Package 'dREG'

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Title Detection of Regulatory DNA using PRO-seq, GRO-seq Data(dREG)

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Depends
R (>= 2.14), bigWig (>= 0.2-9), e1071, rphast, snowfall, data.table, rmutil, mvtnorm, methods

LinkingTo

Suggests data.table, Rgtsvm

Description This package is an analysis pipeline for the analysis of GRO-seq data.

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biocViews Sequencing, Analysis

LazyLoad yes

R topics documented:

`combine.roc` *Combines ROC plots*

Description

Combines ROC plots, interpolating and weighting by nTP.

Usage

```
combine.roc(list.roc,
      weight = rep(1, NROW(list.roc)),
      interp.corners = FALSE,
      use.max = FALSE,
      nvals = 100)
```

Arguments

`list.roc`	List including multiple ROC data frame
`weight`	Weight vector for each ROC dataframe
`interp.corners`	Logical value indicating if the header(1,1) and tail values(0,0) are interpolated to each ROC data frame.
`use.max`	Logical value indicating if maximum value of muliple ROCs at same point are used as TPF values.
`nvals`	Integer value indicating interval number for ROC plot.

Value

A data frame with 2 columns is returned

`FPR`	False Positive Rate
`TPR`	True Positive Rate

References

Danko, C. G., Hyland, S. L., Core, L. J., Martins, A. L., Waters, C. T., Lee, H. W., ... & Siepel, A. (2015). Identification of active transcriptional regulatory elements from GRO-seq data. Nature methods, 12(5), 433-438.

See Also

`roc.calc`, `logreg.roc.calc`, `roc.auc`, `roc.plot`

Examples

```
list.roc<-list();

true <- c(rep(1, 100), rep(0, 100));
scores <- c( rnorm(100, 1, 1 ), rnorm(100, 0, 1 ) );
list.roc[[1]] <- logreg.roc.calc( true, scores );
```

```
true <- c(rep(1, 120), rep(0, 110));
scores <- c( rnorm(120, 1, 0.8 ), rnorm(110, 0, 1.2 ) );
list.roc[[2]] <- logreg.roc.calc( true, scores );

r <- combine.roc(list.roc);
roc.plot(r)
```

`eval_reg_svm` *Evaluates a set of genomic coordinates for regulatory potential using P/GRO-seq data*

Description

Evaluates a set of genomic coordinates for regulatory potential using P/GRO-seq data

Usage

```
eval_reg_svm(gdm,
      asvm,
      positions,
      bw_plus_path,
      bw_minus_path,
      batch_size = 50000,
      ncores = 3,
      use_rgtsvm = FALSE,
      debug = TRUE)
```

Arguments

`gdm`	Genomic data model return by `genomic_data_model`.
`asvm`	A pre-trained SVM model from the e1071 package returned by `regulatory_svm`.
`positions`	Data frame with 2 columns indicating the universe of positions to test and evaluate(chrom,chromCenter). It can be returned by `get_informative_positions`.
`bw_plus_path`	String value indicating file path to bigWig file representing the plus strand.
`bw_minus_path`	String value indicating file path to bigWig file representing the minus strand
`batch_size`	Number of positions to evaluate at once (more might be faster, but takes more memory).
`ncores`	Number of CPU cores in parallel computing
`use_rgtsvm`	Indictating whether the predict will be performed on GPU through the Rgtsvm package.
`debug`	Logical value indicating the process detail is outputted.

Value

Returns the value of the SVM for each genomic coordinate specified.

References

Danko, C. G., Hyland, S. L., Core, L. J., Martins, A. L., Waters, C. T., Lee, H. W., ... & Siepel, A. (2015). Identification of active transcriptional regulatory elements from GRO-seq data. Nature methods, 12(5), 433-438.

See Also

get_informative_positions, get_test_set, read_genomic_data, regulatory_svm

Examples

```
## The following codes cannot run without the bigWig files

# ps_plus_path  <- "bigwig.plus.bw"
# ps_minus_path <- "bigwig.minus.bw"

## Now scan all positions in the genome ...
# positions <- get_informative_positions(ps_plus_path, ps_minus_path,
# depth= 0, step=50, use_ANDOR=TRUE, use_OR=FALSE);

# pred_val<- eval_reg_svm( gdm, asvm, inf_positions, ps_plus_path, ps_minus_path, batch_size=50000)
# write.table( data.frame(inf_positions, pred_val), file="eval.tab",
# row.names=FALSE, col.names=FALSE, quote=FALSE, sep="\t")
```

---

genomic_data_model *Creates a genome data model.*

Description

Creates a genome data model.

