

Assess pH 7 Leave-Out-Out data

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Background info

The objective of this experiment was to reconstruct the Bayley community to have no (or rather minimized) net growth effects. This community eliminates stimulatory effects of deacidification by growing the community at pH 7 and also eliminated inhibitory effects due to *Penicillium* JBC by removing it from the community. Comparison of the growth each species grown alone at pH 7 to growth in the community lacking *Penicillium* will reveal any additional interactions not accounted for by these two growth affectors.

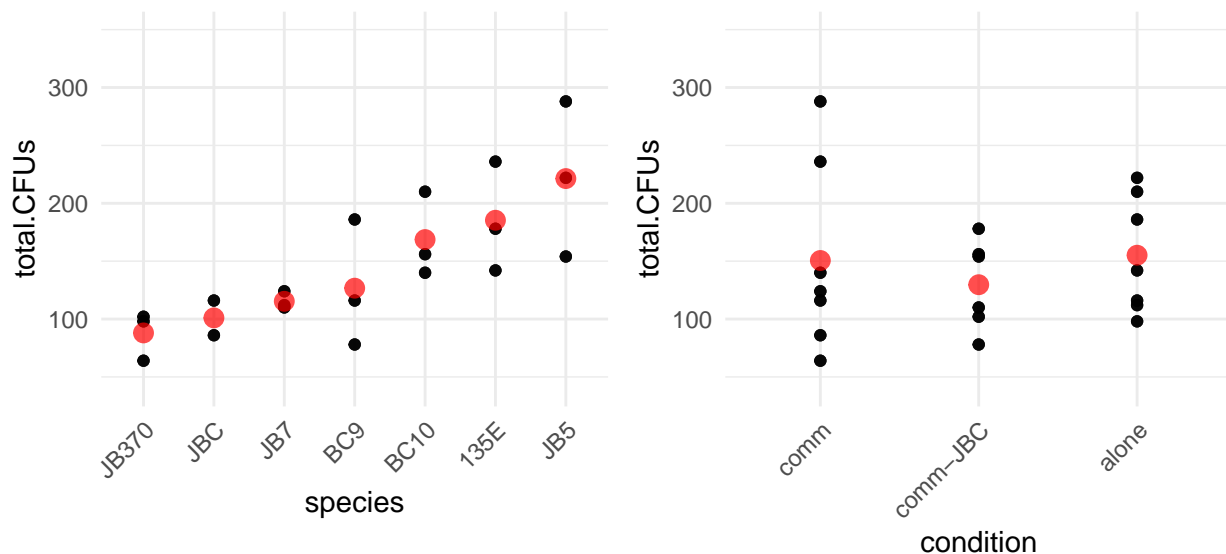
Community conditions

- none (“community” or “comm”)
- *Penicillium* JBC (“comm-JBC”)
- each of 7 Bayley species grown alone

```
d.1007 <- readRDS(here("wrangled_data/leave-one-out_pH7.rds"))  
d.18 <- readRDS(here("wrangled_data/all_pairwise.rds"))
```

Inoculation quality assessment

Are the inoculations around 200 CFUs each? Any notable biases?



Representative inoculation of each species in each condition in black, and the mean within species (left) or within condition (right) in red.

So there's definitely some species that were inoculated lower than target (mean 90-130 CFUs), and some that were just under the target (mean 160-190 CFUs) but none that were super systematically overinoculated

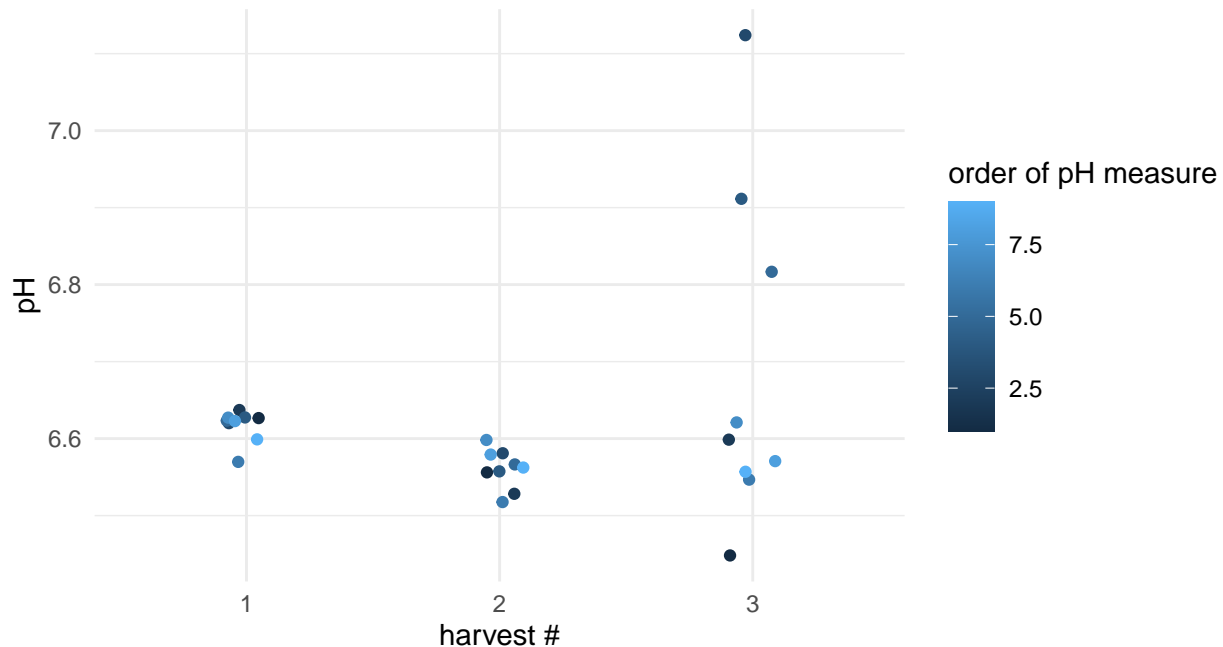
(Brevi average ~220 CFUs).

