

```
library(DESeq2); load("CancerAndParacancer.RData")
```

```
## Loading required package: S4Vectors
```

```
## Loading required package: stats4
```

```
## Loading required package: BiocGenerics
```

```
##
## Attaching package: 'BiocGenerics'
```

```
## The following objects are masked from 'package:stats':
##
##   IQR, mad, sd, var, xtabs
```

```
## The following objects are masked from 'package:base':
##
##   anyDuplicated, aperm, append, as.data.frame, basename, cbind,
##   colnames, dirname, do.call, duplicated, eval, evalq, Filter, Find,
##   get, grep, grepl, intersect, is.unsorted, lapply, Map, mapply,
##   match, mget, order, paste, pmax, pmax.int, pmin, pmin.int,
##   Position, rank, rbind, Reduce, rownames, sapply, saveRDS, setdiff,
##   table, tapply, union, unique, unsplit, which.max, which.min
```

```
##
## Attaching package: 'S4Vectors'
```

```
## The following object is masked from 'package:utils':
##
##   findMatches
```

```
## The following objects are masked from 'package:base':
##
##   expand.grid, I, unname
```

```
## Loading required package: IRanges
```

```
## Loading required package: GenomicRanges
```

```
## Loading required package: GenomeInfoDb
```

```
## Loading required package: SummarizedExperiment
```

```
## Loading required package: MatrixGenerics
```

```
## Loading required package: matrixStats
```

```
##
## Attaching package: 'MatrixGenerics'
```

```
## The following objects are masked from 'package:matrixStats':
##
##   colAlls, colAnyNAs, colAnys, colAvgsPerRowSet, colCollapse,
##   colCounts, colCummaxs, colCummins, colCumprods, colCumsums,
##   colDiffs, colIQRDiffs, colIQRs, colLogSumExps, colMadDiffs,
##   colMads, colMaxs, colMeans2, colMedians, colMins, colOrderStats,
##   colProds, colQuantiles, colRanges, colRanks, colSdDiffs, colSds,
##   colSums2, colTabulates, colVarDiffs, colVars, colWeightedMads,
##   colWeightedMeans, colWeightedMedians, colWeightedSds,
##   colWeightedVars, rowAlls, rowAnyNAs, rowAnys, rowAvgsPerColSet,
##   rowCollapse, rowCounts, rowCummaxs, rowCummins, rowCumprods,
##   rowCumsums, rowDiffs, rowIQRDiffs, rowIQRs, rowLogSumExps,
##   rowMadDiffs, rowMads, rowMaxs, rowMeans2, rowMedians, rowMins,
##   rowOrderStats, rowProds, rowQuantiles, rowRanges, rowRanks,
##   rowSdDiffs, rowSds, rowSums2, rowTabulates, rowVarDiffs, rowVars,
##   rowWeightedMads, rowWeightedMeans, rowWeightedMedians,
##   rowWeightedSds, rowWeightedVars
```

```
## Loading required package: Biobase
```

```
## Welcome to Bioconductor
##
## Vignettes contain introductory material; view with
## 'browseVignettes()'. To cite Bioconductor, see
## 'citation("Biobase")', and for packages 'citation("pkgname")'.
```

```
##
## Attaching package: 'Biobase'
```

```
## The following object is masked from 'package:MatrixGenerics':
##
## rowMedians
```

```
## The following objects are masked from 'package:matrixStats':
##
## anyMissing, rowMedians
```

```
expr <- ExprMat; pdat <- pDataMat; feat <- FeatureMat
same_samples <- intersect(colnames(expr), rownames(pdat))
expr <- expr[, same_samples, drop = F]
pdat <- pdat[same_samples, , drop = F]
expr <- expr[rowSums(expr) >= 954, , drop = F]
expr <- expr[!duplicated(rownames(expr)), , drop = F]
feat <- feat[!duplicated(rownames(feat)), , drop = F]
new_col <- intersect(c("gene_symbol", "symbol", "chromosome", "description", "gene_biotype"), colnames(feat))
feat <- feat[, new_col, drop = F]
feat <- feat[rownames(expr), , drop = F]
colnames(pdat)[1] <- "condition"
pdat$condition <- factor(pdat$condition)
pdat$condition <- relevel(pdat$condition, ref = "paracancerous")
dds <- DESeqDataSetFromMatrix(countData = expr, colData = pdat, design = ~ condition)
dds <- DESeq(dds); res <- results(dds)
```

```
## estimating size factors
```

```
## estimating dispersions
```

```
## gene-wise dispersion estimates
```

```
## mean-dispersion relationship
```

```
## final dispersion estimates
```

```
## fitting model and testing
```

```
## -- replacing outliers and refitting for 1371 genes
## -- DESeq argument 'minReplicatesForReplace' = 7
## -- original counts are preserved in counts(dds)
```

```
## estimating dispersions
```

```
## fitting model and testing
```

```
res_ordered <- res[order(res$padj, na.last = NA), ]
des_order <- as.data.frame(res_ordered)
normal_des <- as.data.frame(counts(dds, normalized = T))
des_order$gene <- rownames(des_order)
top_genes <- rownames(des_order)[1:15]
top_genes <- top_genes[top_genes %in% rownames(FeatureMat)]
cutoff <- min(rowSums(expr)[top_genes]); cat("cutoff is:", cutoff, "\n")
```

```
## cutoff is: 9296
```

```
top <- cbind(GeneID = top_genes, FeatureMat[top_genes, , drop = F],
  des_order[top_genes, c("log2FoldChange", "pvalue", "padj", "stat", "lfcSE", "baseMean"), drop = F])
up_order <- order(des_order$log2FoldChange, decreasing = T)
up_genes <- rownames(des_order)[up_order][1:10]
up_genes <- up_genes[up_genes %in% rownames(FeatureMat)]
cutoff <- min(rowSums(expr)[up_genes]); cat("cutoff is:", cutoff, "\n")
```

```
## cutoff is: 1043
```

```
top_up <- cbind(GeneID = up_genes, FeatureMat[up_genes, , drop = F],
  des_order[up_genes, c("log2FoldChange", "pvalue", "padj", "stat", "lfcSE", "baseMean"), drop = F])
down_order <- order(des_order$log2FoldChange, decreasing = F)
down_genes <- rownames(des_order)[down_order][1:10]
down_genes <- down_genes[down_genes %in% rownames(FeatureMat)]
cutoff <- min(rowSums(expr)[down_genes]); cat("cutoff is:", cutoff, "\n")
```

```
## cutoff is: 954
```

```
top_down <- cbind(GeneID = down_genes, FeatureMat[down_genes, , drop = F],
  des_order[down_genes, c("log2FoldChange", "pvalue", "padj", "stat", "lfcSE", "baseMean"), drop = F])
summary(top); knitr::kable(top)
```

```
##      GeneID      GeneID      Symbol      Description
## Length:15      Min.   : 1589 Length:15      Length:15
## Class :character 1st Qu.: 4898 Class :character Class :character
## Mode  :character Median : 9340 Mode  :character Mode  :character
##                Mean  : 79939
##                3rd Qu.:182952
##                Max.   :286133
##      Synonyms      GeneType      EnsemblGeneID      Status
## Length:15      Length:15      Length:15      Length:15
## Class :character Class :character Class :character Class :character
## Mode  :character Mode  :character Mode  :character Mode  :character
##
##
##      ChrAcc      ChrStart      ChrStop      Orientation
## Length:15      Length:15      Length:15      Length:15
## Class :character Class :character Class :character Class :character
## Mode  :character Mode  :character Mode  :character Mode  :character
##
##
##      Length      GOFunctionID      GOProcessID      GOComponentID
## Min.   : 947      Length:15      Length:15      Length:15
## 1st Qu.: 2186      Class :character Class :character Class :character
## Median : 2429      Mode  :character Mode  :character Mode  :character
## Mean   : 4340
## 3rd Qu.: 5370
## Max.   :13899
##      GOFunction      GOProcess      GOComponent      log2FoldChange
## Length:15      Length:15      Length:15      Min.   : -7.681
## Class :character Class :character Class :character 1st Qu.: -4.934
## Mode  :character Mode  :character Mode  :character Median : -4.777
##                Mean   : -4.722
##                3rd Qu.: -4.088
##                Max.   : -3.357
##      pvalue      padj      stat      lfcSE
## Min.   :0.000e+00 Min.   :0.000e+00 Min.   : -19.64 Min.   :0.2385
## 1st Qu.:0.000e+00 1st Qu.:0.000e+00 1st Qu.: -15.38 1st Qu.:0.2853
## Median :2.520e-45 Median :6.770e-42 Median : -14.13 Median :0.3125
## Mean   :4.419e-42 Mean   :6.613e-39 Mean   : -14.87 Mean   :0.3156
## 3rd Qu.:6.347e-43 3rd Qu.:1.164e-39 3rd Qu.: -13.75 3rd Qu.:0.3529
## Max.   :3.319e-41 Max.   :4.766e-38 Max.   : -13.44 Max.   :0.3911
##      baseMean
## Min.   : 136.3
## 1st Qu.: 217.3
## Median : 435.4
## Mean   :1081.2
## 3rd Qu.:1428.7
## Max.   :4452.2
```

	GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID	Status	ChrAcc
221476	221476	221476	PI16	peptidase inhibitor 16	CD364 CRISP9 MSMBBP PSPBP	protein-coding	ENSG00000164530	active	NC_000
1675	1675	1675	CFD	complement factor D	ADIPSIN ADN DF PFD	protein-coding	ENSG00000197766	active	NC_000
286133	286133	286133	SCARA5	scavenger receptor class A member 5	NET33 Tetr	protein-coding	ENSG00000168079	active	NC_000

	GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID	Status	ChrAcc
7123	7123	7123	CLEC3B	C-type lectin domain family 3 member B	MCDR4 TN TNA	protein-coding	ENSG00000163815	active	NC_000
9340	9340	9340	GLP2R	glucagon like peptide 2 receptor		protein-coding	ENSG00000065325	active	NC_000
11170	11170	11170	FAM107A	family with sequence similarity 107 member A	DRR1 TU3A	protein-coding	ENSG00000168309	active	NC_000
219348	219348	219348	PLAC9	placenta associated 9		protein-coding	ENSG00000189129	active	NC_000
146556	146556	146556	C16orf89	chromosome 16 open reading frame 89		protein-coding	ENSG00000153446	active	NC_000
221091	221091	221091	LRRN4CL	LRRN4 C-terminal like		protein-coding	ENSG00000177363	active	NC_000
1589	1589	1589	CYP21A2	cytochrome P450 family 21 subfamily A member 2	CA21H CAH1 CPS1 CYP21 CYP21B P450c21B	protein-coding	ENSG00000231852	active	NC_000
2674	2674	2674	GFRA1	GDNF family receptor alpha 1	GDNFR GDNFR-alpha-1 GDNFRA GFR-ALPHA-1 GFRalpha-1 RET1L RETL1 RHDA4 TRNR1	protein-coding	ENSG00000151892	active	NC_000
1590	1590	1590	CYP21A1P	cytochrome P450 family 21 subfamily A member 1, pseudogene	CYP21A CYP21P P450c21A	pseudo	ENSG00000290788	active	NC_000
7146	7146	7146	TNXA	tenascin XA (pseudogene)	D6S103E HXBL TNX XA	pseudo		active	NC_000

	GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID	Status	ChrAcc
55022	55022	55022	PID1	phosphotyrosine interaction domain containing 1	HMFN2073 NYGGF4 P-CL1 PCL1	protein-coding	ENSG00000153823	active	NC_000

7148	7148	7148	TNXB	tenascin XB	EDS3 EDSCLL EDSCLL1 HXBL TENX TN-X TNX TNXB1 TNXB2 TNXBS VUR8 XB XBS	protein-coding	ENSG00000168477	active	NC_000
------	------	------	------	-------------	--	----------------	-----------------	--------	--------

summary(top\_up);knitr::kable(top\_up)

```
##      GeneID      GeneID      Symbol      Description
## Length:10      Min.      : 1472      Length:10      Length:10
## Class :character 1st Qu.: 19677      Class :character  Class :character
## Mode  :character Median : 109906      Mode  :character  Mode  :character
##                               Mean  : 41924921
##                               3rd Qu.:101929231
##                               Max.  :109729169
##      Synonyms      GeneType      EnsemblGeneID      Status
## Length:10      Length:10      Length:10      Length:10
## Class :character  Class :character  Class :character  Class :character
## Mode  :character  Mode  :character  Mode  :character  Mode  :character
##
##
##      ChrAcc      ChrStart      ChrStop      Orientation
## Length:10      Length:10      Length:10      Length:10
## Class :character  Class :character  Class :character  Class :character
## Mode  :character  Mode  :character  Mode  :character  Mode  :character
##
##
##      Length      GOFunctionID      GOProcessID      GOComponentID
## Min.      : 679      Length:10      Length:10      Length:10
## 1st Qu.:1236      Class :character  Class :character  Class :character
## Median :1519      Mode  :character  Mode  :character  Mode  :character
## Mean  :1804
## 3rd Qu.:2160
## Max.  :4158
##      GOFunction      GOProcess      GOComponent      log2FoldChange
## Length:10      Length:10      Length:10      Min.      :5.448
## Class :character  Class :character  Class :character  1st Qu.:5.788
## Mode  :character  Mode  :character  Mode  :character  Median :5.931
##                               Mean  :6.141
##                               3rd Qu.:6.124
##                               Max.  :7.555
##      pvalue      padj      stat      lfcSE
## Min.      :0.000e+00      Min.      :0.000e+00      Min.      :3.806      Min.      :0.6162
## 1st Qu.:0.000e+00      1st Qu.:0.000e+00      1st Qu.:5.354      1st Qu.:0.7731
## Median :9.300e-10      Median :1.230e-08      Median :6.512      Median :0.9580
## Mean  :1.763e-05      Mean  :6.626e-05      Mean  :6.558      Mean  :1.0348
## 3rd Qu.:1.525e-07      3rd Qu.:1.219e-06      3rd Qu.:7.230      3rd Qu.:1.1248
## Max.  :1.410e-04      Max.  :5.105e-04      Max.  :9.996      Max.  :1.9849
##      baseMean
## Min.      : 10.84
## 1st Qu.: 21.86
## Median : 33.03
## Mean  : 41.44
## 3rd Qu.: 45.57
## Max.  :132.55
```

	GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID	Status	ChrAcc
51350	51350	51350	KRT76	keratin 76	HUMCYT2A KRT2B KRT2P	protein-coding	ENSG00000185069	active	NC_000011
9119	9119	9119	KRT75	keratin 75	CK-75 K6HF K75 KB18 PFB hK6hf	protein-coding	ENSG00000170454	active	NC_000011
1472	1472	1472	CST4	cystatin S		protein-coding	ENSG00000101441	active	NC_000021
101929412	101929412	101929412	LINC02212	long intergenic non-protein coding RNA 2212		ncRNA	ENSG00000249396	active	NC_000004
101928687	101928687	101928687	LNCAROD	lncRNA activating regulator of DKK1	A-ROD LINC01468 lnc-MBL2-4	ncRNA	ENSG00000231131	active	NC_000011

	GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID	Status	ChrAcc
6704	6704	6704	SPRR2E	small proline rich protein 2E		protein-coding	ENSG00000203785	active	NC_000004
56033	56033	56033	BARX1	BARX homeobox 1		protein-coding	ENSG00000131668	active	NC_000004
163778	163778	163778	SPRR4	small proline rich protein 4		protein-coding	ENSG00000184148	active	NC_000004
105373485	105373485	105373485	LOC105373485	uncharacterized LOC105373485		ncRNA		active	NC_000004
109729169	109729169	109729169	LINC02154	long intergenic non-protein coding RNA 2154		ncRNA	ENSG00000235385	active	NC_000004

