Package 'BigSyn'

March 18, 2020

· · · · · · · · · · · · · · · · · · ·	10
	11
daniRules	12
donors.receptors.check	13
drop_last	13
fitmodel.ctree	14
fitmodel.fn	15
fitmodel.rf	15
fitthemodel	16
GeneralReversetransposefunction	17
GeneralReversetransposefunctiondecoupe	18
Generaltransposefunction	18
	19
getnodesfromrules	20
	20
* • •	21
	22
-	22
6 -	23
	24
	25
$\mathcal{E} = 1$	26
	26
	27
	28
	28
	29
$\epsilon =$	30
	31
	32
	33
	33
	34
	35
••	35
predictor.matrix.default	
predictor.matrix.rate	
	37
• • •	38
-	40
1 —	41
1	42
1	43
1	43
1	44
1 1	 45
	47
	49

augmenti	naxT_f																	3
	Sparameters.va																	
	treedepth																	51
	treetopdf																	52
	Tsampledata .																	52
	TTsampledata															 		53
	%notin%																	
Index																		55

augmentmaxT_f

Creates cell marginal max.

Description

Creates cell marginal max.

Usage

```
augmentmaxT_f(.data, variables, verbose = getOption("verbose"))
```

Arguments

. data a dataframe

variables a vector of character strings

Details

Assume one runs the program

 $augmentmax T_f(.dataBigSyn::STtableA1, variables = c("AA.present")). \label{eq:control} The program looks for all the "cell variables" corresponding to "AA.present", by using the function BigSyn::get_var$

The results is this:

AA.present_La_La_Lrn1, AA.present_La_Lb_Lrn1, AA.present_La_Lc_Lrn1,

...

AA.present_Lc_La_Lrn4, AA.present_Lc_Lb_Lrn4, AA.present_Lc_Ld_Lrn4

The programs computes the number of marginal variables with the function BigSyn::get_cellXXmarginscount. Here it is 3

The program creates the following character matrix, named patterns:

```
"1" "La" ""
"1" "Lb" ""
"1" "Lc" ""
"2" "La_La" "La"
"2" "La_Ld" "La"
"2" "Lb_Lb" "Lb"
```

4 augment $maxT_f$

```
"2" "Lc_Lb" "Lc"
"2" "Lc Ld" "Lc"
"2" "La Lb" "La"
"2" "La Lc" "La"
"2" "Lb La" "Lb"
"2" "Lb Lc" "Lb"
"2" "Lb Ld" "Lb"
"2" "Lc Lc" "Lc"
"3" "La_La_Lrn1" "La_La"
"3" "La_Ld_Lrn1" "La_Ld"
"3" "Lb_Lb_Lrn1" "Lb_Lb"
"3" "Lc_La_Lrn1" "Lc_La"
"3" "Lc_Lb_Lrn1" "Lc_Lb"
"3" "Lc_Ld_Lrn1" "Lc_Ld"
"3" "La_Lb_Lrn1" "La_Lb"
"3" "La_Lc_Lrn1" "La_Lc"
"3" "Lb_La_Lrn1" "Lb_La"
"3" "Lb_Lc_Lrn1" "Lb_Lc"
"3" "Lb_Ld_Lrn1" "Lb_Ld"
"3" "Lc_Lc_Lrn1" "Lc_Lc"
"3" "La_La_Lrn2" "La_La"
"3" "La_Ld_Lrn2" "La_Ld"
"3" "Lb_Lb_Lrn2" "Lb_Lb"
"3" "Lc_La_Lrn2" "Lc_La"
"3" "Lc_Lb_Lrn2" "Lc_Lb"
"3" "Lc_Ld_Lrn2" "Lc_Ld"
"3" "La_Lb_Lrn2" "La_Lb"
"3" "La_Lc_Lrn2" "La_Lc"
"3" "Lb_La_Lrn2" "Lb_La"
"3" "Lb_Lc_Lrn2" "Lb_Lc"
"3" "Lb_Ld_Lrn2" "Lb_Ld"
```

"3" "Lc_Lc_Lrn2" "Lc_Lc"
"3" "La_La_Lrn3" "La_La"
"3" "La_Ld_Lrn3" "La_Ld"
"3" "Lb_Lb_Lrn3" "Lb_Lb"
"3" "Lc_La_Lrn3" "Lc_La"

"2" "Lc_La" "Lc"

augmentmaxT_f 5

```
"3" "Lc_Lb_Lrn3" "Lc_Lb"
"3" "Lc_Ld_Lrn3" "Lc_Ld"
"3" "La_Lb_Lrn3" "La_Lb"
"3" "La_Lc_Lrn3" "La_Lc"
"3" "Lb_La_Lrn3" "Lb_La"
"3" "Lb Lc Lrn3" "Lb Lc"
"3" "Lb Ld Lrn3" "Lb Ld"
"3" "Lc_Lc_Lrn3" "Lc_Lc"
"3" "La_La_Lrn4" "La_La"
"3" "La_Ld_Lrn4" "La_Ld"
"3" "Lb_Lb_Lrn4" "Lb_Lb"
"3" "Lc_La_Lrn4" "Lc_La"
"3" "Lc_Lb_Lrn4" "Lc_Lb"
"3" "Lc Ld Lrn4" "Lc Ld"
Then for all i in 3:1 (starting with the maximum depth) list the different aggregations to the upper
level to perform. So for i=3, aggregating to the second level will be done by computing the variables
```

AA.contl La La, AA.contl La Ld, AA.contl Lb Lb, AA.contl Lc La,

AA.cont1_Lc_Lb, AA.cont1_Lc_Ld, AA.cont1_La_Lb, AA.cont1_La_Lc,

AA.cont1_Lb_La, AA.cont1_Lb_Lc, AA.cont1_Lb_Ld, AA.cont1_Lc_Lc

For example:

AA.cont1_La_La=rowSums(.data([,c("AA.cont1_La_La_Lrn1", "AA.cont1_La_La_Lrn2", "AA.cont1_La_La_Lrn3", "AA.cont1_La_La_Lrn4"),drop=FALSE])

For i=2 aggregating to the upper level will be done by computing the variables:

AA.contl La, AA.contl Lb, AA.contl Lc

AA.cont1_La =rowSums(.data([,c("AA.cont1_La_La", "AA.cont1_La_Ld", "AA.cont1_La_Lb", "AA.cont1_La_Lb", "AA.cont1_La_Lc"),drop=FALSE])

For i=1 aggregating to the upper level will be done by computing the variable:

AA.cont1_=rowSums(.data([,c("AA.cont1_La", "AA.cont1_Lb", "AA.cont1_Lc"),drop=FALSE])

The computation of the marginal totals is done, the second step is the computation of the marginal ratios.

It is done by looping on the rows of the patterns matrix

Line j of pattern is a length 3 character vector. let call patterns[j,2] x and patterns[j,3] y The programs replaces the variable names paste0("AA.cont1",x) by the ration of the variable paste0("AA.cont1",x) by the variable named paste0("AA.cont1",y).

For example for the line "3" "La_Ld_Lrn3" "La_Ld", the following replacement will be made: AA.cont1_La_Ld_Lrn3=AA.cont1_La_Ld_Lrn3/AA.cont1_La_Ld

The same is applied to all the elements of the input parameter variables.

6 augmentpctT_f

Value

a dataframe.

Examples

```
.data=BigSyn::STtableA1
variable="AA.present"
variables=variable
ASTtableA1
ASTtableA1
ASTtableA1[:5,c("AA.present_","AA.present_La","AA.present_La_Lb")]
xx<-ASTtableA1[sort(grep("present",names(ASTtableA1),value=TRUE))]
xx[xx==0]<-NA
StudyDataTools::ggplot_missing(xx)</pre>
```

augmentpctT_f

Convert cell totals to marginal ratios and create overall total.

Description

Convert cell totals to marginal ratios and create overall total.

Usage

```
augmentpctT_f(.data, variables, verbose = getOption("verbose"))
```

Arguments

.data a dataframe

variables a vector of character strings

Details

Assume one runs the program

 $augmentpctT_f(.dataBigSyn::STtableA1, variables = c("AA.cont1", "AA.cont1")). \label{eq:contact} The program looks for all the "cell variables" corresponding to "AA,cont1", by using the function BigSyn::get_var$

The results is this:

```
AA.cont1_La_La_Lrn1, AA.cont1_La_Ld_Lrn1, AA.cont1_Lb_Lb_Lrn1, AA.cont1_Lc_La_Lrn1, AA.cont1_Lc_Lb_Lrn1, AA.cont1_Lc_Lb_Lrn1, AA.cont1_Lb_Lb_Lrn1, AA.cont1_Lb_Lb_Lrn1, AA.cont1_Lb_Lc_Lrn1, AA.cont1_Lb_Ld_Lrn1, AA.cont1_Lb_Ld_Lrn1, AA.cont1_Lb_Lb_Lrn2, AA.cont1_Lb_Lb_Lrn2, AA.cont1_Lb_Lb_Lrn2, AA.cont1_Lc_La_Lrn2, AA.cont1_Lc_Lb_Lrn2, AA.cont1_Lb_Lb_Lrn2, AA.cont1_Lb_Lb_Lrn2, AA.cont1_Lb_Lb_Lrn2, AA.cont1_Lb_Lb_Lrn2, AA.cont1_Lb_Lb_Lrn2, AA.cont1_Lb_Lb_Lrn2, AA.cont1_Lb_Lb_Lrn2, AA.cont1_Lb_Lb_Lrn3, AA.cont1_Lb_Lb_Lrn4, AA.cont1_
```

augmentpctT_f 7

The programs computes the number of marginal variables with the function BigSyn::get_cellXXmarginscount.

