

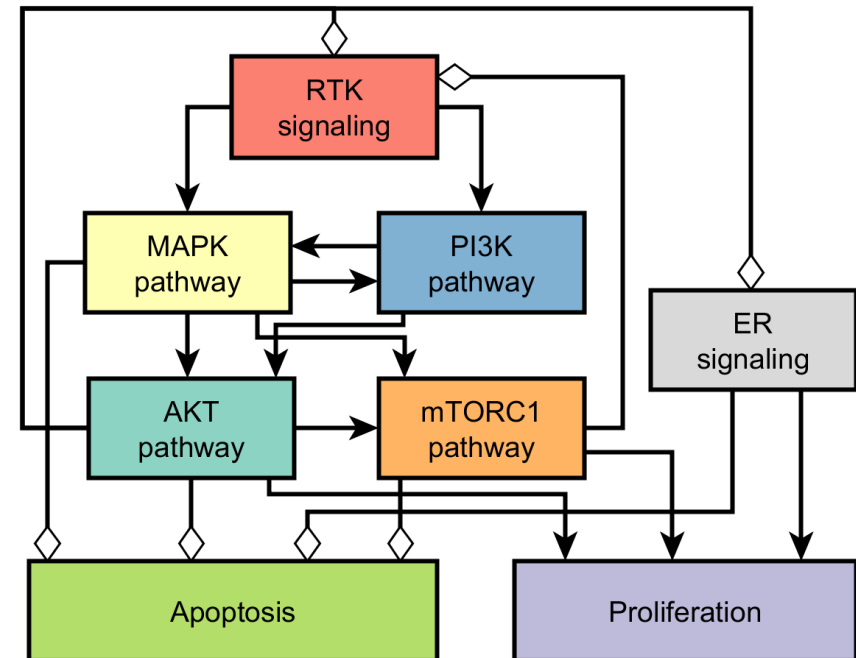
SACB Meeting. MBL.
Nov 9th 2018.



Drug resistance mechanisms and combinatorial drug treatments in breast cancer: a network modeling approach

Jorge Gómez Tejeda Zañudo
Réka Albert
Pennsylvania State University
Dana-Farber Cancer Institute
Broad Institute

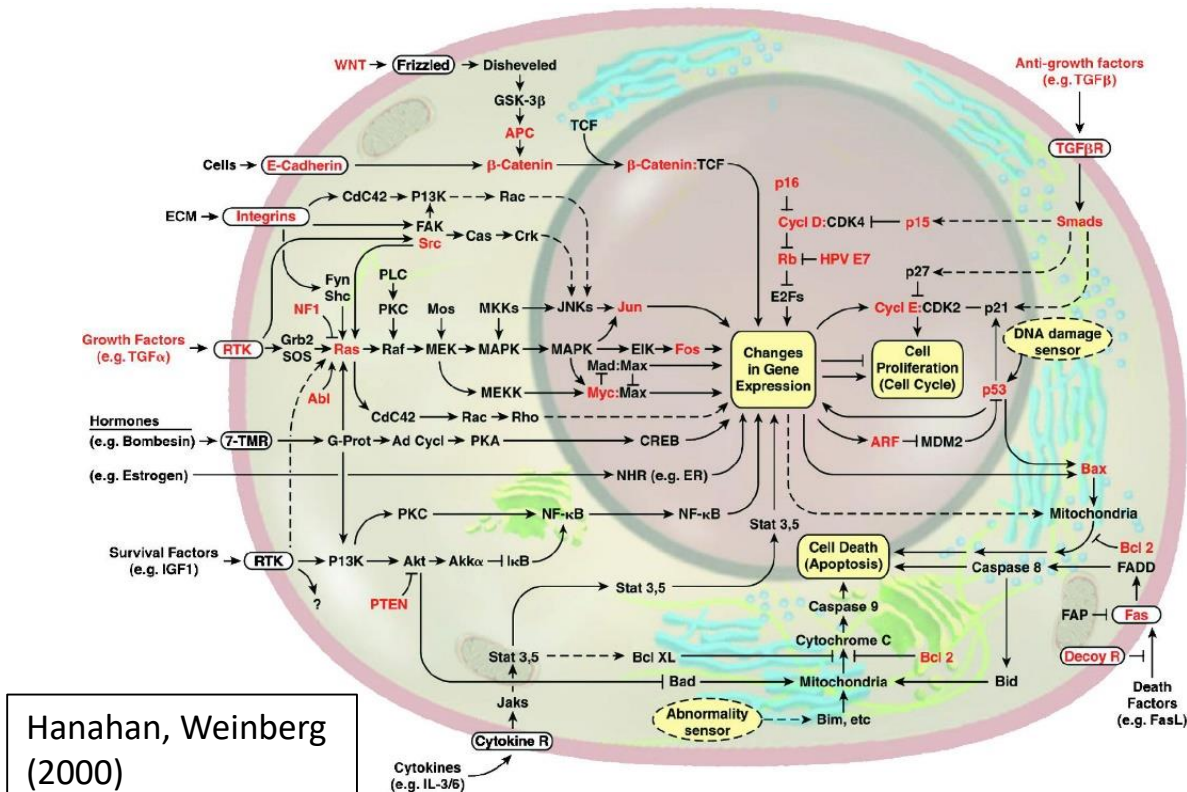
**SU2C Drug Combination
Convergence Team**



