 TIGR	protein-coding	ENSG00000034971
572558	572558	572558 PGM5-AS1	PGM5 antisense RNA 1	FAM233A	ncRNA	ENSG00000224958
5212	5212	5212 VIT	vitrin	VIT1	protein-coding	ENSG00000205221
84366	84366	84366 PRAC1	PRAC1 small nuclear protein	C17orf92 PRAC	protein-coding	ENSG00000159182
70	70	70 ACTC1	actin alpha cardiac muscle 1	ACTC ASD5 CMD1R CMH11 LVNC4	protein-coding	ENSG00000159251

GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID
9340	9340	9340 GLP2R	glucagon like peptide 2 receptor		protein-coding	ENSG00000065325
101927499 101927499 101927499 LOC101927499 uncharacterized LOC101927499						ncRNA
1674	1674	1674 DES	desmin	CDCD3 CSM1 CSM2 LGMD1D LGMD1E LGMD2R	protein-coding	ENSG00000175084
5121 5121 5121 PCP4 Purkinje cell protein 4 PEP-19						protein-coding
ENSG00000183036						

## STEP 4- showing the top 15 genes

```
# find significant DEGs
sum( res$padj < 0.1, na.rm=TRUE )

## [1] 13250

# clean and delete na
res_df <- as.data.frame(res)
res_df <- res_df[!is.na(res_df$padj), ]

#order by significance
res_ordered <- res_df[order(res_df$padj), ]

# extract top15
top15 <- head(res_ordered, 15)
top15

##      baseMean log2FoldChange      lfcSE       stat      pvalue      padj
## 221476 1945.5333 -7.681227 0.3911341 -19.63835 7.272694e-86 1.566175e-81
## 1675  4452.1590 -4.964090 0.2787639 -17.80751 6.180072e-71 6.654392e-67
## 286133 1803.0834 -5.509867 0.3535451 -15.58462 9.260587e-55 6.647558e-51
## 7123  1054.3341 -4.777304 0.3102290 -15.39928 1.654936e-53 8.909761e-50
## 9340  161.6683 -5.950893 0.3872252 -15.36804 2.681586e-53 1.154959e-49
## 11170  844.1215 -4.797705 0.3233931 -14.83552 8.632901e-50 3.098492e-46
## 219348 435.3765 -3.394889 0.2385134 -14.23354 5.673588e-46 1.745439e-42
## 146556 199.0275 -4.732812 0.3349705 -14.12904 2.515499e-45 6.771409e-42
## 221091 235.4840 -3.405595 0.2424912 -14.04420 8.360530e-45 2.000489e-41
## 1589   136.3388 -4.366090 0.3124977 -13.97159 2.323788e-44 5.004278e-41
## 2674   262.9957 -4.177935 0.3030151 -13.78788 3.014997e-43 5.902542e-40
## 1590   172.1416 -3.998853 0.2918134 -13.70346 9.680062e-43 1.737168e-39
## 7146   581.8593 -4.808086 0.3523215 -13.64687 2.107349e-42 3.490905e-39
## 55022  407.2087 -3.356706 0.2495187 -13.45272 2.967443e-41 4.564563e-38
## 7148   3527.1622 -4.903672 0.3647362 -13.44443 3.319385e-41 4.765530e-38

top15_table <- top15[, c("log2FoldChange", "pvalue", "padj")]
top15_table
```

```

##      log2FoldChange      pvalue      padj
## 221476    -7.681227 7.272694e-86 1.566175e-81
## 1675     -4.964090 6.180072e-71 6.654392e-67
## 286133    -5.509867 9.260587e-55 6.647558e-51
## 7123     -4.777304 1.654936e-53 8.909761e-50
## 9340     -5.950893 2.681586e-53 1.154959e-49
## 11170    -4.797705 8.632901e-50 3.098492e-46
## 219348    -3.394889 5.673588e-46 1.745439e-42
## 146556    -4.732812 2.515499e-45 6.771409e-42
## 221091    -3.405595 8.360530e-45 2.000489e-41
## 1589     -4.366090 2.323788e-44 5.004278e-41
## 2674     -4.177935 3.014997e-43 5.902542e-40
## 1590     -3.998853 9.680062e-43 1.737168e-39
## 7146     -4.808086 2.107349e-42 3.490905e-39
## 55022    -3.356706 2.967443e-41 4.564563e-38
## 7148     -4.903672 3.319385e-41 4.765530e-38