summary(top\_down);knitr::kable(top\_down)

```
##      GeneID      GeneID      Symbol      Description
## Length:10      Min.      : 70      Length:10      Length:10
## Class :character 1st Qu.: 4770      Class :character Class :character
## Mode :character Median : 7276      Mode :character Mode :character
##                               Mean  : 10283197
##                               3rd Qu.: 187198
##                               Max.  :101927499
##      Synonyms      GeneType      EnsemblGeneID      Status
## Length:10      Length:10      Length:10      Length:10
## Class :character Class :character Class :character Class :character
## Mode :character Mode :character Mode :character Mode :character
##
##
##      ChrAcc      ChrStart      ChrStop      Orientation
## Length:10      Length:10      Length:10      Length:10
## Class :character Class :character Class :character Class :character
## Mode :character Mode :character Mode :character Mode :character
##
##
##      Length      GOFunctionID      GOProcessID      GOComponentID
## Min.      : 382.0      Length:10      Length:10      Length:10
## 1st Qu.: 767.5      Class :character Class :character Class :character
## Median :1741.0      Mode :character Mode :character Mode :character
## Mean      :2201.6
## 3rd Qu.:2241.2
## Max.      :6769.0
##      GOFunction      GOProcess      GOComponent      log2FoldChange
## Length:10      Length:10      Length:10      Min.      :−7.681
## Class :character Class :character Class :character 1st Qu.:−6.584
## Mode :character Mode :character Mode :character Median :−6.087
##                               Mean      :−6.317
##                               3rd Qu.:−5.924
##                               Max.      :−5.751
##      pvalue      padj      stat      lfcSE
## Min.      :0.000e+00      Min.      :0.000e+00      Min.      :−19.638      Min.      :0.3872
## 1st Qu.:0.000e+00      1st Qu.:0.000e+00      1st Qu.:−12.127      1st Qu.:0.5233
## Median :0.000e+00      Median :0.000e+00      Median :−11.327      Median :0.5599
## Mean      :5.015e−17      Mean      :2.762e−15      Mean      :−11.969      Mean      :0.5514
## 3rd Qu.:1.000e−22      3rd Qu.:1.900e−20      3rd Qu.: −9.968      3rd Qu.:0.6075
## Max.      :5.015e−16      Max.      :2.762e−14      Max.      : −8.111      Max.      :0.7293
##      baseMean
## Min.      : 19.44
## 1st Qu.: 145.78
## Median : 242.39
## Mean      : 4194.07
## 3rd Qu.: 1797.90
## Max.      :34495.23
```

	GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID
221476	221476	221476	PI16	peptidase inhibitor 16	CD364 CRISP9 MSMBBP PSPBP	protein-coding	ENSG00000164530



	GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID
4653	4653	4653	MYOC	myocilin	GLC1A GPOA JOAG JOAG1 TIGR	protein-coding	ENSG00000034971
572558	572558	572558	PGM5-AS1	PGM5 antisense RNA 1	FAM233A	ncRNA	ENSG00000224958
5212	5212	5212	VIT	vitrin	VIT1	protein-coding	ENSG00000205221
84366	84366	84366	PRAC1	PRAC1 small nuclear protein	C17orf92 PRAC	protein-coding	ENSG00000159182
70	70	70	ACTC1	actin alpha cardiac muscle 1	ACTC ASD5 CMD1R CMH11 LVNC4	protein-coding	ENSG00000159251

	GeneID	GeneID	Symbol	Description	Synonyms	GeneType	EnsemblGeneID
9340	9340	9340	GLP2R	glucagon like peptide 2 receptor		protein-coding	ENSG00000065325
101927499	101927499	101927499	LOC101927499	uncharacterized LOC101927499		ncRNA	
1674	1674	1674	DES	desmin	CDCD3 CSM1 CSM2 LGMD1D LGMD1E LGMD2R	protein-coding	ENSG00000175084
5121	5121	5121	PCP4	Purkinje cell protein 4	PEP-19	protein-coding	ENSG00000183036

## STEP 4- showing the top 15 genes

```
# find significant DEGs
sum( res$padj < 0.1, na.rm=TRUE )
```

```
## [1] 13250
```

```
# clean and delete na
res_df <- as.data.frame(res)
res_df <- res_df[!is.na(res_df$padj), ]

#order by significance
res_ordered <- res_df[order(res_df$padj), ]

# extract top15
top15 <- head(res_ordered, 15)
top15
```

```
##      baseMean log2FoldChange      lfcSE      stat      pvalue      padj
## 221476 1945.5333      -7.681227 0.3911341 -19.63835 7.272694e-86 1.566175e-81
## 1675   4452.1590      -4.964090 0.2787639 -17.80751 6.180072e-71 6.654392e-67
## 286133 1803.0834      -5.509867 0.3535451 -15.58462 9.260587e-55 6.647558e-51
## 7123   1054.3341      -4.777304 0.3102290 -15.39928 1.654936e-53 8.909761e-50
## 9340   161.6683      -5.950893 0.3872252 -15.36804 2.681586e-53 1.154959e-49
## 11170   844.1215      -4.797705 0.3233931 -14.83552 8.632901e-50 3.098492e-46
## 219348 435.3765      -3.394889 0.2385134 -14.23354 5.673588e-46 1.745439e-42
## 146556 199.0275      -4.732812 0.3349705 -14.12904 2.515499e-45 6.771409e-42
## 221091 235.4840      -3.405595 0.2424912 -14.04420 8.360530e-45 2.000489e-41
## 1589   136.3388      -4.366090 0.3124977 -13.97159 2.323788e-44 5.004278e-41
## 2674   262.9957      -4.177935 0.3030151 -13.78788 3.014997e-43 5.902542e-40
## 1590   172.1416      -3.998853 0.2918134 -13.70346 9.680062e-43 1.737168e-39
## 7146   581.8593      -4.808086 0.3523215 -13.64687 2.107349e-42 3.490905e-39
## 55022  407.2087      -3.356706 0.2495187 -13.45272 2.967443e-41 4.564563e-38
## 7148   3527.1622      -4.903672 0.3647362 -13.44443 3.319385e-41 4.765530e-38
```

```
top15_table <- top15[, c("log2FoldChange", "pvalue", "padj")]
top15_table
```

```
##          log2FoldChange      pvalue      padj
## 221476      -7.681227 7.272694e-86 1.566175e-81
## 1675        -4.964090 6.180072e-71 6.654392e-67
## 286133      -5.509867 9.260587e-55 6.647558e-51
## 7123        -4.777304 1.654936e-53 8.909761e-50
## 9340        -5.950893 2.681586e-53 1.154959e-49
## 11170       -4.797705 8.632901e-50 3.098492e-46
## 219348      -3.394889 5.673588e-46 1.745439e-42
## 146556      -4.732812 2.515499e-45 6.771409e-42
## 221091      -3.405595 8.360530e-45 2.000489e-41
## 1589        -4.366090 2.323788e-44 5.004278e-41
## 2674        -4.177935 3.014997e-43 5.902542e-40
## 1590        -3.998853 9.680062e-43 1.737168e-39
## 7146        -4.808086 2.107349e-42 3.490905e-39
## 55022       -3.356706 2.967443e-41 4.564563e-38
## 7148        -4.903672 3.319385e-41 4.765530e-38
```