Cellular phenotypes arise from the interactions of molecular components

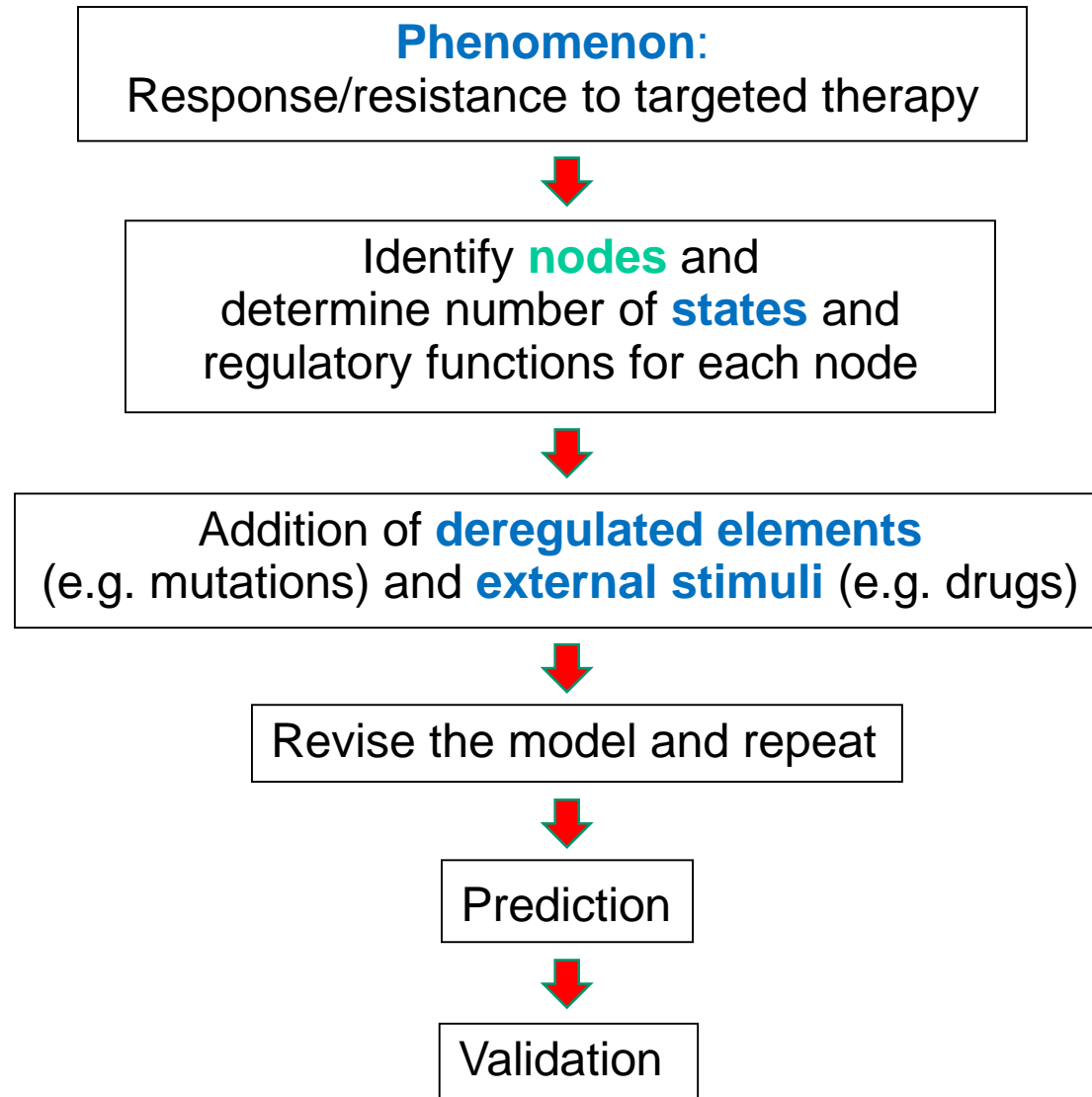
Interconnection of breast cancer signaling pathways → network model

Network model: *Dynamics of information propagation through these pathways and their connection to cellular behaviors*

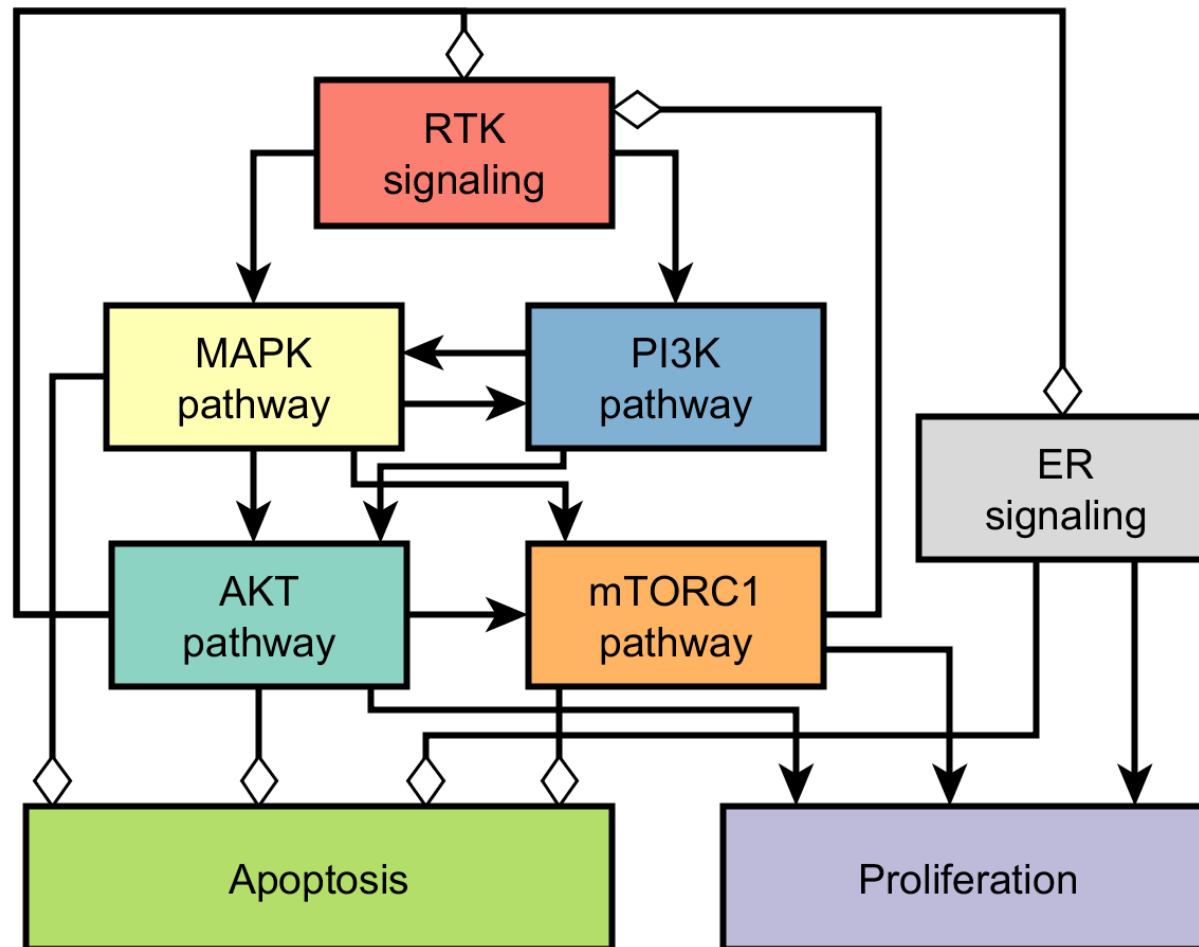


Hanahan, Weinberg
(2000)

