

# BioVLAB-MMIA-NGS: microRNA–mRNA integrated analysis using high-throughput sequencing data

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## ABSTRACT

**Motivation:** It is now well established that microRNAs (miRNAs) play a critical role in regulating gene expression in a sequence-specific manner, and genome-wide efforts are underway to predict known and novel miRNA targets. However, the integrated miRNA–mRNA analysis remains a major computational challenge, requiring powerful informatics systems and bioinformatics expertise.

**Results:** The objective of this study was to modify our widely recognized Web server for the integrated mRNA–miRNA analysis (MMIA) and its subsequent deployment on the Amazon cloud (BioVLAB-MMIA) to be compatible with high-throughput platforms, including next-generation sequencing (NGS) data (e.g. RNA-seq). We developed a new version called the BioVLAB-MMIA-NGS, deployed on both Amazon cloud and on a high-performance publicly available server called MAHA. By using NGS data and integrating various bioinformatics tools and databases, BioVLAB-MMIA-NGS offers several advantages. First, sequencing data is more accurate than array-based methods for determining miRNA expression levels. Second, potential novel miRNAs can be detected by using various computational methods for characterizing miRNAs. Third, because miRNA-mediated gene regulation is due to hybridization of an miRNA to its target mRNA, sequencing data can be used to identify many-to-many relationship between miRNAs and target genes with high accuracy.

**Availability and implementation:** [http://epigenomics.snu.ac.kr/biovlab\\_mmia\\_ngs/](http://epigenomics.snu.ac.kr/biovlab_mmia_ngs/)

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## 1 INTRODUCTION

miRNAs are small (19–24 nt) single-stranded non-coding RNAs that regulate gene expression by specific targeting mechanism to mRNA molecules via complementary sequence pairing. Due to their critical implication in post-transcriptional regulation and impact on the developmental process, a number of (microRNA) miRNA–mRNA integrated analysis tools have been developed. MAGIA (Sales *et al.*, 2010) uses miRNA–mRNA expression profile matrices as input and provides gene set analysis and miRNA target prediction. DIANA-mirExTra (Alexiou *et al.*, 2010) accepts gene sets and computationally compares miRNA-associated motifs. miRGator (Cho *et al.*, 2013)

provides pre-compiled public resources with browser interface to navigate data. However, several limitations exist for these and other existing tools, including the following: (i) support only microarray or sequencing data but not both; (ii) require preprocessing or manual data compiling step; (iii) demand cumbersome installation procedures with interdependent tools and databases; (iv) run on limited computational resources that are not capable of handling large datasets. Here we present the next-generation sequencing (NGS) data-compatible BioVLAB-MMIA-NGS, an updated version of our array-based miRNA–mRNA integrated analysis system mRNA–miRNA analysis (MMIA) (Nam *et al.*, 2009)