The average inoculation within each community was a little low, between 130 - 160 CFUs.

pH probe variability

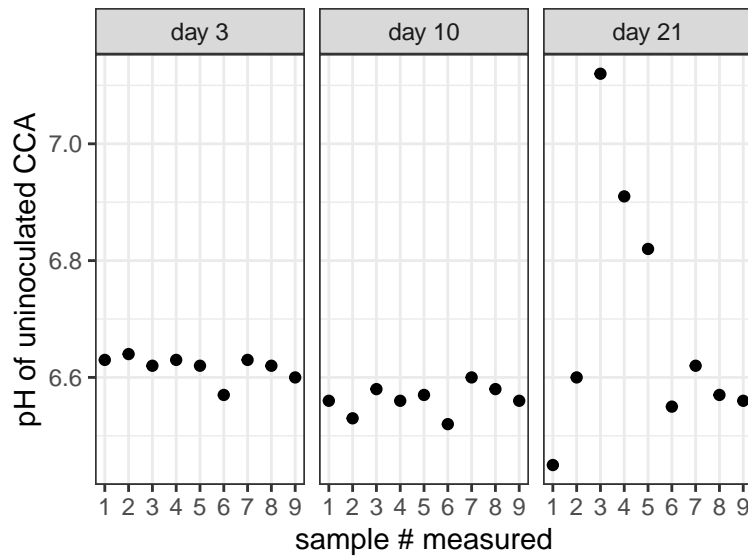
Interested in seeing whether pH of “control” (uninoculated but from same 96-well plate as community samples) cheese at time of harvest is changing at different timepoints, given that I’ve found strong ability of some Bayley species to raise the pH of the media through volatiles alone (namely, *Penicillium*, *Brevibacterium*, and *Scopulariopsis*, and this effect gets stronger between 7 & 10 days of growth).

Note that each “replicate” was the pH of uninoculated cheese at harvest day 3 (rep./harvest # 1), day 10 (rep./harvest # 2), and day 21 (rep./harvest # 3).



So what I’m really seeing here is just a bias according to pH re-calibration (each calibrated at day 3 and day 10 and day 21). Day 21 shows greater variability because of VOC effects after 21 days of neighboring well community growth.

I also want to check that there isn’t some systematic pH change over time of use.

