

Celera Genome Browser

User Guide

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Open Source Release 5.0

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Table of Contents

[Preface](#)

About This Guide	viii
Product Release Notes	viii
Conventions Used in this Guide	ix
Finding Information in this Guide	x
Technical Support	xi

[1.0 Genome Browser Overview](#)

[Introduction 1.2](#)

XML Features	1.3
Specify Assembly Files	1.3
Import Feature Files	1.3
Annotate and Save Workspace Files	1.4
System Requirements	1.5
Troubleshooting Performance Issues	1.5

[2.0 Getting Started](#)

Accessing the Genome Browser	2.2
About the Genome Browser Main Window	2.3
Compacting Memory	2.5
Status Messages from the Genome Browser	2.5
Setting Preferences	2.6
Setting Viewing Preferences	2.6
Creating a Preferences File	2.7
Deleting a Preferences File	2.8
Specifying a User- or Site-Specific Preference File	2.9
Modifying Genomic Axis Annotation View	2.10
Settings	

<u>Specifying SubView Preferences</u>	<u>2.12</u>
<u>Editing Alignment View Settings</u>	<u>2.12</u>
<u>Editing Consensus Sequence View Settings</u>	<u>2.13</u>
<u>Editing Transcript Translation View Settings</u>	<u>2.14</u>
<u>Specifying System Preferences</u>	<u>2.15</u>
<u>Specifying the Data Source Settings</u>	<u>2.16</u>
<u>Setting Login Information</u>	<u>2.17</u>
<u>Adding XML Directories</u>	<u>2.18</u>
<u>Specifying XML Service URLs</u>	<u>2.19</u>
<u>Setting Splice Profiles</u>	<u>2.20</u>
<u>Setting Workspace Backup Options</u>	<u>2.21</u>

[3.0 Genome Browser Data Overview](#)

<u>Genome Browser Data Classes</u>	<u>3.2</u>
<u>Assembly 3.2</u>	
<u>Features 3.2</u>	
<u>Precomputes</u>	<u>3.2</u>
<u>Annotations</u>	<u>3.3</u>
<u>Celera Exchange (XML) Documents</u>	<u>3.4</u>
<u>XML Assembly Files</u>	<u>3.4</u>
<u>XML Feature Files</u>	<u>3.4</u>
<u>XML Workspace Files</u>	<u>3.5</u>

[4.0 Accessing and Viewing Data](#)

<u>Accessing the Data</u>	<u>4.3</u>
<u>Searching for a Specific ID</u>	<u>4.3</u>
<u>Searching for Sequence Data</u>	<u>4.5</u>
<u>Searching for an XML Feature ID</u>	<u>4.6</u>

<u>Opening a Genome Version</u>	<u>4.6</u>
<u>Drilling Down to a Genomic Axis</u>	<u>4.7</u>
<u>Opening an XML Workspace (.gbw) File</u>	<u>4.9</u>
<u>Viewing an XML Workspace or Feature File</u>	<u>4.9</u>
<u>Navigating to a Feature in an XML Workspace File</u>	<u>4.10</u>
<u>Displaying Open Data Sources</u>	<u>4.11</u>
<u>Loading Precompute and Annotation Features</u>	<u>4.12</u>
<u>Showing or Hiding Data Tiers</u>	<u>4.14</u>
<u>Reverse Complementing the Axis</u>	<u>4.15</u>
<u>Displaying a Feature's Properties</u>	<u>4.17</u>
<u>Zooming to a Selected Feature</u>	<u>4.17</u>
<u>Highlighting Evidence</u>	<u>4.18</u>
<u>Showing Edge Matches</u>	<u>4.18</u>
<u>Showing Evidence Edge Mismatches</u>	<u>4.19</u>
<u>Showing non-GT/AG Splices Matches</u>	<u>4.19</u>
<u>Finding a Feature's Matching Subject Sequence</u>	<u>4.20</u>
<u>Displaying Consensus Sequence</u>	<u>4.20</u>
<u>Displaying the Translations or Reverse Complement</u>	<u>4.23</u>
<u>Copying Sequence to the Clipboard</u>	<u>4.24</u>
<u>Setting Color Rules For Feature Display</u>	<u>4.25</u>
<u>Creating a New Color Rule</u>	<u>4.25</u>
<u>Editing an Existing Color Rule</u>	<u>4.28</u>
<u>Deleting an Existing Color Rule</u>	<u>4.30</u>
<u>Applying a Color Rule</u>	<u>4.30</u>
<u>Resetting the View</u>	<u>4.30</u>
<u>Using Property Rules</u>	<u>4.31</u>
<u>Using a Property Rule to Create Links to</u>	
<u>HTML BioMolecule Reports</u>	<u>4.33</u>
<u>Displaying Predicted Features in the Transcript Translation View</u>	<u>4.35</u>
<u>Changing the Translations Display</u>	<u>4.36</u>

<u>Copying Transcript Sequence to the Clipboard</u>	<u>4.36</u>
<u>Evaluating Feature Homologies</u>	<u>4.37</u>
<u>Displaying the Feature Report</u>	<u>4.38</u>
<u>Running BLAST Searches</u>	<u>4.40</u>
<u>Searching for a Nucleotide or Protein Sequence String</u>	<u>4.41</u>
<u>Using Bookmarks</u>	<u>4.42</u>
<u>Creating</u>	<u>4.42</u>
<u>Opening a Bookmark</u>	<u>4.42</u>
<u>Editing or Deleting a Bookmark</u>	<u>4.42</u>
<u>Displaying Multiple Browser Windows</u>	<u>4.43</u>
<u>Closing a Browser Window</u>	<u>4.43</u>
<u>Printing a Page</u>	<u>4.43</u>

5.0 Creating and Modifying Annotation Data

<u>Introduction</u>	<u>5.2</u>
<u>Edit Commands</u>	<u>5.2</u>
<u>Annotation History</u>	<u>5.3</u>
<u>Creating a Transcript or Exon Using a Computed Feature</u>	<u>5.4</u>
<u>Reviewing and Editing Comments</u>	<u>5.5</u>
<u>Adding Evidence to a Transcript or Exon</u>	<u>5.7</u>
<u>Removing Evidence from a Transcript or Exon</u>	<u>5.7</u>
<u>Splitting a Transcript</u>	<u>5.7</u>
<u>Merging Transcripts</u>	<u>5.8</u>
<u>Adjusting Exon Boundaries</u>	<u>5.9</u>
<u>Using the Consensus Sequence Window to Modify an Exon</u>	<u>5.9</u>
<u>Using Evidence to Modify an Exon</u>	<u>5.10</u>
<u>Using a Splice Site to Modify an Exon</u>	<u>5.10</u>
<u>Adding or Deleting Bases in the SubView</u>	<u>5.11</u>

<u>Finding a Long Open Reading Frame</u>	<u>5.11</u>
<u>Evaluating Subject Sequence Alignments</u>	<u>5.12</u>
<u>Deleting a Feature from the Workspace</u>	<u>5.12</u>
<u>Exporting Annotation Data</u>	<u>5.13</u>

[Appendix A Exchange File Format](#)

<u>Introduction A.2</u>	
<u>XML Markup Element Descriptions</u>	<u>A.3</u>
<u>Document Type Definition</u>	<u>A.10</u>
<u>Example Feature File</u>	<u>A.13</u>

[Appendix B Troubleshooting](#)

<u>Troubleshooting</u>	<u>B.2</u>
<u>Memory Problems</u>	<u>B.2</u>
<u>Data Access Problems</u>	<u>B.2</u>
<u>Exception Errors</u>	<u>B.3</u>

P r e f a c e

<u>About This Guide</u>	<u>viii</u>
<u>Product Release Notes</u>	<u>viii</u>
<u>Conventions Used in this Guide</u>	<u>ix</u>
<u>Finding Information in this Guide</u>	<u>x</u>
<u>Technical Support</u>	<u>xi</u>

ABOUT THIS GUIDE

Welcome to Celera's Genome Browser. This guide describes the Genome Browser's features and functions, and provides step-by-step instructions for using the product.

As new features are added to the Genome Browser, we will update the user guide accordingly. This guide is available as an Acrobat PDF from the Genome Browser help menu. Select Help|User Manual from the Main Menu.

This guide was developed for new Genome Browser users.

Product Release Notes

The Genome Browser release notes describe new features, known issues with the product, and additional helpful information. To access the release notes, select Help|Release Notes from the Genome Browser Main Menu.

CONVENTIONS USED IN THIS GUIDE

This guide uses the following typefaces and symbols for presentation purposes:

Bold Text Indicates text that you should type. Brackets surround user-specific text. For example, [**search string**] means that you should type the text for which to search. You do not type the brackets. For short text strings, we also use quotation marks to surround the text you type. You do not type the quotation marks.

Italic text Indicates words that require special emphasis.

NOTE Highlights important information or warnings.

TIP Describes shortcuts.

FINDING INFORMATION IN THIS GUIDE

The following table describes where to find information in this guide:

Chapter 1.0 Genome Browser Overview	Describes the features and functions of the Genome Browser. Also describes the system requirements.
Chapter 2.0 Getting Started	Explains how to access the Genome Browser and describes the Main Window. Explains how to set preferences.
Chapter 3.0 Genome Browser Data Overview	Describes the Celera data that appears in the Genome Browser. Also describes the Celera Exchange (XML) documents.
Chapter 4.0 Accessing and Viewing Data	Explains how to use the Genome Browser to access and view genomic information.
Chapter 5.0 Creating and Modifying Annotation Data	Explains how to use the Genome Browser to create, modify, and export annotation data.
Appendix A Exchange File Format	Describes the Celera Exchange File format that you use to create XML files for importing into the Genome Browser. Includes the Document Type Definition.
Appendix B Troubleshooting	Explains how to troubleshoot problems you may experience with the Genome Browser.

TECHNICAL SUPPORT

Currently, there is no technical support for the open source release.

<u>Introduction</u>	<u>1.2</u>
<u>XML Features</u>	<u>1.3</u>
<u>Specify Assembly Files</u>	<u>1.3</u>
<u>Import Feature Files</u>	<u>1.3</u>
<u>Annotate and Save Workspace Files</u>	<u>1.4</u>
<u>System Requirements</u>	<u>1.5</u>
<u>Troubleshooting Performance Issues</u>	<u>1.5</u>

INTRODUCTION

Genome Browser is a visualization tool that allows you to:

- graphically view Celera genome sequence information and annotation data
- view multiple genome versions at the same time
- overlay and view your mapped feature data with Celera data
- create and save annotation

The Genome Browser is designed to support a fully functional 3-tiered application consisting of:

- an interactive GUI client front-end
- an application server in the middle tier
- a relational database back-end

Currently, the open source release only supports the stand-alone single-tier client functionality. Future versions may provide a standard data server or access to industry standard data protocols. This would allow the user to open the Genome Browser client and then connect it to a variety of different data servers, as well as open local feature files and temporary workspace files for viewing and annotating data on top of the server assembly data.

NOTE: All file import and export functionality takes place on the client side of the application, whether or not the Genome Browser is connected to an application server.

The current, stand-alone release allows you to load genomic data directly into the client using the Celera version of the GAME XML file format known as the Celera Exchange File Format.

XML Features

The Celera Exchange (XML) File Format allows you to import and export data from the Genome Browser in a flexible manner that supports a variety of different workflows and data sources. Using the Celera Exchange File Format with the Genome Browser, you can:

- Specify assembly files
- Import feature files
- Annotate and save workspace files

The following sections provide more detail about these features.

Specify Assembly Files

See [Appendix A](#) for the detailed Celera Exchange File Format Specification and Document Type Definition. Also, see [page 3-4](#) for more details about the XML assembly files.

For 3-tier operation, assembly data is stored in a database, which you access via data server.

However, the open-source release only supports stand-alone operation, so assembly Sequence information must be specified in an assembly file, usually in the form of a FASTA file.

See [page 3-4](#) for more details about XML feature files.

Import Feature Files

You can import either precomputed or human-annotated features as multiple feature files directly into the Genome Browser client as long as they are specified in relation to an assembly Genomic Axis. Since the feature files do not contain any Genomic Axes themselves – only references to them – you may partition feature data among files in any convenient manner. Multiple feature files may contain data on the same Genomic Axis, or a single feature file may contain data on multiple Genomic Axes.

Annotate and Save Workspace Files

See page [3-5](#) for more details about XML workspace files.

The Genome Browser performs all interactive annotation operations in local memory in a data structure called the workspace. You can export the annotation data in the workspace (modified and newly created genes, transcripts, exons, start codons, and evidence relationships) to a single workspace (.gbw) file, which you can reload at a later time to continue annotating.

If you wish to commit your annotation data to your database, you can then feed the workspace file into a promotion utility (not provided by the open-source Genome Browser release).

SYSTEM REQUIREMENTS

Before you access the program, ensure your machine has the following installed:

- Java Runtime Environment (JRE) 1.3.0_
- At least 256 MB RAM
- Adequate virtual memory settings

<u>Accessing the Genome Browser</u>	<u>2.2</u>
<u>About the Genome Browser Main Window</u>	<u>2.3</u>
<u>Compacting Memory</u>	<u>2.5</u>
<u>Status Messages from the Genome Browser</u>	<u>2.5</u>
<u>Setting Preferences</u>	<u>2.6</u>
<u>Setting Viewing Preferences</u>	<u>2.6</u>
<u>Creating a Preferences File</u>	<u>2.7</u>
<u>Deleting a Preferences File</u>	<u>2.8</u>
<u>Specifying a User- or Site-Specific Preference File</u>	<u>2.9</u>
<u>Modifying Genomic Axis Annotation View</u>	<u>2.10</u>
<u>Settings</u>	<u>2.12</u>
<u>Specifying SubView Preferences</u>	<u>2.12</u>
<u>Editing Alignment View Settings</u>	<u>2.13</u>
<u>Editing Consensus Sequence View Settings</u>	<u>2.14</u>
<u>Editing Transcript Translation View Settings</u>	<u>2.15</u>
<u>Specifying System Preferences</u>	<u>2.16</u>
<u>Specifying the Data Source Settings</u>	<u>2.17</u>
<u>Setting Login Information</u>	<u>2.18</u>
<u>Adding XML Directories</u>	<u>2.19</u>
<u>Specifying XML Service URLs</u>	<u>2.20</u>
<u>Setting Splice Profiles</u>	<u>2.21</u>
<u>Setting Workspace Backup Options</u>	

ACCESSING THE GENOME BROWSER

To access the Genome Browser:

- 1 Make sure that a Java virtual machine is installed on your computer
- 2 From your desktop, double-click the Genome Browser icon.

The Genome Browser Main Window appears:

Main Menu

Outline View

Property Inspector
View

Genome Axis Annotation View

Left Frame

Right Frame

Memory Meter

NOTE: The Genome Browser enables only those menu and sub-menu items that are valid in a given context. If an item is greyed out, it is unavailable at that time.

When you access the Genome Browser, the window is the same size and layout and in the same location as when you last closed the application.

About the Genome Browser Main Window

The following table describes the contents of the Genome Browser Main Window and how to navigate within the window.

Genome Browser Main Window

Main Menu Displays the menu options that are available to you. The menu options vary, depending on the data you have open. This guide describes the available options in greater detail when explaining how to perform tasks.

Left Frame



Contains two elements:

- Outline View
- Property Inspector View

If the Outline View contains more data than it can display, Genome Browser inserts vertical and horizontal scroll bars so that you can see the data.

To resize the frame and its elements:

- 1 Place your cursor over a border until the cursor changes to double-arrow.
- 2 Left-click your mouse and drag the frame to the size you want.

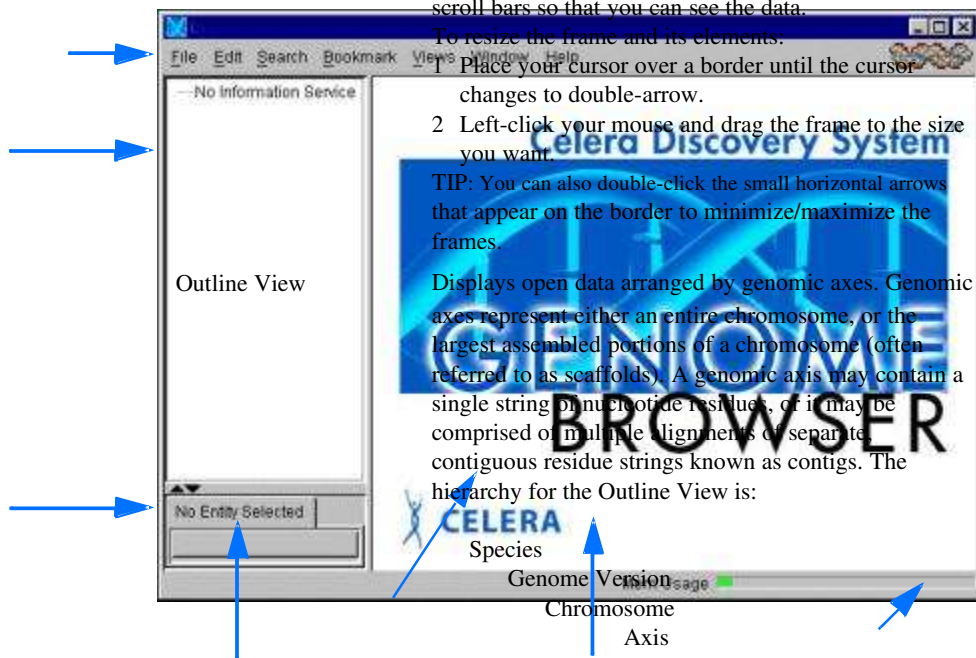
TIP: You can also double-click the small horizontal arrows that appear on the border to minimize/maximize the frames.

Displays open data arranged by genomic axes. Genomic axes represent either an entire chromosome, or the largest assembled portions of a chromosome (often referred to as scaffolds). A genomic axis may contain a single string of nucleotide residues, or it may be comprised of multiple alignments of separate contiguous residue strings known as contigs. The hierarchy for the Outline View is:

Species
Genome Version
Chromosome
Axis

- Click the + sign to expand a folder or folder name.
- Click the - sign to collapse a folder or folder name.