## STEP 4- showing the top 15 genes

```
# find significant DEGs
sum( res$padj < 0.1, na.rm=TRUE )
```

```
## [1] 13250
```

```
# clean and delete na
res_df <- as.data.frame(res)
res_df <- res_df[!is.na(res_df$padj), ]

#order by significance
res_ordered <- res_df[order(res_df$padj), ]

# extract top15
top15 <- head(res_ordered, 15)
top15
```

```
##          baseMean log2FoldChange      lfcSE      stat      pvalue      padj
## 221476 1945.5333      -7.681227 0.3911341 -19.63835 7.272694e-86 1.566175e-81
## 1675   4452.1590      -4.964090 0.2787639 -17.80751 6.180072e-71 6.654392e-67
## 286133 1803.0834      -5.509867 0.3535451 -15.58462 9.260587e-55 6.647558e-51
## 7123   1054.3341      -4.777304 0.3102290 -15.39928 1.654936e-53 8.909761e-50
## 9340   161.6683       -5.950893 0.3872252 -15.36804 2.681586e-53 1.154959e-49
## 11170   844.1215      -4.797705 0.3233931 -14.83552 8.632901e-50 3.098492e-46
## 219348 435.3765      -3.394889 0.2385134 -14.23354 5.673588e-46 1.745439e-42
## 146556 199.0275      -4.732812 0.3349705 -14.12904 2.515499e-45 6.771409e-42
## 221091 235.4840      -3.405595 0.2424912 -14.04420 8.360530e-45 2.000489e-41
## 1589   136.3388      -4.366090 0.3124977 -13.97159 2.323788e-44 5.004278e-41
## 2674   262.9957      -4.177935 0.3030151 -13.78788 3.014997e-43 5.902542e-40
## 1590   172.1416      -3.998853 0.2918134 -13.70346 9.680062e-43 1.737168e-39
## 7146   581.8593      -4.808086 0.3523215 -13.64687 2.107349e-42 3.490905e-39
## 55022  407.2087      -3.356706 0.2495187 -13.45272 2.967443e-41 4.564563e-38
## 7148   3527.1622     -4.903672 0.3647362 -13.44443 3.319385e-41 4.765530e-38
```

```
top15_table <- top15[, c("log2FoldChange", "pvalue", "padj")]
top15_table
```

```
##          log2FoldChange      pvalue      padj
## 221476      -7.681227 7.272694e-86 1.566175e-81
## 1675        -4.964090 6.180072e-71 6.654392e-67
## 286133      -5.509867 9.260587e-55 6.647558e-51
## 7123        -4.777304 1.654936e-53 8.909761e-50
## 9340        -5.950893 2.681586e-53 1.154959e-49
## 11170       -4.797705 8.632901e-50 3.098492e-46
## 219348      -3.394889 5.673588e-46 1.745439e-42
## 146556      -4.732812 2.515499e-45 6.771409e-42
## 221091      -3.405595 8.360530e-45 2.000489e-41
## 1589        -4.366090 2.323788e-44 5.004278e-41
## 2674        -4.177935 3.014997e-43 5.902542e-40
## 1590        -3.998853 9.680062e-43 1.737168e-39
## 7146        -4.808086 2.107349e-42 3.490905e-39
## 55022       -3.356706 2.967443e-41 4.564563e-38
## 7148        -4.903672 3.319385e-41 4.765530e-38
```