Steps in constructing a Boolean model of a signaling network



The general structure of the network model (based on what we know)
ER-positive PIK3CA-mutant breast cancer

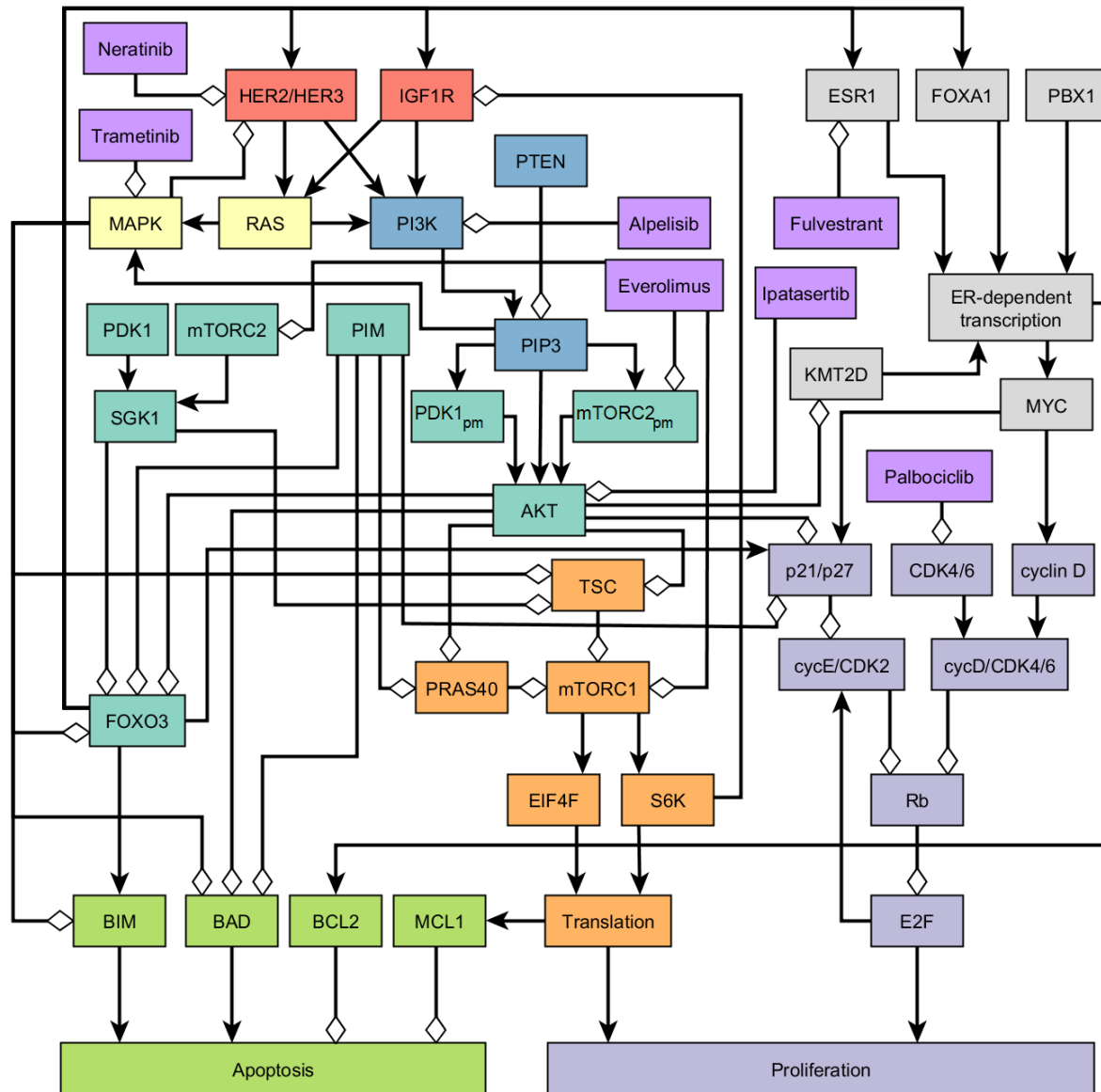


Breast cancer network model

J.G.T. Zañudo

Network modeling of breast cancer to predict drug resistance mechanisms & combinatorial drug treatments

The signaling network: response to pharmacological perturbation



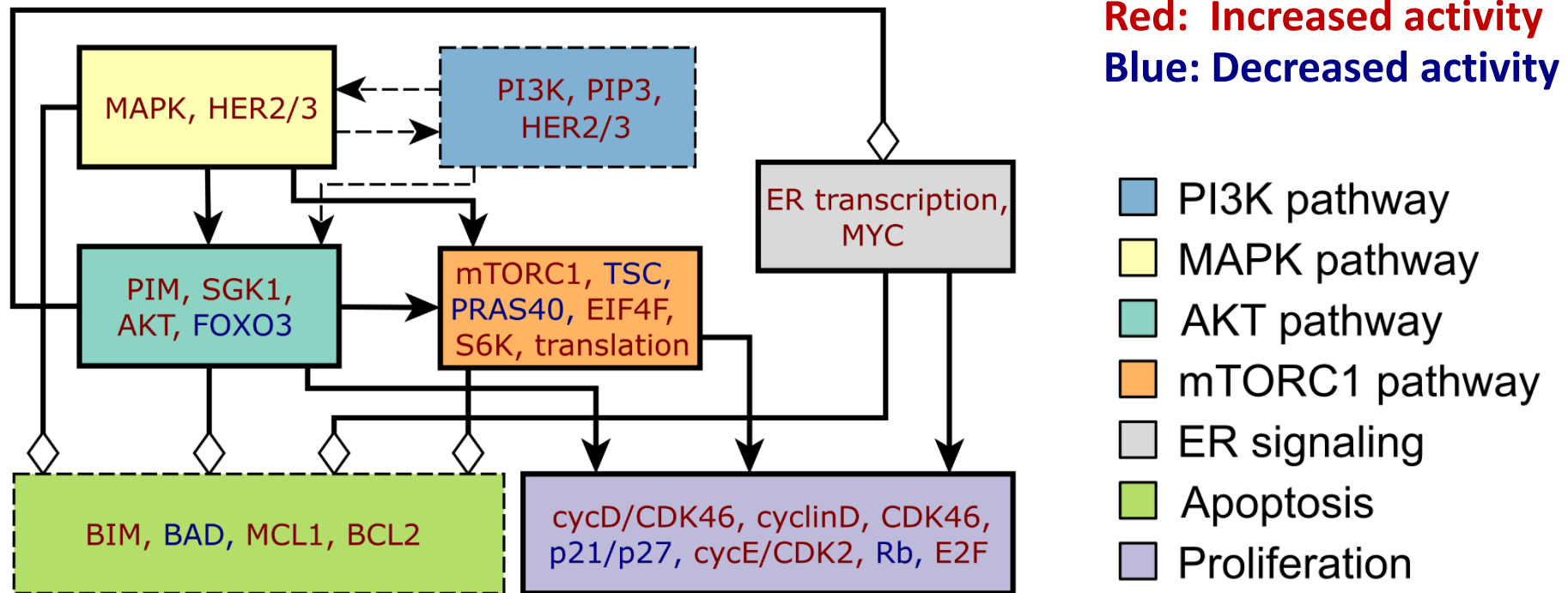
- MAPK pathway
- RTK signaling
- mTORC1 pathway
- PI3K pathway
- ER signaling
- AKT pathway
- Apoptosis
- Proliferation
- Drugs

