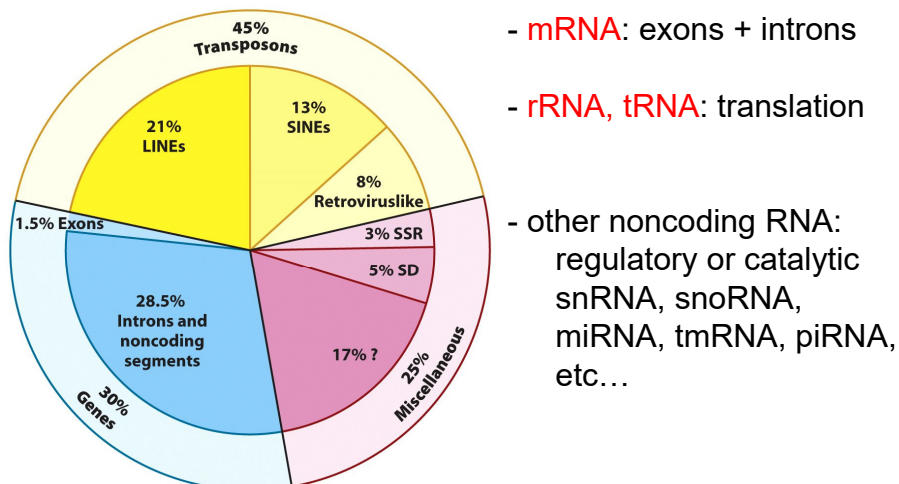


Chapter 26.1: RNA metabolism Part II

Transcription

- Common themes of RNA polymerases
- *E. coli* RNA polymerase and sigma factors
Step-wise control of RNA synthesis
- Eukaryotic RNA polymerases I, II and III
Many protein factors play a role

DNA-dependent synthesis of RNA



- **mRNA**: exons + introns
- **rRNA, tRNA**: translation
- other noncoding RNA:
regulatory or catalytic
snRNA, snoRNA,
miRNA, tmRNA, piRNA,
etc...

Figure 24-8
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RNA polymerases copy one strand of DNA

Conventions: **coding strand, RNA transcript**

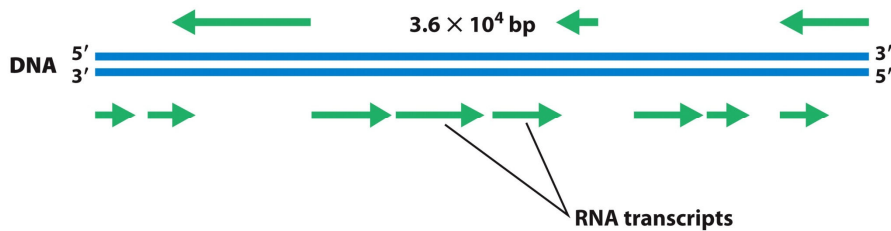
(5') **CGCTATAGCGTTT** (3') DNA nontemplate (coding) strand

(3') **GCGATATCGCAA** (5') DNA template strand

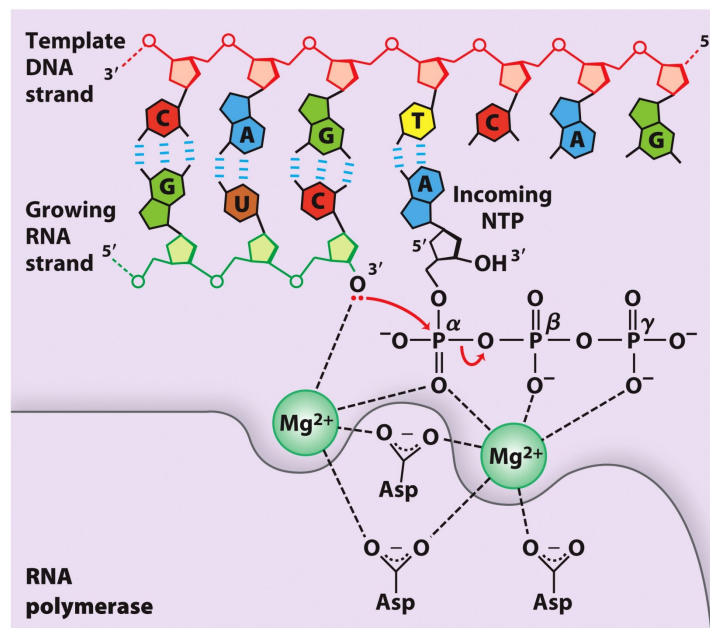
(5') **CGCUAUAGCGUUU** (3') RNA transcript