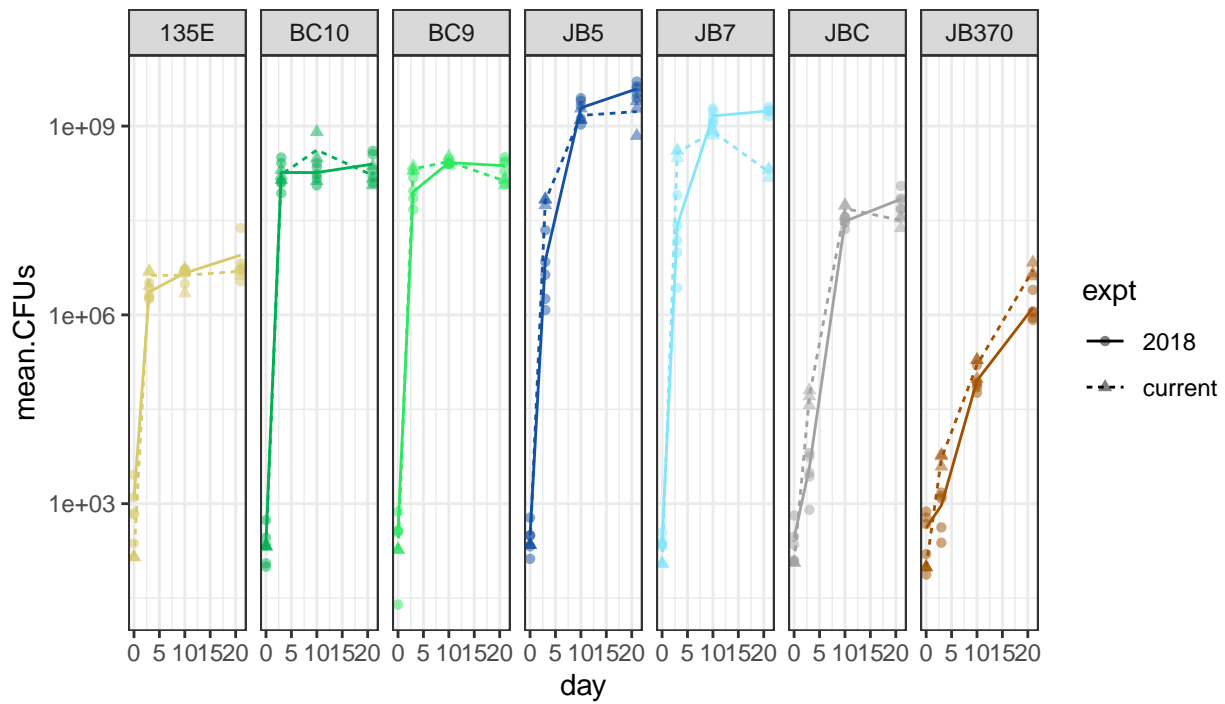


Again, from early timepoints, I can't help but feel there is like some small bias towards pH measurement decline over time. Obviously the effects of volatiles on the pH after 21 days is too great to get a sense for this anymore

Is complete community & alone data replicating what has been seen before (i.e. 08_2018)

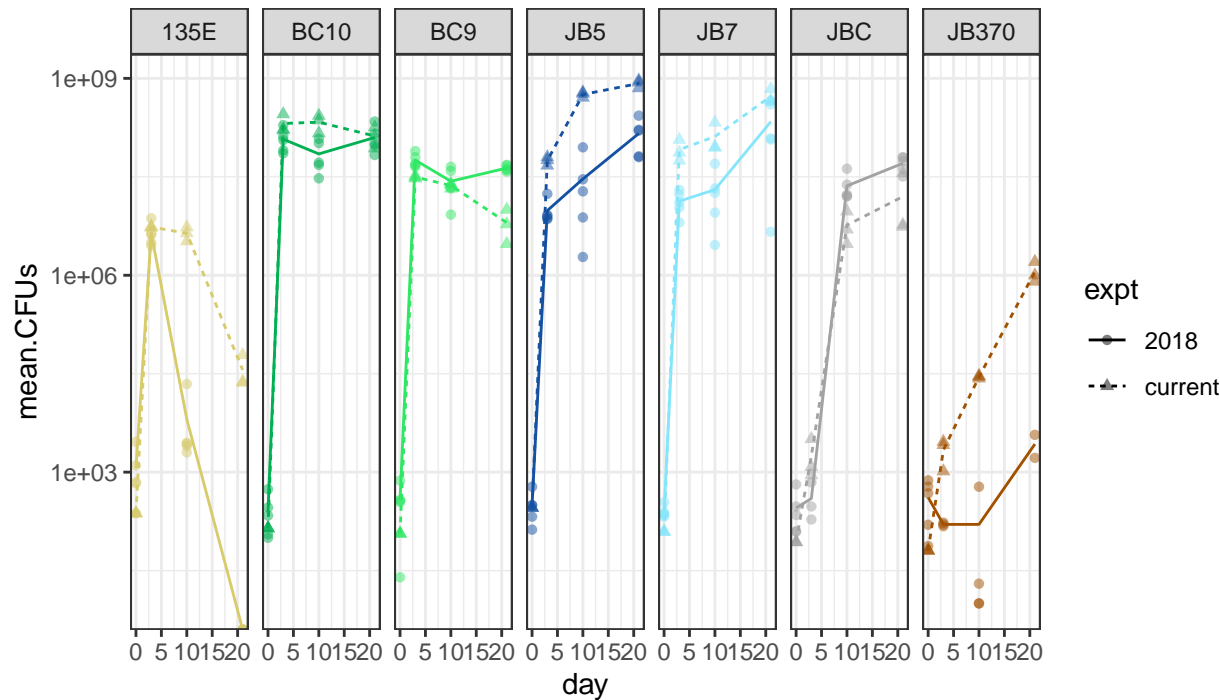
Compare ALONE growth:

Growth alone in this experiment compared to Aug. 2018



Compare community growth:

Growth in community in this experiment compared to Aug. 2018



Compare community succession pattern:

Relative composition of Bayley community bacteria and fungi