Model recapitulates several known experimental and clinical outcomes

Table 4 Illustration of experimental and clinical outcomes in ER+ and HER2+ breast cancer reproduced by the model

Experimental or clinical outcome	References
Drug inhibition of MEK in HER2+ breast cancer cells leads to increased HER2/HER3 heterodimer formation and higher PI3K activation	(Turke et al., 2012)
High HER3 expression induces resistance to PI3K inhibitors, which is overcome by HER3 blockade, in HER2-amplified and/or PIK3CA-mutant breast cancer cell lines and brain metastases of mouse xenografts.	((Kodack et al., 2017); (Chakrabarty et al., 2012))
High PIM expression is a resistance mechanism to PI3K inhibitors in ER+ (PIM1/2/3) and HER2+ (PIM2) breast cancer cell lines. High PIM1/3 expression is observed in biopsies of ER+ human tumors treated with PI3K inhibitors.	((Le et al., 2016); (Zwang et al., 2017))
High PDK1/SGK1 expression is a resistance mechanism to PI3K inhibitors in HER2+ breast cancer cell lines and mouse xenografts tumors. High SGK1 expression and activity in breast cancer tumor samples causes intrinsic resistance to PI3K inhibitors.	(Castel et al., 2016)
High PDK1 and AKT2 expression are putative resistance mechanisms to PI3K inhibitors; they are observed in biopsies of ER+ human tumors treated with PI3K inhibitors.	(Le et al., 2016)
Inhibition of PI3K induces a rapid downregulation of MAPK signaling and induction of apoptosis in ER+ and HER2+ breast cancer cell lines and mouse xenograft tumors. In HER2+ breast cancer cell lines, MAPK activity is reactivated following the induction of RTKs.	((Costa et al., 2015); (Will et al., 2014); (Ebi et al., 2013))
Inhibition of AKT (directly by AKT inhibitors or indirectly by mTOR or PI3K inhibitors) induces the activity of the transcription factor FOXO3, which upregulates a shared set of RTKs, including HER3, IGF1R, in HER2+ breast cancer cell lines and mouse xenografts tumors.	((Chandarlapaty et al., 2011); (Rodrik-Outmezguine et al., 2011); (Chakrabarty et al., 2012))
Inhibition of PI3K in ER+ breast cancer cell lines induces the transcription factor activity of FOXO3, which binds the promoters of ESR1 and HER3, and upregulates their expression. The upregulation of ESR1 expression in response to PI3K inhibitors has also been observed in ER+ mouse xenograft tumors and ER+ human breast cancer tumor biopsies.	((Bosch et al., 2015); (Kodack et al., 2017))

Systematic search for PI3Ki resistance mechanisms



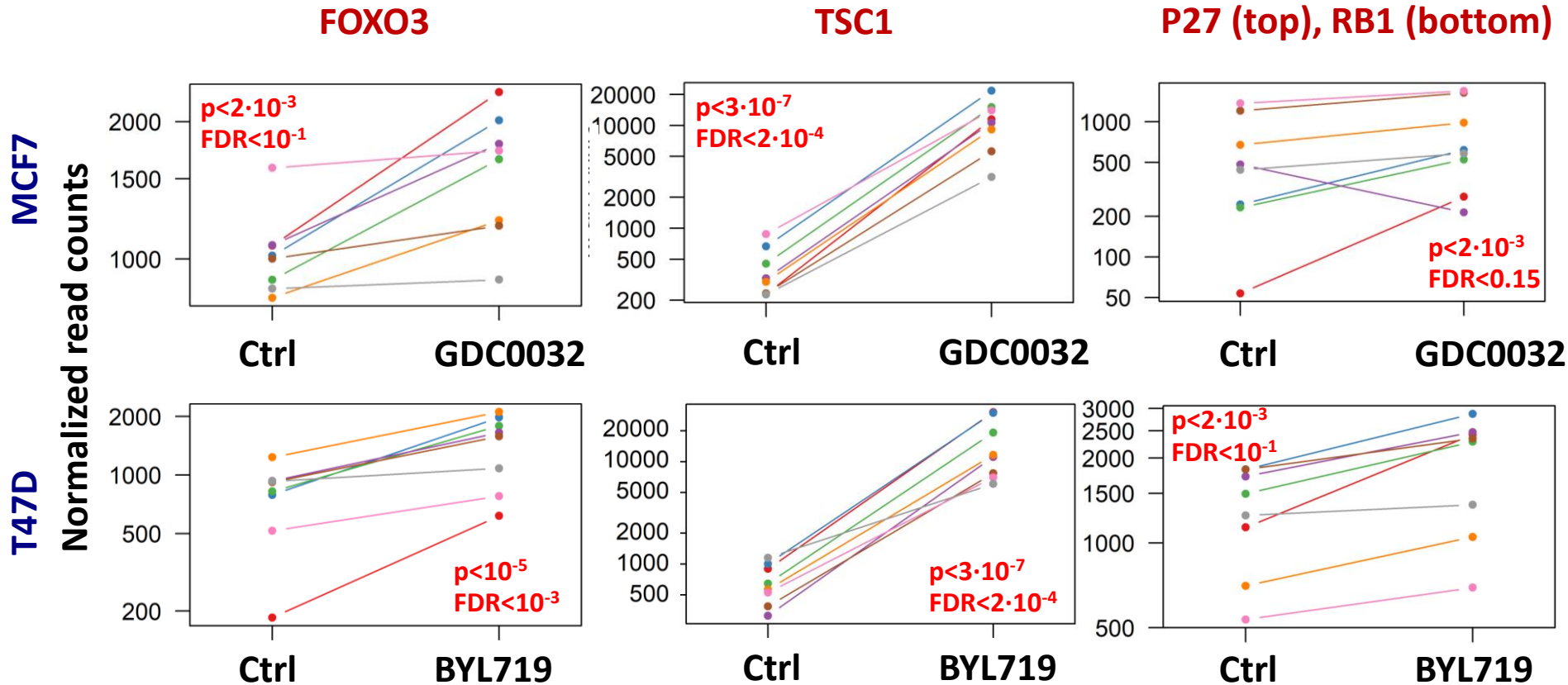
Predicts **loss of FOXO3, TSC, RB, P27, or PRAS40**, increased MAPK signaling, MYC overexpression (among others) as resistance mechanisms