Usage

```
genomic_data_model(window_sizes, half_nWindows)
```

Arguments

window_sizes	Number indicating the width of genomic window.
half_nWindows	Number indicating the count of genomic window at each side(left side or right side).

Value

A s4 object is returned with

n_zooms	Number indicating zoom ratio.
window_sizes	Vector indicating window sizes.
half_nWindows	Vector indicating number of half windows.

References

Danko, C. G., Hyland, S. L., Core, L. J., Martins, A. L., Waters, C. T., Lee, H. W., ... & Siepel, A. (2015). Identification of active transcriptional regulatory elements from GRO-seq data. Nature methods, 12(5), 433-438.

See Also

read_genomic_data, regulatory_svm, eval_reg_svm

Examples

```
gdm <- genomic_data_model( c(10,20,30), c(10, 10, 10) )
```

`get_informative_positions` *Gets center positions that pass a minimum depth filter*

Description

Returns a data frame with center positions that pass a minimum depth filter

Usage

```
get_informative_positions(bw_path,
      bw_minus_path = NULL,
      depth = 0,
      window = 400,
      step = 50,
      use_OR = TRUE,
      use_ANDOR = TRUE,
      debug = TRUE)
```

Arguments

bw_path	String indicating file path to bigwig file representing the plus strand.
bw_minus_path	String indicating file path to bigwig file representing the minus strand, If specified, takes the windows that pass the step in both bigWig files.(intersection)
depth	Integer value indicating minimum number of reads to return.
window	Integer value indicating window distance between to search for #depth reads [bp].
step	Integer value indicating step distance for window list.
use_OR	Logical value indicating if the center positions in minus bigwig file are merged into the results. If false, the intersection operation will be performed to the center positions of plus bigwig and from minus bigwig.
use_ANDOR	Logical value indicating if the center positions will be merged from the two results. a) Intersection operation with the conditions: window interval=1000 depth>=0. b) Union operation with with the conditions: window interval=100 depth >=2.
debug	Logical value indication the process detail is outputted.

Details

The use_ANDOR and use_OR parameter are applied to two Bigwig files as following logical:

```
if(use_ANDOR){
    v1 <- get_window_Or  (window=1000, depth=0);
    v2 <- get_window_and (window=100,  depth=2);
    vals <- c(v1,v2);
}
else {
  if(use_OR){
     vals <- get_window_Or( window=window, depth=depth);
  }
  else {
     vals <- get_window_and( window=window, depth=depth);
  }
}
```

Value

A BED-style data frame will be returned with 3 columns

`chrom`	Chromosome information
`chromStart`	Start position
`chromEnds`	End position

References

Danko, C. G., Hyland, S. L., Core, L. J., Martins, A. L., Waters, C. T., Lee, H. W., ... & Siepel, A. (2015). Identification of active transcriptional regulatory elements from GRO-seq data. Nature methods, 12(5), 433-438.

See Also

`get_test_set`, `read_genomic_data`, `regulatory_svm`, `eval_reg_svm`

`get_test_set` *Returns a genome loci of positive set and negative set for SVM training purpose.*

Description

Returns a genome loci of positive set and negative set for SVM training purpose

Usage

```
get_test_set(positions,
      positive,
      n_samp,
      allow = NULL,
      enrich_negative_near_pos = 0.15,
      extra_enrich_bed = NULL,
      extra_enrich_frac = 0.1,
      avoid_dist = 100)
```

Arguments

positions	Bed-style data frame indicating the universe of positions to test and evaluate (chrom,chromCenter).
positive	Bed-style data frame containing positive positions (chrom,chromStart,chromEnd).
n_samp	Number of training examples
allow	Bed-style data frame containing inverse negative set of positions (chrom,chromStart,chromEnd).
enrich_negative_near_pos	Fraction of training examples chosen to be nearby (<=5kb) a positive example [0,1].
extra_enrich_bed	Bed-style data frame indicating extra bed file to enrich near.
extra_enrich_frac	Fraction of final positions sampled in the negative set which are in the bed file. Unused if extra_enrich_bed is NULL.
avoid_dist	Integer value indicating how long extend avoiding genomic loci.

Details

(1). The parameter of positions can be obtained by get_informative_positions.

Value

Returns a data frame including double number of the _train set(2*n_samp), each sample includes 4 items.

chrom	
chromStart	
chromEnd	
status	1 for positive and 0 for negative.

