

IMPORTANCE OF OPTIMUM WATER QUALITY INDICES IN SUCCESSFUL ORNAMENTAL FISH CULTURE PRACTICES

Dr. S.T. Gopu Kumar, Sreeya Nair


Parishodh Journal

Cite this paper

Downloaded from [Academia.edu](#) 

[Get the citation in MLA, APA, or Chicago styles](#)

Related papers

[Download a PDF Pack](#) of the best related papers 



[Stress-associated impacts of short-term holding on fishes](#)

Donald Portz

[Biological Approaches in Management of Nitrogenous Compounds in Aquaculture Systems](#)

Global Science Books

[Acute Toxicity and Sublethal Effects of Nitrite on Selected Hematological Parameters and Tissues in...](#)

Young-Chae Song

IMPORTANCE OF OPTIMUM WATER QUALITY INDICES IN SUCCESSFUL ORNAMENTAL FISH CULTURE PRACTICES

Sreeya G.Nair^{1*}, V. Vidhya² and S.T. Gopukumar³

¹Department of Zoology, Sree Ayyappa College for Women, Chunkankadai, Tamil Nadu, India.

²Department of Zoology, St.Jude's College, Thoothoor, TamilNadu, India.

³Department of Medical Research, Sarada Krishna Homoeopathic Medical College, Kulasekharam, TN, India.

ABSTRACT

Ornamental fish keeping is the second most preferred hobby in the world and the number of hobbyists for ornamental fish keeping is rising day by day because it provides a great opportunity for entrepreneurship development and income generation. Aquatic ecosystems are dynamic and even in small rearing water tanks, physical and chemical parameters are interrelated. Thus, physical and chemical parameters of water should be considered and analyzed together because all of these factors have a direct impact on the culture systems. Ornamental fish production unit required higher level of expertise for better water quality management as ornamental fishes are more sensitive to poor water quality. As ornamental fish are kept in tanks more numbers than their food fish counterparts, water quality is most critical. Where large numbers of fish are kept in small spaces, the build-up of nitrogenous wastes, most notably ammonia, requires the producer to implement measures to manage it properly. Regular water exchange along with proper aeration overcomes this type of problem in the tanks. This review discussed the relationship between the health and better rearing conditions of fishes.

KEYWORDS: Disease resistance, Fishes, Health, Ornamental, Parameters, Water Quality.

Introduction

Ornamental fish culture ensures socially equitable distribution of benefits along the value chain as this activity is time bound, labour intensive and livelihood options of vulnerable sections of the society like house-wives and unemployed youth (Ghosh *et al.*, 2003). In India, the practice of ornamental fish keeping started in 1951 with the opening of the Taraporevale Aquarium, 'the pride of the nation' at Mumbai and the establishment of several aquarium societies in the city. Since then the practice has become widespread in India, with more than hundred varieties of indigenous species and even more of exotic ones. The trade in live aquatic ornamental animals for the aquarium trade is a global multi-million dollar industry, which can provide economic incentives for habitat conservation. The breeding and culture of freshwater ornamental fishes could generate great employment in almost all parts of India and develop into a local trade. This is an area for employment of women and school dropouts who would find it convenient to indulge in these exercises in their spare time. With increasing supply of ornamental fishes, the demand for aquaria would also go up and production of aquarium tanks could be developed as a small scale entrepreneurship or cottage industry which will be an additional source of employment (Sreeya, 2012).

Aquarium fish often live in suboptimal conditions involving limited volumes of water in aquarium systems with a restricted capacity to maintain adequate water quality. Unlike feral fish, they cannot escape a potentially harmful environment. Even the best-equipped aquarium, combined with rigorous surveillance of water quality parameters, can never truly mimic natural conditions in the wild. Thus, the keeping of fish in aquaria is a compromise that usually has a negative influence on the fish's well-being. In this respect, Snieszko (1981) developed the frequently quoted graphic of three overlapping circles representing the host (i.e., the fish), the pathogens and the environment to show the single and combined influences of infectious agents and non-infectious parameters on fish health. Therefore, only multidisciplinary studies involving the characteristics of potential pathogenic microorganisms for fish, aspects of the biology of the fish hosts as well as a better understanding of the environmental factors affecting such cultures, will allow the application of adequate measures to prevent and control the major diseases limiting the production of fishes.

Water Quality and Fish Diseases

The success of ornamental fish culture depends on the health status of the candidate species (Lipton, 2006). Being aquatic, and secondarily being forced to remain under crowded

conditions, the ornamental fishes are subjected to different diseases of varying nature (Bright and Sreedharan, 2009). Bacterial infections are considered as the major cause for diseases and mortality (Grisez and Ollevier, 1995). A complete understanding of the aetiological agent, the pathogenesis, antigenicity, epizootiology and the inter-relationship of stress-related and environmental factors is essential for successful management and control. The water easily spread most of the pathogens. It is necessary to have an understanding of water quality in order to successfully diagnose and correct aquarium diseases. Stress has been linked as the primary contributing factor of fish disease and mortality in aquaculture (Petric *et al.*, 2006).

