R in NGS 实验 3

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1 实验步骤及结果

```
1 > library(systemPipeR)
    > library(systemPipeRdata)
    > setwd(choose.dir())
    > genWorkenvir(workflow = "chipseq")
    > targetsPath <- system.file("extdata", "targets_chip.txt", package =</pre>
    "systemPipeR")
    > targets <- read.delim(targetsPath, comment.char = "#")</pre>
    > targets[1:4, -c(5, 6)]
                          FileName SampleName Factor SampleLong SampleReference
 9
    1 ./data/SRR446027 1.fastq.gz
                                                  M1 Mock.1h.A
                                          M1A
10
    2 ./data/SRR446028_1.fastq.gz
                                          M1B
                                                  M1 Mock.1h.B
11
    3 ./data/SRR446029_1.fastq.gz
                                          A1A
                                                  A1 Avr.1h.A
                                                                             M1A
12
    4 ./data/SRR446030_1.fastq.gz
                                          A1B
                                                      Avr.1h.B
                                                  Α1
                                                                             M1B
13
    > dir_path <- system.file("extdata/cwl/preprocessReads/trim-se", package =</pre>
    "systemPipeR")
14
    > trim <- loadWF(targets = targetsPath, wf_file = "trim-se.cwl", input_file =</pre>
    "trim-se.yml", dir_path = dir_path)
15
    > trim <- renderWF(trim, inputvars = c(FileName = "_FASTQ_PATH1_", SampleName =
    "_SampleName_"))
16
    > output(trim)[1:2]
17
    $M1A
18
    $M1A$`trim-se`
19
    [1] "./results/M1A.fastq_trim.gz"
20
21
22
    $M1B
23
    $M1B$`trim-se`
24
    [1] "./results/M1B.fastq_trim.gz"
25
    > filterFct <- function(fq, cutoff = 20, Nexceptions = 0) {</pre>
26
          qcount <- rowSums(as(quality(fq), "matrix") <= cutoff, na.rm = TRUE)</pre>
27
          fq[qcount <= Nexceptions]</pre>
28
          # Retains reads where Phred scores are >= cutoff with N
29
          # exceptions
30
    + }
31
    > preprocessReads(args = trim, Fct = "filterFct(fq, cutoff=20, Nexceptions=0)",
32
                       batchsize = 1e+05)
33
    44022 processed reads written to file: ./results/M1A.fastq_trim.gz
34
    44927 processed reads written to file: ./results/M1B.fastq_trim.gz
35
    47793 processed reads written to file: ./results/A1A.fastq_trim.gz
36
    41201 processed reads written to file: ./results/A1B.fastq_trim.gz
37
    38549 processed reads written to file: ./results/V1A.fastq_trim.gz
```

```
38
      49362 processed reads written to file: ./results/V1B.fastq_trim.gz
  39
      58018 processed reads written to file: ./results/M6A.fastq_trim.gz
  40
      41708 processed reads written to file: ./results/M6B.fastq_trim.gz
  41
      54241 processed reads written to file: ./results/A6A.fastq_trim.gz
  42
      62722 processed reads written to file: ./results/A6B.fastq_trim.gz
  43
      52165 processed reads written to file: ./results/V6A.fastq_trim.gz
  44
      50684 processed reads written to file: ./results/V6B.fastq_trim.gz
  45
      41583 processed reads written to file: ./results/M12A.fastq_trim.gz
  46
      49316 processed reads written to file: ./results/M12B.fastq_trim.gz
  47
      56692 processed reads written to file: ./results/A12A.fastq_trim.gz
  48
      58547 processed reads written to file: ./results/A12B.fastq_trim.gz
  49
      42475 processed reads written to file: ./results/V12A.fastq_trim.gz
  50
      56732 processed reads written to file: ./results/V12B.fastq_trim.gz
  51
      > writeTargetsout(x = trim, file = "targets_chip_trim.txt", step = 1,
  52
                       new_col = "FileName", new_col_output_index = 1, overwrite =
      TRUE)
  53
           Written content of 'targetsout(x)' to file: targets_chip_trim.txt
  54
      > library(BiocParallel)
  55
      > library(batchtools)
  56
      > f <- function(x) {</pre>
  57
            targets <- system.file("extdata", "targets_chip.txt", package =</pre>
      "systemPipeR")