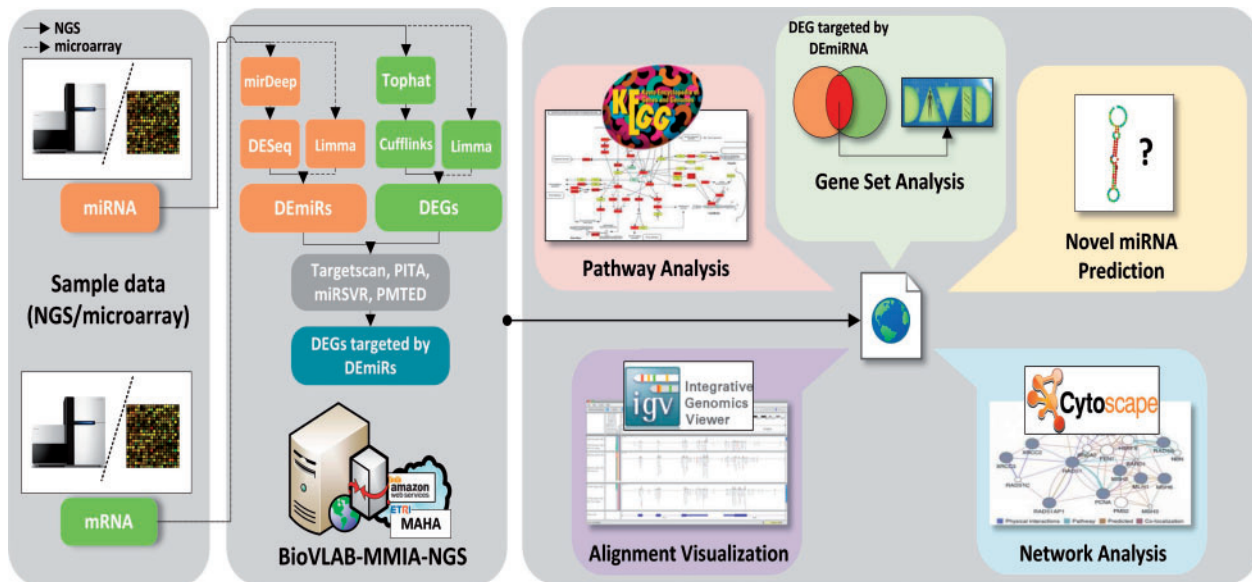
## 2 APPROACH

To perform integrated analyses between miRNA and mRNA using NGS data, we completely redesigned the MMIA Web server. BioVLAB-MMIA-NGS uses sequencing data to directly measure miRNA and mRNA expression levels on a genome scale and accurately detect changes in quantity based on read count. The system accepts raw sequencing data as input without requiring any preprocessing steps. By using RNA-seq and small RNA-seq data, not only can BioVLAB-MMIA-NGS predict novel miRNA candidates, it can also extract new information about miRNA targeting of intragenic regions, exons and introns as well as 3' UTRs, which have recently been used in the integrated analysis in plants (Meng *et al.*, 2013). Furthermore, to completely remove the burden of manually installing additional analysis tools, BioVLAB-MMIA-NGS adopts Java Web start (JAWS), a single click JAVA application deployment technology. Moreover, to support large NGS data analysis, the preprocessing and computational processes in BioVLAB-MMIA-NGS moved data to Amazon cloud and peta-scale super computing system called MAHA ([http://www.etri.re.kr/eng/res/res\\_06020402.etri](http://www.etri.re.kr/eng/res/res_06020402.etri)).

## 3 FEATURES

**Workflow:** In BioVLAB-MMIA-NGS, the integrated analysis workflow begins with extracting differentially and (or) significantly expressed miRNAs (DEmiR) as in our previous published methodology (Xin *et al.*, 2009). To identify miRNAs and their expression level, we adopted the miRDeep (Friedländer *et al.*, 2011) pipeline. The mapper module of the miRDeep package aligns raw sequencing reads to known miRNAs in miRBase

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**Fig. 1.** BioVLAB-MMIA-NGS accepts NGS/microarray data as input and extracts DEGs targeted by DEmiRs. Through the analysis pipelines, the system produces results of pathway analysis, gene set analysis, novel miRNA prediction, alignment visualization and constructed miRNA–mRNA target network

(Kozomara and Griffiths-Jones, 2010), and the quantifier module measures expression levels based on read counts. Quality control and adaptor clipping processes are performed if necessary. Significance of expressed miRNAs is tested and visualized by DESeq (Anders and Huber., 2010) based on read count measures. Differentially expressed mRNAs/genes (DEGs) are extracted by using the Tophat–Cufflinks pipeline (Trapnell *et al.*, 2012) with junction aligning based on the RPKM measure. Statistical significance is visualized by cummeRbund (Goff *et al.*, 2012). For microarray data, DEGs and DEmiRs are detected by Limma package (Smyth, 2005). Once DEGs and DEmiRs are extracted, the next step, miRNA–mRNA combined analysis, is performed. For the combined analysis, we used miRNA target prediction algorithms/databases, TargetScan (Lewis *et al.*, 2005), PITA (Kertesz *et al.*, 2007), miRSVR (Betel *et al.*, 2010) and PMTED (Sun *et al.*, 2013), as well as negative correlations between miRNAs and mRNAs, to extract DEGs targeted by DEmiRs. Gene set analysis is performed using extracted DEGs. Gene sets are automatically submitted to DAVID (Huang *et al.*, 2007) to provide functional annotation and clustering, BioCarta and KEGG (Kanehisa and Goto, 2000) pathways mapping, and disease association. Figure 1 shows BioVLAB-MMIA-NGS workflow.

