

**REPORT FOR END OF YEAR ON PROGRESS ON  
FISH TRACKING SOFTWARE**

by  
Ari Spraggins

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in Partial Fulfillment of the Requirements for the Degree of  
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This thesis was prepared under the direction of the candidate's thesis advisor, Dr. Yaouen Fily, and has been approved by the members of their supervisory committee. It was submitted to the faculty of the Harriet L. Wilkes Honors College and was accepted in partial fulfillment of the requirements for the degree of Bachelor of Science in Liberal Arts and Sciences.

SUPERVISORY COMMITTEE:

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Dr. Yaouen Fily

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[second reader]

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Interim Dean Timothy Steigenga, Harriet L. Wilkes Honors College

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Date

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Some acknowledgements.

## Abstract

Author: Ari Spraggins  
Title: Report for End of Year on Progress on Fish Tracking Software  
Institution: Harriet L. Wilkes Honors College, Florida Atlantic University  
Thesis Advisor: Dr. Yaouen Fily  
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One of the major problem encountered when using the positions of fish in analysis is a tendency for the software to transpose the identities of the two fish whenever they cross. The goal of this program is to analyse that data and to remove that error from it. To do so we are comparing a metric that is unique to the fishes before and after the crossing, in this case a histogram of their brightness, and comparing them.

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# 1 Introduction

The basis of this work is going to be the videos of fish swimming in a tank, more specifically a video of 2 fish. In each frame we use software that identifies dark spots as the fish.

*Example picture of fish*



Figure 1: The two fishes

We can use this identified fish to compare how the fish has moved from frame to frame. Once we have the position of the fish in the next frame we have compare the change between two frames by using a matrix.

*Example picture of two frames*



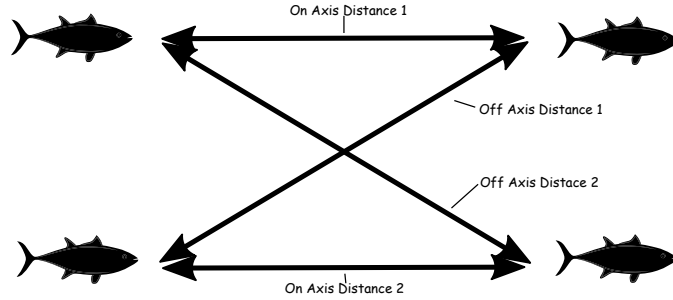


Figure 2: The distance matrix

We can extend this frame by frame comparison over the regions where the fish are visually distinct. However, in ranges where the positions of the fishes are reported as overlapping, this approach won't work for obvious reasons.

***Example picture of crossing***

Instead, during these ranges, we have to take the more involved approach of comparing a visually distinctive features of the fishes from frame. The easiest feature to compare in this case is the brightness of the fish, which we can compare via a histogram.

***Example Picture of histogram***

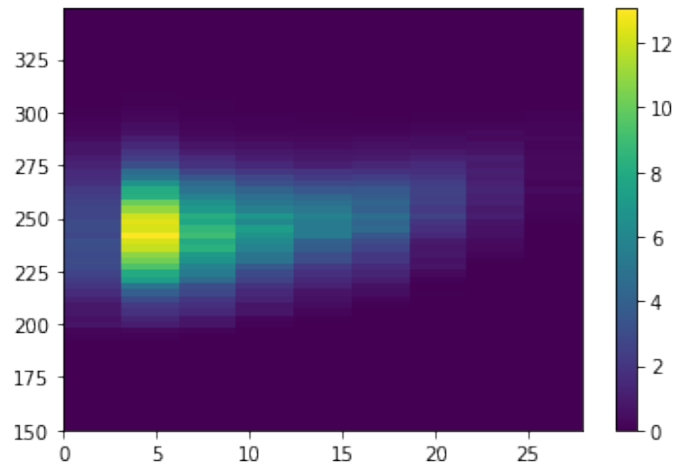


Figure 3: The histogram

## 2 Previous work

The main process by which we are computing the difference between the fish is a process laid out in the paper on idTracker[1]. The authors of the paper had a similar problem, in that they were having issues with the approach that we utilised for the initial unswap in that it both had too much error, and that the error tended to compound in on itself over the length of the tracking process. To combat this, the paper proposed a process by which each fish would be given a unique identifier, their intensity map, and comparing them frame by frame. We are emulating this by using a 2d histogram to plot their intensity maps, and using this to compare the frame data.

