

# One-Way MANOVA in R

---

The **Multivariate Analysis Of Variance (MANOVA)** is an ANOVA with two or more continuous outcome (or response) variables.

The one-way MANOVA tests simultaneously statistical differences for multiple response variables by one grouping variables.

For example, we may conduct an experiment where we give two treatments (A and B) to two groups of mice, and we are interested in the weight and height of mice. In that case, the weight and height of mice are our outcome (or dependent) variables, and our hypothesis is that both together are affected by the difference in treatment. A multivariate analysis of variance could be used to test this hypothesis.

The procedure of MANOVA can be summarized as follow:

1. Create a new composite variable that is a linear combination of all the response variables.
2. Compare the mean values of this new variable between groups.

This article describes how to compute one-way MANOVA in R.

Note that, MANOVA is appropriate in experimental situations, where we have several outcome (dependent) variables which all measure different aspects of some cohesive theme. For example, several exam scores to have a measure of overall academic performance.

Contents:

- [Prerequisites](#)
- [Data preparation](#)
- [Visualization](#)
- [Summary statistics](#)
- Assumptions and preliminary tests
  - [Check sample size assumption](#)
  - [Identify univariate outliers](#)
  - [Detect multivariate outliers](#)
  - [Check univariate normality assumption](#)
  - [Multivariate normality](#)
  - [Identify multicollinearity](#)
  - [Check linearity assumption](#)
  - [Check the homogeneity of covariances assumption](#)
  - [Check the homogeneity of variance assumption](#)
- [Computation](#)
- Post-hoc tests
  - [Compute univariate one-way ANOVA](#)
  - [Compute multiple pairwise comparisons](#)
- [Report](#)

- [Summary](#)
- [References](#)

## Related Book

Practical Statistics in R II - Comparing Groups: Numerical Variables

## Prerequisites

---

Make sure that you have installed the following R packages:

- `tidyverse` for data manipulation and visualization
- `ggpubr` for creating easily publication ready plots
- `rstatix` for easy pipe-friendly statistical analyses
- `car` for MANOVA analyses
- `broom` for printing a nice summary of statistical tests as data frames
- `datarium` contains required data sets for this chapter

Start by loading the following R packages:

```
1 library(tidyverse)
2 library(ggpubr)
3 library(rstatix)
4 library(car)
5 library(broom)
```

## Data preparation

---

We'll use the built-in R dataset `iris`. Select columns of interest:

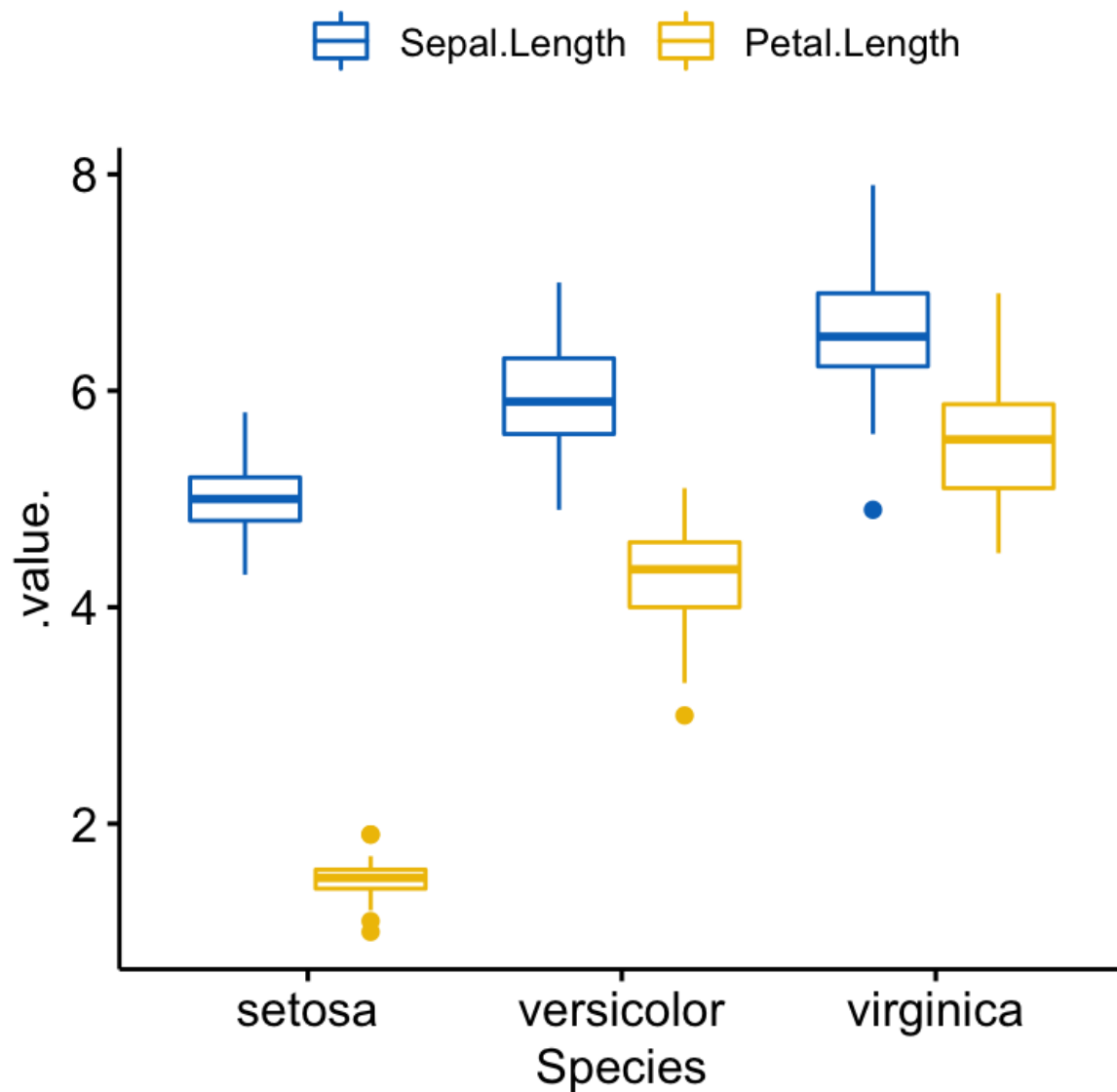
```
1 iris2 <- iris %>%
2   select(Sepal.Length, Petal.Length, Species) %>%
3   add_column(id = 1:nrow(iris), .before = 1)
4 head(iris2)
5 ##   id Sepal.Length Petal.Length Species
6 ## 1  1          5.1          1.4  setosa
7 ## 2  2          4.9          1.4  setosa
8 ## 3  3          4.7          1.3  setosa
9 ## 4  4          4.6          1.5  setosa
10 ## 5  5          5.0          1.4  setosa
11 ## 6  6          5.4          1.7  setosa
```