            dir_path <- system.file("extdata/cwl/preprocessReads/trim-se",</pre>
  58
  59
                                   package = "systemPipeR")
  60
            trim <- loadWorkflow(targets = targets, wf_file = "trim-se.cwl",</pre>
  61
                                input_file = "trim-se.yml", dir_path = dir_path)
            trim <- renderWF(trim, inputvars = c(FileName = "_FASTQ_PATH1_",</pre>
  62
  63
                                               SampleName = "_SampleName_"))
  64
            seeFastq(fastq = infile1(trim)[x], batchsize = 1e+05, klength = 8)
  65
      + }
  66
      > resources <- list(walltime = 120, ntasks = 1, ncpus = 4, memory = 1024)</pre>
      > fqlist <- lapply(seq(along = trim), f)</pre>
     > pdf("./results/fastqReport.pdf", height = 18, width = 4 * length(fqlist))
                                     矢量图, 可放大。
```

> seeFastqPlot(unlist(fqlist, recursive = FALSE))

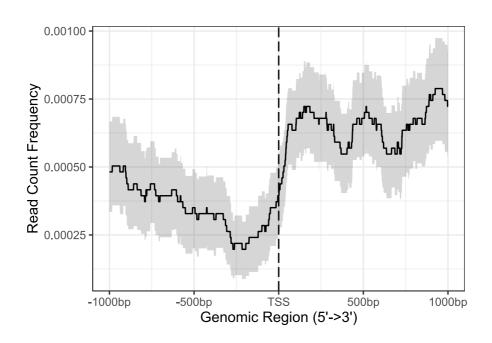
2

> dev.off()

pdf

```
4
      2
    > library(ChIPpeakAnno)
    > library(GenomicFeatures)
 7
    > dir_path <- system.file("extdata/cwl/annotate_peaks", package = "systemPipeR")</pre>
    > dir_path <- system.file("extdata/cwl/annotate_peaks", package = "systemPipeR")</pre>
    > args <- loadWF(targets = "targets_macs.txt", wf_file = "annotate_peaks.cwl",</pre>
10
                       input_file = "annotate_peaks.yml", dir_path = dir_path)
11
    > args <- renderWF(args, inputvars = c(FileName = "_FASTQ_PATH1_",</pre>
12
                                              SampleName = "_SampleName_"))
13
14
    > txdb <- makeTxDbFromGFF(file = "data/tair10.gff", format = "gff",</pre>
15
                                dataSource = "TAIR", organism = "Arabidopsis thaliana")
16
    > for (i in seq(along = args)) {
17
           peaksGR <- as(read.delim(infile1(args)[i], comment = "#"),</pre>
18
                          "GRanges")
19
           annotatedPeak <- annotatePeakInBatch(peaksGR, AnnotationData = genes(txdb))</pre>
20
           df <- data.frame(as.data.frame(annotatedPeak),</pre>
    as.data.frame(values(ge[values(annotatedPeak)$feature,
21
           ])))
22
           df$tx_type <- unlist(df$tx_type)</pre>
23
           tx_name <- c()</pre>
24
           for (j in df$tx_name){
25
               tx_name <- rbind(tx_name, j[1])</pre>
26
           }
27
           df$tx_name <- tx_name</pre>
28
           outpaths <- subsetWF(args, slot = "output", subset = 1, index = 1)</pre>
29
           write.table(df, outpaths[i], quote = FALSE, row.names = FALSE,
                       sep = "\t")
30
31
    + }
32
    > writeTargetsout(x = args, file = "targets_peakanno.txt", step = 1,
33
                       new_col = "FileName", new_col_output_index = 1, overwrite =
    TRUE)
34
    > library(ChIPseeker)
35
    > for (i in seq(along = args)) {
36
           peakAnno <- annotatePeak(infile1(args)[i], TxDb = txdb, verbose = FALSE)</pre>
37
           df <- as.data.frame(peakAnno)</pre>
38
           outpaths <- subsetWF(args, slot = "output", subset = 1, index = 1)</pre>
39
           write.table(df, outpaths[i], quote = FALSE, row.names = FALSE,
40
                       sep = "\t")
41
    + }
42
    > writeTargetsout(x = args, file = "targets_peakanno.txt", step = 1,
43
                       new_col = "FileName", new_col_output_index = 1, overwrite =
    TRUE)
44
    > library(ChIPseeker)
45
    > for (i in seq(along = args)) {
46
           peakAnno <- annotatePeak(infile1(args)[i], TxDb = txdb, verbose = FALSE)</pre>
47
           df <- as.data.frame(peakAnno)</pre>
```

```
48
          outpaths <- subsetWF(args, slot = "output", subset = 1, index = 1)</pre>
49
          write.table(df, outpaths[i], quote = FALSE, row.names = FALSE,
50
                       sep = "\t")
51
    + }
52
    > writeTargetsout(x = args, file = "targets_peakanno.txt", step = 1,
53
                      new_col = "FileName", new_col_output_index = 1, overwrite =
    TRUE)
54
         Written content of 'targetsout(x)' to file: targets_peakanno.txt
55
    > peak <- readPeakFile(infile1(args)[1])</pre>
56
    > covplot(peak, weightCol = "X.log10.pvalue.")