References

Danko, C. G., Hyland, S. L., Core, L. J., Martins, A. L., Waters, C. T., Lee, H. W., ... & Siepel, A. (2015). Identification of active transcriptional regulatory elements from GRO-seq data. Nature methods, 12(5), 433-438.

See Also

get_informative_positions, read_genomic_data, regulatory_svm, eval_reg_svm

`logreg.roc.calc` *Calculates the TPR and FPR for a ROC plot.*

Description

Calculates the TPR and FPR for a ROC plot from the status and score vector.

Usage

```
logreg.roc.calc(true, scores)
```

Arguments

`true`	Vector indicating the two status, 1 and 0.
`scores`	Vector indicating the scores for each status calculated by the predict function.

Details

The function of `roc.calc` calculates a ROC matrix for the genomic loci, whereas the function of `logreg.roc.calc` calculates for a status vector.

Value

A data frame with 3 columns is returned, which is same as `roc.calc`.

`FPR`	False Positive Rate.
`TPR`	True Positive Rate.
`threshold`	Threshold based on the score parameter.

References

Danko, C. G., Hyland, S. L., Core, L. J., Martins, A. L., Waters, C. T., Lee, H. W., ... & Siepel, A. (2015). Identification of active transcriptional regulatory elements from GRO-seq data. Nature methods, 12(5), 433-438.

See Also

`roc.calc`, `combine.roc`, `roc.auc`,`roc.plot`

Examples

```
true <- c(rep(1, 100), rep(0, 100));
scores <- c( rnorm(100, 1, 1 ), rnorm(100, 0, 1 ) );
roc_mat <- logreg.roc.calc( true, scores );
AUC<- roc.auc(roc_mat);
roc.plot(roc_mat, main=AUC );
```

peak_calling *Peak calling based on dREG prediction*

Description

This procedure calls SVR prediction for paired bigWig files using pre-trained SVM model and detects divergent peaks based on the predicted score.

Usage

```
peak_calling(asvm, gdm,
      bw_plus_path, bw_minus_path,
      infp_bed = NULL,
      use_rgtsvm = TRUE,
      min_score = NULL,
      pv_adjust="fdr",
      pv_threshold=0.05,
      smoothwidth = 4,
      ncores = 1 )
```

Arguments

asvm	SVR model pre-trained for dREG package, which can be downloaded from the dREG page in Github.
gdm	Genomic data model return by genomic_data_model. This data is binding with SVR model.
bw_plus_path	File name indicating file path to bigWig file representing the plus strand.
bw_minus_path	File name indicating file path to bigWig file representing the minus strand.
infp_bed	A BED data frame indicating informative sites and scores returned by eval_reg_svm. If NULL is specified, the peak calling starts from the informative sites finding and predicting.
use_rgtsvm	Logical value indictating whether the predict will be performed on GPU through the Rgtsvm package.
min_score	Numerical value indicating the minimum dREG score applied to the peak calling procedure. If NULL is specified, this value is calculated based on the predicted scores.
pv_adjust	String value indictating which correction method is used to do multiple comparison, see details in p.adjust, default is 'fdr'.
pv_threshold	Numerical value indicating the threshold is used to report the dREG peaks.
smoothwidth	Numerical value indicating the parameter of curve smooth in the moving average.
ncores	Number of CPU cores in parallel computing.

Value

This function returns a list containing 6 items, including:

1) dREG peaks: peak_bed

chr	Chromosome
start	Start position
end	End position
score	Maxmimum score in the peak region
prob	Probability of multivariate Laplace distribution indicating the probability of the peak points belonging to negative set (No divergent peak).
center	the center position in original peak

2) Informative sites with score infp_bed

chr	Chromosome
start	Start position
end	End position
score	predicted score
infp	indicating the informative site or dense site

3) Broad peak regions peak_broad

chr	Chromosome
start	Start position
end	End position
no	index
min	minimum score in this region
max	maximum score in this region
mean	score mean in this region
sum	score um in this region
stdev	standard deviation of scores in this region
count	informative site in this region

4) Threshold of dREG score min_score

5) Raw results of peak calling raw_peak

6) Summary of peak calling peak_sum

adjust.none.0.05	The count of dREG peak withou p-value correction.
adjust.fdr.0.05	The count of dREG peak adjusted by the 'fdr' method.
adjust.BH.0.05	The count of dREG peak adjusted by the 'BH' method.
adjust.bonferroni.0.05	The count of dREG peak adjusted by the 'bonferroni' method.
adjust.holm.0.05	The count of dREG peak adjusted by the 'holm' method.
adjust.hochberg.0.05	The count of dREG peak adjusted by the 'hochberg' method.
adjust.BY.0.05	The count of dREG peak adjusted by the 'BY' method.
peak.sig.score	The score range of significant dREG peaks.

peak.narrow100 The ignored narrow peaks which length are less than 100.

peak.narrow100.sig
The ignored narrow peaks which may be significant based on 'peak.sig.score'.

peak.narrow100.score
The score range of the ignored narrow peaks.