Figure 26-2
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RNA can be transcribed in either direction, often overlapping



Conserved two metal ion mechanism



Differences from DNA polymerases

- RNA polymerases can initiate with free NTP-
don't need primers
- Multiple transcripts but limited segment of DNA to
be copied
- Only one strand is copied at a time (don't have to
worry about semi-discontinuous synthesis)

E. coli RNA polymerase

- Core: 5 subunits ($\alpha_2\beta\beta'\omega$) (plus one σ)

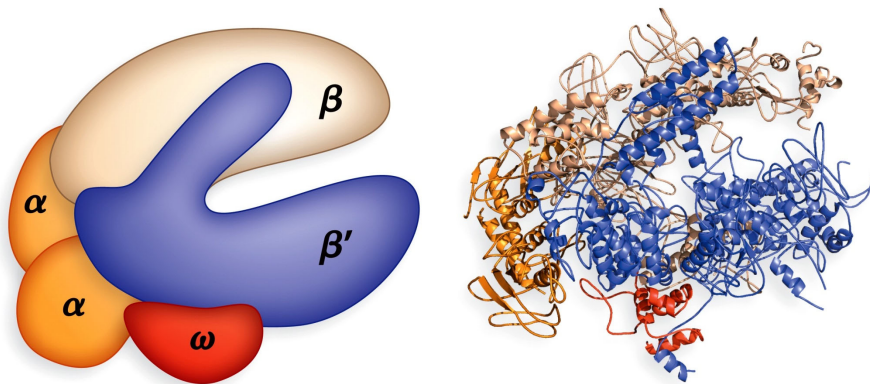


Figure 26-4
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Anatomy of the **Transcription bubble** in *E. coli*

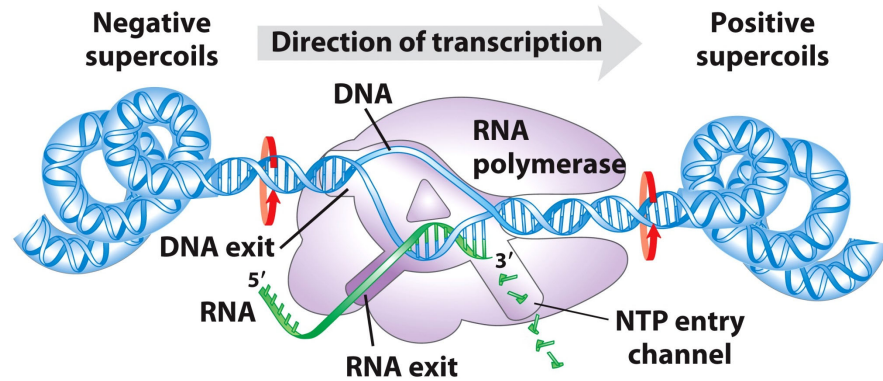
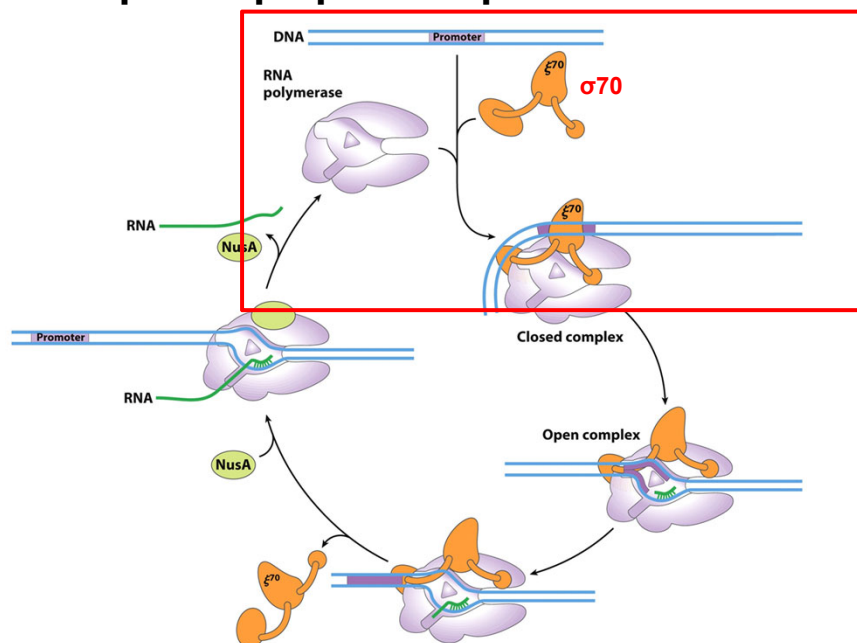