NOTE: When you first open the Genome Browser, the Outline View displays “No Information Service.”



Genome Browser Main Window (cont'd)

Property Inspector View	<p>Displays the properties associated with the entity currently highlighted in the outline view or in the annotation view.</p> <p>NOTE: When you first open the Genome Browser, or if you do not have an object selected, the Property Inspector View displays “No Entity Selected.”</p>
Right Frame	<p>Contains the Genomic Axis Annotation View.</p> <p>If you enable the SubView view, it will appear in the bottom portion of the right frame area.</p> <p>You can resize this frame, too.</p>
Memory Usage Meter	<p>Roughly indicates the amount of available memory while you are using the application. The meter color changes to a yellow line if 70% of available memory is used and a red line if 90% of available memory is used. Depending upon what the application is doing, this meter may fluctuate.</p> <p>To find out exactly how much memory the Genome Browser client is using:</p> <ol style="list-style-type: none">1 Double-click the memory meter. <p>The Memory Usage dialog box appears, which indicates:</p> <p>Total Memory — the total memory that the Genome Browser requires, up to the maximum it is allowed to consume.</p> <p>Free Memory — how much memory remains available for the Genome Browser client to use.</p> <p>Used Memory — how much memory the Genome Browser is consuming at the moment.</p>

Compacting Memory

- 1 Double-click the Memory Usage Meter in the lower right of the Genome Browser Main Window.

The Memory Usage dialog box appears:

- 2 Click Yes.

Status Messages from the Genome Browser

NOTE: You will receive this message only if you have the Genome Browser open and are connected to a Celera database.

If the Genome Browser has downtime scheduled, Celera will broadcast a Status Message with the appropriate information across the server to the Genome Browser client.

SETTING PREFERENCES

Before you begin using the Genome Browser, you should review and set or change, if necessary, the Genome Browser Preferences.

Setting Viewing Preferences

The following sections explain how to:

- Create a preferences file and specify the location for group preference files
- Modify the Genomic Axis Annotation View Settings

Creating a Preferences File

You may save changes you make to the View settings in a file to use for yourself or to share with a group of users.

- 1 From the Main Menu, select Edit|Preferences|Genomic Axis Annotation View.
- 2 Click the View Preference File tab.

- 3 Click New File.

The Create New View Setting File dialog box appears.

- 4 Type a name in the text box.
- 5 Click OK.

The new preference becomes the default.

Deleting a Preferences File

To remove a preferences file:

- 1 From the Main Menu, select Edit|Preferences|Genomic Axis Annotation View.
- 2 Click the View Preference File tab.
The Preferences: Genomic Axis Annotation View dialog box appears.
- 3 Click Delete File.
- 4 In the Delete File dialog box, select the file you want to delete and click OK.

The Deleting file dialog box appears.

- 5 Click Yes to delete the file or No to retain it.

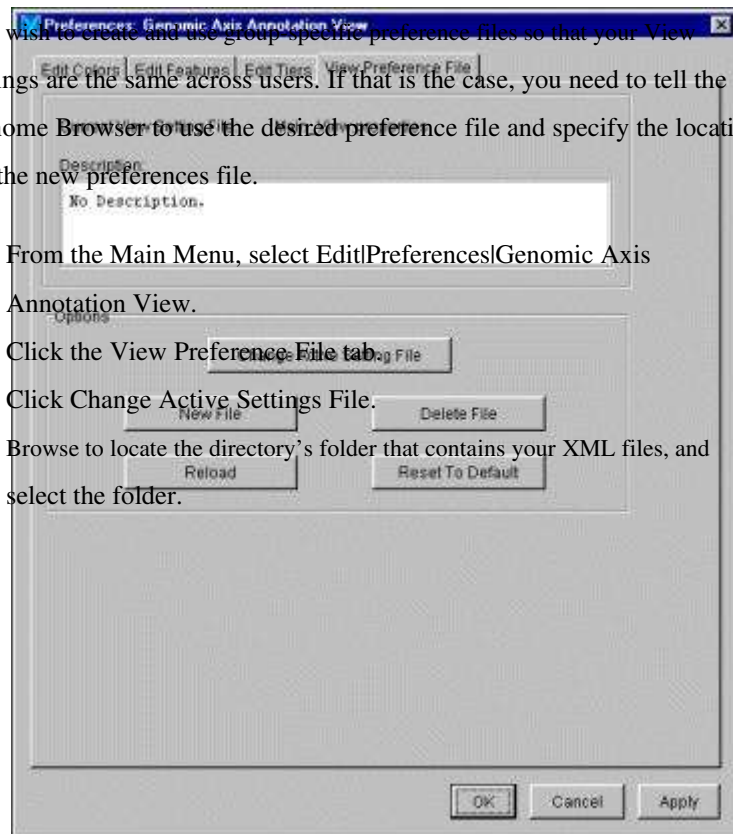
To remove any changes (e.g., color choices) from the current session, click Reload.

To reset to default, click Reset to Default.

Specifying a User- or Site-Specific Preference File

You may wish to specify a preference file other than the default, or your site may wish to create and use group-specific preference files so that your View settings are the same across users. If that is the case, you need to tell the Genome Browser to use the desired preference file and specify the location for the new preferences file.

- 1 From the Main Menu, select Edit|Preferences|Genomic Axis Annotation View.
- 2 Click the View Preference File tab.
- 3 Click Change Active Settings File.
- 4 Browse to locate the directory's folder that contains your XML files, and select the folder.



- 5 Click Select File.


Modifying Genomic Axis Annotation View Settings

The Genome Browser provides a variety of color schemes to help you view genomic data. You modify default settings in the Preferences: Genomic Axis Annotation View dialog box.

- 1 From the Main Menu, select Edit|Preferences|Genomic Axis Annotation View.

The Preferences: Genomic Axis Annotation View dialog box appears.

The following table describes how to edit colors, features, and tiers using

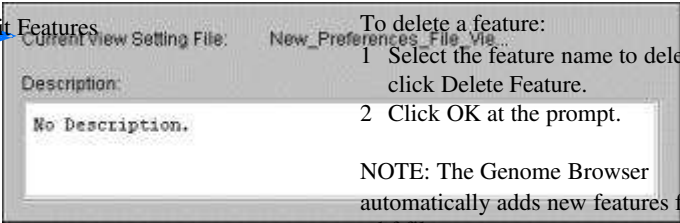
this dialog box:


To add a color:
 1 Click Add Color.
 2 Select a color and click OK.
 The new color appears in the list.
 3 Click on the new color's row in the Color Name column and type a name for the new color.

To delete a color:

- 1 Select the color name to delete, and click Delete Color.
- 2 Click OK at the prompt.

Edit Features



To delete a feature:
 1 Select the feature name to delete, and click Delete Feature.
 2 Click OK at the prompt.

NOTE: The Genome Browser automatically adds new features from a .gbf file.

Edit Tiers

To reorder how the tiers appear in the Genomic Axis Annotation View:

- 1 Click in the Position column for the tier(s) you wish to reorder.
- 2 Specify the new position(s) for the tier(s).
- 3 Press Enter to save your change.

To delete a tier:

- 1 Select the tier name to delete, and click Delete Tier.
- 2 Click OK at the prompt.

NOTE: The Genome Browser automatically adds tier names that appear in XML feature (.gbf) files if the feature group name is new.

Specifying Sub View Preferences

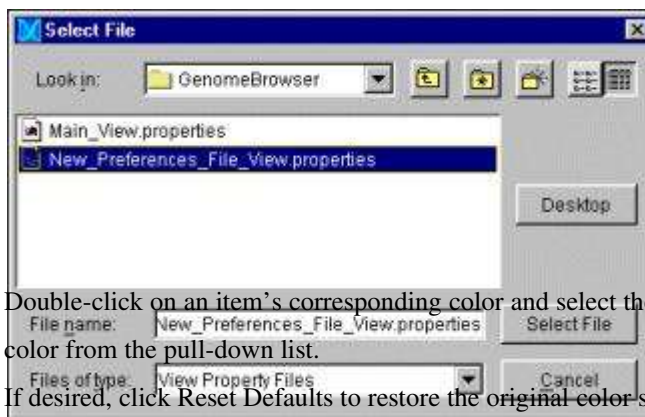
The following sections explain how to:

- Edit Alignment View Settings
- Edit Consensus Sequence View Settings
- Edit Transcript Translation View Settings

Editing Alignment View Settings

You can change the Alignment View settings to accommodate your color preferences.

- 1 From the Main Menu, select Edit|Preferences|SubViews.
- 2 Click the Edit Query Alignment View Settings tab:



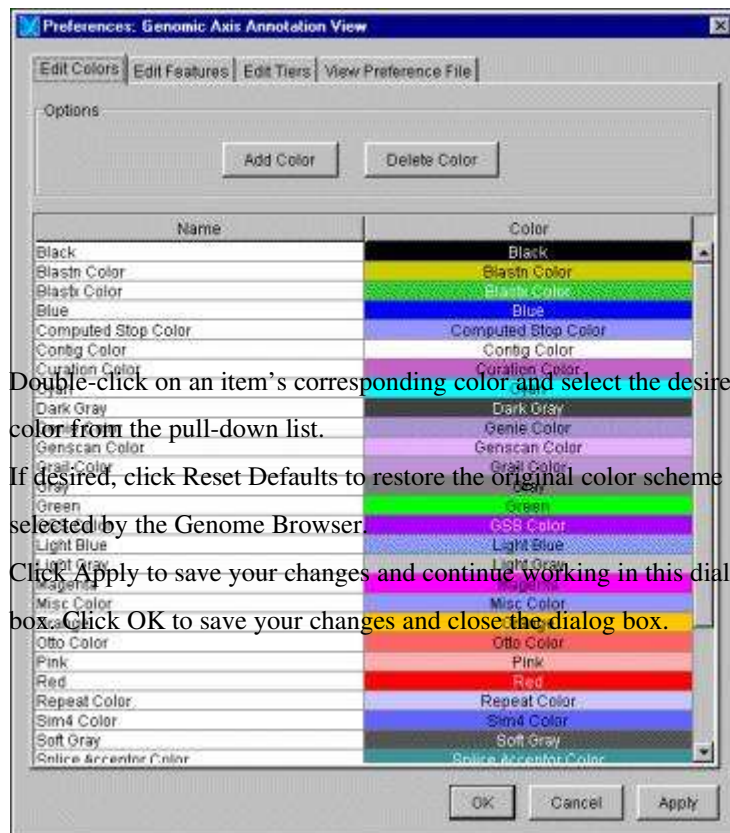
- 3 Double-click on an item's corresponding color and select the desired color from the pull-down list.
- 4 If desired, click Reset Defaults to restore the original color scheme selected by the Genome Browser.
- 5 Click Apply to save your changes and continue working in this dialog box. Click OK to save your changes and close the dialog box.

Editing Consensus Sequence View Settings

You can change the Consensus Sequence View settings to accommodate your color preferences.

- 1 From the Main Menu, select Edit|Preferences|SubViews.
- 2 Click Edit Consensus Sequence View Settings.

- 3 Double-click on an item's corresponding color and select the desired color from the pull-down list.
- 4 If desired, click Reset Defaults to restore the original color scheme selected by the Genome Browser.
- 5 Click Apply to save your changes and continue working in this dialog box. Click OK to save your changes and close the dialog box.



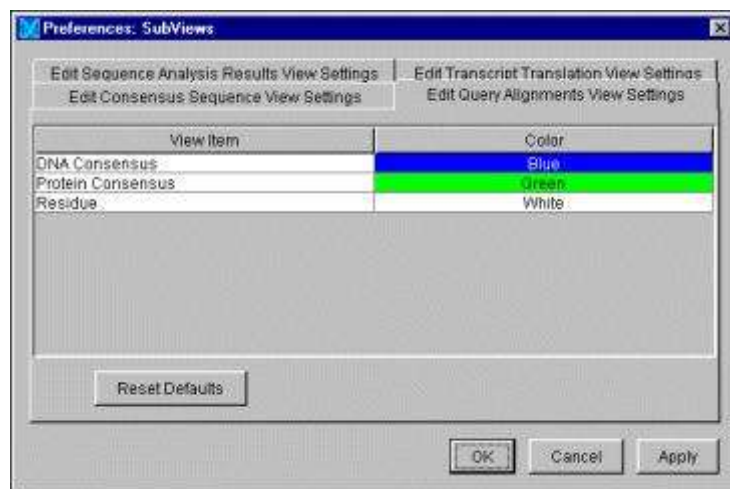
Editing Transcript Translation View Settings

You can change the Transcript Translation View settings to accomodate your color preferences.

- 1 From the Main Menu, click Edit|Preferences|SubViews.
 - 2 Click Edit Transcript Translation View Settings:
-
- 3 Double-click on an item's corresponding color and select the desired color from the pull-down list.
 - 4 If desired, click Reset Defaults to restore the original color scheme selected by the Genome Browser.
 - 5 Click Apply to save your changes and continue working in this dialog box. Click OK to save your changes and close the dialog box.

Specifying System Preferences

- 1 From the Main Menu, select Edit|Preferences|System .
- 2 Click the Applications Settings tab:



- 3 Select the desired options. Use the following table to guide you through your choices.

Option	Description
Display SubViews When Available	Check this box to display the available SubView windows. These windows are context-sensitive: only the relevant windows display, depending upon what you have selected in the Genomic Axis Annotation View.
Focus SubViews Upon Navigation	Check this box to automatically define the SubView range when you navigate to a feature from search results. This will also automatically activate the Consensus Sequence panel.
Display Memory Usage Meter	Check this box to display the Memory Usage Meter in the lower-right corner. See page 2.4 for more information about the Memory Usage Meter.
Show Navigation/Search complete messages	Check this box to show complete messages when searching for data.
Look and Feel Options	Click a radio button to change the overall appearance of the Genome Browser's windows and dialog boxes: <ul style="list-style-type: none">• Metal• CDE/Motif• Windows (default)

Specifying the Data Source Settings

You must specify the data source settings before you can access any data in the Genome Browser. Depending on the data you will access, you may do one or more of the following:

- Set login information
- Specify XML directories
- Specify XML Service URLs

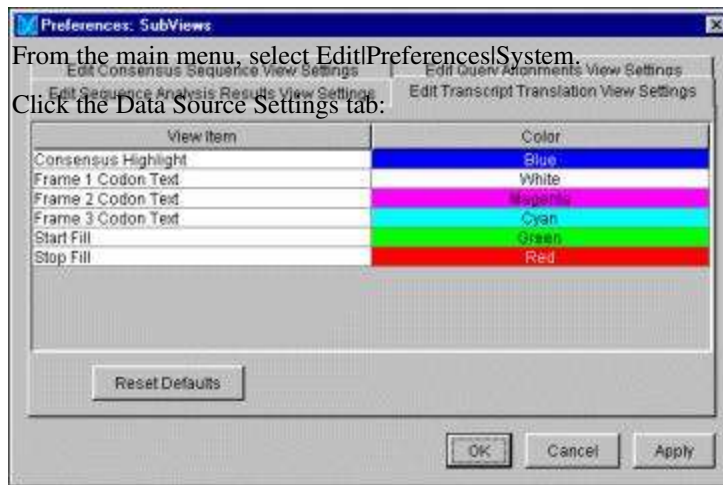
Setting Login Information

NOTE: If you are not connecting to a Celera database, you do not need to login.

You will need to set your login information before you may connect to a database. You do this from the Preferences: System dialog box.

NOTE: When you select File/Open Genome, the Preferences dialog box appears automatically if you have never set your login information.

- 1 From the main menu, select Edit/Preferences/System.
- 2 Click the Data Source Settings tab:



- 3 In the User Name text box, type your database user name.
 - 4 Type your database Password in the text box.
 - 5 If desired, select the "Save Login Information" box.
- If you do not check this box, you will need to login each time you connect to a database.

- 6 Click Apply to save your changes and continue working in this dialog box. Click OK to save your changes and close the dialog box.

Adding XML Directories

Before you may open an XML assembly or load an XML feature file's data, you need to tell the Genome Browser where to look for the files.

- 1 From the Main Menu, select Edit|Preferences|System .
- 2 Click the Data Source Settings tab.
- 3 Click Add to Current Directories.

The Select Directory dialog box appears:

- 4 Browse to locate the directory's folder that contains your XML files, and select the folder.
- 5 Click Select Directory.
- 6 Repeat steps 3 through 5 for each directory that you wish to add.
- 7 If desired, specify a validation option.
- 8 Click Apply to save your changes and continue working in this dialog box. Click OK to save your changes and close the dialog box.

NOTE: To delete an XML directory, select the directory to delete and click Remove Selected Directory.

Specifying XML Service URLs

NOTE: For more information about the Genome Browser XML Service, refer to the [Celera Programmer Reference Guide for the Genome Browser](#).

NOTE: To delete a URL, select the URL you want to delete in the Current URLs box, and click Remove Selected URL.

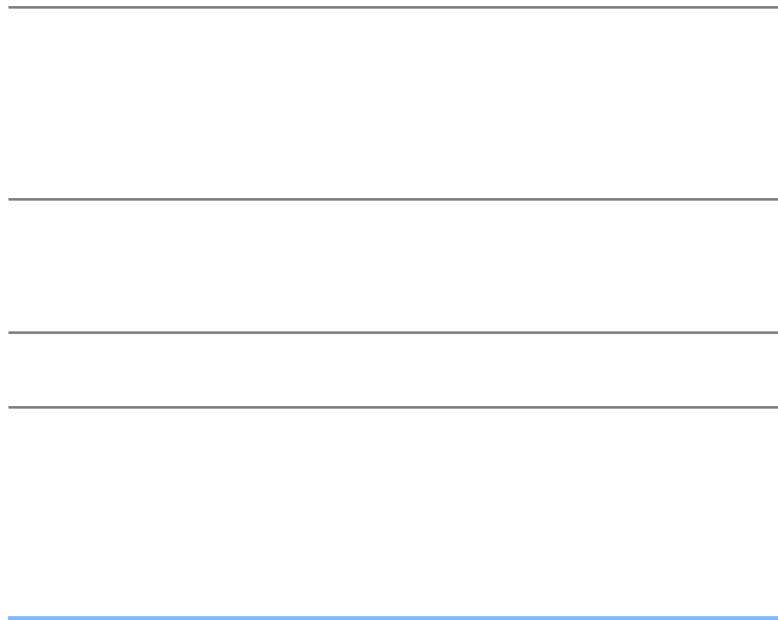
If your site takes advantage of the Genome Browser's XML Service capability, you should specify the URLs in the Preferences dialog box.