Here it is 3

The program creates the following character matrix, named patterns:

```
"1" "La" ""
"1" "Lb" ""
"1" "Lc" ""
"2" "La La" "La"
"2" "La Ld" "La"
"2" "Lb Lb" "Lb"
"2" "Lc_La" "Lc"
"2" "Lc_Lb" "Lc"
"2" "Lc_Ld" "Lc"
"2" "La Lb" "La"
"2" "La Lc" "La"
"2" "Lb La" "Lb"
"2" "Lb Lc" "Lb"
"2" "Lb_Ld" "Lb"
"2" "Lc_Lc" "Lc"
"3" "La_La_Lrn1" "La_La"
"3" "La_Ld_Lrn1" "La_Ld"
"3" "Lb_Lb_Lrn1" "Lb_Lb"
"3" "Lc_La_Lrn1" "Lc_La"
"3" "Lc_Lb_Lrn1" "Lc_Lb"
"3" "Lc_Ld_Lrn1" "Lc_Ld"
"3" "La_Lb_Lrn1" "La_Lb"
"3" "La_Lc_Lrn1" "La_Lc"
"3" "Lb_La_Lrn1" "Lb_La"
"3" "Lb_Lc_Lrn1" "Lb_Lc"
"3" "Lb_Ld_Lrn1" "Lb_Ld"
"3" "Lc_Lc_Lrn1" "Lc_Lc"
"3" "La_La_Lrn2" "La_La"
"3" "La_Ld_Lrn2" "La_Ld"
"3" "Lb_Lb_Lrn2" "Lb_Lb"
"3" "Lc_La_Lrn2" "Lc_La"
"3" "Lc_Lb_Lrn2" "Lc_Lb"
```

"3" "Lc_Ld_Lrn2" "Lc_Ld"

8 augmentpctT_f

```
"3" "La_Lb_Lrn2" "La_Lb"
"3" "La_Lc_Lrn2" "La_Lc"
"3" "Lb_La_Lrn2" "Lb_La"
"3" "Lb_Lc_Lrn2" "Lb_Lc"
"3" "Lb_Ld_Lrn2" "Lb_Ld"
"3" "Lc Lc Lrn2" "Lc Lc"
"3" "La La Lrn3" "La La"
"3" "La_Ld_Lrn3" "La_Ld"
"3" "Lb_Lb_Lrn3" "Lb_Lb"
"3" "Lc_La_Lrn3" "Lc_La"
"3" "Lc_Lb_Lrn3" "Lc_Lb"
"3" "Lc_Ld_Lrn3" "Lc_Ld"
"3" "La_Lb_Lrn3" "La_Lb"
"3" "La_Lc_Lrn3" "La_Lc"
"3" "Lb La Lrn3" "Lb La"
"3" "Lb Lc Lrn3" "Lb Lc"
"3" "Lb_Ld_Lrn3" "Lb_Ld"
"3" "Lc Lc Lrn3" "Lc Lc"
"3" "La_La_Lrn4" "La_La"
"3" "La_Ld_Lrn4" "La_Ld"
"3" "Lb_Lb_Lrn4" "Lb_Lb"
"3" "Lc_La_Lrn4" "Lc_La"
"3" "Lc_Lb_Lrn4" "Lc_Lb"
"3" "Lc Ld Lrn4" "Lc Ld"
```

Then for all i in 3:1 (starting with the maximum depth) list the different aggregations to the upper level to perform. So for i=3, aggregating to the second level will be done by computing the variables: AA.contl_La_La, AA.contl_La_Ld, AA.contl_Lb_Lb, AA.contl_Lc_La, AA.contl_Lc_Lb, AA.contl_Lc_Ld, AA.contl_La_Lb, AA.contl_Lb_Lb, AA.contl_Lb_Lb, AA.contl_Lb_Lb, AA.contl_Lb_Lb, AA.contl_Lb_Lb, AA.contl_Lb_Lc, AA

For example AA.cont1_La_La =rowSums(.data([,c("AA.cont1_La_La_Lrn1", "AA.cont1_La_La_Lrn2", "AA.cont1_La_La_Lrn3", "AA.cont1_La_La_Lrn4"),drop=FALSE])

For i=2 aggregating to the upper level will be done by computing the variables : AA.cont1_La, AA.cont1_Lb, AA.cont1_Lc AA.cont1_La=rowSums(.data([,c("AA.cont1_La_La", "AA.cont1_La_Ld", "AA.cont1_La_Lb", "AA.cont1_Lb", "AA.cont1_Lb",

For i=1 aggregating to the upper level will be done by computing the variable AA.cont1_=rowSums(.data([,c("AA.cont1_La", "AA.cont1_Lb", "AA.cont1_Lc"),drop=FALSE])

The computation of the marginal totals is done, the second step is the computation of the marginal ratios.

It is done by looping on the rows of the patterns matrix

augmentT_f

Line j of pattern is a length 3 character vector. let call patterns[j,2] x and patterns[j,3] y The programs replaces the variable names paste0("AA.cont1",x) by the ration of the variable paste0("AA.cont1",x) by the variable named paste0("AA.cont1",y).

For example for the line "3" "La_Ld_Lrn3" "La_Ld", the following replacement will be made: AA.cont1_La_Ld_Lrn3=AA.cont1_La_Ld_Lrn3/AA.cont1_La_Ld

The same is applied to all the elements of the input parameter variables.

Value

a dataframe.

Examples

```
.data=BigSyn::STtableA1
variable="AA.cont1"
variables=variable
STAtableA1
STAtableA1
STAtableA1
STAtableA1
STatableA1
STtableA1
STAtableA1
STATABLE
```

 $augmentT_f$

Creates cell marginal max and percentages.

Description

Creates cell marginal max and percentages.

Usage

```
augmentT_f(.data, variablesmax, variablespct, verbose = getOption("verbose"))
```

Arguments

```
.data a dataframe
```

variablesmax a vector of character strings variablespct a vector of character strings

Details

applys the functions augmentmaxT_f and augmentpctT_f to .data

Value

a dataframe.

10 compilefits

Examples

```
.data=BigSyn::STtableA1
variablesmax="AA.present";variablespct="AA.cont1"
ASTtableA1<-augmentT_f(.data,variablesmax,variablespct,verbose=F)
ASTtableA1[c("AA.present_","AA.cont1_","AA.present_La","AA.cont1_La","AA.present_La_Lb","AA.cont1_La_Lb")]</pre>
```

BigSyn: Some non confidential R functions developped for the MLDSC synthetic data project

Description

The BigSyn package allows to synthesize big hierarchical databases by opposition to just a single small "rectangular" table. The general idea is to - provide tools to transpose the data and back transpose the synthetic version of the transposed data. - provide a synthetisation procedure that runs the modeling and the sampling separately - provide tools to operate a reasonable pre-selection of predictors. - provide tools to visualize the synthetisation.

BigSyn functions

The main BigSyn functions are SDPSYN2 Generaltransposefunction GeneralReversetransposefunction

Shiny application

runCompare() launches a shiny application to

General approach

A step by step example is provided in the Synthesize_database demo file

Examples

demo(Synthesize_database)

compilefits

Save a pdf image of each regression tree grown in the modeling phase and discard useless information

Description

For each element of save parameters, look at the tree and produces the corresponding pdf. It also removes all the information that is stored in the ouptut of parykit::Ctree, e.g. the data. It only keeps the tree and the rules to get it.

compilesamplereports 11

Usage

```
compilefits(
   Sparameters,
   fitmodelsavepath,
   pdfpath = fitmodelsavepath,
   .progress = "text"
)
```

Arguments

Details

Depends on plyr. Partykit output contain all the data that was used to grow the tree. This function removes the unwanted information.