Temperature

Several biotic and abiotic factors influence the growth of fish (Jobling, 1996). Among the various physical factors affecting the aquatic environment, temperature is of paramount importance and is considered as the “abiotic master factor” for fishes (Brett, 1971). Global climate change is suggested to potentially affect freshwater fisheries by lowering productivity in wild fish populations and in intensive aquaculture systems worldwide (Ficke *et al.*, 2007). As fishes are poikilotherms, drastic change in their surrounding water temperature will influence their metabolic processes, behaviour, migration, growth, reproduction, and survival (Fry, 1971; Portner, 2001). Researchers are making continuous efforts to define thermal tolerance of various fish species of aquaculture importance (Rishikesh *et al.*, 2009). Long-term changes in the environmental temperature induce ectothermic animals to display compensatory responses (which include changes in the metabolic enzymes and tissue chemistry) that are suggested to mitigate the effect of temperature on metabolism (Hazel and Prosser, 1974; Hochachka and Somero, 1971).

Temperature beyond the optimum limits of a particular species, however, adversely affects fish health by increasing metabolic rate and subsequent oxygen demand, invasiveness and virulence of bacteria and other pathogens which in turn may cause a variety of pathophysiological disturbances in the host (Wedemeyer *et al.*, 1999). Temperature affects virtually all biochemical, physiological activities of fishes. The survival and growth of poikilothermal teleosts are immediately influenced by temperature fluctuations in their environments. All teleost species have developed their own specific adaptive mechanism, both behavioural and physiological, to cope up with temperature fluctuations (Prosser and Heath, 1991). These adaptive capabilities enable them to survive through acclimation and adaptation to stressful temperature conditions (Hazel and Prosser, 1974). Identifying the range of temperatures tolerated by a species is important to determine the viability of its

growth. Up to a species-specific maximum, fish growth rates will accelerate with increasing temperature, after which they sharply decline (Fielder *et al.*, 2005; Jobling, 1996).

Stocking Density

Crowding is a factor involved in physiological stress (Barton, 2002). The increase in stocking densities can alter the immunological responses and physiological processes, mainly those related to metabolism and behavior (Vijayan *et al.*, 1990; Irwin *et al.*, 1999; Barcellos *et al.*, 2004; Kristiansen *et al.*, 2004 and Schram *et al.*, 2006). It has been noticed that inappropriate stocking densities can alter lipid metabolism, mainly of triglycerides, in brook charr, *Salvelinus fontinalis* (Vijayan *et al.*, 1990). In gilthead sea bream, *Sparus aurata*, different stocking densities altered fatty acid metabolism, with a decrease in hepatic oleic acid, a monounsaturated fatty acid important as energy source, mainly in higher stocking densities (Montero *et al.*, 1999). Also, crowding is responsible for the increase in plasma cortisol, which plays an important role in the low efficiency of immunological responses under these conditions (Mommsen *et al.*, 1999; Di Marco *et al.*, 2008).

Water pH

Water pH affects metabolism and physiology of fish. Alkaline pH 7 to 8 is highly suitable for better growth of fish. Robert and William (1986) found that in channel catfish, excretion of ammonia at pH 6 increased; whereas, it decreased with increased pH. Saha *et al.* (2002) and Scott *et al.*, (2005) indicated that ammonia excretion increased with increasing pH (alkalinity), while growth decreased.

Total Alkalinity

Total Alkalinity is the measure of the capacity of water to neutralize or buffer acids using carbonate, bicarbonate ions, and in rare cases, by hydroxide, thus protecting the organisms from major fluctuations in pH. Without a buffering system, free carbon dioxide will form large amounts of a weak acid (carbonic acid) that may potentially decrease the night-time pH level to 4.5. During peak periods of photosynthesis, most of the free carbon dioxide will be consumed by the phytoplankton and, as a result, drive the pH levels above 10.0.

Dissolved Oxygen

Dissolved oxygen is the most important and critical parameter, requiring continuous monitoring in ornamental fish culture systems. This is due to the fact that fish aerobic metabolism requires dissolved oxygen (Timmons *et al.*, 2001). Optimum level of dissolved oxygen recommended is 4 to 5 mg/l for warm water fishes (Wedemeyer and Goodyear, 1984). Studies of freshwater teleosts indicated that low dissolved oxygen concentrations also

can modify juvenile and adult growth rates, feeding rates, habitat use and susceptibility to predation, as well as adult reproductive activities (Magnuson *et al.*, 1985; Suthers and Gee, 1986; US-EPA, 1986; Kramer, 1987; Poulin *et al.*, 1987; Saint-Paul and Soares, 1987).