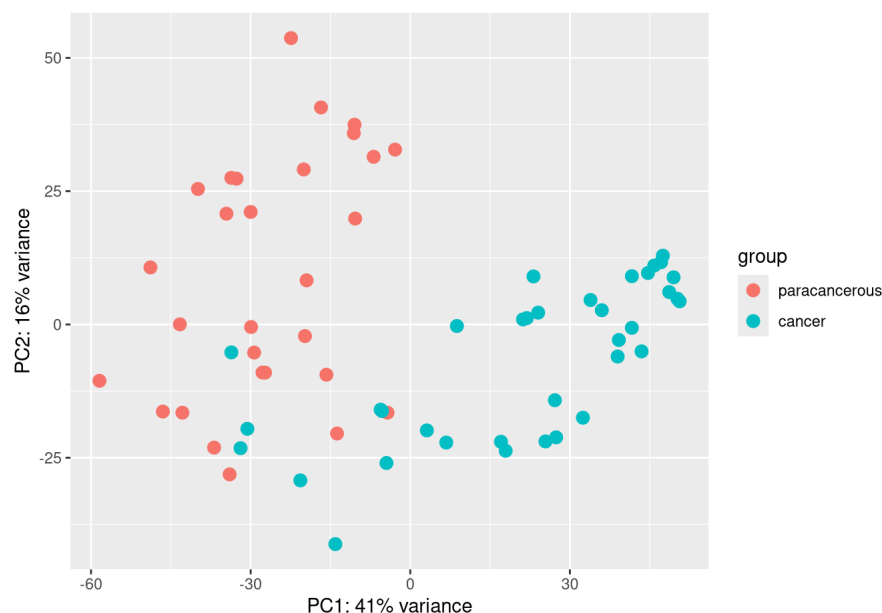
## STEP 5- plotting for visualization

```
library(pheatmap)
library(ggplot2)

# 1-Data quality control plots
# PCA plot

vsd <- vst(dds, blind=FALSE)
plotPCA(vsd, intgroup = "condition", ntop = 500)
```

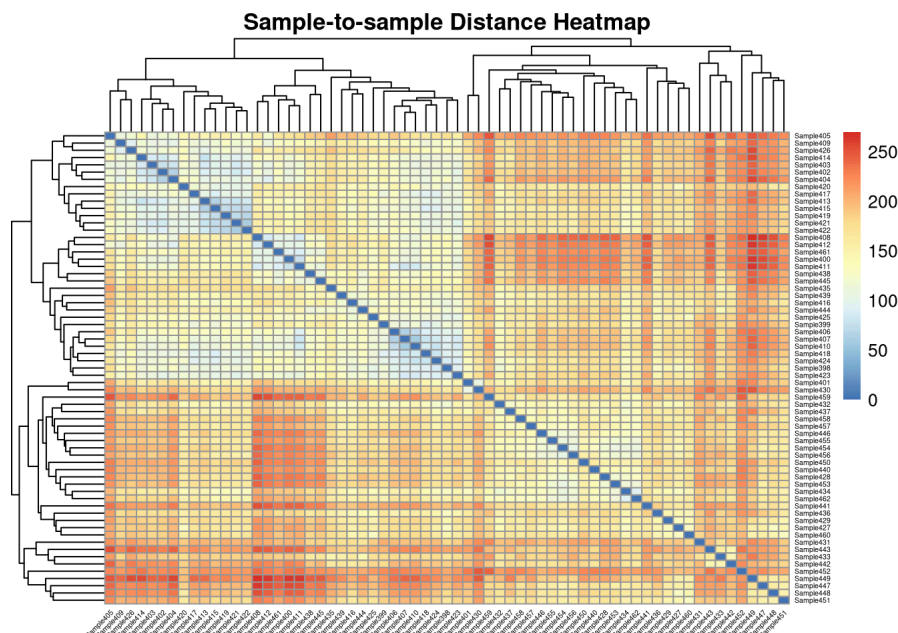
```
## using ntop=500 top features by variance
```



```
# simple distance heatmap- overall gene-expression profiles.
# compute sample-to-sample distances
dists <- dist(t(assay(vsd)))

# convert to matrix
mat <- as.matrix(dists)

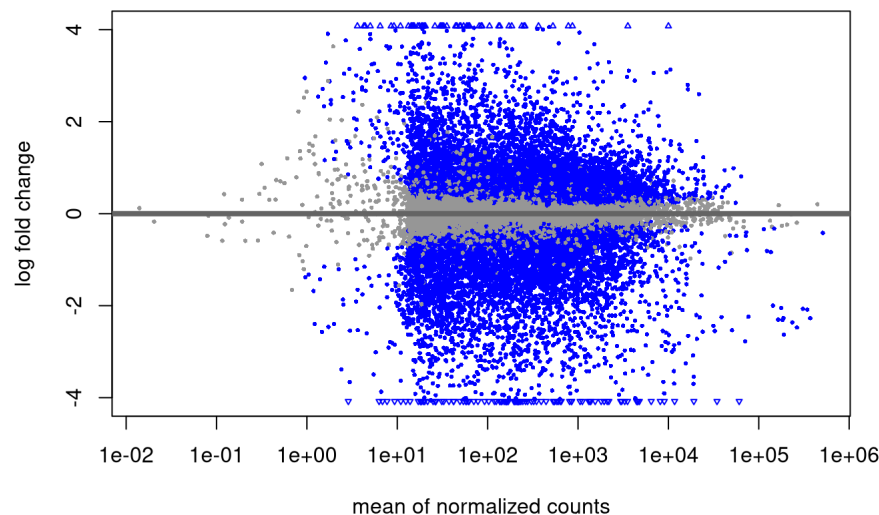
pheatmap(mat,
  clustering_distance_rows = dists,
  clustering_distance_cols = dists,
  show_rownames = TRUE,
  show_colnames = TRUE,
  fontsize_row = 4,
  fontsize_col = 4,
  angle_col = 45,
  main = "Sample-to-sample Distance Heatmap")
```



```
## 2 - Differential expression plots

# MA plot: log2 fold change versus mean expression
# shows the pattern of genes that are up- or down-regulated
plotMA(
  res,
  main = "MA plot: Cancer vs Paracancerous"
)
```