- 1 From the Main Menu, select Edit|Preferences|System .
- 2 Click the Data Source Settings tab.
- 3 Type the URL provided by your site programmer in the New URL text box, and click Add to Current URLs.
- 4 Click Apply to save your changes and continue working in this dialog box. Click OK to save your changes and close the dialog box.

Setting Splice Profiles

Celera provides splice profiles for Human, Mouse and Drosophila data. You can override these default splice profiles and specify different “Acceptor,” “Donor,” and “Neither” splice profile files you have stored on your computer.

- 1 From the Main Menu, select Edit|Preferences|System .
- 2 Click the Splice Profiles tab.



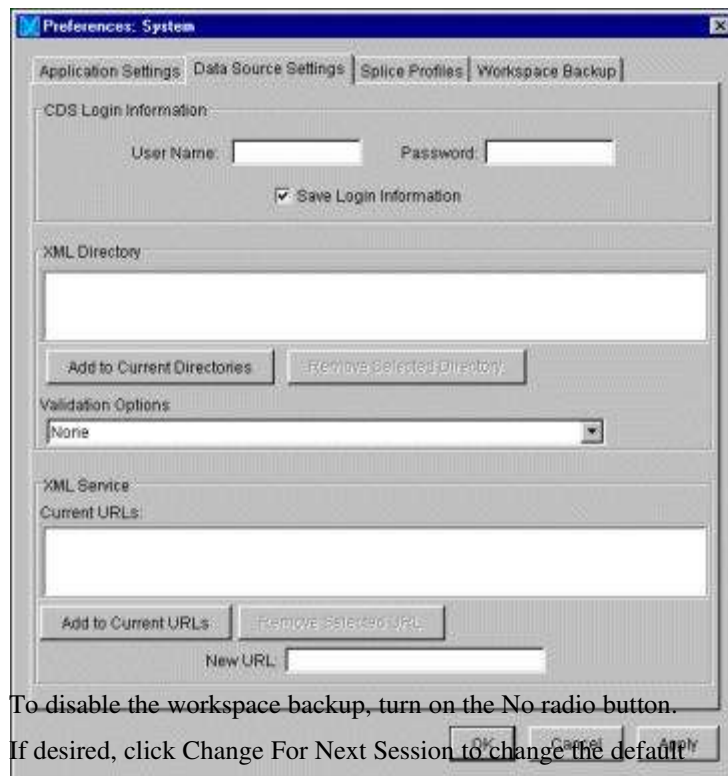
If desired, check the Override Default Splice Profiles box. This will activate the Choose buttons that allow you to upload Donor, Acceptor and Neither splice files.

- 3 Click Apply to save your changes and continue working in this dialog box. Click OK to save your changes and close the System Preferences dialog box.

Setting Workspace Backup Options

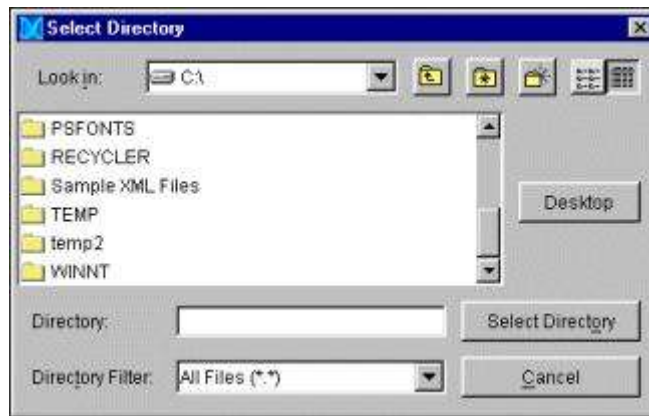
The Genome Browser is set to periodically backup your Workspace. If you like, you can disable this feature.

- 1 From the Main Menu, select Edit|Preferences|System .
- 2 Click the Workspace Backup tab..



- 3 To disable the workspace backup, turn on the No radio button.
- 4 If desired, click Change For Next Session to change the default directory and/or file name for the workspace backup file..
- 5 Click Apply to save your changes and continue working in this dialog box. Click OK to save your changes and close the dialog box.

<u>Genome Browser Data Classes</u>	<u>3.2</u>
<u>Assembly 3.2</u>	
<u>Features 3.2</u>	
<u>Precomputes</u>	<u>3.2</u>
<u>Annotations</u>	<u>3.3</u>
<u>Celera Exchange (XML) Documents</u>	<u>3.4</u>
<u>XML Assembly Files</u>	<u>3.4</u>
<u>XML Feature Files</u>	<u>3.4</u>
<u>XML Workspace Files</u>	<u>3.5</u>



GENOME BROWSER DATA CLASSES

See [page 3.6](#) for a diagram that illustrates the Genome Browser data object flow.

The Genome Browser supports these major data classes:

- Assembly
- Features, comprised of precomputes and annotations

Assembly

Assembly is the raw nucleotide residue data, stored as sequences of A, T, C and G, which have been ordered together to form single base-pair coordinate systems known as Genomic Axes. Genomic Axes represent either an entire chromosome, or the largest assembled portions of a chromosome (often referred to as scaffolds). A Genomic Axis may also represent a set of scaffolds that are ordered and oriented relative to one another. A Genomic Axis may contain either a single string of nucleotide residues, or it may be comprised of multiple, separate, contiguous-residue strings known as contigs, ordered and oriented within a scaffold.

Features

Features are comprised of precomputes and annotations.

Precomputes

Precomputes are features, aligned to a sequence, that have been determined by some algorithmic process (e.g., BLAST). Precomputes may be either:

- simple form —single ranges along a genomic axis, with descriptive information (e.g., HSP)
- compound form — adjacent simple features that are grouped by the analysis program (e.g., BlastHits)

Annotations

Annotations are features, aligned to a sequence, that have been created by a human or automated curation process. The Genome Browser supports three annotation feature types:

- Exon — a single range on a nucleotide sequence, together with descriptive information and zero or more evidence relationships. Evidence relationships are references to precompute objects used as evidence of the existence of an exon.
- Transcript — a sequence of exons separated by intron regions, translated into a protein sequence. A transcript may also have a Start_Codon or a Translation_Start_Position (if the Start_Codon can not be determined), together with an optional Stop_Codon to indicate the translation frame of the transcript.
- Gene — one or more transcripts that are associated with a single gene.

CELERA EXCHANGE (XML) DOCUMENTS

The Genome Browser supports the following types of XML exchange documents:

NOTE: You may open only Celera-signed XML assembly files.

- Assembly
- Feature
- Workspace

These files all use the same data file format, but they contain different kinds of data. They are distinguished by using different file name extensions to indicate the type of data they contain.

The following sections provide more detail.

XML Assembly Files

Assembly files (.gba extension) are the only files that can contain genomic assembly data. An assembly file may contain features as well, but for large data sets it is more practical to break the features into multiple feature (.gbf) files that you can load on an as-needed basis. Celera-signed assembly files do not contain features.

XML Feature Files

NOTE: Although there is no constraint on how to organize features in feature files, your computer may experience memory problems if a feature file is too large.

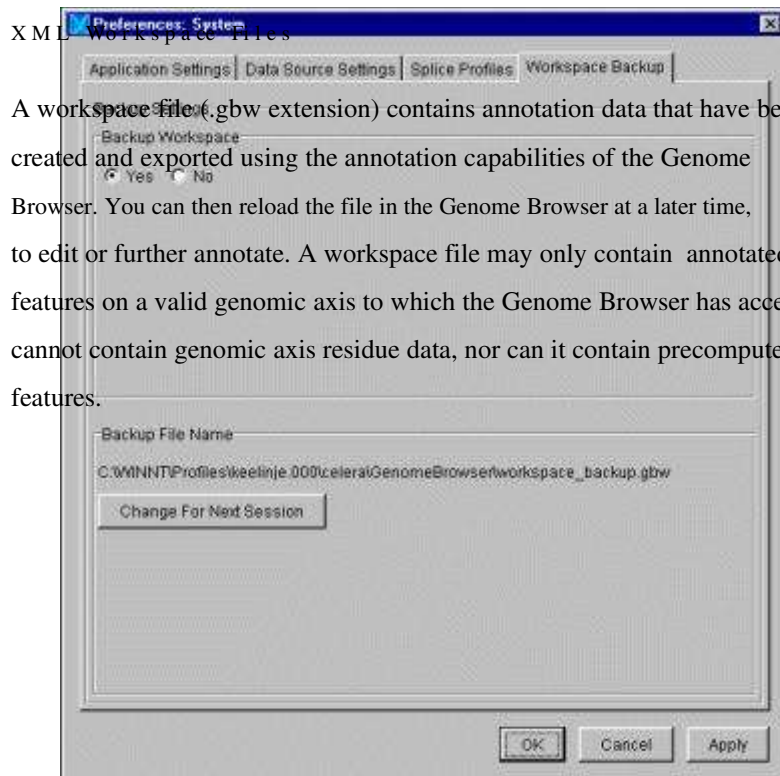
Feature files (.gbf extension) contain either precomputed or human-annotated features calculated along a genomic axis. However, they are not allowed to contain assembly (i.e., genomic axis residue) data. There is no limit to how many or what type of features can be contained in a feature file, as long as they each reference a valid genomic axis to which the Genome

Browser has access. You can open any number of feature files in the Genome Browser at one time, and a single feature file may contain features on many different genomic axes.

Human-curated features contained in the feature files show up in the “Promoted” tier of the browser.

NOTE: You may load only one workspace file in to the Genome Browser at a time.

A workspace file (.gbw extension) contains annotation data that have been created and exported using the annotation capabilities of the Genome Browser. You can then reload the file in the Genome Browser at a later time, to edit or further annotate. A workspace file may only contain annotated features on a valid genomic axis to which the Genome Browser has access. It cannot contain genomic axis residue data, nor can it contain precomputed features.



The following diagram illustrates the Genome Browser data object flow.

<u>Accessing the Data</u>	<u>4.3</u>
<u>Searching for a Specific ID</u>	<u>4.3</u>
<u>Searching for Sequence Data</u>	<u>4.5</u>
<u>Searching for an XML Feature ID</u>	<u>4.6</u>
<u>Opening a Genome Version</u>	<u>4.6</u>
<u>Drilling Down to a Genomic Axis</u>	<u>4.7</u>
<u>Opening an XML Workspace (.gbw) File</u>	<u>4.9</u>
<u>Viewing an XML Workspace or Feature File</u>	<u>4.9</u>
<u>Navigating to a Feature in an XML Workspace File</u>	<u>4.10</u>
<u>Displaying Open Data Sources</u>	<u>4.11</u>
<u>Loading Precompute and Annotation Features</u>	<u>4.12</u>
<u>Showing or Hiding Data Tiers</u>	<u>4.14</u>
<u>Reverse Complementing the Axis</u>	<u>4.15</u>
<u>Displaying a Feature's Properties</u>	<u>4.17</u>
<u>Zooming to a Selected Feature</u>	<u>4.17</u>
<u>Highlighting Evidence</u>	<u>4.18</u>
<u>Showing Edge Matches</u>	<u>4.18</u>
<u>Showing Evidence Edge Mismatches</u>	<u>4.19</u>
<u>Showing non-GT/AG Splices Matches</u>	<u>4.19</u>
<u>Finding a Feature's Matching Subject Sequence</u>	<u>4.20</u>
<u>Displaying Consensus Sequence</u>	<u>4.20</u>
<u>Displaying the Translations or Reverse Complement</u>	<u>4.23</u>
<u>Copying Sequence to the Clipboard</u>	<u>4.24</u>
<u>Setting Color Rules For Feature Display</u>	<u>4.25</u>
<u>Creating a New Color Rule</u>	<u>4.25</u>
<u>Editing an Existing Color Rule</u>	<u>4.28</u>
<u>Deleting an Existing Color Rule</u>	<u>4.30</u>
<u>Applying a Color Rule</u>	<u>4.30</u>
<u>Resetting the View</u>	<u>4.30</u>
<u>Using Property Rules</u>	<u>4.31</u>

<u>Using a Property Rule to Create Links to</u>	
<u>HTML BioMolecule Reports</u>	<u>4.33</u>
<u>Displaying Predicted Features in the</u>	
<u>Transcript Translation View</u>	<u>4.35</u>
<u>Changing the Translations Display</u>	<u>4.36</u>
<u>Copying Transcript Sequence to the Clipboard</u>	<u>4.36</u>
<u>Evaluating Feature Homologies</u>	<u>4.37</u>
<u>Displaying the Feature Report</u>	<u>4.38</u>
<u>Running BLAST Searches</u>	<u>4.40</u>
<u>Searching for a Nucleotide or Protein Sequence String</u>	<u>4.41</u>
<u>Using Bookmarks</u>	<u>4.42</u>
<u>Creating 4.42</u>	
<u>Opening a Bookmark</u>	<u>4.42</u>
<u>Editing or Deleting a Bookmark</u>	<u>4.42</u>
<u>Displaying Multiple Browser Windows</u>	<u>4.43</u>
<u>Closing a Browser Window</u>	<u>4.43</u>
<u>Printing a Page</u>	<u>4.43</u>

ACCESSING THE DATA

To access data in the Genome Browser, you may either:

- Search for a specific ID and navigate to its region on the genome
- Open one or more genome versions and drill down to a region on a genome

Searching for a Specific ID

- 1 From the Main Menu, select Search|Features...

The Search Known Features dialog box appears:

NOTE: If you are not already connected to an information service, the first query type is the only available option.

2 Select a query type:

- Search the available Genome Version(s) — Searches all genome versions to which you have access, whether you are connected to a genome version or not.
- Search the loaded Genome Version(s) — Searches only the genome versions to which you are connected. This type is unavailable if you do not have a genome version loaded.
- Search the current Genome Version — Searches only the genome version you have selected in the Outline View. This type is unavailable if you do not have a folder selected in the Outline View.

NOTE: The last ID type used will be the topmost available option in the list.

3 Select one of the following ID types from the pull-down list:

- Transcript (Celera Accession)
- Gene (Celera Accession)
- Genomic Axis Name
- Protein (Celera Accession)
- Feature ID
- Unknown ID
- Chromosome
- Celera Variant (SNP)
- External ID (gi or accession)
- STS Name
- BAC accession or clone name
- Subject Sequence ID
- Gene Ontology
- Gene Family
- Regulatory Region Accession
- Conserved Segment Accession
- Gene Index Accession

NOTE: Open New Browser is unchecked by default. Check this box to display the data in a separate browser window.

- 4 Type or paste the ID for which to search in the blank text box and click OK. With the exception of GA names, this search is not case-sensitive.
- 5 Select the desired result and click Navigate to view the data, or Bookmark to mark the results for later use.

Searching for Celera Discovery System Sequence Data

This section describes how to search for a GA (Genomic Axis) number in the Genome Browser:

- 1 From the Main Menu, select Search|Features...
The Search Known Features dialog box appears.
- 2 Select a query type.
- 3 Select Genomic Axis Name from the pull-down list, and type or paste the desired GA number (e.g., GA_x9V1BB6) in the blank text box.

TIP: Right-click to paste the GA number in the text box.

- 4 Click OK.
If more than one result appears, select the desired GA ID.
- 5 Click Navigate.
The information for this genomic axis appears in the Property Inspector View.

From the Main Menu, select Views|Genomic Axis Annotation View to see the data in the Genomic Axis Annotation View or double-click on the GA number in the Outline View.

See [page 4.4](#) for more information about the available query types.

Searching for an XML Feature ID

You can also search for a Feature ID that appears in a XML Feature file (.gbf).

- 1 From the Main Menu, select Search|Features...
The Search Known Features dialog box appears.
- 2 Select a query type.
- 3 Select Feature ID from the pull-down list and type (or paste) the unique Feature ID in the blank text box.
- 4 Click OK.
The Choose Path dialog box appears with the feature selected or a list of feature IDs. If more than one appears, select the desired ID.
- 5 Click Navigate.
The information for this feature appears in the Property Inspector View.
- 6 From the Main Menu, select Views|Genomic Axis Annotation to see the data in the Genomic Axis Annotation View.

See [page 4.4](#) for more information about the query types.

Opening a Genome Version

You may open one or more genome versions to drill down to the assembly and feature data that you want to view. You can either:

- Connect to a Celera database to view an assembly and its feature data
- Open an XML assembly (.gba) file

NOTE: You may open only Celera-signed XML assembly files. Also, before you can open a .gba file, you must specify the Data Source Settings from the Preferences dialog box. Refer to [“Setting Preferences” on page 2.6](#) for instructions.

To open a genome version:

- 1 From the Main Menu, select File|Open Genome Version.
The Selection Species/Assembly dialog box appears, displaying a list of available genome versions.

- 2 Select the desired assembly from displayed list, and click OK, or double-click on the selected assembly.

TIP: To open more than one genome version at a time, use Ctrl-click to select multiple genome versions and then click OK.

The Outline View now displays the unexpanded species folder. For example:

Drilling Down to a Genomic Axis

- 1 Connect to the Celera database or open an assembly file.
- 2 Expand the species folder and the sub-folders to open the folder that contains desired axis.