The physiological activities of fish are also subjected to changes in environmental factors such as dissolved O₂ level (Jordan and Steffensen, 2007;The physiological activities of fish are also subjected to changes in environmental factors such as dissolved O₂ level (Jordan and Steffensen, 2007;Petersen and Gamperl, 2010). A recent study found that environmental temperature had profound effects on the metabolic competition mode of southern catfish (*Silurus meridionalis* Chen), possibly due to the increased oxygen demand and decreased availability of environmental dissolved oxygen at high temperatures (Pang *et al.*, 2010). Conformers, such as the Adriatic sturgeon (*Acipenser naccarii*), cannot maintain their resting O₂ consumption rate during hypoxia, and it will decrease linearly with decreasing dissolved O₂ content (Mc Kenzie *et al.*, 2007).

Hypoxia can cause physiological stress and cellular damage as well as inhibit repair mechanisms (Jones, 1985). Spot and pinfish can detect a variety of DO concentrations but they do not necessarily avoid hypoxia (Wannamaker and Rice, 2000). Wannamaker and Rice (2000) suggested that these fishes may have relatively lower physiological costs when occupying hypoxic areas. Shultz *et al.*, (2011) recommended that the dissolved O₂ concentrations during holding of bonefish in the context of live-release angling tournaments do not deviate from that of ambient sea water, which was typically 6 mg/l. Similar studies on live-release bass tournaments have recommended to the anglers and tournament organizers to monitor DO concentrations so as to maintain their required levels of dissolved O₂ for recovery (Suski *et al.*, 2006; Furimsky *et al.*, 2003; Suski *et al.*, 2003).

Ammonia

Ammonia is the principal nitrogenous waste product of fishes that represents 60% to 80% of nitrogenous excretion of fish (Handy and Poxton, 1993; Salin and Williot, 1991). It is also, the main nitrogenous waste material excreted by gills beside urea and amines and an end product of the protein catabolism (De Croux *et al.*, 2004). Among all the water quality parameters, which affect fish, ammonia is considered as one of the most important after oxygen (Francis – Floyd and Watson, 1996). Under intensive rearing conditions, and particularly when effluent is reused, ammonia concentrations may reach levels that limit fish survival and growth (Haywood, 1983). Ammonia can cause reductions in growth or even death (EPA, 1998; Meade, 1985; Salin, and Williot, 1991).

In water, total ammonia consists of non toxic (ionized ammonia) referred to as ammonium (NH_4^+) and toxic un-ionized ammonia (NH_3). The equilibrium between these two forms is dependant on the pH and temperatures. Ammonia is measured as total ammonia nitrogen (TAN) which represents the sum of NH_4^+ and NH_3 . The NH_3 molecule is soluble in lipids which is 300 to 400 times more toxic than NH_4^+ (Haywood, 1983; Thurston *et al.*, 1981). Un-ionized ammonia (UIAN) can readily diffuse across the gill membranes due to its lipid solubility and lack of charge (Aysel and Koksall, 2005). When ammonia accumulates to toxic levels, fish cannot extract energy from feed and will fall into a coma and die (Hargreaves and Tucker, 2004).

Ammonia tends to block oxygen transfer from the gills to the blood and can cause both immediate and long term gill damage (Joel and Amajuoyi, 2010). Also it can cause impairment of cerebral energy metabolism, damage to gill, liver, kidney, spleen and thyroid tissue in fish, crustaceans and mollusks (Smart, 1978). Chronic un-ionized ammonia exposure may affect fish and other organisms in several ways, e.g. gill hyperplasia, muscle depolarization, hyper excitability, convulsions and finally death (Ip *et al.*, 2001). Toxicity of ammonia to fish has been intensively investigated in numerous fish species (Aysel and Koksall, 2005; El-Shafai *et al.*, 2004; Lamarie *et al.*, 2004). The acute and chronic toxicities of ammonia have been reviewed for fresh water species (Tomasso, 1994; Handy and Poxton, 1993; Russo and Thurston, 1991; Haywood, 1983; Ruffier *et al.*, 1981). Uncontrolled level of ammonia in culture environment may not only lead to mortality but may prevent the fish from achieving its full genetic potential in terms of growth and reproductive capability. At the sub-lethal value of ammonia, it can compromise the well being of fish by jeopardizing its health (Ajani, 2008).

Smith and Piper (1975) and Smart (1976) found that the most characteristic feature for chronic exposure of rainbow trout to ammonia was the appearance of swollen, rounded secondary gill lamellae or telangiectatic capillaries in the secondary lamellae. Also, Kirk and Lewis (1993) reported that the gills of rainbow trout exposed to 0.1 mg/l ammonia for 2 h exhibited deformation of the lamellae. Ammonia concentrations of above 0.2 mg/l in fish ponds have a tendency to harm the fishes and is recommended that the UIA-N concentrations be maintained below 0.1 mg/l (Abdalla and Heba Allah, 2011).