## Visualization

---

The R code below creates a merged box plots of `Sepal.Length` and `Petal.Length` by `Species` groups.

```
1 ggboxplot(
2   iris2, x = "Species", y = c("Sepal.Length", "Petal.Length"),
3   merge = TRUE, palette = "jco"
4 )
```



## Summary statistics

Compute summary statistics (mean, SD) by groups for each outcome variable:

```
1 iris2 %>%
2   group_by(Species) %>%
3   get_summary_stats(Sepal.Length, Petal.Length, type = "mean_sd")
4 ## # A tibble: 6 x 5
5 ##   Species    variable      n mean    sd
6 ##   <fct>      <chr>      <dbl> <dbl> <dbl>
7 ## 1 setosa    Petal.Length    50  1.46 0.174
8 ## 2 setosa    Sepal.Length    50  5.01 0.352
9 ## 3 versicolor Petal.Length    50  4.26 0.47
10 ## 4 versicolor Sepal.Length    50  5.94 0.516
11 ## 5 virginica Petal.Length    50  5.55 0.552
12 ## 6 virginica Sepal.Length    50  6.59 0.636
```

## Assumptions and preliminary tests

MANOVA makes the following assumptions about the data:

- **Adequate sample size.** Rule of thumb: the n in each cell > the number of outcome variables.

- **Independence of the observations.** Each subject should belong to only one group. There is no relationship between the observations in each group. Having repeated measures for the same participants is not allowed. The selection of the sample should be completely random.
- **Absence of univariate or multivariate outliers.**
- **Multivariate normality.** The R function `mshapiro_test()` [in the `rstatix` package] can be used to perform the Shapiro-Wilk test for multivariate normality.
- **Absence of multicollinearity.** The dependent (outcome) variables cannot be too correlated to each other. No correlation should be above  $r = 0.90$  [Tabachnick and Fidell (2012)].
- **Linearity** between all outcome variables for each group.
- **Homogeneity of variances.** The **Levene's test** can be used to test the equality of variances between groups. Non-significant values of Levene's test indicate equal variance between groups.
- **Homogeneity of variance-covariance matrices.** The **Box's M Test** can be used to check the equality of covariance between the groups. This is the equivalent of a multivariate homogeneity of variance. This test is considered as highly sensitive. Therefore, significance for this test is determined at  $\alpha = 0.001$ .

## Check sample size assumption

```
1 iris2 %>%
2   group_by(Species) %>%
3   summarise(N = n())
4 ## # A tibble: 3 x 2
5 ##   Species      N
6 ##   <fct>    <int>
7 ## 1 setosa      50
8 ## 2 versicolor  50
9 ## 3 virginica   50
```

As the table above shows 50 observations per group, the assumption of adequate sample size is satisfied.

## Identify univariate outliers

Univariate outliers can be easily identified using box plot methods, implemented in the R function `identify_outliers()` [rstatix package].

Group the data by `Species` and then, identify outliers in the `Sepal.Length` variable:

```
1 iris2 %>%
2   group_by(Species) %>%
3   identify_outliers(Sepal.Length)
4 ## # A tibble: 1 x 6
5 ##   Species      id Sepal.Length Petal.Length is.outlier is.extreme
6 ##   <fct>    <int>         <dbl>         <dbl> <lgl>      <lgl>
7 ## 1 virginica  107           4.9           4.5 TRUE      FALSE
```

Group the data by `Species` and then, identify outliers in the `Petal.Length` variable:

```

1 iris2 %>%
2   group_by(Species) %>%
3   identify_outliers(Petal.Length)
4 ## # A tibble: 5 x 6
5 ##   Species      id Sepal.Length Petal.Length is.outlier is.extreme
6 ##   <fct>      <int>      <dbl>      <dbl> <lgl>      <lgl>
7 ## 1 setosa      14         4.3         1.1 TRUE      FALSE
8 ## 2 setosa      23         4.6         1 TRUE      FALSE
9 ## 3 setosa      25         4.8         1.9 TRUE      FALSE
10 ## 4 setosa      45         5.1         1.9 TRUE      FALSE
11 ## 5 versicolor 99         5.1         3 TRUE      FALSE

```

There were no univariate extreme outliers in the Sepal.Length and Petal.Length variable, as assessed by box plot methods.

Note that, in the situation where you have extreme outliers, this can be due to: 1) data entry errors, measurement errors or unusual values.

You can include the outlier in the analysis anyway if you do not believe the result will be substantially affected. This can be evaluated by comparing the result of the MANOVA with and without the outlier.

Remember to report in your written results section any decisions you make regarding any outliers you find.

## Detect multivariate outliers

Multivariate outliers are data points that have an unusual combination of values on the outcome (or dependent) variables.

In MANOVA setting, the **Mahalanobis distance** is generally used to detect multivariate outliers. The distance tells us how far an observation is from the center of the cloud, taking into account the shape (covariance) of the cloud as well.