Figure 26-1c
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Multiple steps provide points to control



σ^{70} promoter sequences in *E. coli*

Numbering **convention**: transcription start site = +1

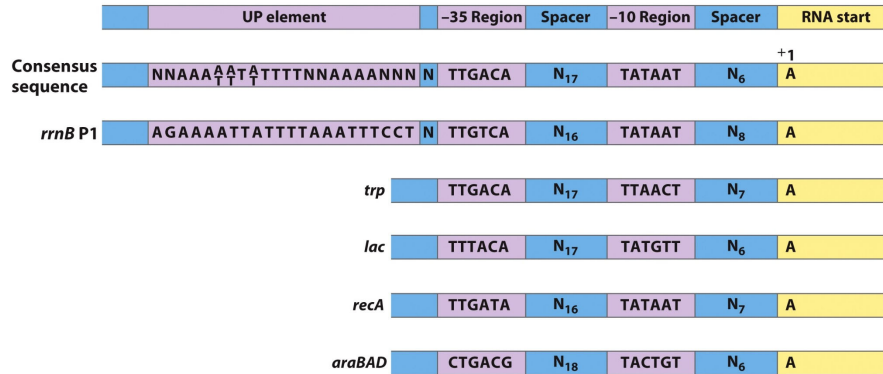


Figure 26-5
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Consensus: most preferred (ideal) sequence, but real promoters vary from this sequence

Sigma subunits recognize a variety of genes

TABLE 26-1 The Seven σ Subunits of *Escherichia coli*

σ subunit	K_d (nM)	Molecules/cell*	Holoenzyme ratio (%)*	Function
σ^{70}	0.26	700	78	Housekeeping
σ^{54}	0.30	110	8	Modulation of cellular nitrogen levels
σ^{38}	4.26	<1	0	Stationary phase genes
σ^{32}	1.24	<10	0	Heat shock genes
σ^{28}	0.74	370	14	Flagella and chemotaxis genes
σ^{24}	2.43	<10	0	Extracytoplasmic functions; some heat shock functions
σ^{18}	1.73	<1	0	Extracytoplasmic functions, including ferric citrate transport

Source: Adapted from Maeda, H., Fujita, N., & Ishihama, A. (2000) *Nucleic Acids Res.* 28, 3500.

Note: σ factors are widely distributed in bacteria; the number varies from a single σ factor in *Mycoplasma genitalium* to 63 distinct σ factors in *Streptomyces coelicolor*.