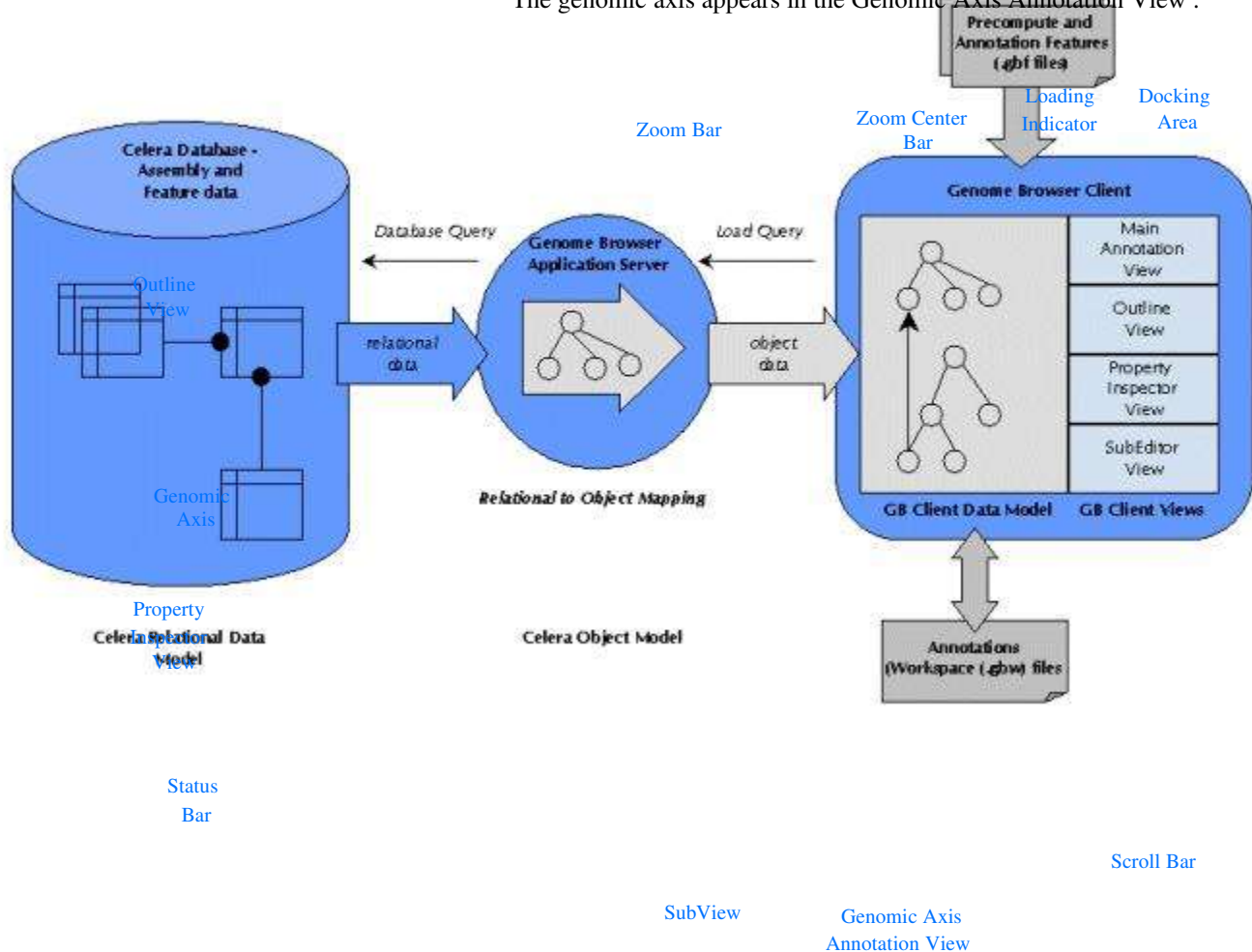
TIP: Right-click your mouse on the genomic axis, chromosome, or scaffold position on the chromosome; and select a sort option for viewing by size:

- 3 Select the genomic axis' folder.

The axis information appears in the Property Inspector View.

- 4 Double-click the genomic axis' folder.

The genomic axis appears in the Genomic Axis Annotation View :



- To center around a specific region before you zoom in or out, position the vertical Zoom Center Bar in the desired region.
- To zoom out or in, slide the zoom bar up or down.
- To pan across the genomic axis, slide the horizontal scroll bar left or right.

Opening an XML Workspace (.gbw) File

You can open your own workspace file, as long as this file was created using the Celera Exchange File Format. The Genome Browser displays the data with the Celera data, allowing you to create or modify annotations on your data.

See [Appendix A](#) for the detailed Exchange File Format Specification and Document Type Definition.

You may work with only one XML workspace file at a time.

- 1 Ensure that the file was created using the Celera Exchange File format.
- 2 From the Main Menu, select File|Open Workspace File.

If you have never opened an XML workspace file, the Open dialog box appears automatically. Browse to locate the desired file, and click Open.

TIP: If you have opened one or more XML workspace files, the sub-menu allows you to access the Open dialog box or to select from a list of recently opened files. Select a file from this list to open it, or select Open File... to access the Open dialog box, browse to locate the desired file, and click Open.

Viewing an XML Workspace or Feature File

- 1 Open the desired XML workspace file.
- 2 From the Main Menu, select File|View Celera Exchange File.
The Celera Exchange File Viewer displays.

- 3 Drill down to the desired object in the viewer to display its attributes and properties:

Navigating to a Feature in an XML Workspace File

- 1 Open the desired XML workspace file.
- 2 From the Main Menu, select File|View Celera Exchange File.
The Celera Exchange File Viewer displays.
- 3 Drill down to the desired feature and select it.
- 4 From the Viewer's menu, select Genome Browser|Navigate.
The Genome Browser displays the selected feature in the Genomic Axis Annotation View and the Property Inspector View.

Displaying Open Data Sources

You can easily find out which assembly, feature, and workspace files you have open.

- 1 From the Main Menu, select File/List Open Data Sources.

A dialog box appears, listing all data sources that the Genome Browser has open now.

- 2 Close the dialog box.



LOADING PRECOMPUTE AND ANNOTATION FEATURES

To conserve system resources, Genome Browser does not automatically load contig or feature information when you navigate to an axis. You must select a portion of the genomic axis to view, and then load the features for the selected portion.

See [“Adding XML Directories” on page 2.18](#) for instructions.

- 1 If you wish to load features that you created using the Celera Exchange File Format, ensure that you have specified the XML directory that contains the feature file(s).
- 2 Before you select a region, you should use the zoom and horizontal scroll bars to zoom in on the desired region.
- 3 Left-click and drag your mouse to select a 500Kb or less region on the axis. The Genome Browser allows you to select a larger region, but it will load data from the start of the selected region up to the maximum range.

To select the entire visible region, select Options|Select Visible Region from the Main Menu.

- 4 From the Main Menu, select Data Manipulation|Load Data, or right-click on the axis and select Load Data from the sub-menu.
- 5 Select the annotation and precompute feature types to load from the menu.

TIP: Loading predicted splice site, start, and stop locations can consume a great deal of memory. We suggest that you select a smaller region from a genomic axis when loading these feature sets.

NOTE: Mouse over the Loading Indicator to see the Progress Meter.

The following table shows the Celera feature types (Your feature file may contain different feature types).

Celera Feature Types

Feature Type	Options
<ul style="list-style-type: none"> • Human Curated Features • High Pri Computed Features • Low Pri Computed Features • Contigs 	<p>Select any or none to load information, including underlying contig data, computational analysis results, and annotation (automatic and human-reviewed).</p> <p>NOTE: The Genome Browser loads all pre-computed features aligned to the genomic axis' selected region from the relevant feature (.gbf) file(s) when you select High Pri Computed Features.</p> <p>TIP: The high-priority computed features are computationally determined to be the most important subset of computed features, and have undergone redundancy filtering. If your system experiences memory problems, you may choose to not load the low-priority computed features.</p>
Predicted Splice Sites	<p>Calculates splice site locations. You can load splice sites on either genomic axis' strand or on both strands. Select one or both of the following:</p> <ul style="list-style-type: none"> • Forward • Reverse
Predicted Start Codons	<p>Calculates start codon location. Select one or both of the following:</p> <ul style="list-style-type: none"> • Forward • Reverse
Predicted Stop Codons	<p>Calculates stop codon location. Select one or both of the following:</p> <ul style="list-style-type: none"> • Forward • Reverse

As you load the features, the Genomic Axis Annotation View displays them. The features on the forward strand are above the axis, and those on the reverse are below the axis. The exception would be if the axis was reverse-complemented.

See [“Showing or Hiding Data Tiers” on page 4.14](#) for more information.

Each set of features appears on a “tier.” Genome Browser only displays tier labels for those feature sets relevant to the selected region. The tiers are collapsed by default.

When you click on a feature in the Genomic Axis Annotation View, the information for the selected feature appears in the Property Inspector View.

Showing or Hiding Data Tiers

Depending on the number of available feature sets, you may decide to hide some tiers to better see the data that most interests you. Genome Browser defaults to having all tiers available.

To hide a tier, right click on a tier label and select Hide to remove the tier from the display.

NOTE: You should always expand a data tier when you are performing annotation.

To expand or collapse a data tier, right click on the tier’s label and select Expand or Collapse. You can single-click on a tier name to toggle between the Expand and Collapse options.

To see long tier names, adjust the tier column by pulling the window margin to the right, as if to widen the window.

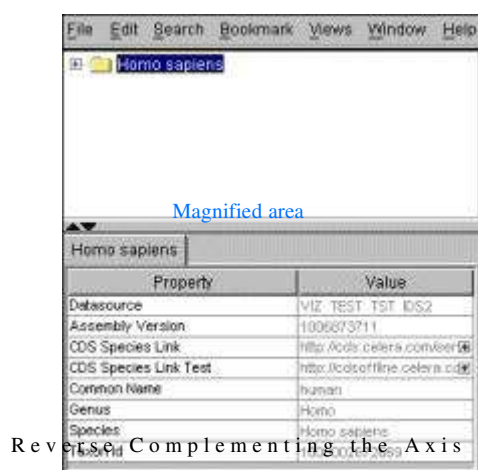
To move a particular feature close to the axis, right click on the feature and select Dock. To move a feature (e.g., consensus) away from the axis, right click on the feature and select Undock.

To move the axis tier up and down over precomputed tiers, click and hold the top of the axis tier up/down, or until the double-arrow becomes visible.

Clicking a double-arrow at the top of the axis will move the axis to the top of the View. Clicking the double-arrow at the bottom of the axis moves the axis to the bottom of the View.

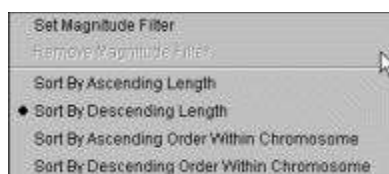
To change the order of the tiers, click on a tier name and drag it up or down.

To magnify results, select the area to magnify. Press the Alt key and right-click at the same time, and you will see the image magnified in the View.



Reverse Complementing the Axis

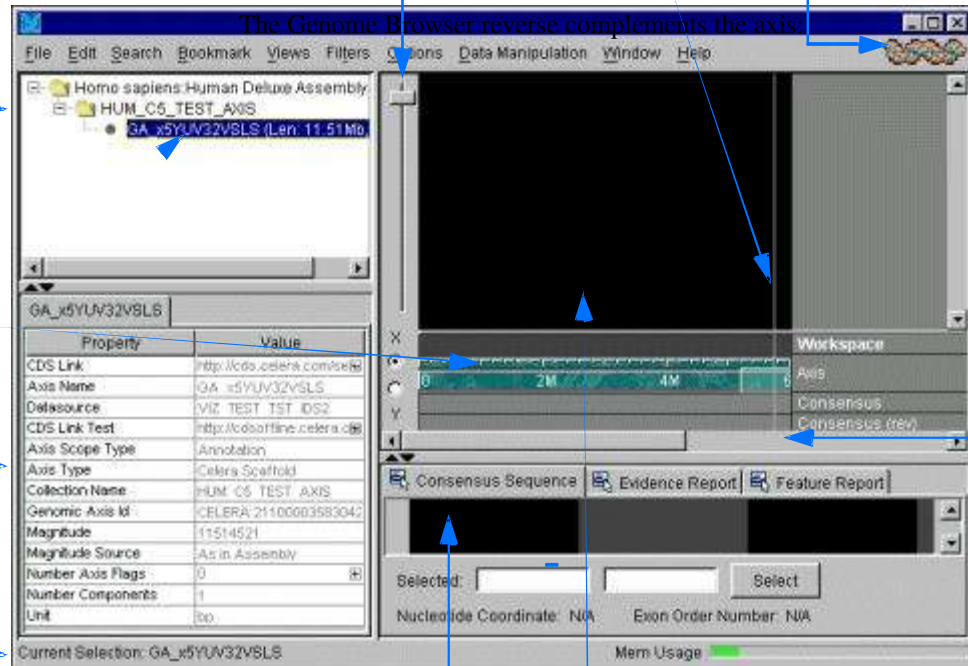
By default, the Genome Browser displays the forward strand from left to right above the axis, and the reverse strand from right to left below the axis:



When you are reviewing or annotating the reverse strand, it may be easier to flip the display. You can reverse complement the axis to display the reverse strand from left to right on the top of the Genomic Axis Annotation View:

NOTE: To restore the default view, select Options from the Main Menu, and uncheck Reverse Complement Axis.

- 1 From the Main Menu, select Options.
- 2 Select Reverse Complement Axis.



DISPLAYING A FEATURE'S PROPERTIES

To display any feature's properties, you use your mouse to select it in the Genomic Axis Annotation View.

To select a simple feature, click on the rectangle that represents the feature in the Genomic Axis Annotation View.

A red box appears around the selected feature, and its properties appear in the Property Inspector View.

To select a compound feature, shift-click anywhere on the feature.

- If the feature is a transcript, then simply click on an intron region to select it.
- If the feature is a precompute, click the line linking the glyphs.

A yellow box appears around the feature, and its properties appear in the Property Inspector View.

To select a gene, Ctrl-click on a transcript.

A green box appears around the gene, and its properties appear in the Property Inspector View.

Zooming to a Selected Feature

To zoom in on a selected feature:

From the Main Menu, double-click on the feature, or select Options/Zoom to Axis Selection or Options/Zoom to Subview Selection.

The Genome Browser centers and zooms in on the feature.

Highlighting Evidence

The Genome Browser allows you to highlight precompute features that were used to support a promoted feature. To display this information:

- 1 From the Main Menu, select Options.
- 2 Select Highlight Evidence.

To see an annotated feature's evidence, click the feature in the Promoted tier. The Genome Browser puts a red box around the evidence for that feature:

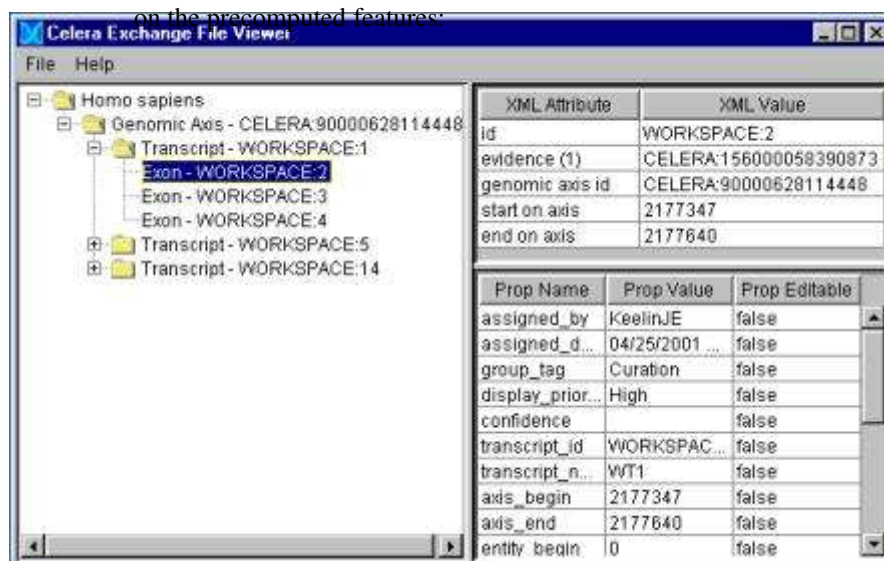
Showing Edge Matches

The Genome Browser allows you to highlight a promoted feature's edge matches to precomputed features. To display this information:

- 1 From the Main Menu, select Options.
- 2 Select Show Edge Matches.

To see the edge matches for a promoted feature, click the feature in the Promoted tier. The Genome Browser shows edge matches as yellow bars

on the precomputed features:



Showing Evidence Edge Mismatches

The Genome Browser allows you to highlight where the evidence is different than the transcript feature. To display this information:

- 1 From the Main Menu, select Options.
- 2 Select Show Evidence Edge Mismatches .

Showing non-GT/AG Splices Matches

The Genome Browser allows you to highlight exon edges that are non-canonical in blue. To display this information:

- 1 From the Main Menu, select Options.
- 2 Select Show non-GT/AG Splices Matches .

Finding a Feature's Matching Subject Sequence

The Genome Browser allows you to find features that have the same subject sequence on the selected portion of the genomic axis with features loaded.

This can be useful when you are evaluating LAP, Genewise, or SIM4 features.

For example, you can select a LAP feature and use this function to locate the corresponding BLASTx feature.

- 1 In the Genomic Axis Annotation View, select a range on the genomic axis.

TIP: If you have the SubView open, click the Subject Sequence Report tab to review matching subject sequence information for a selected feature.

- 2 Press <Shift> and click to select the feature.

A yellow box appears around the feature.

- 3 From the Main Menu, select Options|Find Matching Subject Sequence.

The Genome Browser locates features with the same subject sequence and places a box around these features.

Displaying Consensus Sequence

You can display a region's consensus sequence:

- in the Genomic Axis Annotation View
- in the SubView

To display a region's consensus sequence in the Genomic Axis Annotation View:

- 1 In the Genomic Axis Annotation View, center the Vertical Zoom Center Bar around the region.
- 2 Slide the Zoom Bar, which appears on the left side of the Genomic Axis Annotation View, down until the consensus residues appear:

To display a region's consensus sequence in the SubView:

- 1 If the SubView is not displayed, select Views|Display SubViews When Available from the Main Menu.