**User interface:** BioVLAB-MMIA-NGS keeps the user-friendly Web-based interface for sample information, analysis options and parameters, and computing nodes. To provide extended analysis interface, we integrated IGV (Thorvaldsdóttir *et al.*, 2013) for visualizing the alignment results with zoom in/out functionality with annotation tracks and Cytoscape (Shannon *et al.*, 2003) for illustrating identified miRNA–mRNA target networks. By using JAWS, IGV and Cytoscape automatically visualize the results; manual installation and data handling processes are not required. A Web page summarizing all the results helps the users to view and further investigate the data.

**System:** The information system architecture has also adopted several important changes. The analysis begins from the Web interface, and the graphical workflow composer shows progress status. In addition, BioVLAB, the cloud infrastructure used in our previous system, has been completely rebuilt using Apache Airavata (<http://airavata.apache.org/>), generating a highly flexible and extensible three-layered BioVLAB-MMIA-NGS architecture. Moreover, BioVLAB-MMIA-NGS now supports human, mouse and rice genomes.

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## REFERENCES

- Alexiou,P. *et al.* (2010) The DIANA-mirExTra web server: from gene expression data to microRNA function. *PLoS One*, **5**, e9171.
- Anders,A. and Huber,W. (2010) Differential expression analysis for sequence count data. *Genome Biol.*, **11**, R106.
- Betel,D. *et al.* (2010) Comprehensive modeling of microRNA targets predicts functional non-conserved and non-canonical sites. *Genome Biol.*, **11**, R90.
- Cho,S. *et al.* (2013) miRGator v3.0: a microRNA portal for deep sequencing, expression profiling and mRNA targeting. *Nucleic Acids Res.*, **41**, D252–D257.
- Friedländer,M. *et al.* (2011) miRDeep2 accurately identifies known and hundreds of novel microRNA genes in seven animal clades. *Nucleic Acids Res.*, **40**, 37–52.
- Goff,L. *et al.* (2012) cummeRbund: Analysis, exploration, manipulation, and visualization of Cufflinks high-throughput sequencing data. R package version 2.4.1, <http://bioconductor.org/packages/2.11/bioc/html/cummeRbund.html>.

- Huang,D.W. *et al.* (2007) DAVID Bioinformatics Resources: expanded annotation database and novel algorithms to better extract biology from large gene lists. *Nat. Protoc.*, **35**, 169–175.
- Kanehisa,M., Goto,S. *et al.* (2000) KEGG: Kyoto Encyclopedia of Genes and Genomes. *Nucleic Acids Res.*, **28**, 27–30.
- Kertesz,M. *et al.* (2007) The role of site accessibility in microRNA target recognition. *Nat. Genet.*, **39**, 1278–1284.
- Kozomara,A. and Griffiths-Jones,S. (2010) miRBase: integrating microRNA annotation and deep-sequencing data. *Nucleic Acids Res.*, **39**, D152–D157.
- Lewis,B.P. *et al.* (2005) Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. *Cell*, **120**, 15–20.
- Meng,Y. *et al.* (2013) Introns targeted by plant microRNAs: a possible novel mechanism of gene regulation. *Rice*, **6**, 8.
- Nam,S. *et al.* (2009) MicroRNA and mRNA integrated analysis (MMIA): a web tool for examining biological functions of microRNA expression. *Nucleic Acids Res.*, **37**, W356–W362.
- Sales,G. *et al.* (2010) MAGIA, a web-based tool for miRNA and genes integrated analysis. *Nucleic Acids Res.*, **38**, W352–W359.
- Shannon,P. *et al.* (2003) Cytoscape: a software environment for integrated models of biomolecular interaction networks. *Genome Res.*, **13**, 2498–2504.
- Smyth,G.K. (2010) Limma: linear models for microarray data. In: Gentleman,R. *et al.* (ed.) *Bioinformatics and Computational Biology Solutions Using R and Bioconductor*. Springer, New York, pp. 397–420.
- Sun,X. *et al.* (2013) PMTED: a plant microRNA target expression database. *BMC Bioinformatics*, **14**, 174.
- Thorvaldsdóttir,H. *et al.* (2013) Integrative Genomics Viewer (IGV): high-performance genomics data visualization and exploration. *Bioinformatics*, **14**, 178–192.
- Trapnell,C. *et al.* (2012) Differential gene and transcript expression analysis of RNA-seq experiments with TopHat and Cufflinks. *Nat. Protoc.*, **7**, 562–578.
- Xin,F. *et al.* (2009) Computational analysis of miRNA profiles and their target genes suggests significant involvement in breast cancer antiestrogen resistance. *Bioinformatics*, **25**, 430–434.