Ammonia induces detrimental changes in tissue structure, cell function, blood chemistry, osmoregulation, disease resistance, growth and reproductive capacity (Jeney *et al.*, 1992). Chronic exposure can result in the deterioration of several physiological functions any one of which may be the ultimate cause of death (Russo, 1985). Ammonia may

affect gill structure (Smart, 1976), respiratory function (Chen and Lin, 1992; Knoph, 1996) and oxygen consumption (Smart, 1978) in aquatic animals. Keeping animals healthy in intensive aquaculture depends on preventing accumulation of toxic waste products such as ammonia. Ammonia concentration of 1–2mM is common in the blood of marine invertebrates (Haberfield *et al.*, 1975). Higher concentrations are presumably toxic because they perturb acid–base balance with too much alkalinity (Hammen, 1980).

Nitrite

Nitrite (NO_2^-) is a potential contaminant in aquatic environments that receives nitrogenous waste (Grosell and Jensen, 1999). Nitrite is formed from ammonia and may accumulate in aquatic systems as a result of imbalance of nitrifying bacterial activity (*Nitrosomonas sp.* and *Nitrobacter sp.*) (Masser *et al.*, 1999). High levels of nitrite in the water is a potential factor triggering stress and cause high mortality in aquatic organisms (Ferreira da Costa *et al.*, 2004; Wang *et al.*, 2004; Jensen, 2003; Martinez and Souza, 2002). Several studies have examined the toxicity and physiological effects of NO_2^- in fish (Das *et al.*, 2004; Knudsen and Jensen, 1997; Doblander and Lackner, 1996). When NO_2^- reaches the blood, it crosses the erythrocyte membrane and oxidizes haemoglobin to methaemoglobin.

The toxicity of nitrite may result from a combination of effects rather than from a simple effect, such as methaemoglobinaemia, in particular. An elevated ambient nitrite concentration is problematic for freshwater fish, as nitrite is actively taken up across the gills in competition with chloride (Eddy and Williams, 1987). The principal effect of such nitrite loading is a progressive oxidation of haemoglobin to methaemoglobin, but several other physiological changes occur (Jensen, 1990).

The interference with branchial ion exchange together with methaemoglobinemia and likely tissue hypoxia suggests that major changes may arise in blood O_2 transport and respiratory properties resulting in perturbations of electrolyte and acid–base status (Jensen *et al.*, 1987). Knudsen and Jensen (1997) showed that nitrite interferes with K^+ homeostasis in carp leading to an extracellular hyperkalaemia. Aggergaard and Jensen (2001) have shown among rainbow trout exposed to nitrite increased in plasma K^+ with a concomitant decrease in plasma Cl^- . This rise in plasma K^+ is suggested due to the release of K^+ from intracellular compartments (Knudsen and Jensen, 1997). Nitrite binds competitively to haemoglobin oxidising it to meet Hb, a variant causing the blood to appear brown in colour (hence the

name “brown blood disease”) and vastly reduce the ability to bind and transport oxygen (Jensen, 2003; Martinez and Souza, 2002; Hargreaves, 1998).

Nitrate

Acute and chronic effects of nitrate have been reported in several fresh waterfish species (Hamlin, 2006; Camargo *et al.*, 2005) and marine invertebrates (Kuhn *et al.*, 2010; Romano and Zeng, 2007; Camargo *et al.*, 2005; Hirayama, 1974). The mechanisms of nitrate toxicity to aquatic animals are due mainly to methemoglobinemia, caused by the oxidation of hemoglobin (Hb) to methemoglobin (MetHb) in blood (Camargo *et al.*, 2005) consequently reducing oxygen binding capacity and ultimately resulting in respiro-circulatory constraints.

In fish a MetHb reductase system compensates for MetHb formation by conversion of MetHb to Hb (Freeman *et al.*, 1983). Nitrate is taken up via the branchial system in fishes. However due to the low permeability of the gills to nitrate, uptake is limited and other mechanisms are suggested (Stormer *et al.*, 1996). Another possible pathway might be transdermal uptake of nitrate in the gastro intestinal tract, as was reported for nitrite in European flounder, *Platichthys flesus* (Grosell and Jensen, 2000).

CONCLUSION

Water quality affects growth and well-being of fish, therefore, water quality should be of great importance to the aquaculturist. The disease management in ornamental fish sector requires focus on preventive measures related to water quality and other husbandry activation. It is equally important to know how to interpret the water quality parameters that are measured to maintain the health and well-being of their fish stock. Exploration and implementation of prophylactic measures in aquaria are essential to prevent induction and spreading of disease and minimize the loss of valuable fish stock.

REFERENCES

- Grisez L, Ollevier F. *Vibrio* (Listonella) anguillarum infection in marine fish larviculture. *Larvi*. 1995;91:497.
- El-Shebly AA, Gad HA. Effect of chronic ammonia exposure on growth performance, serum growth hormone (GH) levels and gill histology of Nile tilapia (*Oreochromis niloticus*). *J Microbiol Biotechnol Res*. 2011;1(4):183-97.
- Ajani F, Olukunle OA, Agbede SA. Hormonal and haematological responses of *Clarias*

gariepinus (Burchell 1822) to nitrite toxicity.

Benli AÇ, Köksal G. The acute toxicity of ammonia on tilapia (*Oreochromis niloticus* L.) larvae and fingerlings. Turkish Journal of Veterinary and Animal Sciences. 2005 May 10;29(2):339-44.