Resistance to PI3K inhibitors

J.G.T. Zañudo

Network modeling of breast cancer to predict drug
resistance mechanisms & combinatorial drug treatments

CRISPR screens backup **FOXO3**, **TSC1/2**, **RB1**, **P27**, **PRAS40** as putative resistance mechanisms



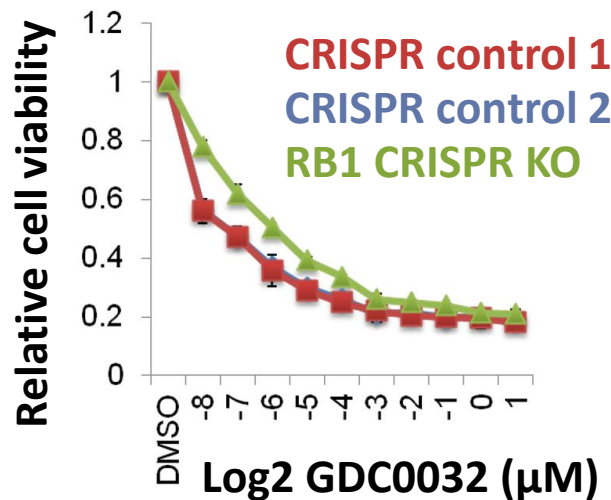
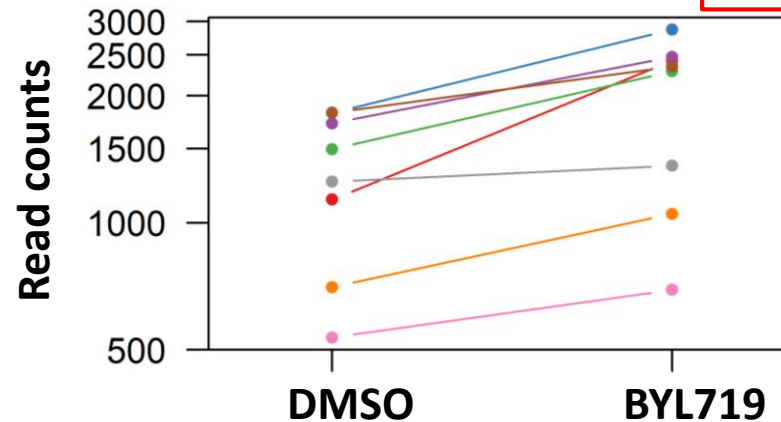
Unpublished:
From Guotai Xu
(Scaltriti, Baselga labs)

Resistance to PI3K inhibitors

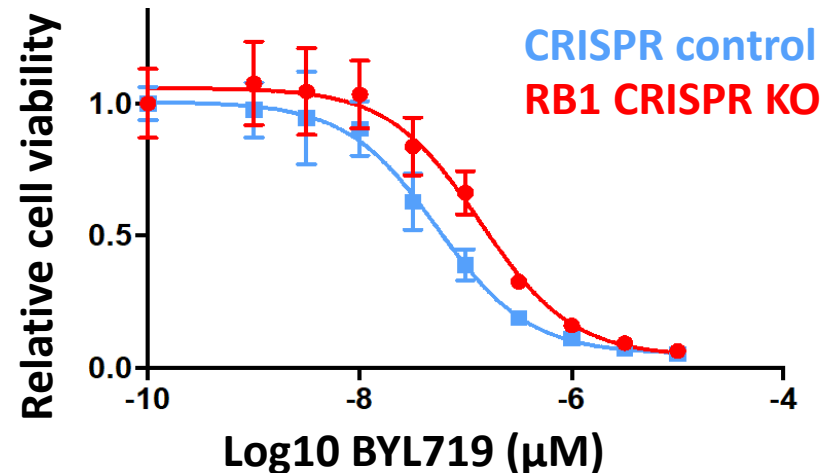
Drug response experiments confirm loss of RB1 decreases sensitivity of T47D cells to PI3K inhibitors

RB1 (from CRISPR screen) – T47D

$p < 2 \cdot 10^{-3}$
 $FDR < 10^{-1}$



Guotai Xu (Scaltriti, Baselga labs)



Seth Wanders, Gaby Johnson (Wagle lab)
Flora Luo (Garraway lab)

Resistance to PI3K inhibitors

J.G.T. Zañudo

Network modeling of breast cancer to predict drug
resistance mechanisms & combinatorial drug treatments