The function `mahalanobis_distance()` [rstatix package] can be easily used to compute the Mahalanobis distance and to flag multivariate outliers. Read more in the documentation of the function.

This metric needs to be calculated by groups:

```

1 # Compute distance by groups and filter outliers
2 # Use -id to omit the id column in the computation
3 iris2 %>%
4   group_by(Species) %>%
5   mahalanobis_distance(-id) %>%
6   filter(is.outlier == TRUE) %>%
7   as.data.frame()
8 ## [1] id      Sepal.Length Petal.Length mahal.dist  is.outlier
9 ## <0 rows> (or 0-length row.names)

```

There were no multivariate outliers in the data, as assessed by Mahalanobis distance ( $p > 0.001$ ).

If you have multivariate outliers, you could consider running MANOVA before and after removing the outlier to check whether or not their presence alters the results. You should report your final decision.

## Check univariate normality assumption

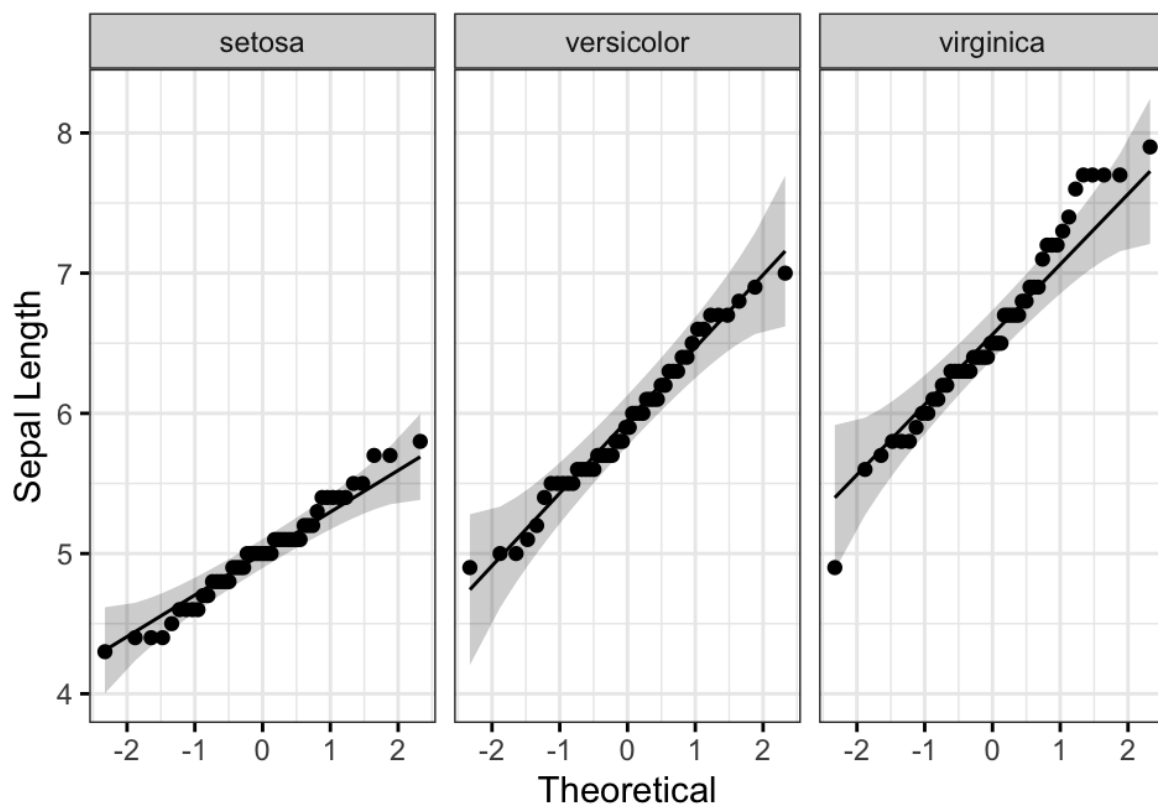
The normality assumption can be checked by computing Shapiro-Wilk test for each outcome variable at each level of the grouping variable. If the data is normally distributed, the p-value should be greater than 0.05.

```
1 iris2 %>%
2   group_by(Species) %>%
3   shapiro_test(Sepal.Length, Petal.Length) %>%
4   arrange(variable)
5 ## # A tibble: 6 x 4
6 ##   Species    variable    statistic      p
7 ##   <fct>     <chr>         <dbl>   <dbl>
8 ## 1 setosa    Petal.Length    0.955 0.0548
9 ## 2 versicolor Petal.Length    0.966 0.158
10 ## 3 virginica Petal.Length    0.962 0.110
11 ## 4 setosa    Sepal.Length    0.978 0.460
12 ## 5 versicolor Sepal.Length    0.978 0.465
13 ## 6 virginica Sepal.Length    0.971 0.258
```

Sepal.Length and Petal.length were normally distributed for each Species groups, as assessed by Shapiro-Wilk's test ( $p > 0.05$ ).

You can also create QQ plot for each group. QQ plot draws the correlation between a given data and the normal distribution.