### 3 Methods

One of the things that quickly becomes apparent is that the fish are present as a series of dark pixels against a white background, which means that we can feed the image of a frame to a different software (in this case trilab-tracker) to generate a list of pixels that compose the fishes for tracking. This software returns the fish as either a pair of arrays for the regions where it detects two fish, and a single array where it detects one fish of the form [frame][fish][xpixels, ypixels][color]. Once we have the fish in an array form for ease of operation, we can begin performing an initial unswap by tracking the distance between the fish to determine if there was a swap, by comparing the on and off axis distances of the two possible positions of the fish. The downside of this approach is that it only works in areas in which the tracker returns that there is two fish, so we will first need to determine what regions are overlapping and nonoverlapping. Once we have these nonoverlapping regions, we can begin performing the swap check.

```

1 def swapStatus(pos,i):
2     '''
3     Detect swaps between consecutive frames based on
4     proximity.
5
6     Input:
7     pos:Postionts. Array with shape (Nframes,Nfish
8     ,Ndimensions),
9     i: Frame index. Int.
10
11     Output:
12     Int. 0 if no swaps, 1 if swapped, 2 if
13     overlapping.
14     '''
15     nFish=pos.shape[1] #Number of fish
16     distanceMatrix=[np.linalg.norm(pos[i+1][0]-pos[i
17     ][0]),
18                     np.linalg.norm(pos[i+1][1]-pos[i
19     ][1]),
20                     np.linalg.norm(pos[i+1][0]-pos[i
21     ][1]),
22                     np.linalg.norm(pos[i+1][1]-pos[i
23     ][0])]
24     swapCriteron=(distanceMatrix[0]+distanceMatrix[1])
25     -(distanceMatrix[2]+distanceMatrix[3])
26     if abs(swapCriteron)<1e-10:
27         return 2 #Overlapping
28     elif swapCriteron>0:
29         return 1 #Swapped
30     elif swapCriteron<0:
31         return 0 #Normal
32     else:
33         return -1

```

Once we have this data for the nonoverlapping ranges, we have to switch approaches for the overlapping regions. Since we can't compare the distances with the software only reporting a single fish, we are forced to use a different technique., we are using the technique of comparing the histograms of the brightness of the fishes before and after an overlapping range, as proposed by the paper on idTracker[1]. The process for this is for us to feed the arrays directly into numpy's histogram2d, which allows us to compute the histograms with a minimal amount of effort other than determining the correct bins. After that we need to manipulate the data slightly so that the histograms are taken as the average over the nonoverlapping regions for more accuracy, and are then saved out for comparison.

```

1 for i in tnrange(60, desc='nonOverlappingRange'):
2     for k in range(2):
3         countSum=0
4         countDif=0
5         pairData=[]
6         for j in range(*nonOverlappingRange[i]):
7             fishPixels = fishU[j][k]
8             m,l=np.triu_indices(fishPixels.shape[0], k
=1)
9             d=np.sqrt((fishPixels[l,0]-fishPixels[m
,0])**2+(fishPixels[l,1]-fishPixels[m,1])**2)
10            bSum=fishPixels[l,2]+fishPixels[m,2]
11            bDif=fishPixels[l,2]-fishPixels[m,2]
12
13            heightValuesSum,_,_=np.histogram2d(d,bSum,
bins=(binsDist,binsSum))
14            histSum+=heightValuesSum
15            countSum+=1
16            heightValuesDif,_,_=np.histogram2d(d,bDif,
bins=(binsDist,binsDif))
17            histDif+=heightValuesDif
18            countDif+=1
19            histSum/=countSum
20            histSumList[i,k]=histSum.copy()
21            histDif/=countDif
22            histDifList[i,k]=histDif.copy()

```

We can then feed this representation into a distance matrix to perform the final check for swaps.

## A An appendix

## B An other appendix

## References

- [1] Alfonso Pérez-Escudero et al. “idTracker: Tracking Individuals in a Group by Automatic Identification of Unmarked Animals”. In: *Nature Methods* 11.7 (July 2014), pp. 743–748. ISSN: 1548-7105. DOI: 10.1038/nmeth.2994. URL: <https://www.nature.com/articles/nmeth.2994> (visited on 06/25/2020).