Examples

```
# load("../asvm.6.6M.20170828.rdata");
# gdm <- genomic_data_model(window_sizes= c(10, 25, 50, 500, 5000), half_nWindows= c(10, 10, 30, 20, 20) )
# bw_plus_path <- "K562.chr21.plus.bw"
# bw_minus_path <- "K562.chr21.minus.bw"
# x <- peak_calling( svm, gdm, bw_plus_path, bw_minus_path, ncores=12, use_rgtsvm=T)
# show(x$peak_bed);
```

---

read_genomic_data *Gets read data from the specified genomic position.*

Description

Gets read data from the specified genomic position.

Usage

```
read_genomic_data( gdm,
      bed,
      file_bigwig_plus,
      file_bigwig_minus,
      as_matrix = TRUE,
      scale.method = c("logistic", "linear"),
      batch_size = 50000,
      ncores = 1 )
```

Arguments

gdm	Genomic data model return by genomic_data_model.
bed	bed-style data frame of genomic regions.(at least 3 columns including chrom, start, end).
file_bigwig_plus	String value indicating file path to bigwig file representing GRO-seq/ PRO-seq reads on the plus strand.
file_bigwig_minus	String value indicating file path to bigwig file representing GRO-seq/ PRO-seq reads on the minus strand.
as_matrix	Logical type,if true, returns a matrix object, otherwise returns a list() object, where each element in the list is the zoom data.
scale.method	String value indicating the normalize method of read counts. Two options are available, "logistic" or "linear", default value is logistic. See details

batch_size	Number of genomic positions to evaluate at once (more might be faster, but takes more memory)
ncores	Number of CPU cores in parallel computing

Details

Data normalize method:
(1): Logistic function: F(x) = 1/(1+exp(-a*(x-b))
(2): Linear function: F(x) = x / tootal_reads

Value

A matrix of normalized read count, the columns are windows list specified by gdm object.

References

Danko, C. G., Hyland, S. L., Core, L. J., Martins, A. L., Waters, C. T., Lee, H. W., ... & Siepel, A. (2015). Identification of active transcriptional regulatory elements from GRO-seq data. Nature methods, 12(5), 433-438.

See Also

get_informative_positions, get_test_set, regulatory_svm, eval_reg_svm

Examples

```
file_bigwig_plus <- "";
file_bigwig_minus <- "";
gdm <- genomic_data_model(20, 10);
#mat <- read_genomic_data(gdm, bed, file_bigwig_plus, file_bigwig_minus);
#summary(mat);
```

---

regulatory_svm *Trains a SVM to recognize a certain pattern of regulatory positions*

Description

Trains a SVM to recognize a certain pattern of regulatory positions.

Usage

```
regulatory_svm(gdm,
      bw_plus_path,
      bw_minus_path,
      positions, positive,
      allow = NULL,
      n_train = 25000,
      n_eval = 1000,
      pdf_path = "roc_plot.pdf",
```

```
        plot_raw_data = TRUE,
        extra_enrich_bed = NULL,
        extra_enrich_frac = 0.1,
        enrich_negative_near_pos = 0.15,
        use_rgtsvm = FALSE,
        svm_type = "SVR",
        ncores = 1,
        ...,
        debug = TRUE)
```

Arguments

gdm	Genomic data model returned by genomic_data_model.
bw_plus_path	String indicating file path to bigWig file representing the plus strand.
bw_minus_path	String indicating file path to bigWig file representing the minus strand.
positions	Data frame with two columns(chrom,chromCenter), indicating the universe of positions to test and evaluate. It can be generated by get_informative_positions.
positive	Bed-style data frame containing positive positions(chrom,chromStart,chromEnd).
allow	Bed-style data frame containing positions to avoid in the negative set(chrom,chromStart,chromEnd).
n_train	Number of training examples.
n_eval	Number of examples on which to test performance.
pdf_path	String value indicating a PDF file. Set to NULL if no PDF should be printed.
plot_raw_data	If TRUE (default), and if a PDF file is specified, plots the raw data used to train the model.
extra_enrich_bed	Bed-style data frame indicating extra bed file to enrich near. Used by get_test_set.
extra_enrich_frac	Fraction of final positions sampled in the negative set which are in the bed file. Unused if extra_enrich_bed is NULL. Used by get_test_set.
enrich_negative_near_pos	Fraction of training examples chosen to be nearby (<=5kb) a positive example [0,1].
use_rgtsvm	Indictating whether the predict will be performed on GPU through the Rgtsvm package.
svm_type	Two options, "SVR" for support vecctor regression (epsilon-regression). "P_SVM" for probabilistic SVM (C-classification).
ncores	Integer indicating how many cores are used to improve the performance.
...	The parameters for plot function
debug	Logical value indication the process detail is outputted.