```

## STEP 4- showing the top 15 genes

```
# find significant DEGs
sum( res$padj < 0.1, na.rm=TRUE )
```

```
## [1] 13250
```

```
# clean and delete na
res_df <- as.data.frame(res)
res_df <- res_df[!is.na(res_df$padj), ]

#order by significance
res_ordered <- res_df[order(res_df$padj), ]

# extract top15
top15 <- head(res_ordered, 15)
top15
```

```

##      baseMean log2FoldChange      lfcSE      stat      pvalue      padj
## 221476 1945.5333    -7.681227 0.3911341   -19.63835 7.272694e-86 1.566175e-81
## 1675  4452.1590    -4.964090 0.2787639   -17.80751 6.180072e-71 6.654392e-67
## 286133 1803.0834    -5.509867 0.3535451   -15.58462 9.260587e-55 6.647558e-51
## 7123  1054.3341    -4.777304 0.3102290   -15.39928 1.654936e-53 8.909761e-50
## 9340  161.6683     -5.950893 0.3872252   -15.36804 2.681586e-53 1.154959e-49
## 11170 844.1215     -4.797705 0.3233931   -14.83552 8.632901e-50 3.098492e-46
## 219348 435.3765     -3.394889 0.2385134   -14.23354 5.673588e-46 1.745439e-42
## 146556 199.0275     -4.732812 0.3349705   -14.12904 2.515499e-45 6.771409e-42
## 221091 235.4840     -3.405595 0.2424912   -14.04420 8.360530e-45 2.000489e-41
## 1589   136.3388     -4.366090 0.3124977   -13.97159 2.323788e-44 5.004278e-41
## 2674   262.9957     -4.177935 0.3030151   -13.78788 3.014997e-43 5.902542e-40
## 1590   172.1416     -3.998853 0.2918134   -13.70346 9.680062e-43 1.737168e-39
## 7146   581.8593     -4.808086 0.3523215   -13.64687 2.107349e-42 3.490905e-39
## 55022  407.2087     -3.356706 0.2495187   -13.45272 2.967443e-41 4.564563e-38
## 7148   3527.1622     -4.903672 0.3647362   -13.44443 3.319385e-41 4.765530e-38

```

```
top15_table <- top15[, c("log2FoldChange", "pvalue", "padj")]
top15_table
```

```

##      log2FoldChange      pvalue      padj
## 221476    -7.681227 7.272694e-86 1.566175e-81
## 1675     -4.964090 6.180072e-71 6.654392e-67
## 286133    -5.509867 9.260587e-55 6.647558e-51
## 7123     -4.777304 1.654936e-53 8.909761e-50
## 9340     -5.950893 2.681586e-53 1.154959e-49
## 11170    -4.797705 8.632901e-50 3.098492e-46
## 219348    -3.394889 5.673588e-46 1.745439e-42
## 146556    -4.732812 2.515499e-45 6.771409e-42
## 221091    -3.405595 8.360530e-45 2.000489e-41
## 1589     -4.366090 2.323788e-44 5.004278e-41
## 2674     -4.177935 3.014997e-43 5.902542e-40
## 1590     -3.998853 9.680062e-43 1.737168e-39
## 7146     -4.808086 2.107349e-42 3.490905e-39
## 55022    -3.356706 2.967443e-41 4.564563e-38
## 7148     -4.903672 3.319385e-41 4.765530e-38