57
    > outpaths <- subsetWF(args, slot = "output", subset = 1, index = 1)
58
    > peakHeatmap(outpaths[1], TxDb = txdb, upstream = 1000, downstream = 1000,
59
                  palette = 'Reds')
60
    > plotAvgProf2(outpaths[1], TxDb = txdb, upstream = 1000, downstream = 1000,
    conf=0.05.
61
                   xlab = "Genomic Region (5'->3')", ylab = "Read Count Frequency")
```



```
1
   > library(GenomicRanges)
 2
    > dir_path <- system.file("extdata/cwl/count_rangesets", package = "systemPipeR")</pre>
 3
    > args <- loadWF(targets = "targets_macs.txt", wf_file = "count_rangesets.cwl",</pre>
 4
                      input_file = "count_rangesets.yml", dir_path = dir_path)
 5
    > args <- renderWF(args, inputvars = c(FileName = "_FASTQ_PATH1_",</pre>
 6
                                             SampleName = " SampleName "))
 7
    > targets <- system.file("extdata", "targets_chip.txt", package = "systemPipeR")</pre>
 8
    > dir_path <- system.file("extdata/cwl/bowtie2", package = "systemPipeR")</pre>
 9
    > args_bam <- loadWF(targets = targets, wf_file = "bowtie2-mapping-se.cwl",</pre>
10
                           input_file = "bowtie2-mapping-se.yml", dir_path = dir_path)
11
    > args_bam <- renderWF(args_bam, inputvars = c(FileName = "_FASTQ_PATH1_",
12
                                                      SampleName = "_SampleName_"))
13
    > args_bam <- output_update(args_bam, dir = FALSE, replace = TRUE,
14
                                  extension = c(".sam", ".bam"))
```

```
15
    > outpaths <- subsetWF(args_bam, slot = "output", subset = 1, index = 1)</pre>
16
    > bfl <- BamFileList(outpaths, yieldSize = 50000, index = character())</pre>
17
    > # countDFnames <- countRangeset(bfl, args, mode = "Union", ignore.strand =
    TRUE) # skipped
18
    > # writeTargetsout(x = args, file = "targets_countDF.txt", step = 1, new_col =
    "FileName", new col output index = 1, overwrite = TRUE) # skipped
19
    > library(GenomicRanges)
20
    > dir_path <- system.file("extdata/cwl/count_rangesets", package = "systemPipeR")</pre>
21
    > args <- loadWF(targets = "targets_macs.txt", wf_file = "count_rangesets.cwl",</pre>
22
                      input_file = "count_rangesets.yml", dir_path = dir_path)
23
    > args <- renderWF(args, inputvars = c(FileName = "_FASTQ_PATH1_",</pre>
24
                                             SampleName = "_SampleName_"))
25
26
    > ## Bam Files
27
    > targets <- system.file("extdata", "targets_chip.txt", package = "systemPipeR")</pre>
28
    > dir_path <- system.file("extdata/cwl/bowtie2", package = "systemPipeR")</pre>
29
    > args_bam <- loadWF(targets = targets, wf_file = "bowtie2-mapping-se.cwl",</pre>
30
                          input_file = "bowtie2-mapping-se.yml", dir_path = dir_path)
31
    > args_bam <- renderWF(args_bam, inputvars = c(FileName = "_FASTQ_PATH1_",
32
                                                     SampleName = "_SampleName_"))
33
    > args_bam <- output_update(args_bam, dir = FALSE, replace = TRUE,
34
                                 extension = c(".sam", ".bam"))
35
    > outpaths <- subsetWF(args_bam, slot = "output", subset = 1, index = 1)</pre>