Examples

compilesamplereports compilesamplereports

Description

Sample reports are the output of the function ReportonSample

Usage

```
compilesamplereports(Sparameters, samplereportssavepath)
```

12 daniRules

Arguments

```
Sparameters: a list, that has the same structure than the outputs of samplereportssavepath:

a file path where to store the sample reports
```

Details

depends on plyr

See Also

ReportonSample

Examples

```
y=iris$Species;x=iris[,-5]
partykitctree <- partykit::ctree(y ~ ., data=cbind(y=y,x))</pre>
```

daniRules

daniRules

Description

the rules for the tree. Add an extra rule. For example, if the left branch rule is X in (a,b,c) and the right branch rule is X in (e,f), the right branch rule is modified by X not in (a,b,c). The extreme right branch rule is replaced by the negation of any of the other branches, for each node.

Usage

```
daniRules(x, i = NULL, ...)
```

Arguments

x same format than output of partykit::ctree i (default: NULL)

Details

Adapted from partykit:::.list.rules.party

Value

for each terminal node of the tree, give the definition. For example: node 11 corresponds to 'x>1 and y in ("a","b")'

```
x=partykit::ctree(Petal.Width~.,iris)
daniRules(x)
```

donors.receptors.check 13

```
donors.receptors.check
```

For each leave of each tree, counts the number of donors in the gold set vs the number of receptors in the synthetic dataset.

Description

For each leave of each tree, counts the number of donors in the gold set vs the number of receptors in the synthetic dataset.

Usage

```
donors.receptors.check(Rules, gold.data, syn.data)
```

Arguments

Rules a named list (names correspond to the gold.data and syn.data variable names) of

lists of logical expressions (for each variable, the logical expression corresponds

to a terminal leaf in a tree).

gold.data a data frame

syn.data a data frame containing the same variables than gold.data

Examples

drop_last

Drop last margin position (Trims all strings of a vector of strings after the last "_")

Description

Drop last margin position (Trims all strings of a vector of strings after the last "_")

14 fitmodel.ctree

Usage

```
drop_last(x)
```

Arguments

Х

a vector of character strings

Details

```
if x is "AA.char1_La_Ld_Lrn1" returns "AA.char1_La_Ld", if x contains no "_", returns empty string
```

Value

a vector of character strings

Examples

```
drop_last("AA.char1_La_Ld_Lrn1")
drop_last("iojoij")
drop_last("aa.iojoij")
```

fitmodel.ctree

Function to fit a ctree model.

Description

Function to fit a ctree model.

Usage

```
fitmodel.ctree(x, y, treeplotsavepath = NULL, ...)
```

Arguments

```
x a dataframe of predictors
y a vector :dependent variable
treeplotsavepath:
```

a path to save the graph as a pdf. if NULL, no pdf is saved

Value

a named list of 4 elements: "Rules" a data.frame with two variables: terminalnode (a integer vector) and condition a string that gives the rule for each terminal node. "y" the values of the predictor "terminalnodes" a vector: the terminal nodes for each element of \$y\$. "shortlist" a character string giving the names of the variables in x that were used for the classification

fitmodel.fn 15

Examples

```
fitmodel.ctree(x=iris[,-5],y=iris$Species)
```

fitmodel.fn

Fit a model with a specific function

Description

Fit a model with a specific function

Usage

```
fitmodel.fn(method, x, y, treeplotsavepath = NULL, ...)
```

Arguments

```
method a string. currently only method="ctree" or "rf" (random forest).

x a predictors, a dataframe.

y variable to predict, a vector

treeplotsavepath

a

... synthetic parameters to pass to the right fit model function. the fit model function name is the concatenation of "fit.model" and method
```

Value

a sublist of synparameters, which names are possible arguments of synthpop::syn.ctree if method="ctree".

Examples

```
fitmodel.fn(method="ctree",x=iris[,-5],y=iris$Species,nbuckets=30,tutu="not a good argument")
```

fitmodel.rf

Function to fit a ctree model.

Description

Function to fit a ctree model.

Usage

```
fitmodel.rf(x, y, treeplotsavepath = NULL, ...)
```

16 fitthemodel

Arguments

```
x a dataframe of predictors
y a vector :dependent variable
treeplotsavepath:
a path to save the graph as a pdf. if NULL, no pdf is saved
```

Value

a named list of 4 elements: "Rules" a data.frame with two variables: terminalnode (a integer vector) and condition a string that gives the rule for each terminal node. "y" the values of the predictor "terminalnodes" a vector: the terminal nodes for each element of \$y\$. "shortlist" a character string giving the names of the variables in x that were used for the classification

Examples

```
fitmodel.ctree(x=iris[,-5],y=iris$Species)
```

fitthemodel

Function to fit the model.

Description

Function to fit the model.

Usage

```
fitthemodel(
   Sparameters_i,
   fitmodelsavepath,
   TtableANAto0,
   redocomputationsevenifexists = FALSE,
   treeplotsavefolder = NULL
)
```

Arguments

Sparameters_i an element of the list output from Sparameters.default.f fitmodelsavepath

a folder path. Results will either be read from or stored in this folder. If the file exists, by default it is not replaced.

TtableANAto0 a table containing the predictors without NAs as well as the outcome

Value

a list.

Examples

GeneralReversetransposefunction

General Reverse Transpose function

Description

General Reverse Transpose function

Usage

GeneralReversetransposefunction(TtableA, key)

Arguments

key A list of variables (columns of the transposed table)

table A dataframe

Value

A list: first element of the list is a dataframe, the transposed version of the orioginal table. Second element is a key to allow back transposition

```
data(tableA);data(TtableA);data(XKA);key<-XKA$key
RtableA=GeneralReversetransposefunction(TtableA,key)
ordertableA <-do.call(order,tableA[c(id1,id2)])
orderRtableA<-do.call(order,RtableA[c(id1,id2)])
identical(nrow(tableA),nrow(RtableA))
identical(lapply(tableA,class),lapply(RtableA,class))
identical(tableA[ordertableA,],RtableA[orderRtableA,])
identical(names(tableA),names(RtableA))
all (lapply(names(tableA),function(x){identical(tableA[orderRtableA,x],RtableA[orderRtableA,x])}))</pre>
```

GeneralReversetransposefunctiondecoupe

General Reverse Transpose function with split

Description

General Reverse Transpose function with split

Usage

```
GeneralReversetransposefunctiondecoupe(.data, key, nrowmax = 10000)
```

Arguments

key A list of variables (columns of the transposed table)

table A dataframe

Value

A list: first element of the list is a dataframe, the transposed version of the orioginal table. Second element is a key to allow back transposition

Examples

```
data(tableA);data(TtableA);data(XKA);key<-XKA$key
RtableA=GeneralReversetransposefunctiondecoupe(TtableA,key,10)</pre>
```

Generaltransposefunction

General Transpose function

Description

General Transpose function

Usage

```
Generaltransposefunction(
  tableA,
  id1,
  id2,
  origin = deparse(substitute(tableA))
)
```

Arguments

id1 A list of variables (rows)

id2 A list of variables (columns of the transposed table), id2 can contain as a last

element the strint "rn", if the variable rn is an index for the cells formed by the

variables listed first in id2

table A dataframe

Value

A list: first element of the list is a dataframe, the transposed version of the orioginal table. Second element is a key to allow back transposition

Examples

```
tableA<-sampledata(TRUE)
id1=c("id1a","id1b")
id2=c("id2a","id2b")
TtableA<-Generaltransposefunction(tableA,id1,id2)</pre>
```

Generaltransposefunctionsimple

Simple General Transpose function

Description

Simple General Transpose function

Usage

```
Generaltransposefunctionsimple(tableA, id1, id2)
```

Arguments

tableA A dataframe

id1 A list of variables (rows)

id2 A list of variables (columns of the transposed table)

Value

A data frame

```
tableA<-sampledata(TRUE)
id1=c("id1a","id1b")
id2=c("id2a","id2b")
TtableA<-Generaltransposefunctionsimple(tableA,id1,id2)</pre>
```

getnodesfromrules

Function to get terminal node from a set of partitioning rules and new predictors

Description

Function to get terminal node from a set of partitioning rules and new predictors

Usage

```
getnodesfromrules(x, Rules)
```

Arguments

x a dataframe of predictors

Rules a data frame containing 2 character variables: "terminalnode" and "condition"

Value

a vector of lenth the number of rows of x indicating the terminal nodes.

Examples

```
getnodes from rules (x=iris[1:3,-5], Rules=fit model.ctree (x=iris[,-5], y=iris\$Species)\$ Rules)
```

```
getpredictorsfromcaptureoutput
```

getpredictorsfromcaptureoutput

Description

for each terminal node of the tree, give the elements of "predictors" who appear in the node definition

Usage

```
getpredictorsfromcaptureoutput(tree, predictors)
```

Arguments

tree same format than output of partykit::ctree

predictors a vector of character strings, indicating variable names.

Details

A basic 'grep' function is applied to each potential predictor. It is returned if it appears in the rules.

getpredictorsfromtree 21

Value

a named vector of booleans. The names correspond to 'predictors', and the boolean value indicates if the variables was actually used in the fitted model or not.