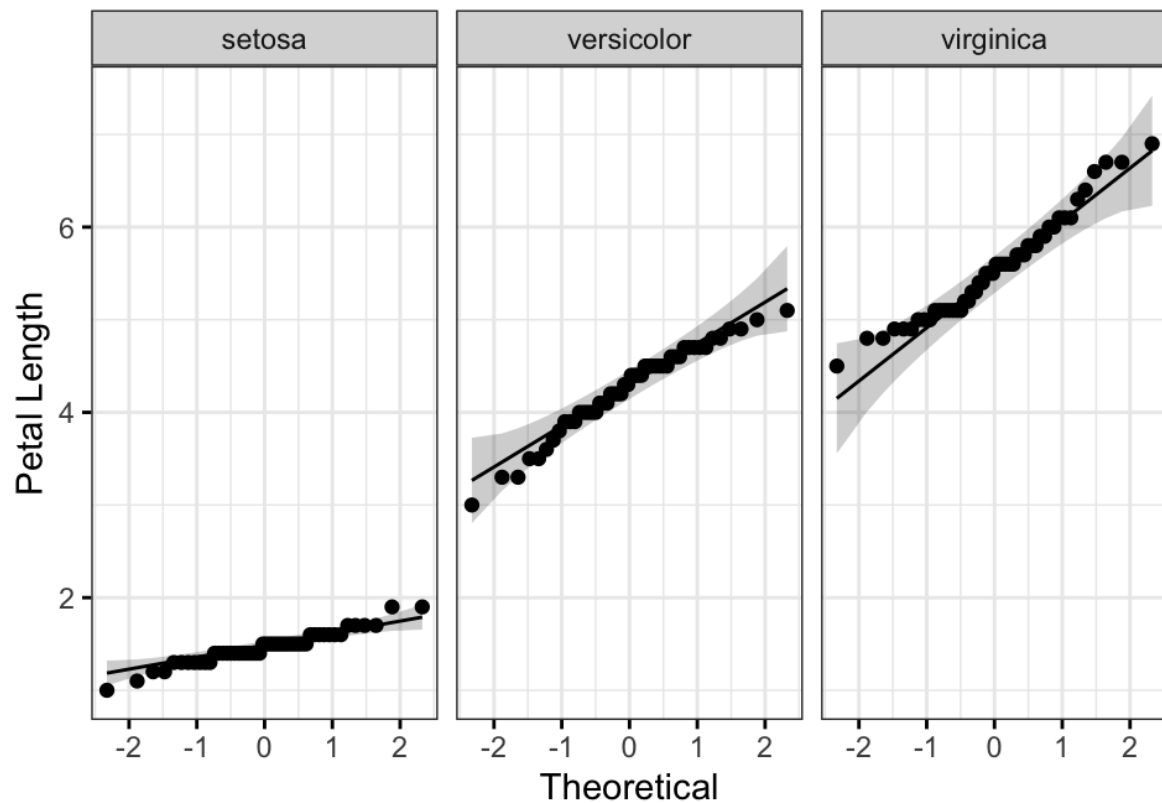
```
1 # QQ plot of Sepal.Length
2 ggqqplot(iris2, "Sepal.Length", facet.by = "Species",
3   ylab = "Sepal Length", ggtheme = theme_bw())
```



```

1 # QQ plot of Petal.Length
2 ggqqplot(iris2, "Petal.Length", facet.by = "Species",
3           ylab = "Petal Length", ggtheme = theme_bw())

```



All the points fall approximately along the reference line, for each group. So we can assume normality of the data.

Note that, if your sample size is greater than 50, the normal QQ plot is preferred because at larger sample sizes the Shapiro-Wilk test becomes very sensitive even to a minor deviation from normality.

In the situation where the assumptions are not met, you could consider running MANOVA on the data after transforming the outcome variables. You can also perform the test regardless as MANOVA is fairly robust to deviations from normality.

## Multivariate normality

```

1 iris2 %>%
2   select(Sepal.Length, Petal.Length) %>%
3   mshapiro_test()
4 ## # A tibble: 1 x 2
5 ##   statistic p.value
6 ##   <dbl>     <dbl>
7 ## 1     0.995     0.855

```

The test is not significant ( $p > 0.05$ ), so we can assume multivariate normality.

## Identify multicollinearity

Ideally the correlation between the outcome variables should be moderate, not too high. A correlation above 0.9 is an indication of multicollinearity, which is problematic for MANOVA.

In other hand, if the correlation is too low, you should consider running separate one-way ANOVA for each outcome variable.

Compute pairwise Pearson correlation coefficients between the outcome variable. In the following R code, we'll use the function `cor_test()` [rstatix package]. If you have more than two outcome variables, consider using the function `cor_mat()`:

```
1 iris2 %>% cor_test(Sepal.Length, Petal.Length)
2 ## # A tibble: 1 x 8
3 ##   var1          var2          cor statistic      p conf.low conf.high
4 ##   <chr>         <chr>         <dbl>     <dbl>   <dbl>   <dbl>   <dbl>
5 ## 1 Sepal.Length Petal.Length  0.87      21.6 1.04e-47  0.827  0.906
   Pearson
```

There was no multicollinearity, as assessed by Pearson correlation ( $r = 0.87$ ,  $p < 0.0001$ ).