36
    > bfl <- BamFileList(outpaths, yieldSize = 50000, index = character())</pre>
37
    > countDFnames <- countRangeset(bfl, args, mode = "Union", ignore.strand = TRUE)
38
    > dir_path <- system.file("extdata/cwl/rundiff", package = "systemPipeR")</pre>
39
    > args_diff <- loadWF(targets = "targets_countDF.txt", wf_file = "rundiff.cwl",</pre>
40
                           input_file = "rundiff.yml", dir_path = dir_path)
41
    > args_diff <- renderWF(args_diff, inputvars = c(FileName = "_FASTQ_PATH1_",
42
                                                       SampleName = "_SampleName_"))
43
44
    > cmp <- readComp(file = args_bam, format = "matrix")</pre>
45
    > dbrlist <- runDiff(args = args_diff, diffFct = run_edgeR, targets =</pre>
    targets.as.df(targets(args_bam)),
46
                          cmp = cmp[[1]], independent = TRUE, dbrfilter = c(Fold = 2,
47
                                                                               FDR = 1)
48
    Disp = 0.25073 , BCV = 0.5007
49
    Disp = 0.15223 , BCV = 0.3902
50
    Disp = 0.19792 , BCV = 0.4449
    Disp = 0.11692 , BCV = 0.3419
51
52
    Disp = 0.09286 , BCV = 0.3047
53
    Disp = 0.14312 , BCV = 0.3783
54
    Disp = 0.17049 , BCV = 0.4129
55
    Disp = 0.09671 , BCV = 0.311
56
    Disp = 0.14805 , BCV = 0.3848
57
    Wrote count result 1 to M1A_peaks.edgeR.xls
58
    Saved plot 1 to M1A_peaks.edgeR.xls.pdf
```

```
59
     Disp = 0.23843 , BCV = 0.4883
 60
     Disp = 0.11547, BCV = 0.3398
 61
     Disp = 0.21655 , BCV = 0.4653
 62
     Disp = 0.04269 , BCV = 0.2066
 63
     Disp = 0.15216 , BCV = 0.3901
     Disp = 0.12235 , BCV = 0.3498
 64
 65
     Disp = 0.20886 , BCV = 0.457
 66
     Disp = 0.12322 , BCV = 0.351
 67
     Disp = 0.20853 , BCV = 0.4567
 68
     Wrote count result 2 to A1A_peaks.edgeR.xls
 69
     Saved plot 2 to A1A_peaks.edgeR.xls.pdf
 70
     Disp = 0.10458 , BCV = 0.3234
 71
     Disp = 0.13358 , BCV = 0.3655
 72
     Disp = 0.09085 , BCV = 0.3014
 73
     Disp = 0.06552 , BCV = 0.256
     Disp = 0.13023 , BCV = 0.3609
 74
 75
     Disp = 0.1174 , BCV = 0.3426
 76
     Disp = 0.09355 , BCV = 0.3059
 77
     Disp = 0.08648 , BCV = 0.2941
 78
     Disp = 0.0827 , BCV = 0.2876
 79
     Wrote count result 3 to V1A_peaks.edgeR.xls
 80
     Saved plot 3 to V1A_peaks.edgeR.xls.pdf
 81
     Disp = 0.19174 , BCV = 0.4379
 82
     Disp = 0.12306 , BCV = 0.3508
 83
     Disp = 0.15138 , BCV = 0.3891
 84
     Disp = 0.13047 , BCV = 0.3612
 85
     Disp = 0.09703 , BCV = 0.3115
 86
     Disp = 0.16206 , BCV = 0.4026
 87
     Disp = 0.17529 , BCV = 0.4187
 88
     Disp = 0.1089 , BCV = 0.33
 89
     Disp = 0.15879 , BCV = 0.3985
 90
     Wrote count result 4 to M6A_peaks.edgeR.xls
 91
     Saved plot 4 to M6A peaks.edgeR.xls.pdf
 92
     Disp = 0.10874 , BCV = 0.3298
 93
     Disp = 0.17742 , BCV = 0.4212
 94
     Disp = 0.12824 , BCV = 0.3581
 95
     Disp = 0.24666 , BCV = 0.4967
 96
     Disp = 0.10608 , BCV = 0.3257
 97
     Disp = 0.20981 , BCV = 0.4581
 98
     Disp = 0.23416 , BCV = 0.4839
 99
     Disp = 0.10848 , BCV = 0.3294
100
     Disp = 0.21298 , BCV = 0.4615
101
     Wrote count result 5 to A6A_peaks.edgeR.xls
102