DANA-FARBER
CANCER INSTITUTE

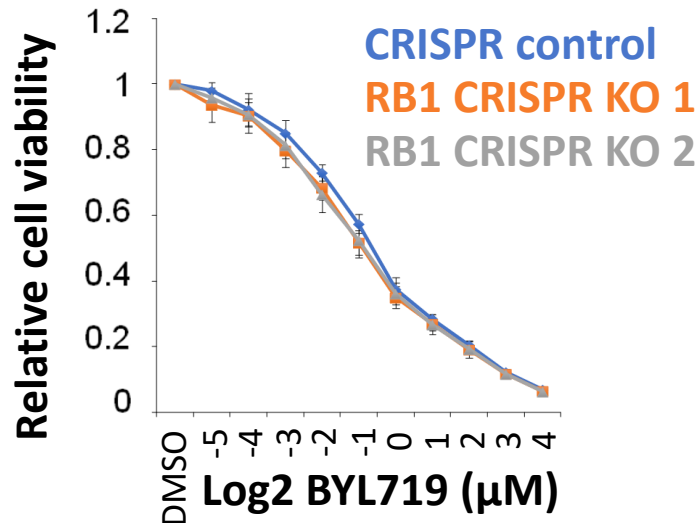
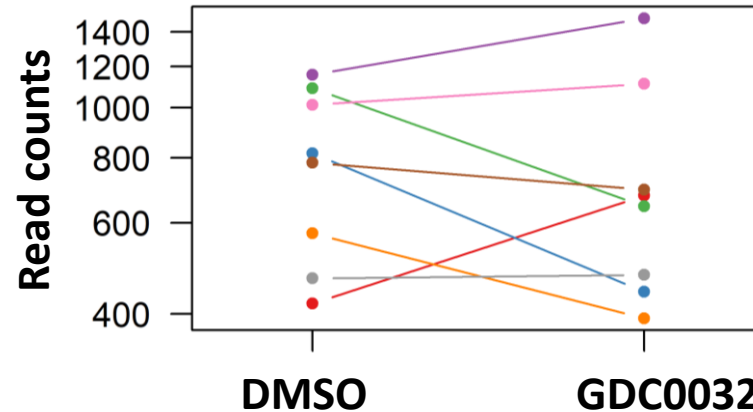
PENN STATE

BROAD
INSTITUTE

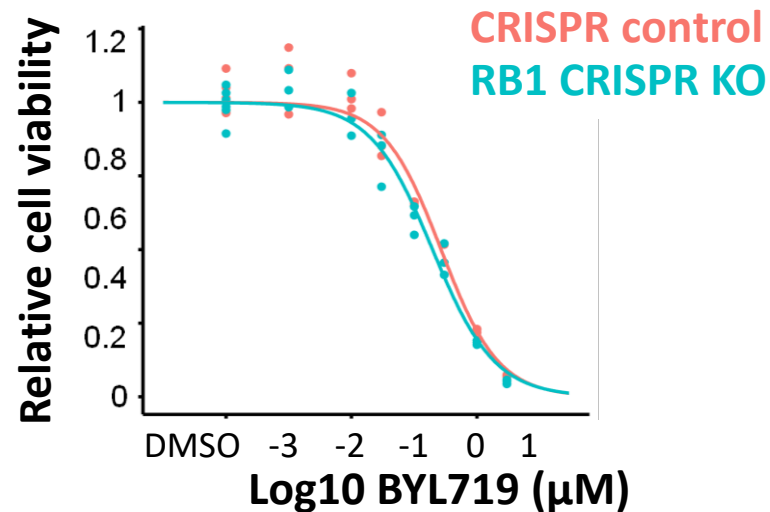
Drug response experiments confirm loss of RB1 has no effect in sensitivity of MCF7 cells to PI3K inhibitors

RB1 (from CRISPR screen) – MCF7

$p < 2 \cdot 10^{-1}$
FDR < 0.92

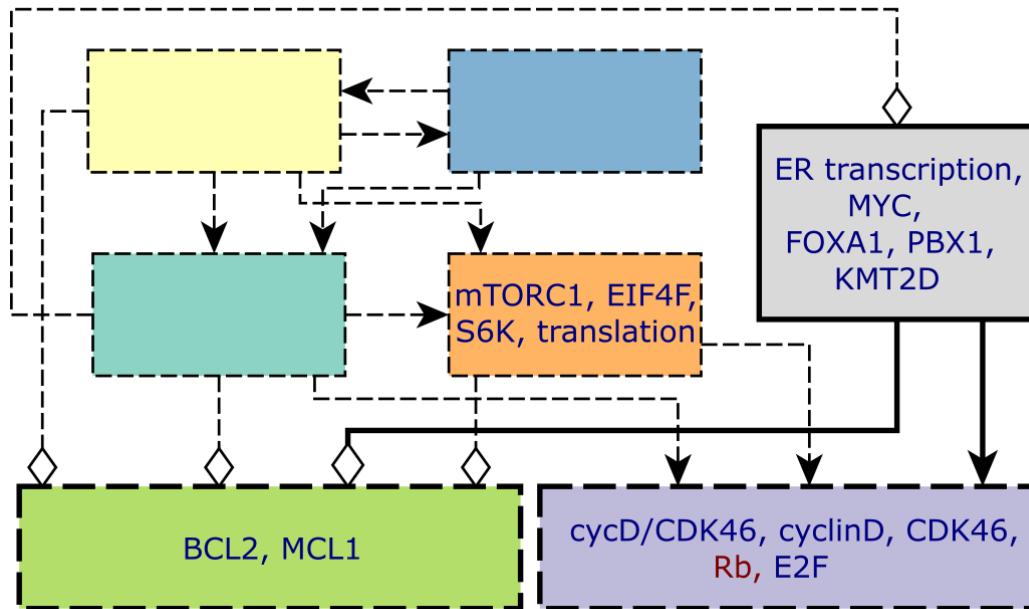


Guotai Xu (Scaltriti, Baselga labs)



Seth Wanders, Gaby Johnson (Wagle lab)
Flora Luo (Garraway lab)