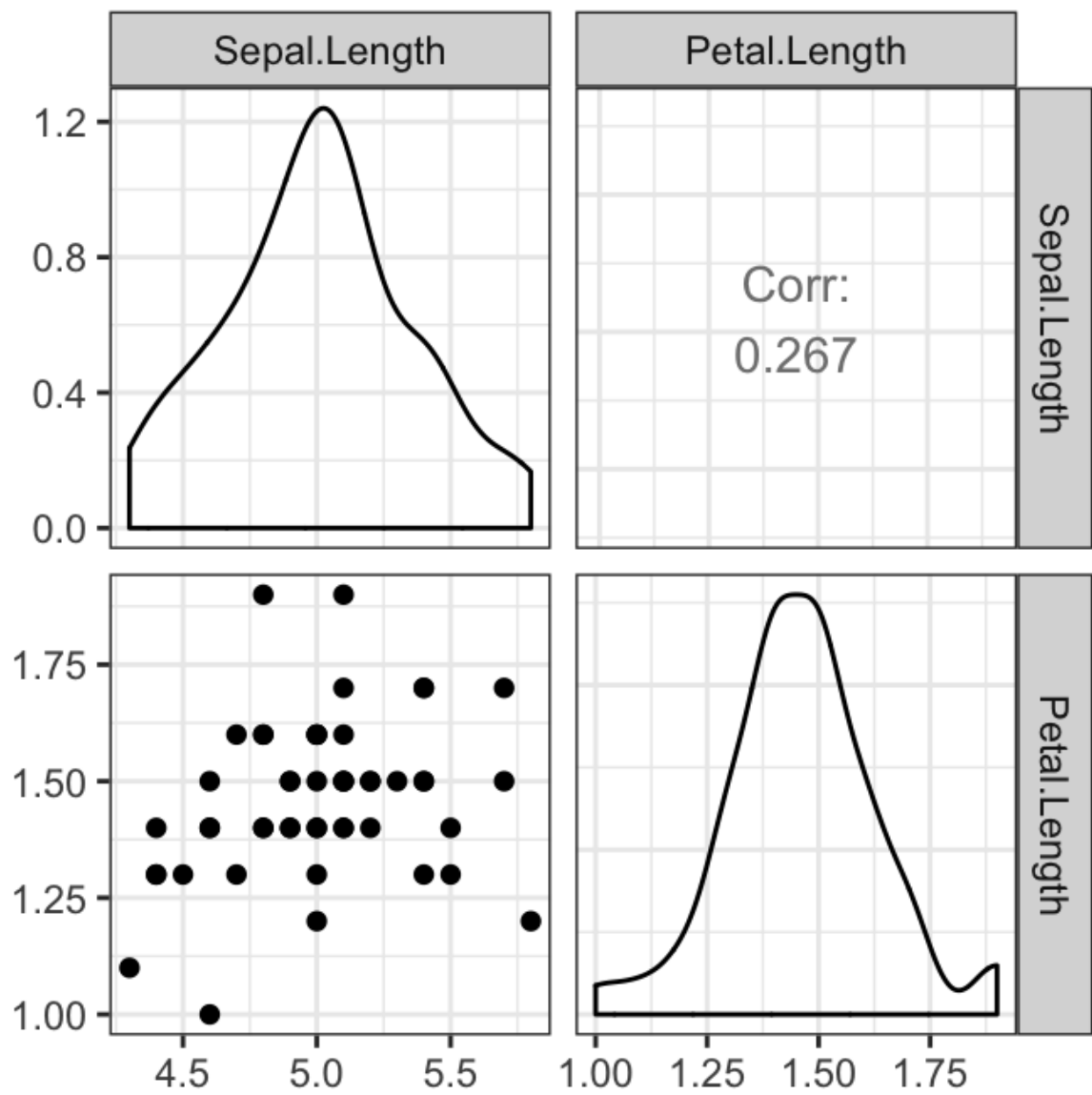
In the situation, where you have multicollinearity, you could consider removing one of the outcome variables that is highly correlated.

## Check linearity assumption

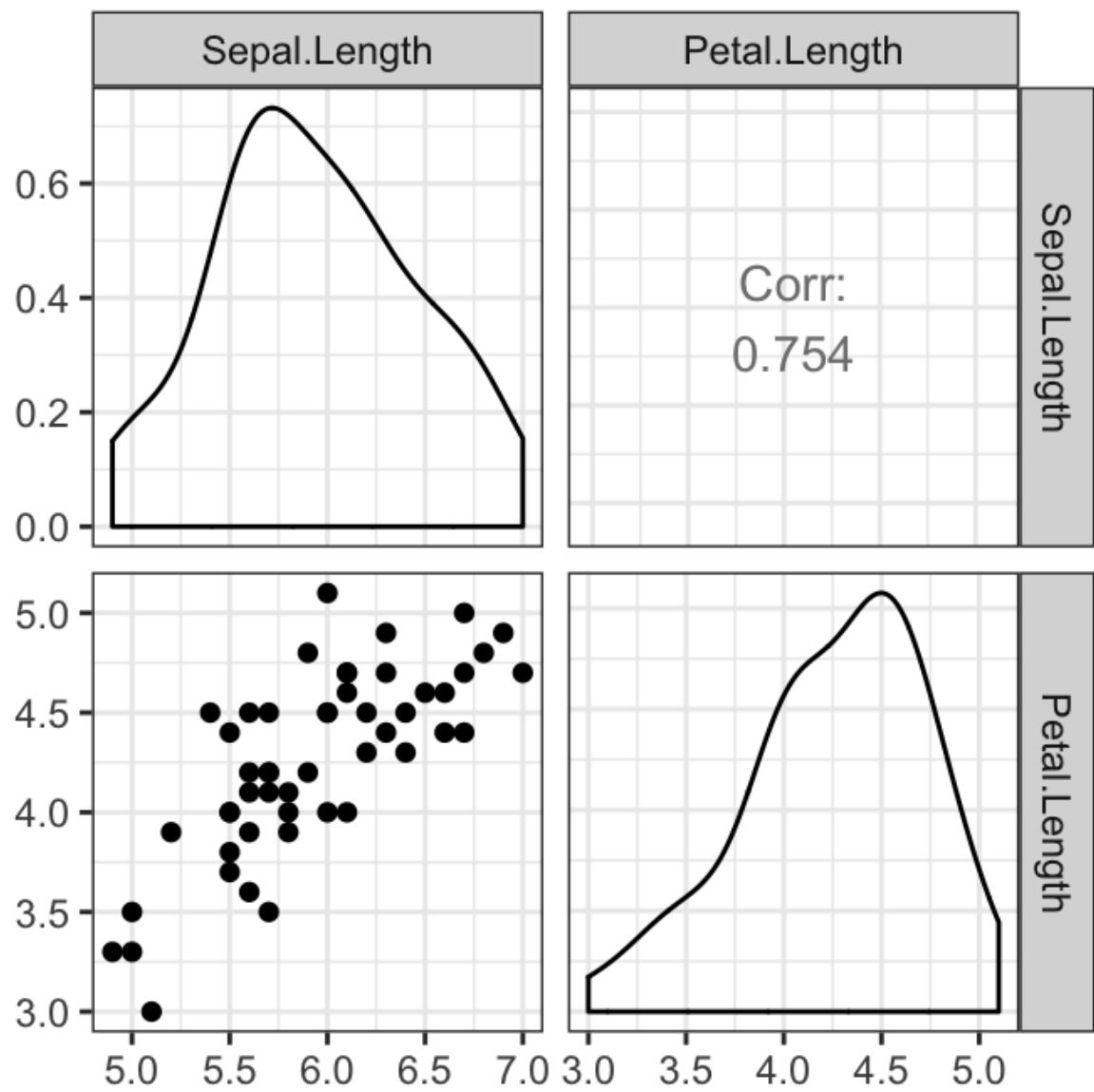
The pairwise relationship between the outcome variables should be linear for each group. This can be checked visually by creating a scatter plot matrix using the R function `ggpairs()` [GGally package]. In our example, we have only one pair:

```
1 # Create a scatterplot matrix by group
2 library(GGally)
3 results <- iris2 %>%
4   select(Sepal.Length, Petal.Length, Species) %>%
5   group_by(Species) %>%
6   doo(~ggpairs(.) + theme_bw(), result = "plots")
7 results
8 ## # A tibble: 3 x 2
9 ##   Species    plots
10 ##   <fct>     <list>
11 ## 1 setosa    <gg>
12 ## 2 versicolor <gg>
13 ## 3 virginica <gg>
14 # Show the plots
15 results$plots
16 ## [[1]]
```