*Approximate number of each σ subunit per cell and the fraction of RNA polymerase holoenzyme complexed with each σ subunit during exponential growth. The numbers change as growth conditions change. The fraction of RNA polymerase complexed with each σ subunit reflects both the amount of the particular subunit and its affinity for the enzyme.

Table 26-1
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Identification of promoter sequences

Sequence analysis

- look for most preferred sequences (**consensus**)

Mutational analysis

- do mutations affect expression?

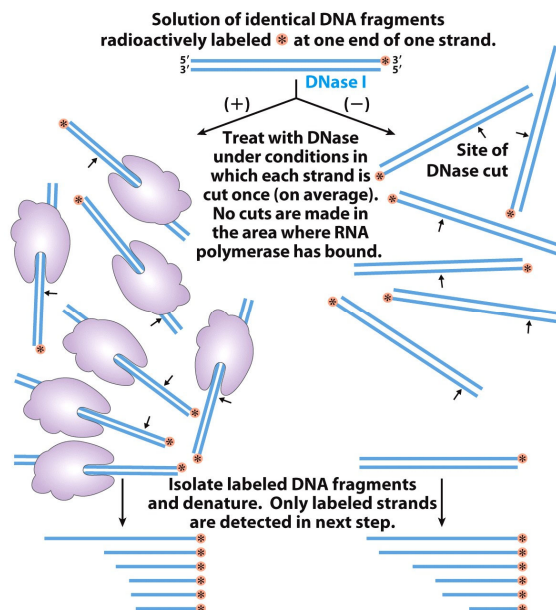
DNA binding and nuclease protection assays

- show that RNAP binds to sequence

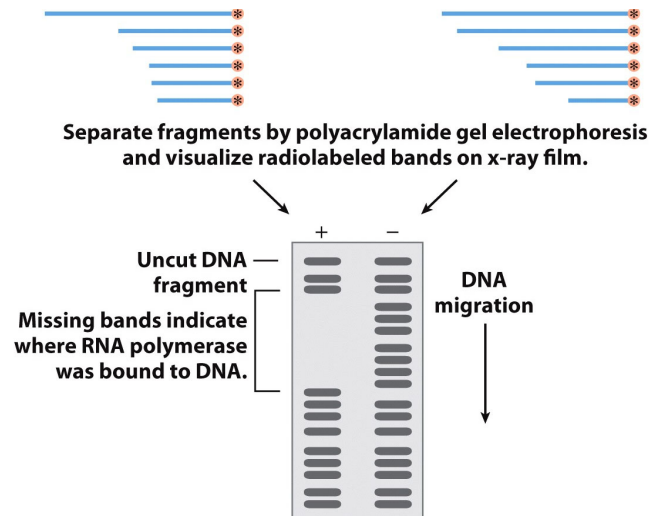
Reporter genes: functional in vivo assay

- has to be easy to detect and sensitive
- must be able to quantify (β -Gal, CAT)

Nuclease protection assay (DNA footprinting)



Nuclease protection assay (DNA footprinting)