NOTE: If you move the blue glyph, the vertical scroll in the Consensus Sequence window will move in unison.

- 2 When the SubView displays, select a Genomic Axis Range. Right-click over the axis and select Select SubView Range. The SubView Range will be initially set to the Annotation view range. The range will be highlighted by a yellow bar, and a blue glyph will appear that allows you to scroll across the consensus sequence. The underlying genomic

sequence for the axis' selected region appears in the Consensus

Sequence window:



Move the blue glyph to
scroll across the
sequence

Underlying genomic
sequence for the axis'
selected region



Vertical scroll

- 3 If desired, click on a feature in the Genomic Axis Annotation View.



A yellow box indicates the selected feature, and the color of the feature's sequence in the Consensus Sequence window changes to the same color that appears in the Main Annotation:

Yellow box indicates the selected feature

Color of the feature's sequence matches the same color in the Main Annotation

Displaying the Translations or Reverse Complement

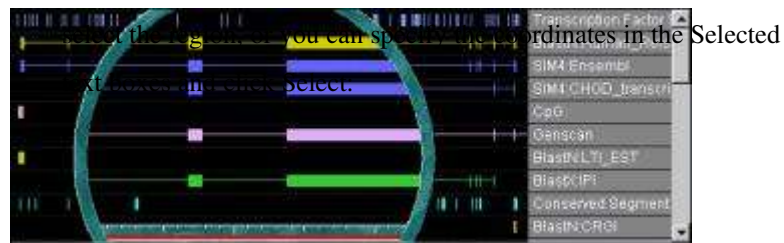
To do this from the SubView:

- 1 Right-click on the Consensus Sequence tab and select Show.
- 2 Click the desired translation and/or Reverse Complement from the sub-menu.
A check appears next to your selection.

Copying Sequence to the Clipboard

To copy consensus residues from the SubView to the clipboard for further use:

- 1 Select the desired region in the SubView. You can either click and drag to



- 2 Right-click on the Consensus Sequence tab and click Copy Selected Sequence to Clipboard or Copy Selected Sequence to Sequence Analysis.

- 3 Select one of the following from the sub-menu:

- Nucleotide Sequence
- Amino Acid Sequence

- 4 If you select Amino Acid Sequence, select either:

- +1 Translation



To copy sequence from the Genomic Axis Annotation View to the clipboard for further use:

- 1 Select the desired range on the genomic axis.
- 2 From the Main Menu, select Edit/Copy Sequence.
- 3 Select one of the following from the sub-menu:

- Copy forward sequence

Setting Color Rules For Feature Display

You can use color rules to change some or all features' color intensity based on the value of a specific feature property.

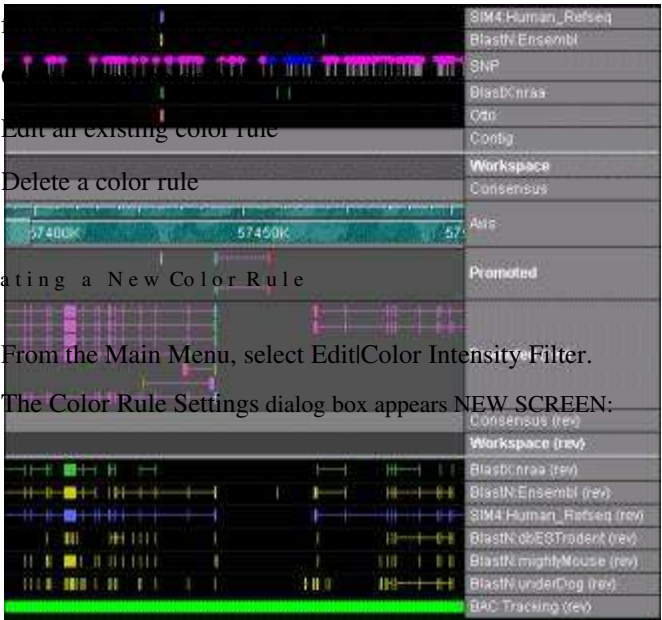
The

-
- Edit an existing color rule
- Delete a color rule

Creating a New Color Rule

1 From the Main Menu, select EditColor Intensity Filter.

The Color Rule Settings dialog box appears NEW SCREEN:



NOTE: The Save and Delete buttons are active only if you have already defined a color rule. Also, the Apply Rule and Reset View buttons are active only if you have a region selected on the genomic axis.

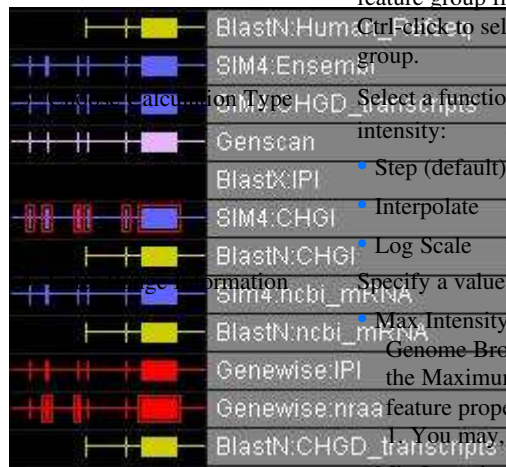
2 Click New.

The Color Rule Settings|Create a New Color Rule dialog box appears:

3 Use the following table to complete this dialog box:

Step	Instructions
1 Select Property to Trigger From	Select a Feature Property from the pull-down list. The Genome Browser will apply the color rule based on this feature's property value.
2 Select Effected Feature Groups	Check one of the following boxes: <ul style="list-style-type: none"> • Apply To All — All features that have the selected feature property will display in the Genomic Axis Annotation View using this color rule. • Apply To Specific — Only the feature group that you select will display using this color rule.

If you check Apply To Specific, select a feature group from the pull-down list box.



Select a function to use for shading

intensity:

• Step (default)

• Interpolate

• Log Scale

Specify a value for each of the following:

• Max Intensity At Value — The Genome Browser suggests values for the Maximum Intensity, based on the feature property that you select in Step 1. You may, however, change this value.

• Min Intensity At Value — The Genome Browser suggests values for the Minimum Intensity, based on the feature property that you select in Step 1. You may, however, change this value.

• Ranges — Specify the number of ranges. This is applicable for the Step and Interpolate functions only. The Log Scale function uses the extreme values.

5 Enter The Rule Name Type a name for the color rule in the text box.

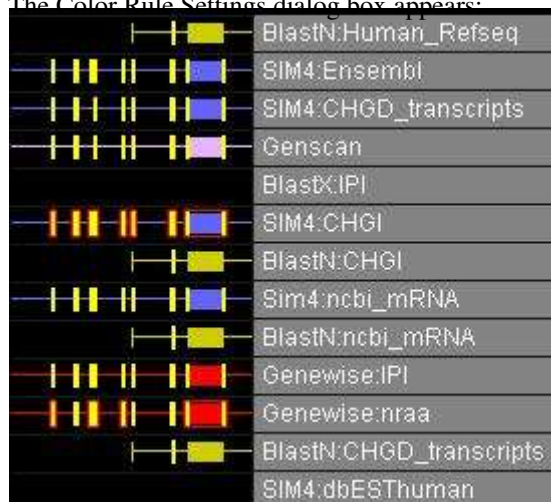
6 When you have completed the information in this dialog box, click OK.

7 Click OK to exit the Color Rule Settings dialog box.

Editing an Existing Color Rule

- 1 From the Main Menu, select Edit|Color Rules.

The Color Rule Settings dialog box appears:



Select a Rule to edit from the pull-down list.

- 2 Change the existing preferences as desired. The following table describes the settings you may change:

Color Rule Setting	Instructions
Rule Settings Group	<p>Select a function to use for shading intensity:</p> <ul style="list-style-type: none"> • Step (default) • Interpolate • Log Scale <p>Then, check one of the following boxes:</p> <ul style="list-style-type: none"> • Apply To All — All features that have the selected feature property will display in the Genomic Axis Annotation View using this color rule. • Apply To Specific — Only the feature group that you select will display using this color rule. <p>If you check Apply To Specific, select a feature group from the pull-down list box.</p> <p>Finally, select a Feature Property from the pull-down list. The Genome Browser will apply the color rule based on this feature's property value.</p>
Data Points Group	<p>To edit the Value or Intensity for a specific data point, click in the row to edit, type your changes, and then press <Enter>.</p> <p>To add a new data point, complete the Value and Intensity text boxes and click Add Row.</p> <p>To delete a data point, click in the row to delete and click Remove Row.</p>
Select Example Color	<p>To use a different example color, click Select Example Color and then select a new color. The new color will appear in the Example column.</p>

- 3 When you have made the desired changes, click Save.
- 4 Click OK to exit the Color Rule Settings dialog box.

Deleting an Existing Color Rule

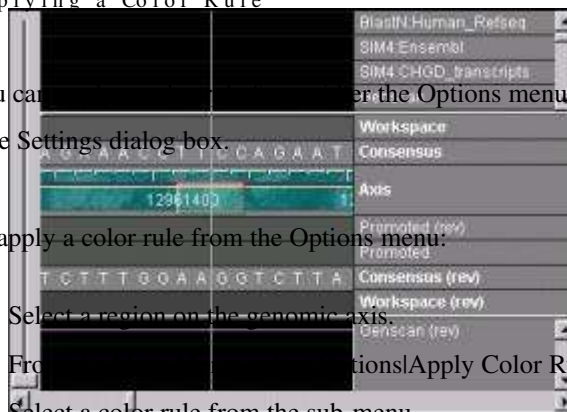
- 1 From the Main Menu, select Edit|Color Rules.
- 2 Select a Rule to delete from the pull-down list.
- 3 Click Delete.
- 4 When the Rule Deletion confirmation message appears, click Yes.

Applying a Color Rule

You can apply a color rule from the Options menu or from the Color Rule Settings dialog box.

To apply a color rule from the Options menu:

- 1 Select a region on the genomic axis.
- 2 From the Main Menu, select Options|Apply Color Rule.
- 3 Select a color rule from the sub-menu.



To apply a color rule from the Color Rule Settings dialog box:

- 1 Select a region on the genomic axis.
- 2 From the Main Menu, select Edit|Color Rules.
- 3 Click Apply Rule.
- 4 Click OK to exit the dialog box.

Resetting the View

You can reset the color intensity to 100% from either the Options menu or from the Color Rule Settings dialog box.

To reset the color intensity to 100% from the Options menu:

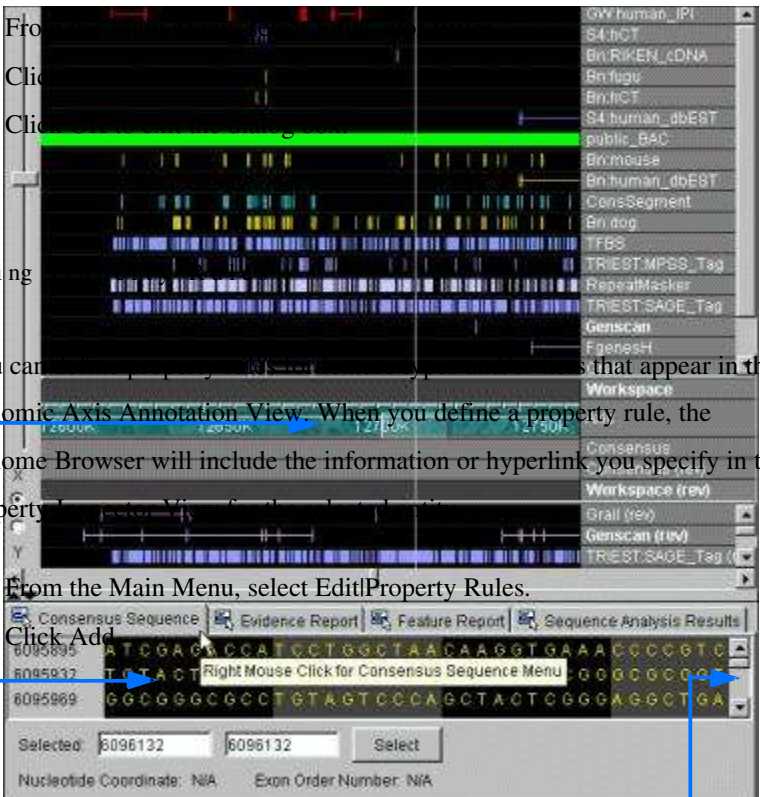
- 1 From the Main Menu, select Options|Apply Color Rule.
- 2 Select Reset Intensities from the sub-menu.

To reset the color intensity to 100% from the Color Rule Settings dialog box:

- 1 From the Main Menu, select EditProperty Rules.
- 2 Click Add.
- 3 Click the feature you want to add.

Using the Genome Browser, you can view the properties of a feature that appear in the Genomic Axis Annotation View. When you define a property rule, the Genome Browser will include the information or hyperlink you specify in the Property Rule.

- 1 From the Main Menu, select EditProperty Rules.
- 2 Click Add.



The screenshot shows the Genome Browser interface with a genomic track. The track displays various annotations, including gene models, exons, and introns. A red arrow points to the 'Add' button in the 'EditProperty Rules' dialog box. Another red arrow points to the 'Consensus Sequence' menu item. A third red arrow points to the 'Right Mouse Click for Consensus Sequence Menu' option.

- 3 The Add Property Rule dialog box appears:

The following table explains how to complete this dialog box:

S e c t i o n	I n s t r u c t i o n s
Selection	This displays the current selection's ID, Entity Type (feature type), and Discovery Environment (tier name).
Rule Name = text box	Type a name for the rule you are creating in this text box. You may use spaces.
Conditions	<ol style="list-style-type: none">1 Select one or more feature types from the When Entity Type = section.2 Select one or more tier names from the When Discovery Env = section.3 Click either the And or the Or radio button.

Section

Instructions

Create Property

- 1 Type a name for the property in the Name = text box. Do not type spaces or hyphens.
- 2 Type a value for the property in the Value = text box. For example type an internal or external ID number for a property as a value. To select a value from a list, select a value from the list, and click OK.

- 3 When the dialog box, click OK.

Using a Property Rule to Create Links to CDS BioMolecule Reports

The following example explains how to add a property rule that will create a link from a Transcript feature in the Genome Browser to a web-based BioMolecule Report — mRNA view.

- 1 From the Main Menu, select EditProperty Rules.
- 2 Click Add.
- 3 In the Rule Name = text box, type a name for the rule; for example, **"BioMolecule Report link"**.
- 4 In the When Entity Type = section, select "Transcript".
- 5 In the Name = text box, type a property name; for example, **"BMR_mRNA_View"**.
- 6 In the Value = text box, type the URL to use as follows:
`http://myserver/servlet/
com.celera.gateway.GatewayURL?&page=bmr
&species=Homo+sapiens&id=<transcript_accession>`

When you have finished completing the dialog box, it should appear similar to this:

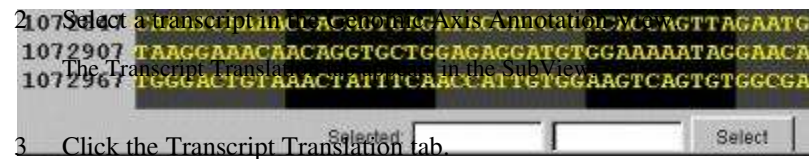
7 Click OK.

Double-click link to
access the HTML
BioMolecule Report for
this transcript.

Displaying Predicted Features in the Transcript Translation View

To review a predicted or promoted feature in the Transcript Translation View:

- 1 If the SubView is not displayed, select Views|Display SubViews when available from the Main Menu.



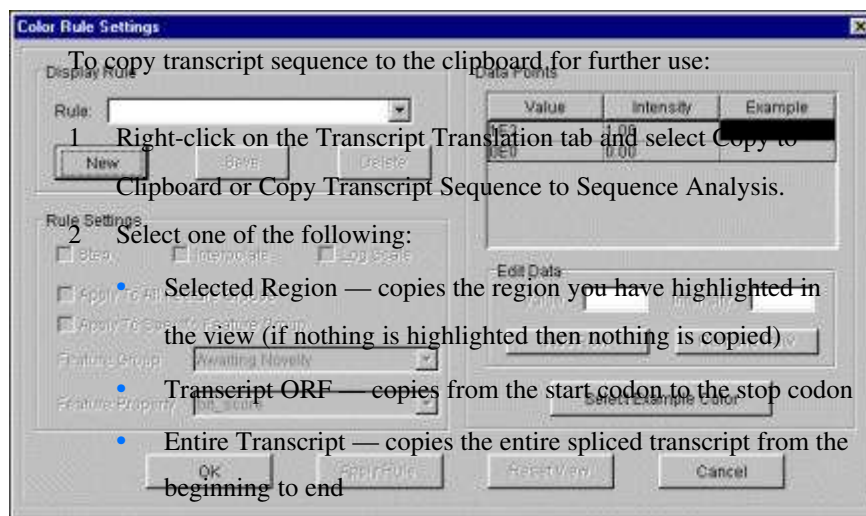
- 3 Click the Transcript Translation tab.

Changing the Translations Display

If you wish to change the display:

- 1 Right-click on the Transcript Translation tab and select Show.
- 2 Select or deselect a menu item.
- 3 Repeat steps 1 through 2 until the display is as you wish.

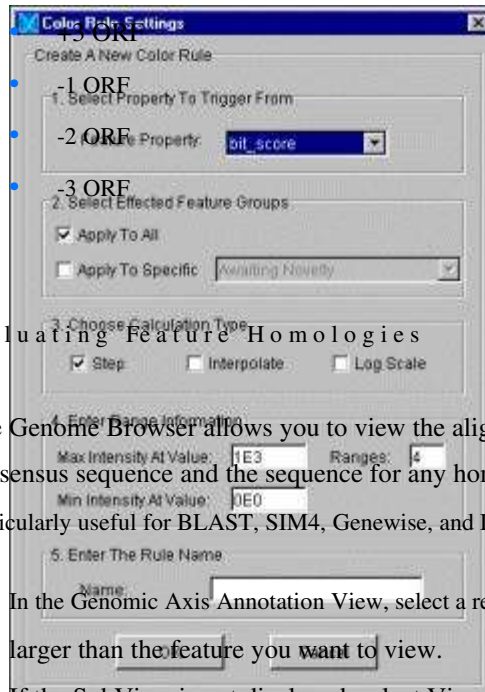
Copying Transcript Sequence to the Clipboard



- 3 Select one of the following from the sub-menu:
 - Nucleotide Sequence
 - Amino Acid Sequence

4 If you select Amino Acid Sequence, select one of the following:

- +1 ORF
- +2 ORF



Evaluating Feature Homologies

The Genome Browser allows you to view the alignment between the Celera consensus sequence and the sequence for any homology feature. This is particularly useful for BLAST, SIM4, Genewise, and LAP.