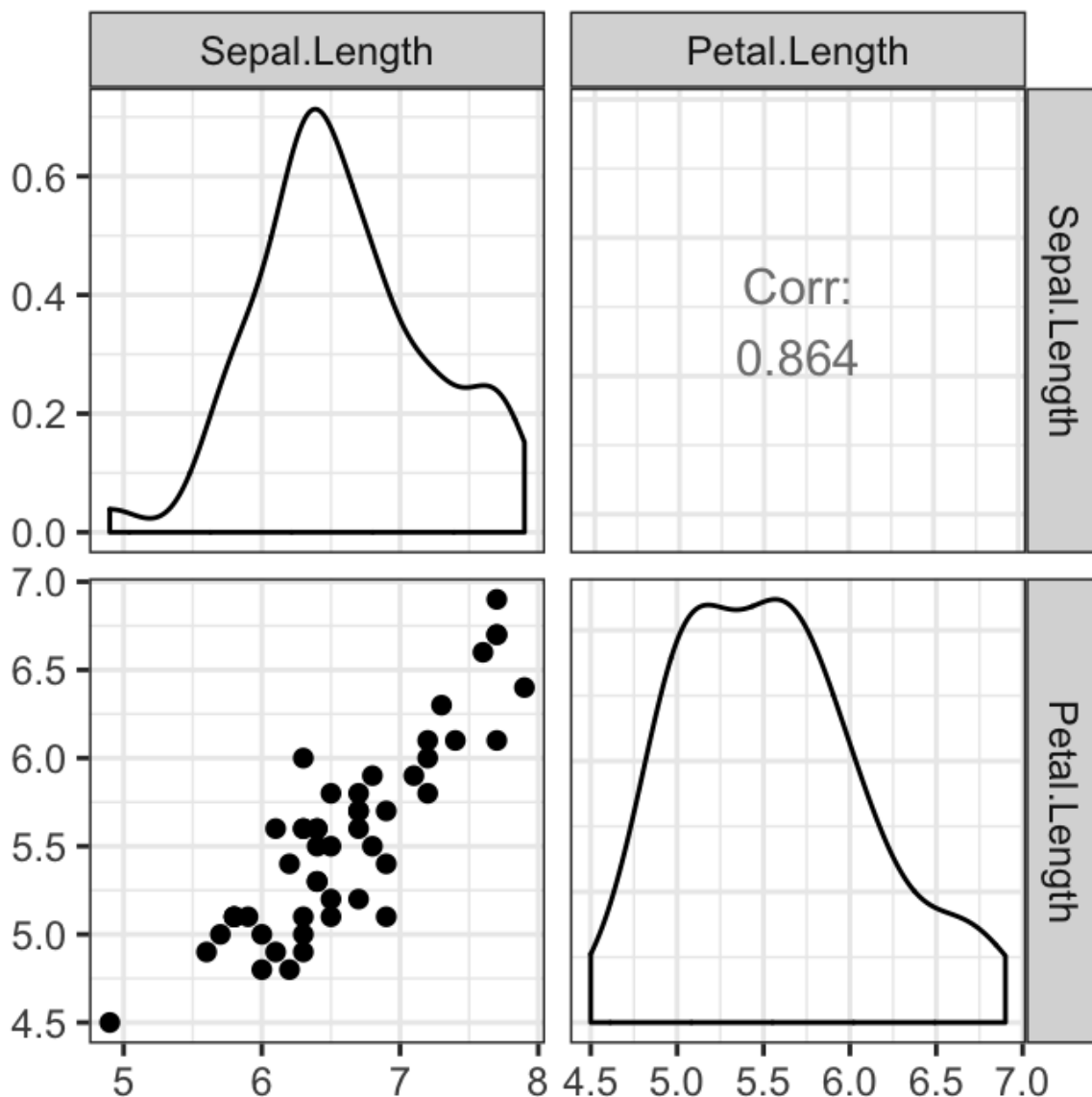




```
1 ##
2 ## [[2]]
```



```
1 ##
2 ## [[3]]
```



There was a linear relationship between Sepal.Length and Petal.Length in each Species group, as assessed by scatter plot.

In the situation, where you detect non-linear relationships, You can:

1. transform or remove the concerned outcome variables;
2. run the analysis anyway. You will loss some power.