     Saved plot 5 to A6A peaks.edgeR.xls.pdf
103
     Disp = 0.12458 , BCV = 0.353
104
     Disp = 0.12455 , BCV = 0.3529
105
     Disp = 0.1462 , BCV = 0.3824
```

```
106
     Disp = 0.0498 , BCV = 0.2232
107
     Disp = 0.05905 , BCV = 0.243
108
     Disp = 0.06298 , BCV = 0.251
109
     Disp = 0.10963 , BCV = 0.3311
110
     Disp = 0.09929 , BCV = 0.3151
     Disp = 0.10645 , BCV = 0.3263
111
112
     Wrote count result 6 to V6A_peaks.edgeR.xls
113
     Saved plot 6 to V6A_peaks.edgeR.xls.pdf
114
     Disp = 0.19246 , BCV = 0.4387
115
     Disp = 0.10992 , BCV = 0.3315
116
     Disp = 0.15488 , BCV = 0.3935
117
     Disp = 0.12168 , BCV = 0.3488
118
     Disp = 0.09003 , BCV = 0.3
119
     Disp = 0.14578 , BCV = 0.3818
120
     Disp = 0.16995 , BCV = 0.4122
     Disp = 0.09506 , BCV = 0.3083
121
122
     Disp = 0.15563 , BCV = 0.3945
123
     Wrote count result 7 to M12A_peaks.edgeR.xls
124
     Saved plot 7 to M12A_peaks.edgeR.xls.pdf
125
     Disp = 0.29779 , BCV = 0.5457
126
     Disp = 0.16126 , BCV = 0.4016
127
     Disp = 0.26423 , BCV = 0.514
128
     Disp = 0.28419 , BCV = 0.5331
129
     Disp = 0.11531 , BCV = 0.3396
130
     Disp = 0.28049 , BCV = 0.5296
131
     Disp = 0.27675 , BCV = 0.5261
132
     Disp = 0.13993 , BCV = 0.3741
133
     Disp = 0.25 , BCV = 0.5
134
     Wrote count result 8 to A12A_peaks.edgeR.xls
135
     Saved plot 8 to A12A_peaks.edgeR.xls.pdf
136
     Disp = 0.22277 , BCV = 0.472
137
     Disp = 0.19447, BCV = 0.441
138
     Disp = 0.17158 , BCV = 0.4142
139
     Disp = 0.23746 , BCV = 0.4873
140
     Disp = 0.14669 , BCV = 0.383
141
     Disp = 0.24076 , BCV = 0.4907
142
     Disp = 0.20954 , BCV = 0.4578
143
     Disp = 0.11004 , BCV = 0.3317
144
     Disp = 0.21206 , BCV = 0.4605
145
     Wrote count result 9 to V12A_peaks.edgeR.xls
146
     Saved plot 9 to V12A_peaks.edgeR.xls.pdf
147
     > writeTargetsout(x = args_diff, file = "targets_rundiff.txt",
148
                       step = 1, new_col = "FileName", new_col_output_index = 1,
149
                       overwrite = TRUE)
150
          Written content of 'targetsout(x)' to file: targets_rundiff.txt
151
     > writeTargetsout(x = args_diff, file = "targets_rundiff.txt",
152
                       step = 1, new_col = "FileName", new_col_output_index = 1,
```

```
153 +
                        overwrite = TRUE)
154
           Written content of 'targetsout(x)' to file: targets_rundiff.txt
155
     > dir_path <- system.file("extdata/cwl/annotate_peaks", package = "systemPipeR")</pre>
156
     > args <- loadWF(targets = "targets_bam_ref.txt", wf_file = "annotate_peaks.cwl",</pre>
157
                       input_file = "annotate_peaks.yml", dir_path = dir_path)
158
     > args <- renderWF(args, inputvars = c(FileName1 = " FASTQ PATH1 ",</pre>
159
                                              SampleName = " SampleName "))
160
161
     > args_anno <- loadWF(targets = "targets_macs.txt", wf_file =</pre>
      "annotate_peaks.cwl",
162
                             input_file = "annotate_peaks.yml", dir_path = dir_path)
163
     > args_anno <- renderWF(args_anno, inputvars = c(FileName = "_FASTQ_PATH1_",
164
                                                         SampleName = "_SampleName_"))