1 In the Genomic Axis Annotation View, select a region from the axis that is larger than the feature you want to view.

2 If the SubView is not displayed, select Views | Display SubViews when available from the Main Menu.

NOTE: You must select a feature that is completely within the range you have selected on the axis. The double-arrow in the upper left of the the Subview allows you to expand or collapse the View.

3 Click on a homology feature in the Genomic Axis Annotation View. The Query Sequence Alignments tab appears in the SubView.

4 The Query Sequence Alignments includes a zoom and zoom center bar. Zoom in the Query Sequence Alignments to review the sequence homologies.

TIP: If a feature has more than one exon, the Genome Browser will automatically zoom in on the first exon. To zoom in on the second or other exon, click on it before you zoom in.

NOTE: To review an HTML report, right-click on the DbAlignView tab and select HTML Report | Show HTML Report.

Depending upon the analysis program for the selected feature, the window displays either nucleotides or amino acids (i.e., a BLASTN feature displays DNA sequence, and a BLASTX feature displays protein sequence). The Celera sequence is on top, and the subject sequence is

below. The frames are relative to the entire sequence of the genomic axis.

~~The Query Sequence Alignments indicates homology by varying shades~~ of color. For protein sequences, bright green indicates a perfect match, and for nucleotide sequences, bright blue indicates a perfect match.

- 5 Select another feature, if desired.
- 6 To delete the contents of the Query Sequence Alignments window, right-click on the Query Sequence Alignments tab and select Clear.

Displaying the Feature Report

NOTE: The Feature Report displays information for only the data within the selected SubView range on the axis.

- 1 ~~In the Genomic Axis Annotation View, select a region from the axis that~~ contains the features you wish to view in the report, and load the features for the range.
 - 2 If the SubView is not displayed, select Views|Display SubViews when available from the Main Menu.
 - 3 In the SubView, click the Feature Report tab.
 - 4 Select Options|Populate.
-

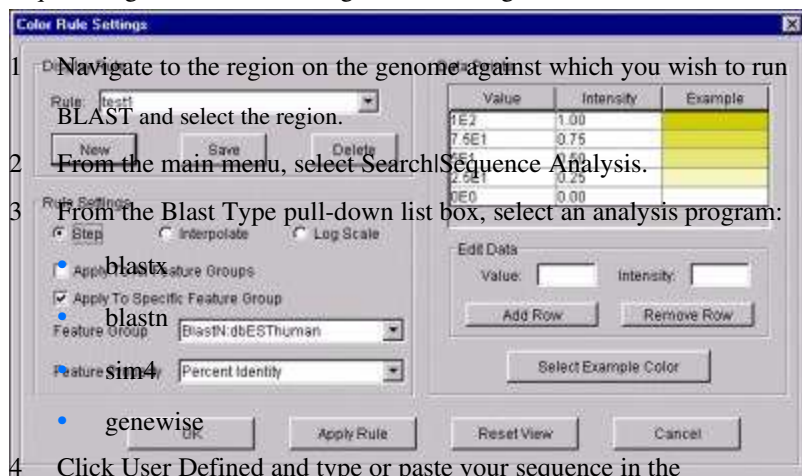
NOTE: Clicking on a feature in the Genomic Axis Annotation View highlights the corresponding row in the Feature Table (if selected).

The Genome Browser populates the Feature Report. The following table explains how to find information in this report.

To :	Do this :
Sort data by a column heading in descending order.	Click on the column heading.
Sort data by a column heading in ascending order.	Shift-click on the column heading.
Highlight report data in the Genomic Axis Annotation View.	Click in the row that you want to highlight to select an individual feature, or shift-click to select the composite feature. A colored box appears around the feature in the Genomic Axis Annotation View.

RUNNING BLAST SEARCHES

The Genome Browser allows you to compare a public or proprietary sequence against a selected range on a Celera genomic axis.



Sequence text box. It does not matter whether you include the definition line.

5 Click Search.

When the Search button becomes active again, the search is complete.

6 Click the Sequence Analysis Results tab in the SubView to review your results.

SEARCHING FOR A NUCLEOTIDE OR PROTEIN SEQUENCE STRING

You can search for a nucleotide or amino acid sequence string in either the Consensus Sequence or Transcript Translation window.

- 1 If the SubView is not displayed, select Views|Display SubViews when available.
- 2 Select either the Consensus Sequence or Transcript Translation tab.
- 3 Right-click your mouse and select Find... or press <Ctrl-F>.
- 4 Type or paste a search string in the Find what box.
TIP: You can use keyboard shortcuts for copying (<Ctrl-C>) and pasting in this box (<Ctrl-V>).
- 5 Select an option from the Look in pull-down list:
 - Nucleotide
 - Nucleotide Reverse Complement
 - ORF +1
 - ORF +2

 - ORF +3
 - All ORF's
- 6 If you selected an Amino Acid option from the Look in pull-down list, select either one-letter or three-letter from the Translation box.
- 7 Click Find Next or Find All.
If you click Find Next, the Genome Browser will return the next occurrence of the search string.

If you click Find All, the Genome Browser will return a table that displays all occurrences of the search string, with the corresponding locations.

NOTE: If you select Nucleotide Reverse Complement or an Amino Acid option, you do not need to be displaying that type of data in the Consensus Residues or Transcript Translation window. For example, the Genome Browser will find the reverse complement of ATG, even if the window is not displaying the Nucleotide Reverse Complement when you search.

USING BOOKMARKS

Genome Browser allows you to create and edit bookmarks. You can bookmark contigs and precomputed and annotated features.

Creating

- 1 Click the item you want to bookmark to select it.
- 2 From the Main Menu, select Bookmark|Bookmark Current Selection.

Opening a Bookmark

- 1 Ensure that you have an information service open.
- 2 From the Main Menu, select Bookmark|Select Bookmark .
- 3 Select your bookmark from the sub-menu.

Editing or Deleting a Bookmark

- 1 From the Main Menu, select Bookmark|Select Bookmark .
- 2 Select the bookmark that you wish to delete or modify.
- 3 Select Delete or Edit.

If you select Delete, the bookmark disappears from the window. If you select Edit, the Bookmark Properties box appears, and you can type over the existing name.

- 4 Click Close when you are done.

DISPLAYING MULTIPLE BROWSER WINDOWS

Genome Browser allows you to display more than one browser window at a time. This allows you to review multiple views of the same genome.

- 1 From the Main Menu, select Window.
- 2 Select New Browser Window.

NOTE: Depending on your system's resources, displaying multiple browser windows may consume more memory than is acceptable for your computer.

Genome Browser displays a duplicate of the window that you already have open. You can either review different features for the same data, or you can select different data to review.

Closing a Browser Window

- 1 Click in the browser window you want to close to activate it.
- 2 From the Main Menu, select Window.
- 3 Select Close Browser Window.

PRINTING A PAGE

To print any screen while you're in Genome Browser, select File|Print Screen.

Add Property Rule

Selection
Current Selection: hCT12345 - CELERA:104000074035889
Entity Type: Transcript
Discovery Environment: Edited Features

Define Rule
Rule Name =

Conditions
When Entity Type = ☐ And ☐ Or

When Discovery Env =

Create Property
Name =
Value =
Insert

Example:

OK Cancel

	<u>5.2</u>
<u>Introduction</u>	<u>5.2</u>
<u>Edit Commands</u>	<u>5.3</u>
<u>Annotation History</u>	<u>5.4</u>
<u>Creating a Transcript or Exon Using a Computed Feature</u>	<u>5.5</u>
<u>Reviewing and Editing Comments</u>	<u>5.7</u>
<u>Adding Evidence to a Transcript or Exon</u>	<u>5.7</u>
<u>Removing Evidence from a Transcript or Exon</u>	<u>5.7</u>
<u>Splitting a Transcript</u>	<u>5.8</u>
<u>Merging Transcripts</u>	<u>5.9</u>
<u>Adjusting Exon Boundaries</u>	
<u>Using the Consensus Sequence Window to Modify an Exon</u>	<u>5.9</u>
<u>Using Evidence to Modify an Exon</u>	<u>5.10</u>
<u>Using a Splice Site to Modify an Exon</u>	<u>5.10</u>
<u>Adding or Deleting Bases in the SubView</u>	<u>5.11</u>
<u>Finding a Long Open Reading Frame</u>	<u>5.11</u>
<u>Evaluating Subject Sequence Alignments</u>	<u>5.12</u>
<u>Deleting a Feature from the Workspace</u>	<u>5.12</u>
<u>Exporting Annotation Data</u>	<u>5.13</u>

INTRODUCTION

As you review the Celera assembly and annotation information, you may need to create, modify, and export annotation data. When you enable curation, the Genome Browser allows you to create new features or modify existing features in the Workspace area:

- transcripts
- genes
- exons
- start and stop codons
- evidence relationships

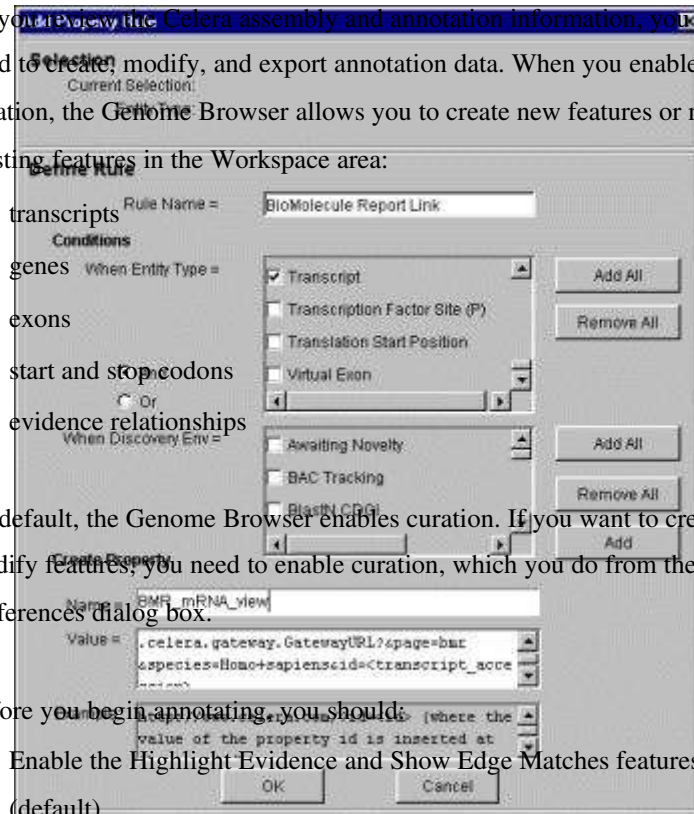
See [“Specifying System Preferences” on page 2.15](#) if you do not know how to disable curation.

By default, the Genome Browser enables curation. If you want to create or modify features, you need to enable curation, which you do from the Preferences dialog box.

Before you begin annotating, you should:

- Enable the Highlight Evidence and Show Edge Matches features (default).
- Expand the data tiers with which you are working.
- Open the SubView.

See [page 4.18](#) to learn how to highlight evidence and [page 4.18](#) to learn how to show edge matches. See [page 4.14](#) to learn how to expand the data tiers.



Edit Commands

Command	Value
BMR_mRNA_view	http://cds.premier.celera.com/service/cds
CDS_link	http://gb2cds.premier.celera.com/service
aliases	1
assigned_date	05/23/2001 08:43:38
axis_begin	12/59607
axis_end	12/59607
curation_flags	2
display_priority	High
entity_begin	0
entity_orientation	Forward
entity_end	12/59607
entity_flags	2
gene_accession	U00001.258
genomic_axis_id	CELERA:12800000144666
group_tag	Edited Features
is_child	true
is_deleted	false
in frame	

As you are creating and modifying annotation data, you can use the following edit commands:

To undo a change, select Edit|Undo from the Main Menu, or use the keyboard shortcut, Ctrl-Z.

To redo an undone change, select Edit|Redo from the Main Menu, or use the keyboard shortcut, Ctrl-Y.

To copy information from the Property Inspector View to the clipboard, select the information and use the keyboard shortcut, Ctrl-C. To paste the contents of the clipboard, position your cursor in the appropriate place, and use the keyboard shortcut, Ctrl-V.

Annotation History

The Genome Browser tracks the annotation that you create and modify. You can review the annotation history for either:

- the current annotation session
- the open Workspace file

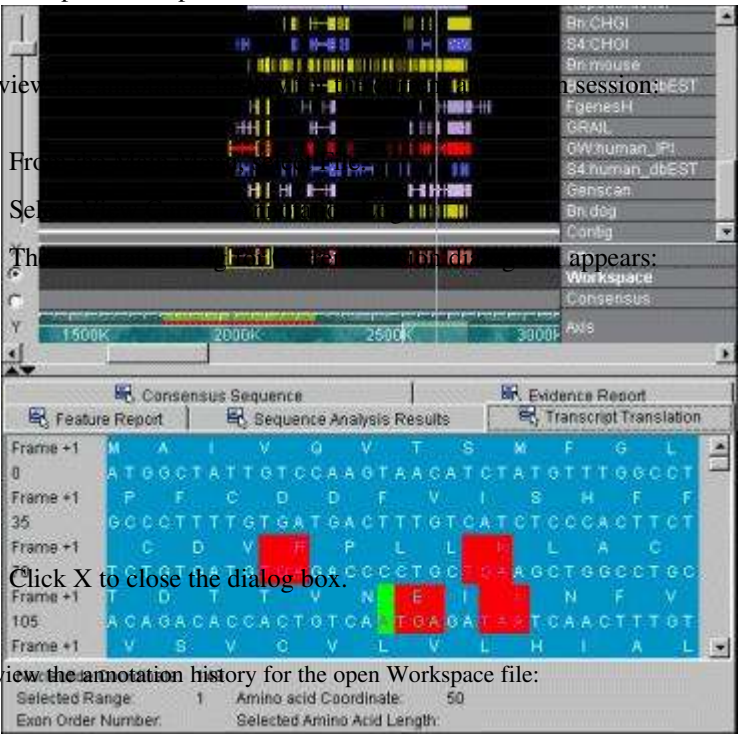
To view the annotation history for the open Workspace file:

1 From the Main Menu, select File.

2 Select View Open Workspace Annotation Log.

The Workspace Log dialog box appears:

3 Click X to close the dialog box.



The screenshot shows a complex interface. At the top, there's a track view with various colored bars representing annotations. Below that, a list of annotations is shown, including 'Br/CHGI', 'B4/CHOI', 'Br/mouse', 'n-session:NEST', 'FgenesH', 'IGRIL', 'QW/human_F1', 'B4/human_dbEST', 'Genscan', 'Br/dog', and 'Contig'. Below the list, there's a 'Workspace' section with a 'Consensus' track. At the bottom, there's a 'Sequence Analysis Results' section with a table showing sequence data across multiple frames. The table has columns for 'Frame +1', '0', '35', '70', '105', and 'Frame +1'. The rows show sequence data, with some cells highlighted in red and green. The table also includes 'Selected Range', 'Amino acid Coordinate', 'Exon Order Number', and 'Selected Amino Acid Length'.

1 From the Main Menu, select File.

2 Select View Open Workspace Annotation Log.

The Workspace Log dialog box appears:

TBA

3 Click X to close the dialog box.

Release 5.0

Genome Browser

CREATING A TRANSCRIPT OR EXON USING A COMPUTED FEATURE

To create a transcript or exon using a predicted feature:

- 1 Zoom in and click on a predicted feature.
- 2 Drag it down to the Workspace and release your mouse button. The feature appears in the Workspace. If you drag a promoted feature into the Workspace, the Genome Browser includes the gene and everything that is attached to it, including multiple transcripts, and the transcripts' exons and start and stop codons.
- 3 Click the new feature to select it.

The Property Inspector View displays the information for this feature.

NOTE: To select a transcript, click on an intron region, or shift-click on one of its exons.

The Genome Browser automatically sets the assigned by and assigned date fields in the Property Inspector View. The Genome Browser uses your system login id for the modified by value. Also, if you drag a transcript from the Promoted tier into the Workspace, the modified by and modified date are automatically set, too. You can modify the release status, reviewed by, and reviewed date fields.

REVIEWING AND EDITING COMMENTS

You may create and edit comments for transcripts and exons in the workspace. Precompute and promoted feature comments are read-only.

NOTE: If a feature does not have comments to display, the comments field does not appear.

To access comments for any feature, select it in the Genomic Axis Annotation View and click the + that appears in the comments field value column in the Property Inspector view. You may need to scroll down to see it.

If the comments are read-only, the following dialog box appears:

If the comments are not read-only, the following dialog box appears:

The following table describes this dialog box.

Element	Description/Instructions
New Comments	Allows you to add comments to this feature: Type the information in the New Comments text box, and click Add Comment. You can add multiple comments in this text box; simply click Add Comment after each item.
Existing Comments	Displays existing comments for this gene.

ADDING EVIDENCE TO A TRANSCRIPT OR EXON

To add computed evidence to a transcript or exon that appears in the Workspace:

- NOTE:** The Genome Browser will not add the evidence to the transcript if you place the evidence on an intron region. You must drag it on top of an exon.
- 1 Click the evidence in the Genomic Axis Annotation View to select it.
 - 2 Drag it down to the Workspace on top of an exon.
 - 3 Release your mouse button.