**MA plot: Cancer vs Paracancerous**



```
install.packages("BiocManager")
```

```
## Installing package into '/opt/app-root/src/Rpackages/4.3'
## (as 'lib' is unspecified)
```

```
BiocManager::install("EnhancedVolcano")
```

```
## 'getOption("repos")' replaces Bioconductor standard repositories, see
## 'help("repositories", package = "BiocManager")' for details.
## Replacement repositories:
##   CRAN: https://cran.rstudio.com/
```

```
## Bioconductor version 3.20 (BiocManager 1.30.27), R 4.4.3 (2025-02-28)
```

```
## Warning: package(s) not installed when version(s) same as or greater than current; use
## `force = TRUE` to re-install: 'EnhancedVolcano'
```

```
## Installation paths not writeable, unable to update packages
## path: /usr/lib64/R/library
## packages:
##   AER, aplot, arrow, bayesplot, bigD, BiocManager, BiocParallel, bookdown,
##   boot, BradleyTerry2, brglm, broom, Cairo, checkmate, cli, clock, cluster,
##   colorspace, commonmark, covr, cowplot, credentials, crosstalk, Cubist,
##   curl, data.table, dbplyr, dbscan, dendextend, DEoptimR, Deriv, devtools,
##   dials, diffobj, digest, doBy, doFuture, doRNG, DOSE, downlit, DT, dtplyr,
##   effects, emdbook, emmeans, entropy, Epi, epiR, etm, evaluate, extrafont,
##   extrafontdb, FactoMineR, fastcluster, fgsea, fitdistrplus, flextable,
##   foghorn, forcats, forecast, foreign, fs, future, future.apply,
##   future.callr, gam, gapminder, gargle, gclus, gcovbook, gdtools, generics,
##   gert, GGally, gganimate, ggforce, ggfortify, ggfun, ggnewscale, ggplot2,
##   ggplotify, ggpp, ggpubr, ggraph, ggridges, ggsci, ggstats, ggtangle,
##   ggvenn, gh, glmnet, globals, googledrive, googlesheets4, GPfit, gprofiler2,
##   gss, gt, hardhat, harmony, haven, here, Hmisc, HMM, hms, httpuv, httr2,
##   hunspell, igraph, insight, jsonlite, keras, KFAS, kinship2, knitr,
##   labelled, Lahman, later, lattice, lava, leidenbase, lgr, listenv, littler,
##   lme4, lobster, logger, magrittr, mapproj, maps, markdown, mathjaxr, Matrix,
##   MatrixModels, mclust, memisc, mgcv, mime, miniUI, mlr, mlr3, mlr3measures,
##   mlr3misc, mockery, mockr, modeltools, multcomp, MuMIn, nanotime, ncd4,
##   nlme, nloptr, officer, openssl, openxlsx, optimx, ottr, parallelly,
##   parsnip, partykit, patchwork, pbapply, pbdZMQ, pbkrtest, permute, pheatmap,
##   pillar, pixmap, pkgbuild, pkgdown, pkgload, plotly, plotrix, poppr, pracma,
##   pROC, prodlim, profmem, progressr, promises, PRROC, ps, purrr, qtl2,
##   quantmod, questionr, QuickJSR, R.cache, R.oo, ragg, raster, rbibutils,
##   Rcpp, RcppArmadillo, RcppDate, RcppParallel, RCurl, Rdpack, readr, recipes,
##   reformulas, rentrez, reshape, reshape2, restfulr, reticulate, rgl, rlang,
##   rmarkdown, rms, RMySQL, robustbase, roxygen2, RPostgreSQL, rprojroot,
##   rsample, RSQLite, rstantools, rstatix, Rttf2pt1, RUnit, rversions, rvest,
##   s2, S7, safetensors, sass, scales, scatterpie, sctransform, seriation,
##   Seurat, SeuratObject, sf, sfsmisc, shapr, shiny, slider, sparsevctr,
##   spatstat, spatstat.data, spatstat.explore, spatstat.geom, spatstat.linnet,
##   spatstat.model, spatstat.random, spatstat.univar, spatstat.utils, spelling,
##   spls, statmod, statnet.common, stringi, stringr, styler, survey, svglite,
##   systemfonts, tensor, tensorflow, terra, testthat, textshaping, tfruns,
##   TH.data, threejs, tibble, timeDate, tinytex, TSP, tune, tzdb, units,
##   usethis, utf8, uwot, V8, vcdExtra, vegan, visNetwork, vroom, waldo, warp,
##   workflows, WriteXLS, xfun, xgboost, XML, xml2, yulab.utils, zeallot, zip,
##   zoo
```

```
# Volcano plot
library(EnhancedVolcano)
```