Barcellos LJ, Kreutz LC, Souza C, Rodriguez LB, Fioreze I, Quevedo RM, Cericato L, Soso AB, Fagundes M, Conrad J, Lacerda LA. Haematological changes in Jundia (*Rhamdia quelen*) after acute and chronic stress caused by usual aquacultural management, with emphasis on immunosuppressive effects. Aquaculture. 2004;237:229-36.

Barcellos LJ, Kreutz LC, Souza C, Rodriguez LB, Fioreze I, Quevedo RM, Cericato L, Soso AB, Fagundes M, Conrad J, Lacerda LA. Haematological changes in Jundia (*Rhamdia quelen*) after acute and chronic stress caused by usual aquacultural management, with emphasis on immunosuppressive effects. Aquaculture. 2004;237:229-36.

Barton BA. Stress in fishes: a diversity of responses with particular reference to changes in circulating corticosteroids. Integrative and comparative biology. 2002 Jul 1;42(3):517-25.

Brett JR. Growth responses of young sockeye salmon (*Oncorhynchus nerka*) to different diets and planes of nutrition. Journal of the Fisheries Board of Canada. 1971 Oct 1;28(10):1635-43.

Bright Singh, I. S and Sreedharan, K. Ornamental fish diseases and their management measures. CMFRI - Winter School Course Manual on "Recent Advances in Breeding and Larviculture of Marine Finfish and Shellfish. 2009. 244-253.

Camargo JA, Alonso A, Salamanca A. Nitrate toxicity to aquatic animals: a review with new data for freshwater invertebrates. Chemosphere. 2005 Mar 1;58(9):1255-67.

Chen JC, Lin CY. Oxygen consumption and ammonia-N excretion of *Penaeus chinensis* juveniles exposed to ambient ammonia at different salinity levels. Comparative biochemistry and physiology. C, Comparative pharmacology and toxicology. 1992 Jun;102(2):287-91.

Das T, Pal AK, Chakraborty SK, Manush SM, Chatterjee N, Mukherjee SC. Thermal tolerance and oxygen consumption of Indian Major Carps acclimated to four temperatures. Journal of Thermal Biology. 2004 Apr 1;29(3):157-63.

Julieta M, Loteste A. Lethal effects of elevated pH and ammonia on juveniles of

- neotropical fish *Colosoma macropomum* (Pisces, Caracidae). *Journal of environmental biology*. 2004 Jan;25(1):7-10.
- Di Marco P, Priori A, Finoia MG, Massari A, Mandich A, Marino G. Physiological responses of European sea bass *Dicentrarchus labrax* to different stocking densities and acute stress challenge. *Aquaculture*. 2008 Mar 31;275(1-4):319-28.
- Doblender C, Lackner R. Metabolism and detoxification of nitrite by trout hepatocytes. *Biochimica et Biophysica Acta (BBA)-General Subjects*. 1996 Mar 15;1289(2):270-4.
- Eddy FB, Williams EM. Nitrite and freshwater fish. *Chemistry and Ecology*. 1987 Aug 1;3(1):1-38.
- Gijzen H. Chronic ammonia toxicity to duckweed-fed tilapia. *Aquaculture*. 2004; 232: 117- 127.
- Delos C, Erickson R. Update of ambient water quality criteria for ammonia. EPA/822/R-99/014. Final/technical Report. 1999.
- da Costa OT, dos Santos Ferreira DJ, Mendonça FL, Fernandes MN. Susceptibility of the Amazonian fish, *Colossoma macropomum* (Serrasalminae), to short-term exposure to nitrite. *Aquaculture*. 2004 Apr 5;232(1-4):627-36.
- Ficke AD, Myrick CA, Hansen LJ. Potential impacts of global climate change on freshwater fisheries. *Reviews in Fish Biology and Fisheries*. 2007 Nov 1;17(4):581-613.
- Fielder DS, Bardsley WJ, Allan GL, Pankhurst PM. The effects of salinity and temperature on growth and survival of Australian snapper, *Pagrus auratus* larvae. *Aquaculture*. 2005 Nov 14;250(1-2):201-14.
- Freeman L, Beitinger TL, Huey DW. Methemoglobin reductase activity in phylogenetically diverse piscine species. *Comparative Biochemistry and Physiology Part B: Comparative Biochemistry*. 1983 Jan 1;75(1):27-30.
- Fry FE. The effect of environmental factors on the physiology of fish. *Fish physiology*. 1971:1-98.
- Furimsky M, Cooke SJ, Suski CD, Wang Y, Tufts BL. Respiratory and circulatory responses to hypoxia in Largemouth Bass and Smallmouth Bass: implications for “live-release” angling tournaments. *Transactions of the American Fisheries Society*. 2003 Nov;132(6):1065-75.
- Ghosh A, Mahapatra BK, Datta NC. Ornamental fish farming-successful small scale aqua business in India. *Aquaculture Asia*. 2003 Jul;8(3):14-6.