See Also

daniRules

Examples

```
getpredictorsfromcaptureoutput(party::ctree(Petal.Width~.,iris),names(iris))
```

```
getpredictorsfromtree getpredictorsfromtree
```

Description

for each terminal node of the tree, give the elements of "predictors" who appear in the node definition

Usage

```
getpredictorsfromtree(tree, predictors)
```

Arguments

tree same format than output of partykit::ctree

predictors list of variables

See Also

daniRules

```
tree<-partykit::ctree(Petal.Width~.,iris)
getpredictorsfromtree(tree,names(iris))</pre>
```

22 get_cellrn

get_cell

 $get\ cell\ without\ the\ row\ number$

Description

get cell without the row number

Usage

```
get_cell(x, iscellrn = FALSE, iscell = FALSE)
```

Arguments

Х

a vector of character strings

Details

```
if x is "aa.xoijj_a_1_f_1_" returns "a_1_f"
```

Value

a vector of character strings

Examples

```
get_cell("aa.x_1_2_3_4")#default
get_cell("1_2_3",TRUE)
get_cell("1_2_3",FALSE,TRUE)
unique(Tsampledata(TRUE)$variables))
unique(get_cell(Tsampledata(FALSE)$variables))
```

get_cellrn

Get cell and row number

Description

Get cell and row number

Usage

```
get_cellrn(x)
```

Arguments

Х

a vector of character strings

get_cellXXgroup 23

Details

```
if x is "aa.x_a_1_f_1" returns "a_1_f_1"
```

Value

a vector of character strings

Examples

```
get_cellrn("AA.char1_La_Ld_Lrn1")
data(TtableA);
unique(get_cellrn(names(TtableA)))
#Second example: no transposing variables
data(TtableB);data(XKB)
unique(get_cellrn(names(XKB)))
```

get_cellXXgroup

Get cell group

Description

Get cell group

Usage

```
get_cellXXgroup(x, marginpos, iscellXX = TRUE)
```

Arguments

x a vector of character strings

marginpos a vector of integer

Details

```
#' if x is "a_1_f_2_aa.xoijj",marginpos=2 returns "1"; if x is "a_1_f_2_aa.xoijj",marginpos=-2 returns "a_f_2"; if x is "a_1_f_2_aa.xoijj",marginpos=c(1:2) returns "a_1"
```

Value

a vector of character strings

Examples

```
get_cellXXgroup(c("aa.x_1_2_3_4","bb.x_1_2_3_4"),2,iscellXX=FALSE)
get_cellXXgroup(c("1_2_3_4","1_2_3_4"),2:3,iscellXX=TRUE)
variables<-Tsampledata(TRUE)$variables
unique(get_cellXXgroup(variables,2,iscellXX=FALSE))
unique(get_cellXXgroup(variables,-2,iscellXX=FALSE))
get_cellXXgroup(variables[50],2,iscellXX=FALSE)
get_cellXXgroup(variables[50],-2,iscellXX=FALSE)
#Second example: no transposing variables
TK<-Tsampledata(FALSE)
unique(get_cellXXgroup(TK$variable,1,iscell=FALSE))</pre>
```

get_cellXXmarginscount

Get the number of margins for a cell

Description

Get the number of margins for a cell

Usage

```
get_cellXXmarginscount(x, iscellXX = FALSE)
```

Arguments

x a vector of character stringsiscell a boolean indicating if x is a variable name or a cell name.

Details

```
if x is "aa.xoijj_a_1_f_1", cell=FALSE returns 4"; if x is "a_1_f_1", cell=TRUE returns 4"
```

Value

a vector of integers.

```
get_cellXXmarginscount("1_2_3_4",iscellXX=TRUE)
get_cellXXmarginscount("aa.x_1_2_3_4",iscellXX=FALSE)
data(TtableA)
unique(get_cellXXmarginscount(names(TtableA),iscellXX=FALSE))
#Second example: no transposing variables
TK<-Tsampledata(FALSE)
unique(get_cellXXmarginscount(TK$variables))</pre>
```

get_cellXXsplit 25

get_cellXXsplit

split a cell

Description

split a cell

Usage

```
get_cellXXsplit(x, marginpos = NULL, iscellXX = FALSE)
```

Arguments

x a vector of character strings

iscell x a boolean indicating if x is a cell

Details

```
if x is "aa.xoijj_a_1_f_1" returns c("a","1","f","1")
```

Value

a vector of character strings

```
get_cellXXsplit("aa.x_1_2_3_4",iscellXX=FALSE)
get_cellXXsplit("1_2_3_4",iscellXX=TRUE)
get_cellXXsplit("1_2_3_4",2:3,iscellXX=TRUE)
get_cellXXsplit("1_2_3_4",-(2:3),iscellXX=TRUE)
variables<-Tsampledata(TRUE)$variables
unique(get_cellXXsplit(variables,iscell=FALSE))
get_cellXXsplit(variables[50],iscell=FALSE)
get_cellXXsplit(variables[50],-(2:3),iscell=FALSE)
unique(get_cellXXsplit(variables,2,iscell=FALSE))
#Second example: no transposing variables
TK<-Tsampledata(FALSE)
unique(get_cellXXsplit(TK$variables,iscell=FALSE))</pre>
```

get_natural.predictors

 ${\tt get_missingind}$

Get missing indicator for a cell or variable

Description

Get missing indicator for a cell or variable

Usage

```
get_missingind(x, variables)
```

Arguments

Х

a vector of character strings

Details

```
if x is "a_1_f_1_aa.xoijj" returns c("a","1","f","1")
```

Value

a vector of character strings

Examples

```
variables<-Tsampledata(TRUE)$variables
unlist(unique(get_missingind(variables,variables)))
variables<-Tsampledata(FALSE)$variables
unlist(unique(get_missingind(variables,variables)))</pre>
```

```
{\tt get\_natural.predictors}
```

Get natural predictors

Description

Get natural predictors

Usage

```
get_natural.predictors(x, variables = x, predictors = NULL)
```

Arguments

```
x a vector of character strings
variables a vector of character strings
```

get_origin 27

Details

```
if x is "a_1_f_1_aa.xoijj" returns c("a","1","f","1")
```

Value

a vector of character strings

Examples

```
TK<-TtableA
get_natural.predictors(x=sample(names(TtableA),5),variables=names(TtableA))</pre>
```

get_origin

Get origin table

Description

Get origin table

Usage

```
get_origin(x)
```

Arguments

Х

a vector of character strings

Details

```
if x is "aa.xoijj_a_1_f_1_" returns c("aa")
```

Value

a vector of character strings

```
get_origin("tableA.cont1_1_Lrn1")
variables<-Tsampledata(TRUE)$variables
unlist(unique(get_origin(variables,variables)))
variables<-Tsampledata(FALSE)$variables
unlist(unique(get_origin(variables,variables)))</pre>
```

28 get_presentind

get_parent

get parent if any

Description

```
get parent if any
```

Usage

```
get_parent(variables, variable_ref)
```

Arguments

```
variables a character strings
variable_ref a vector of character strings
```

Details

```
if x is "aa.xoijj_a_1_f_1_" returns "a_1_f"
```

Value

a vector of character strings

Examples

```
get_parent("aa.x_1_2_3_4","aa.x_1_2_3")#default
```

 ${\tt get_presentind}$

get the present indicator for a cell

Description

get the present indicator for a cell

Usage

```
get_presentind(
  variables,
  refvariables = variables,
  rns = unlist(unique(get_cellrn(refvariables)))
)
```

Arguments

Х

a vector of character strings

get_var 29

Details

```
if x is "a_1_f_1_aa.xoijj" returns c("a","1","f","1")
```

Value

a vector of character strings

Examples

```
get_presentind("AA.x_1_2_3_4","AA.present_1_2_3_4")
get_presentind("AA.present_1_2_3_4",c("AA.present_1_2_3_3","AA.present_1_2_3"))
get_presentind("AA.present_1_2_3_4",c("AA.present_1_2_3_3","AA.present_1_2_3_4"))
variables<-Tsampledata(TRUE)$variables
variable<-"AA.present_La_La_Lrn1"
get_presentind(variable,variables)
unlist(unique(get_presentind(variables)))
variables<-Tsampledata(FALSE)$variables
unlist(unique(get_presentind(variables,variables)))</pre>
```

get_var

Get variable name

Description

Get variable name

Usage

```
get_var(x)
```

Arguments

Х

a vector of character strings

Details

```
if x is "aa.xoijj_a_1_f_1" returns "aa.xoijj"
```

Value

a vector of character strings

```
get_var("aa.x_1_2_3_4")
data(TtableA)
unique(get_var(names(TtableA)))
#Second example: no transposing variables
TK<-Tsampledata(FALSE)
unique(get_var(TK$variables))</pre>
```

```
get_XXpredecessoratmargin

get cell predecessors at margin
```