Value

A svm model trained by svm function in e1071 package.

References

Danko, C. G., Hyland, S. L., Core, L. J., Martins, A. L., Waters, C. T., Lee, H. W., ... & Siepel, A. (2015). Identification of active transcriptional regulatory elements from GRO-seq data. Nature methods, 12(5), 433-438.

See Also

get_informative_positions, get_test_set, read_genomic_data, eval_reg_svm

---

roc.auc *Computes the AUC of a ROC plot.*

Description

Computes the AUC of a ROC plot.

Usage

```
roc.auc(ROC)
```

Arguments

ROC	A matrix with 3 columns (FPR, TPR and threshold) calculated by logreg.roc.calc.

Details

The parameter of ROC is a matrix or data frame including 3 columns, FPR(False Positive Rate), TPR(True Positive Rate) and threshold.

Value

AUC value is returned.

References

Danko, C. G., Hyland, S. L., Core, L. J., Martins, A. L., Waters, C. T., Lee, H. W., ... & Siepel, A. (2015). Identification of active transcriptional regulatory elements from GRO-seq data. Nature methods, 12(5), 433-438.

See Also

roc.calc, logreg.roc.calc, combine.roc, roc.plot

Examples

```
roc_mat <- data.frame( FPR=c(0, 0.25, 0.5, 0.75, 1),
TPR=c(0, 0.5, 0.8, 0.95, 1),
threshold=c(1, 1, 1, 1, 1) );
AUC<- roc.auc( roc_mat );
roc.plot( roc_mat, main=AUC );
```

`roc.calc` *Calculates the TPR and FPR for a ROC plot.*

Description

Calculates the TPR and FPR for a ROC plot.

Usage

```
roc.calc(true,
      possible,
      scores,
      filterPossible = TRUE,
      n_points = 100)
```

Arguments

`true`	Bed-style data frame, a set of 'true' genomic intervals (e.g. ChIP-seq peaks).
`possible`	Bed-style data frame, A set of 'possible' genomic intervals (e.g. DNAse-1 peaks).
`scores`	Vector indicating the scores for each possibe genomic interval in parameter of `possible`.
`filterPossible`	Vector indicating indexes which be removed.
`n_points`	Integer indicating how many points for the ROC plot.

Value

A data frame with 3 columns is returned

`FPR`	False Positive Rate
`TPR`	True Positive Rate
`threshold`	Threshold based on the score parameter.

References

Danko, C. G., Hyland, S. L., Core, L. J., Martins, A. L., Waters, C. T., Lee, H. W., ... & Siepel, A. (2015). Identification of active transcriptional regulatory elements from GRO-seq data. Nature methods, 12(5), 433-438.

See Also

`logreg.roc.calc`, `combine.roc`, `roc.auc`, `roc.plot`

`roc.plot` *Draws a ROC figure.*

Description

Draws a ROC figure.

Usage

```
roc.plot(ROC, ...)
```

Arguments

`ROC`	Matrix or data frame with 3 columns, FPR, TPR and threshold.
`...`	The parameters for `plot` function

Value

None

References

Danko, C. G., Hyland, S. L., Core, L. J., Martins, A. L., Waters, C. T., Lee, H. W., ... & Siepel, A. (2015). Identification of active transcriptional regulatory elements from GRO-seq data. Nature methods, 12(5), 433-438.

See Also

`roc.calc`, `logreg.roc.calc`, `combine.roc`, `roc.auc`

Examples

```
true <- c(rep(1, 100), rep(0, 100));
scores <- c( rnorm(100, 1, 1 ), rnorm(100, 0, 1 ) );
roc_mat <- logreg.roc.calc( true, scores );
AUC<- roc.auc(roc_mat);
roc.plot(roc_mat, main=AUC );
```

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