```

## STEP 5- plotting for visualization

```

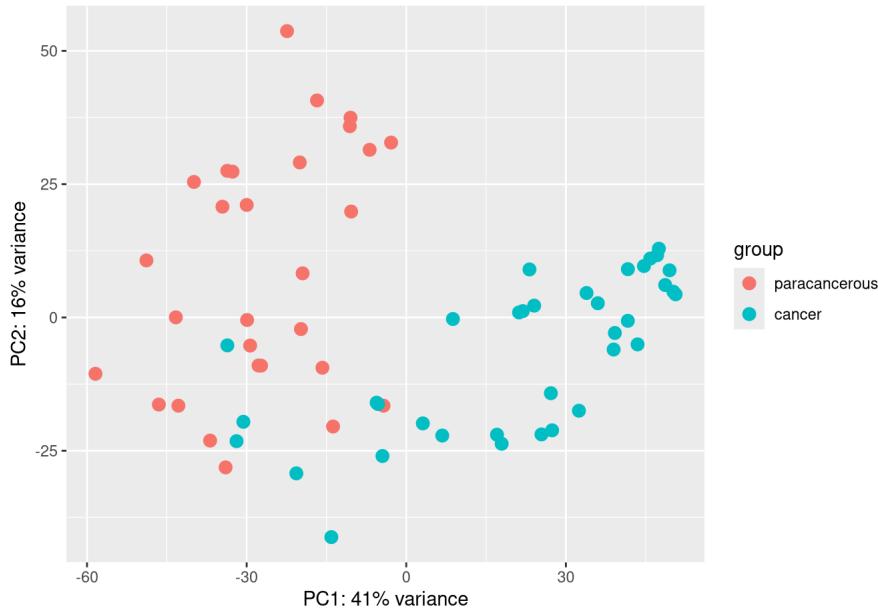
library(pheatmap)
library(ggplot2)

# 1-Data quality control plots
# PCA plot

vsd <- vst(dds, blind=FALSE)
plotPCA(vsd, intgroup = "condition", ntop = 500)

## using ntop=500 top features by variance

```



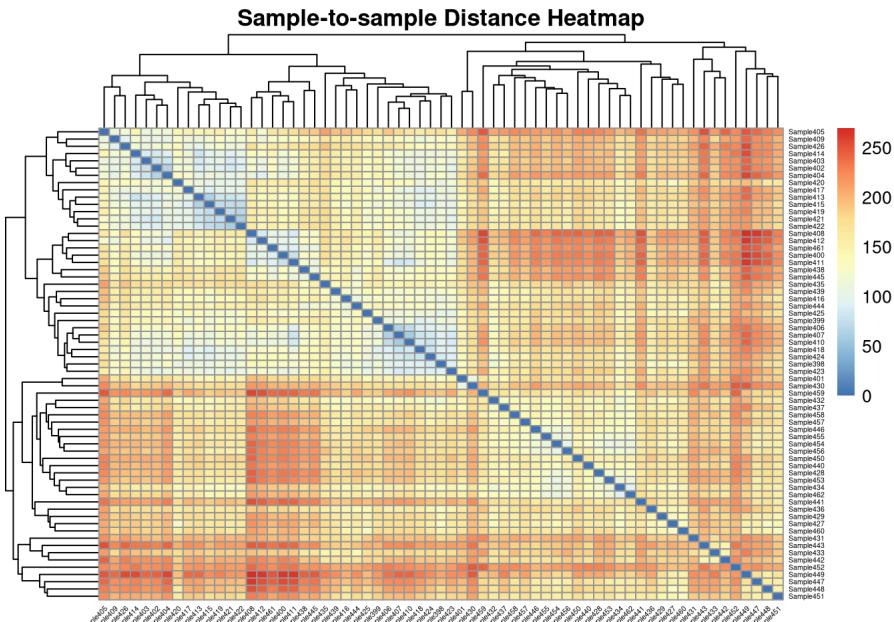
```

# simple distance heatmap- overall gene-expression profiles.
# compute sample-to-sample distances
dists <- dist(t(assay(vsd)))

# convert to matrix
mat <- as.matrix(dists)

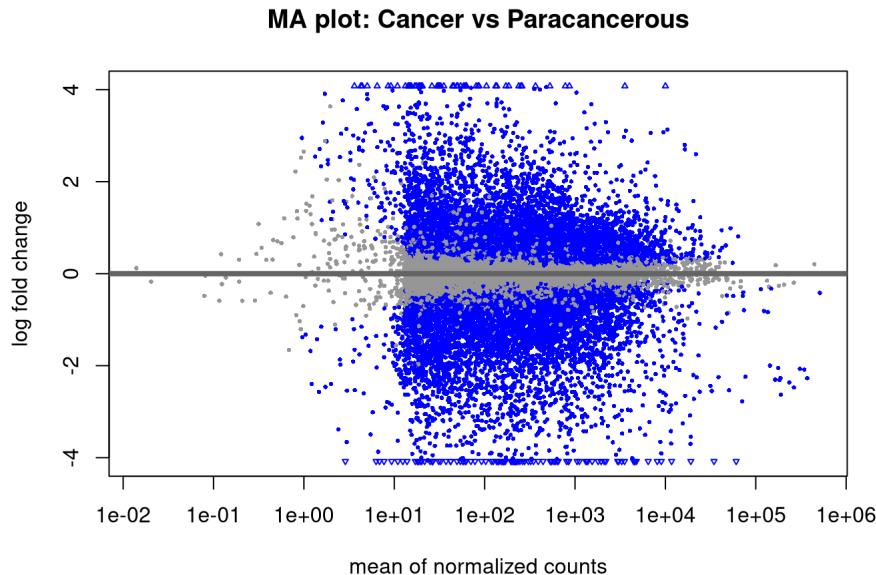
pheatmap(mat,
          clustering_distance_rows = dists,
          clustering_distance_cols = dists,
          show_rownames = TRUE,
          show_colnames = TRUE,
          fontsize_row = 4,
          fontsize_col = 4,
          angle_col = 45,
          main = "Sample-to-sample Distance Heatmap")