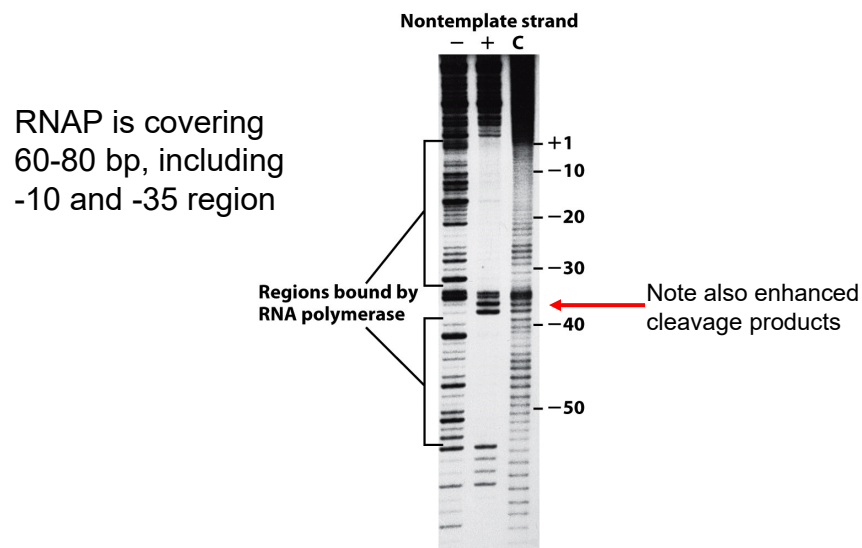


Box 26-1 Figure 1

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Nuclease protection assay (DNA footprinting)



Box 26-1 figure 2

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Central problem: specificity vs activity

How does RNAP exhibit 10^6 -fold specificity for promoter over non-promoter site and then leave?

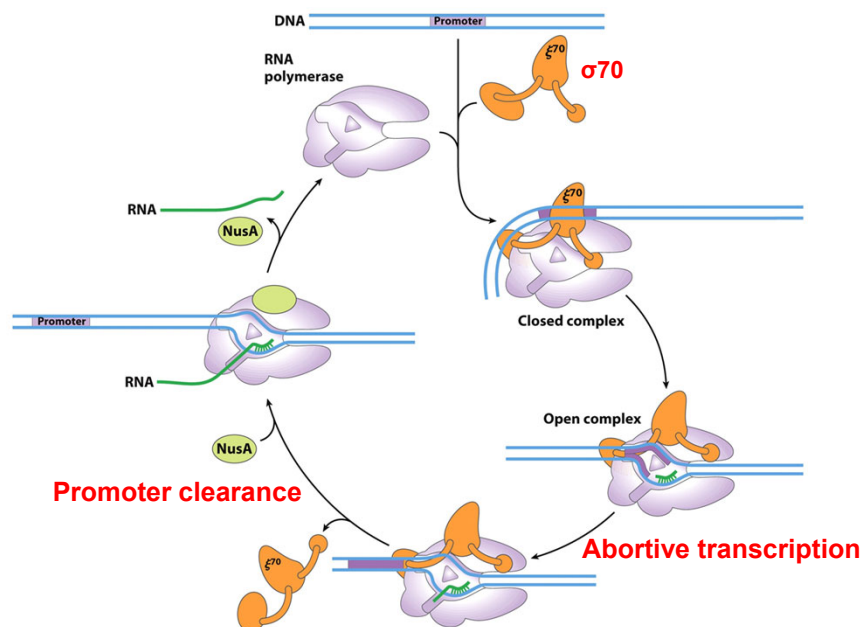
Answer: Steps

- Pol recognizes duplex DNA → **closed complex**
- Unwinds DNA → **open complex** (stable)
- Begins transcribing → **abortive transcripts**
- Transition from initiating pol to elongating pol → **promoter clearance**