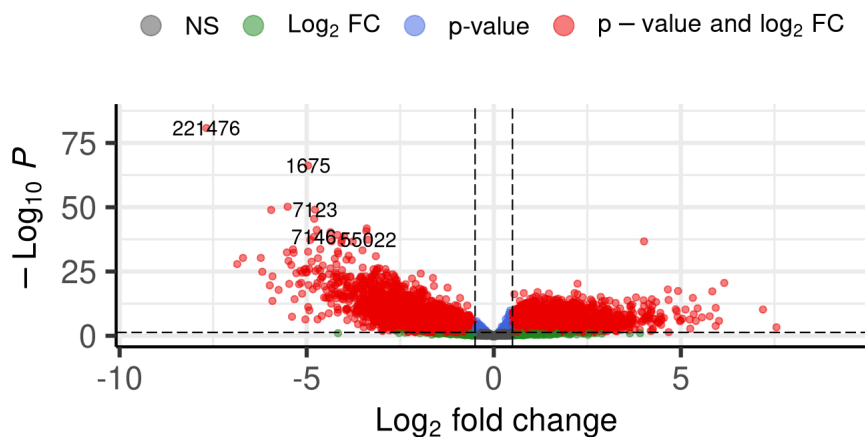
```
## Loading required package: ggrepel
```

```
# get the row names of the top 15 genes.
# these row names correspond to the gene IDs in the DESeq2 result object.
top15_genes <- rownames(top15_table)

# making a volcano plot, highlighting only the top 15 most significant genes
EnhancedVolcano(
  res,
  lab      = rownames(res),      # labels for all genes (by row name)
  x        = "log2FoldChange",   # x-axis: effect size (Cancer / Paracancerous)
  y        = "padj",             # y-axis: adjusted p-value
  pCutoff  = 0.05,               # significance threshold for padj
  FCcutoff = 0.5,                # threshold for |log2 fold change|
  selectLab = top15_genes,       # only these genes are labelled on the plot
  pointSize = 1.5,
  labSize  = 4,
  col      = c("grey30", "forestgreen", "royalblue", "red2"),
  title    = "Volcano Plot (Top 15 Most Significant Genes)",
  subtitle = "Top genes labelled based on lowest adjusted p-values"
)
```

## Volcano Plot (Top 15 Most Significant Genes)

Top genes labelled based on lowest adjusted p-values



```
# 3- Biological interpretation plots

library(RColorBrewer)

# Heatmap for Top 15 DEGs

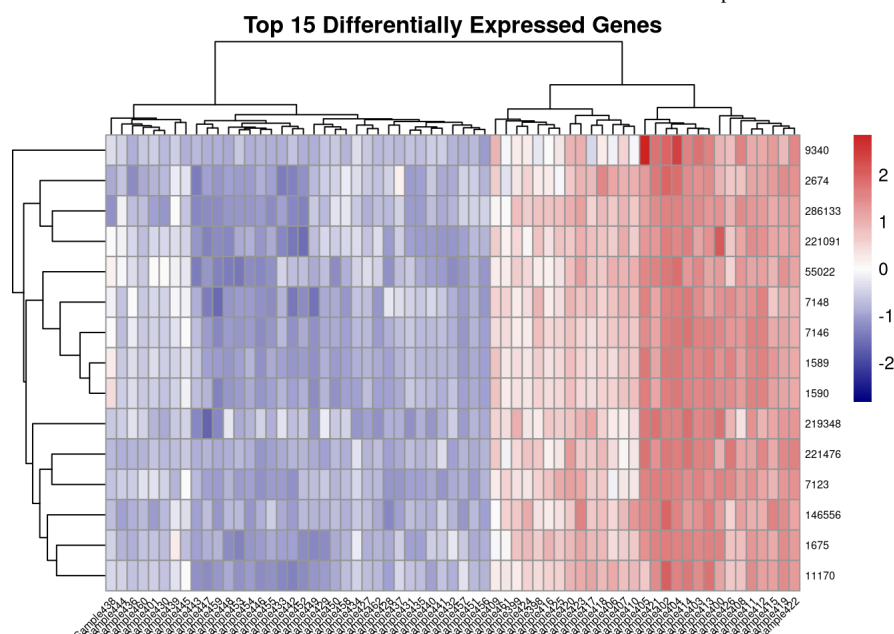
# extract expression matrix from VST
vsd_mat <- assay(vsd)

# taking the rownames of top15 genes
top15_genes <- rownames(top15_table)

# intersect to avoid missing genes (just in case)
top15_genes <- intersect(top15_genes, rownames(vsd_mat))

# subset the VST matrix
heatmap_top15 <- vsd_mat[top15_genes, ]

# plot heatmap
pheatmap(
  heatmap_top15,
  scale = "row",
  clustering_distance_rows = "euclidean",
  clustering_distance_cols = "euclidean",
  clustering_method = "complete",
  show_rownames = TRUE,
  show_colnames = TRUE,
  fontsize_row = 6,
  fontsize_col = 6,
  angle_col = 45,
  color = colorRampPalette(c("navy", "white", "firebrick3"))(100),
  main = "Top 15 Differentially Expressed Genes"
)
```



```
library(reshape2)
library(ggplot2)

# Choosing how many genes to show in the boxplots (we went with top 6)
top_genes <- rownames(top15_table)[1:6]

# subset the VST expression matrix to only these genes
expr_top <- vsd_mat[top_genes, ]

# convert the matrix to long format for ggplot2
df <- melt(expr_top)
colnames(df) <- c("Gene", "Sample", "Expression")

# add group (condition) information for each sample
# 'condition' is the column we created earlier in pdat / colData(dds)
df$condition <- colData(vsd)$condition[match(df$Sample, rownames(colData(vsd)))]

# making boxplots of expression per condition for each gene
ggplot(df, aes(x = condition, y = Expression, fill = condition)) +
  geom_boxplot(outlier.shape = NA, alpha = 0.8) +
  geom_jitter(width = 0.2, size = 1.3, alpha = 0.7) +
  facet_wrap(~ Gene, scales = "free_y", ncol = 3) +
  theme_bw(base_size = 12) +
  theme(
    strip.text = element_text(size = 12, face = "bold"),
    legend.position = "none"
  ) +
  ylab("VST normalized expression") +
  ggtitle("Expression Boxplots of Top Differentially Expressed Genes")
```

Expression Boxplots of Top Differentially Expressed Genes