```



**## 2 – Differential expression plots**

```
# MA plot: log2 fold change versus mean expression
# shows the pattern of genes that are up- or down-regulated
plotMA(
  res,
  main = "MA plot: Cancer vs Paracancerous"
)
```



```
install.packages("BiocManager")
```

```
## Installing package into '/opt/app-root/src/Rpackages/4.3'
## (as 'lib' is unspecified)
```

```
BiocManager::install("EnhancedVolcano")
```

```
## 'getOption("repos")' replaces Bioconductor standard repositories, see
## 'help("repositories", package = "BiocManager")' for details.
## Replacement repositories:
##   CRAN: https://cran.rstudio.com/
```

```
## Bioconductor version 3.20 (BiocManager 1.30.27), R 4.4.3 (2025-02-28)
```

```
## Warning: package(s) not installed when version(s) same as or greater than current; use
##   `force = TRUE` to re-install: 'EnhancedVolcano'
```

```
## Installation paths not writeable, unable to update packages
## path: /usr/lib64/R/library
## packages:
##   AER, aplot, arrow, bayesplot, bigD, BiocManager, BiocParallel, bookdown,
##   boot, BradleyTerry2, brglm, broom, Cairo, checkmate, cli, clock, cluster,
##   colorspace, commonmark, covr, cowplot, credentials, crosstalk, Cubist,
##   curl, data.table, dbplyr, dbscan, dendextend, DEoptimR, Deriv, devtools,
##   dials, diffobj, digest, doBy, doFuture, doRNG, DOSE, downlit, DT, dtplyr,
##   effects, emdbook, emmeans, entropy, Epi, epiR, etm, evaluate, extrafont,
##   extrafontdb, FactoMineR, fastcluster, fgsea, fitdistrplus, flextable,
##   foghorn,forcats, forecast, foreign, fs, future, future.apply,
##   future.callr, gam, gapminder, gangle, gclus, gcookbook, gdtools, generics,
##   gert, GGally, gganimate, ggforce, ggfortify, ggfun, ggnewscale, ggplot2,
##   ggplotify, ggpp, ggpublisher, ggraph, ggridges, ggsci, ggstats, ggtangle,
##   ggvenn, gh, glmnet, globals, googledrive, googlesheets4, GPfit, gprofiler2,
##   gss, gt, hardhat, harmony, haven, here, Hmisc, HMM, hms, httpuv, httr2,
##   hunspell, igraph, insight, jsonlite, keras, KFAS, kinship2, knitr,
##   labelled, Lahman, later, lattice, lava, leidenbase, lgr, listenv, littler,
##   lme4, lobstr, logger, magrittr, mapproj, maps, markdown, mathjaxr, Matrix,
##   MatrixModels, mclust, memisc, mgcv, mime, minuiUI, mlr, mlr3, mlr3measures,
##   mlr3misc, mockery, mockr, modeltools, multcomp, MuMin, nanotime, ncdf4,
##   nlme, nloptr, officer, openssl, openxlsx, optimx, ottr, parallelly,
##   parsnip, partykit, patchwork, pbapply, pbdZMQ, pbkrtest, permute, pheatmap,
##   pillar, pixmap, pkgbuild, pkgdown, pkgload, plotly, plotrix, popr, pracma,
##   pROC, proddim, profmem, progressr, promises, PRRROC, ps, purrr, qtl2,
##   quantmod, questionr, QuickJSR, R.cache, R.oo, ragg, raster, rbibutils,
##   Rcpp, RcppArmadillo, RcppDate, RcppParallel, RCurl, Rdpack, readr, recipes,
##   reformulas, rentrez, reshape, reshape2, restfulr, reticulate, rgl, rlang,
##   rmarkdown, rms, RMySQL, robustbase, roxygen2, RPostgreSQL, rprojroot,
##   rsample, RSQLite, rstantools, rstatix, Rttf2pt1, RUnit, rversions, rvest,
##   s2, S7, safetensors, sass, scales, scatterpie, sctransform, seriation,
##   Seurat, SeuratObject, sf, sfsmisc, shapr, shiny, slider, sparsevctrs,
##   spatstat, spatstat.data, spatstat.explore, spatstat.geom, spatstat.linnet,
##   spatstat.model, spatstat.random, spatstat.univar, spatstat.utils, spelling,
##   spls, statmod, statnet.common, stringi, stringr, styler, survey, svglite,
##   systemfonts, tensor, tensorflow, terra, testthat, textshaping, tfruns,
##   TH.data, threejs, tibble, timeDate, tinytex, TSP, tune, tzdb, units,
##   usethis, utf8, uwot, V8, vcdExtra, vegan, visNetwork, vroom, waldo, warp,
##   workflows, WriteXLS, xfun, xgboost, XML, xml2, yulab.utils, zeallot, zip,
##   zoo
```

```
# Volcano plot
library(EnhancedVolcano)
```

```
## Loading required package: ggrepel
```