- Grosell M, Jensen FB. Uptake and effects of nitrite in the marine teleost fish *Platichthys flesus*. *Aquatic Toxicology*. 2000 Aug 1;50(1-2):97-107.
- Grosell M, Jensen FB. NO₂-uptake and HCO₃-excretion in the intestine of the European flounder (*Platichthys flesus*). *Journal of Experimental Biology*. 1999 Aug 1;202(15):2103-10.
- Haberfield EC, Haas LW, Hammen CS. Early ammonia release by a polychaete *Nereis virens* and a crab *Carcinus maenas* in diluted sea water. *Comparative Biochemistry and Physiology Part A: Physiology*. 1975 Jan 1;52(3):501-3.
- Hamlin HJ. Nitrate toxicity in Siberian sturgeon (*Acipenser baeri*). *Aquaculture*. 2006 Mar 31;253(1-4):688-93.
- Hammen CS. *Marine invertebrates: comparative physiology*. University Press of New England; 1980. 127 pp.
- Handy RD, Poxton MG. Nitrogen pollution in mariculture: toxicity and excretion of nitrogenous compounds by marine fish. *Reviews in Fish Biology and Fisheries*. 1993 Sep 1;3(3):205-41.
- Hargreaves JA, Tucker CS. *Managing ammonia in fish ponds*. Stoneville, MS: Southern Regional Aquaculture Center; 2004 Dec.
- Hargreaves JA. Nitrogen biogeochemistry of aquaculture ponds. *Aquaculture*. 1998 Jul 15;166(3-4):181-212.
- Haywood GP. Ammonia toxicity in teleost fishes: a review. *Can. Tech. Rep. Fish. Aquat. Sci.*. 1983;1177:1-35.
- Hazel JR, Prosser CL. Molecular mechanisms of temperature compensation in poikilotherms. *Physiological reviews*. 1974 Jul;54(3):620-77.
- Hochachka PW, Somero GN. *Biochemical adaptation to the environment*. *American Zoologist*. 1971 Feb 1;11(1):159-67.
- Ip YK, Chew SF, Randall DJ. Ammonia toxicity, tolerance, and excretion. *Fish physiology*. 2001 Jan 1;20:109-48.
- Irwin S, O'halloran J, FitzGerald RD. Stocking density, growth and growth variation in juvenile turbot, *Scophthalmus maximus* (Rafinesque). *Aquaculture*. 1999 Jul 15;178(1-2):77-88.
- Jeney G, Nemcsok J, Jeney ZS, Olah J. Acute effect of sublethal ammonia concentrations on common carp (*Cyprinus carpio* L.). II. Effect of ammonia on blood plasma transaminases (GOT, GPT), G1DH enzyme activity, and ATP value.

- Aquaculture. 1992 Jun 1;104(1-2):149-56.
- Jensen FB. Nitrite disrupts multiple physiological functions in aquatic animals. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*. 2003 May 1;135(1):9-24.
- Jensen FB. Nitrite and red cell function in carp: control factors for nitrite entry, membrane potassium ion permeation, oxygen affinity and methaemoglobin formation. *Journal of Experimental Biology*. 1990 Sep 1;152(1):149-66.
- Jensen FB, Andersen NA, Heisler N. Effects of nitrite exposure on blood respiratory properties, acid-base and electrolyte regulation in the carp (*Cyprinus carpio*). *Journal of Comparative Physiology B*. 1987 Sep 1;157(5):533-41.
- Jobling MA. Temperature and growth: modulation of growth rate via temperature change. In *Seminar series-society for experimental biology* 1997 Jan 1 (Vol. 61, pp. 225-254). Cambridge University Press.
- Ogbonna J, Chinomso A. Determination of the concentration of ammonia that could have lethal effect on fish pond. *Journal of Engineering and Applied Sciences (Asian Research Publishing Network)*. 2010;5:1-5.
- Jones DP. The role of oxygen concentration in oxidative stress: hypoxic and hyperoxic models. *Oxidative stress*. 1985 Jan 1;151-95.
- Jordan AD, Steffensen JF. Effects of ration size and hypoxia on specific dynamic action in the cod. *Physiological and Biochemical Zoology*. 2007 Mar;80(2):178-85.
- Kirk RS, Lewis JW. An evaluation of pollutant induced changes in the gills of rainbow trout using scanning electron microscopy. *Environmental Technology*. 1993 Jun 1;14(6):577-85.
- Knoph MB. Gill ventilation frequency and mortality of Atlantic salmon (*Salmo salar* L.) exposed to high ammonia levels in seawater. *Water Research*. 1996 Apr 1;30(4):837-42.
- Knudsen PK, Jensen FB. Recovery from nitrite-induced methaemoglobinaemia and potassium balance disturbances in carp. *Fish Physiology and Biochemistry*. 1997 Jan 1;16(1):1-0.
- Kramer DL. Dissolved oxygen and fish behavior. *Environmental biology of fishes*. 1987 Feb 1;18(2):81-92.
- Kristiansen TS, Fernö A, Holm JC, Privitera L, Bakke S, Fosseidengen JE. Swimming behaviour as an indicator of low growth rate and impaired welfare in Atlantic halibut