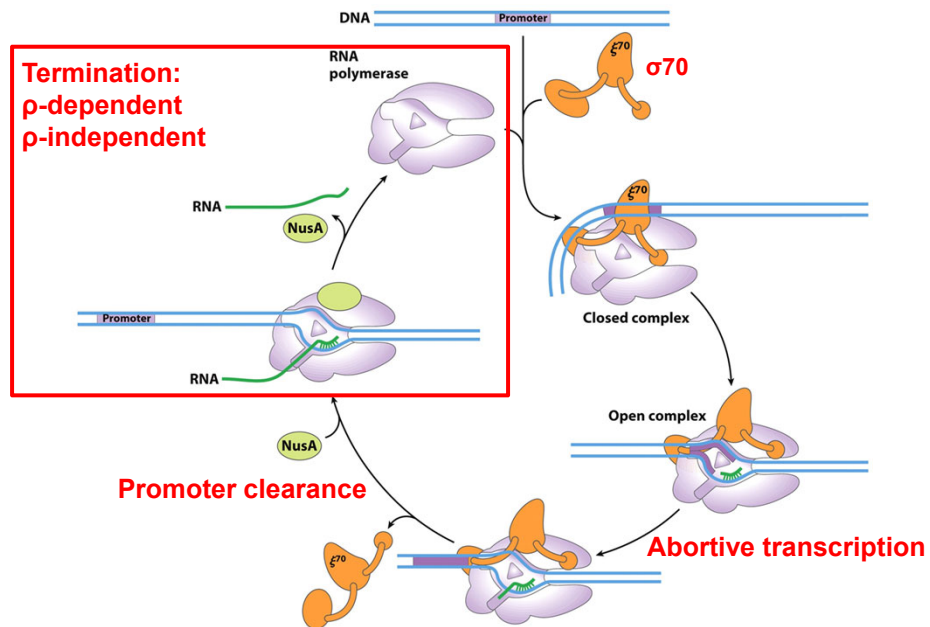
Elongating polymerase

- physical change in structure
- becomes non-specific

Multiple steps provide points to control



Multiple steps provide points to control



ρ -independent termination

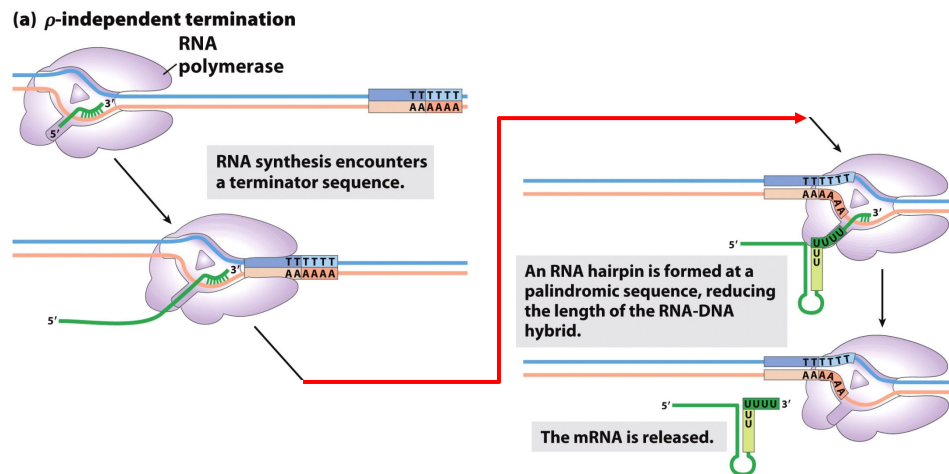


Figure 26-7
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Eukaryotes: three major RNA polymerases

Pol I: rRNA (18S, 5.8S and 28S precursor transcript)

Pol III: tRNA, 5S rRNA, other small RNAs

Pol II: mRNA, snRNA, miRNA

- Similarities to bacterial RNAP, but much more complicated DNA template (chromatin)
- Requires many additional protein factors (transcription factors)
- Still step-wise process: assembly, initiation, elongation, termination

Eukaryotic RNA Pol II promoters

Several types:

- **TATA-box, initiator (Inr)** (although not most Pol II)
- TATA-less, initiator
- snRNA

How does Pol II recognize a promoter?