Description

get cell predecessors at margin

Usage

```
get_XXpredecessoratmargin(
   XXs,
   refXXs = XXs,
   marginpos = NULL,
   iscellXX = FALSE
)
```

Arguments

XXs a vector of character strings

refXXs a vector of character strings containing the potential predecessors

marginpos a vector of integers

Details

```
if XXs is "aa.xoijj_a_1_f_1" and refXXs contains "aa.xoijj_a_1_e_1" and marginpos=3 returns "aa.xoijj_a_1_e_1" if XXs is "aa.xoijj_a_1_f_2" and refXXs contains "aa.xoijj_a_1_f_1" and marginpos=NULL returns "aa.xoijj_a_1_f_1" if XXs is "id1" and iscellXX=FALSE whatever refXXs returns character(0) if XXs is "" and iscellXX=FALSE whatever refXXs returns character(0) if XXs is "b_1_f_1" and iscellXX=TRUE and refXXs contains "a_1_f_1" returns "a_1_f_1"
```

Value

a vector of character strings

```
get_XXpredecessoratmargin(XXs="aa.x_1_2_3_4", refXXs=c("bb.x_1_2_2_4", "aa.x_1_2_2_4", "aa.x_1_1_3_4"),2,iscellXX
get_XXpredecessoratmargin(XXs=c("1_2_2_4", "1_2_2_4", "1_1_3_4", "1_1_3_3"),iscellXX=TRUE)
get_XXpredecessoratmargin(XXs="1_1_3_4", refXXs=c("1_2_2_4", "1_2_2_4", "1_1_3_4", "1_1_3_3"),iscellXX=TRUE)
data(XKA)
cells<-unique(get_cellrn(XKA$variables))
get_XXpredecessoratmargin(cells, marginpos=1, iscellXX=TRUE)
get_XXpredecessoratmargin(cells[10], cells, 1, iscellXX=TRUE)</pre>
```

Description

get cell predecessors at margin

Usage

```
get_XXpredecessorsatmargin(
   XXs,
   marginpos,
   refXXs = XXs[order(get_cellXXgroup(XXs, marginpos))],
   iscellXX = FALSE,
   cellXXgroup = get_cellXXgroup(refXXs, marginpos2, iscellXX),
   CompcellXXgroup = get_cellXXgroup(refXXs, -marginpos2, iscellXX))
```

Arguments

XXs a vector of character strings

marginpos a vector of integers

refXXs a vector of character strings containing the potential predecessors

Details

```
if XXs is "aa.xoijj_a_1_f_1" and refXXs contains "aa.xoijj_a_1_e_1" and marginpos=3 returns "aa.xoijj_a_1_e_1" if XXs is "aa.xoijj_a_1_f_2" and refXXs contains "aa.xoijj_a_1_f_1" and marginpos=NULL returns "aa.xoijj_a_1_f_1" if XXs is "id1" and iscellXX=FALSE whatever refXXs returns character(0) if XXs is "" and iscellXX=FALSE whatever refXXs returns character(0) if XXs is "b_1_f_1" and iscellXX=TRUE and refXXs contains "a_1_f_1" returns "a_1_f_1"
```

Value

a vector of character strings

```
get_XXpredecessoratmargin(
    XXs="aa.x_1_2_3_4",
    refXXs=c("bb.x_1_2_2_4","aa.x_1_2_2_4","aa.x_1_1_3_4"),
    2,iscellXX=FALSE)
get_XXpredecessoratmargin(
```

32 good.fit.parameters

```
XXs=c("1_2_2_4","1_2_2_4","1_1_3_4","1_1_3_3"),
  iscellXX=TRUE)
get_XXpredecessoratmargin(
  XXs="1_1_3_4",
  refXXs=c("1_2_2_4","1_2_2_4","1_1_3_4","1_1_3_3"),
  iscellXX=TRUE)
data(XKA)
cells<-unique(get_cellrn(XKA$variables))
get_XXpredecessoratmargin(cells,marginpos=1,iscellXX=TRUE)
get_XXpredecessoratmargin(cells[10],cells,1,iscellXX=TRUE)</pre>
```

good.fit.parameters Select only arguments for a specific fitting function

Description

Select only arguments for a specific fitting function

Usage

```
good.fit.parameters(method, synparameters)
```

Arguments

```
method a string. currently only method="ctree". synparameters a named list.
```

Details

Currently only works with method="ctree" Only selects the arguments that match the function partykit::ctree_control

Value

a sublist of synparameters, which names are possible arguments of partykit::ctree_control if method="ctree".

```
good.fit.parameters(method="ctree",list(teststat=30,tutu=3))
```

good.sample.parameters

```
good.sample.parameters
```

Select only arguments for sampling.

Description

Select only arguments for sampling.

Usage

```
good.sample.parameters(method, synparameters)
```

Arguments

synparameters

```
method
                   a string. currently only method="ctree".
                  a named list.
```

Details

In prevision of future developments. returns NULL for the moment

Examples

```
good.sample.parameters(method="ctree",list(teststat=30,tutu=3))
```

```
Select only arguments for a specific fitting function This function is not
good.syn.parameters
                           used anymore
```

Description

Select only arguments for a specific fitting function This function is not used anymore

Usage

```
good.syn.parameters(method, synparameters)
```

Arguments

```
method
                  a string. currently only method="ctree".
synparameters
                  a named list.
```

Details

Currently only works with method="ctree" Only selects the arguments that match the function synthpop::syn.ctree

34 NAto0

Value

a sublist of synparameters, which names are possible arguments of synthpop::syn.ctree if method="ctree".

Examples

```
good.syn.parameters(\texttt{method="ctree",list(y=c(1:2),smoothing=TRUE,tutu="not in synthpop::syn.ctree arguments"))}
```

NAto0

Recoding of NAs to 0 or "NA"

Description

Recoding of NAs to 0 or "NA"

Usage

NAto0(tableA)

Arguments

tableA

a dataframe

Details

for synthetisation to run, missing values are treated as a special factor level for factor variables, or as 0 for continuous variables. To avoid issues, for continuous variables, a missing indicator is also created.

Value

a dataframe

```
toto<-cars
toto$speed[sample(nrow(cars),3)]<-NA
NAto0(toto)</pre>
```

onlygoodargs 35

onlygoodargs	Generic function: remove all the elements of a named list which names
	are not arguments of a specific function.

Description

Generic function: remove all the elements of a named list which names are not arguments of a specific function.

Usage

```
onlygoodargs(fun, L)
```

Arguments

```
fun a function L a list
```

Details

remove all the elements of a list which names are not arguments of a specific function.

Value

a list.

Examples

```
onlygoodargs(lm,list(data=cars,formula=speed~dist,tutu="not an arg from lm"))
```

posixcttonumeric

Converts all posixct variables of a dataframe into a numeric variable

Description

Converts all posixct variables of a dataframe into a numeric variable

Usage

```
posixcttonumeric(tableA)
```

Arguments

tableA

a dataframe

Value

a list.

Examples

```
toto<-cars
toto$now<-Sys.time()
posixcttonumeric(toto)</pre>
```

```
predictor.matrix.default
```

Define a default predictor matrix

Description

Define a default predictor matrix

Usage

```
predictor.matrix.default(variables)
```

Arguments

variables a vector of character strings

Details

Returns the lower diagonal matrix with ones.

Value

a matrix

```
variables<-Tsampledata(TRUE)$variables
predictor.matrix.default(TK$variables)</pre>
```

predictor.matrix.rate 37

```
predictor.matrix.rate predictor.matrix.rate
```

Description

predictor.matrix.rate

Usage

```
predictor.matrix.rate(
  variables,
  nopredictor = character(0),
  allpredictor = character(0),
 marginposs = integer(0)
)
```

Arguments

a vector of character strings Χ

Details

```
if x is "aa.xoijj_a_1_f_1_" returns c("a","1","f","1")
```

Value

a vector of character strings

preparepredictorsforctreefit

Prepare predictors for ctree fit

Description

Prepare predictors for ctree fit

Usage

```
preparepredictorsforctreefit(x, keep = NULL)
```

Arguments

a predictors, a dataframe. method a string. currently only method="ctree".

variable to predict, a vector У

synthetic parameters to pass to the right fit model function. the fit model function . . .

name is the concatenation of "fit.model" and method

38 reduceT_f

Value

a sublist of synparameters, which names are possible arguments of synthpop::syn.ctree if method="ctree".