165
     > annofiles <- subsetWF(args_anno, slot = "output", subset = 1,
166
                              index = 1
167
     > gene_ids <- sapply(names(annofiles), function(x)</pre>
     unique(as.character(read.delim(annofiles[x])[,
168
                    "geneId"])), simplify = FALSE)
169
     > load("data/GO/catdb.RData")
170
     > BatchResult <- GOCluster_Report(catdb = catdb, setlist = gene_ids,
171
                                         method = "all", id_type = "gene", CLSZ = 2,
     cutoff = 0.9,
172
                                         gocats = c("MF", "BP", "CC"), recordSpecGO =
     NULL)
173
     > library(Biostrings)
174
     > library(seqLogo)
175
     > library(BCRANK)
176
     > dir_path <- system.file("extdata/cwl/annotate_peaks", package = "systemPipeR")</pre>
177
     > args <- loadWF(targets = "targets_macs.txt", wf_file = "annotate_peaks.cwl",</pre>
178
                       input_file = "annotate_peaks.yml", dir_path = dir_path)
179
     > args <- renderWF(args, inputvars = c(FileName = "_FASTQ_PATH1_",</pre>
180
                                              SampleName = " SampleName "))
181
182
     > rangefiles <- infile1(args)</pre>
183
     > for (i in seq(along = rangefiles)) {
184
            df <- read.delim(rangefiles[i], comment = "#")</pre>
185
            peaks <- as(df, "GRanges")</pre>
186
            names(peaks) <- pasteO(as.character(seqnames(peaks)), "_",</pre>
187
                                    start(peaks), "-", end(peaks))
188 +
            peaks <- peaks[order(values(peaks)$X.log10.pvalue., decreasing = TRUE)]</pre>
189
            pseq <- getSeq(FaFile("./data/tair10.fasta"), peaks)</pre>
190
            names(pseq) <- names(peaks)</pre>
191
            writeXStringSet(pseq, paste0(rangefiles[i], ".fasta"))
192
     + }
193
     > set.seed(0)
194
     > BCRANKout <- bcrank(paste0(rangefiles[1], ".fasta"), restarts = 25,
```

```
195 +
                         use.P1 = TRUE, use.P2 = TRUE)
196
     > toptable(BCRANKout)
197
          Consensus
                      Score
198
          AGTCAHTT 87.08268
199
          ATGTNAGA 78.86206
200 3 CACAHDBAAM 78.43056
201 4
          MGGTATC 77.63990
202 5 AMRCABAAR 69.68058
203
     6 AWAARBCAA 69.00625
204 7 DCCDDGAAAS 62.46803
205 8 RYNTGNCTCT 61.84035
206 9 DHGABWGGAA 61.19199
207 10 TGNSHTTCHT 60.77090
208
     11 GHTGABNTTM 60.46803
209
     12 GCHGHTVTVT 58.78979
210
     13 CABKBTGBHA 58.76985
211 | 14 AAHTBHCTVC 57.98167
212 | 15 DDBCGBCCDT 57.18527
213
    16 TTVGMNAGWTC 56.98050
214 | 17 GATTKGBNGAA 55.79763
215
     18 DDHCNGHCTTG 52.71424
216 | 19 AKAGAHAAGC 52.56690
217
     20 TBRTAKCTNC 52.18433
218
     21 ACGAWNTBRK 51.60559
219
     22 VWCGTVNDVTT 50.63590
220
     23 CVDGDTCAVVC 50.49847
221
     24 GAHNAMASAC 49.08524
222
     25 CCBASGYNDG 46.22657
223
     > topMotif <- toptable(BCRANKout, 1)</pre>
224
     > weightMatrix <- pwm(topMotif, normalize = FALSE)</pre>
225
     > weightMatrixNormalized <- pwm(topMotif, normalize = TRUE)</pre>
226
     > pdf("results/seqlogo.pdf")
227
     > seqLogo(weightMatrixNormalized)
228
     > dev.off()
229
     RStudioGD
230
             2
```