## Check the homogeneity of covariances assumption

This can be evaluated using the Box's M-test implemented in the `rstatix` package.

```
1 box_m(iris2[, c("Sepal.Length", "Petal.Length")], iris2$Species)
2 ## # A tibble: 1 x 4
3 ##   statistic p.value parameter method
4 ##       <dbl>   <dbl>   <dbl> <chr>
5 ## 1      58.4 9.62e-11      6 Box's M-test for Homogeneity of Covariance
  Matrices
```

The test is statistically significant (i.e.,  $p < 0.001$ ), so the data have violated the assumption of homogeneity of variance-covariance matrices.

Note that, if you have balanced design (i.e., groups with similar sizes), you don't need to worry too much about violation of the homogeneity of variances-covariance matrices and you can continue your analysis.

However, having an unbalanced design is problematic. Possible solutions include: 1) transforming the dependent variables; 2) running the test anyway, but using **Pillai's** multivariate statistic instead of Wilks' statistic.

## Check the homogeneity of variance assumption

For each of the outcome variables, the one-way MANOVA assumes that there are equal variances between groups. This can be checked using the Levene's test of equality of variances. Key R function: `levene_test()` [rstatix package].

Procedure:

1. Gather the outcome variables into key-value pairs
2. Group by variable
3. Compute the Levene's test

```
1 iris2 %>%
2   gather(key = "variable", value = "value", Sepal.Length, Petal.Length) %>%
3   group_by(variable) %>%
4   levene_test(value ~ Species)
5 ## # A tibble: 2 x 5
6 ##   variable      df1    df2 statistic      p
7 ##   <chr>      <int> <int>      <dbl>    <dbl>
8 ## 1 Petal.Length      2    147      19.5 0.0000000313
9 ## 2 Sepal.Length      2    147      6.35 0.00226
```

The Levene's test is significant ( $p < 0.05$ ), so there was no homogeneity of variances.

Note that, if you do not have homogeneity of variances, you can try to transform the outcome (dependent) variable to correct for the unequal variances.

Alternatively, you can continue, but accept a lower level of statistical significance (alpha level) for your MANOVA result. Additionally, any follow-up univariate ANOVAs will need to be corrected for this violation (i.e., you will need to use different post-hoc tests).

## Computation

There are four different types of multivariate statistics that can be used for computing MANOVA. These are: "Pillai", "Wilks", "Hotelling-Lawley", or "Roy".

The most commonly recommended multivariate statistic to use is **Wilks' Lambda**.

However, **Pillai's Trace** is more robust and is recommended when you have unbalanced design and also have a statistically significant Box's M result (as in our example, see previous section).

Note that, "Pillai" is the default in the R `manova()` function [car package].

Compute MANOVA:

```

1 model <- lm(cbind(Sepal.Length, Petal.Length) ~ Species, iris2)
2 Manova(model, test.statistic = "Pillai")
3 ##
4 ## Type II MANOVA Tests: Pillai test statistic
5 ##           Df test stat approx F num Df den Df Pr(>F)
6 ## Species    2      0.989      71.8      4    294 <2e-16 ***
7 ## ---
8 ## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

There was a statistically significant difference between the Species on the combined dependent variables (Sepal.Length and Petal.Length),  $F(4, 294) = 71.829$ ,  $p < 0.0001$ .

## Post-hoc tests

A statistically significant one-way MANOVA can be followed up by **univariate one-way ANOVA** examining, separately, each dependent variable. The goal is to identify the specific dependent variables that contributed to the significant global effect.

## Compute univariate one-way ANOVA

Procedure:

1. Gather the outcome variables into key-value pairs
2. Group by variable
3. Compute one-way ANOVA test

Note that, there are different R function to compute one-way ANOVA depending whether the assumptions are met or not:

- `anova_test()` [rstatix]: can be used when normality and homogeneity of variance assumptions are met
- `welch_anova_test()` [rstatix]: can be used when the homogeneity of variance assumption is violated, as in our example.
- `kruskal_test()` [rstatix]: Kruskal-Wallis test, a non parametric alternative of one-way ANOVA test

The following R codes shows how to use each of these functions:

```

1 # Group the data by variable
2 grouped.data <- iris2 %>%
3   gather(key = "variable", value = "value", Sepal.Length, Petal.Length) %>%
4   group_by(variable)
5
6 # Do welch one way anova test
7 grouped.data %>% welch_anova_test(value ~ Species)
8 # or do kruskal-wallis test
9 grouped.data %>% kruskal_test(value ~ Species)
10 # or use aov()
11 grouped.data %>% anova_test(value ~ Species)

```

Here, we show the results of `anova_test()`:

```

1 ## # A tibble: 2 x 8
2 ##   variable      Effect    DFn   DFd     F      p `p<.05` ges
3 ##   <chr>         <chr>   <dbl> <dbl> <dbl>   <dbl> <chr>   <dbl>
4 ## 1 Petal.Length Species      2    147 1180. 2.86e-91 *    0.941
5 ## 2 Sepal.Length Species      2    147  119. 1.67e-31 *    0.619

```

There was a statistically significant difference in Sepal.Length ( $F(2, 147) = 119, p < 0.0001$ ) and Petal.Length ( $F(2, 147) = 1180, p < 0.0001$ ) between iris Species.

Note that, as we have two dependent variables, we need to apply Bonferroni multiple testing correction by decreasing the level we declare statistical significance.

This is done by dividing classic alpha level (0.05) by the number of tests (or dependent variables, here 2). This leads to a significance acceptance criteria of  $p < 0.025$  rather than  $p < 0.05$  because there are two dependent variables.

## Compute multiple pairwise comparisons

A statistically significant univariate ANOVA can be followed up by multiple pairwise comparisons to determine which groups are different.

The R functions `tukey_hsd()` [rstatix package] can be used to compute Tukey post-hoc tests if the homogeneity of variance assumption is met.

If you had violated the assumption of homogeneity of variances, as in our example, you might prefer to run a Games-Howell post-hoc test. It's also possible to use the function

`pairwise_t_test()` [rstatix] with the option `pool.sd = FALSE` and `var.equal = FALSE`.