Examples

```
\label{eq:prepare} prepare predictors for ctree fit (x, \\ keep=NULL)
```

reduceT_f

Reverse augmentT_f: Function that will convert cell and marginal ratios and overall total to cell values

Description

Reverse augmentT_f: Function that will convert cell and marginal ratios and overall total to cell values

Usage

```
reduceT_f(.data, variables, verbose = FALSE, hack = TRUE, recalibrateonly = F)
```

Arguments

data frame to "reduce" variables list of variable names roots

verbose (default FALSE) if verbose, the formulae to compute the new variables is printed.

hack (default TRUE)

Details

this functions looks for the Augmentation parameters in the package object Augmentparameters[[tablename]]\$percent For each variable listed in Augmentparameters[[tablename]]\$percent, it looks for the corresponding variable in .data and computes cell values from cell and marginal ratios and overall total

```
library(BigSyn)
library(reshape2)
library(data.table)
hack=TRUE
verbose=TRUE
data(TtableA,package="BigSyn")
variablemax="AA.present"
variablesmax=variablemax
variablepct="AA.cont1"
variablespct=variablepct
variables=variablespct
ATtableA<-augmentT_f(TtableA,</pre>
```

reduceT_f

```
variablesmax=variablesmax,
                     variablespct=variablespct)
.data=ATtableA
RATtableA<-reduceT_f(.data = ATtableA, variables=variablespct)</pre>
all(sapply(1:nrow(TtableA),function(i){
jj<-NAto0(TtableA)[i,]!=NAto0(RATtableA)[i,names(TtableA)]</pre>
identical(signif(NAto0(TtableA)[i,jj],3),
          signif(NAto0(RATtableA)[i,names(TtableA)[jj]],3))}))
randomcheck<-function(i=NULL){if(is.null(i)){</pre>
         i<-sample(1:nrow(TtableA),1)};</pre>
variablex="AA.cont1_La_La";
vx=c("AA.cont1_La_La_Lrn1"
     "AA.cont1_La_La_Lrn2"
     "AA.cont1_La_La_Lrn3",
     "AA.cont1_La_La_Lrn4");
BigSyn::get_presentind(variables = vx,refvariables = names(TtableA))->px
BigSyn::get_missingind(x=vx,variables = names(TtableA))->mx
list(i=i,
     total=ATtableA[i,"AA.cont1_"],
     LaRatio=ATtableA[i,"AA.cont1_La"],
     LaLaRatio=ATtableA[i, "AA.cont1_La_La"],
     LaLaTotal=ATtableA[i,"AA.cont1_"]*
     ATtableA[i,"AA.cont1_La"]*
     ATtableA[i,"AA.cont1_La_La"],
rbind(rat=RATtableA[i,vx],at=ATtableA[i,vx],t=TtableA[i,vx]),
rbind(ratp=RATtableA[i,px],atp=ATtableA[i,px],tp=TtableA[i,px]),
rbind(ratp=RATtableA[i,mx],atp=ATtableA[i,mx],tp=TtableA[i,mx]))}
randomcheck(19)
randomcheck(109)
randomcheck(57)
nrep=1
SATtableA<-SDPSYN2(ATtableA,asis=c("id1a","id1b"),
                  fitmodelsavepath = NULL,treeplotsavefolder = NULL)[[1]]
CSATtableA<-resampleT_f(SATtableA, "AA.cont1")
RSATtableA<-reduceT_f(.data = SATtableA,variables="AA.cont1",verbose=TRUE)
RCSATtableA<-reduceT_f(.data = CSATtableA, variables="AA.cont1", verbose=TRUE)</pre>
toto=function(.datareduced,.data){
w<-merge(.datareduced[c("id1a","id1b","AA.cont1_")],</pre>
          .data[c("id1a","id1b","AA.cont1_")],by=c("id1a","id1b"))
plot(w$AA.cont1_.x,w$AA.cont1_.y)}
toto(.datareduced,.data)
toto2=function(.datareduced){
 .datareduced$AA.cont1_check<-
     rowSums(.datareduced[grep("Lrn",grep("AA.cont1_",names(.datareduced),value=T),value=T)],
                na.rm=T)
with(.datareduced,plot(AA.cont1_,AA.cont1_check))
}
toto2(RATtableA)
toto2(RSATtableA)
toto2(RCSATtableA)
```

40 resampleT_f

resampleT_f	resample when synthetisation created incoherent high and low level
	aggregates

Description

resample when synthetisation created incoherent high and low level aggregates

Usage

```
resampleT_f(.data, variables, verbose = FALSE)
```

Arguments

.data data frame to "reduce"
variables list of variable names roots

verbose (default FALSE) if verbose, the formulae to compute the new variables is printed.

Details

In the case where marginal presence indicator equals 1 but all cell presence indicators where synthetised to 0, then presence indicators and other variables are resampled from synthetic units with coherent values.

(CSATtableA\$AA.cont1_Lb_La_Lrn3==0|is.na(CSATtableA\$AA.cont1_Lb_La_Lrn3)))

```
library(BigSyn)
library(reshape2)
library(data.table)
data(TtableA,package="BigSyn")
variablepct="AA.cont1"
variablespct=variablepct
variablemax="AA.present"
variablesmax=variablemax
ATtableA<-augmentT_f(TtableA, variablesmax=variablesmax, variablespct=variablespct)
set.seed(1)
SATtableA<-BigSyn::SDPSYN2(ATtableA,asis=c("id1a", "id1b"))[[1]]
problems<-SATtableA$AA.cont1_Lb_La>0&!is.na(SATtableA$AA.cont1_Lb_La)&((SATtableA$AA.cont1_Lb_La_Lrn1==0|is.na
varcell=c("AA.cont1_Lb_La_Lrn1", "AA.cont1_Lb_La_Lrn2", "AA.cont1_Lb_La_Lrn3")
varcellandpresenceind<-unlist(c(varcell,get_missingind(c(varcell,"AA.cont1_Lb_La"),names(SATtableA)),get_prese
replacements < -SATtableA\$AA.cont1\_Lb\_La > 0\&! is.na(SATtableA\$AA.cont1\_Lb\_La)\&!((SATtableA\$AA.cont1\_Lb\_La\_Lrn1 == 0) = 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.000 + 0.00
SATtableA[problems,c("AA.cont1_Lb_La",varcellandpresenceind)][1:3,]
SATtableA[replacements,c("AA.cont1_Lb_La",varcellandpresenceind)][1:3,]
CSATtableA<-resampleT_f(SATtableA, variablespct)
CSATtableA[problems,c("AA.cont1_Lb_La",varcellandpresenceind)][1:3,]
problems2<-CSATtableA$AA.cont1_Lb_La>0&!is.na(CSATtableA$AA.cont1_Lb_La)&
((CSATtableA$AA.cont1_Lb_La_Lrn1==0|is.na(CSATtableA$AA.cont1_Lb_La_Lrn1))&
  (CSATtableA$AA.cont1_Lb_La_Lrn2==0|is.na(CSATtableA$AA.cont1_Lb_La_Lrn2))&
```

runCompare 41

```
any(problems2); sum(problems2)
RCSATtableA<-reduceT_f(CSATtableA, variablespct)</pre>
RCSAT table A [problems, intersect (c("AA.cont1\_Lb\_La", varcelland presence ind), names (RCSAT table A))][1:3,] \\
problems3<-RCSATtableA$AA.cont1_Lb_La>0&!is.na(RCSATtableA$AA.cont1_Lb_La)&
((RCSATtableA$AA.cont1_Lb_La_Lrn1==0|is.na(RCSATtableA$AA.cont1_Lb_La_Lrn1))&
 (RCSATtableA$AA.cont1_Lb_La_Lrn2==0|is.na(RCSATtableA$AA.cont1_Lb_La_Lrn2))&
 (RCSATtableA$AA.cont1_Lb_La_Lrn3==0|is.na(RCSATtableA$AA.cont1_Lb_La_Lrn3)))
any(problems3); sum(problems3)
AA < -rbind(RCSATtableA[problems3, intersect(c("AA.cont1\_Lb\_La", varcellandpresenceind), names(RCSATtableA))], \\
CSATtableA[problems3,intersect(c("AA.cont1_Lb_La",varcellandpresenceind),names(RCSATtableA))],
SATtableA[problems3,intersect(c("AA.cont1_Lb_La",varcellandpresenceind),names(RCSATtableA))])
AA$y=rep(c("RCSA","CSA","SA"),each=sum(problems3))
AA$x=rep(1:sum(problems3),3)
AA[order(AA$x),]
library(ggplot2);library(dplyr)
xx<-function(x){xxx<-x[sort(grep("present",names(x),value=TRUE))]</pre>
xxx[xxx==0]<-NA
StudyDataTools::ggplot_missing(xxx)}
xx(ATtableA)
xx(SATtableA)
xx(CSATtableA)
xx(RCSATtableA)
```

 ${\tt runCompare}$

runCompare

Description

Shiny App to visualize regression trees and compare synthetic vs non synthetic data

Usage

Arguments

```
data1 a dataframe data2 a dataframe
```

42 sample.ctree

```
listofpackage1 a vector of character strings
listofpackage2 a vector of character strings
package1 a character string
package2 a character string
Sparameters
```

Examples

```
package1<-NULL
package2<-NULL
runCompare()</pre>
```

sample.ctree

Function to sample from a ctree fitted model

Description

Function to sample from a ctree fitted model

Usage

```
sample.ctree(xp, fit.model, smoothing = "none", ...)
```

Arguments

```
y a vector of values to pull from terminalnodes a vector of terminal nodes newterminalnodes:

a path to save the graph
```

Value

a vector of the same size than terminal nodes, obtained by sampling between the values of y such for the same terminal node.

```
y < -iris \\ Species; x < -xp < -iris[, -5]; fit.model < -fitmodel.ctree(x,y); sample.ctree(x,fit.model) \\ \\ + (x,fit.model) \\ + (x,fit.m
```

sample.fn 43

sample.fn

Sample a model with a specific function

Description

Sample a model with a specific function

Usage

```
sample.fn(method, xp, fit.model, smoothing, ...)
```

Arguments

method a string. currently only method="ctree".