- (*Hippoglossus hippoglossus* L.) reared at three stocking densities. *Aquaculture*. 2004 Feb 1;230(1-4):137-51.
- Lemarié G, Dosdat A, Coves D, Dutto G, Gasset E, Person-Le Ruyet J. Effect of chronic ammonia exposure on growth of European seabass (*Dicentrarchus labrax*) juveniles. *Aquaculture*. 2004 Jan 12;229(1-4):479-91.
- Lipton, A. P. "Diseases of ornamental fishes and their control." (2006): 109-114.
- Magnuson JJ, Beckel AL, Mills K, Brandt SB. Surviving winter hypoxia: behavioral adaptations of fishes in a northern Wisconsin winterkill lake. *Environmental Biology of Fishes*. 1985 Dec 1;14(4):241-50.
- Martinez CB, Souza MM. Acute effects of nitrite on ion regulation in two neotropical fish species. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*. 2002 Sep 1;133(1):151-60.
- Masser MP, Rakocy J, Losordo TM. Recirculating aquaculture tank production systems. *Management of recirculating systems*. SRAC Publication. 1999;452.
- McKenzie DJ, Steffensen JF, Korsmeyer K, Whiteley NM, Bronzi P, Taylor EW. Swimming alters responses to hypoxia in the Adriatic sturgeon *Acipenser naccarii*. *Journal of Fish Biology*. 2007 Feb;70(2):651-8.
- Meade JW. Allowable ammonia for fish culture. *The Progressive Fish-Culturist*. 1985 Jul;47(3):135-45.
- Mommsen TP, Vijayan MM, Moon TW. Cortisol in teleosts: dynamics, mechanisms of action, and metabolic regulation. *Reviews in Fish Biology and Fisheries*. 1999 Sep 1;9(3):211-68.
- Montero D, Izquierdo MS, Tort L, Robaina L, Vergara JM. High stocking density produces crowding stress altering some physiological and biochemical parameters in gilthead seabream, *Sparus aurata*, juveniles. *Fish Physiology and Biochemistry*. 1999 Jan 1;20(1):53-60.
- Pang X, Cao ZD, Peng JL, Fu SJ. The effects of feeding on the swimming performance and metabolic response of juvenile southern catfish, *Silurus meridionalis*, acclimated at different temperatures. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*. 2010 Feb 1;155(2):253-8.
- Petersen LH, Gamperl AK. Effect of acute and chronic hypoxia on the swimming performance, metabolic capacity and cardiac function of Atlantic cod (*Gadus morhua*). *Journal of Experimental Biology*. 2010 Mar 1;213(5):808-19.

- Petri D, Glover CN, Ylving S, Kolås K, Fremmersvik G, Waagbø R, Berntssen MH. Sensitivity of Atlantic salmon (*Salmo salar*) to dietary endosulfan as assessed by haematology, blood biochemistry, and growth parameters. *Aquatic Toxicology*. 2006 Dec 1;80(3):207-16.
- Poulin R, Wolf NG, Kramer DL. The effect of hypoxia on the vulnerability of guppies (*Poecilia reticulata*, Poeciliidae) to an aquatic predator (*Astronotus ocellatus*, Cichlidae). *Environmental Biology of Fishes*. 1987 Dec 1;20(4):285-92.
- Prosser CL. Introduction: Definition of comparative physiology: theory of adaptation. *Environmental and metabolic animal physiology*. 1991;1:1-citation_lastpage.
- Rishikesh S D, Asim K P, Lalchand R T, Tilak D, Kartik B. Thermal tolerance and oxygen consumption rates of the catfish *Horabagrus brachysoma* (Günther) acclimated to different temperatures. *Aquaculture*. 2009 Oct 1;295(1-2):116-9.
- Robert JS, Lewis WM. Influence of pH and ammonia salts on ammonia toxicity and water balance in young channel catfish. *Transactions of the American Fisheries Society*. 1986 Nov 1;115(6):891-9.
- Ruffier PJ, Boyle WC, Kleinschmidt J. Short-term acute bioassays to evaluate ammonia toxicity and effluent standards. *Journal (Water Pollution Control Federation)*. 1981 Mar 1;367-77.
- Russo RC, Thurston RV. Toxicity of ammonia, nitrite and nitrate to fishes. *Aquaculture and water quality*. 1991;3:58-89.
- Russo RC. *Fundamentals of aquatic toxicology*. Hemisphere Publishing Corporation, Washington DC. 1985.
- Saha N, Kharbuli ZY, Bhattacharjee A, Goswami C, Häussinger D. Effect of alkalinity (pH 10) on ureogenesis in the air-breathing walking catfish, *Clarias batrachus*. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*. 2002 Jun 1;132(2):353-64.
- Saint-Paul U, Soares GM. Diurnal distribution and behavioral responses of fishes to extreme hypoxia in an Amazon floodplain lake. *Environmental Biology of Fishes*. 1987 Oct 1;20(2):91-104.
- Salin D, Williot P. Acute toxicity of ammonia to Siberian sturgeon, *Acipenser baeri*. Williot, Ed. *Acipenser*, Cemagref Publ. 1991:153-67.
- Schram E, Van der Heul JW, Kamstra A, Verdegem MC. Stocking density-dependent growth of Dover sole (*Solea solea*). *Aquaculture*. 2006 Mar 10;252(2-4):339-47.