Systematic search for synergistic combinations with PI3K inhibitors



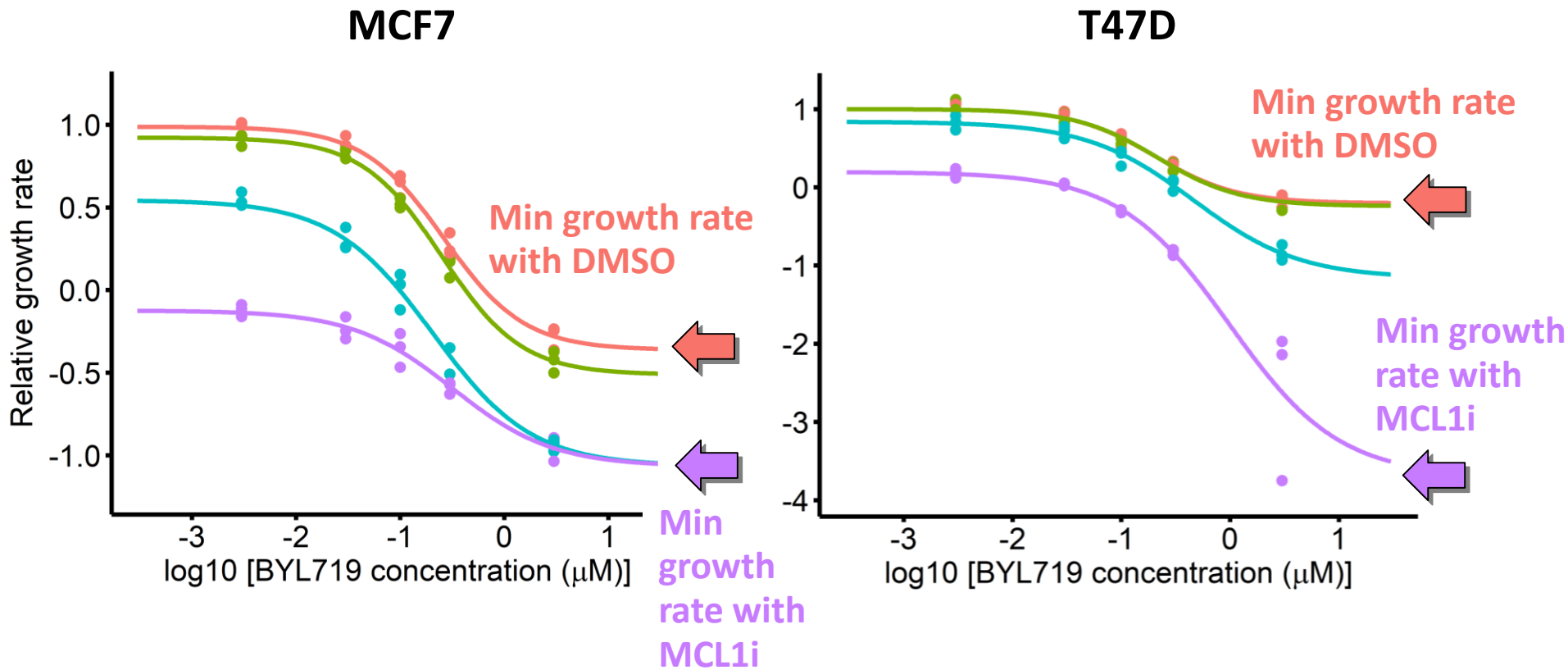
Red: Increased activity
Blue: Decreased activity

- PI3K pathway
- MAPK pathway
- AKT pathway
- mTORC1 pathway
- ER signaling
- Apoptosis
- Proliferation

**Model predicts synergy with the inhibition of anti-apoptotic proteins
MCL1 and BCL2**

Synergy with PI3K inhibitors

MCL1 inhibitor (S63845) is synergistic with PI3K inhibitor (BYL719)



MCF7	T47D
DMSO	DMSO
MCL1i (0.03 μM)	MCL1i (0.03 μM)
MCL1i (0.3 μM)	MCL1i (0.3 μM)
MCL1i (3 μM)	MCL1i (3 μM)
+ BCLXL/BCL2i (0.3 μM)	

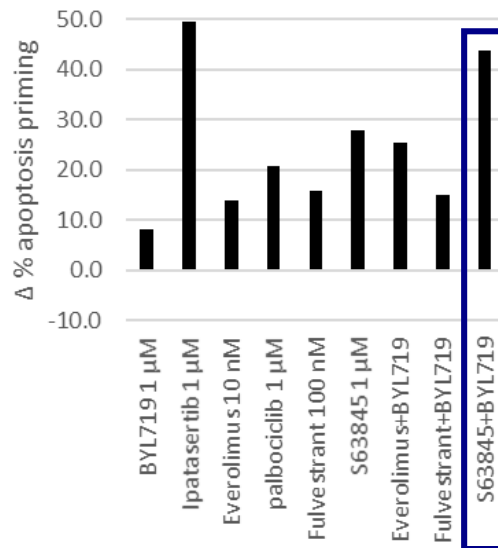
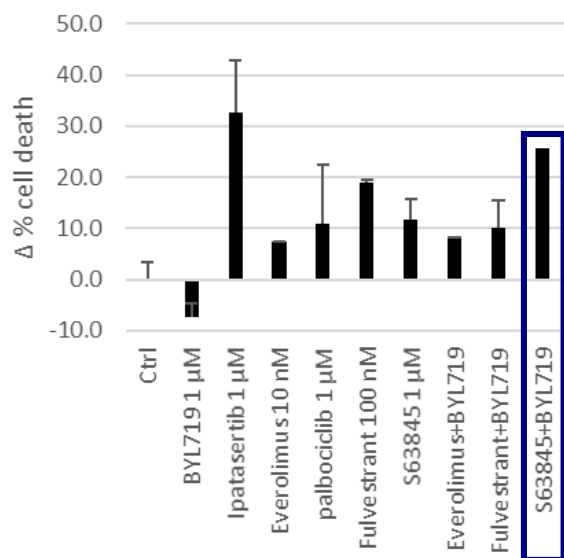
In collaboration with
Pingping Mao, Kailey
Kowalski (Wagle lab)

Synergy with PI3K inhibitors

J.G.T. Zañudo

Network modeling of breast cancer to predict drug
resistance mechanisms & combinatorial drug treatments