... synthetic parameters to pass to the right fit model function. the fit model function

name is the concatenation of "fit.model" and method

x a predictors, a dataframe.y variable to predict, a vector

Value

a sublist of synparameters, which names are possible arguments of synthpop::syn.ctree if method="ctree".

Examples

sampledata

Sample data for transposition

Description

Sample data for transposition

Usage

```
sampledata(transposingvariables = TRUE)
```

Arguments

```
transposingvariables
```

a boolean. If TRUE, transposing id variables are created.

samplefrompool

Value

a data frame with id variables, numeric, factor and character variables.

samplefrompool

Function to sample from a set of partitioning rules

Description

Function to sample from a set of partitioning rules

Usage

```
samplefrompool(y, terminalnodes, newterminalnodes)
```

Arguments

```
y a vector of values to pull from terminalnodes a vector of terminal nodes newterminalnodes:

a path to save the graph
```

Value

a vector of the same size than terminalnodes, obtained by sampling betweenn the values of y such for the same terminal node.

```
y=iris$Species;x=iris[,-5];fit.mod<-fitmodel.ctree(x,y);terminalnodes<-getnodesfromrules(x,fit.mod$Rules);
newterminalnodes<-sample(unique(terminalnodes),10,replace=TRUE);
samplefrompool(y,terminalnodes,newterminalnodes)
y<-y[terminalnodes!=7]
terminalnodes<-terminalnodes[terminalnodes!=7]
samplefrompool(y,terminalnodes,newterminalnodes)</pre>
```

SDPSYN2 45

SDPSYN2

General SDP function.

Description

General SDP function.

Usage

```
SDPSYN2(
       TtableA,
       asis = NULL,
      notpredictor = asis,
      nrep = 1,
       synparameters = NULL,
     Sparameters = Sparameters.default.f(ref.table = TtableA, asis = asis, notpredictor =
              notpredictor, preferredmethod = "ctree", defaultsynparameters =
              c(as.list(synparameters),
         eval (formals (Sparameters. default.f) \$ default symparameters) [setdiff (names (formals (Sparameters. default.f) \$ default symparameters] [setdiff (names (formals (Sparameters. default.f) \] [setdiff (na
               c("", names(synparameters)))])),
     STtableA = if (is.null(asis)) {
                                                                                                                               data.frame(.n = rep(nrep, each = nrow(TtableA))) }
         else {
                                          plyr::ddply(data.frame(.n = nrep), ~.n, function(d) {
                                                                                                                                                                                                                                                            TtableA[asis]
                             }) },
       fitmodelsavepath = NULL,
       treeplotsavefolder = NULL,
       samplereportsavepath = NULL,
       stepbystepsavepath = NULL,
       doparallel = TRUE,
       recode = NULL,
       saveeach = 200,
       randomfitorder = TRUE,
       fitonly = FALSE
)
```

Arguments

```
TtableA
                  a dataframe to synthesize
                  list of variable names from TtableA to keep as is (e.g. not to synthesize)
asis
notpredictor
                  list of variable names which should not be used as predictors
                  number of synthetic replicates wanted
nrep
                  general synthetisation paramters
synparameters
Sparameters
                  a list, Specific (variable by variable) synthetisation parameters, splits ...
STtableA
                  a dataframe
fitmodelsavepath
                  a path where to save the fitted models
```

46 SDPSYN2

```
treeplotsavefolder
                  a path where to save the tree plots
samplereportsavepath
                  a path where to save the sampling report
stepbystepsavepath
                  a path where to backup the synthetised in case of a crash
doparallel
                  a boolean indicating whether sampling should be done in parallel for each repli-
                  acte
                  : a vector of character strings or NULL, list of variables to be recoded
recode
                  an integer, indicating every how many variables a backup is done
saveeach
randomfitorder a boolean: fitting for each variable can be done in the order of appearance of
                  each variables or at random
fitonly
                  a boolean, if TRUE, no sampling is done.
```

Details

This function is doing both the fitting and the sampling.

```
data(TtableA,package="BigSyn")
ATtableA=augmentT_f(TtableA, variablesmax="AA.present",
                   variablespct="AA.cont1")
asis=NULL; notpredictor=asis; nrep=1; synparameters=NULL;
Sparameters=
Sparameters.default.f(ref.table=TtableA.
                       asis=asis,
                       notpredictor=notpredictor,
                       preferredmethod="ctree",
                       defaultsynparameters=
                          c(as.list(synparameters),
                             eval(formals(Sparameters.default.f)$defaultsynparameters)[
                     setdiff(names(formals(Sparameters.default.f)$defaultsynparameters),
                              c("",names(synparameters)))]));
SATtableA=plyr::rdply(nrep,ATtableA[asis]);
samplereportsavepath=NULL;
stepbystepsavepath=NULL;
doparallel=FALSE;
recode=NULL;
randomfitorder=TRUE;
fitonly=FALSE;
fitmodelsavepath=tempdir()
treeplotsavefolder=tempdir()
sapply(list.files(tempdir(),full.names = TRUE ),file.remove)
SATtableA<-SDPSYN2(ATtableA, asis=NULL,
                  fitmodelsavepath = fitmodelsavepath,
                  treeplotsavefolder=treeplotsavefolder)
todisplay<-grep("La_La_Lrn1", names(STtableA[[1]]), value=T);</pre>
STtableA[[1]][1:3,todisplay];TtableA[1:3,todisplay]
```

sorttablewithingroup 47

```
# Controling that AA.present_La=0=>AA.present_La_Lb=0 in synthetic data
library(BigSyn)
library(reshape2)
library(data.table)
data(TtableA,package="BigSyn")
variablepct="AA.cont1"
variablespct=variablepct
variablemax="AA.present"
variablesmax=variablemax
set.seed(1)
asis=c("id1a", "id1b")
                 fitmodelsavepath=NULL
                 treeplotsavefolder=NULL
                 samplereportsavepath=NULL
                 stepbystepsavepath=NULL
                 doparallel=TRUE
                 recode=NULL
                 saveeach=200
                 randomfitorder=TRUE
                 fitonly=FALSE
variablemax="AA.present"
variablesmax=variablemax
variablepct="AA.cont1"
variablespct=variablepct
ATtableA<-augmentT_f(TtableA,
                   variablesmax=variablesmax,variablespct=variablespct)
TtableA<-ATtableA
STtableA<-ATtableA[asis]</pre>
Sparameters=Sparameters.default.f(
  ref.table=ATtableA,asis=c("id1a", "id1b"),
  notpredictor=NULL,
  preferredmethod="ctree",
  defaultsynparameters=
    eval(formals(Sparameters.default.f)$defaultsynparameters))
SATtableA<-BigSyn::SDPSYN2(ATtableA,asis=c("id1a", "id1b"))[[1]]
problems<-SATtableA$AA.present_Lb_La==1&SATtableA$AA.present_Lb==0</pre>
mean(problems)
Sparameters[["AA.present_Lb_La"]]
library(dplyr)
library(ggplot2)
xx<-function(x){</pre>
xxx<-x[sort(grep("present",names(x),value=TRUE))]</pre>
xxx[xxx==0]<-NA
StudyDataTools::ggplot_missing(xxx)}
xx(ATtableA)
xx(SATtableA)
```

48 sorttablewithingroup

Description

sort table within group without changing the group position

Usage

```
sorttablewithingroup(.data, groupvar, sortvar, decreasing = FALSE)
```

Arguments

```
    .data a dataframe
    groupvar a vector of character strings that are names of variables from .data
    sortvar a vector of character strings that are names of variables from .data
    decreasing a boolean if TRUE, decreasing order is used.
```

Details

Groups are defined by unique values of .data[groupvar]. Within each group, data is sorted according to sortvar.