- Scott DM, Lucas MC, Wilson RW. The effect of high pH on ion balance, nitrogen excretion and behaviour in freshwater fish from an eutrophic lake: a laboratory and field study. *Aquatic Toxicology*. 2005 Jun 1;73(1):31-43.
- Shultz AD, Murchie KJ, Griffith C, Cooke SJ, Danylchuk AJ, Goldberg TL, Suski CD. Impacts of dissolved oxygen on the behavior and physiology of bonefish: implications for live-release angling tournaments. *Journal of Experimental Marine Biology and Ecology*. 2011 Jun 15;402(1-2):19-26.
- Smart GR. Investigations of the toxic mechanisms of ammonia to fish—gas exchange in rainbow trout (*Salmo gairdneri*) exposed to acutely lethal concentrations. *Journal of Fish Biology*. 1978 Jan;12(1):93-104.
- Smart G. The effect of ammonia exposure on gill structure of the rainbow trout (*Salmo gairdneri*). *Journal of Fish Biology*. 1976 Jun;8(6):471-5.
- Smith CE. Lesions associated with chronic exposure to ammonia. The pathology of fishes. 1975:497-514.
- Snieszko SF. Bacterial gill disease of freshwater fishes. US Department of the Interior, Fish and Wildlife Service, Division of Fishery Research; 1981.
- Sreeya G.Nair. Studies on Pathophysiology of fresh water ornamental fishes, Ph. D. Thesis, Manonmaniam Sundaranar University, Tirunelveli; 2012.
- Stormer J, Jensen FB, Rankin JC. Uptake of nitrite, nitrate, and bromide in rainbow trout, (*Oncorhynchus mykiss*): effects on ionic balance. *Canadian Journal of Fisheries and Aquatic Sciences*. 1996 Sep 1;53(9):1943-50.
- Suski CD, Killen SS, Kieffer JD, Tufts BL. The influence of environmental temperature and oxygen concentration on the recovery of largemouth bass from exercise: implications for live-release angling tournaments. *Journal of Fish Biology*. 2006 Jan;68(1):120-36.
- Suski CD, Killen SS, Morrissey MB, Lund SG, Tufts BL. Physiological changes in largemouth bass caused by live-release angling tournaments in southeastern Ontario. *North American Journal of Fisheries Management*. 2003 Aug 1;23(3):760-9.
- Suthers IM, Gee JH. Role of hypoxia in limiting diel spring and summer distribution of juvenile yellow perch (*Perca flavescens*) in a prairie marsh. *Canadian Journal of Fisheries and Aquatic Sciences*. 1986 Aug 1;43(8):1562-70.
- Thurston RV, Russo RC, Vinogradov GA. Ammonia toxicity to fishes. Effect of pH on the toxicity of the unionized ammonia species. *Environmental science & technology*.

1981 Jul;15(7):837-40.

Timmons MB, Ebeling JM, Wheaton FW, Summerfelt ST, Vinci BJ. Recirculating Aquaculture Systems. Cayuga Aqua Ventures, Ithaca. NY0-9712646-0-0; 2001.

Tomasso JR. Toxicity of nitrogenous wastes to aquaculture animals. Reviews in Fisheries Science. 1994 Jan 1;2(4):291-314.

US-EPA. 1986. Ambient water quality criteria for dissolved oxygen. Office of Water Regulations and Standards, Criteria and Standards Division, Washington. DC. EPA 440/5-86-003

Vijayan MM, Ballantyne JS, Leatherland JF. High stocking density alters the energy metabolism of brook charr, *Salvelinus fontinalis*. Aquaculture. 1990 Aug 1;88(3-4):371-81.

Wang WN, Wang AL, Zhang YJ, Li ZH, Wang JX, Sun RY. Effects of nitrite on lethal and immune response of *Macrobrachium nipponense*. Aquaculture. 2004 Apr 5;232(1-4):679-86.

Wannamaker CM, Rice JA. Effects of hypoxia on movements and behavior of selected estuarine organisms from the southeastern United States. Journal of Experimental Marine Biology and Ecology. 2000 Jun 28;249(2):145-63.

Wedemeyer G, A. and Goodyear, C. P. 1984 .Diseases caused by environmental stressors in O. Kinne (ed.), Diseases of Marine Animals, vol.4, part 1: Introduction. 424-434

Wedemeyer GA, Yasutake W. Clinical methods for the assessment of the effects of environmental stress on fish health. Department of the Interior, Fish and Wildlife Service; 1977.