- Transcription factors (not Pol subunit like σ) bind to DNA

TBP (TATA binding protein) plays a central role

- even at promoters that don't have a TATA box

TBP induces structural alteration in DNA

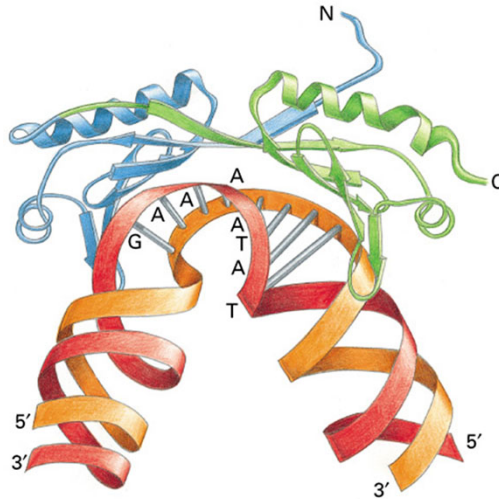
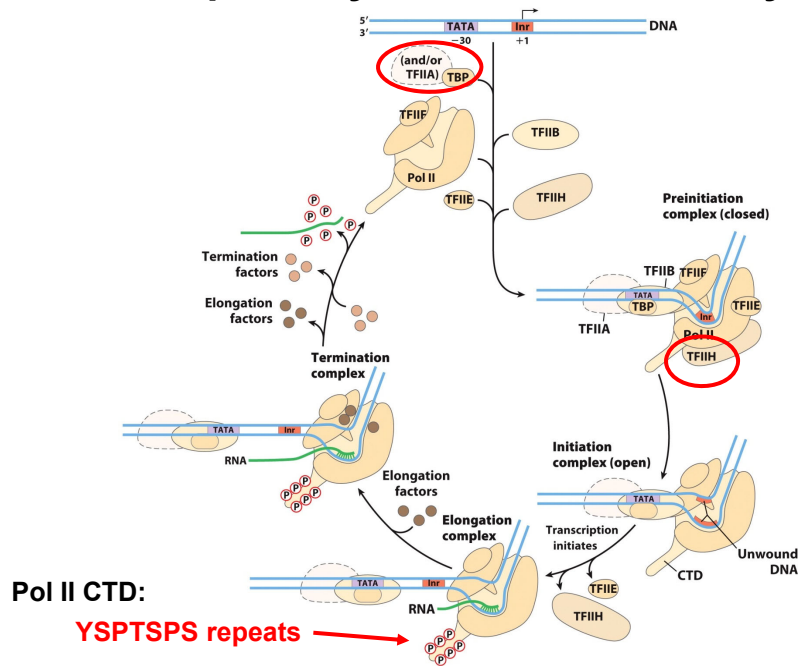


Figure 6-18. Molecular Biology of the Cell, 4th Edition.

Transcription by RNA Pol II in eukaryotes



Transcription by RNA Pol II in eukaryotes

TABLE 26-2 Proteins Required for Initiation of Transcription at the RNA Polymerase II (Pol II) Promoters of Eukaryotes

Transcription protein	Number of subunits	Subunit(s) M_r	Function(s)
Initiation			
Pol II	12	10,000–220,000	Catalyzes RNA synthesis
TBP (TATA-binding protein)	1	38,000	Specifically recognizes the TATA box
TFIIA	3	12,000, 19,000, 35,000	Stabilizes binding of TFIIB and TBP to the promoter
TFIIB	1	35,000	Binds to TBP; recruits Pol II–TFIIF complex
TFIIE	2	34,000, 57,000	Recruits TFIIH; has ATPase and helicase activities
TFIIF	2	30,000, 74,000	Binds tightly to Pol II; binds to TFIIB and prevents binding of Pol II to nonspecific DNA sequences
TFIIH	12	35,000–89,000	Unwinds DNA at promoter (helicase activity); phosphorylates Pol II (within the CTD); recruits nucleotide-excision repair proteins
Elongation*			
ELL [†]	1	80,000	Phosphorylates Pol II (within the CTD)
pTEFb	2	43,000, 124,000	
SII (TFIIS)	1	38,000	
Elongin (SIII)	3	15,000, 18,000, 110,000	

*The function of all elongation factors is to suppress the pausing or arrest of transcription by the Pol II–TFIIF complex.

[†]Name derived from eleven-nineteen lysine-rich leukemia. The gene for ELL is the site of chromosomal recombination events frequently associated with acute myeloid leukemia.

Table 26-2

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