REMOVING EVIDENCE FROM A TRANSCRIPT OR EXON

To remove evidence:

- 1 Click the exon in the Workspace.
- 2 Right-click on the evidence to display the Evidence Options menu, and select Remove as evidence for current curation.

SPLITTING A TRANSCRIPT

To split a transcript in the Workspace:

- 1 Click on the intron region in the area where you wish to split the transcript.
- 2 Right-click your mouse and select Split Transcript.
The Genome Browser splits the transcript in the location where you right-clicked.

MERGING TRANSCRIPTS

First, you need to ensure that the transcripts are attached to the same gene:

- 1 Click the transcript in the Workspace to select it.
- 2 Right-click on an intron region and select Create New Gene from the context menu.
- 3 Ctrl-click to select the newly created gene to which the transcript will be attached.
- 4 Right-click your mouse on the transcript to attach and then click Attach Transcript to Selected Gene.
- 5 Ctrl-click on either feature, and a green box appears around both.

Next, you can merge the two transcripts in the Workspace:

- 1 Select the first transcript in the Workspace.
- 2 Right-click on an intron region in the second transcript.
The context menu appears:

- 3 Select Merge with Selected Transcript.

The Genome Browser merges the transcripts in the Workspace.

ADJUSTING EXON BOUNDARIES

You can modify exon boundaries using:

- the Consensus Sequence window
- Evidence
- Splice sites

The following sections provide instructions for each method.

Using the Consensus Sequence Window to Modify an Exon

- 1 If the SubView is not displayed, select Views|Display SubViews when available from the Main Menu.
- 2 Select a range along the axis. A red bar appear marking the range you selected.

NOTE: The SubView Range should be greater or equal to the Parent Feature range to view the colored nt's.

- 3 Select the Subview range by selecting Data Manipulation|SubView Range. A yellow bar marks the range you selected.
- 4 Select the exon to modify from the Axis.
- 5 Click the Consensus Residues tab, if it is not on top.

The underlying genomic sequence information appears in the box, in the area of the selected feature. A yellow bar on the Genomic Axis indicates the relative position. Exon sequence appears as purple, and intron sequence appears as yellow.

NOTE: You must begin with the first/ last base of the exon, or the boundary will not change. You can not move exon boundaries past a Start or Stop codon.

- 6 Starting at the exon's edge, drag the edge to the desired position. The boundary change is reflected on the axis.

Using Evidence to Modify an Exon

If you have added evidence to a transcript in the Workspace, you can tell the Genome Browser to use the evidence to modify an exon:

- NOTE:** The Genome Browser will not add the evidence if you drag it to an intron region. You must drag it on top of an exon.
- 1 Click the evidence in the Genomic Axis Annotation View to select it.
 - 2 Drag it down to the Workspace on top of the exon.
 - 3 Click the exon to modify in the Workspace to select it.
 - 4 Right-click on the evidence to display the Evidence Options menu:

NOTE: To undo a change, select [File|Edit|Undo...](#) from the Main Menu.

- 5 Select an option from the menu.
You can set the left and right edge of a current curation, as well as remove any evidence.

Using a Splice Site to Modify an Exon

- 1 Load the Predicted Splice Sites data for the selected axis range.
TIP: Loading predicted splice site locations can consume a great deal of memory. We suggest that you select a smaller region from the genomic axis before you load splice sites.
- 2 Click on the splice site to use.
- 3 Drag it down, and place your mouse pointer over the exon.
- 4 Release your mouse button.

ADDING OR DELETING BASES IN THE SUBVIEW

You can add or delete bases to a transcript in the SubView to see the effect on a translation.

NOTE: You can not save these changes.

- 1 If the SubView is not displayed, select Views|Display SubViews when available from the Main Menu.
- 2 In the Genomic Axis Annotation View, click on a feature.
- 3 Select the Transcript Translation tab in the SubView.
- 4 In the SubView, type or delete bases in the sequence.

FINDING A LONG OPEN READING FRAME

- 1 If the SubView is not displayed, select Views|Display SubViews when available from the Main Menu.
- 2 In the Genomic Axis Annotation View, click on a feature in the workspace to select it.
- 3 Select the Transcript Translation tab in the SubView.
- 4 In the SubView, right-click your mouse any where in the sequence.
The Curation Options sub-menu appears:

5 Select one of the following:

NOTE: The changes may not be apparent in the Genomic Axis Annotation View.

- Set Longest Open Reading Frame — calculates the longest open reading frame from Stop to Stop codon.
- Set Longest ATG to Stop — calculates the longest open reading frame from Start to Stop codon.
- Stop Codon

EVALUATING SUBJECT SEQUENCE ALIGNMENTS

- 1 If the SubView is not displayed, select Views|Display SubViews when available.
- 2 In the Genomic Axis Annotation View, click on a feature to select it.

NOTE: Query Sequence Alignments View allows you to see nt-nt HSPs and aa-aa HSPs in color.

- 3 Select the Subject Sequence Alignments tab.
The Genome Browser displays the Celera data hits along the subject sequence's axis.

DELETING A FEATURE FROM THE WORKSPACE

To delete a transcript, exon, and gene from the Workspace:

- 1 Click the feature to select it, and right-click your mouse.
- 2 Select Delete Transcript (or Exon) from Workspace.

EXPORTING ANNOTATION DATA

When you export the data, the Genome Browser saves the Workspace file as XML in the Celera Exchange File Format. You can then reload this file at a later time to modify the annotation data.

- 1 From the Main Menu, select File|Save Workspace .
- 2 In the Save dialog box, browse to locate the desired folder.
- 3 Type a file name, and click Save.

A dialog box appears asking whether you want to clear the workspace.

Appendix A. Exchange File Format

<u>Introduction</u>	<u>A.2</u>
<u>XML Markup Element Descriptions</u>	<u>A.3</u>
<u>Document Type Definition</u>	<u>A.10</u>
<u>Example Feature File</u>	<u>A.13</u>

INTRODUCTION

The Celera Data Model is based on the concept of data objects with identity. This means that every piece of genomic data has a permanent independent existence and can be referenced via a unique object id, known as a UID. Data objects come in various types, which can form relationships with other data objects via references to their UIDs.

UIDs are alphanumeric strings that can uniquely identify any data object created. Since they are by definition unique over all data sources and over all time, UIDs allow a data object to reference any other data object, even when they are located in separate data storage media or created at different times.

Not all data providers have access to a common source of unique identifiers, so the UID format is broken into parts separated by a colon: an alphanumeric prefix, called the UID Space, and a decimal suffix which represents an integer less than 2^{64} power. The prefix, which is case-insensitive, identifies the UID space and represents the authority guaranteeing the uniqueness of the suffix integer. UIDs generated by the Celera UID server, for example, all have the “CELERA” prefix. For example:

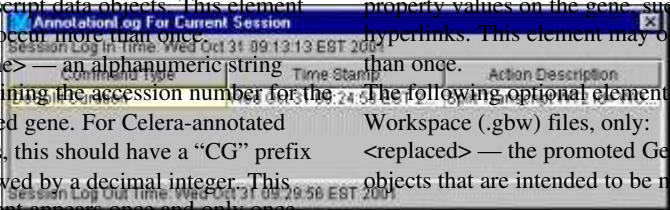
CELERA:nnnnn

You can open data objects from other sources with unique identifiers into the Genome Browser without fear of UID collisions, simply by picking a unique UID space prefix. The Genome Browser can open data objects with up to 2^{16} distinct UID spaces simultaneously. This UID specification is consistent with the XML attribute definitions of ids and idrefs that can fail to pass a validating parser if they are simple numeric strings.

XML MARKUP ELEMENT DESCRIPTIONS

The Celera Exchange File Format is based on a subset of the GAME DTD, with a small number of minor extensions. The following table describes the syntax of each XML element, or tag, and any file (.gbf, or .gbw) restrictions.

XML Markup Descriptions		
Element and Description	Attributes and Elements	
	Required	Optional
<game> The root node, or document element, for a valid Celera Exchange Format file.	ATTRIBUTES: version — the version of game DTD to which this document conforms.	ATTRIBUTES: assembly_version — the version of the assembly data in the database. This attribute is for feature (.gbf) and workspace (.gbw) files that import data on top of the database. taxon — the scientific name of the species of organism with which the data is associated. ELEMENTS: <annotation> <computational_analysis> <feature_set> <seq> — a single feature (.gbf) or workspace (.gbw) file may contain any number of elements of type “SUBJECT”.
<annotation> This corresponds directly to a Gene data object, and is used to represent a human-curated gene.	ATTRIBUTES: id — the UID for the gene. It must appear once and only once. ELEMENTS: <feature_set> — represents Transcript data objects. This element may occur more than once. <name> — an alphanumeric string containing the accession number for the curated gene. For Celera-annotated genes, this should have a “CG” prefix followed by a decimal integer. This element appears once and only once. Annotations saved out in (.gbw) files have <name> elements with “WG” prefix followed by decimal integer, representing workspace genes generated on the client. The Genome Browser automatically generates these elements.	ELEMENTS: <description> — descriptive information about the curation, such as similarity with a known gene. <property> — stores arbitrary property values on the gene, such as URL hyperlinks. This element may occur more than once. The following optional element is for Workspace (.gbw) files, only: <replaced> — the promoted Gene objects that are intended to be modified.

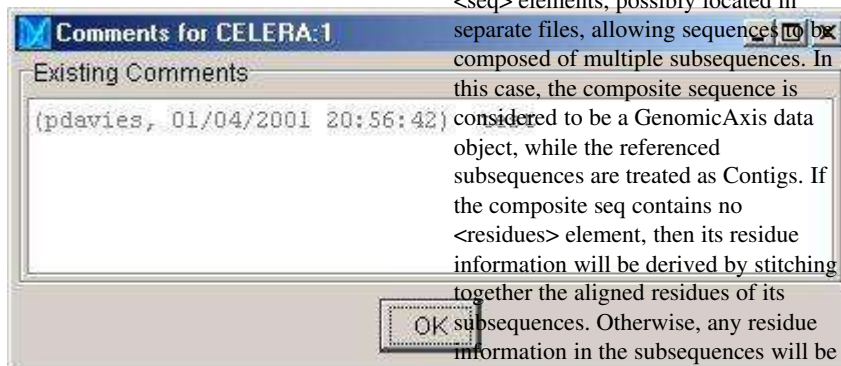


XML Markup Descriptions

Element and Description	Attributes and Elements	Optional
<p><code><computational_analysis></code> This represents a group of precomputed features generated by a particular algorithm or analysis program.</p>	<p>Required</p> <p>ATTRIBUTES: id — stores a UID, which can be used by evidence relationships that refer to an entire computational algorithm, rather than an individual result set.</p> <p>ELEMENTS: <code><result_set></code> — Represents the precomputed features. <code><program></code> — This must contain a name, which is used by the Genome Browser to establish a horizontal tier for displaying the precomputed features.</p>	<p>ELEMENTS: <code><date></code> — indicates when the program was run. <code><database></code> — indicates against which database the analysis program was run. <code><parameter></code> — represents the type of algorithm (e.g. blastx, blastn) used by each run of the program.</p>
<p><code><feature_set></code> This corresponds directly to a Transcript data object and is used to describe a human-curated transcript. Feature sets contained within <code><annotation></code> elements correspond to Transcript data objects that have been assigned to a specific Gene data object. However, <code><feature_set></code> elements can also exist directly under a <code><gene></code> element, in which case they describe Transcript data objects not assigned to any Gene data object.</p>	<p>Attributes and Elements</p> <p>ATTRIBUTES: id — the UID for the transcript.</p> <p>ELEMENTS: <code><type></code> — must contain the string “transcript” and must occur once and only once. <code><name></code> — an alphanumeric string containing the accession number for the curated transcript. For Celera-annotated transcripts, this should have a “CT” prefix followed by a decimal integer. Feature_sets saved out in (.gbw) files have <code><name></code> elements with “WT” prefix followed by decimal integer, representing workspace transcripts generated on the client. <code><feature_span></code> — represents the Exon, Start_Codon, Stop_Codon, and Translation_Start_Position data objects that make up the transcript. This element may occur more than once.</p>	<p>ELEMENTS: <code><replaced></code> — refers to the promoted Transcript data objects that are intended to be modified. This element is for Workspace (.gbw) files only, and it may occur more than once.</p>

XML Markup Descriptions

Element and Description	Attributes and Elements	Optional
<p><seq> This represents a piece of nucleotide sequence against which <result_span> and <feature_span> elements may be aligned. In a feature (.gbf) or a workspace (.gbw) file, the <seq> element may represent some other subject sequence against which a feature is aligned.</p>	<p>Required</p> <p>ATTRIBUTES:</p> <p>id — The UID for the data object.</p> <p>type — Use one of the following values:</p> <ul style="list-style-type: none"> • DNA • RNA • AA 	<p>Optional</p> <p>ATTRIBUTES:</p> <p>length — the number of nucleotides in the sequence.</p> <p>ELEMENTS:</p> <p><residues> — the raw sequence data stored as characters.</p> <p><dbxref> — the database source for this sequence data.</p> <p><description> — general information. The following optional may be used more than one time:</p> <p><seq_alignment> — refers to other <seq> elements, possibly located in separate files, allowing sequences composed of multiple subsequences. In this case, the composite sequence is considered to be a GenomicAxis data object, while the referenced subsequences are treated as Contigs. If the composite seq contains no <residues> element, then its residue information will be derived by stitching together the aligned residues of its subsequences. Otherwise, any residue information in the subsequences will be ignored.</p>
<p><feature_span> This describes simple human-curated features that can be combined to make a feature set. The <feature_span> element may not stand alone. It must be contained within a <feature_set> element.</p>	<p>ATTRIBUTES:</p> <p>id — the UID for the data object.</p> <p>ELEMENTS:</p> <p><seq_relationship> — indicates the <feature_span> element's location on an assembly sequence.</p> <p><type> — contains one of the following values:</p> <ul style="list-style-type: none"> • exon • start-codon • stop-codon • frame <p>These values correspond to the Exon, Start_Codon, Stop_Codon and Translation_Start_Position data objects.</p>	<p>ELEMENTS:</p> <p><evidence> — refers to precomputed results on which the feature span is based.</p> <p><replaced> — refers to the promoted data objects that are intended to be modified. This element is for workspace (.gbw) files only, and it may occur more than once.</p>



XML Markup Descriptions

Element and Description

<seq_relationship>
This describes the geometric relationship between a <feature_span> or <result_span> element and its <seq> element.

<result_set>
This represents a compound-precomputed feature data object, and is a precomputed analog to the <feature_set> element for human-curated features.

<result_span>
This describes simple precomputed features that can be combined to make a feature set , and is a precomputed analog of the <feature_span> element. The <result_span> element may not stand alone. It must be contained within a <result_set> element.

Attributes and Elements

Comments for WORKSPACE:5

New Comments

Optional

ATTRIBUTES:

id — a UID reference to the <seq> element representing the GenomicAxis data object to which it is aligned.

type — contains one of the following values:

ELEMENTS:

<alignment> — contains a sequence string, formatted with gaps and spaces that allow it to be aligned to other <seq_relationship> elements.

Existing Comments

Test comment

ELEMENTS:

 — defines the start and end points on the sequence.

ATTRIBUTES:

id — the UID for the precomputed feature.

ELEMENTS:

<result_span> — represents a simple precomputed feature from which the compound feature is composed. It may occur more than once.

<type> — represents the type of analysis tool used to generate the feature (e.g. Genscan, Blast).

<description> — describes information such as similarity to specific proteins.

<score> — indicates the quality of the match.

<dbxref> — refers to model organism database where the annotations are generated and maintained. This element may occur more than once.

<output> — Of particular interest are outputs of type “expect”. This element may occur more than once.

ATTRIBUTES:

id — the UID for the data object.

ELEMENTS:

<span_type> — the type of data object (e.g., HSP).

<seq_relationship> — positions the <result_span> on a <seq> element.

Seq relationships of type “subject” and “query” refer to spans on a subject and query sequences respectively

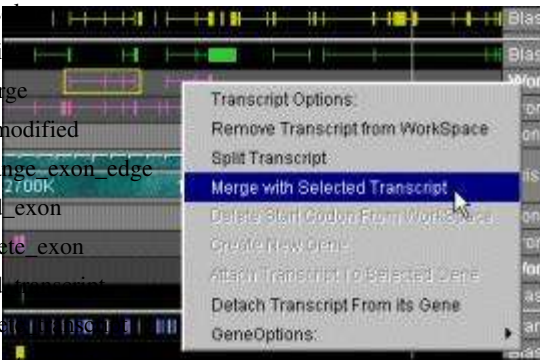
<score> — indicates the quality of the match. This element may occur more than once.

<output> — This takes sub-elements <type> and <value>, when type is “Expect” <value> is the expect value. This element may occur more than once.