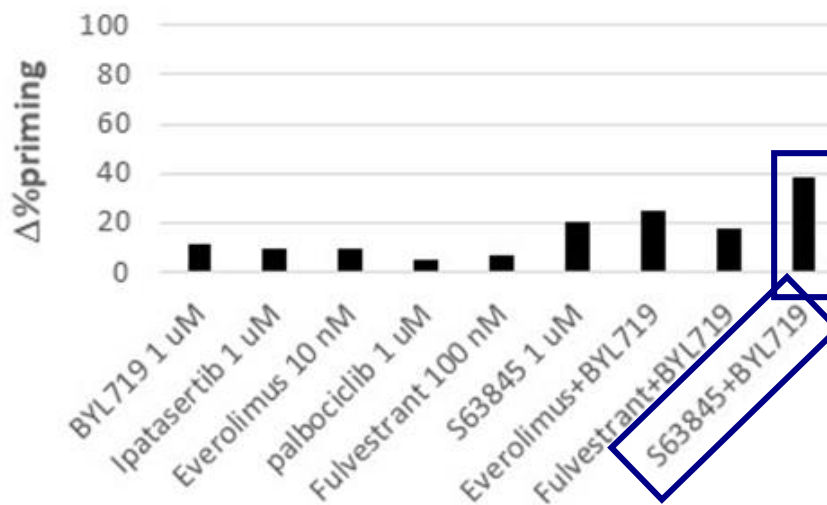
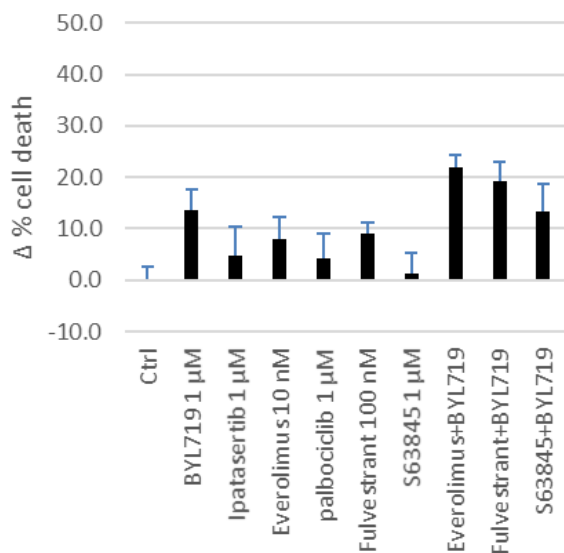
MDAMB415



MCL1 inhibitor (S63845) and PI3K inhibitors (BYL719) induce cell death in MDAMB415 and MCF7 cells

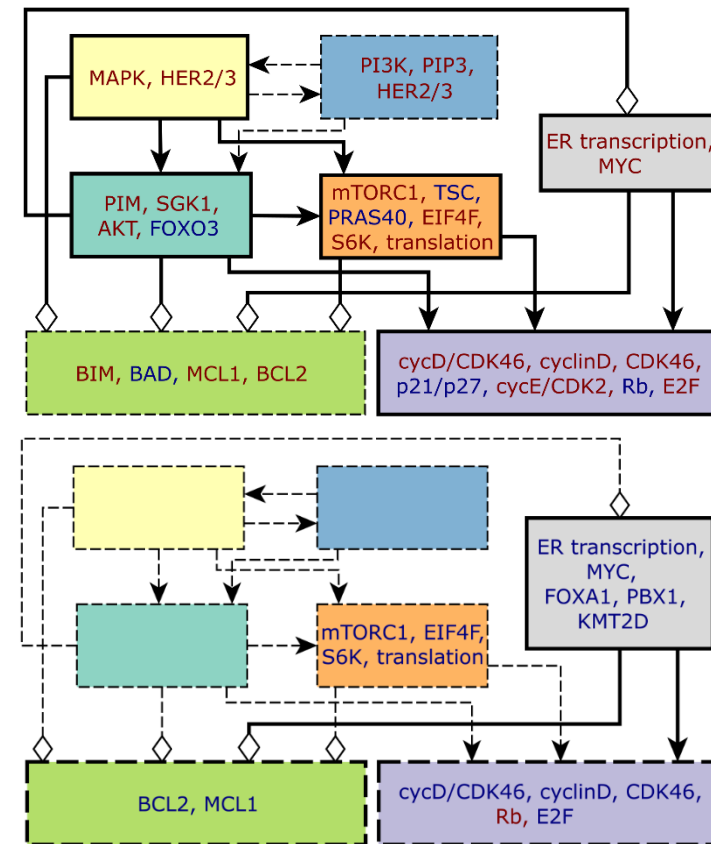
In collaboration with
Joan Montero and
the Letai lab

MCF7



Conclusions

- Constructed breast cancer network model, integrates known pathways and deregulations
- Identifies PI3Ki resistance mechanisms (**RB1**, **PRAS40**, **FOXO3**, **P27**), and predicts combinatorial drug interventions (**PI3Ki + MCL1i**)
- Several predicted resistance mechanisms were confirmed in CRISPR screens and drug-response experiments



Outlook

- Verify resistance mechanisms and combinations:
 - BH3 profiling, cell viability (Montero lab, Letai lab, Wagle lab)
 - CRISPR KOs (Wagle lab, Scaltriti lab, Baselga lab)

Acknowledgements

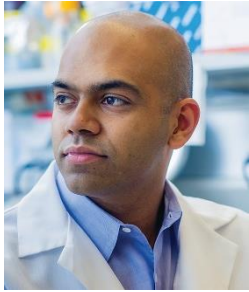
J.G.T. Zañudo
*Network modeling of breast cancer to predict drug
resistance mechanisms & combinatorial drug treatments*



ALBERT LAB WAGLE LAB



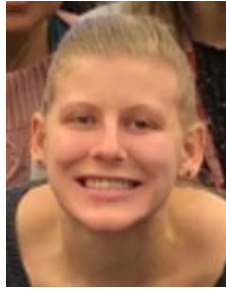
Réka Albert



Nick Wagle



Pingping Mao



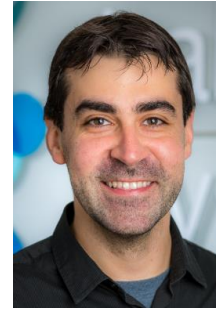
Kailey
Kowalski



Seth Wander
Gabi Johnson



Tony Letai



Joan
Montero



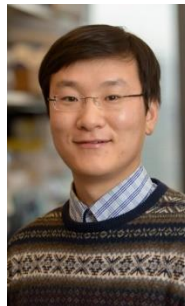
SCALTRITI LAB



Maurizio Scaltriti



Jose Baselga

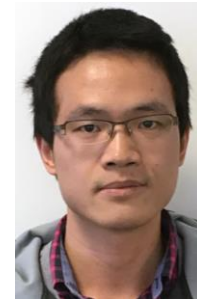


Guotai Xu

BASELGA LAB



Raul Rabadan



Junfei Zhao

RABADAN LAB



Memorial Sloan Kettering
Cancer Center

Funding from NSF and The V foundation/
SU2C Convergence Scholar Award.



Thank you for your time!

Questions, comments, suggestions?

Complaints???

Contact:

jgtz@broadinstitute.org

jgtz@phys.psu.edu

@jgtzanudo (Twitter)

Read more:

SpringerOpen blog post

*Using physics, math and models to fight cancer
drug resistance*

<http://blogs.springeropen.com/springeropen/2018/01/04/physics-math-to-fight-cancer-drug-resistance/>