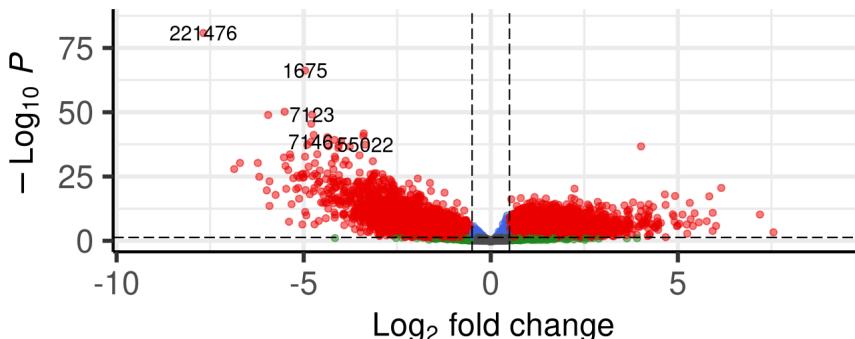
```
# get the row names of the top 15 genes.
# these row names correspond to the gene IDs in the DESeq2 result object.
top15_genes <- rownames(top15_table)

# making a volcano plot, highlighting only the top 15 most significant genes
EnhancedVolcano(
  res,
  lab      = rownames(res),      # labels for all genes (by row name)
  x        = "log2FoldChange",    # x-axis: effect size (Cancer / Paracancerous)
  y        = "padj",             # y-axis: adjusted p-value
  pCutoff  = 0.05,              # significance threshold for padj
  FCcutoff = 0.5,               # threshold for |log2 fold change|
  selectLab = top15_genes,      # only these genes are labelled on the plot
  pointSize = 1.5,
  labSize   = 4,
  col       = c("grey30", "forestgreen", "royalblue", "red2"),
  title     = "Volcano Plot (Top 15 Most Significant Genes)",
  subtitle  = "Top genes labelled based on lowest adjusted p-values"
)
```