XML Markup Descriptions		
Element and Description	Attributes and Elements	
	Required	Optional
<p><seq_alignment></p> <p>This indicates that the containing <seq> should derive a portion of its sequence from a referenced <seq> element.</p>	<p>ATTRIBUTES:</p> <p>seq_id — references the UID of the subsequence <seq> element.</p>	<p>ATTRIBUTES:</p> <p>start and end — indicate the range and orientation on the composite sequence where the subsequence is to be aligned</p> <p>entity_start and entity_end — the corresponding range and orientation on the subsequence.</p> <p>file_path — the path string for an assembly (.gba) file that contains the subsequence. If no file is specified, the subsequence is assumed to be in the same file as the composite sequence.</p>

XML Markup Descriptions

Element and Description	Attributes and Elements	
	Required	Optional
<property> This stores a name-value attribute pair representing some piece of information attached to a specific instance of a data object (e.g., a URL). The properties are displayed in the Genome Browser in the Property Panel, and properties containing valid URLs can be used to hyperlink via a web browser.	ATTRIBUTES: name value	ATTRIBUTES: editable — indicates whether the attribute can be modified by the user, and is by default “false”. Only human-curated features (e.g., Gene, Transcript) may have modifiable properties. ELEMENTS: <property> — enables hierarchies of <property> elements to any depth.
<comments> This stores a single comment text. An <annotation> element may contain multiple comments.		ATTRIBUTES: author date
<replaced> This is used only by human-curated features in workspace (.gbw) files.	ATTRIBUTES: id — the UID of the reference to a promoted human-curated feature, that is intended to be modified. type — indicates the nature of the modification. Use one of the following values: <ul style="list-style-type: none">• update• observed• split• merge• unmodified• change_exon_edge• add_exon• delete_exon• add_transcript• delete_transcript• set_start_codon• remove_start_codon• changed_properties• added_comment	



```
<?xml encoding='UTF-8' ?>

<!ELEMENT game      ((annotation | computational_analysis |
feature_set | seq )*)>
<!--ATTLIST game          version           CDATA    #IMPLIED
                        assembly_version     CDATA    #IMPLIED
                        taxon                 CDATA    #IMPLIED -->
<!--ELEMENT annotation    (name , description? ,
annotation_source? , comments* , feature_set+ , replaced*
property* )>
<!--ATTLIST annotation    id CDATA    #REQUIRED -->
<!--ELEMENT computational_analysis    (date? , program ,
version? , parameter* , database? , result_set+ )>
<!--ATTLIST computational_analysis    id CDATA    #REQUIRED -->
<!--ELEMENT feature_set    (name , type , annotation_source? ,
author? , creation_date? , version? , replaced* , property*
, feature_span+ )>
<!--ATTLIST feature_set    id CDATA    #REQUIRED -->
<!--ELEMENT seq    (dbxref? , description , residues? ,
seq_alignment* )>
<!--ATTLIST seq          length CDATA    #IMPLIED
                        id       CDATA    #REQUIRED
                        type      CDATA    #REQUIRED -->
<!--ELEMENT name        (#PCDATA )>

<!--ELEMENT description    (#PCDATA )>

<!--ELEMENT date EMPTY>
<!--ATTLIST date          day       CDATA    #REQUIRED
                        year        CDATA    #REQUIRED
                        month       CDATA    #REQUIRED -->
<!--ELEMENT progra•    (#PCDATA )>

<!--ELEMENT version      (#PCDATA )>

<!--ELEMENT parameter    (type , value )>

<!--ELEMENT database    (name , date? , version? )>
<!--ATTLIST database    id CDATA    #IMPLIED -->
<!--ELEMENT result_set    (description? , type? , score? ,
dbxref* , output* , result_span* , property* )>
<!--ATTLIST result_set    id CDATA    #REQUIRED -->
<!--ELEMENT type        (#PCDATA )>

<!--ELEMENT author      (#PCDATA )>

<!--ELEMENT creation_date    (#PCDATA )>

<!--ELEMENT feature_span    (type , annotation_source? ,
seq_relationship , property* , replaced* , evidence* )>
<!--ATTLIST feature_span    id CDATA    #REQUIRED -->
<!--ELEMENT dbxref    (xref_db , db_xref_id , version? )>
```

```

<!ELEMENT value      (#PCDATA )>

<!ELEMENT score      (#PCDATA )>

<!ELEMENT output     (type , value )>

<!ELEMENT result_span (span_type? , score? , output* ,
value? , seq_relationship* , property* )>
<!ATTLIST result_span id CDATA #REQUIRED >
<!ELEMENT seq_relationship (span , alignment? )>
<!ATTLIST seq_relationship id CDATA #REQUIRED
                        type (query | subjct ) #IMPLIED >
<!ELEMENT evidence     (#PCDATA )>
<!ATTLIST evidence type (homology | gene_prediction )
#IMPLIED >
<!ELEMENT evidence_options (result CDATA #IMPLIED
                        (use_to_set_left_edge_of_current_curation? )
                        (use_to_set_right_edge_of_current_curation? )
                        (generate_evidence_for_next_curation? ))
<!ELEMENT xref_db      (#PCDATA )>
<!ELEMENT xref_id      (#PCDATA )>
<!ELEMENT span         (start , end )>
<!ELEMENT alignment    (#PCDATA )>
<!ELEMENT start        (#PCDATA )>
<!ELEMENT end          (#PCDATA )>

<!ELEMENT replaced     (#PCDATA )>
<!ATTLIST replaced type (unmodified |
                        modified |
                        obsolete |
                        new |
                        split |
                        merge |
                        deep-modified ) #REQUIRED
id CDATA #REQUIRED >
<!ELEMENT property     (property* )>
<!ATTLIST property name CDATA #REQUIRED
value CDATA #REQUIRED
editable (true | false ) #IMPLIED >
<!ELEMENT seq_alignment (#PCDATA )>
<!ATTLIST seq_alignment start CDATA #IMPLIED
end CDATA #IMPLIED
entity_start CDATA #IMPLIED
entity_end CDATA #IMPLIED
file_path CDATA #IMPLIED
seq_id CDATA #REQUIRED >
<!ELEMENT residues     (#PCDATA )>
<!ELEMENT comments     (#PCDATA )>
<!ATTLIST comments author CDATA #IMPLIED
date CDATA #IMPLIED >

```

<!ELEMENT annotation_source (#PCDATA)>



EXAMPLE FEATURE FILE

This section provides an example Celera Exchange Format feature (.gbf) file.

There are additional example files on the Genome Browser download page:

```
<game version="1.001">
  <!-- transport -->
  <computational_analysis
id="CLIENT:155000040997423">
  <date year="100" day="12" month="8"></date>
  <program>
    Test_Program
  </program>
  <version>
    Test
  </version>
  <parameter>
    <type>
      Sim4
    </type>
    <value>
      /work/chrom/tools/sim4/bin/sim4 none
given none given A=3
    </value>
  </parameter>
  <database>
    <name>
      CHGI_V3_TA.fasta
    </name>
    <version>
      Test
    </version>
  </database>
  <result_set id="CLIENT:155000040991938">
    <property name="link" value="http://
www.celera.com/" editable="true">
    </property>
    <description>
      CRA|335000014763425 /altid=TA|67966 /
dataset=chgi_v3 /def=NOT ASSIGNED /taxon=9606 /org=Homo
sapiens
    </description>
    <parent>
      <type>
        blastn_parent
      </type>
      <value>
        155000040915142
      </value>
    </parent>
    <score>
      0
    </score>
  </result_span id="CLIENT:155000040991939">
```

```

        <span_type>
            MSSP
        </span_type>
        <seq_relationship type="query"
id="CELERA:285347054">
            <span>
                <start>
                    147743
                </start>
                <end>
                    149268
                </end>
            </span>
            <alignment>

TGACACACACCAGGTTAATATCTTTAATATATACCGAGCTCATACAAATTTATCTGACCC
CAGAAGATAGAGCAAAGGACACTGACAAATCCACAAAGGAGGAAATACAATCGCTCCACAT
GAAAGATGTTCAACCGTGCTAGAAAGTCAGTGAAATGCCAACTGAAACACGATGCTATTTT
TTCATTATCAAATTAGCAAATATTTAGCCAGATGATAATTTCCCAATACGGGCAAGATGGA
CATGTTTATCCTTGCTGGTGGCAGTGTAAGGGGATACAAATATTTCCGAAAACCATTTTG
TCAATATGTAGTAAAAGCCTCAAAGGTTTTCATATTCTTTAGTTGAAGAATCCACCTCC
TGGAGTCTACCTAATAACTTGACATTGGGAAACAGCCTGTGCACAAAGATATTCATCAA
TTTATAACTGAAAAGTTAGTAACAGCATTAGTGATTGGCAACTGGCACAATGGTCCATCT
AGTCAATGGGAGAGTACACAGCCACTCCAAAATAAAATATTTAGTAATCATAGGGGAGA
TGCCACGATATGCTAAGTGAACAGCTGCACAAACTGTATCATTACAGAAGACAGCA
ACAAAACATATGTTAAAACAAAAGCAAGAAACCATAGTTGGGGGGGAAAGCCTGAGAGA
CACCAAAATGTGATGTTTTATTTCTTCTTTCTCTACTTGCAAAAATTTATCTAACATG
TGTTTTACATGGGTAATGAGAAGACCTAGTAGGTTTGGTGCTTGTGTATGCAAAGGCATG
AGGCACACGGAAGTGAGGAGCTGGGTTTCTCCCATCTCTACCGCATCTGCGCTGTGCCA
ACTGCCACCCACCCAACTCAGGTCCCAAGAGACAGAGACACAGGGGCTGTCTGTTCC
ACGTGGAAGGCCCTACACCCCAACCTTCTCCCTACTCTGGGGAGCTTCTCTGTTGGCC
CAACAGTCTGCTCTTGCTTCCCTTACCAGGAGACCCACCTTGTGCCTACCTCTCTAAGT
CCCTAAACCAGTGCTTTCCAAGCAGAGGTGCCATGAGCCTTACAACGTGTTGTCAGATG
GAGGCGCCTCCATTAGGATCTCTTCCCTCTTCCACCCAGAGCTCTGCCCCAGTCAGTGG
CTGCAGCATGGGCCCTTTTGTCCAATCCCAGGGCTTGTTCCTAAGTCAACCTCAGCACA
TGCCCAGCTCAGCCATGTCCCTTCACTCCCTTACAGGCTGGAAGTGCAGGGTGGTGCCC
ACCCAAGGCGGCTGTCTGCCATGACAACAGGAGCGTGTGCGTCAGGCTGGTCTGCCGCA
CTTTCTGGGTTTTGTTTTCCACTCCTCCTCATTGCTCATCACATACTTACCTGGGGACAA
TGAAATAGGTCTTTGCTATTCCCTAGAAGTCACCAAAATGCTTTAGGACATTAAGTTAGT
CCCAGGGTCACACTATACTAGGCCAGGGAGTCACCCCTTCACTAACTCCCATCTCCAGTG
ATACCTGGTATGCACTGAAGCCTA

            </alignment>
        </seq_relationship>
        <seq_relationship type="subj"
id="CLIENT:335000014763425">
            <span>
                <start>
                    9
                </start>
                <end>
                    1534
                </end>
            </span>
            <alignment>

TGACACACACCAGGTTAATATCTTTAATATATACCGAGCTCATACAAATTTATCTGACCC

```

```

CAGAAGATAGAGCAAAGGACACTGACAATCCACAAAGGAGGAAATACAATCGCTCCACAT
GAAAGATGTTCAACCGTGCTAGAAAGTCAGTGAAATGCCAACTGAAACACGATGCTATTTT
TTCATTATCAAATTAGCAAATATTTAGCCAGATGATAATCCCAATACGGGCAAGATGGA
CATGTTTATCCTTGCTGGTGGCAGTGTAAAGGGATACAAATATTTCCGGAACCAATTTTG
TCAATATGTAGTAAAGCCTCAAAGGTTTTCATATCTTTAGTTGAAGAATCCCACCTCC
TGGAGTCTACCCTAATAACTTGACATTGGGAAACAGCCTGTGCACAAAGATATTCATCAA
TTTATAACTGAAAAGTTAGTAACAGCATTAGTGATTGGCAACTGGCACAATGGTCCATCT
AGTCAATGGGAGAGTACACAGCCACTCCAAAATAAAATATTTAGTAATCATAAGGGGAGA
TGCCACAGTATGCTAAGTGAAAACAGCTGCACAAAACGTATCATTACAGAAGACAGCA
ACAAAACATATGTTAAACAAAAGCAAAGAAACCATAGTTGGGGGGGAAAGCCTGAGAGA
CACAAAATGTGATGTTTATTTCTCTTCTTCTCTACTTGCAAAAATTTATCTAACATG
TGGTTTACATGGGTAAAGAGAAGACCTAGTAGGTTGGTGCTTGTTGATGCAAGGCATG
AGGCACACGGAAGTGAGGAGCTGGGTTTCTCCCATCTCTACCGCATCTGCGCTGTGCCA
ACTGCCACCCCAACCAACTCAGGTCCCCAAGAGACAGAGACACAGGGGCTGTCTGTGCC
ACGTGGAAGGCCCTACACCCCAACCTTCTCCCTACTCTGGGGAGCTTCTCTGTGGCC
CAACAGTCTGCTCTTCTTCCCTTACCCAGGAGACCCACCTTGTGCTTACCTCTCTAAGT
CCCTAAACAGTGCTTTCCAAGCAGAGGTGCCATGAGCCTTACAACTGTGGTTGCAGATG
GAGGCGCCTCCATTAGGATCTCTTCCCTCTTCCACCCAGAGCTCTGCCCCCAGTCAGTGG
CTGCAGCATGGGCCCTTTTGTCCAATCCAGGGCTTGTTCCTAAGTCAACCTCAGCACA
TGCCAGCTCAGCCATGTCCCTTCACTCCCTTACAGGCTGGAAC TGCCAGGGTGGTGCCC
ACCCAAGGCGGCTGTCTGCCATGACAACAGGAGCGTGTGCGTCAGGCTGGTCTGCCGCA
CTTTCTGGGTTTTGTTTTCCACTCCTCTCATTGCTCATCATACTTACCTGGGGACAA
TGAAATAGGTCTTTGCTATTCCCTAGAGTCACCAAAATGCTTTAGGACATTAAGTTAGT
CCCAGGTACACTATACTAGGCCAGGGAGTCACCCCTTCACTAACTCCCATCTCCAGTG
ATACCCTGGTATGCACTGAAGCCTA

```

```

    </alignment>
  </seq_relationship>
</result_span>
</result_set>
<result_set id="CLIENT:155000040991940">
  <property name="link" value="http://
www.celera.com/" editable="true">
    </property>
    <description>
      CRA|335000014716891 /altid=TA|21432 /
dataset=chgi_v3 /def=NOT ASSIGNED /taxon=9606 /org=Homo
sapiens
    </description>
    <parent>
      <type>
        blastn_parent
      </type>
      <value>
        155000040915145
      </value>
    </parent>
    <score>
      0
    </score>
    <result_span id="CLIENT:155000040991941">
      <span_type>
        MSSP
      </span_type>
      <seq_relationship type="query"
id="CELERA:285347054">
        <span>

```

```

        <start>
            142075
        </start>
        <end>
            141452
        </end>
    </span>
    <alignment>

AATCTCAGTATTTTATAAATTGTAAAGTTATTTTCCGTATCTCTTGGACATAATGGTAT
TTTTGTGACTGTGATATTTTGCATTTTATTTCTTCTCTCACAGCGTCAGTGGAGGTCA
GCAGGAGGTCCcTGCAGgCTTGCACTGCCCTGCTGGTTATCACTGCTGGTGACTCATGCA
GCCAGCCTGCTGATGCCCTGAGCTGGCCTGTTTCACTTGGTGTCAGAGGGCTCTGCTGT
GGGCCCCGGGGGAGGGAACACGCACCTTCCTCTGTGGAACGTGGCTGCTTTCCCCCTT
CCTCATCTGTTCTTTCATCTGCCCCAGCCTGTGAAGGGAACCTCGTGCCTGGGCAGGC
TTATGGGTGCCGGGAGGCTGGGGAGCTGGcTCACGCTCTGAGTGGGCTTCCCTGGCCAGT
CCTGTGCCCCATCACCAGCCTCGCTCTGTGGTCTTTGCCCTGGTGCACTGGGCGGTACCA
CTTCACCACGTCTTTTCAGTGGTCCTCCGTGTGTTCCAAAGCTTCTGGATTTCAAATGTT
GCTGTGCTCACAGAAACCTGATGGTTTACACACCAAGCAATACCACCACCACCACCGCCA
CCACCACAACAACAAAAA

        </alignment>
    </seq_relationship>
    <seq_relationship type="sjct"
id="CLIENT:335000014716891">
        <span>
            <start>
                623
            </start>
            <end>
                0
            </end>
        </span>
        <alignment>

AATCTCAGTATTTTATAAATTGTAAAGTTATTTTCCGTATCTCTTGGACATAATGGTAT
TTTTGTGACTGTGATATTTTGCATTTTATTTCTTCTCTCACAGCGTCAGTGGAGGTCA
GCAGGAGGTCCcTGCAGgCTTGCACTGCCCTGCTGGTTATCACTGCTGGTGACTCATGCA
GCCAGCCTGCTGATGCCCTGAGCTGGCCTGTTTCACTTGGTGTCAGAGGGCTCTGCTGT
GGGCCCCGGGGGAGGGAACACGCACCTTCCTCTGTGGAACGTGGCTGCTTTCCCCCTT
CCTCATCTGTTCTTTCATCTGCCCCAGCCTGTGAAGGGAACCTCGTGCCTGGGCAGGC
TTATGGGTGCCGGGAGGCTGGGGAGCTGGCTCACGCTCTGAGTGGGCTTCCCTGGCCAGT
CCTGTGCCCCATCACCAGCCTCGCTCTGTGGTCTTTGCCCTGGTGCACTGGGCGGTACCA
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<u>Troubleshooting</u>	<u>B.2</u>
<u>Memory Problems</u>	<u>B.2</u>
<u>Data Access Problems</u>	<u>B.2</u>
<u>Exception Errors</u>	<u>B.3</u>

TR O U B L E S H O O T I N G

This appendix contains answers to some common troubleshooting questions.

If after reviewing this appendix, you still can't solve your issue, you should contact your Celera Technical Support Specialist.

M e m o r y P r o b l e m s

In general, you can solve many issues by ensuring that you have allocated adequate memory for the Genome Browser. For instructions, please refer to "Troubleshooting Performance Issues" on page 1-5.

D a t a A c c e s s P r o b l e m s

If you are not properly logged in, you will receive the following message when you attempt to open an information service:

You will need to restart the Genome Browser and re-enter your login information. For instructions, please refer to "Setting Preferences" on page 2-6.

Exception Errors

If the Genome Browser experiences an exception error, an error dialog box will appear that allows you to send a message to the Celera support staff. If this happens, you should:

- 1 Click Send email to Celera Support.
- 2 Type your e-mail address in the box provided, and click OK.
- 3 Type a description of what you were doing when the exception occurred in the box provided, and click OK.
A message appears indicating that your problem was sent to the Celera Support staff.
- 4 Click OK.
A Celera Support representative will contact you.

NOTE: If you prefer to be contacted by telephone, you should include your name and telephone number in the description box.