Value

a list.

```
set.seed(1)
N=10
.data=data.frame(
   .group=sample(letters[1:2],N,replace=TRUE),
  y=runif(N),
  origorder=1:N)
  groupvar=".group"
  sortvar="v"
.data2=plyr::ddply(.data,".group",
                                                           function(d){d$intraorder=order(d[[sortvar]]);
d$neworder=d$origorder[order(order(d[[sortvar]]))];d})
.data4=.data2[.data2[[groupvar]]=="a",];.data4[.data4$intraorder,]
cbind(.data,"|",.data2)
cbind(.data,"|",.data2[order(.data2$origorder),])
cbind(.data[order(order(.data2$origorder)),],"|",.data2)
.data3=cbind(.data,"|",
                                           .data2[order(.data2$neworder),]);
.data3[.data[[groupvar]]=="a",];.data3
.data3=cbind(.data,"|",
                                           .data[order(order(.data2$origorder)),][order(.data2$neworder),]);
.data3[.data[[groupvar]]=="a",]
.data3=cbind(.data,"|",
                                           .data[order(order(.data2$origorder))[order(.data2$neworder)],]);
.data3[.data[[groupvar]]=="a",]
cbind(.data, I="|", sorttablewithingroup(.data, groupvar, sortvar), I="|", sortvar), I="|
```

Sparameters.default.f 49

```
sorttablewithingroup(.data,groupvar,sortvar,decreasing=TRUE),
I="|",sorttablewithingroup(.data,NULL,groupvar,decreasing=TRUE))
```

Sparameters.default.f Default synthetisation parameters based on variable names

Description

Default synthetisation parameters based on variable names

Usage

```
Sparameters.default.f(
    ref.table,
    asis = NULL,
    notpredictor = NULL,
    variables = Sparameters.variables.reorder.default(names(ref.table)),
    predictors.matrix = predictor.matrix.default(variables)[!is.element(variables, asis),
       !is.element(variables, notpredictor)],
    splittingvar = NULL,
    moresplits = NULL,
    preferredmethod = "ctree",
    splithreshold = 100,
    defaultsynparameters = list(smoothing = "none", importance = TRUE, keep.forest = TRUE,
       minbucket = 30)
)
```

Arguments

ref.table a dataframe asis a vector of character strings, indicating which variables to keep as is. notpredictor a vector of character strings, indicating which variables are not supposed to be used as predictors. variables a vector of character strings, indicating the variables to synthesize. Order is important. predictors.matrix a predictor matrix. Number of rows is the number of variables to synthesize, number of columns is all the variables from ref.table moresplits an object of class more splist (not defined yet) preferredmethod: "rf" for random forest or "ctree" for classification tree defaultparameters a list indicating default parameters for synthesisation functions, for example ntree=5, smoothing="none"

Details

creates default synthetisation parameters Some rules: parents variable are potential predictors of their children, synthetisation is conditional to missingindicators, synthetisation is conditional to presence in cell

Examples

```
data(TtableA)
ATtableA<-augmentT_f(TtableA,variablespct="AA.cont1",variablesmax="AA.present")
ref.table<-ATtableA
Spa<-Sparameters.default.f(ref.table=ATtableA)
names(Spa)<-lapply(Spa,function(x){x$variable})
Spa$AA.present_La_Lb
Spa$AA.cont1_La_Lb</pre>
```

```
Sparameters.variables.reorder.default
```

General Default ordering of variables for synthetisation based on name of the variable.

Description

General Default ordering of variables for synthetisation based on name of the variable.

Usage

```
Sparameters.variables.reorder.default(
  variables,
  orderwithinorigin = NULL,
  id = NULL,
  extrasort = NULL
)
```

Arguments

treedepth 51

Details

After transposition, variable names follow this format: origin.variablename_margin1_margin2....lastmargin Some rules have to be followed:

- Missing indicators have to be synthesised before the corresponding variables, for example AA.factor1missingind_L1_L2_L1 needs to be synthesised before AA.factor1missingind_L1_L2_L1
- Cell indicators must be synthesised before the corresponding variables. For example AA.present_L1_L2_L1 must be synthesised before AA.factor1_L1_L2_L1 and before AA.cont1_L1_L2_L1
- Parent variables (aggregated) must be synthesised before their children: For example AA.present_L1 must be synthesised before AA.present_L1_L2, AA.cont2_L1_L2 must be synthesised before AA.cont2_L1_L2_L3 AA.present_L1 must be synthesised before AA.present_L1_L3 AA.cont2missingind_L1 must be synthesised before AA.cont2missingind_L1_L3
- if for examples variable AA.cont1 in each cell has to be synthesised before AA.cont2, this can be specified with the orderwithinorigin argument
- for the use of the argument extrasort, refer to sorttable within group

Value

a list.

Examples

```
TK<-Tsampledata(TRUE)$TtableA
Sparameters.variables.reorder.default(names(TK$TtableA))
#Second example: no transposing variables
TtableA<-Tsampledata(TRUE)$TtableA
orderwithinorigin=c("AA.factor1","AA.factor2")
variables<-names(TtableA)
Sparameters.variables.reorder.default(variables,orderwithinorigin)
```

treedepth

Compute depth of a "party" tree

Description

this function takes the output of the partykit::ctree function and prints the tree into a pdf file, located in the specified folder. It computes the depth and width and tries to create a pdf with the right dimensions.

Usage

treedepth(x)

Arguments

Х

a tree

Tsampledata Tsampledata

Details

recursive function

Examples

```
y=iris$Species;x=iris[,-5]
partyctree <- party::ctree(y ~ ., data=cbind(y=y,x))
treedepth(partyctree@tree)
partyctree <- party::ctree(y ~ ., data=cbind(y=y,x))
treedepth(partykit::ctree(y ~ ., data=cbind(y=y,x)))</pre>
```

treetopdf

Ctree to pdf graph

Description

this function takes the output of the partykit::ctree function and prints the tree into a pdf file, located in the specified folder. It computes the depth and width and tries to create a pdf with the right dimensions.

Usage

```
treetopdf(partykitctree, savepath)
```

Arguments

partykitctree: an output of partykit::ctree

savepath: a file path where to store the pdf of the plot

Examples

```
y=iris$Species;x=iris[,-5]
partykitctree <- partykit::ctree(y ~ ., data=cbind(y=y,x))
treetopdf(partykitctree,"./x.pdf")</pre>
```

Tsampledata

Transposed sample data.

Description

Transposed sample data.

Usage

```
Tsampledata(transposingvariables = TRUE)
```

TTsampledata 53

Arguments

```
transposingvariables
```

a boolean. If TRUE, stransposing id variables are created.

Details

Tsampledata(x) is Generaltransposefunction(Tsampledata(x))

Value

a data frame with id variables, numeric, factor and character variables.

TTsampledata

Transposed sample data.

Description

Transposed sample data.

Usage

TTsampledata(transposingvariables = TRUE)

Arguments

transposingvariables

a boolean. If TRUE, stransposing id variables are created.

Details

Tsampledata(x) is Generaltransposefunction(Tsampledata(x))

Value

a data frame with id variables, numeric, factor and character variables.

54 %notin%

%notin%

Not in operator

Description

```
negation de '%in%'
```

Usage

```
... %notin% NA
```

Details

This function is used in the function 'daniRules'.

See Also

daniRules

Examples

1%notin%2:3 1%notin%1:3

Index

%notin%, 54	<pre>good.fit.parameters, 32 good.sample.parameters, 33</pre>
<pre>augmentmaxT_f, 3</pre>	good.syn.parameters, 33
augmentpctT_f, 6	good. Syn. par ameter 5, 33
augmentT_f, 9	NAto0, 34
BigSyn, 10	onlygoodargs, 35
compilefits, 10	posixcttonumeric, 35
compilesamplereports, 11	predictor.matrix.default, 36
1 10 1 10	predictor.matrix.rate, 37
daniRules, 12	preparepredictors for ctree fit, 37
donors.receptors.check, 13	
drop_last, 13	reduceT_f, 38
fitmodel.ctree, 14	resampleT_f, 40
fitmodel.fn, 15	runCompare, 41
fitmodel.rf, 15	comple atmos 42
fitthemodel, 16	sample.ctree, 42
Tittlemodel, 10	sample.fn, 43 sampledata, 43
GeneralReversetransposefunction, 17	
GeneralReversetransposefunctiondecoupe,	samplefrompool, 44 SDPSYN2, 45
18	•
Generaltransposefunction, 18	sorttablewithingroup, 47
Generaltransposefunctionsimple, 19	Sparameters.default.f, 49
get_cell, 22	Sparameters.variables.reorder.default,
get_cellrn, 22	50
get_cellXXgroup, 23	treedepth, 51
get_cellXXmarginscount, 24	treetopdf, 52
get_cellXXsplit, 25	Tsampledata, 52
get_missingind, 26	TTsampledata, 53
get_natural.predictors, 26	
get_origin, 27	
get_parent, 28	
get_presentind, 28	
get_var, 29	
get_XXpredecessoratmargin, 30	
<pre>get_XXpredecessorsatmargin, 31</pre>	
getnodesfromrules, 20	
getpredictorsfromcaptureoutput, 20	
getnredictorsfromtree 21	