## Volcano Plot (Top 15 Most Significant Genes)

Top genes labelled based on lowest adjusted p-values

● NS ● Log<sub>2</sub> FC ● p-value ● p – value and log<sub>2</sub> FC



total = 21535 variables

```
# 3- Biological interpretation plots

library(RColorBrewer)

# Heatmap for Top 15 DEGs

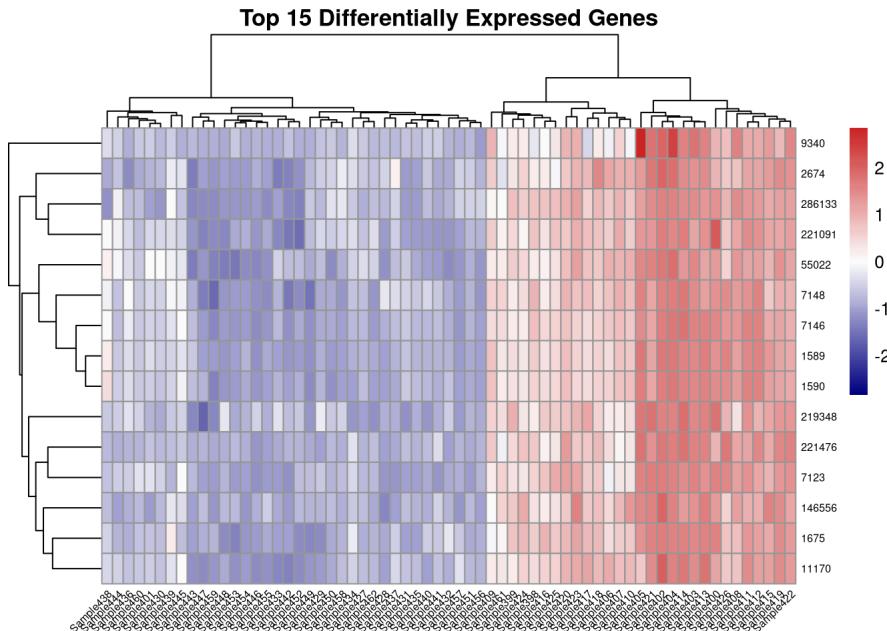
# extract expression matrix from VST
vsd_mat <- assay(vsd)

# taking the rownames of top15 genes
top15_genes <- rownames(top15_table)

# intersect to avoid missing genes (just in case)
top15_genes <- intersect(top15_genes, rownames(vsd_mat))

# subset the VST matrix
heatmap_top15 <- vsd_mat[top15_genes, ]

# plot heatmap
pheatmap(
  heatmap_top15,
  scale = "row",
  clustering_distance_rows = "euclidean",
  clustering_distance_cols = "euclidean",
  clustering_method = "complete",
  show_rownames = TRUE,
  show_colnames = TRUE,
  font_size_row = 6,
  font_size_col = 6,
  angle_col = 45,
  color = colorRampPalette(c("navy", "white", "firebrick3"))(100),
  main = "Top 15 Differentially Expressed Genes"
)
```



```

library(reshape2)
library(ggplot2)

# Choosing how many genes to show in the boxplots (we went with top 6)
top_genes <- rownames(top15_table)[1:6]

# subset the VST expression matrix to only these genes
expr_top <- vsd_mat[top_genes, ]

# convert the matrix to long format for ggplot2
df <- melt(expr_top)
colnames(df) <- c("Gene", "Sample", "Expression")

# add group (condition) information for each sample
# 'condition' is the column we created earlier in pdat / colData(dds)
df$condition <- colData(vsd)$condition[match(df$Sample, rownames(colData(vsd)))] 

# making boxplots of expression per condition for each gene
ggplot(df, aes(x = condition, y = Expression, fill = condition)) +
  geom_boxplot(outlier.shape = NA, alpha = 0.8) +
  geom_jitter(width = 0.2, size = 1.3, alpha = 0.7) +
  facet_wrap(~ Gene, scales = "free_y", ncol = 3) +
  theme_bw(base_size = 12) +
  theme(
    strip.text      = element_text(size = 12, face = "bold"),
    legend.position = "none"
  ) +
  ylab("VST normalized expression") +
  ggtitle("Expression Boxplots of Top Differentially Expressed Genes")

```