```

1 pwc <- iris2 %>%
2   gather(key = "variables", value = "value", Sepal.Length, Petal.Length) %>%
3   group_by(variables) %>%
4   games_howell_test(value ~ Species) %>%
5   select(-estimate, -conf.low, -conf.high) # Remove details
6 pwc
7 ## # A tibble: 6 x 6
8 ##   variables .y. group1      group2      p.adj p.adj.signif
9 ## * <chr>   <chr> <chr>      <chr>      <dbl> <chr>
10 ## 1 Petal.Length value setosa      versicolor 1.85e-11 ****
11 ## 2 Petal.Length value setosa      virginica 1.68e-11 ****
12 ## 3 Petal.Length value versicolor virginica 4.45e-10 ****
13 ## 4 Sepal.Length value setosa      versicolor 2.86e-10 ****
14 ## 5 Sepal.Length value setosa      virginica 0.      ****
15 ## 6 Sepal.Length value versicolor virginica 5.58e- 7 ****

```

All pairwise comparisons were significant for each of the outcome variable (Sepal.Length and Petal.Length).

## Report

A one-way multivariate analysis of variance was performed to determine the effect of iris Species on Sepal.Length and Petal.Length. There are three different species: setosa, versicolor and virginica.

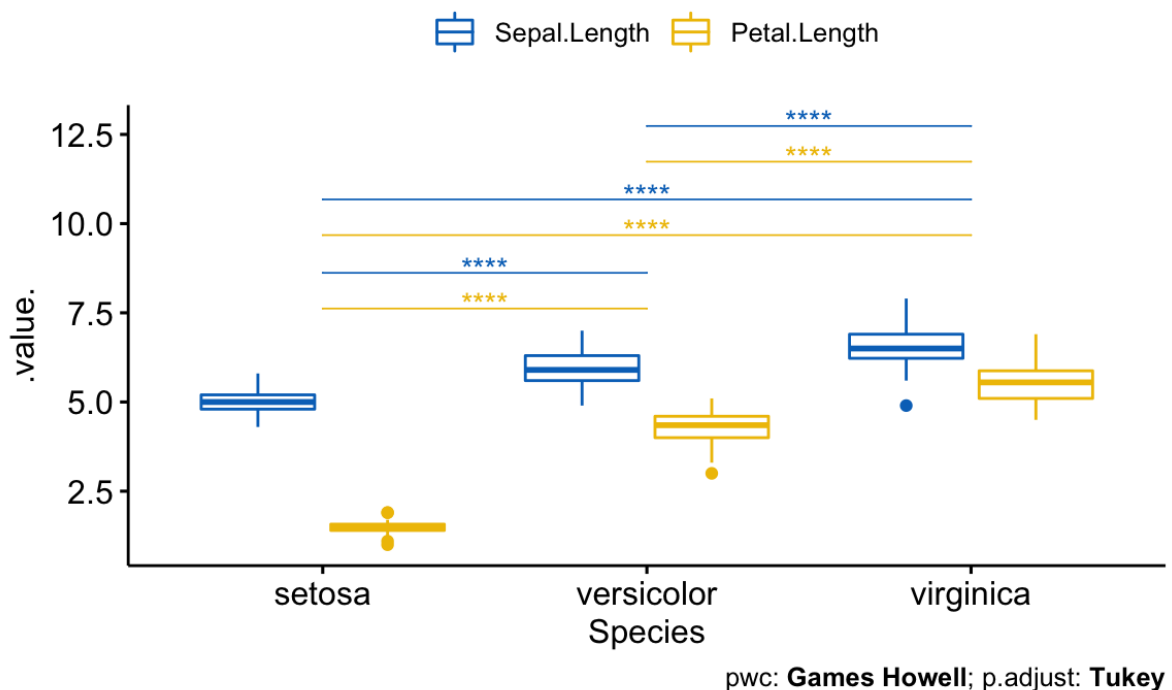
There was a statistically significant difference between the Species on the combined dependent variables (Sepal.Length and Petal.Length),  $F(4, 294) = 71.829$ ,  $p < 0.0001$ .

Follow-up univariate ANOVAs, using a Bonferroni adjusted alpha level of 0.025, showed that there was a statistically significant difference in Sepal.Length ( $F(2, 147) = 119$ ,  $p < 0.0001$ ) and Petal.Length ( $F(2, 147) = 1180$ ,  $p < 0.0001$ ) between iris Species.

All pairwise comparisons between groups were significant for each of the outcome variable (Sepal.Length and Petal.Length).

```
1 # visualization: box plots with p-values
2 pwc <- pwc %>% add_xy_position(x = "Species")
3 test.label <- create_test_label(
4   description = "MANOVA", statistic.text = quote(italic("F")),
5   statistic = 71.83, p = "<0.0001", parameter = "4,294",
6   type = "expression", detailed = TRUE
7 )
8 ggboxplot(
9   iris2, x = "Species", y = c("Sepal.Length", "Petal.Length"),
10  merge = TRUE, palette = "jco"
11 ) +
12  stat_pvalue_manual(
13    pwc, hide.ns = TRUE, tip.length = 0,
14    step.increase = 0.1, step.group.by = "variables",
15    color = "variables"
16  ) +
17  labs(
18    subtitle = test.label,
19    caption = get_pwc_label(pwc, type = "expression")
20  )
```

MANOVA,  $F(4,294) = 71.83$ ,  $p = <0.0001$



## Summary

This article describes how to compute and interpret one-way MANOVA in R. We show how to check the test assumptions and to perform post-hoc analyses.